

1 **Title:** *Ralstonia* spp: Emerging Global Opportunistic Pathogens

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3 **Authors:** Michael P Ryan<sup>1</sup> and Catherine C Adley<sup>1\*</sup>

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5 **Name and Address of Institute at which the work was performed:**

6 <sup>1</sup>Microbiology Laboratory Department of Chemical and Environmental Sciences, University

7 of Limerick, Limerick, Ireland

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11 **\* Correspondent footnote:**

12 Michael P Ryan

13 Department of Chemical and Environmental Sciences

14 University of Limerick

15 Limerick

16 Ireland

17 Tel: +353 61 202448

18 Fax: +353 61 202568

19 E-mail: [Michael.P.Ryan@ul.ie](mailto:Michael.P.Ryan@ul.ie)

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## 26 **Abstract**

27 The bacterial genus *Ralstonia* (Gram negative non-fermenters) is becoming more prevalent in  
28 cases of infection with three bacterial species *Ralstonia pickettii*, *Ralstonia insidiosa* and  
29 *Ralstonia mannitolilytica* making up all cases reported (in the literature) to date. These  
30 organisms are prevalent in many different types of water supplies (including hospital water  
31 supplies) being well adapted to survive in low nutrient conditions. They have been shown to  
32 cause infections, sometimes serious such as osteomyelitis and meningitis, in hospital settings.  
33 Seventy cases of *R. pickettii*, 13 cases of *R. mannitolilytica* and 3 cases of *R. insidiosa*  
34 infection have been identified from the literature. Insight is given into the types of infections  
35 that are caused by these bacteria, the underlying conditions that are associated with these  
36 infections and potential treatments.

## 37 **Introduction**

38 Gram-negative non-fermenting bacteria are a growing concern in clinical environments, being  
39 one of the most common causes of nosocomial infections. The major opportunistic pathogens  
40 from this group comprise of many bacterial species including: *Acinetobacter baumannii*,  
41 *Pseudomonas aeruginosa*, *Burkholderia cepacia*, and *Stenotrophomonas maltophilia*. These  
42 bacteria take advantage of underlying medical conditions and diseases to cause infection.  
43 *Ralstonia* spp are amongst these non-fermenting gram-negative bacteria that are emerging as  
44 opportunistic pathogens.

45 *Ralstonia* is a genus which includes former members of *Burkholderia* spp. (*Burkholderia*  
46 *pickettii* and *Burkholderia solanacearum*). *Ralstonia* spp. are aerobic Gram-negative, non-  
47 fermentative rods that can be found in water and soil (1). *Ralstonia pickettii*, the type species  
48 of the genus, was until recently regarded as the only member of the genus of clinical  
49 importance (2) however two novel *Ralstonia* spp. have been described recently that are  
50 considered to be of minor clinical importance, *Ralstonia insidiosa* (3) and *Ralstonia*

51 *mannitolilytica* (2). Investigation of instances of *Ralstonia* spp. infections in the literature  
52 showed many different types of infections (mostly with *R. pickettii* as the cause). This  
53 indicates that the genus may be a more extensive pathogen than was assumed previously with  
54 the types of infections being more invasive and severe.

## 55 **Bacteriology**

### 56 ***Ralstonia pickettii***

57 As stated above *R. pickettii* is the type species of the *Ralstonia* genus. The bacterium has been  
58 recovered from a wide variety of clinical specimens including blood, urine and cerebrospinal  
59 fluid (4). The bacteria has been isolated from numerous water sources including municipal  
60 drinking water supplies (5), bottled water (6), dental water supplies (7), hospital water  
61 supplies (8), space shuttle water systems (9), standard purified water (10), laboratory based  
62 high-purity water systems (11) and industrial Ultra-pure/High Purity water (12, 13, 14). *R.*  
63 *pickettii* has been identified in biofilm formation in plastic industrial water piping (13, 15). *R.*  
64 *pickettii* is not thought of as a major pathogen and its virulence is thought to be low. It is  
65 consequently not routinely looked for in hospital analysis (2). All reported instances of  
66 infection attributed to *R. pickettii* infection are described in Table 1.

### 67 ***Ralstonia insidiosa***

68 *Ralstonia insidiosa* is the closest related bacteria to *R. pickettii* (16) and has also been isolated  
69 from river and pond water, soil, activated sludge (16), laboratory purified water systems (3),  
70 and industrial Ultra-pure/High Purity water (16) and water distribution systems (18). The  
71 bacteria has also been found in the lung sputum of cystic fibrosis suffers (2). All reported  
72 instances of infection attributed to *R. insidiosa* infection are described in Table 2.

### 73 ***Ralstonia mannitolilytica***

74 *R. mannitolilytica* is also closely related to *R. pickettii*. *R. mannitolilytica* had previously been  
75 called “*Pseudomonas thomasi*” and *R. pickettii* biovar 3/“*thomasi*” (19) and has been

76 described in a number of hospital outbreaks (20, 21, 22, 23). Initial reports (24) described  
77 cases of bacteremia and bacteriuria in twenty patients. These infections were linked to “*P.*  
78 *thomasi*” contaminated parenteral fluids that had been prepared with deionized water  
79 contaminated with “*P. thomasi*”. Pan *et al.*, reported an epidemic involving twenty-four  
80 patients that was caused by “*P. thomasi*.” contaminated saline solution that had been  
81 prepared by the hospital pharmacy (23). Pseudo-outbreaks linked to *R. mannitolilytica* have  
82 also been described (25). Reports have described it as causing meningitis and  
83 hemoperitoneum infection (26), renal transplant infection (27) and bacteremia (28). *R.*  
84 *mannitolilytica* is the most prevalent species of the *Ralstonia* genus to be found in Cystic  
85 Fibrosis suffers (29). All reported instances of infection attributed to *R. mannitolilytica*  
86 infection are described in Table 3.

### 87 **Identification of *Ralstonia* spp.**

88 The three *Ralstonia* spp. are difficult to identify and differentiate from one another using  
89 routine hospital analysis (3, 28). The three bacteria have very similar biochemical patterns to  
90 each other and to other bacterial species such as the *Burkholderia cepacia* complex and  
91 *Pseudomonas fluorescens* (2, 3, 28). The existing commercial biochemical identification  
92 systems on the market, e.g. API 20NE, RapID NE Vitek etc., do not give the best  
93 identification, especially for some genera or species including Gram-negative non-fermenting  
94 rods (30). Some identification methods may confound epidemiology as *Ralstonia* spp. can be  
95 easily misidentified as *Burkholderia cepacia* complex and *Pseudomonas fluorescens* using  
96 routine methods as stated above.

97 *R. pickettii* has been shown to give variable results using the standard biochemical test kits,  
98 e.g. API 20NE and *R. mannitolilytica* and *R. insidiosa* have not been added to test panels (13).

99 The best way to tell the bacteria apart is though PCR with Species -Specific PCR primers  
100 available for all three bacteria (17, 31).

## 101 **Epidemiology**

102 The population at risk from these bacteria comprises almost exclusively immunocompromised  
103 patients with for example those with hematological malignancies (bacteremia, septicemia,  
104 etc.), patients in intensive care, patients with indwelling devices such as central venous  
105 catheters and neonates (32, 33, 34, 35, 36, 37, 38). Examples of underlying conditions and  
106 *Ralstonia* spp. infection includes: a 53-year-old man who had suffered a myocardial infarction  
107 and who contracted *R. pickettii*-related bacteremia while recuperating in hospital; (30) a 32-  
108 year-old man who was suffering from hepatitis-C-related liver cirrhosis who contracted *R.*  
109 *pickettii*-related peritonitis (3), a 41-year-old man with underlying diabetes mellitus who  
110 developed *R. pickettii* related pneumonia (39), a 7-year-old boy who received a cord blood  
111 transplant and then suffered an episode of *R. pickettii*-related bacteremia (40) and a 71-year-  
112 old man with underlying chronic renal failure, diabetes mellitus, hypertension and alcoholic  
113 cirrhosis who developed *R. pickettii*-related osteomyelitis (41).

114 *R. pickettii* and *R. mannitolilytica* have also been associated in infections with permanent  
115 indwelling intravenous devices such as catheters (33, 42).

116 *Ralstonia* spp. have been shown to be the causative agent of severe invasive infections  
117 including osteomyelitis, septic arthritis, meningitis, etc. (26, 41, 43). Another example of  
118 severe infection due to *R. pickettii* is the case of a 38-year-old female who contracted both  
119 bacteremia and meningitis (44). In many of the more serious infections the aetiology was  
120 unknown.

121 Reports that have indicated the occurrences of *Ralstonia* spp. infections (Tables 1, 2, 3) are  
122 increasing. This rise could be associated with the increase in the patient population at risk  
123 (older populations, people surviving with serious illness for longer, etc.), potentially as a  
124 result of the widespread use of antibiotics and the implementation of more invasive medical  
125 practices (2).

126

127 **Factors responsible for infection**

128 **Contaminated solutions**

129 Many of the cases of infection with *Ralstonia* spp. are due to contaminated solutions  
130 including water for injection, saline solutions made with purified water, respiratory solutions  
131 and sterile drug solutions (37, 45, 46, 47). Contamination of the products listed above  
132 generally occurs at the manufacturing stage. This contamination can occur through many  
133 different means but one of the most important is due to the ability of *Ralstonia* spp to pass  
134 through 0.2 µm filters that are used for the sterilization of many medicinal products, such as  
135 saline solution (48, 49). Another example of how solutions can become contaminated during  
136 the manufacturing stage was seen in 1983, when five infants became infected with *R. pickettii*  
137 that was associated with a contaminated respiratory therapy solution. *R. pickettii*  
138 contaminated the solution due to the bypassing of an 82°C holding tank during the  
139 manufacture of distilled water. This contaminated solution was then used for endotracheal  
140 suctioning and this allowed colonization of the patients (45).

141 **Disinfectants**

142 *R. pickettii* has been shown to survive in different hospital disinfectants including  
143 chlorhexidine. In 1983, six patients contracted *R. pickettii* related septicemia. The source of  
144 the contamination was traced back to the bidistilled water used to make up the 0.05% aqueous  
145 solution of chlorhexidine. This was then used for skin antisepsis before the insertion of a  
146 venous catheter and lead to infection (50). Contaminated chlorhexidine was also implicated in  
147 the recovery of *R. pickettii* from six pediatric patients in 1995 (51). As before the  
148 contaminated chlorhexidine was used for topical disinfection. *R. pickettii* has also been shown  
149 to survive ethacridine lactate (aromatic organic compound based on acridine) (52).

150 **Pseudo-outbreaks**

151 *Ralstonia* spp. have been linked with pseudo-outbreaks, with seven such outbreaks being  
152 described in the literature (Table 1 and 3). These pseudo-outbreaks have caused unnecessary

153 treatments to be given to patients (e.g. wrong or unneeded antibiotics or the removal of  
154 indwelling devices) and are also a waste of valuable time and resources in hospital  
155 laboratories. Pseudo-outbreaks may be due to many different causes including contaminated  
156 distilled water used in the bacterial testing procedures, phlebotomist error or contamination at  
157 the manufacturing stage of materials used in laboratory testing. Verschraegen *et al.* reported  
158 that *R. pickettii* was the cause of pseudobacteraemia in a surgical ward. This was the first such  
159 case reported in the literature (53). The patients did not exhibit any signs of bacteremia, even  
160 though the organism was isolated from blood samples. The contamination was traced to  
161 distilled water (used in the testing procedures) and a 0.5% chlorhexidine solution (used to  
162 wipe the bench) prepared using the distilled water. Both of these were found in the hospital  
163 pharmacy. *R. mannitolilytica* has also been found to be the causative agent of pseudo-  
164 outbreaks in special-care baby wards (25).

## 165 **Treatment**

166 The treatment and management of *Ralstonia* spp. infections is often challenging as these  
167 pathogens are frequently resistant to numerous different types of antibiotics, including several  
168  $\beta$ -lactams and most of the aminoglycosides. A major study into the antibiotic resistance of  
169 numerous strains of *R. pickettii* and *R. insidiosa* showed both bacteria were highly resistant to  
170 the aminoglycoside gentamicin and the  $\beta$ -lactam antibiotic aztreonam and variably resistant to  
171 the ticarcillin-clavulanic acid mix (54). Two inducible  $\beta$ -lactamases in *R. pickettii*, *bla*<sub>OXA-60</sub>  
172 and *bla*<sub>OXA-22</sub> are considered to be the reason for the high level of resistance to  $\beta$ -lactams (55,  
173 56). The presence of an aminoglycoside acetyl-transferase in *R. pickettii* genomes 12J and  
174 12D also has an aminoglycoside acetyl-transferase accounts for the widespread resistance to  
175 aminoglycosides (see Table 1, and reference 54). The study carried out by Ryan and Adley  
176 showed that most strains were susceptible to the carbapenem meropenem and all isolates were  
177 susceptible to the quinolones (ciprofloxacin and ofloxacin), the tetracyclines (tetracycline and

178 minocycline), the cephalosporins (cefotaxime and ceftazidime), the folate pathway inhibitors  
179 (trimethoprim/sulfamethoxazole) and the extended spectrum beta-lactam antibiotic of the  
180 ureidopenicillin class (piperacillin) (56). Various different antibiotics have been used to treat  
181 infections of *Ralstonia* spp. including trimethoprim/sulfamethoxazole, ciprofloxacin,  
182 piperacillin, cefotaxime, ceftriaxone, imipenem, meropenem, levofloxacin and cefepime  
183 (Table 1, 2 and 3). What can be seen from this data is the failure of meropenem and  
184 ceftriaxone to treat infection in several instances and the need to use an alternative which was  
185 mostly ciprofloxacin.

## 186 **Conclusions**

187 *Ralstonia* spp. are not recognized as major pathogens. However in this paper we have  
188 identified over a hundred examples of *Ralstonia* spp. infections. These species have certain  
189 characteristics such as resistance to disinfection practices and ability to survive in water  
190 supplies that allows them to cause many potentially harmful infections and death.

## 191 **Methodology**

192 All available publications discussing *R. pickettii*, *R. insidiosa* and *R. mannitolilytica*  
193 infections were searched for using the PubMed, Web of Knowledge, Science Direct and  
194 Google Scholar search databases from 1970 to 2013. The terms '*Ralstonia pickettii*,  
195 '*Pseudomonas pickettii*', '*Burkholderia pickettii*', '*Ralstonia*' and '*pickettii*' '*Ralstonia*  
196 '*insidiosa*' '*insidiosa*' '*Ralstonia mannitolilytica*' '*mannitolilytica*' '*Pseudomonas thomasi*'  
197 were all searched for and any references that discussed infection were set aside. These  
198 papers/abstracts were then read and the required information extracted (year of outbreak,  
199 number/age/sex of patients, geographic location, underlying condition, condition caused by  
200 infection, susceptibility/resistance to antibiotics, antibiotics used to treat condition, clinical  
201 outcome). The reference sections of these publications were also checked for any infection  
202 related references that may have been missed during the electronic searches.



203

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208 ***Conflict of interest***

209 The authors declare that they have no competing interests.

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