



Newborn screening for cystic fibrosis

Jeffrey S. Wagener^{a,b}, Edith T. Zemanick^{a,b}, and Marci K. Sontag^c

Purpose of review

Newborn screening for cystic fibrosis (CF) is now universal in the US and many other countries. The rapid expansion of screening has resulted in numerous publications identifying new challenges for healthcare providers. This review provides an overview of these publications and includes ideas on managing these challenges.

Recent findings

Most CF newborn screening algorithms involve DNA mutation analysis. As screening has expanded, new challenges have been identified related to carrier detection and inconclusive diagnoses. Early descriptions of infants with CF-related metabolic syndrome (CRMS) indicate that the natural history of this condition cannot be predicted. Early identification has also provided an opportunity to better understand the pathophysiology of CF. However, few studies have been conducted in infants with CF to determine optimal therapy and recommendations are largely anecdotal.

Summary

Newborn screening provides an opportunity to identify and begin treatment early in individuals with CF. Whereas a single, optimal approach to screening does not exist, all programs can benefit from new findings regarding sweat testing, carrier detection, early pathophysiology, and clinical outcomes.

Keywords

CFTR related metabolic syndrome, cystic fibrosis, immunoreactive trypsinogen, mutation analysis, newborn screening

INTRODUCTION

The US reached a milestone in 2010 when Texas established a newborn screening (NBS) program for cystic fibrosis (CF), making screening universal across all 50 states and the District of Columbia. This achievement followed the statement by the Centers for Disease Control (CDC) in 2004 that universal newborn screening for CF was justified based on the long-term benefits from early nutritional treatment [1]. Additionally, the majority of parents support the value of newborn screening and have encouraged states to institute universal screening programs [2]. However, with universal CF screening new issues have arisen. Multiple different CF screening protocols exist, although most include genetic mutation analyses. Choice of protocol is often based on the balance between the risk of false-negative results and the added costs of managing false-positive results. Genetic testing necessarily includes asymptomatic carrier detection, which newborn screens for other diseases generally do not include. Additionally, some mutations are associated with milder disease phenotypes and others are associated with inconclusive diagnoses

for which the natural history is unknown. Finally, whereas screening for CF allows early patient identification and has expanded our understanding of early pathophysiology, there is a paucity of therapeutic studies in CF infants, and management guidelines are based on limited scientific data.

IMMUNOREACTIVE TRYPsinOGEN AND SWEAT TESTING

The first stage of all CF newborn screening programs involves measuring immunoreactive trypsinogen (IRT), a pancreatic-enzyme precursor whose concentrations are persistently elevated in the blood of

^aUniversity of Colorado School of Medicine, ^bChildren's Hospital Colorado and ^cColorado School of Public Health, University of Colorado Denver, Aurora, Colorado, USA

Correspondence to Jeffrey S. Wagener, MD, University of Colorado School of Medicine, Children's Hospital Colorado, 13123 east 16th Ave. Aurora, CO 80045, USA. Tel: +1 720 777 6181; e-mail: jeffrey.wagener@ucdenver.edu

Curr Opin Pediatr 2012, 24:329–335

DOI:10.1097/MOP.0b013e328353489a

KEY POINTS

- Newborn screening for CF algorithms vary, based on the balance between the risk of false-positive and false-negative results.
- Carrier detection, unique to algorithms with DNA mutation analysis, requires additional attention to educating affected individuals.
- CF-related metabolic syndrome includes individuals with gene mutations and intermediate (or normal) sweat chloride values.
- Anatomic and physiologic changes occur early in the lives of infants with CF.
- Care guidelines for infants with CF are based on limited scientific data and studies of therapies in infants are needed.

infants with CF [3]. The second stage involves either DNA mutation analysis (IRT/DNA) or obtaining a second IRT (IRT/IRT) and looking for persistent elevation. Whereas there is no standardized IRT detection cut-off level, using a lower cut-off improves test sensitivity and adding the mutation analysis improves positive predictive value. However, using lower cut-off levels increases the number of false-positive tests and each program must determine the balance of sensitivity and false-positive results [4]. There are two isoforms of IRT and programs vary as to which form they measure. Lindau-Shepard and Pass [5] showed that measuring both isoforms resulted in comparable performance. To further aid in reducing the number of false-positive tests, pancreatitis-associated protein (PAP) is being investigated in combination with the IRT [6]. PAP is a secretory protein which increases after sustained pancreatic stress and is elevated in neonates with CF [7]. Whereas false-positive results for both IRT and PAP have been reported in infants with renal failure [8], this approach has been proposed to avoid using DNA analysis. A second approach to decreasing false-positive results is to add mutation analysis to the IRT/IRT method [9]. This also allows a lower IRT cut-off level, improving sensitivity while reducing the number of false-positive results related to carrier detection [10¹¹]. In a recent review of CF newborn screening, Castellani and Massie [11¹²] commented that 'although there is not one universal CF newborn screening protocol that will suit the heterogeneous needs of diverse regions, many options for adjusting algorithms to local conditions are now available'. Most importantly, every program needs to monitor its data regularly and conduct ongoing

quality improvement to optimize patient outcomes [4,12,13].

Following detection by newborn screening, the diagnosis of CF should be confirmed by measuring sweat electrolytes, even when two disease-causing mutations are identified [14]. The sweat test, however, may be nondiagnostic due to intermediate chloride values or insufficient quantity of sweat (quantity not sufficient (QNS)). The problem of intermediate chloride values may be a greater problem in programs using the IRT/DNA method since CF carriers (who naturally have slightly higher sweat chloride values) are detected [15]. Whereas the goal of screening is to identify patients with CF at the earliest age, younger and smaller patients are more likely to have QNS results [16]. Thus special attention is needed to perform accurate sweat testing so that QNS rates are at acceptable levels [17¹⁸]. For infants with an initial QNS sweat test, a nasal potential difference can be successfully measured at select referral centers [18]. Otherwise repeat sweat testing can be done, assuring that the child is well hydrated at the time of the test. Whereas some programs have added sweat conductivity measurement to quantitative chloride measurements to help with diagnosis, conductivity has a higher rate of false-positive results [19]. Importantly, sweat test results need to be interpreted in view of the patient's genotype and phenotype to make the diagnosis of CF [20,21].

FALSE-POSITIVE SCREENING AND CARRIER DETECTION

The most common CF gene mutation worldwide is the F508del. Other mutations vary greatly between different populations, creating a need for different DNA screening algorithms [22–25]. Screen sensitivity improves by increasing the number of mutations, but this also increases the number of carriers detected (false-positive results). Studies of the emotional impact of false-positive screen results show that parents may have persisting concerns about the test's accuracy, their child's health, and the implications of having a genetic mutation [26²⁷]. This perception of health vulnerability persists with the parents for at least the first year, during which time infants who are CF carriers have a higher frequency of reported medical problems compared with noncarrier controls [27²⁸]. One way to alleviate anxiety is to shorten the time between notifying the family of a positive screen and confirming a diagnosis [28]. During this time parents often seek information from the internet or their family physician [29], and programs need to provide ongoing education to assure that healthcare providers are knowledgeable about false-positive and negative screen results [30–32].

False-positive results happen with both IRT/DNA and IRT/IRT screening algorithms; however, carrier detection is a challenge unique to programs using DNA testing. Since the initial IRT cut-off level can be lower, DNA testing decreases false-negative rates, but this must be balanced with the need to address carrier detection. Some studies suggest that one way to reduce carrier misconceptions and to improve a family's understanding is to provide structured genetic counseling at the time of the sweat test [33–35]. However, follow-up after genetic counseling with parents of carriers indicates that, whereas 94% understand their child does not have CF, only 79% understand that their child carries the CF gene and fewer than half of the parents or relatives of a carrier infant expressed any interest in personal testing [36]. Additional telephone follow-up by the program is supported by families and appears necessary to provide information and correct misconceptions about carrier status [37].

Some CF screening programs avoid the problems of carrier detection by performing two IRT measurements followed by a sweat test. This requirement for two samples (IRT/IRT) results in slight delays before the definitive diagnosis can be made (median 4.0 weeks vs. 2.3 weeks with IRT/DNA) [38]. However, the median time to diagnosis is still below the 7 weeks reported in the Wisconsin randomized, controlled newborn screening trial in which clinical benefit was demonstrated [39]. The IRT/IRT approach also has a higher risk of false-negative results due to higher initial IRT cut-off values, although there does not appear to be a significant delay in diagnosing patients with false-negative newborn screening results [40]. What is clear is the need for efficient follow-up of positive screening results and a rapid referral to a care center where not only the patient, but also the family, can have their medical and emotional needs met [41].

MILD PHENOTYPES AND INCONCLUSIVE DIAGNOSTIC RESULTS

DNA testing algorithms may include mutations which result in milder disease phenotypes or are associated with inconclusive diagnoses for which the natural history is unknown. Inclusion of these genetic variants can create confusion, the potential for misdiagnosis, and unclear implications of the diagnosis [42,43]. For programs with little genetic variation in the population, only a limited number of mutations need to be included in the screen [44]. Expanding the panel of genetic variants in these populations will have little impact on detection rate due to the infrequency of additional mutations [42]. In other populations with greater racial and genetic

variation, expanding the panel may allow other changes in the program such as negating the need for a 'failsafe' testing of particularly elevated IRT values [4,45]. However, in both situations careful consideration of which mutations to include is essential for appropriate genetic counseling and management. Of particular interest has been the inclusion of the R117H mutation. In some populations this mutation has a naturally high frequency and results in an excess number of false-positive screen results [46]. Additionally, even in combination with more typical CF mutations the impact of the R117H is altered by the intron-8 poly-T status of the patient [47].

Another consequence of expanded DNA testing is identifying infants with an inconclusive diagnosis. Specifically, these are infants with a positive newborn screen which includes one or two mutations, but physiologic measures such as the sweat test are not diagnostic of classical CF. This condition has been referred to as the cystic fibrosis-related metabolic syndrome (CRMS). Over time the sweat test may become abnormal and some of these individuals develop clinical signs of CF in later life, although the disease is generally mild [48**]. Close monitoring has been proposed, although avoiding exposure to other CF patients may be important to reduce exposure to infectious agents [49].

CLINICAL OUTCOMES FROM CYSTIC FIBROSIS NEWBORN SCREENING

Improved nutrition was the primary benefit of CF newborn screening identified in the randomized, controlled trial conducted by Farrell *et al.* [39] and continues to be the most significant outcome of early detection [50]. Using additional data from this trial, Tluczek *et al.* [51] reported no difference in pulmonary function and quality-of-life outcomes between screened and control patients when controlling for pancreatic function and mucoid *Pseudomonas aeruginosa* infection. These results, however, differ from previous epidemiologic data indicating a protective benefit of newborn screening for nutritional state and pulmonary function, as well as reduced complications [52,53]. There is some evidence of improved survival in CF patients identified by newborn screening; however, this study used historical controls during a period in which there have been other significant improvements in CF care [54*].

Even with early diagnosis and long-term preventive care in specialized CF care centers, differences exist in outcomes between programs. A comparison of US and French patients with CF detected by newborn screening demonstrated differences in

the extent of lung disease [55[■]]. The screening protocols, median age at diagnosis, pancreatic function and genotypes were similar; however, the French children had a more rapid progression of lung disease based on chest radiographs when compared with CF patients in Wisconsin. This finding was further supported by the patients having significant differences in lung function (forced expiratory volume in 1 s) between 6 and 12 years of age (83 ± 19 vs. $93 \pm 18\%$ predicted at age 12 for France and Wisconsin, respectively). These differences partially disappeared when patient weight or hospitalization data was added to the estimating model, suggesting there might be other risk factors for deterioration associated with region. The authors did not identify any specific risk factors, but suggested that nutrition, environment, or the healthcare delivery system might be contributing.

One unexpected potential consequence of newborn screening is the potential increased cost of caring for young children with CF. Between 2001 and 2007 the annual cost of treating a child below 11 years of age with CF increased nearly 10-fold based on private insurance payments [56]. Whereas these increased costs were not the result of newborn screening, the earlier age at diagnosis and recommendations for close monitoring of all infants with CF predict that the overall cost for care will continue to increase [57,58[■]].

An additional outcome of newborn screening may be a decreasing prevalence of CF due to at-risk couples obtaining prenatal testing [59[■]]. Early detection of an infant with CF provides the opportunity to identify adult relatives at risk for having a child with CF, but, interestingly, nonparent adult relatives, particularly males, do not commonly pursue testing [60].

EARLY PATHOPHYSIOLOGY

Newborn screening and early identification provide the ability to evaluate CF patients before they develop clinical disease. Chest computed tomography (CT) scanning has the strongest association with later development of lung disease in patients with CF [61]. Limited CT scans have a lower ionizing radiation dose and quantitative anatomic changes associate with future lung disease, plus these anatomic changes correlate with early airway inflammation [62]. Lung function can be measured by a variety of techniques, including the lung clearance index (LCI), which measures distribution of ventilation and detects early differences between infants with CF and healthy controls [63]. Chest CT and LCI are only weakly associated in infants with CF, suggesting that early

anatomic changes precede functional changes [64].

Infection and airway inflammation have been identified in asymptomatic infants with CF [65]. Airway inflammation is associated with eventual lower lung function, and airway infection is associated with a greater decline in lung function over time [66[■]]. Additionally, airway inflammation is associated with the nutritional status of infants and young children with CF [67[■]]. Surveillance bronchoscopy demonstrates bacterial infection in 27%, neutrophilic inflammation in 67% and reflux in 42% of infants with CF during the first 6 months of life [68]. However, in a well controlled clinical trial of routinely scheduled bronchoscopy, Wainwright *et al.* [69[■]] showed no difference between patients managed with and without routine bronchoscopy.

Infants detected by CF newborn screening develop early infection with *Staphylococcus aureus* followed often by *P. aeruginosa* [70]. Bacterial acquisition does not seem to be affected by cohorting patients into *P. aeruginosa*-positive and negative clinics [71[■]], although previous studies have indicated a higher risk of infection when infants are seen in the same location as older patients [72]. Early *P. aeruginosa* detection may be aided by the use of antibody testing, although several tests exist and the optimal antibody is still not defined [73].

Finally, whereas the earliest identified benefit of NBS for CF is improved nutrition, even with screening, infants have less than normal growth [39]. Low levels of insulin-like growth factor 1, first identified in the pig model of CF and confirmed in infants identified by newborn screening, may partially account for this finding [74[■]].

THERAPY

There are almost no studies of specific therapies in infants with CF. Whereas guidelines have been developed to recommend management, these are based mainly on expert recommendations and not evidence-based results [57,58[■],75].

Jadin *et al.* [76[■]] recently noted that infants fed exclusively breast milk during the first 2 months of life, compared with exclusively formula-fed infants, appear to have a long-term respiratory benefit with fewer positive cultures for *P. aeruginosa*. However, infants who were exclusively breast fed for the first 6 months of life had less growth compared with formula-fed infants. Interestingly, mothers of CF infants are less likely to breastfeed than mothers of non-CF infants [77]. In addition to possible benefits for respiratory health, breast feeding may improve the quality of the mother-child

relationship and should be encouraged by newborn screening programs, even though growth may be slightly less. In addition, all infants with CF should receive supplemental vitamins, since vitamin deficiency is one morbidity which can be potentially prevented with newborn screening [78].

No recent controlled studies of any respiratory treatment have been published and the only controlled trial of antibiotic therapy in infants identified by newborn screening suggested that preventive antibiotics had only limited value [79]. At this point there is a profound need for clinical studies of therapies in infants with CF. Recently Accurso *et al.* [80] reported effective, targeted therapy for CF patients with cystic fibrosis transmembrane conductance regulator gating mutations. Targeted therapies for other mutation classes are currently underway, and studies of extending therapy to infants with gating mutations are being started. Therapy targeted at the basic defect holds great promise, especially when initiated in the newborn before the onset of significant lung disease [81].

CONCLUSION

Newborn screening for CF has come a long way since the first CDC workshop in 1997 recommended further study [82]. Based on modern criteria, CF clearly should be included in newborn screening programs [83]. Screening for CF has created some new challenges, particularly related to DNA testing, carrier detection, and inconclusive diagnoses. But excellent resources are available to assist program development [10^{***}]. Even with these new challenges, the future for the CF patient is improving as new therapies directed at the primary genetic and biochemical abnormalities become available [84].

Acknowledgements

E. Z. is funded by grants (ZEMANI08A0 and ZEMANI11A0) and M. S. by grant (SONTAG07A0) from the US Cystic Fibrosis Foundation.

Conflicts of interest

There are no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

Additional references related to this topic can also be found in the Current World Literature section in this issue (pp. 430–431).

1. Grosse SD, Boyle CA, Botkin JR, *et al.* Newborn screening for cystic fibrosis: evaluation of benefits and risks and recommendations for state newborn screening programs. *MMWR Recomm Rep* 2004; 53 (RR-13):1–36.
 2. de Monestrol I, Brucefors AB, Sjöberg B, Hjelte L. Parental support for newborn screening for cystic fibrosis. *Acta Paediatr* 2011; 100:209–215.
 3. Crossley JR, Smith PA, Edgar BW, *et al.* Neonatal screening for cystic fibrosis, using immunoreactive trypsin assay in dried blood spots. *Clin Chim Acta* 1981; 113:111–121.
 4. Hale JE, Parad RB, Dorkin HL, *et al.* Cystic fibrosis newborn screening: using experience to optimize the screening algorithm. *J Inherit Metab Dis* 2010; 33 (Suppl 2):S255–S261.
 5. Lindau-Shepard BA, Pass KA. Newborn screening for cystic fibrosis by use of a multiplex immunoassay. *Clin Chem* 2010; 56:445–450.
 6. Sommerburg O, Lindner M, Muckenthaler M, *et al.* Initial evaluation of a biochemical cystic fibrosis newborn screening by sequential analysis of immunoreactive trypsinogen and pancreatitis-associated protein (IRT/PAP) as a strategy that does not involve DNA testing in a Northern European population. *J Inherit Metab Dis* 2010; 33 (Suppl 2):S263–S271.
 7. Sarles J, Barthelmy S, Ferec C, *et al.* Blood concentrations of pancreatitis associated protein in neonates: relevance to neonatal screening for cystic fibrosis. *Arch Dis Child Fetal Neonatal Ed* 1999; 80:F118–F122.
 8. Oosterveld MJ, Schilperoot JV, Lilien MR, Arets HG. Positive neonatal screening for cystic fibrosis in neonates with renal failure. *Thorax* 2010; 65:652–653.
 9. Sontag MK, Wright D, Beebe J, *et al.* A new cystic fibrosis newborn screening algorithm: IRT/IRT1 upward arrow/DNA. *J Pediatr* 2009; 155:618–622.
 10. Newborn screening for cystic fibrosis; Approved guideline, Vol. 31. Wayne, PA: Clinical and Laboratory Standards Institute; 2011.
- This manual represents the most comprehensive information available on CF newborn screening, including program design, algorithms, and all supporting aspects from selection of mutation analysis to sweat testing and psychological support.
11. Castellani C, Massie J. Emerging issues in cystic fibrosis newborn screening. ■ *Curr Opin Pulm Med* 2010; 16:584–590.
- Excellent review of issues pertaining to choice of algorithm.
12. Goose MK, Reynolds R, Li Z, Farrell PM. Opportunities for quality improvement in cystic fibrosis newborn screening. *J Cyst Fibros* 2010; 9:284–287.
 13. Earley MC, Laxova A, Farrell PM, *et al.* Implementation of the first worldwide quality assurance program for cystic fibrosis multiple mutation detection in population-based screening. *Clin Chim Acta* 2011; 412:1376–1381.
 14. Farrell PM, Rosenstein BJ, White TB, *et al.* Guidelines for diagnosis of cystic fibrosis in newborns through older adults: Cystic Fibrosis Foundation consensus report. *J Pediatr* 2008; 153:S4–S14.
 15. Nelson MR, Adamski CR, Tluczek A. Clinical practices for intermediate sweat tests following abnormal cystic fibrosis newborn screens. *J Cyst Fibros* 2011; 10:460–465.
 16. Kleyn M, Korzeniewski S, Grigorescu V, *et al.* Predictors of insufficient sweat production during confirmatory testing for cystic fibrosis. *Pediatr Pulmonol* 2011; 46:23–30.
 17. Legrys VA, McColley SA, Li Z, Farrell PM. The need for quality improvement in ■ sweat testing infants after newborn screening for cystic fibrosis. *J Pediatr* 2010; 157:1035–1037.
- Important discussion of issues related to sweat testing in infants.
18. Sermet-Gaudelus I, Girodon E, Roussel D, *et al.* Measurement of nasal potential difference in young children with an equivocal sweat test following newborn screening for cystic fibrosis. *Thorax* 2010; 65:539–544.
 19. Sands D, Oltarzewski M, Nowakowska A, Zybert K. Bilateral sweat tests with two different methods as a part of cystic fibrosis newborn screening (CF NBS) protocol and additional quality control. *Folia Histochem Cytobiol* 2010; 48:358–365.
 20. Kirk JM. A continuing role for sweat testing in an era of newborn screening for cystic fibrosis. *Clin Biochem* 2011; 44:487–488.
 21. Sermet-Gaudelus I, Girodon E, Sands D, *et al.* Clinical phenotype and genotype of children with borderline sweat test and abnormal nasal epithelial chloride transport. *Am J Respir Crit Care Med* 2010; 182:929–936.
 22. Lilley M, Christian S, Hume S, *et al.* Newborn screening for cystic fibrosis in Alberta: two years of experience. *Paediatr Child Health* 2010; 15:590–594.
 23. Ivady G, Madar L, Nagy B, *et al.* Distribution of CFTR mutations in Eastern Hungarians: relevance to genetic testing and to the introduction of newborn screening for cystic fibrosis. *J Cyst Fibros* 2011; 10:217–220.
 24. Makukh H, Krenkova P, Tyrkus M, *et al.* A high frequency of the Cystic Fibrosis 2184insA mutation in Western Ukraine: genotype-phenotype correlations, relevance for newborn screening and genetic testing. *J Cyst Fibros* 2010; 9:371–375.
 25. Loukas YL, Soumelas GS, Dotsikas Y, *et al.* Expanded newborn screening in Greece: 30 months of experience. *J Inherit Metab Dis* 2010. [Epub ahead of print]
 26. Tluczek A, Orland KM, Cavanagh L. Psychosocial consequences of false-positive newborn screens for cystic fibrosis. ■ *Qual Health Res* 2011; 21:174–186.
- Excellent study of the impact of false-positive newborn screening.

27. Tluczek A, McKechnie AC, Brown RL. Factors associated with parental perception of child vulnerability 12 months after abnormal newborn screening results. *Res Nurs Health* 2011; 34:389–400.
- Important study concerning the vulnerable child syndrome.
28. Beucher J, Leray E, Deneuve E, *et al*. Psychological effects of false-positive results in cystic fibrosis newborn screening: a two-year follow-up. *J Pediatr* 2010; 156:771–776; 776.e1.
29. Dillard JP, Shen L, Robinson JD, Farrell PM. Parental information seeking following a positive newborn screening for cystic fibrosis. *J Health Commun* 2011; 15:880–894.
30. Stark AP, Lang CW, Ross LF. A pilot study to evaluate knowledge and attitudes of Illinois pediatricians toward newborn screening for sickle cell disease and cystic fibrosis. *Am J Perinatol* 2011; 28:169–176.
31. Collaco JM, Panny SR, Hamosh A, Mogayzel PJ Jr. False negative cystic fibrosis newborn screen. *Clin Pediatr (Phila)* 2010; 49:214–216.
32. Dunn CT, Skrypek MM, Powers AL, Laguna TA. The need for vigilance: the case of a false-negative newborn screen for cystic fibrosis. *Pediatrics* 2011; 128:e446–e449.
33. Tluczek A, Zaleski C, Stachiw-Hietpas D, *et al*. A tailored approach to family-centered genetic counseling for cystic fibrosis newborn screening: the Wisconsin model. *J Genet Couns* 2011; 20:115–128.
34. Culling B, Ogle R. Genetic counselling issues in cystic fibrosis. *Paediatr Respir Rev* 2010; 11:75–79.
35. Cavanagh L, Compton CJ, Tluczek A, *et al*. Long-term evaluation of genetic counseling following false-positive newborn screen for cystic fibrosis. *J Genet Couns* 2010; 19:199–210.
36. Lang CW, McColley SA, Lester LA, Ross LF. Parental understanding of newborn screening for cystic fibrosis after a negative sweat-test. *Pediatrics* 2011; 127:276–283.
37. La Pean A, Collins JL, Christopher SA, *et al*. A qualitative secondary evaluation of statewide follow-up interviews for abnormal newborn screening results for cystic fibrosis and sickle cell hemoglobinopathy. *Genet Med* 2011. [Epub ahead of print]
38. Sanders DB, Lai HJ, Rock MJ, Farrell PM. Comparing age of cystic fibrosis diagnosis and treatment initiation after newborn screening with two common strategies. *J Cyst Fibros* 2012; 11:150–153.
39. Farrell PM, Kosorok MR, Laxova A, *et al*. Nutritional benefits of neonatal screening for cystic fibrosis. Wisconsin Cystic Fibrosis Neonatal Screening Study Group. *N Engl J Med* 1997; 337:963–969.
40. Maclean JE, Solomon M, Corey M, Selvadurai H. Cystic fibrosis newborn screening does not delay the identification of cystic fibrosis in children with negative results. *J Cyst Fibros* 2011; 10:333–337.
41. Mehta A. The how (and why) of disease registers. *Early Hum Dev* 2010; 86:723–728.
42. Strom CM, Redman JB, Peng M. The dangers of including nonclassical cystic fibrosis variants in population-based screening panels: p.L997F, further genotype/phenotype correlation data. *Genet Med* 2011; 13:1042–1044.
43. Burgel PR, Fajac I, Hubert D, *et al*. Nonclassical cystic fibrosis associated with D1152H CFTR mutation. *Clin Genet* 2010; 77:355–364.
44. Baker MW, Groose M, Hoffman G, *et al*. Optimal DNA tier for the IRT/DNA algorithm determined by CFTR mutation results over 14 years of newborn screening. *J Cyst Fibros* 2011; 10:278–281.
45. Korzeniewski SJ, Young WI, Hawkins HC, *et al*. Variation in immunoreactive trypsinogen concentrations among Michigan newborns and implications for cystic fibrosis newborn screening. *Pediatr Pulmonol* 2011; 46:125–130.
46. Thauvin-Robinet C, Munck A, Huet F, *et al*. The very low penetrance of cystic fibrosis for the R117H mutation: a reappraisal for genetic counselling and newborn screening. *J Med Genet* 2009; 46:752–758.
47. Massie RJ, Poplawski N, Wilcken B, *et al*. Intron-8 polythymidine sequence in Australasian individuals with CF mutations R117H and R117C. *Eur Respir J* 2001; 17:1195–1200.
48. Ren CL, Desai H, Platt M, Dixon M. Clinical outcomes in infants with cystic fibrosis transmembrane conductance regulator (CFTR) related metabolic syndrome. *Pediatr Pulmonol* 2011; 46:1079–1084.
- First study of longitudinal evaluation in patients with CRMS. Results indicate a variable outcome.
49. Borowitz D, Parad RB, Sharp JK, *et al*. Cystic Fibrosis Foundation practice guidelines for the management of infants with cystic fibrosis transmembrane conductance regulator-related metabolic syndrome during the first two years of life and beyond. *J Pediatr* 2009; 155 (6 Suppl):S106–S116.
50. Salvatore D, Buzzetti R, Baldo E, *et al*. An overview of international literature from cystic fibrosis registries 2. Neonatal screening and nutrition/growth. *J Cyst Fibros* 2010; 9:75–83.
51. Tluczek A, Becker T, Laxova A, *et al*. Relationships among health-related quality of life, pulmonary health, and newborn screening for cystic fibrosis. *Chest* 2011; 140:170–177.
52. Rosenfeld M, Emerson J, McNamara S, *et al*. Baseline characteristics and factors associated with nutritional and pulmonary status at enrollment in the cystic fibrosis EPIC observational cohort. *Pediatr Pulmonol* 2010; 45:934–944.
53. Accurso FJ, Sontag MK, Wagener JS. Complications associated with symptomatic diagnosis in infants with cystic fibrosis. *J Pediatr* 2005; 147 (3 Suppl):S37–S41.
54. Dijk FN, McKay K, Barzi F, *et al*. Improved survival in cystic fibrosis patients diagnosed by newborn screening compared to a historical cohort from the same centre. *Arch Dis Child* 2011; 96:1118–1123.
- Comparison of survival in children identified by newborn screening compared with historical controls.
55. Walsh AC, Rault G, Li Z, *et al*. Pulmonary outcome differences in U.S. and French cystic fibrosis cohorts diagnosed through newborn screening. *J Cyst Fibros* 2010; 9:44–50.
- Interesting comparison of different outcomes in similar newborn screening populations.
56. Briesacher BA, Quittner AL, Fouayzi H, *et al*. Nationwide trends in the medical care costs of privately insured patients with cystic fibrosis (CF), 2001–2007. *Pediatr Pulmonol* 2011; 46:770–776.
57. Borowitz D, Robinson KA, Rosenfeld M, *et al*. Cystic Fibrosis Foundation evidence-based guidelines for management of infants with cystic fibrosis. *J Pediatr* 2009; 155 (6 Suppl):S73–S93.
58. Sermet-Gaudelus I, Mayell SJ, Southern KW. Guidelines on the early management of infants diagnosed with cystic fibrosis following newborn screening. *J Cyst Fibros* 2010; 9:323–329.
- European-based guidelines for managing infants with CF. Most, but not all, suggestions are similar to US-based guidelines.
59. Massie J, Curnow L, Gaffney L, *et al*. Declining prevalence of cystic fibrosis since the introduction of newborn screening. *Arch Dis Child* 2010; 95:531–533.
- Intellectually stimulating discussion on whether newborn screening has altered the prevalence of CF.
60. McClaren BJ, Metcalfe SA, Aitken M, *et al*. Uptake of carrier testing in families after cystic fibrosis diagnosis through newborn screening. *Eur J Hum Genet* 2010; 18:1084–1089.
61. Sanders DB, Li Z, Brody AS, Farrell PM. Chest computed tomography scores of severity are associated with future lung disease progression in children with cystic fibrosis. *Am J Respir Crit Care Med* 2011; 184:816–821.
62. Pillarisetti N, Linnane B, Ranganathan S. Early bronchiectasis in cystic fibrosis detected by surveillance CT. *Respirology* 2010; 15:1009–1011.
63. Lum S, Gustafsson P, Ljungberg H, *et al*. Early detection of cystic fibrosis lung disease: multiple-breath washout versus raised volume tests. *Thorax* 2007; 62:341–347.
64. Hall GL, Logie KM, Parsons F, *et al*. Air trapping on chest CT is associated with worse ventilation distribution in infants with cystic fibrosis diagnosed following newborn screening. *PLoS One* 2011; 6:e23932.
65. Khan TZ, Wagener JS, Bost T, *et al*. Early pulmonary inflammation in infants with cystic fibrosis. *Am J Respir Crit Care Med* 1995; 151:1075–1082.
66. Pillarisetti N, Williamson E, Linnane B, *et al*. Infection, inflammation, and lung function decline in infants with cystic fibrosis. *Am J Respir Crit Care Med* 2011; 184:75–81.
- Prospective study of early lung abnormalities in infants with CF detected by newborn screening. Findings of infection, inflammation, and abnormal lung function are compared at various ages in children studied longitudinally.
67. Ranganathan SC, Parsons F, Gangell C, *et al*. Evolution of pulmonary inflammation and nutritional status in infants and young children with cystic fibrosis. *Thorax* 2011; 66:408–413.
- Valuable addition to the findings of early nutritional deficiency and airway inflammation.
68. Staffer P, Davies JC, Balfour-Lynn IM, *et al*. Bronchoscopy in cystic fibrosis infants diagnosed by newborn screening. *Pediatr Pulmonol* 2011; 46:696–700.
69. Wainwright CE, Vidmar S, Armstrong DS, *et al*. Effect of bronchoalveolar lavage-directed therapy on *Pseudomonas aeruginosa* infection and structural lung injury in children with cystic fibrosis: a randomized trial. *JAMA* 2011; 306:163–171.
- Pivotal study showing that clinical outcomes are not altered by surveillance bronchoscopy and lavage.
70. Abman SH, Ogle JW, Harbeck RJ, *et al*. Early bacteriologic, immunologic, and clinical courses of young infants with cystic fibrosis identified by neonatal screening. *J Pediatr* 1991; 119:211–217.
71. Hayes D Jr, West SE, Rock MJ, *et al*. *Pseudomonas aeruginosa* in children with cystic fibrosis diagnosed through newborn screening: assessment of clinic exposures and microbial genotypes. *Pediatr Pulmonol* 2010; 45:708–716.
- These results suggest that separate clinics for *Pseudomonas* positive and negative patients are not necessary when good infection control policies are in effect.
72. West SE, Zeng L, Lee BL, *et al*. Respiratory infections with *Pseudomonas aeruginosa* in children with cystic fibrosis: early detection by serology and assessment of risk factors. *JAMA* 2002; 287:2958–2967.
73. Hayes D Jr, Farrell PM, Li Z, West SE. *Pseudomonas aeruginosa* serological analysis in young children with cystic fibrosis diagnosed through newborn screening. *Pediatr Pulmonol* 2010; 45:55–61.
74. Rogan MP, Reznikov LR, Pezzulo AA, *et al*. Pigs and humans with cystic fibrosis have reduced insulin-like growth factor 1 (IGF1) levels at birth. *Proc Natl Acad Sci USA* 2010; 107:20571–20575.
- Important finding of abnormal growth factors in the CF pig model and correlating values in infants detected by newborn screening.

75. Robinson KA, Saldanha IJ, McKoy NA. Management of infants with cystic fibrosis: a summary of the evidence for the cystic fibrosis foundation working group on care of infants with cystic fibrosis. *J Pediatr* 2009; 155 (6 Suppl): S94–S105.
76. Jadin SA, Wu GS, Zhang Z, *et al.* Growth and pulmonary outcomes during the first 2 y of life of breastfed and formula-fed infants diagnosed with cystic fibrosis through the Wisconsin Routine Newborn Screening Program. *Am J Clin Nutr* 2011; 93:1038–1047.
- Important study comparing outcomes for CF infants fed breast milk or infant formula.
77. Tluczek A, Clark R, McKechnie AC, *et al.* Task-oriented and bottle feeding adversely affect the quality of mother-infant interactions after abnormal newborn screens. *J Dev Behav Pediatr* 2010; 31:414–426.
78. Obeid M, Price J, Sun L, *et al.* Facial palsy and idiopathic intracranial hypertension in twins with cystic fibrosis and hypovitaminosis A. *Pediatr Neurol* 2011; 44:150–152.
79. Weaver LT, Green MR, Nicholson K, *et al.* Prognosis in cystic fibrosis treated with continuous flucloxacillin from the neonatal period. *Arch Dis Child* 1994; 70:84–89.
80. Accurso FJ, Rowe SM, Clancy JP, *et al.* Effect of VX-770 in persons with cystic fibrosis and the G551D-CFTR mutation. *N Engl J Med* 2010; 363:1991–2003.
81. Welsh MJ. Targeting the basic defect in cystic fibrosis. *N Engl J Med* 2010; 363:2056–2057.
82. Newborn screening for cystic fibrosis: a paradigm for public health genetics policy development. *Morb Mortal Wkly Rep* 1997; 46: 1–24.
83. Petros M. Revisiting the Wilson-Jungner criteria: how can supplemental criteria guide public health in the era of genetic screening? *Genet Med* 2012; 14:129–134.
84. Bush A, Davies J. Cystic fibrosis: to ion transport and beyond. *Eur Respir J* 2011; 36:991–992.