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Isolation and identification of Enterobacter spp from pet animals Ear Infections in Al-Hilla Veterinary clinic, Iraq

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ABSTRACT	ARTICLE DETAI
Objective: To isolation and identification of Enterobactor spp. from ear infection in pet animals in	Published On:
Babylon province by traditional method Automated Vitek 2 system as well as determined the antibiotic sensitivity test.	18 August 2023
Methods; One hundred swab of ear infection were isolated from dogs and cats ear infections in	
different veterinary clinic in Babylon province, at a period between 1 january to 1 July 2022.	
Results: The results of pet animals swab collection revealed that the percentage of gram negative was	
71(71%) while the positive was 29 (29%) as well as the biochemical test showed that the enterococcal	
cloaca percent was (64%), Klebsiella (19%), positive bacteria (6%), E. coli 3% and no growth was	
7%. The biochemical tests showed positive for Catalase, Citrate. They gave negative reaction for	
Urease, Oxidase, MR (Methyl Red), Indole. By the other hand, the result of isolated E. cloaca that	
was plated onto culture media. then subcultures on MacConkey agar (MAC) and XLD agar. The	
Antimicrobial susceptibility test of E.cloaca against 55 conventional antimicrobial revealed that	
E.cloaca varied to a large extent. The isolated bacteria showed sensitivity for the most commonly	
used antimicrobials for treatment of bacterial otitis media in dog and cat which were amoxycillin,	
amoxycillin+clavulanic acid, cephalospins, gentamicin and fluoroquinolones (ciprofloxacin,	
ofloxacin, levofloxacin) azithromycin, clarithromycin, neomycin and ploymyxin-B. In contrast, the	
present study showed that E.cloaca resistant for three antibiotic belongs from beta lactam group	

(Cefsulodin, Loracarbet and Cefpodoxime). **Conclusion:** the production of inhibitor-resistant β -lactamases by E. cloacae isolates could be increasingly common in nosocomial pathogens other than E. coli or K. pneumoniae in pet animals ear infection in Al-Hilla city Babylon province, Iraq.

KEYWORDS: Enterobacter spp., ear infection, Vitek2, Cat and Dog.

INTRODUCTION

In the underdeveloped world, ear infections are the leading cause of hearing loss, although being entirely preventable (1,2). Ear infections affect about 10-20 % of the dogs and cats, as well as it is one of the most common health problems of the pet animals (3). Ear infections are usually caused by yeast, ear mites and bacteria, with bacteria being the most common cause. Ear problem in dog and cat are usually identified by observing head shaking/ tilting, smelly ears, ear scratching, lack of balance, unusual back-and-forth eye movements, redness inside the ear, swelling of the ear and/or brown, yellow, or bloody discharge from the ear. The problem may be in one or both the ears at any age. Ear infections are usually caused by Staphylococcus,

Pseudomonas and Streptococcus species (4). The most common bacterial causes of otitis media in animals and human beings include Escherichia coli, Proteus spp., Staphylococcus Streptococcus aureus, pneumoniae, Moraxella (Branhamella) catarrhalis, and Haemophilus influenzae (5). Besides Staphylococcus intermedius, Staphylococcus hyicus, Corynebacterium spp., and Enterobacter spp,. isolates, Proteus mirabilis was identified as the most frequent cause of otitis in dogs (6). Enterobacter cloacae are pathogens that are commonly isolated from human clinical specimens. They cause an extensive range of nosocomial infections involving skin and soft tissue, the lower respiratory tract, urinary tract, and the CNS infections. The antibiotic-resistance properties of these bacteria had been

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described by several authors worldwide (7). We also look into the prevalence of methicillin-resistant S. aureus (MRSA) among S. aureus isolates and the presence of the mecA gene in MRSA samples from Babylon, Iraq.

MATERIALS AND METHODS

Study design: One hundred ear infection swabs were recruited from the veterinary clinics in Babylon province (Hilla city), Iraq, for this study. Al-Qasim green University in Iraq was home to the study's Faculty of Microbiology.

Sample collection: One or both ears may have been swabbed aseptically to gather samples. Medical professionals collected samples, which were then transferred to the microbiological lab in an ice box within an hour after being immersed in nutrient broth (Oxoid, UK) as a transport media.

Isolation and identification: Each clinical sample was inoculated onto plates of XLD and MacConkey Agar (Oxoid, UK) and then incubated at 35-37°C for 20 hours. E. cloaca was identified as colonies of color.

Antibiotic susceptibility of Staphylococcus aureus isolates by VITEK2 system

Antimicrobial sensitivity testing of staph. Aureus was performed by Vitek-2 automated susceptibility system according to manufacturer's instructions. Antibiotic susceptibility of staph. Aureus against 66 antibiotics belonging to 9 classes of antimicrobial agents was tested, as shown in Table (2).

Statistical Analysis: SPSS (Armonk, New York: IBM Corp.) version 17 (Statistical Package for the Social Sciences) was used for the calculations **(8)**.

RESULTS

Sample collection and Geographical isolation: The results of pet animals swab collection illustrated in table (1) and figure (1) revealed that the percentage of gram negative was 71(71%) while the positive was 29 (29%) as well as the biochemical test showed that the enterococcal cloaca percent was (64%), Klebsiella (19%), positive bacteria (6%), E. coli 3% and no growth was 7%.

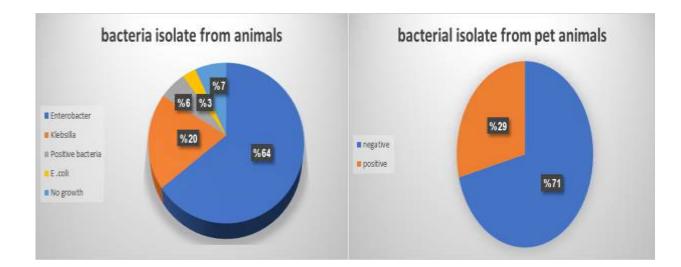


Table (1): Geographically distribution of different swab collection from ear infection in pet animals at Babylon province.

No	Locat	tion Cat	dog	Total
1	Babylon veterinary hospital	3	1	4
2	Janet ard babil clinic	9	2	11
3	Flamango clinic	10	5	15
4	Al awaeal clinic	11	6	17
5	Safary clinic	10	3	13
6	Muayad clinic	6	3	9
7	Dr. Enas clinic	9	4	13
8	Dr Ali alsharaa clinic	8	4	12
9	Nelover clinic	3	3	6
	Total	69	31	100

Biochemical Identification of Enterobacter spp.

The biochemical tests for pet animal samples all isolates showed positive for Catalase, Citrate. They gave negative reaction for Urease, Oxidase, MR (Methyl Red), Indole. In the present study, these conventional biochemical tests that carried out, and the results were compared with the results documented by Probert et al., (9). The result of isolated E. cloaca that was plated onto culture media. Fig (2, A) then subcultures on MacConkey agar (MAC) and XLD agar in Fig (2, B). The current finding strongly consistent with (9, 10 11).

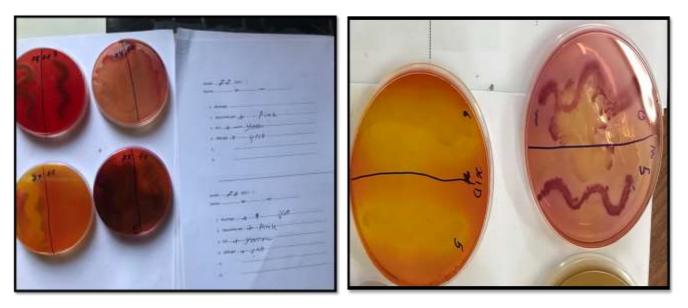


Figure (2): A- Enterobacter on culture media; B- Enterobacter on XLD and MacConkey.

Identification and Antibiotic susceptibility of Enterobacter cloaca isolates by VITEK2 system:

The VITEK 2 system has proven the identification of the Enterobactor.cloaca that have been taken from cat and dog ear swabs. The main thing that sets VITEK 2 devices apart is that they can diagnose bacterial isolates faster and more accurately. Using (64) molecular tests to make a diagnosis with special kits and away from contamination, which could make it hard to find a pathogen.

Table (2): Antibiotic sensitivit	v of isolated E.cloaca	by using VITEK2 system
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Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
Ticarcillin	<=8	S	Gentamicin		S
Ticarcillin /clavulanic	<=8	S	Isepumicin		S
acid					
Piperacillin	<=4	S	Netilmicin		S
+ Piperacillin/sulbactam			Tobramycin		S
Piperacillin/tazobactam	<=4	S	Cinoxacin		
+cefsulodin		R	Nalidixic Acid		
+ceftobiprole		S	Ciprofloxacin		S
+loracarbet		R	Delafloxacin		
+cefpodoxime		R	Enrolloxacin		
+ceftibutin			Flerosacin		
+cefotaxime		S	Gutilloxacin		S
+cefovecin			Lev ofloxacin		S
+ceftazidime	<=1	S	Lomefloxacin		
+ceftiofur			Marbofloxacin		
Ceftizoxime		S	moxifloxacin		
Cettriaxone		S	Norfloxacin		
Cefoperazone		S	ofloxacin		S
/Sulbactam					
Ceftazidime:Avibactam		S	pefloxacin		
Cefepime	<=1	S	Pradofloxacin		
+Cefpirome		S	Tosufloxacin		
Aztreonam	<=1	S	Doxyeycline		

Doripenem+		S	Minocycline	2	S
+Faropenem	+Faropenem		Tetracyeline		S
Imipenem	<=0,25	S	Colistin		
.Meropenem	<=0,25	S	Rifampicin		
Panipenem+			Trimethoprim		
+Meropenem/Vaborbact		S	Trimethoprim/	<=20	S
am			Sulfamethosazole		
Amikacin	<=2	S			

DISCUSSION

In this study, 64 bacterial sample were identified as Enterobacter spp. Using commercialized systems, such as biochemical tests and Vitek2 system. These techniques present limitations in species' identification, particularly within the E. cloacae complex and between E. hormaechei, E. cloacae, E. asburiae, E. kobei, and E. ludwigii, owing to the high similarity between them (12). For the right drug treatment, it is important to correctly identify the species, and wrong identifications could hurt the patient's health (13). Antimicrobial susceptibility test of E.cloaca in table (2) against 55 conventional antimicrobial revealed that E.cloaca varied to a large extent. The isolated bacteria showed sensitivity for the most commonly used antimicrobials for treatment of bacterial otitis media in dog and cat which were amoxycillin, amoxycillin+clavulanic acid, cephalospins (cefalexin, cefdinir, cefixime, ceftriaxone, cefuroxime), gentamicin and fluoroquinolones (ciprofloxacin, ofloxacin, levofloxacin) azithromycin, clarithromycin, neomycin and ploymyxin-B (13). These findings agreed with Singh et al., (3). In contrast, the present study showed that E.cloaca resistant for three antibiotic belongs from beta lactam group (Cefsulodin, Loracarbet and Cefpodoxime) .According to a recent meta-analysis report, an estimated 48.9% of Gramnegative (Enterobacteriaceae) clinical isolates in Ethiopia are ESBL producers, most of them are classified as MDR because they usually carry genes encoding resistance to antibiotics other than beta-lactams (14). Results of this study are in agreement with a study of Alzaidi and Mohammed in Iraq (15), who founded that all E. cloacae isolates recovered to be susceptible to cefoxitin disks as per CLSI guidelines (CLSI, 2019), confirming their ability to produce AmpC β lactamases. For our knowledge, the present study is the first study that isolated Enterobacor. cloaca as the most prominent bacteria caused otitis in dog and cat in Iraq. It is the same as that reported in Italy by Piccolo et al., (2020) when they isolated E.cloaca in cat when they reported that 34.5% from all isolated bacteria was Enterobacter spp.

CONCLUSION

From this study, we can conclude that the production of inhibitor-resistant β -lactamases by E. cloacae isolates could be increasingly common in nosocomial pathogens other than

E. coli or K. pneumoniae in pet animals ear infection in Al-Hilla city Babylon province, Iraq.

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Conflict of interest: None.

REFERENCES

- I. Ullauri, A., Smith, A., Espinel, M., Jimenez, C., Salazar, C., & Castrillon, R. (2014). WHO ear and hearing disorders survey: Ecuador national study 2008-2009. In Conference Papers in Science (Vol. 2014, pp. 1-13). Hindawi Limited.
- II. Sonbol, F. I., Abdelaziz, A. A., El-banna, T. E. S., & Farag, O. (2022). Detection and characterization of staphylococcus aureus and methicillin-resistant S. aureus (MRSA) in ear infections in Tanta, Egypt. Journal of Advanced Medical and Pharmaceutical Research, 3(2), 36-44
- III. Singh, B. R., Pawde, A. M., Singh, S. V., Agri, H., Sinha, D. K., & Vinodhkumar, O. R. (2019). Ear Infections in Animals in Bareilly: Common causes and Effective Antimicrobials. Austin J Vet Sci & Anim Husb, 6(3), 1061.
- IV. Campbell, J. J., Coyner, K. S., Rankin, S. C., Lewis, T. P., Schick, A. E., & Shumaker, A. K. (2010). Evaluation of fungal flora in normal and diseased canine ears. Veterinary dermatology, 21(6), 619-625.
- V. Kraijer-Huver, I. M. G., Ter Haar, G., Djajadiningrat-Laanen, S. C., & Boevé, M. H. (2009). Peri-and retrobulbar abscess caused by chronic otitis externa, media and interna in a dog. The Veterinary Record, 165(7), 209.
- VI. Neves, R. C., Makino, H., Cruz, T. P., Silveira, M. M., Sousa, V. R., Dutra, V., ... & Belli, C. B. (2018). In vitro and in vivo efficacy of tea tree essential oil for bacterial and yeast ear infections in dogs. Pesquisa Veterinária Brasileira, 38, 1597-1607.
- VII. Otto, C. M., Franz, M. A., Kellogg, B., Lewis, R., Murphy, L., & Lauber, G. (2002). Field treatment of search dogs: lessons learned from the World Trade

Center disaster. Journal of Veterinary Emergency and Critical Care, 12(1), 33-41.

- VIII. Al-Ameedi, A. I., Ayad, Z. M., Mohammed, W. A., & Hajwal, S. K. (2023). Ginkgo Biloba extract's efficacy to mitigate the genotoxicity that hydroxyurea induces in mice. Adv. Anim. Vet. Sci, 11(4), 552-557.
 - IX. Probert, W. S., McQuaid, C., & Schrader, K. (2014). Isolation and identification of an Enterobacter cloacae strain producing a novel subtype of Shiga toxin type 1. Journal of clinical microbiology, 52(7), 2346-2351.
 - X. Dierikx, C. M., van Duijkeren, E., Schoormans, A. H. W., van Essen-Zandbergen, A., Veldman, K., Kant, A., ... & Mevius, D. J. (2012). Occurrence and characteristics of extended-spectrum-β-lactamase-and AmpC-producing clinical isolates derived from companion animals and horses. Journal of Antimicrobial Chemotherapy, 67(6), 1368-1374.
 - XI. Piccolo, F. L., Belas, A., Foti, M., Fisichella, V., Marques, C., & Pomba, C. (2020). Detection of multidrug resistance and extendedspectrum/plasmid-mediated AmpC beta-lactamase genes in Enterobacteriaceae isolates from diseased cats in Italy. Journal of feline medicine and surgery, 22(7), 613-622.
- XII. Davin-Regli, A., Lavigne, J. P., & Pagès, J. M. (2019). Enterobacter spp.: update on taxonomy, clinical aspects, and emerging antimicrobial resistance. Clinical microbiology reviews, 32(4), 10-1128.
- XIII. Intra, J., Carcione, D., Sala, R. M., Siracusa, C., Brambilla, P., & Leoni, V. (2023). Antimicrobial Resistance Patterns of Enterobacter cloacae and Klebsiella aerogenes Strains Isolated from Clinical Specimens: A Twenty-Year Surveillance Study. Antibiotics, 12(4), 775.
- XIV. Getaneh, A., Ayalew, G., Belete, D., Jemal, M., & Biset, S. (2021). Bacterial etiologies of ear infection and their antimicrobial susceptibility pattern at the university of Gondar comprehensive specialized hospital, Gondar, Northwest Ethiopia: a six-year retrospective study. Infection and drug resistance, 4313-4322.
- XV. Alzaidi, J. R., & Mohammed, A. S. (2022). First Record of Dissemination of BLBLI-Resistant Enterobacter cloacae from Public Hospitals in Baghdad, Iraq, The open microbiology journal, vol 16(3):1-8.