

# Unnatural evolutionary processes of SARS-CoV-2 variants and possibility of deliberate natural selection

1 **Atsushi Tanaka**<sup>1,2,\*</sup> †, **Takayuki Miyazawa**<sup>2,3,\*</sup> †

2 <sup>1</sup>Division of Research Animal Laboratory and Translational Medicine, Research and Development  
3 Center Osaka Medical and Pharmaceutical University, Takatsuki, Osaka 569-8686, Japan

4 <sup>2</sup>Laboratory of Virus-Host Coevolution, Institute for Life and Medical Sciences, Kyoto University,  
5 Sakyo-ku, Kyoto 606-8507, Japan

6 <sup>3</sup>Resilience Research Unit, Kyoto University, Katsura, Nishikyo-ku, Kyoto 615-8540, Japan

7

8 **\* Correspondence:**

9 **Atsushi Tanaka\***

10 atsushi.tanaka@ompu.ac.jp

11

12 **Takayuki Miyazawa\***

13 takavet@infront.kyoto-u.ac.jp

14

15 **Keywords:** SARS-CoV-2, evolution, Omicron BA.1, Omicron BA.2, SARS-CoV-2 Omicron, Puerto Rico.

## 16 **Abstract**

17 Over the past three years, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has repeatedly  
18 caused pandemics, generating various mutated variants ranging from Alpha to Omicron. In this study, we  
19 aimed to clarify the evolutionary processes leading to the formation of SARS-CoV-2 Omicron variants,  
20 focusing on Omicron variants with many amino acid mutations in the spike protein among SARS-CoV-2  
21 isolates. To determine the order of mutations leading to the formation of the SARS-CoV-2 Omicron variants,  
22 we compared the sequences of 129 Omicron BA.1-related, 141 BA.1.1-related, and 122 BA.2-related isolates,  
23 and attempted to clarify the evolutionary processes of SARS-CoV-2 Omicron variants, including the order of  
24 mutations leading to their formation and the occurrence of homologous recombination. As a result, we  
25 concluded that the formation of a part of Omicron isolates BA.1, BA.1.1, and BA.2 was not the product of  
26 genome evolution, as is commonly observed in nature, such as the accumulation of mutations and homologous  
27 recombinations. Furthermore, the study of 35 recombinant isolates of Omicron variants BA.1 and BA.2  
28 confirmed that Omicron variants were already present in 2020. The analysis showed that Omicron variants  
29 were formed by an entirely new mechanism that cannot be explained by previous biology, and knowing how  
30 the SARS-CoV-2 variants were formed prompts a reconsideration of the SARS-CoV-2 pandemic.

## 31 **1 Introduction**

32

33 COVID-19, the coronavirus disease 2019, caused by severe acute respiratory syndrome coronavirus 2 (SARS-  
34 CoV-2), was first reported in December 2019 in Wuhan, China (1). This emerging infectious disease was  
35 unprecedentedly fast, spreading worldwide, leading the World Health Organization (WHO) to declare a global  
36 pandemic of COVID-19 on March 11, 2020 (<https://www.who.int/>). SARS-CoV-2, belonging to  
37 betacoronavirus subgroup B, has a single-stranded positive-sense RNA genome that codes for ten genes,  
38 ultimately producing 26 proteins according to an annotation of NCBI Reference Sequence: NC\_045512.2. Its  
39 genome size varies from 29.8 to 29.9 kb. It consists of four structural proteins: spike (S), envelope (E),  
40 membrane (M), and nucleocapsid (N) proteins (2, 3). In the structural proteins, the S protein as the surface

41 glycoprotein is the largest protein, being approximately 180 kDa, and consisting of two subunits, S1 and S2. It  
42 mediates membrane fusion and ultimately facilitates virus entry. The receptor-binding domain (RBD) (amino  
43 acid residues 319-541) of the S1 subunit interacts with angiotensin - converting enzyme 2 (ACE2), binding to  
44 its peptidase domain (4, 5).

45  
46 Over the three years from 2019 to 2022, SARS-CoV-2 was re-accelerated by new variants that emerged over  
47 several months in various geographical regions and disseminated throughout the world, to induce the pandemic  
48 repeatedly.

49  
50 In the early stage of the first pandemic, the most impactful mutation of SARS-CoV-2 was the non-synonymous  
51 mutation D614G in the S protein. This mutation, which was not present in the ancestral lineage that caused the  
52 Wuhan outbreak, quickly became dominant worldwide (6). Soon after, the variant of concern, B.1.1.7 : 20I  
53 (Alpha, V1), the lineage B.1.1.7 (clade 501.YV1), or Alpha, characterized by 17 unique mutations containing  
54 ten amino acid differences in the S protein, was first detected in southeastern England in late September 2020  
55 (7) and expanded rapidly across the United Kingdom to become predominant during early 2021, spreading to  
56 most European countries with similar success. By November 2021, local transmission of this lineage had been  
57 reported in 175 countries (8). Shortly after, the emergence of variant strains of SARS-CoV-2 Alpha, variants  
58 B.1.351 : 20H (Beta, V2), was identified in October 2020, which was first detected in the Eastern Cape province  
59 of South Africa from specimens collected in early August. This Beta variant spread within South Africa and was  
60 considered to have displaced the other SARS-CoV-2 lineages circulating there (9). Then, the variant P.1: 20J  
61 (Gamma, V3) was identified in Brazil in December 2020, thought to have evolved in Brazil. Health officials in  
62 Japan first reported it publicly on January 10, 2021, after identifying the Gamma variant in four Brazilian  
63 travelers at Haneda Airport in Tokyo, Japan (10).

64  
65 At about the same time, the Delta variant (Pango lineage designation B.1.617.2), which was first detected in  
66 India in February 2021, and the Mu variant, also known as lineage B.1.621 first detected in Colombia in January  
67 2021, were reported (11, 12).

68  
69 Almost one year later, regarding these emergences of variants of concern, Omicron (phylogenetic assignment  
70 of named global outbreak (Pango) lineage designation B.1.1.529; BA.1, Nextstrain clade 21K) was a variant of  
71 SARS-CoV-2 first reported to WHO by the Network for Genomics Surveillance in South Africa on November  
72 24, 2021 (13, 14) with more than 50 amino acids changes when compared with the first reported strain Wuhan-  
73 Hu-H1 (NCBI Reference Sequence: NC\_045512.2.), and 39 of these amino acids difference were observed in  
74 the S protein. This variant was first detected in Botswana and became the predominant circulating variant  
75 worldwide (15).

76 In the United States, the San Francisco Department of Public Health confirmed that a case of COVID-19 among  
77 individuals in California was caused by Omicron variant BA.1, carried by a traveler who returned from South  
78 Africa on November 22, 2021 (<https://www.cdc.gov/media/releases/2021/s1201-Omicron-variant.html>). Then,  
79 the first Omicron sub-lineage BA.1 expanded rapidly and replaced the Delta variant (16).

80 Less than two weeks later, the Omicron variant BA.1, the new Omicron variant, BA.2 lineage, showing 31  
81 amino acids changes in the S protein when compared with the Wuhan-Hu-H1, was initially identified in  
82 Denmark on December 5, 2021 (17). On February 22, 2022, WHO mentioned the Omicron sublineage BA.2  
83 (<https://www.who.int/news/item/22-02-2022-statement-on-Omicron-sublineage-ba.2>), whereby the Omicron  
84 variant of concern was currently the dominant variant circulating globally, replacing the Delta variant (Pango  
85 lineage designation B.1.617.2) ([https://www.who.int/docs/default-source/coronaviruse/2022-01-07-global-technical-brief-and-priority-action-on-Omicron---corr2.pdf?sfvrsn=918b09d\\_20](https://www.who.int/docs/default-source/coronaviruse/2022-01-07-global-technical-brief-and-priority-action-on-Omicron---corr2.pdf?sfvrsn=918b09d_20)), accounting for nearly all  
86 sequences reported to GISAID. Then, as of March 16, 2023, WHO stated that the Omicron variants accounted  
87 for over 98% of the publicly available viral sequences after February 2022 (<https://www.who.int/news/item/16-03-2023-statement-on-the-update-of-who-s-working-definitions-and-tracking-system-for-SARS-CoV-2-variants-of-concern-and-variants-of-interest>).

91 Omicron variants BA.1 and BA.2 were suggested to have expanded and diverged around October to December  
92 2021, respectively. These mutants were estimated to have diverged from a common ancestor around February  
93 to March 2021 (18). Since Omicron variants BA.1 and BA.2 share a common 14-amino acid mutation in the S  
94 protein, the common ancestor of Omicron variants BA.1 and BA.2 may have already acquired the 14-amino  
95 acid mutation in the S protein region around February to March 2021; however, no common ancestral strain has  
96 been found in the international databases, and the strains may have acquired their mutations through different  
97 pathways.

98 In this study, we attempted to clarify the evolutionary processes of the Omicron variant, which has two-times  
99 more amino acid mutations in the S protein than other variants, by examining the rank order of introduced amino  
100 acid mutations in the S protein.

## 101 2 Results

102 Each variant is considered to have arisen through an independent evolutionary pathway from isolates with the  
103 D614G mutation in the S protein. Concerning the genetic variation in the S protein of these variants, most of the  
104 mutations were non-synonymous (Fig. 1). There were no synonymous mutations in the Alpha, Beta, Gamma,  
105 Delta, or Mu variants, but only one each in the Lambda and Omicron variants. Among these variants, the  
106 Omicron variant (BA.1 lineage), which shows the greatest accumulation of mutations in the S protein, is  
107 primarily non-synonymous in the S protein and has only one synonymous mutation, at c25000u. The  
108 synonymous/non-synonymous ratio is abnormal, considering how human coronaviruses have mutated (See  
109 Supplemental Figure 1).

110 At first, we considered the existence of the isolate of SARS-CoV-2, whose amino acid sequence in the S protein  
111 contains the Omicron-BA.1-type mutation subsets, but one mutation position was not mutated and comprised  
112 the original Wuhan-type amino acid sequence. We designated these isolates as BA.1-01. Each amino acid  
113 sequence of the S protein region was named BA.1-0.1\_S: amino acids of the Omicron-BA.1 type (Oaa) and  
114 Wuhan type (Waa) and its position number (XXX) (Ex., BA.1-0.1\_S:OaaXXXWaa), as described in Methods.  
115 Then, the putative isolates bearing BA.1-0.1\_S:OaaXXXWaa were searched for using the BLAST program  
116 based on their amino acid sequences. In this search, we obtained the isolates whose identities showed 100%  
117 matches with the query amino acid sequence except for SARS-CoV-2\_human\_USA\_NY-  
118 PV55373\_2022(GenBank: ON246090.1), whose identity was 99.92%.

119 Surprisingly, we found that Omicron BA.1-0.1 isolates were detected at all mutation sites except N501Y (Fig.  
120 2A). In the BA.1 lineage of the Omicron variant, there are Omicron isolates (BA.1.1) with the R346K mutation  
121 seen in the Mu(m) variant (termed B.1.621), *i.e.*, BA.1\_S can be defined as BA.1.1\_S:K346R. We also  
122 performed a BLAST search for isolates with amino acid sequences of BA.1-0.1.1\_S:OaaXXXWaa, as described  
123 in Methods. As a result, Omicron BA.1.1-subset-0.1 isolates were detected at all mutation sites except S373P  
124 (Fig. 2B). Similar to the BA.1 lineage of the Omicron variant, in the BA.2 lineage of the Omicron variant,  
125 isolates of BA.2-0.1 were found at all mutant sites except T478K and P681H in the S protein (Supplemental  
126 Figure 2). The presence of these isolates refutes the establishment of Omicron strains through a continuous  
127 evolutionary process by accumulating mutations. So, we could not determine which mutation occurred first or  
128 last to form the Omicron variants. As shown in Fig. 2B, over half of the BA.1.1-0.1 isolates have the synonymous  
129 mutation c21595u detected in the S protein. However, this does not help explain the order in which the c21595u  
130 mutation arose. Curiously, in BA.1 strain isolates, this c21595u mutation was only detected in SARS-CoV-  
131 2\_human\_USA\_ID-CDC-LC0481844\_2022 (GenBank: OM409228.1) and SARS-CoV-2\_human\_USA\_MI-  
132 CDC-ASC210597972\_2022 (GenBank: OM396816.1). These isolates commonly lack the G339D mutation.  
133 This synonymous mutation is in a mutation-prone hotspot location. Still, if the same mutation occurred  
134 independently in different isolates, it is highly unnatural for the proportion of c21595u occurrences to be  
135 significantly biased in the Omicron variants BA.1.1-0.1.

136 It has been reported that two different variants were infected in a single cell while establishing various SARS-  
137 CoV-2 variants, causing homologous recombination in the process of viral RNA synthesis, resulting in multiple  
138 variants. On considering that homologous recombination caused the isolates shown in Fig. 2, some of the

139 breakpoints at which strand changes occur by homologous recombination are too short (1nt, 2nt, 3nt, etc.) (Fig.  
140 3 and Supplemental Figure 3). Therefore, it is unreasonable to employ homologous recombination as the basis  
141 for establishing these isolates. Also, most of these isolates were found in the USA between 2021 and 2022;  
142 however, considering that the most prevalent variant in the USA in August 2021 was the Delta variant, it is most  
143 unlikely that it did not cause mutations such as T478K and D614G, which were already prevalent. It is  
144 inconceivable that the oldest variants (such as T478K and D614G), which were no longer prevalent, were  
145 sufficiently present to cause superinfection and be involved in homologous recombination. Also, most of these  
146 isolates were isolated in the USA between 2021 and 2022. Still, given that the isolates primarily prevalent in the  
147 USA in August 2021 were Delta variants, it is unlikely that an older type of variant without the T478K or D614G  
148 mutation was present to cause superinfection and be involved in homologous recombination. This consideration  
149 is supported by the fact that all of these BA.1-0.1 and BA.1.1-0.1 isolates retained the sequence of the BA.1  
150 lineage in all regions except the S protein (Fig. 4). In addition, the fact that all of these BA.1-0.1 and BA.1.1-  
151 0.1 strains retained the sequence of Omicron strain BA.1 except for the S protein also makes it unreasonable to  
152 consider that these isolates arose by homologous recombination with an older type of mutant without the T478K  
153 or D614G mutations (Fig. 4).

154 Furthermore, some of the BA.1 and BA.1-0.1 isolates have mutation subsets (synonymous: u10135c, ORF3:  
155 L106F, N: D343G) up- and downstream of the S gene, and the u10135c and L106F (ORF3) mutations  
156 correspond perfectly. Therefore, it is considered that homologous recombination between the BA.1 variant and  
157 variants without these mutations did not occur during the mutants' formation processes (Fig. 4). The synonymous  
158 mutation c2470u occurred in BA.1.1 compared with BA.1, and this c2470u mutation was retained by most,  
159 excluding a few of the BA.1.1-0.1 isolates (SARS-CoV-2\_human\_USA\_IL-CDC-ASC210695497\_2022 :  
160 GenBank: OM770362.1; SARS-CoV-2\_human\_USA\_NY-CDC-LC0450936\_2021: GenBank: OM228453.1) .  
161 The synonymous mutation c2470u has also only been observed in a minimal number of BA.1-0.1 isolates  
162 ( SARS-CoV-2\_human\_USA\_OR-CDC-LC0470830\_2022: GenBank: OM367679.1; SARS-CoV-  
163 2\_human\_USA\_ID-CDC-LC0481844\_2022: GenBank: OM409228.1; SARS-CoV-2\_human\_USA\_MI-CDC-  
164 ASC210597972\_2022: GenBank:OM396816.1; SARS-CoV-2\_human\_USA\_WI-CDC-LC0494047\_2022:  
165 GenBank: OM500517.1) . These results suggest that the establishment of BA.1-0.1 and BA.1.1-0.1 isolates  
166 occurred independently. On the other hand, if reversion mutations caused each of these isolates with one amino  
167 acid different to the Wuhan-type, these isolates could be detected by examining an astronomical number of  
168 isolates. However, these virus strains were detected in the number of sequenced whole genomes (a limited  
169 number), rather than in astronomical numbers examined. The fact that most of these mutations occurred without  
170 synonymous mutations (Fig. 2) suggests that none of them arose as a result of trial-and-error random mutations  
171 in nature. Few synonymous mutations are detected in some BA.1-0.1, BA.1.1-0.1, and BA.2-0.1 isolates (Fig.  
172 2 and Supplemental Figure 2), as seen in other viruses (Supplemental Figure 1). The c25000u is the only  
173 synonymous mutation that did not occur until BA.1, BA.1.1, BA.2, BA.1-0.1 BA.1.1-0.1, and BA.2-0.1 isolates  
174 were formed and was not observed in previous variants such as alpha, beta, gamma, delta, etc. Nevertheless, it  
175 is curious to find the occurrence of mutants with synonymous mutations such as c22120u, c24034u, c23635u,  
176 c24448u, c21811u, a23884g, c22987u, c23609a, c23413u, c23896u, c22879u, u24097a, c23893u, c24442u,  
177 u24847c, c24382u, c22264u, c22879u, c22480u, u21976c, c22480u, g24577a, and u23101c in BA.1.1, BA.1-  
178 0.1, and BA.1.1-0.1 isolates (Fig. 2 Synonymous Others), and a22948g, c23635u, c21859u, c22945u, c23701u,  
179 c22987u, a24433g, c23347u, u24640c, a24619g, c24865u, a23989g, u23047c, u24346c, c21811u, c21952u,  
180 a22753u, c23635u, c24023u, c24382u, and c22572u in BA.2-0.1 isolates (Supplemental Figure 2 Synonymous  
181 Others) after the formation of mutants with these subsets.

182 If two different viral variants infect a single cell simultaneously in the process of establishing various SARS-  
183 CoV-2 variants, and if homologous recombination occurs during viral RNA synthesis between the Omicron  
184 variant BA.1 lineage and BA.2 lineage, it is expected that there are variants caused by homologous  
185 recombination between the BA.1 and BA.2 lineages.

186 Therefore, we also performed BLAST searches for isolates with mutant amino acid subsets of both the Omicron  
187 variant BA.1 and BA.2 strains. We detected recombinant isolates of Omicron BA.1 and BA.2 lineages.  
188 Surprisingly, the recombinant Omicron BA.1 and BA.2 lineages, SARS-CoV-2/human/PRI/PR-PR-UPRRP-

189 582/2020 (GenBank: ON928946.1), were already present in Puerto Rico in 2020. Omicron (B.1.1.529) is a  
190 variant of SARS-CoV-2 first reported to WHO by the Network for Genomics Surveillance in South Africa on  
191 November 24, 2021 (13, 14). It was first detected in Botswana and spread to become the predominant variant in  
192 circulation worldwide (15). Following the appearance of the original B.1.1.529 variant, several subvariants of  
193 Omicron emerged, including BA.1, BA.2, BA.3, BA.4, and BA.5 (19). After October 2022, two subvariants of  
194 BA.5 called BQ.1 and BQ.1.1 emerged.

195 The question then arose about why a recombinant strain, SARS-CoV-2/human/PRI/PR-UPRRP-582/2020  
196 (GenBank: ON928946.1), already existed in 2020. We searched for SARS-CoV-2 isolates prevalent in Puerto  
197 Rico using the keywords "PRI", "PR-UPRRP", and "2020". Consequently, we found 29 Omicron-associated  
198 sequences in the 127 hits obtained (Fig. 5B). These results suggest that the SARS-CoV-2 variants bearing the  
199 amino acid sequences of the S protein are identical to Omicron BA.1 and Omicron BA.2 variants, which were  
200 already prevalent in Puerto Rico in 2020, with 15 isolates showing the complete Omicron BA.1+ R346K\_mut-  
201 subset (BA1.1) , and 14 isolates showing a synonymous substitution of c21595u. Four isolates had an amino  
202 acid sequence of the S protein that perfectly matched that of Omicron BA2 (BA.2\_S), five isolates were Omicron  
203 BA.2-0.1 (BA.2-S:K440N) and four isolates were Omicron BA.2-0.1 (BA.2-S:K440N)+F79S, BA.2-0.1 (BA.2-  
204 S:K440N)+Q613H, BA.2-0.1 (BA.2-S:K440N)+212S+D215E and BA.2-0.1 (BA.2-S:K440N)+212S (Fig. 5B).

205

### 206 3 Discussion

207 Several hypotheses have been proposed in which the original SARS-CoV-2 virus resulted from an accidental  
208 laboratory spill. With recent developments in biotechnology, many viruses, including coronaviruses, have been  
209 artificially synthesized and used in various experiments (20-22). The artificial generation of mutant viruses in  
210 laboratories and study of viral phenotypes by introducing mutations is called "reverse genetics", being a common  
211 technique in virology. It has been claimed that SARS-CoV-2 must have been artificially generated because of  
212 the unnatural presence of a codon (CGG) encoding a contiguous arginine at the furin cleavage site of SARS-  
213 CoV-2. This claim has been refuted based on the following facts: 1) there is no logical reason for a genetically  
214 engineered virus to utilize such a suboptimal furin cleavage site; 2) The only previous study on artificial insertion  
215 of furin cleavage sites at the S1/S2 boundary of the S protein of SARS-CoV-1 using the pseudotype virus  
216 experimental system utilized the optimal "RRSRR" sequence, which is different from the furin cleavage site's  
217 sequence present in SARS-CoV-2; 3) There is no evidence of previous studies at the Wuhan Institute of Virology  
218 that artificially inserted a complete furin cleavage site in coronaviruses; 4) Unnatural CGG codons adjacent to  
219 arginine at the furin cleavage site are rare in coronaviruses but are observed at a particular frequency in SARS-  
220 CoV-1, SARS-CoV-2, and other human coronaviruses. However, these are only declarations and are not logical.  
221 No one has offered an explanation why a naturally occurring virus would utilize a suboptimal furin cleavage  
222 site. There has been no mention of the technical possibility of inserting this furin cleavage site or a CGG codon  
223 artificially. The insertion of a polybasic furin cleavage site into the S protein makes it impossible to conclude  
224 whether SARS-CoV-2 is a naturally occurring or an artificial virus.

225 Despite the accumulation of many mutations in the S protein of Omicron mutants, most of the mutations are  
226 non-synonymous, with only one synonymous mutation of c25000u, which is highly unnatural, leading to the  
227 hypothesis that the Omicron mutants were artificially synthesized. The following results presented in this study  
228 may support the hypothesis that the Omicron variants were artificially synthesized rather than naturally  
229 occurring: 1) the presence of Omicron variant-associated isolates with one mutation site being the Wuhan-type;  
230 2) the almost complete absence of synonymous mutations in the S protein in these isolates; 3) the Omicron  
231 variant, which should have been first reported to WHO from South Africa on November 24, 2021, was already  
232 endemic in Puerto Rico in 2020, and there were isolates that were recombinants between Omicron strains BA1  
233 and BA2. In addition, the Omicron mutant-related isolates (BA.1-0.1, BA.1.1-0.1, and BA.2-0.1 isolates) with  
234 a Wuhan-type mutation at one of the mutation sites were established. Some had synonymous mutations after  
235 establishing the Omicron mutant-related isolates (Fig. 2 and Supplemental Figure 2 Synonymous Others). It is  
236 reasonable to assume that viruses with the reversion amino acid mutations caused by non-synonymous mutations

237 in the S protein were artificially synthesized and then acquired further synonymous mutations in the natural  
238 environment.

239 Assuming that artificially synthesized mutants with only non-synonymous mutations are spread globally, this  
240 would explain how mutants with non-synonymous mutations without previous synonymous mutations develop  
241 synonymous mutations under natural circumstances. Considering the current epidemic situation of SARS-CoV-  
242 2, it is unlikely that these viruses arose spontaneously. Based on our efforts to explain the formation of the  
243 SARS-CoV-2 isolates, they were formed by a completely new mechanism that cannot be explained by previous  
244 biology.

245 One idea, the hypothesis that these viruses were artificially generated, is more reasonable than proposing a novel  
246 mutation acquisition mechanism. However, is there any reason to artificially create these mutants, which are  
247 unlikely to have occurred naturally, given the current SARS-CoV-2 epidemic?

248 It is known that the pathogenicity, host specificity, cell tropism, and immunogenicity of numerous viruses can  
249 be altered by mutation of a single (or several) amino acid(s) of a viral protein on the viral envelope (envelope  
250 protein, HA protein, spike protein, etc.). A single-amino-acid substitution in the HA protein of the 2009  
251 pandemic A (H1N1) influenza viruses changed their replication and pathogenicity (23). In the Chikungunya  
252 virus, single amino acid changes in the E2 glycoprotein influenced glycosaminoglycan utilization for target-cell  
253 binding (24), and a single amino acid change in the E1 glycoprotein affected mosquito vector specificity and  
254 epidemic potential (25). In previous coronaviruses such as MERS-CoV and SARS-CoV-1, point mutations have  
255 been demonstrated to confer resistance to neutralizing antibodies (26-28).

256 Consider that the SARS-CoV-2 Omicron variant and its one-amino-acid reversion mutants were artificially and  
257 systematically generated. In that case, we should suspect that the other variants (Alpha to Delta) were also  
258 artificially generated viruses. Indeed, the lack of findings to date that many of the various mutations seen,  
259 especially in the early variants, are indeed associated with increased viral infection (29) supports the hypothesis  
260 that each variant was artificially synthesized to identify the amino acids of the S protein responsible for  
261 infectivity and pathogenicity. The possibility that the set of mutants was artificially generated to identify amino  
262 acids of the S protein involved in the infectivity and virulence is supported.

263 Reverse genetics experiments are an essential part of virus research, and it is inimical to virus research to  
264 consider that artificially synthesized viruses were deliberately spread throughout the world. However, now that  
265 reverse genetics has become common in virus research, we believe it is not scientific to discuss the mutation  
266 process of SARS-CoV-2 without excluding the possibility of artificially synthesized viruses.

267 Finally, we would like to add that while artificially synthesized viruses may have spread, we are not criticizing  
268 reverse genetics technology, as such technology has led to marked advances in virology. In addition, our analysis  
269 employed databases with a limited number of viral sequences, and we cannot deny the possibility that unreliable  
270 data may have been registered due to technical problems in sequencing or some other issues. Furthermore, we  
271 do not conclude that these viruses were artificially synthesized and distributed based on malicious intent. This  
272 study aims to point out that SARS-CoV-2 has undergone unthinkable mutations based on conventional  
273 coronavirus mutation mechanisms, and we hope that the possibility of artificial creation is included in serious  
274 discussions on the formation of SARS-CoV-2 variants.

275 Nonetheless, the analysis we have shown here concludes that the Omicron variants were formed by a completely  
276 new mechanism that cannot be explained by previous biology. The process of how SARS-CoV-2 mutations  
277 occurred should prompt a reconsideration of the SARS-CoV-2 pandemic.

278 **4 Methods**

279 **4. 1 Data acquisition**

280 The SARS-CoV-2 RNA genome, genes, and proteins according to an annotation of SARS-CoV-2 Wuhan-Hu-  
281 H1 (COVID-19/Wuhan-Hu-1CHN/2019/First Isolate) NCBI Reference Sequence: NC\_045512.2 were used as  
282 references for the definition of mutations, and provided a basis for the numbering of nucleotides and amino acids  
283 of each protein. Genome data of SARS-CoV-2 isolates described in this article were obtained from the NCBI  
284 Nucleotide database (<https://www.ncbi.nlm.nih.gov/>) on 25/11/2022 to 17/03/2023.

285 **4. 2 Query of representative SARS-CoV-2 variant genome**

286 Amino acid sequences of spike protein of SARS-CoV-2 variants (Alpha:B.1.1.7, Beta:B.1.351, Gamma:P1,  
287 Delta:B.1.617.2.63, Lambda:C.37, Mu:B.1.621, Omicron:BA.1, BA.1.1, and BA.2) were obtained from an  
288 Internet site, Stanford Coronavirus Antiviral & Resistance Database (<https://covdb.stanford.edu/>) or Covariant  
289 (<https://covariants.org/>), and used as a query sequence for an NCBI protein BLAST search (blastp: protein-  
290 protein BLAST,  
291 [https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&PAGE\\_TYPE=BlastSearch&LINK\\_LOC=blasthome](https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&PAGE_TYPE=BlastSearch&LINK_LOC=blasthome)). Then, the whole genome sequence of each isolate bearing the query spike sequence was derived from the  
292 BLAST search result, identified with query amino acid sequences of 100%. The nucleotide sequences of the  
293 detected SARS-CoV-2 genome were as follows: GenBank Accession No.: GenBank: MW423686.2;  
294 MW430966.1; MW430967.1; MW422256.1; MW598419.1; MW667552.1; MW667553.1; MW721502.1;  
295 MW721504.1; MW520923.1; MW642248.1; MW642249.1; MW642250.1; MZ182066.1; MZ155303.1;  
296 MZ155230.1; MZ170364.1; MZ179869.1; MW666666.1; MW696612.1; MW699217.1; MW644498.1;  
297 MZ727706.1; MZ620161.1; MZ637380.1; MZ637401.1; MZ780550.1; OL672836.1; OL677199.1;  
298 OP769083.1; OL764360.1; OL815447.1; ON762438.1; OL849989.1; OL897126.1; OL896945.1;  
299 OL896936.1; OL896931.1; OM233931.1; OM072551.1; OM072822.1; OM296922.1.

302 **4. 3 Query of SARS-CoV-2 Omicron variant genome bearing an S protein amino acid sequence in**  
303 **which one of the Omicron-type nucleotide mutation subsets was not mutated and retains the original**  
304 **SARS-CoV-2 Wuhan-Hu-H1-type arrangement.**

305 For each of the Omicron variants, BA.1, BA.1.1, and BA.2, the isolate series bearing an S protein amino acid  
306 sequence in which one of the Omicron-type nucleotide mutation subsets is not mutated and retains the original  
307 SARS-CoV-2 Wuhan-Hu-H1-type arrangement were named BA.1-0.1, BA.1.1-0.1 and BA.2-0.1, respectively.  
308 In addition, in this article, we named the amino acid sequences of spike protein of BA.1, BA.1.1, and BA.2 as  
309 BA.1\_S, BA.1.1\_S, and BA.2\_S, respectively, and then the series of amino acid sequences of spike protein of  
310 BA.1-0.1, BA.1.1-0.1, and BA.2-0.1 were named, respectively, as follows: Omicron BA.1-0.1 spike series  
311 (BA.1-0.1\_Ss) were named as BA.1\_S:V67A; BA.1\_S:69H\_70V; BA.1\_S:I95T;  
312 BA.1\_S:D142G\_143V\_144Y\_145Y; BA.1\_S:I211N\_212L; BA.1\_S:ΔEPE; BA.1\_S:D339G; BA.1\_S:L371S;  
313 BA.1\_S:P373S; BA.1\_S:F375S; BA.1\_S:N417K; BA.1\_S:K440N; BA.1\_S:S446G; BA.1\_S:N477S;  
314 BA.1\_S:K478T; BA.1\_S:A484E; BA.1\_S:R493Q; BA.1\_S:S496G; BA.1\_S:R498Q; BA.1\_S:Y501N;  
315 BA.1\_S:H505Y; BA.1\_S:K547T; BA.1\_S:G614D; BA.1\_S:Y655H; BA.1\_S:K679N; BA.1\_S:H681P;  
316 BA.1\_S:K764N; BA.1\_S:Y796D; BA.1\_S:K856N; BA.1\_S:H954Q; BA.1\_S:K969N and BA.1\_S:F981L /  
317 Omicron BA.1.1-0.1 spike series (BA.1.1-0.1\_Ss) were named as BA.1.1\_S:V67A; BA.1.1\_S:69H\_70V;  
318 BA.1.1\_S:I95T; BA.1.1\_S:D142G\_143V\_144Y\_145Y; BA.1.1\_S:I211N\_212L; BA.1.1\_S:ΔEPE;  
319 BA.1.1\_S:D339G; BA.1.1\_S:L371S; BA.1.1\_S:P373S; BA.1.1\_S:F375S; BA.1.1\_S:N417K;  
320 BA.1.1\_S:K440N; BA.1.1\_S:S446G; BA.1.1\_S:N477S; BA.1.1\_S:K478T; BA.1.1\_S:A484E;  
321 BA.1.1\_S:R493Q; BA.1.1\_S:S496G; BA.1.1\_S:R498Q; BA.1.1\_S:Y501N; BA.1.1\_S:H505Y;  
322 BA.1.1\_S:K547T; BA.1.1\_S:G614D; BA.1.1\_S:Y655H; BA.1.1\_S:K679N; BA.1.1\_S:H681P;  
323 BA.1.1\_S:K764N; BA.1.1\_S:Y796D; BA.1.1\_S:K856N; BA.1.1\_S:H954Q; BA.1.1\_S:K969N;  
324 BA.1.1\_S:F981L / Omicron BA.2-0.1 spike series (BA.2-0.1\_Ss) were named as BA.2\_S:I19T;

325 BA.2\_S:24L\_25P\_26P\_S27A; BA.2\_S:D142G; BA.2\_S:V213G; BA.2\_S:D339G; BA.2\_S:F371S;  
326 BA.2\_S:P373S; BA.2\_S:F375S; BA.2\_S:A376T; BA.2\_S:N405D; BA.2\_S:S408R; BA.2\_S:N417K;  
327 BA.2\_S:K440N; BA.2\_S:N477S; BA.2\_S:K478T; BA.2\_S:A484E; BA.2\_S:R493Q; BA.2\_S:R498Q;  
328 BA.2\_S:Y501N; BA.2\_S:H505Y; BA.2\_S:G614D; BA.2\_S:Y655H; BA.2\_S:K679N; BA.2\_S:H681P;  
329 BA.2\_S:K764N; BA.2\_S:Y796D; BA.2\_S:H954Q; BA.2\_S:K969N, and these constructs are shown in Figs. 2,  
330 4 and supplemental Figure 1. These amino acids sequences of spike protein of SARS-CoV-2 Omicron variants,  
331 BA.1-0.1, BA.1.1-0.1, and BA.2-0.1, were used as query sequences for an NCBI protein BLAST search. Then,  
332 the whole genome sequences of BA.1-0.1, BA.1.1-0.1, and BA.2-0.1 isolates bearing the query spike sequence  
333 were derived from the BLAST search results, identified with a query amino acid sequence of 100%. The  
334 nucleotide sequences of the detected SARS-CoV-2 genome were as follows: GenBank Accession No.:  
335 OM117411.1; OP797378.1; OM789835.1; OP928789.1; OP928803.1; OP929381.1; OP929396.1;  
336 OP929417.1; OM173977.1; OM518459.1; OM566981.1; ON019560.1; OM097227.1; OM096937.1;  
337 OM099902.1; OM117114.1; OM096685.1; OM354436.1; OM646886.1; OM472901.1; OM364511.1;  
338 OM131858.1; OL815451.1; OL896986.1; OL897116.1; OL897118.1; OL896964.1; OM367679.1;  
339 OM343778.1; OM409228.1; OM396816.1; OM134162.1; OM075886.1; OM123427.1; OM122677.1;  
340 OM121681.1; OM224850.1; ON246090.1; OM931599.1; OM864873.1; OM906519.1; OM906587.1;  
341 OM464776.1; OM015999.1; OM015958.1; OM015597.1; OM016329.1; OL898806.1; OL898861.1;  
342 OM016937.1; OM016186.1; OM036549.1; OM051171.1; OM126493.1; OM079115.1; OM099199.1;  
343 OM134489.1; OM098796.1; ON618279.1; ON618009.1; OM627701.1; OM356511.1; OM295457.1;  
344 ON700063.1; OM033824.1; ON368355.1; OM084700.1; ON208126.1; OM566593.1; OM945690.2;  
345 ON030252.1; ON019844.1; OM890075.1; ON020044.1; OM833954.1; ON376082.1; OM084604.1;  
346 OP795273.1; ON066609.1; OM352882.1; OM290510.1; OM369978.1; OM199342.1; OM011974.1;  
347 OM090274.1; OM043984.1; OM121683.1; OM121624.1; OM175506.1; OM360429.1; OM360221.1;  
348 OM358058.1; OM500517.1; OM135027.1; OM742858.1; OM521685.1; OM896558.1; ON694155.1;  
349 OM686755.1; OM484260.1; OM332056.1; OM156397.1; OM079447.1; OM134645.1; OM173298.1;  
350 OM123082.1; OM116023.1; OM652943.1; OL994299.1; OL994920.1; OM122027.1; OM121015.1;  
351 OL898817.1; OM527504.1; OM225320.1; OM931491.1; OM931575.1; OM931587.1; OM034409.1;  
352 OM036283.1; OL996129.1; OM035680.1; OM096996.1; ON065532.1; OM968098.1; OM816604.1;  
353 ON235452.1; ON334146.1; OP024162.1; OP209732.1; OM354578.1; OM099080.1; OM297301.1;  
354 OM297438.1; OM365368.1; OM449159.1; OM078863.1; OM096959.1; OM117155.1; OM133880.1;  
355 OM077358.1; OM372005.1; OM770362.1; OM897488.1; OM918459.1; OM918478.1; OL897115.1;  
356 OL897114.1; OL986779.1; OL986696.1; OL987046.1; ON831866.1; OM864099.1; OM863888.1;  
357 OP745925.1; ON831672.1; OM043643.1; OM176192.1; OM226685.1; OM343689.1; OM295527.1;  
358 OM894975.1; OM846676.1; OM822024.1; OM846844.1; OM906550.1; OM015933.1; OM016323.1;  
359 OM016331.1; OM035685.1; OM022498.1; OM156115.1; OM036875.1; OM099560.1; OM199246.1;  
360 OM067048.1; OM079299.1; OM099911.1; OM116588.1; OM097010.1; OM173300.1; OM805961.1;  
361 OM983266.1; OM983325.1; ON618010.1; OM084691.1; ON021265.1; ON039239.1; ON056981.1;  
362 ON144127.1; OM770527.1; OM156164.1; OM155119.1; OM199353.1; OM084630.1; OM084605.1;  
363 OM084621.1; OM359369.1; OM411574.1; OM584789.1; OM720486.1; OM429777.1; ON047062.1;  
364 ON065416.1; OP415118.1; OM954373.1; ON042406.1; OM335528.1; OM332335.1; OM353626.1;  
365 OM332813.1; OM197398.1; OM226919.1; OM228399.1; OM225859.1; OM271353.1; OM159454.1;  
366 OM224473.1; OM358278.1; OM361030.1; OM412141.1; OM496298.1; OM044048.1; OM121864.1;  
367 OM224477.1; OM227379.1; OM228453.1; OM622156.1; OM906370.1; OM970683.1; ON117965.1;  
368 OM198667.1; OM357800.1; OM357161.1; OM335230.1; OM261124.1; OM077578.1; OM497172.1;  
369 OM625194.1; OM907131.1; ON047464.1; OM911851.1; OM042846.1; OM155337.1; OM097339.1;  
370 OM116805.1; OM134409.1; OM686782.1; OM695863.1; OM724725.1; OM174366.1; OM822132.1;  
371 OM822106.1; OM822105.1; OM822485.1; OM135143.1; OM125829.1; OM098855.1; OM156118.1;  
372 OM155114.1; OM863926.1; OP359104.1; ON209298.1; ON232806.1; ON421981.1; ON811217.1;  
373 OM698275.1; ON052769.1; ON060006.1; ON060007.1; ON060009.1; OM843171.1; OM843276.1;  
374 OM843550.1; OM843316.1; OM843340.1; ON049267.1; ON450720.1; ON250163.1; ON256603.1;  
375 ON480422.1; OM888844.1; OM890089.1; ON009425.2; ON082904.1; OM901275.1; OM877094.2;  
376 OM877095.2; OM877096.2; OM877097.2; ON378542.1; ON389858.1; ON389889.1; ON390359.1;  
377 OM936703.1; ON352711.1; ON378000.1; ON177702.1; ON205494.1; ON378633.1; ON617689.1;



378 ON619375.1; OM567618.1; OM659585.1; OM770913.1; OM781641.1; OM533441.1; OM533458.1;  
379 OM570235.1; OM570252.1; OM570249.1; OM283361.1; OM283362.1; OM283320.1; OM283343.1;  
380 ON618014.1; ON618018.1; ON618019.1; ON618363.1; ON311615.1; ON383919.1; OP579158.1;  
381 OP054411.1; ON633107.1; ON414693.1; ON422887.1; OP364296.1; OP629673.1; ON363097.1;  
382 OP633561.1; ON458445.1; ON592247.1; ON549687.1; ON067040.1; ON321116.1; ON199452.1;  
383 ON200331.1; OM861064.1; OM969592.1; ON019120.1; ON221861.1; OM861619.1; ON091288.1;  
384 ON151370.1; ON233850.1; ON236456.1; ON296711.1; ON535443.1; ON624524.1; ON377450.1;  
385 ON397268.1; ON239032.1; ON373649.1; ON481637.1; ON701163.1; ON312677.1; ON349263.1;  
386 ON377487.1; ON377609.1; OM638574.1; OM911616.1; OM988767.1; ON019770.1; OM988769.1;  
387 ON468158.1; ON608924.1; ON604965.1; ON535763.1; ON378227.1; ON378238.1; ON728470.1.

#### 388 4. 4 Query of recombinant SARS-CoV-2 Omicron variant between BA.1 and BA.2 genome

389 Deduced recombinant spike protein between Omicron variants, BA.1 and BA.2 shown as BA.1\_S:T19I\_L24-  
390 \_P25- \_P26- \_A27S\_V213G\_S371F\_T376A\_D405N\_R408S was used as a query sequence for an NCBI  
391 protein BLAST search. The whole genome sequence of BA.1 and BA.2 recombinant-Omicron isolates showed  
392 some of the specific amino acid mutations observed in variant BA.1 and BA.2 in the S protein. The nucleotide  
393 sequences of the detected SARS-CoV-2 genome were as follows: GenBank Accession No.: OM360636.1;  
394 OM410816.1; OM429902.1; OM497964.1; OM565587.1; OM628132.1; ON549899.1; ON449685.1;  
395 ON176765.1; OM628094.1; ON099844.1; OM942313.1; ON395480.1; ON171854.1; ON172005.1;  
396 ON076710.1; ON928946.1; OM932113.1; OM942438.1; OM989528.1; OM499181.1; ON414822.1;  
397 OM878325.1; ON103067.1; ON103153.1; ON419036.1; ON928719.1; ON337887.1; ON420444.1;  
398 ON146520.1; OM469541.1; OM904085.1; ON254531.1; OM881098.1; ON373310.1.

#### 399 4. 5 Query of SARS-CoV-2 Omicron variant genome detected in Puerto Rico in 2020

400 Nucleotide sequences were searched using the keywords PRI PR-UPRRP 2020 (Search details: PRI[All  
401 Fields] AND (PR[All Fields] AND UPRRP[All Fields]) AND 2020[All Fields]). The search results were all  
402 SARS-CoV-2 isolate genome sequences. Among these sequences, SARS-CoV-2 Omicron variant-related  
403 sequences were picked up as follows: GenBank Accession No.: ON928761.1; ON928660.1; ON928794.1;  
404 ON928762.1; ON928848.1; ON928741.1; ON928918.1; ON928680.1; ON928975.1; ON928949.1;  
405 ON928673.1; ON928865.1; ON928716.1; ON928663.1; ON928779.1; ON928896.1; ON928946.1;  
406 ON928912.1; ON928704.1; ON928873.1; ON928813.1; ON928898.1; ON928765.1; ON928912.1;  
407 ON928883.1; ON928957.1; ON928880.1; ON928699.1; ON928724.1; ON928941.1.

408 Genomes were aligned using SnapGene software or GENETYX software. Numbering of nucleotides and  
409 amino acids of each protein was determined using Wuhan-Hu-1 (NC\_045512.2; COVID-19/Wuhan-Hu-  
410 1CHN/2019/First Isolate) as a reference strain for the definition of mutations.

411 5. References

- 412 1. F. Wu, S. Zhao, B. Yu, Y. M. Chen, W. Wang, Z. G. Song, Y. Hu, Z. W. Tao, J. H. Tian, Y. Y.  
413 Pei, M. L. Yuan, Y. L. Zhang, F. H. Dai, Y. Liu, Q. M. Wang, J. J. Zheng, L. Xu, E. C. Holmes and Y.  
414 Z. Zhang: A new coronavirus associated with human respiratory disease in China. *Nature*, 579(7798),  
415 265-269 (2020) doi:10.1038/s41586-020-2008-3
- 416 2. R. A. Khailany, M. Safdar and M. Ozaslan: Genomic characterization of a novel SARS-CoV-  
417 2. *Gene Rep*, 19, 100682 (2020) doi:10.1016/j.genrep.2020.100682
- 418 3. Y. M. Bar-On, A. Flamholz, R. Phillips and R. Milo: SARS-CoV-2 (COVID-19) by the  
419 numbers. *Elife*, 9 (2020) doi:10.7554/eLife.57309
- 420 4. J. Lan, J. Ge, J. Yu, S. Shan, H. Zhou, S. Fan, Q. Zhang, X. Shi, Q. Wang, L. Zhang and X.  
421 Wang: Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor.  
422 *Nature*, 581(7807), 215-220 (2020) doi:10.1038/s41586-020-2180-5
- 423 5. D. Wrapp, N. Wang, K. S. Corbett, J. A. Goldsmith, C. L. Hsieh, O. Abiona, B. S. Graham and  
424 J. S. McLellan: Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science*,  
425 367(6483), 1260-1263 (2020) doi:10.1126/science.abb2507
- 426 6. B. Korber, W. M. Fischer, S. Gnanakaran, H. Yoon, J. Theiler, W. Abfalterer, N. Hengartner,  
427 E. E. Giorgi, T. Bhattacharya, B. Foley, K. M. Hastie, M. D. Parker, D. G. Partridge, C. M. Evans, T.  
428 M. Freeman, T. I. de Silva, C.-G. G. Sheffield, C. McDanal, L. G. Perez, H. Tang, A. Moon-Walker,  
429 S. P. Whelan, C. C. LaBranche, E. O. Saphire and D. C. Montefiori: Tracking Changes in SARS-CoV-  
430 2 Spike: Evidence that D614G Increases Infectivity of the COVID-19 Virus. *Cell*, 182(4), 812-827 e19  
431 (2020) doi:10.1016/j.cell.2020.06.043
- 432 7. A. Carrasco-Montalvo, A. Bruno, D. de Mora, M. Olmedo, J. Garces, M. Paez, M. Regato-  
433 Arrata, M. Gonzalez, J. Romero, O. Mestanza, B. Freire-Paspuel, A. Gaviria, S. A. Orlando, M. A.  
434 Garcia-Bereguian and L. Patino: First Report of SARS-CoV-2 Lineage B.1.1.7 (Alpha Variant) in  
435 Ecuador, January 2021. *Infect Drug Resist*, 14, 5183-5188 (2021) doi:10.2147/IDR.S319439
- 436 8. S. Zarate, B. Taboada, J. E. Munoz-Medina, P. Isa, A. Sanchez-Flores, C. Boukadida, A.  
437 Herrera-Estrella, N. Selem Mojica, M. Rosales-Rivera, B. Gomez-Gil, A. G. Salas-Lais, C. E.  
438 Santacruz-Tinoco, H. Montoya-Fuentes, J. E. Alvarado-Yaah, G. M. Molina-Salinas, G. E. Espinoza-  
439 Ayala, J. A. Enciso-Moreno, R. M. Gutierrez-Rios, A. Loza, J. Moreno-Contreras, R. Garcia-Lopez,  
440 X. Rivera-Gutierrez, A. Comas-Garcia, R. M. Wong-Chew, M. E. Jimenez-Corona, R. M. Del Angel,  
441 J. A. Vazquez-Perez, M. Matias-Florentino, M. Perez-Garcia, S. Avila-Rios, H. G. Castelan-Sanchez,  
442 L. Delaye, L. P. Martinez-Castilla, M. Escalera-Zamudio, S. Lopez and C. F. Arias: The Alpha Variant  
443 (B.1.1.7) of SARS-CoV-2 Failed to Become Dominant in Mexico. *Microbiol Spectr*, 10(2), e0224021  
444 (2022) doi:10.1128/spectrum.02240-21
- 445 9. H. Tegally, E. Wilkinson, M. Giovanetti, A. Iranzadeh, V. Fonseca, J. Giandhari, D. Doolabh,  
446 S. Pillay, E. J. San, N. Msomi, K. Mlisana, A. von Gottberg, S. Walaza, M. Allam, A. Ismail, T. Mohale,  
447 A. J. Glass, S. Engelbrecht, G. Van Zyl, W. Preiser, F. Petruccione, A. Sigal, D. Hardie, G. Marais, N.-  
448 y. Hsiao, S. Korsman, M.-A. Davies, L. Tyers, I. Mudau, D. York, C. Maslo, D. Goedhals, S. Abrahams,  
449 O. Laguda-Akingba, A. Alisoltani-Dehkordi, A. Godzik, C. K. Wibmer, B. T. Sewell, J. Lourenço, L.  
450 C. J. Alcantara, S. L. Kosakovsky Pond, S. Weaver, D. Martin, R. J. Lessells, J. N. Bhiman, C.  
451 Williamson and T. de Oliveira: Detection of a SARS-CoV-2 variant of concern in South Africa. *Nature*,  
452 592(7854), 438-443 (2021) doi:10.1038/s41586-021-03402-9
- 453 10. T. Fujino, H. Nomoto, S. Kutsuna, M. Ujiie, T. Suzuki, R. Sato, T. Fujimoto, M. Kuroda, T.  
454 Wakita and N. Ohmagari: Novel SARS-CoV-2 Variant in Travelers from Brazil to Japan. *Emerg Infect*  
455 *Dis*, 27(4), 1243-5 (2021) doi:10.3201/eid2704.210138
- 456 11. M. Joshi, M. Kumar, V. Srivastava, D. Kumar, D. S. Rathore, R. Pandit, D. W. Graham and C.  
457 G. Joshi: Genetic sequencing detected the SARS-CoV-2 delta variant in wastewater a month prior to

458 the first COVID-19 case in Ahmedabad (India). *Environ Pollut*, 310, 119757 (2022)  
459 doi:10.1016/j.envpol.2022.119757

460 12. K. Laiton-Donato, C. Franco-Munoz, D. A. Alvarez-Diaz, H. A. Ruiz-Moreno, J. A. Usme-  
461 Ciro, D. A. Prada, J. Reales-Gonzalez, S. Corchuelo, M. T. Herrera-Sepulveda, J. Naizaque, G.  
462 Santamaria, J. Rivera, P. Rojas, J. H. Ortiz, A. Cardona, D. Malo, F. Prieto-Alvarado, F. R. Gomez, M.  
463 Wiesner, M. L. O. Martinez and M. Mercado-Reyes: Characterization of the emerging B.1.621 variant  
464 of interest of SARS-CoV-2. *Infect Genet Evol*, 95, 105038 (2021) doi:10.1016/j.meegid.2021.105038

465 13. J. Quarleri, V. Galvan and M. V. Delpino: Omicron variant of the SARS-CoV-2: a quest to  
466 define the consequences of its high mutational load. *Geroscience*, 44(1), 53-56 (2022)  
467 doi:10.1007/s11357-021-00500-4

468 14. A. Gowrisankar, T. M. C. Priyanka and S. Banerjee: Omicron: a mysterious variant of concern.  
469 *Eur Phys J Plus*, 137(1), 100 (2022) doi:10.1140/epjp/s13360-021-02321-y

470 15. A. Vitiello, F. Ferrara, A. M. Auti, M. Di Domenico and M. Boccellino: Advances in the  
471 Omicron variant development. *J Intern Med*, 292(1), 81-90 (2022) doi:10.1111/joim.13478

472 16. Y. Fan, X. Li, L. Zhang, S. Wan, L. Zhang and F. Zhou: SARS-CoV-2 Omicron variant: recent  
473 progress and future perspectives. *Signal Transduct Target Ther*, 7(1), 141 (2022) doi:10.1038/s41392-  
474 022-00997-x

475 17. J. Fonager, M. Bennedbaek, P. Bager, J. Wohlfahrt, K. M. Ellegaard, A. C. Ingham, S. M.  
476 Edslev, M. Stegger, R. N. Sieber, R. Lassauniere, A. Fomsgaard, T. Lillebaek, C. W. Svarrer, F. T.  
477 Moller, C. H. Moller, R. Legarth, T. V. Sydenham, K. Steinke, S. J. Paulsen, J. A. S. Castruita, U. V.  
478 Schneider, C. H. Schouw, X. C. Nielsen, M. Overvad, R. T. Nielsen, R. L. Marvig, M. S. Pedersen, L.  
479 Nielsen, L. L. Nilsson, J. Bybjerg-Grauholm, I. H. Tarpgaard, T. S. Ebsen, J. U. H. Lam, V. Gunalan  
480 and M. Rasmussen: Molecular epidemiology of the SARS-CoV-2 variant Omicron BA.2 sub-lineage  
481 in Denmark, 29 November 2021 to 2 January 2022. *Euro Surveill*, 27(10) (2022) doi:10.2807/1560-  
482 7917.ES.2022.27.10.2200181

483 18. L. B. Shrestha, C. Foster, W. Rawlinson, N. Tedla and R. A. Bull: Evolution of the SARS-  
484 CoV-2 omicron variants BA.1 to BA.5: Implications for immune escape and transmission. *Rev Med*  
485 *Virol*, 32(5), e2381 (2022) doi:10.1002/rmv.2381

486 19. L. Yao, K. L. Zhu, X. L. Jiang, X. J. Wang, B. D. Zhan, H. X. Gao, X. Y. Geng, L. J. Duan, E.  
487 H. Dai and M. J. Ma: Omicron subvariants escape antibodies elicited by vaccination and BA.2.2  
488 infection. *Lancet Infect Dis*, 22(8), 1116-1117 (2022) doi:10.1016/S1473-3099(22)00410-8

489 20. S. Torii, C. Ono, R. Suzuki, Y. Morioka, I. Anzai, Y. Fauzyah, Y. Maeda, W. Kamitani, T.  
490 Fukuhara and Y. Matsuura: Establishment of a reverse genetics system for SARS-CoV-2 using circular  
491 polymerase extension reaction. *Cell Rep*, 35(3), 109014 (2021) doi:10.1016/j.celrep.2021.109014

492 21. T. Y. Taha, I. P. Chen, J. M. Hayashi, T. Tabata, K. Walcott, G. R. Kimmerly, A. M. Syed, A.  
493 Ciling, R. K. Suryawanshi, H. S. Martin, B. H. Bach, C. L. Tsou, M. Montano, M. M. Khalid, B. K.  
494 Sreekumar, G. Renuka Kumar, S. Wyman, J. A. Doudna and M. Ott: Rapid assembly of SARS-CoV-  
495 2 genomes reveals attenuation of the Omicron BA.1 variant through NSP6. *Nat Commun*, 14(1), 2308  
496 (2023) doi:10.1038/s41467-023-37787-0

497 22. W. Wang, X. Peng, Y. Jin, J. A. Pan and D. Guo: Reverse genetics systems for SARS-CoV-2.  
498 *J Med Virol*, 94(7), 3017-3031 (2022) doi:10.1002/jmv.27738

499 23. L. Xu, L. Bao, Q. Lv, W. Deng, Y. Ma, F. Li, L. Zhan, H. Zhu, C. Ma and C. Qin: A single-  
500 amino-acid substitution in the HA protein changes the replication and pathogenicity of the 2009  
501 pandemic A (H1N1) influenza viruses in vitro and in vivo. *Virol J*, 7, 325 (2010) doi:10.1186/1743-  
502 422X-7-325

503 24. L. A. Silva, S. Khomandiak, A. W. Ashbrook, R. Weller, M. T. Heise, T. E. Morrison and T.  
504 S. Dermody: A single-amino-acid polymorphism in Chikungunya virus E2 glycoprotein influences  
505 glycosaminoglycan utilization. *J Virol*, 88(5), 2385-97 (2014) doi:10.1128/JVI.03116-13

- 506 25. K. A. Tsetsarkin, D. L. Vanlandingham, C. E. McGee and S. Higgs: A single mutation in  
507 chikungunya virus affects vector specificity and epidemic potential. *PLoS Pathog*, 3(12), e201 (2007)  
508 doi:10.1371/journal.ppat.0030201
- 509 26. X. C. Tang, S. S. Agnihothram, Y. Jiao, J. Stanhope, R. L. Graham, E. C. Peterson, Y. Avnir,  
510 A. S. Tallarico, J. Sheehan, Q. Zhu, R. S. Baric and W. A. Marasco: Identification of human  
511 neutralizing antibodies against MERS-CoV and their role in virus adaptive evolution. *Proc Natl Acad*  
512 *Sci U S A*, 111(19), E2018-26 (2014) doi:10.1073/pnas.1402074111
- 513 27. J. Sui, D. R. Aird, A. Tamin, A. Murakami, M. Yan, A. Yammanuru, H. Jing, B. Kan, X. Liu,  
514 Q. Zhu, Q. A. Yuan, G. P. Adams, W. J. Bellini, J. Xu, L. J. Anderson and W. A. Marasco: Broadening  
515 of neutralization activity to directly block a dominant antibody-driven SARS-coronavirus evolution  
516 pathway. *PLoS Pathog*, 4(11), e1000197 (2008) doi:10.1371/journal.ppat.1000197
- 517 28. J. ter Meulen, E. N. van den Brink, L. L. Poon, W. E. Marissen, C. S. Leung, F. Cox, C. Y.  
518 Cheung, A. Q. Bakker, J. A. Bogaards, E. van Deventer, W. Preiser, H. W. Doerr, V. T. Chow, J. de  
519 Kruijff, J. S. Peiris and J. Goudsmit: Human monoclonal antibody combination against SARS  
520 coronavirus: synergy and coverage of escape mutants. *PLoS Med*, 3(7), e237 (2006)  
521 doi:10.1371/journal.pmed.0030237
- 522 29. L. van Dorp, D. Richard, C. C. S. Tan, L. P. Shaw, M. Acman and F. Balloux: No evidence for  
523 increased transmissibility from recurrent mutations in SARS-CoV-2. *Nat Commun*, 11(1), 5986 (2020)  
524 doi:10.1038/s41467-020-19818-2

525

## 526 **Conflict of Interest**

527 The authors declare that the research was conducted in the absence of any commercial or financial  
528 relationships that could be construed as a potential conflict of interest.

529 **Figure legends**

530 **Fig. 1. Mutation subsets of S protein of SARS-CoV-2 variants.**

531 Sequences of S protein of SARS-CoV-2 variants (variants of concern, VOCs: Alpha:B.1.1.7, Beta:B.1.351,  
532 Gamma:P1, Delta:B.1.617.2.63, and Omicron:BA.1; BA.2 and variants of interest, VOIs: Lambda:C.37,  
533 Mu:B.1.621) are compared with the SARS-CoV-2 Wuhan-Hu-H1 reference sequence, and different amino acids  
534 (amino acid change, deletion, and insertion) and synonymous changes of nucleotides are shown. Non-  
535 synonymous changes are shown by amino acid changes (capital letters), and synonymous changes are shown by  
536 nucleotide changes (small letters). Amino acids different from Wuhan-Hu-H1 found in each variant: Alpha:  
537 B.1.1.7, Beta: B.1.351, Gamma: P1, Delta: B.1.617.2.63, Lambda: C.37, Mu: B.1.621, and Omicron: BA.1,  
538 BA.2 are highlighted with red, orange, green, yellow, aquamarine, lime, deep sky blue, and blue violet,  
539 respectively. Amino acid changes common to Omicron: BA.1 and BA.2 are shown in purple.

540

541 **Fig. 2. Mutations of S proteins of SARS-CoV-2 Omicron isolates.**

542 **(A)** Different amino acids and synonymous changes of nucleotides in S proteins of SARS-CoV-2 Omicron BA.1,  
543 BA.1.1 isolates, and BA.1-0.1s compared with SARS-CoV-2 Wuhan-Hu-H1. Nucleotide deletions and  
544 insertions were "deletion<sup>1</sup>" (deletion: nt 21,766-21,771), "deletion<sup>2</sup>" (deletion: nt 21,987-21,995), "deletion<sup>3</sup>"  
545 (deletion: nt 22,194-22,196), and "insertion<sup>4</sup>" (insertion between 22,206-22,196), and introduced amino acid  
546 changes were H69\_V70-, G142D\_V143\_Y144\_Y145-, N211I\_L212-, and 215ins.EPE, respectively. **(B)**  
547 Different amino acids and synonymous nucleotide changes in S proteins of SARS-CoV-2 Omicron BA.1.1-0.1  
548 isolates. Amino acids different from Wuhan-Hu-H1 found in each variant: Alpha: B.1.1.7, Beta: B.1.351,  
549 Gamma: P1, Delta: B.1.617.2.63, Mu: B.1.621, and Omicron: BA.1, BA.2 are highlighted with red, orange,  
550 green, yellow, lime, deep sky blue, and blue violet, respectively. Amino acid changes common to Omicron:BA.1  
551 and BA.2 are highlighted with purple.

552

553 **Fig. 3. Estimated homologous recombination breakpoints of the SARS-CoV-2 S gene of Omicron BA.1-**  
554 **0.1 or BA.1.1-0.1.**

555 Sequence alignment of amino acids and their coding nucleotides (nt.21,746-21,787; nt.22,658-22,702;  
556 nt.22,976-23,011, and nt.23,582-23,620) containing the mutation point of the SARS-CoV-2 S gene of the  
557 Omicron BA.1 variant compared with SARS-CoV-2 Wuhan-Hu-H1. Changed nucleotides and amino acids of  
558 Omicron BA.1 are shown in red letters. Estimated homologous recombination breakpoints of the SARS-CoV-  
559 2 S gene of Omicron BA.1-0.1 or BA.1.1-0.1 are shown by asterisks.

560

561 **Fig. 4. Representative mutations of SARS-CoV-2 Omicron isolates other than S protein.**

562 **(A)** Representative amino acids and synonymous changes of nucleotides of SARS-CoV-2 Omicron BA.1,  
563 BA.1.1 isolates, and BA.1-0.1 compared with SARS-CoV-2 Wuhan-Hu-H1. **(B)** Representative amino acids  
564 and synonymous changes of nucleotides of SARS-CoV-2 Omicron BA.1.1-0.1 compared with SARS-CoV-2  
565 Wuhan-Hu-H1. Amino acids different from Wuhan-Hu-H1 found in each variant: Alpha: B.1.1.7, Lambda: C.37,  
566 Mu: B.1.621, and Omicron: BA.1 are highlighted with red, aquamarine, deep sky blue, and blue violet,  
567 respectively.

568 Amino acid changes common to Omicron: BA.1 and BA.2 are shown in purple. Synonymous nucleotide  
569 changes: c2470u observed in Omicron:BA.1.1 mainly shown with blue. Synonymous and non-synonymous  
570 changes: u10135c of nsp5, L106F in ORF3, and D343G in N protein subset observed in ~40% of Omicron;

571 BA.1-0.1 are highlighted with **emerald-green**. Undetermined nucleotides or amino acids are shown as UD or X,  
572 respectively.

573

574 **Fig. 5. Mutations of S proteins of SARS-CoV-2 Omicron BA.1-BA.2 recombinant isolates and SARS-CoV-2 Omicron BA.1 and BA.2 isolates detected in Puerto Rico in 2020.**

576 **(A)** Different amino acids and synonymous change of nucleotides in S proteins of SARS-CoV-2 Omicron BA.1-  
577 BA.2 recombinant isolates compared with SARS-CoV-2 Wuhan-Hu-H1. Nucleotide deletions, "deletion<sup>5</sup>"  
578 (deletion: nt 21,633-21,641), introduced the amino acids changes L24- P25- P26- A27S. **(B)** Different amino  
579 acids and synonymous change of nucleotides in S proteins of SARS-CoV-2 Omicron BA.1.1 and Omicron  
580 BA.1-BA.2 recombinant isolate, highlighted with magenta (GenBank: ON928946.1), Omicron BA.2, and  
581 Omicron 2-0.1(K440N), detected in Puerto Rico in 2020. Amino acids different from Wuhan-Hu-H1 found in  
582 each variant: Alpha: B.1.1.7, Beta: B.1.351, Gamma: P1, Delta: B.1.617.2.63, Mu: B.1.621, and Omicron: BA.1,  
583 BA.2 are highlighted with red, orange, green, yellow, lime, deep sky blue, and blue-violet, respectively. Amino  
584 acid changes common to Omicron: BA.1 and BA.2 are highlighted with purple.

585

586 **Supplemental Figure 1**

587 **Human coronavirus 229E strains detected in Seattle, USA, in 2015 and 2019.**

588 Alignment of nucleotide **(A)** and amino acid **(B)** sequences of the S protein of Human coronavirus 229E strains,  
589 HCoV\_229E/Seattle/USA/SC3112/2015 (GenBank: KY983587.1), and CoV\_229E/Seattle/USA/SC0865/2019  
590 (GenBank: MN306046.1). The number of nucleotide substitutions observed between them was 32, amino acid  
591 substitutions numbered 18 between them, and the synonymous (14: 32-18)-non-synonymous mutation (18) rate  
592 between them was 1.285

593

594 **Supplemental Figure 2**

595 **Different amino acids and synonymous changes of nucleotides in S proteins of SARS-CoV-2 Omicron**  
596 **BA.2 isolates and BA.2-0.1s compared with SARS-CoV-2 Wuhan-Hu-H1.**

597 Nucleotide deletions, "deletion<sup>5</sup>" (deletion: nt 21,633-21,641), introduced the amino acid changes L24- P25-  
598 P26- A27S. Amino acids different from Wuhan-Hu-H1 found in each variant: Alpha: B.1.1.7, Beta: B.1.351,  
599 Gamma: P1, Delta: B.1.617.2.63, Mu: B.1.621, and Omicron: BA.1, BA.2 are highlighted with red, orange,  
600 green, yellow, lime, deep sky blue, and blue-violet, respectively. Amino acid changes common to Omicron:  
601 BA.1 and BA.2 are highlighted with purple.

602

603 **Supplemental Figure 3**

604 **Estimated homologous recombination breakpoints of the SARS-CoV-2 S gene of the Omicron BA.2-0.1**  
605 **or BA.1-BA.2 recombinant.**

606 **(A)** Sequence alignment of the amino acids and coding nucleotides (nt. 22,658-22,702) containing the mutation  
607 point of the SARS-CoV-2 S gene of Omicron BA.2 variants compared with SARS-CoV-2 Wuhan-Hu-H1. **(B)**  
608 Sequence alignment of the amino acids and coding nucleotides (nt. 22,178-22,222) containing the mutation point  
609 of the SARS-CoV-2 S gene of Omicron BA,1, BA.2 variant and BA.1-BA.2 recombinant isolate compared with  
610 SARS-CoV-2 Wuhan-Hu-H1. Changed nucleotides and amino acids of Omicron variants BA.1, BA.2, and

611 BA.1-BA.2 recombinant isolates compared with SARS-CoV-2 Wuhan-Hu-H1 sequences are shown in red  
612 letters. Asterisks show an estimated homologous recombination breakpoint of the SARS-CoV-2 S gene of  
613 Omicron BA.2-0.1.









Fig. 3

21, 750    21, 760    21, 770    21, 780  
 SARS-CoV-2\_Wuhan-Hu-1    GUUACUUGGUUCCAUGCUAUACAUGUCUCUGGGACCA AUGGU  
 SARS-CoV-2\_Omicron\_BA. 1    GUUACUUGGUUCCAUGUUAU-----CUCUGGGACCA AUGGU  
 break point                    \*\*\*  
 V U W F H A I H V S G U N G  
 V U W F H V I - - S G U N G  
                                   A67V H69- V70-

22, 660    22, 670    22, 680    22, 690    22, 700  
 UCUGUCCUAUAUAAUCCGCAUCAUUUCCACUUUUAAGUGUU AU  
 UCUGUCCUAUAUAAUCUCGCACCAUUUUCACUUUUAAGUGUU AU  
                                   \*\*\*\*\*  
 S V L Y N S A S F S T F K C Y  
 S V L Y N L A P F F T F K C Y  
                                   S371L S373P S375F

22, 980    22, 990    23, 000    23, 010  
 AUCUAUCAGGCCGGUAGCACACCUUGUAAUGGUGUU  
 AUCUAUCAGGCCGGUAACAACCUUGUAAUGGUGUU  
                                   \*\*  
 I Y Q A G S T P C N G V  
 I Y Q A G N K P C N G V  
                                   S477N T478K

23, 590    23, 600    23, 610    23, 620  
 UAUCAGACUCAGACUAAUUCUCCUCGGCGGGCACGUAGU  
 UAUCAGACUCAGACUAGUCUCUAUCGGCGGGCACGUAGU  
                                   \*\*\*\*\*  
 Y Q T Q T N S P R R A R S  
 Y Q T Q T K S H R R A R S  
                                   N679K P681H







# Supplemental Figure 1

## A

HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1 ATGTTGTTTACTGTGTCATATGCTTGTCATATTGTGCTGTTGCAACATCAAA  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1 ATGTTGTTTACTGTGTCATATGCTTGTCATATTGTGCTGTTGCAACATCAAA

61 GGAGCAACACTAGTCACCTTGGTCAACGGCTGTGCTGGTCGGAAAGATGTTT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 120  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 120

123 GCTGTGGAAGTGCTGGATATACCTCCCMACTTCAATTAATAGTTCGGTCTCTA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 180  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 180

181 AATAAATCACTCTGTGTCATGGTGTTGTAGAGTTCAGGCTGTTGCTGTTAA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 240  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 240

241 TGCTATGCTGTTGCTGCTGCGCACTTAATGCTGTTGCTTAAATGACTACT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 300  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 300

300 GGAAGAGCTGTTAAAGGTTTTTAACTAACTTCTGCTGATGTCATGCTAAGC  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 360  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 360

361 ATCAATTGGAGTCTGAGCCTGAGCCTGAGCCTTGTAAACTGATTAAGTGCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 420  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 420

420 TGTGTTGTTTTAGCTCACAACAGACATTTTGTTCAGATGTCATGACTCATCTG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 480  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 480

481 GATGTTTATGTCACCAAGAGCAATGTTGCTGGTGGTCACTAATCACTGCTG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 540  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 540

541 GATGTTTGGCGCTATTTTACTTGTGTAATCTACTGCTGGTGGTGGTGGT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 600  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 600

601 TTTTATATAGCCTGACTTCTGTTTTGCTGTTTTGCTGTTTTGCTGTTTTG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 660  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 660

661 GCTACTATCTGCAGCAGCCTCTTCTGACTGTCGCTTACTTACTTACTGCGT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 720  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 720

721 ATTAATGTGTCAGCAACTGCTGCTATAATATATTGCAATTTGTTATAGACA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 780  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 780

781 CTGAGATGTCAGCAGCCTCTTGATGTCAGCAAGTGTGTTTTTCAAGACCCAT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 840  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 840

841 CACCCCTGGAGTACTTGTCTATTTGCTGCTACTGTTATCAACAAACAGCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 900  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 900

900 ATGTTGTTGTTTTAACTTCGAACATAGCCTGGCTGGAGGCTACTAACTGCTG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 960  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 960

961 CTTGTGTTAATAATACCTGCGCAATTTTTAACTAAAGCCGCTGCTGTGT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1020  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1020

1021 GACACATCACTTCACTACCAATTCGTCATGCTCAAGCTTCAAGTGAGTCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1080  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1080

1081 AGTATACAGGGAAATGCCCTTTCTTCTGCGAAATTAATCACTGTTAAATTT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1140  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1140

1141 GCGACTGATGTTTTGGTAAAGAGATACCCGGTGGTGGCAGCTTAAATAGGCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1200  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1200

1201 AATTTGGTTAATTAATCATATATAGGCTCATGATGTTCTGGAGTATGTT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1260  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1260

1261 GATGTAATCTAGCCTCGCAAGCTTGTAGGGTTGAGTCTGTTAACTGAATGATCA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1320  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1320

1321 TGAAATAATCTAAATATACATATTAATGATGCTGCTGGTCTTATGGTAAT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1380  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1380

1381 AGCAAGTCACTTTTATGATAATGATGATCAGCCTCACTCACTGATGTTCTGG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1440  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1440

1441 TTTAAGGATGCTACTGATGACATCTGCTACTGCTGCTACTCCAGCCAGACG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1500  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1500

1501 CTTGTTTATACGACAGCTGTTGGCTGATGCTGTTGCGATTCATAGCTACT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1560  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1560

1561 GCTTTCTTATGTGAGGAATGGTAAATTTATGCTGCTACAGGCACTTAAAT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1620  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1620

1621 TGACAGAGCCTGTTTAACTTATGTTTACTGGTGTGCTGACAGATCTATAAT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1680  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1680

1681 CTTGTCACACAGATTTTTGATGATGATGTTGACTGTTGCACACTATTTG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1740  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1740

## B

HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1741 TCTATACCCCTCAATTGGACCATTCTGTCCAGTTGAGTATTAAGAATCAACATA  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1741 TCTATACCCCTCAATTGGACCATTCTGTCCAGTTGAGTATTAAGAATCAACATA

1742 CTTATACCTGCCATTGGACCACTCTGTCCAGTTGAGTATTAAGAATCAACATA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1800  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1800

1801 CCTATGCTAGTATTGCTGCTCACTTATGTTCAGATGATGATGCTGTGTGAATG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1860  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1860

1861 CTTAGACGATATCTCTGCTTAAACATTAAGACGCCTTAAAGAAATAGCCGATC  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1920  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1920

1921 CTGAGCTGTCAGATGATGATGATGATGCTCTCTTTCAGAGAAAGGCTTTACACTCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 1980  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 1980

1981 AATGTTATGATTTTTGGTACTAACACTTAGACTGTATATCACTTGGCCAGAA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2040  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2040

2041 GGTATGACTGGCTGCTGCTGTGCTGATAGAGACATCTTTTACAAACTTTTAC  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2100  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2100

2101 TCTGACTTGGCTCTGACGCAGCATAAAAGTCTATAAGGCTTCTGCTACTCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2160  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2160

2161 GACTGCTGTCCCAATTATATAGGATGATGTTTCTGGCTGGCCCTGATGT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2220  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2220

2221 GACGTTACGCCATATACAGTCTTATAATGCGAAATTTTAGGGAGCTTACA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2280  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2280

2281 TGACCTGTCAATCTTTTACATAGGATGATGCTGTAAATATTTGGTGTG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2340  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2340

2341 CAGCAGTATTTTAAAGAAATACAGCAATTCTTCTGCTGATCTTTAAAGAAAT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2400  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2400

2401 ACCACATGACTGCTGCTTACGTGGTGTATGATGATGATACACAACTTACAGC  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2460  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2460

2461 CTAACAAGACTTACTGACTGATTAAGATCAGAGTGTGTTAACACAGGGCAG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2520  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2520

2521 TATTGAACATTAACCTTAGTCAGGGCAAAATTTGAGCAGCTTTAGCTCTATT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2580  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2580

2581 CAGCTATATACAGCACTGATATGATTTCAAGCTTTCAAGAGTATGAGCTGATT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2640  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2640

2641 ACTGTTAAGTGGCTGCTGATGATTTGCTCTATACATCTGAATCAACTCAAG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2700  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2700

2701 TTTGCTGTCAGCAGCAGCTCACAAAAAGTGAAGTGTTCAACTCACGCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2760  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2760

2761 AAGGCTATGGCTCTGTGGAATGAGCGACACTTTCTCACTGTAACTGCTCTCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2820  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2820

2821 GAGGGCTGTTTTTTCCATATCTTCTTGTGGCAGCAATATAGGATTTGAGGC  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2880  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2880

2881 TGCTGTGGTTGCTGGTATGATTAAGAATTGTATGTCAGACACATTAATCTGCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 2940  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 2940

2941 CTTACAAAGAGCCATTTATAGGATATGACTTCTGCTAATGTTTCAGCAGCTTT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3000  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3000

3001 CTCTACATAGGATTTTTGCAAGATTAATGATGATTTGCAATCTGATTATGACTT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3060  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3060

3061 CTTGCTGTGAAACTTTTCCGAGATATGCTGCTGCTGATTAATGAGCTCGGAGA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3120  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3120

3121 TTAGTAAATGCACTAATTAACATCTTGACTGATGCTGTAAATGAACTAAGCAG  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3180  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3180

3181 AACTTATTAAGTGAACGGTAATGATGCTTTCAAGTTGAATAGCTGGGAGTCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3240  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3240

3241 TACTATGCTCAAAATGCAAAATGACTGACAGCAATAGACAGCAATGTCGACTA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3300  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3300

3301 AAGTGGCTCAAGCTGTGAGACTATCAAGCGCTTGTGGGTGCTGTGACTT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3360  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3360

3361 TCACTGCTGCTACTTTGGTGGATATGTTGCTATTGATTGTTGCTCTAGCTGT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3420  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3420

3421 TGCCCTTTTAGTGTTTGACTGCTTAAAGGTTGTTGATGCACTAATCCT  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3480  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3480

3481 CTTATACGACTGTGAAAGATCCACATACATA  
HCoV-229E\_Seattle\_USA\_SC3112\_2015\_seq 3516  
HCoV-229E\_Seattle\_USA\_SC0865\_2019\_seq 3516

HCoV\_229E\_SC3112\_2015\_Spike\_seq 1 MFLVLLVAYALHIAAGCQTTNGTNTSHVNCVGVGHSENFVAFSGGYIPSNFNNIFLL 60  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 1 MFLVLLVAYALHIAAGCQTTNGTNTSHVNCVGVGHSENFVAFSGGYIPSNFNNIFLL 60

61 TNTSSVVDGVRFPOLLNCLVWSGSQFTGVFVYNGTRGKACGKYFNSSADVIRYN 120  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 61 TNTSSVVDGVRFPOLLNCLVWSGSQFTGVFVYNGTRGKACGKYFNSSADVIRYN 120  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 61 TNTSSVVDGVRFPOLLNCLVWSGSQFTGVFVYNGTRGKACGKYFNSSADVIRYN 120

121 INFENLRRLGTLFKTSYGAVVFYCNMTNLVSDAHIPSGTVLGNFYCVNMTIGNETS 180  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 121 INFENLRRLGTLFKTSYGAVVFYCNMTNLVSDAHIPSGTVLGNFYCVNMTIGNETS 180  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 121 INFENLRRLGTLFKTSYGAVVFYCNMTNLVSDAHIPSGTVLGNFYCVNMTIGNETS 180

181 AFVGLPKTVREVFISRTGHFYNGYRFLSLGAEVNFVNTAAITVCTVALASYADVL 240  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 181 AFVGLPKTVREVFISRTGHFYNGYRFLSLGAEVNFVNTAAITVCTVALASYADVL 240  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 181 AFVGLPKTVREVFISRTGHFYNGYRFLSLGAEVNFVNTAAITVCTVALASYADVL 240

241 WWSQTAAITNYCNVSNLRCQSLFDWPDGFYSTSPQPVELHVSIVLPYKHFTF 300  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 241 WWSQTAAITNYCNVSNLRCQSLFDWPDGFYSTSPQPVELHVSIVLPYKHFTF 300  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 241 WWSQTAAITNYCNVSNLRCQSLFDWPDGFYSTSPQPVELHVSIVLPYKHFTF 300

301 TIVLHVFHQHQRGPKYCNRPVINTILANFNETKPLCVDSHFHFTQFDNNKLARWSA 360  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 301 TIVLHVFHQHQRGPKYCNRPVINTILANFNETKPLCVDSHFHFTQFDNNKLARWSA 360  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 301 TIVLHVFHQHQRGPKYCNRPVINTILANFNETKPLCVDSHFHFTQFDNNKLARWSA 360

361 SIVTGPCFSGKVNIVFKGVSVCFLKDPGGCAMPIMANLVNKHSHNIGLSVWSRSG 420  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 361 SIVTGPCFSGKVNIVFKGVSVCFLKDPGGCAMPIMANLVNKHSHNIGLSVWSRSG 420  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 361 SIVTGPCFSGKVNIVFKGVSVCFLKDPGGCAMPIMANLVNKHSHNIGLSVWSRSG 420

421 DVTGVPKPKVEGVSFMMVLLNKTKYNI DVSGOVRISNDTLNGLTYTSSONLIG 480  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 421 DVTGVPKPKVEGVSFMMVLLNKTKYNI DVSGOVRISNDTLNGLTYTSSONLIG 480  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 421 DVTGVPKPKVEGVSFMMVLLNKTKYNI DVSGOVRISNDTLNGLTYTSSONLIG 480

481 FKDVTNGTYSITPCNPQDLVYQAVGMALSENFYSYGFVSNVEMPKFFYASNGTYN 540  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 481 FKDVTNGTYSITPCNPQDLVYQAVGMALSENFYSYGFVSNVEMPKFFYASNGTYN 540  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 481 FKDVTNGTYSITPCNPQDLVYQAVGMALSENFYSYGFVSNVEMPKFFYASNGTYN 540

541 CTADVLTYSSFGVCGADSIIZAVQRNYSYVSAIVANLSPNSNNTSVOQEVLQITST 600  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 541 CTADVLTYSSFGVCGADSIIZAVQRNYSYVSAIVANLSPNSNNTSVOQEVLQITST 600  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 541 CTADVLTYSSFGVCGADSIIZAVQRNYSYVSAIVANLSPNSNNTSVOQEVLQITST 600

601 PIVDCTSYVCGNVRCVLLKQYTSACKTEEDALRNSAMLESADVSEM LFDKKAFTLA 660  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 601 PIVDCTSYVCGNVRCVLLKQYTSACKTEEDALRNSAMLESADVSEM LFDKKAFTLA 660  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 601 PIVDCTSYVCGNVRCVLLKQYTSACKTEEDALRNSAMLESADVSEM LFDKKAFTLA 660

661 NVSFDGYNLSVIVPSLRSGRVAGSATEDLFLSKLVTSGLGTVDADYKCKTKGLSIA 720  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 661 NVSFDGYNLSVIVPSLRSGRVAGSATEDLFLSKLVTSGLGTVDADYKCKTKGLSIA 720  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 661 NVSFDGYNLSVIVPSLRSGRVAGSATEDLFLSKLVTSGLGTVDADYKCKTKGLSIA 720

721 DLACAQYINGIMLPGADAEARMMYVTLGGIALLGLTSAISIFPFLSQRLNRYVAL 780  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 721 DLACAQYINGIMLPGADAEARMMYVTLGGIALLGLTSAISIFPFLSQRLNRYVAL 780  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 721 DLACAQYINGIMLPGADAEARMMYVTLGGIALLGLTSAISIFPFLSQRLNRYVAL 780

781 QTDVQENRQQLLAASFNKMTNVDAFVGDNDATQSTQALQVATALNKIQDVNQGN 840  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 781 QTDVQENRQQLLAASFNKMTNVDAFVGDNDATQSTQALQVATALNKIQDVNQGN 840  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 781 QTDVQENRQQLLAASFNKMTNVDAFVGDNDATQSTQALQVATALNKIQDVNQGN 840

841 SLNHLTSQLRQNFQAISSSIQAIDRLDIQADQVDRILTRLAALNWFVSHLTLYKTE 900  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 841 SLNHLTSQLRQNFQAISSSIQAIDRLDIQADQVDRILTRLAALNWFVSHLTLYKTE 900  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 841 SLNHLTSQLRQNFQAISSSIQAIDRLDIQADQVDRILTRLAALNWFVSHLTLYKTE 900

901 VRASRLAQQVKEVKSQSKRYGFCNGTHIFLSLNAEPGLVFLHTLPTQYKQVEA 960  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 901 VRASRLAQQVKEVKSQSKRYGFCNGTHIFLSLNAEPGLVFLHTLPTQYKQVEA 960  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 901 VRASRLAQQVKEVKSQSKRYGFCNGTHIFLSLNAEPGLVFLHTLPTQYKQVEA 960

961 WSLGCVDIGINGVLRQPLNLKYEQNYRITSRIEMFRRIPRIADPVEIENCVTFVNI 1020  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 961 WSLGCVDIGINGVLRQPLNLKYEQNYRITSRIEMFRRIPRIADPVEIENCVTFVNI 1020  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 961 WSLGCVDIGINGVLRQPLNLKYEQNYRITSRIEMFRRIPRIADPVEIENCVTFVNI 1020

1021 RSELQTLVPEIDWIKTLQELSKYPNVTVPDLVWQYVLLNLESTSEIEMKSAELN 1080  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 1021 RSELQTLVPEIDWIKTLQELSKYPNVTVPDLVWQYVLLNLESTSEIEMKSAELN 1080  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 1021 RSELQTLVPEIDWIKTLQELSKYPNVTVPDLVWQYVLLNLESTSEIEMKSAELN 1080

1081 YTVQKQLTLDNINSTVLKWNKRVVQYLWVQWQVLSLNLVLLCCSSTGCG 1140  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 1081 YTVQKQLTLDNINSTVLKWNKRVVQYLWVQWQVLSLNLVLLCCSSTGCG 1140  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 1081 YTVQKQLTLDNINSTVLKWNKRVVQYLWVQWQVLSLNLVLLCCSSTGCG 1140

1141 GFFCSFASSIIGCCESLTKPYVDKRIHQ 1171  
HCoV\_229E\_SC3112\_2015\_Spike\_seq 1141 GFFCSFASSIIGCCESLTKPYVDKRIHQ 1171  
HCoV\_229E\_SC0865\_2019\_Spike\_seq 1141 GFFCSFASSIIGCCESLTKPYVDKRIHQ 1171





