

Research Article

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Synopsis of tailed Myxobolidae (Cnidaria, Myxozoa, Myxosporea) infecting Indian fishes

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Abstract: A synopsis of 43 nominal species from five genera of tailed Myxobolidae infecting Indian freshwater and marine fishes is presented. The main characteristic of this group is the presence of at least one tail-like caudal process. For each species, relevant morphological and morphometric data are provided, such as the host(s), site(s) of infection within the host and sampling state. A key for the identification of 13 genera of tailed Myxobolidae is also included.

Keywords: Fish parasites, species diversity, taxonomy, identification key

Species of the family Myxobolidae Thélohan, 1892 of the phylum Cnidaria, subphylum Myxozoa, are characterised by spores usually flattened parallel to the straight sutural plane, bilaterally symmetrical body with two or sometimes one polar capsule. This synopsis accounts the genera of tailed Myxobolidae from fishes of the Indian subcontinent that have at least one caudal process or caudal appendages, which look like ‘tails’ (Thélohan 1892).

Till now nine genera, i.e., *Henneguya* Thélohan, 1892, *Hennegoides* Lom, Tonguthai et Dyková, 1991, *Neohenneguya* Tripathi, 1953, *Unicauda* Davis, 1944, *Dicauda* Hoffman et Walker, 1978, *Laterocaudata* Chen et Hsieh, 1984, *Phlogospora* Qadri, 1962, *Tetrauronema* Wu, Wang et Jiang, 1988, and *Trigonosporus* Hoshina, 1952, have been reported as tailed Myxobolidae all over the world.

In India, species of only five genera, namely *Henneguya*, *Neohenneguya*, *Hennegoides*, *Unicauda* and *Phlogospora*, have been reported. In this paper, we provide for the first time an updated synopsis of tailed Myxobolidae in Indian fishes. Most of the research on this group has been carried out mainly in four states (West Bengal, Andhra Pradesh, Punjab and Orissa) on both freshwater and marine fishes.

A number of authors (Chakravarty 1939, Ganapati 1941, Tripathi 1952, Qadri 1962, 1965, Bhatt and Siddiqui 1964, Lalitha Kumari 1965, 1969, Chaudhuri and Chakravarty 1970, Narasimhamurti and Kalavati 1975, Kalavati and Narasimhamurti 1981, 1984, Seenappa et al. 1981, Bajpai and Halder 1982a,b, Halder et al. 1983, 1997, Halder and

Mukherjee 1985, Sarkar 1985, Sarkar et al. 1985, Menon 1986, Gupta and Khera 1987, Wu et al. 1988, Kalavati et al. 1992, Susha and Janardhan 1995, Acharya et al. 2004, Gupta and Kaur 2018, Nissa and Kaur 2021) described numerous new species of tailed Myxobolidae from freshwater and marine fishes of West Bengal, Andhra Pradesh, Orissa, Kerala and Uttar Pradesh.

Hemanand et al. (2008), and Kaur and Attri (2015) recently described one species from the freshwater fish of Manipur and Punjab, respectively. Kalavati and Nandi (2007) listed 34 species of tailed Myxobolidae.

The present synopsis updates the number of species infecting Indian fishes in a tabulated arrangement, giving details of morphology and dimensions of cyst, spore body and polar capsules. Site(s) of infection within the host, type host and sampling state are also given.

MATERIALS AND METHODS

The collection of data about myxobolid parasites in Indian fishes was based on a detailed internet search to detect references to fish infection by tailed Myxobolidae. A careful check of the references of each paper was done to maximise the probability of getting all the data available. Fish names in Table 1 follow Froese and Pauly (2022).

Key to tailed genera of Myxobolidae

1a. Spores ovoid or pyriform, with two polar capsules and a single tail-like caudal process 2

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Table 1. Synopsis of *Henneguya*, *Hennegoides*, *Neohenneguya*, *Phlogospora* and *Unicauda* spp. infecting Indian fishes. *Abbreviations:* C – cyst; SB – spore shape; D – diameter; SBL – spore body length; STL – total length of spore; WS – width of spore; TS – spore thickness; CA – caudal appendage length; PC – polar capsule; LPC – length of large polar capsule; SPC – length of small polar capsule; NC – number of coils of polar filament inside polar capsule. AP – Andhra Pradesh; K – Kerala; KR – Karnataka; M – Manipur; O – Orissa; P – Punjab; TN – Tamilnadu; WB – West Bengal; UP – Uttar Pradesh. All the measurements are in μm ; dimensions of polar capsules are given as length \times width.

| Species | Infected organ | Spores | Type-host | State |
|--|----------------------|--|---|-------|
| <i>Henneguya basari</i> Gupta et Khera, 1987 | Pharyngeal cavity | C oval, white, 500–1000 \times 300; SB oval, STL 25.6–39.2 (34.3); WS 3.2–4.4 (3.7); CA 10.0–20.3 (15.3); PC equal and pyriform, 5.0–7.4 (6.3) \times 1.4–2.0 (1.7). | <i>Channa punctata</i> Bloch | UP |
| <i>H. bengalensis</i> Gupta et Khera, 1987 | Buccal cavity | C oval, white, 1000–1250; SB elongate, STL 26.4–29.5 (28.4); WS 2.8–3.9 (3.3); CA 11.6–13.8 (12.61); PC equal and pyriform, 3.3–3.6 (3.5) \times 1.1–1.4 (1.2). | <i>Channa punctata</i> Bloch | WB |
| <i>H. bicaudi</i> Kaur et Attri, 2015 | Gill filament | C round, 500–700 (D); SB oblongate, SBL 20–22 (21); WS 5.9–6.5 (6.2), CA 8.7–9.0 (8.9); PC equal and pyriform, 3.1 \times 2.0; NC 7–8. | <i>Cirrhinus mrigala</i> Hamilton | P |
| <i>H. bicornuata</i> Gupta et Khera, 1987 | Branchial epithelium | C oval, milky white, 1200–1500; SB oblongate, STL 26.4–29.2; WS 2.8–3.9; CA 11.6–13.8; PC equal and pyriform, 3.3–3.6 \times 1.1–1.4. | <i>Channa punctata</i> Bloch | WB |
| <i>H. bleekeri</i> Halder et Mukherjee, 1985 | Kidney | C round, 1500 \times 1000; SB oblongate, STL 20–25.5 (21.9); WS 3.3–5.0 (4.3); CA 10.1–15.5 (13.0); PC equal and pyriform, 3.3–4.4 (3.6) \times 1.1–2.2 (1.7); NC 6–8. | <i>Mystus bleekeri</i> Day | WB |
| <i>H. chaudhuryi</i> Gupta et Khera, 1987 | Gill filaments | SB oblongate, STL 26.3–33.2 (30.0); WS 3.3–4.1 (3.7); CA 14.5–20.0 (17.7); PC equal and pyriform, 5.0–7.5 (6.0) \times 1.6. | <i>Channa punctata</i> Bloch | WB |
| <i>H. ganapatiae</i> Qadri, 1970 | Gills | SB oval, SBL 9.3–10.0 (9.7); WS 4.0–4.8 (3.7); CA 22–25 (22.3); PC equal and elongate, 3.2–2.6 (3.32) \times 1.4–1.8 (1.6). | <i>Notopterus notopterus</i> Pallas | AP |
| <i>H. latesi</i> Tripathi, 1952 | Gills | C small; SB pyriform, SBL 9.0–10.8; WS 4.0–4.8 (4.5); CA 17.2–25.0; PC equal and pyriform, slightly convergent, 3.6 \times 2.0. | <i>Lates calcarifer</i> Bloch | WB |
| <i>H. latiusi</i> Gupta et Kaur, 2018 | Gills | C spherical, 200–900 (D); S oblongate, 12 \times 4.3; CA 12–14 long; PC equal and pyriform, 4.0 \times 2.5. | <i>Crossocheilus latiusi</i> Hamilton | P |
| <i>H. manipurensis</i> Hemanand, Meiti, Bandyopadhyay et Mitra, 2008 | Body tissue | SB elongated; STL 23.8–28.0 (25); SBL 12.6–15.4 (14.0); WS 5.6–7.0 (6.3); CA 11.2–12.6 (11.7); PC equal and pyriform, 5.6–6.3 (5.8) \times 2.1–2.8 (2.3). | <i>Anabas testudineus</i> Bloch | M |
| <i>H. megalopsi</i> Kalavati, Venkateswara Rao et Vaidehi, 1992 | Gall bladder | SB oval; STL 36.8–40.0 (37.6); SBL 11.2–12.8 (12.4); WS 3.2–3.6 (3.24); TS 2.5–3.0; CA 25.6–37.2 (34.2); PC equal and pyriform, 4.4 \times 0.8. | <i>Megalops cyprinoides</i> Broussonet | O |
| <i>H. mystasi</i> Halder, Samal et Mukhopadhyay, 1997 | Gills | SB pyriform; STL 35.8–53.8 (46.3); SBL 13.0–18.7 (16.14); WS 1.6–4.0 (2.92); CA 19.5–37.4 (29.85); PC equal and pyriform, 4.0–4.9 (4.36) \times 1.6–3.2 (1.92). | <i>Mystus gulio</i> Hamilton | O |
| <i>H. mystusia</i> Sarkar, 1985 | Gill filament | C small, creamy white, 200 \times 100; SB fusiform; STL 27.0–40.0 (32.33); SBL 12.0–15.0 (13); WS 3.0–4.0 (3.8); TS 2.5–3.0 (2.9); CA 17.0–25.0 (19.3); PC equal and tubular, 5.0–6.0 (5.04) \times 1.0–1.3 (1.2). | <i>Mystus</i> sp. | WB |
| <i>H. namae</i> Halder, Das et Sharma, 1983 | Gills and heart | C oval, creamy white, 180–220 \times 150–190; SB oblongate, SBL 17.6–19.3 (18.5); WS 5.5–6.6 (6.4); CA 17.6–18.7 (18.5); PC unequal and pyriform, LPC 5.5–6.0 (5.8) \times 1.1; SPC 4.4–5.0 (4.9) \times 1.1; NC 9–10. | <i>Chanda nama</i> Hamilton | WB |
| <i>H. nandi</i> Gupta et Khera, 1987 | Gills and heart | C round 399–450 \times 342–402; SB oblongate; STL 18–25 (20.2); SBL 9.0–11.0 (9.9); WS 3.0–5.0 (3.9); CA 9.0–15.0 (10.3); PC equal and pyriform, 3.2–4.0 (3.6) \times 1.4–2.0 (1.6). | <i>Nandus nandus</i> Hamilton | P |
| <i>H. notopteriae</i> Qadri, 1965 | Gills | SB ellipsoidal or lanceolate; SBL 11.5–13.0; WS 4.5–5.0 (4.8); CA 40.5–42.0; PC unequal, LPC golf stick like, 5.0–5.5 \times 1.0–1.5; SPC pyriform, 3.5–4.5 \times 1.0–1.3. | <i>Notopterus notopterus</i> Pallas | AP |
| <i>H. ophiocephali</i> Chakravarty, 1939 | Gills and muscles | C spherical, 2000 (D); SB oval; STL 41.5–52.5 (37.6); WS 6.2–7.2 (3.2); CA 26.0–32.0; PC unequal and pyriform, LPC 6.2–9.3 \times 2.1–3.0; SPC 5.2–8.2 \times 2.1–3.0. | <i>Channa gachua</i> Hamilton | WB |
| <i>H. otolithi</i> Ganapati, 1941 | Bulbus arteriosus | SB oval; SBL 10.0–12.0 (16.1); WS 6.0–8.5; CA 35.0–40.0; PC equal and pyriform, 3.0–4.0 \times 2.0–2.5; NC 5–6. | <i>Otolithes ruber</i> Bloch et Schneider | TN |
| <i>H. qadrii</i> Lalitha Kumari, 1965 | Intestine | SB oval; SBL 9.2–12.3 (11.0); WS 4.6–5.4; CA 6.2–14.3 (9.8); PC unequal & pyriform, LPC 4.6–6.2 (5.3) \times 1.2–1.8 (1.5); SPC 3.9–4.6 (4.5) \times 1.2–1.9 (1.5). | <i>Channa gachua</i> Hamilton | AP |
| <i>H. renalis</i> Sarkar, Mazumder et Pramanik, 1985 | Kidney | SB broadly oval, SBL 14.4–19.2 (16.8); WS 8.0–11.2 (9.8); CA 4.8–12.8 (7.9); PC equal and pyriform, 4.6–7.6 (5.74) \times 2.5–5.1 (4.26). | <i>Channa marulius</i> Hamilton | WB |
| <i>H. ritae</i> Gupta et Khera, 1987 | Branchial filament | SB oblongate, STL 39.9–44.8 (42.4); WS 5.0–5.8 (5.2); CA 27.2–31.5 (29.4); PC equal and pyriform, 5.0–5.8 (5.3) \times 1.7–2.5 (1.9); NC 6–7. | <i>Rita rita</i> Hamilton | WB |
| <i>H. rubicundi</i> Halder et Mukherjee, 1985 | | C creamy yellowish, 2000–1500 (D); SB elongate oval, STL 24.4–36.7 (31.2); WS 3.3–8.1 (5.5); CA 13.3–24.5 (19.7); PC equal and pyriform, 3.3–5.1 (3.7) \times 2.2–3.0 (2.3); NC 6–8. | <i>Odontambliopus rubicundus</i> Hamilton | WB |
| <i>H. singhi</i> Lalitha Kumari, 1969 | Gill filaments | SB elongate; SBL 11.1–13 (12.3); WS 3.9–5.7 (4.4); CA 30–48 (39); PC unequal and pyriform, LPC 4.8–6.4 (5.7) \times 0.8–1.4 (1.1); SPC 3.5–5.5 (4.1) \times 1.4–2.0 (1.6); NC 4–5. | <i>Notopterus notopterus</i> Pallas | AP |
| <i>H. tachysuri</i> Menon, 1986 | Branchial epithelium | C round or oval, white, 1000–3000 (D); SB ovoid, STL 47–60; SBL 12.0–15.0; WS 7.0–8.0; TS 5.0–6.0; CA 35.0–44.0; PC equal and pyriform, 6.0–7.0 \times 2.0–3.0; NC 3–5. | <i>Channa punctata</i> Bloch | WB |

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|---|-----------------------------|--|---|-------|
| <i>H. thermalis</i> Seenappa, Manohar et Prabhu, 1981 | Brain tissue | SB pyriform; STL 47.0–60.0; SBL 12.0–13.2; WS 6.0–8.0; TS 5.0; CA 11.0–13.0; PC unequal; LPC pyriform, 4.0–5.0 × 2.0–3.0; SPC ovoid, 3.0–4.0 × 2.0. | <i>Lepidocephalichthys thermalis</i> Valenciennes | K |
| <i>H. waltirensis</i> Narasimhamurti et Kalavati, 1975 | Gill filaments | SB oval; STL 55.0; SBL 14.6–15.5; WS 3.2–4.0; CA 40.0–50.0; PC equal and pyriform, 10.0–12.0 × 1.6–2.5, NC 6–7. | <i>Channa punctata</i> Bloch | AP, K |
| <i>H. zahoori</i> Bhatt et Siddiqui, 1964 | Gill filaments | C spherical, whitish, 500 (D); SB biconvex; STL 20.0–30.6 (23.6); SBL 8.0–12.0 (9.6); WS 2.1–3.0 (2.6); CA 12.0–18.6 (13.9); PC equal and pyriform, 4.9–6.7 (5.8) × 0.7–1.1 (0.9). | <i>Channa punctata</i> Bloch | WB |
| <i>Hennegoides senghalae</i> Nissa et Kaur, 2021 | Gills | C pale, thread like, 3000–5000; SB inverted pyriform; STL 7.5; WS 3.0–4.5 (3.8); CA (27.0–47.3); PC equal, 1.5 × 0.75, NC 3–4. | <i>Sperata seenghala</i> Sykes | P |
| <i>Neohenneguya tetraradiata</i> Tripathi, 1952 | Gills | C irregular, 500; SB spindle-shaped 16.2–21.6 × 5.4; CA two long, equal at both ends 63.0–72.0; PC spherical, 2.0–2.7. | <i>Odontamblyopus rubicundus</i> Hamilton | WB |
| <i>Phlogospora gulio</i> Acharya, 2004 | Gill lamellae | S pyriform; STL 40.2–56.5 (53.5); SBL 16.2–20.5 (19.8); WS 4.1–6.2 (5.9); CA 24.0–36.0; PC 9.1–12.3 (11.8) × 3.04.0 (3.5); NC 10–14. | <i>Mystus gulio</i> Hamilto | WB |
| <i>P. mysti</i> Qadri, 1962 | Gills | S Bunsen flame like; STL 35.0–42.0; SBL 14.0–18.0; | <i>Mystus bleekeri</i> Day | AP |
| <i>P. oculatus</i> Susha et Janardanan, 1995 | Gills | S pear-shaped; STL 45.0–90.75 (65.7); SBL 13.5–24.0 (18.5); WS 2.7–3.5 (3.1); CA 30.8–68.3; PC single, ovoid, 7.5–13.2 (9.6) × 1.5–3.1 (2.4); NC 10–13. | <i>Mystus oculatus</i> Valenciennes | K |
| <i>Unicauda andharae</i> Kalavati et Narasimhamurti, 1981 | Gills | SB oval, STL 38.4–47.3 (42.5); WS 4.0–5.4 (4.9); CA 20.8–24.5 (22.8); PC equal and pyriform, 5.0–6.4 (5.9) × 1.0–1.6 (1.2); NC 4–5. | <i>Channa gachua</i> Hamilton | AP |
| <i>U. aplocheili</i> Kalavati, Venkateswara, Rao et Vaidehi, 1992 | Scales | SB pyriform, STL 16.8–18.2 (16.9); SBL 9.8–11.2 (10.3); WS 5.6–7.0 (5.7); CA 5.6–8.4 (7.5); PC equal and elongated, 5.0–6.4 (5.9) × 1.2–1.6 (1.4); NC 4–5. | <i>Aplocheilus panchar</i> Hamilton | O |
| <i>U. armati</i> Gupta et Khera, 1987 | Gills | C yellow, round, 510 × 391; SB oval, SBL 7.5–9.0 (8.4); WS 5.6–7.0 (5.7); CA 8.0–15.0 (11.5); PC equal and pyriform, 2.0–3.0 (2.7) × 1.5–2.0 (1.6). | <i>Mastacembalus armatus</i> Lacépède | P |
| <i>U. basiri</i> Bhatt et Siddiqui, 1964 | | SB ovoid, STL 25.6–39.2 (34.3); WS 3.2–4.4 (3.7) CA 10.0–20.3 (15.3); PC equal, 5.0–7.4 (6.3) × 1.4–2.0 (1.7). | <i>Channa punctata</i> Bloch | WB |
| <i>U. bengalensis</i> Chaudhuri et Chakravarty, 1970 | Upper buccal cavity | SB oblongate, STL 35.7–45.7 (39.4); WS 2.9–4.3 (4.0) CA 14.3–27.2 (21.5); PC equal, 6.0–7.2 (6.5) × 1.4–1.8 (1.5). | <i>Channa punctata</i> Bloch | WB |
| <i>U. bicornuata</i> Chaudhuri et Chakravarty, 1970 | Branchial epithelium, Gills | SB oblongate, STL 26.4–29.2 (28.4); WS 2.8–3.9 (3.3); CA 11.5–13.8 (12.6); PC equal and pyriform, 3.3–3.6 (3.5) × 1.1–1.4 (1.2). | <i>Channa punctata</i> Bloch | AP |
| <i>U. chaudhuryi</i> Bajpai et Haldar, 1982 | Gill filament | SB oblongate, STL 26.6–33.2 (30.0); WS 3.3–4.1 (3.7); CA 14.5–20.0 (17.7); PC equal and pyriform, 5.0–7.5 (6.0) × 1.6. | <i>Channa punctata</i> Bloch | WB |
| <i>U. irregularis</i> Haldar, Samal et Mukhopadhyay, 1997 | Body muscles | SB pyriform, STL 24.0–39.1 (28.0); SBL 9.7–19.5 (13.3), WS 4.9–6.5 (5.1); CA 9.8–22.8 (14.8); PC equal and pyriform, 4.0–6.5 (5.0) × 1.6–4.0 (2.3). | <i>Mystus vittatus</i> Bloch | O |
| <i>U. minuta</i> Haldar, Samal et Mukhopadhyay, 1997 | Gills | SB broadly elongated, STL 16.3–26.0 (20.9); SBL 8.1–13.0 (9.4); WS 3.2–5.7 (4.1); CA 6.5–16.3 (11.4); PC equal and elongated, 1.6–4.9 (3.9) × 1.6–3.5 (2.2). | <i>Etroplus suratensis</i> Bloch | O |
| <i>U. ritae</i> Bajpai et Haldar, 1982 | Gill filaments | SB oblongate, STL 39.9–44.8 (42.4); WS 5.0–5.8 (5.2); CA 27.2–31.5 (29.4); PC equal and pyriform, 5.0–5.8 (5.3) × 1.7–2.5 (1.9). | <i>Rita rita</i> Hamilton | WB |
| <i>U. theraponi</i> Haldar, Samal et Mukhopadhyay, 1997 | Gills | SB pyriform, STL 27.7–39.1 (33.3); SBL 14.6–22.8 (19.8) WS 4.0–8.1 (5.5); CA 8.1–19.5 (12.9); PC equal and pyriform, 4.9–8.1 (6.2) × 1.6–3.2 (2.5). | <i>Therapon jarbua</i> Forsskål | O |

- 1b. Spores with two or more tail-like caudal processes or bifurcated tail; composed of same materials as shell valve 3
- 2a. Single tail-like caudal process; caudal process symmetrically located at the posterior end *Unicauda* Davis, 1944
- 2b. Single tail-like caudal process, which may be forked at the very end, originating posterolaterally and asymmetrically *Laterocaudata* Chen et Hsieh, 1984
- 3a. Spores tear-shaped shaped, with a single polar capsule, caudal process bifurcated *Phlogospora* Qadri, 1962
- 3b. Spores with two polar capsules and 2 caudal processes ... 4
- 3c. Spores with two polar capsules and 4 caudal processes ... 5
- 4a. Spores with two caudal processes *Henneguya* Thélohan, 1892
- 4b. Spores with two caudal processes, asymmetric *Hennegoides* Lom, Tonguthai et Dyková, 1991
- 4c. Spores with two caudal processes, extending in opposite directions *Dicauda* Hoffman et Walker, 1978
- 5a. Spores ellipsoidal with two caudal processes at each end of the spore; polar capsules situated asymmetrically *Neohenneguya* Tripathi, 1953

- 5b. Spores with four posterolateral caudal processes, two from each shell valve *Tetrauronema* Wu, Wang et Jiang, 1988
- 5c. Spores rhomboidal with four caudal processes pointing in the opposite direction, connected by transverse filaments *Trigonosporus* Hoshina, 1952

DISCUSSION

This study provides a synopsis on tailed Myxobolidae infecting freshwater and marine fishes of the Indian sub-continent based on available data along with their host, state records and brief spore description. In total 43 species in five genera are reported. It is noteworthy than more than half of the species (27) belongs to the genus *Henneguya*. This is not surprising because this genus is the second most-species rich of myxozoans and its species infect fish worldwide (Eiras 2002, Eiras and Adriano 2012). The present checklist may help in future studies for taxonomic identification and comparison with their related species of tailed Myxobolidae.

Concerning the pathogenicity, three species of *Henneguya*, *H. mystusia* Sarkar, 1985, *H. otolithi* Ganapati, 1941 and *H. waltirensis* Narasimhamurti et Kalavati, 1945, and one species of *Unicauda*, *U. aplocheili* Kalavati, Venkateswara Rao et Vaidehi, 1992, were found to be highly pathogenic. Ulcerative body tissue in *Anabas testudineus* Bloch affected by *H. manipurensis* Hemanand, Meiti et Bandyopadhyay, 2008 was also observed. In *Mystus* sp. functional disruption of gill lamellae was found, including dilatation of capillaries of primary gill lamellae, and atrophy of secondary gill lamellae caused by *H. mystusia* Sarkar, 1985. *Henneguya otolithi* Ganapati, 1941 infecting the heart of *Otolithus ruber* (Bloch et Schneider) causes damage in the heart, destruction of muscle fibres and is associated with fibroblast infiltration. *Henneguya waltirensis* induces hypertrophy of the gill epithelium and vacuolisation of the cell cytoplasm in *Channa punctata* (Bloch).

REFERENCES

- ACHARYA S., DAS M.K., HALDAR D.P., DUTTA T. 2004: On a new myxosporean parasite, *Phlogospora gulio* sp. n. from the cat fish, *Mystus gulio* (Hamilton-Buchanan) in West Bengal. *Geobios* 31: 261–264.
- BAJPAI R.R., HALDAR D.P. 1982a: A new myxosporidian, *Unicauda chaudhuryi* n. sp. (Myxozoa: Myxosporidia) from the fish *Ophiocephalus punctatus* Bloch. *Riv. Parassitol.* 43: 142–152.
- BAJPAI R.R., HALDAR D.P. 1982b: Observations on a new species of *Unicauda* Davis, 1944 (Myxozoa: Myxosporidia) from the teleost *Rita rita* Hamilton. *Arch. Protistenkd.* 125: 79–86.
- BHATT V.S., SIDDIQUI W.A. 1964: Four new species of Myxosporidia from the Indian fresh water fish, *Ophiocephalus punctatus* Bloch. *J. Protozool.* 11: 314–316.
- CHAKRAVARTY M.M. 1939: Studies on Myxosporidia from the fishes of Bengal, with a note on myxosporidian infections in aquaria fishes. *Arch. Protistenkd.* 92: 169–178.
- CHAUDHURI S.R., CHAKRAVARTY M.M. 1970: Studies on Myxosporidia (Protozoa: Sporozoa) from the food fishes of Bengal. I. Three new species from *Ophiocephalus punctatus* Bloch. *Acta Protozool.* 8: 167–173.
- EIRAS J.C. 2002: Synopsis of the species of the genus *Henneguya* Thélohan, 1892 (Myxozoa: Myxosporidia: Myxobolidae). *Syst. Parasitol.* 52: 43–54.
- EIRAS J.C., ADRIANO E.A. 2012: A checklist of new species of *Henneguya* Thélohan, 1892 (Myxozoa, Myxosporidia, Myxobolidae) described between 2002 and 2012. *Syst. Parasitol.* 83: 95–104.
- FROESE R., PAULY D. (EDS.) 2022: FishBase. World Wide Web electronic publication. www.fishbase.org, version (08/2022).
- GANAPATI P.N. 1941: On a new myxosporidian *Henneguya otolithi* n. sp., a tissue parasite from the bulbus arteriosus of two species of fish of the genus *Otolithus*. *Proc. Ind. Acad. Sci., Section B* 13: 135–150.
- GUPTA A., KAUR H. 2018: 18S and 28S rDNA identity and phylogeny of two novel myxosporeans infecting gills of cyprinid carps inhabiting a coldwater wetland in northern India. *Microb. Path.* 120: 97–108.
- GUPTA S., KHERA S. 1987: On the genera *Henneguya* Thélohan, 1892 and *Unicauda*, Davis, 1944. *Res. Bull. Punjab Univ.* 38: 153–163.
- HALDAR D.P., DAS M.K., SHARMA B.K. 1983: Studies on protozoan parasites from fishes. Four new species of the genera *Henneguya* Thélohan, 1882, *Thelohanellus* Kudo, 1933, and *Myxobolus* Bütschli, 1892. *Arch. Protistenkd.* 127: 283–296.
- HALDAR D.P., MUKHERJEE M. 1985: Studies on Bivalvulida (Myxozoa: Myxosporidia) observations on two new species of *Henneguya* from food fishes of West Bengal, India. *Arch. Protistenkd.* 130: 419–425.
- HALDAR D.P., SAMAL K.K., MUKHOPADHYAY D. 1997: Studies on the protozoan parasites of fishes in Orissa: five new species of the genera *Henneguya*, *Thelohanellus* and *Unicauda* (Myxozoa: Bivalvulida). *J. Bengal. Nat. Hist. Soc.* 16: 50–63.
- HEMANAND TH., MEITEI N.M., BANDYOPADHYAY P.K., MITRA A.K. 2008: A new species of *Henneguya*, a gill parasite of a freshwater fish *Anabas testudineus* (Bloch) affected with ulcerative disease syndrome from Manipur, India. *Türkiye Parazit. Derg.* 32: 82–85.
- KALAVATI C., NARASIMHAMURTI C.C. 1981: *Unicauda andhrae* n. sp. (Myxosporidia) in the mucus on the gills of *Ophiocephalus gachua*. *Z. Parasitenkd.* 65: 89–93.
- KALAVATI C., NARASIMHAMURTI C.C. 1984: Histopathological changes in the gills of *Channa punctatus* Bl. infected with *Henneguya waltirensis*. *Arch. Protistenkd.* 129: 199–202.
- KALAVATI C., VENKATESWARA RAO J., VAIDEHI J. 1992: Myxosporidian parasites (Protozoa) of fishes of Chilka Lake, east coast of India: three new species of *Henneguya* Thélohan, *Rudicapsula* Kalavati and Narasimhamurti and *Unicauda* Auerbach. *Ind. J. Parasitol.* 16: 77–83.
- KALAVATI C., NANDI N.C. 2007: Handbook on Myxosporean Parasites of Indian Fishes. Zoological Survey of India, Kolkata, New Delhi, 294 pp.
- KAUR H., ATTRI R. 2015: Morphological and molecular characterization of *Henneguya bicaudi* n. sp. (Myxosporidia: Myxobolidae) infecting gills of *Cirrhinus mrigala* (Ham.) in Harike Wetland, Punjab (India). *Parasitol. Res.* 114: 4161–4167.
- LALITHA KUMARI P.S. 1965: On a new species of *Henneguya* (Protozoa: Myxosporidia) from an Indian fresh water fish, *Ophiocephalus gachua*. *Riv. Parassitol.* 26: 79–84.
- LALITHA KUMARI P.S. 1969: Studies on parasitic Protozoa (Myxosporidia) of fresh water fishes of Andhra Pradesh, India. *Riv. Parassitol.* 30: 153–226.
- MENON N.G. 1986: On a new myxosporidian *Henneguya tachysuri* sp. nov., from the marine cat fish *Tachysurus thalassinus* (Rüppell) from the Gulf of Mannar. *J. Marine Biol. Ass. India* 21: 196–199.
- NARASIMHAMURTI C.C., KALAVATI C. 1975: A new myxosporidian parasite, *Henneguya waltirensis* n. sp., from the gills of *Ophiocephalus punctatus* Bl. *Riv. Parassitol.* 36: 255–259.

- NISSA K., KAUR H. 2021: First record of the genus *Hennegoides* Lom, Tonguthai and Dyková, 1991 from Punjab (India) infecting the catfish, *Sperata seenghala* (Sykes, 1839). *Int. J. Parasitol. Parasites Wildl.* 14: 7–12.
- QADRI S.S. 1962: On a new myxosporidian parasite, *Phlogospora mysti*, gen. n., sp. n., from Indian fresh water fish, *Mystus bleekeri*. *Arch. Protistenkd.* 106: 211–217.
- QADRI S.S. 1965: Study on a new myxosporidian parasite from the fresh water fish *Notopterus notopterus*. *Zool. Anz.* 175: 225–228.
- SARKAR N.K. 1985: Myxosporidan *Henneguya mystusii* sp. n. (Myxozoa: Myxosporidia) from the gill of a fresh water teleost fish *Mystus* sp. *Acta Protozool.* 24: 55–58.
- SARKAR N.K., MAZUMDER S.K., PRAMANIK A. 1985: Observations on 4 new species of Myxosporidia (Myxozoa) from channid (ophiocephalid) fishes of West Bengal, India. *Arch. Protistenkd.* 130: 289–296.
- SEENAPPA D., MANOHAR L., PRABHU R.N. 1981: *Henneguya thermalis* n. sp., parasitic in the brain tissue of the loach, *Lepidocephalichthys thermalis* (Hamilton). *Curr. Sci.* 50: 295–296.
- SUSHA T.K., JANARDHAN K.P. 1995: On a new myxosporidian parasite, *Phlogospora oculatus* n. sp., from *Mystus oculatus* (Hamilton) in Kerala. *Proc. Zoo. Soc. Cal.* 48: 11–14.
- THÉLOHAN P. 1892: Observations about the Myxosporidia and essay of classification of these organisms. *Bull. Soc. Phil. Paris* 4: 165–178. (In French).
- TRIPATHI Y.R. 1952: Studies on parasites of Indian fishes I. Protozoa: Myxosporidia together with a check list of parasitic protozoa described from Indian fishes. *Rec. Ind. Mus.* 50: 63–88.
- WU B.H., WANG S.X., JIANG N.C. 1988: [A new species *Tetrauronea macropodus* sp. nov. (gen. et fam. nov.) from freshwater fishes of China]. *Acta Zootaxon. Sin.* 13: 315–316. (In Chinese.)

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