

BIBLIOGRAPHIC INFORMATION SYSTEM

Journal Full Title: Journal of Biomedical Research & Environmental Sciences

Journal NLM Abbreviation: J Biomed Res Environ Sci

Journal Website Link: <https://www.jelsciences.com>

Journal ISSN: 2766-2276

Category: Multidisciplinary

Subject Areas: Medicine Group, Biology Group, General, Environmental Sciences

Topics Summation: 128

Issue Regularity: Monthly

Review Process type: Double Blind

Time to Publication: 7-14 Days

Indexing catalog: [Visit here](#)

Publication fee catalog: [Visit here](#)

DOI: 10.37871 ([CrossRef](#))

Plagiarism detection software: iThenticate

Managing entity: USA

Language: English

Research work collecting capability: Worldwide


Organized by: [SciRes Literature LLC](#)

License: Open Access by Journal of Biomedical Research & Environmental Sciences is licensed under a Creative Commons Attribution 4.0 International License. Based on a work at SciRes Literature LLC.

Manuscript should be submitted in Word Document (.doc or .docx) through

Online Submission

form or can be mailed to support@jelsciences.com

 **Vision:** Journal of Biomedical Research & Environmental Sciences main aim is to enhance the importance of science and technology to the scientific community and also to provide an equal opportunity to seek and share ideas to all our researchers and scientists without any barriers to develop their career and helping in their development of discovering the world.

RESEARCH ARTICLE

Evaluation of Levels of Phosphatases in the Lindane Exposed Fish, *Channa punctatus*

Aradhna Gupta and Bechan Sharma*

Department of Biochemistry, University of Allahabad, India

ABSTRACT

The alterations in the activities of acid and alkaline phosphatase are good indicators of stress due to the exposure of fish to any toxicant. The perturbations in their activities may result in the decreased immune responses and intestinal microflora activity. In the present study, the effect of sublethal concentrations of lindane (0.025, 0.05 and 0.1 mg/l) in different tissues of *C. punctatus* exposed for 96h has been studied. The data showed decrease in the acid phosphatase (AcP) activity in the following order: gills>muscle>heart>brain>kidney>liver, respectively, and for the Alkaline phosphatase (ALP) the order was gills>liver>muscle>heart>brain>kidney, respectively, for 96h treatment. At highest concentration of lindane (0.1 mg/l) tested; the maximum decrease in AcP activity was recorded in gills (47.11%) and minimum in liver (15.02%). The other organs such as muscle, heart, brain and kidney exhibited 38.12, 30.85, 29.97 and 16.85% decrease, respectively. The activity of ALP showed maximum decrease in gills (27.93%) and minimum in kidney (0.98%). At highest concentration of lindane (0.1 mg/l) tested, the brain, heart, liver and muscle registered 1, 6.29, 18.84 and 14.25% decrease in ALP activity, respectively, under this condition.

Introduction

The freshwater organisms act as the sink of both natural and anthropogenic inputs of contaminants into the environment. Fish are essential components of healthy aquatic systems. They are ecologically and economically valuable providing humans with recreation and nutritive food. The release of toxic substances in the environment by either human activities or natural cases have been reported to cause excessive fish mortality [1]. The exposure of aquatic organism to even sublethal concentrations of contaminants may prove to be equally devastating [2]. Pesticides like glyphosate, Deet, Propoxur, Malathion have been exhaustively used in agriculture to combat the pest menace to produce higher yield of crops improving the economy. Among these pesticides, HCH (1,2,3,4,5,6 hexachlorocyclohexane, commonly known as lindane) is the most indiscriminately used insecticide, which has been shown to cause toxicity into the non-target aquatic organisms as well. Due to the non-biodegradable and persistent nature of HCH in the environment, it undergoes biomagnification through food-chain thereby causing serious concern to the human health [3].

The phosphatases (both the acid and alkaline) are known to be active

*Corresponding author(s)

Bechan Sharma, Department of Biochemistry, University of Allahabad, India

Email: sharmabi@yahoo.com

DOI: 10.37871/jbres1710

Submitted: 15 March 2023

Accepted: 29 March 2023

Published: 31 March 2023

Copyright: © 2023 Gupta A, et al. Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

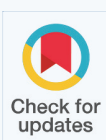
Keywords

- > Lindane
- > HCH
- > Acid phosphatases
- > Alkaline phosphatases
- > Toxicity

BIOLOGY GROUP

TOXICOLOGY

VOLUME: 4 ISSUE: 3 - MARCH, 2023



at acidic and alkaline pH ranges, respectively, and are usually termed as phosphomonoesterases. The changes in the activities of these enzymes due to pesticide exposure in the fish tissues have been reported as indicators of pesticide toxicity [4-6].

Alkaline phosphatase (ALP) is an enzyme that is found in various tissues throughout the body, such as the liver, bones, and intestines. It plays a crucial role in the metabolism of various compounds in the body, including proteins, carbohydrate metabolism, transphosphorylation reactions, secretory activity and nucleotides metabolism [7]. It affects the intestinal microbiota, causes systemic inflammation inhibiting tissue infiltration of neutrophils at the site of inflammation and induces nutrient absorption like calcium, phosphorus, fatty acids. Tissue non-specific alkaline phosphatase (TNAP) is a form of enzyme found in various tissues throughout the body, including bone, liver, kidney, and intestine. It is also involved in various other physiological processes, such as cell signaling and lipid metabolism, and abnormalities. Its expression or activity has been linked to a range of other medical conditions, including liver disease, kidney disease, and cancer. Studies have proved that it is also present in fish mucus and protects it from water-borne pathogens, detoxify proinflammatory barriers by dephosphorylation like lipopolysaccharides, and hence not recognised by their specific receptors to show any proinflammatory response involving NF- κ B activation pathway, gene activation and release of pro-inflammatory cytokines (IL-8) [8]. Its increased activity has been associated with the colonic tissue inflammation [9].

Acid phosphatase (AcP) is a lysosomal enzyme surrounded by lipoproteins that hydrolyses the phosphorous esters in acidic medium. AcP is involved in autolytic process of the cell after its death [10].

The impact of organochlorine group of pesticides on the activities of phosphatases (AcP and TNAP) in different tissues of the aquatic organisms including the fresh water fish, *Channa punctatus*, has not been properly studied. The present study, was therefore carried out by exposing *C. punctatus* with different sublethal concentrations of lindane for 96hr and evaluating the impact of the pesticide on the levels of phosphatases in the fish tissues.

Materials and Methods

Lindane was procured from Rallis India Ltd. Bangalore, India and was dissolved in acetone (AR

grade) for use. All other reagents used in this study were analytical grade.

The teleost fish, *C. punctatus* (length 6.0 cm to 8.0 cm and weight 25-30 g) were procured from the local fish market. The fish were thoroughly washed with running tap water and acclimatised in aquatic environment for 7 days. The fish were divided in four groups each containing 10 fish to be used as (i) control, treated with (ii) 0.025 mg/l, (iii) 0.05 mg/l and (iv) 0.1 mg/l lindane. Another group of fish treated with the corresponding concentration of acetone has been used in our earlier studies and it did not exert any adverse effect and the results were comparable to that of control (i).

C. punctatus, treated with different sublethal concentrations of lindane (0.025, 0.05 and 0.1 mg/l) were sacrificed after the stipulated treatment period i.e. 96h and its different body organs such as brain, gills, heart, kidney, liver and muscles were dissected out for estimation of protein and assay of enzyme activities.

Tissue homogenization

The tissues were homogenised to prepare their homogenates (10%, w/v) in normal saline under cold conditions. The homogenates of different fish tissues were centrifuged at 10,000 xg at 6°C and the supernatants were collected for estimations of different biochemical parameters.

Protein determination

The quantitative estimation of total protein in various tissue extracts were done according to Lowry OH, et al. [11]. The Bovine Serum Albumin (BSA) was used as standard. A blank was prepared which contained all reagents but no protein. The intensity of blue colour was measured calorimetrically at 620 nm.

Assays of the activities of acid and alkaline phosphatases

The activities of Acid and alkaline phosphatases (AcP, EC 3.1.3.2; ALP, EC 3.1.3.2) were assayed by the method of Salomon LL, et al. [12]. For assay of AcP activity, the reaction mixture (3.0 ml) contained sodium acetate buffer (100 mM, pH 4.5), p-nitrophenyl phosphate (2.5 mM) and cell-free extract (100-200 μ g protein). The mixture was incubated for 20 min at 37°C with intermittent gentle shaking. The reaction was terminated by addition of

NaOH (2.0 ml, 1 N). The intensity of yellow colour was measured calorimetrically at λ_{\max} 405 nm.

For assay of ALP, the reaction mixture contained p-nitrophenyl phosphate (2.0 mM) prepared in bicarbonate buffer (50 mM, pH 9.5) and cell free extract (100–200 μg protein). The mixture was incubated for 30 min at 37°C with intermittent gentle shaking. The reaction was stopped by addition of NaOH (0.4 ml, 0.1 N) which developed yellow colour. The intensity of yellow colour was measured calorimetrically at λ_{\max} 410 nm. The Optical Density (OD) observed with the control was corrected from the experimental observations. The p-nitrophenyl phosphate was used as standard.

The activity of AcP was calculated as following: $\Delta\text{OD} \times 621.1 / 0.24 \times \text{incubation time (min)} \times \text{protein (mg)}$ and the unit was expressed as nanomoles p-nitrophenol formed $\text{min}^{-1} \text{mg}^{-1}$ protein. Where 621.1 nM p-nitrophenol corresponds to OD equal to 0.24 at λ_{\max} 405 nm.

Similarly, the activity of ALP was calculated as following: $\Delta\text{OD} \times 621.1 / 0.36 \times \text{incubation time (min)} \times \text{protein (mg)}$ and the unit was expressed as nanomoles p-nitrophenol formed $\text{min}^{-1} \text{mg}^{-1}$ protein. Where 621.1 nM p-nitrophenol corresponds to OD equal to 0.36 at λ_{\max} 410nm. Spectrophotometric measurements were made using UV-VIS spectrophotometer with silica glass/quartz cuvettes (1 cm light- path, capacity 3 ml) against enzyme blank containing all the reactants at room temperature.

Results and Discussion

The effects of lindane on the activities of AcP and ALP in different organs of *C. punctatus* were monitored according to the procedure as mentioned in Materials and Methods. The results reflecting the impact of the pesticide on the enzymes' activities are shown in tables 1,2.

The data shows highest activity of AcP in kidney (50.55 \pm 2.89 units/mg protein) and lowest in gills (18.36 \pm 3.47 units/mg protein) of the control fish. The values in other tissues such as in liver, heart, muscle and brain were recorded to be 44.53 \pm 2.33, 38.116 \pm 1.73, 27.41 \pm 4.15, 20.34 \pm 2.32 units/mg protein, respectively (Table 1). The effect of lindane was more prominent at higher concentration (0.1 mg/l) than at lower concentrations (0.025, 0.05 mg/l) for 96h. At lowest concentration (0.025 mg/l), the activity of AcP was not affected in liver, whereas brain, heart and muscles displayed around 15% inhibition in the enzyme activity. Under this condition, kidney and gills showed about 5 and 20% reduction in the enzyme activity, respectively. This data indicated that at lowest concentration of the pesticide, gills were maximally affected whereas liver showed complete resistance towards pesticide toxicity. At the highest concentration of lindane (0.1 mg/l) the maximum decrease in AcP activity was recorded in gills (47.11%) and minimum in liver (15.02%). The other organs such as muscle, heart, brain and kidney exhibited 38.12, 30.85, 29.97 and 16.85% decrease in enzyme activity, respectively. Under this condition, the order of diminution in the activity of acid phosphatase in

Table 1: Effect of Lindane on the specific activity of acid phosphatase in different tissues of *C. punctatus* exposed for 96h.

Organ	Activity of Acid Phosphatase (Units/mg protein)			
	Lindane (mg/l)			
	0	0.025	0.05	0.1
Brain	20.34 \pm 2.32	17.25 \pm 1.15* (-15.23)	17.19 \pm 1.73* (-12.33)	14.23 \pm 3.47* (-29.97)
Gills	18.36 \pm 3.47	14.72 \pm 4.05 (-19.84)	11.38 \pm 1.16* (-37.96)	9.70 \pm 2.54* (-47.11)
Heart	38.116 \pm 1.73	32.34 \pm 1.74 (-15.11)	30.86 \pm 2.87 (-18.96)	26.40 \pm 3.45* (-30.85)
Kidney	50.55 \pm 2.89	47.95 \pm 2.31* (-4.96)	44.27 \pm 2.29* (-12.42)	42.03 \pm 1.15* (-16.85)
Liver	44.53 \pm 2.33	42.44 \pm 2.31 (-4.7)	39.08 \pm 2.32 (-12.26)	37.86 \pm 4.04 (-15.02)
Muscle	27.41 \pm 4.15	23.42 \pm 1.85 (-14.57)	18.84 \pm 1.18 (-31.24)	16.96 \pm 1.18* (-38.12)

Values are presented as nM p-nitrophenol formed $\text{min}^{-1} \text{mg}^{-1}$ protein. Each value represents the mean \pm SEM of three different observations. Values in parenthesis are percent change over control. The (-) sign represents decrease over control. Significance of data is shown in superscripts. Significantly different from control at ** $p < 0.01$, * $p < 0.05$ (Student's t test). h: hour, SEM: Standard Error of Mean.

different tissues of the fish was recorded as following: gills>muscle>heart> brain>kidney>liver. The data indicated that at highest concentration (0.1 mg/l), gills displayed maximum sensitivity to lindane whereas the liver was affected to minimum possible extent (Table 1).

The results of the effect of sublethal concentrations of lindane (0.025, 0.05 and 0.1 mg/l) on the activity of ALP in different tissues of *C. punctatus* is shown in table 2. The data demonstrated highest activity of ALP to be present in kidney (82.03 ± 1.49 units/mg protein) and lowest in brain (30.4 ± 2.88 units/mg protein) of the control fish. The activities of ALP in other fish tissues such as liver, heart, muscle and gills were recorded as 70.84 ± 2.92, 53.60 ± 2.26, 4.23 ± 4.44 and 36.82 ± 3.45 units/mg protein, respectively. At the highest concentration of lindane treatment, the order of decrease in the activity of ALP in different fish tissues was as follows: gills>liver>muscle>heart >brain>kidney (Table 2).

The comparison of the data presented in tables 1,2 indicated that lindane at subacute concentrations caused marked decrease in the activities of enzymes in all the organs of the fish tested. In both of the cases, the inhibitory effect of lindane was concentration dependent and organ specific. However, the effect was more marked on the AcP as compared to ALP. Lindane caused sharp decrease in the activity of AcP and ALP in the gills of the fish at highest concentration of lindane (0.1 mg/l). The activity of AcP was least affected in liver whereas the activity of ALP was least affected in kidney of the fish exposed to the pesticide for 96h.

The activities of AcP and ALP are good indicators of stress of a toxicant on fish [13]. Any damage or dysfunction in experimental organs is manifestation of alteration in phosphatase activity [14]. The reduction in the activities of these enzymes could be due to increased utilization of protein to meet out the increased energy cost of homeostasis, tissue repair and detoxification during stress [15,16].

The decrease in AcP activity correlates its role in cellular activity and cellular damage leading to autolytic breakdown or cellular necrosis resulting in insufficient production of enzymes or leakage of enzyme into extracellular compartment due to pesticide treatment [17]. Necrotic changes in different organs might be responsible for increase or decrease in the activity of phosphatases [18–20]. The altered enzyme activities can be due to factors that influence the rate at which enzyme enters the circulation from the cells or rate of enzyme production by individual cell type or proliferation of a particular type of enzyme producing cell. It is generally accepted that an increase or decrease in the activity of enzymes in the extracellular fluid or plasma is a sensitive indicator of minor cellular damage [21].

It was observed in *Cyprinus carpio* that a herbicide, 2,4 diamin at sublethal concentrations (50 and 80 ppm) did not cause any significant alteration in the activities of both phosphatases after different exposure period (1–30 days) and they have proposed that this herbicide does not cause destruction of cell membranes and lysosomes, which could liberate their hydrolases into the cell system of tissues to

Table 2: Effect of Lindane on the specific activity of alkaline phosphatase in different tissues of *C. punctatus* exposed for 96h

Organ	Activity of Acid Phosphatase (Units/mg protein)			
	Lindane (mg/l)			
	0	0.025	0.05	0.1
Brain	30.4 ± 2.88	35.5 ± 3.78* (17)	33.7 ± 2.36 (11)	30.09 ± 2.86 (-1)
Gills	36.82 ± 3.45	34.80 ± 2.42 (-5.48)	30.62 ± 2.86 (-16.78)	26.56 ± 2.03 (-27.93)
Heart	53.60 ± 2.26	56.145 ± 4.05* (4.75)	57.50 ± 1.23* (7.29)	50.22 ± 2.86 (-6.29)
Kidney	82.03 ± 1.49	80.02 ± 3.28 (-2.45)	83.28 ± 2.60 (1.52)	81.23 ± 3.20 (-0.98)
Liver	70.84 ± 2.92	67.96 ± 2.23 (-3.99)	58.22 ± 1.84* (-17.78)	57.50 ± 4.14* (-18.84)
Muscle	47.23 ± 4.44	44.19 ± 4.36 (-6.42)	42.30 ± 2.03* (-9.42)	40.48 ± 2.88* (-14.25)

Values are represented as nM p-nitrophenol formed min⁻¹ mg⁻¹protein. Each value represents the mean± SEM of three different observations. Values in parenthesis are percent change over control. The (-) sign represents decrease over control. Significance of data is shown in superscripts. Significantly different from control at **p < 0.01, *p < 0.05 (Student's t test). h: hour; SEM: Standard Error of Mean.

facilitate autolysis [22]. The decrease in activity of AcP in the tissues of *C. punctatus* exposed to sublethal concentrations of lindane in present investigation is supported by the results obtained with *C. gachua* exposed to thiodan and rogan for 30 days [23]. A reduction in the gill AcP activity was reported in rosy barb, *Puntius conchonius* (Hamilton) exposed to copper and cadmium toxicity [24], in the ovary and liver of *C. punctatus* exposed to zinc (10–25 mg/l) for 8, 10 and 15 days [25], in *C. punctatus* exposed to monocotrophs [26], and in *Clarius gariepinus* exposed to aqueous leaf extract of *Lepidagathis alopecuroides* [27].

Apart from pesticides, the sharp decline in the activities of AcP and ALP has been reported by many workers for example in *H. fossilis* treated with cadmium; *Sarotherodon mossambicus* with mercury [28]; in *Labeo rohita* with copper, and zinc [29] and in rainbow trout (*O. Mykiss*) with chromium [30]. However, *Perna viridis* has been shown to get affected by the exposure to copper, lead and zinc causing elevation in the ALP activity [31].

The decrease in activity of ALP in the present study could be due to uncoupling of oxidative phosphorylation. Its activity has been shown to be reduced by 56% in Crucian carp (*Carassius carassius*) [32] when water temperature changed from 18°C to 2°C. The use of alkaline water with pH 8.0–7.5 has resulted into decreased activity by 22% at 22°C in Flat fish larvae (*S. senegalensis*) [33], and in the liver of *Sarotherodon mossambicus* [34]. The acidic water may inactivate ALP to get energy via anaerobic breakdown of glycogen. In saline water its activity was found to be increased in juveniles of *Alosa sapidissima* [35]. Its activity in general was found to be higher in summer and lower in winter in Crucian carp (*C. carassius*) [36]. However, the impact of physico chemical properties of water on the activity needs more study to reach to any conclusion. It may also depend upon food intake, oxygen concentration of water; season, and the place from where the fish has been taken etc. Several studies have reported increased enzyme activity in fish blood due to the leakage from liver [37,38]. The reduction in the ALP activity in the brain may be attributable to the alteration in the nucleic acid content because of pollutants. A statistically significant decline in AcP and ALP activities during 30 days exposure of fish to pesticide has been reported [39]. In contrast to our findings some workers have reported the increase in the activities of AcP and ALP, which has been attributed to onset of hepatic disorder and inflammatory reactions etc. [40–44].

Conclusion

The data suggests that lindane even at the very small concentration was able to pose threats to the life of *C. punctatus*. The sub-lethal concentrations of lindane caused perturbations in the levels of phosphatases in all the organs of the fish but maximum was in gills of the fish exposed for 96h. The perturbations in the levels of phosphatases may be used as early signals for assessment of toxic effects of the pesticides in aquatic system. The results from this study may be useful towards adequate formulation of pesticides, and policy makers for the sustainable management of ecophysiological and the environmental health of aquatic animals. These results also may offer a sound understanding of the toxicological endpoint of the aquatic life. Further research is needed to fully understand the effects of lindane on fish and other organisms and their potential implications for the environment and human health.

Acknowledgment

The authors are grateful to University of Allahabad for providing facilities for carrying out the present work. AG acknowledges UGC–New Delhi for providing the financial in the form of a scholarship.

Conflicts of Interest

There is no conflict of interest to be disclosed.

References

1. EPA. Protecting water quality from agricultural runoff. Fact Sheet No. EPA-841-F-05-001. 2005.
2. Gupta A, Rai DK, Pandey RS, Sharma B. Analysis of some heavy metals in the riverine water, sediments and fish from river Ganges at Allahabad. *Environ Monit Assess.* 2009 Oct;157(1-4):449-58. doi: 10.1007/s10661-008-0547-4. Epub 2008 Oct 11. PMID: 18850290.
3. Chaudhuri K, Selvaraj S, Pal AK. Studies on the genotoxicity of endosulfan in bacterial systems. *Mutat Res.* 1999 Feb 2;439(1):63-7. doi: 10.1016/s1383-5718(98)00174-0. PMID: 10029677.
4. Bhatnagar MC, Tyagi M, Tamata S. Pyrethroid induced toxicity to phosphatases in *C. batrachus*. *J Environ Biol.* 1995;16:11-14.
5. Velišek J, Jurčíková J, Dobšíková R, Svobodová Z, Piačková V, Máčková J, Novotný L. Effects of deltamethrin on rainbow trout (*Oncorhynchus mykiss*). *Environ Toxicol Pharmacol.* 2007 May;23(3):297-301. doi: 10.1016/j.etap.2006.11.006. Epub 2006 Nov 18. PMID: 21783771.
6. Bhattacharya M, Kaviraj A. Toxicity of the pyrethroid

- pesticide fenvalerate to freshwater catfish *Clarias gariepinus*: lethality, biochemical effects and role of dietary ascorbic acid. *J Environ Sci Health B*. 2009 Aug;44(6):578-83. doi: 10.1080/03601230903000602. PMID: 20183065.
7. Kumar A, Sharma B, Pandey RS. Toxicological assessment of the pyrethroids insecticides with special reference to cypermethrin and λ -cyhalothrin in fresh water fishes. *Int J Biol Med Res*. 2010;1(4):315-325.
 8. Guardiola FA, Cuesta A, Esteban MÁ. Using skin mucus to evaluate stress in gilthead seabream (*Sparus aurata* L.). *Fish Shellfish Immunol*. 2016 Dec;59:323-330. doi: 10.1016/j.fsi.2016.11.005. Epub 2016 Nov 3. PMID: 27818341.
 9. Lallès JP. Biology, environmental and nutritional modulation of skin mucus alkaline phosphatase in fish: A review. *Fish Shellfish Immunol*. 2019 Jun;89:179-186. doi: 10.1016/j.fsi.2019.03.053. Epub 2019 Mar 27. PMID: 30928666.
 10. Khan S, Sharma N. A study of enzymes acid phosphatase and alkaline phosphatase in the liver and kidney of fish *Gambusia affinis* exposed to the chlorpyrifos, an organophosphate. *Int J Pharm Sci Rev Res*. 2012;13(1):88-90.
 11. LOWRY OH, ROSEBROUGH NJ, FARR AL, RANDALL RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951 Nov;193(1):265-75. PMID: 14907713.
 12. Salomon LL, James J, Weaver PR. Assay of phosphatase activity by direct spectrophotometric determination of phenolate ion. *Anal Chem*. 1964;36(6):1162-1164. doi: 10.1021/ac60212a071.
 13. Plummer DT. An introduction to practical biochemistry. New Delhi, India: Tata Publishing Company; 1971.
 14. Velisek J, Inlason T, Gomulka P, Svoboda Z, Dobsikova R, Noyoty L, Dudzik M. Effects of cypermethin on rainbow trout (*Onchorhynchus mykiss*). *Veterinari Medicina*. 2006;51(10):469-476.
 15. Ogueji EO, Auta J. Investigation of biochemical effects of acute concentrations of λ -cyhalothrin on African catfish *Clarias batrachus*-Teugels. *Journal of Fisheries International*. 2007;2(1):86-90.
 16. Thatheyus AJ, Selvanayagam M, Sebastin Raj S. Toxicity of nickel on protein content of *Cyprinus carpio* var. *Communis* (Linn.). *Indian J Environ Hlth*. 1992;34(3):236-238.
 17. Agrahari S, Gopal K. Fluctuations of certain biochemical constituents and marker enzymes as a consequence of monocotrophs toxicity in the Indian edible freshwater fish *C. punctatus*. *Pestic Biochem Physiol*. 2009;94:5-9.
 18. Gill TS, Pande J, Tewari H. Sublethal effects of an organophosphorus insecticide on certain metabolite levels in a freshwater fish, *Puntius conchonius* Hamilton. *Pesticide Biochemistry and Physiology*. 1990;36:290-299.
 19. El-Sayed YS, Saad TT. Subacute intoxication of a deltamethrin-based preparation (Butox) 5% EC in monosex Nile tilapia, *Oreochromis niloticus* L. *Basic Clin Pharmacol Toxicol*. 2008 Mar;102(3):293-9. doi: 10.1111/j.1742-7843.2007.00157.x. Epub 2007 Dec 5. PMID: 18053029.
 20. Bhattacharya M, Kaviraj A. Toxicity of the pyrethroid pesticide fenvalerate to freshwater catfish *Clarias gariepinus*: lethality, biochemical effects and role of dietary ascorbic acid. *J Environ Sci Health B*. 2009 Aug;44(6):578-83. doi: 10.1080/03601230903000602. PMID: 20183065.
 21. Kori-Siakpere O, Ikomi RB, Ogbe MG. Variations in acid phosphatase and alkaline phosphatase activities in the plasma of the African catfish *Clarias gariepinus* exposed to sublethal concentration of potassium permanganate. *Asian J Exp Biol Sci*. 2010;1:170-174.
 22. Velíšek J, Jurčíková J, Dobšíková R, Svobodová Z, Piačková V, Máčková J, Novotný L. Effects of deltamethrin on rainbow trout (*Oncorhynchus mykiss*). *Environ Toxicol Pharmacol*. 2007 May;23(3):297-301. doi: 10.1016/j.etap.2006.11.006. Epub 2006 Nov 18. PMID: 21783771.
 23. Oruc EO, Uner N. Effects of 2,4 diamine on some parameters of protein and carbohydrate metabolism in the serum, muscle and liver of *Cyprinus carpio*. *Environ Poll A*. 1999;41:165-177.
 24. Bhatnagar MC, Bana AK. Pesticides induced histophysiological alterations in liver of *Channa gachua*. *Proc Acad Environ Biol*. 1993;2:115-118.
 25. Gill TS, Pande J, Tewari H. Individual and combined toxicity of common pesticides to teleost *Puntius conchonius* Hamilton. *Indian J Exp Biol*. 1991 Feb;29(2):145-8. PMID: 1869298.
 26. Verma H, Srivastava N. Changes in certain enzyme of the ovary and liver in *Channa punctatus*. *Electronic J. Ichthyol*. 2010;2:85-91.
 27. Agrahari S, Gopal K. Inhibition of Na⁺-K⁺-ATPase in different tissues of fresh water fish *Channa punctatus* (Bloch) exposed to monocotrophs. *Pesticide Biochemistry and Physiology*. 2008;92:57-60.
 28. Obamanu FG, Gabriel UU, Edori OS, Emetonjr JN. Biomarker enzymes in muscle tissue and organs of *Clarias gariepinus* after intramuscular injection with aqueous extracts of *Lepidagathis alopecuroides* leaves. *Journal of Medicinal Plants Research*. 2009;3(12):995-1001.
 29. Naidu KA, Naidu KA, Ramamurthi R. Acute effect of mercury toxicity on some enzymes in liver of teleost *Sarotherodon mossambicus*. *Ecotoxicol Environ Saf*. 1984 Jun;8(3):215-8. doi: 10.1016/0147-6513(84)90024-1. PMID: 6329640.
 30. Trivedi R, Rajbanshi VK. Effect of zinc and copper on the Akaine phosphatases of *Labeo rohita* (Hamilton). *The fifth Indian Fisheries forum proceedings*. 2002.
 31. Boge G, Leydet G, Houvet D. The effects of hexavalent chromium on the activity of alkaline phosphatase in the intestine of rainbow trout (*Onchorhynchus mykiss*). *Aquatic Toxicology*. 1992;23:247-260.

32. Aanand S, Purushthamam CS, Pal AK, Rajendra KV. Toxicological studies on the effect of copper, lead and zinc on selected enzymes in the adductor muscle and intestinal diverticula of the green mussel *Perna viridis*. Indian Journal of Marine Sciences. 2010;39(2):299-302.
33. Varis J, Haverinen J, Vornanen M. Lowering Temperature is the Trigger for Glycogen Build-Up and Winter Fasting in Crucian Carp (*Carassius carassius*). Zoolog Sci. 2016 Feb;33(1):83-91. doi: 10.2108/zs150072. PMID: 26853873.
34. Pimentel MS, Faleiro F, Diniz M, Machado J, Pousão-Ferreira P, Peck MA, Pörtner HO, Rosa R. Oxidative Stress and Digestive Enzyme Activity of Flatfish Larvae in a Changing Ocean. PLoS One. 2015 Jul 29;10(7):e0134082. doi: 10.1371/journal.pone.0134082. PMID: 26221723; PMCID: PMC4519323.
35. Shaikila BI, Thangavel P, Ramaswamy M. Adaptive trends in tissue acid and alkaline phosphatase of *Sarotherodon mossambica* (Peters) under sevin toxicity. Indian Journal of Environmental Health. 1993;35(1):36-39.
36. Liu ZF, Gao XQ, Yu JX, Qian XM, Xue GP, Zhang QY, Liu BL, Hong L. Effects of different salinities on growth performance, survival, digestive enzyme activity, immune response, and muscle fatty acid composition in juvenile American shad (*Alosa sapidissima*). Fish Physiol Biochem. 2017 Jun;43(3):761-773. doi: 10.1007/s10695-016-0330-3. Epub 2016 Dec 24. PMID: 28013424.
37. Penttinen OP, Holopainen IJ. Seasonal feeding activity and ontogenetic dietary shifts in crucian carp, *Carassius carassius*. Environmental Biology of Fishes. 1992;33:215-221. doi: 10.1007/BF00002566.
38. Firat O, Cogun HY, Yüzereroğlu TA, Gök G, Firat O, Kargin F, Kötemen Y. A comparative study on the effects of a pesticide (cypermethrin) and two metals (copper, lead) to serum biochemistry of Nile tilapia, *Oreochromis niloticus*. Fish Physiol Biochem. 2011 Sep;37(3):657-66. doi: 10.1007/s10695-011-9466-3. Epub 2011 Jan 13. PMID: 21229307; PMCID: PMC3146979.
39. Meenambal M, Pugazhendy K, Vasantharaja C, Venkatesan S. Ameliorative property of *Delonix elata* supplementary feed against cypermethrin induced serum biochemical changes in freshwater fish *Cyprinus carpio* (Linn). Journal of Pharmacy Research. 2012;5(5):2489-2492.
40. Sharma SD, Maya K. Alkaline and acid phosphatases in different tissues of *Channa punctatus* exposed to metanil yellow. Indian J Fish. 1994;41(1):48-52.
41. Ram RN, Singh SK. Carbofuran-induced histopathological and biochemical changes in liver of the teleost fish, *Channa punctatus* (Bloch). Ecotoxicol Environ Saf. 1988 Dec;16(3):194-201. doi: 10.1016/0147-6513(88)90050-4. PMID: 3229378.
42. Ayalogu OE, Lgbon NM, Dede EB. Biochemical changes in the serum and liver of albino rats exposed to petroleum samples (gasoline, kerosene and crude petroleum). J Appl Sci Environ Mgt. 2001;5(1):97-100.
43. Svoboda M, Luskova V, Drastichova J, Llabok V. The effect of diazinon on haematological indices of common carp (*Cyprinus carpio*). Acta Vet Brno. 2001;70:457-465.
44. Obamanu FG, Gabriel UU, Edori OS, Emetonjr JN. Biomarker enzymes in muscle tissue and organs of *Clarius gariepinus* after intramuscular injection with aqueous extracts of *Lepidagathis alopecuroides* leaves. Journal of Medicinal Plants Research. 2009;3(12):995-1001.

How to cite this article: Gupta A, Sharma B. Evaluation of Levels of Phosphatases in the Lindane Exposed Fish, *Channa punctatus*. 2023 Mar 31; 4(3): 555-561. doi: 10.37871/jbres1710, Article ID: JBRES1710, Available at: <https://www.jelsciences.com/articles/jbres1710.pdf>