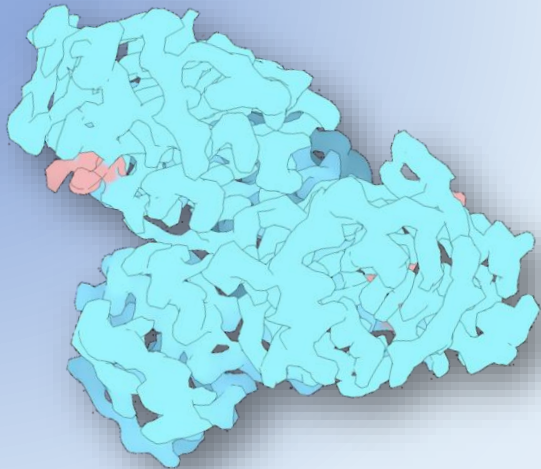


Structural insights on the SARS-CoV-2 Main Protease maturation process and inhibition



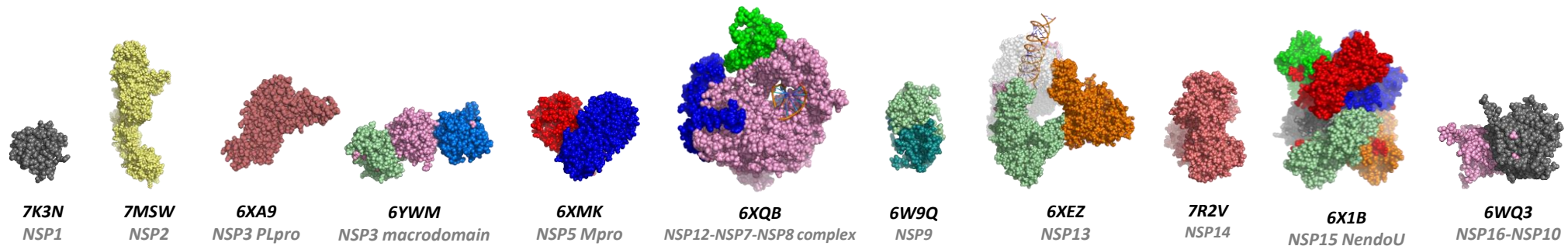
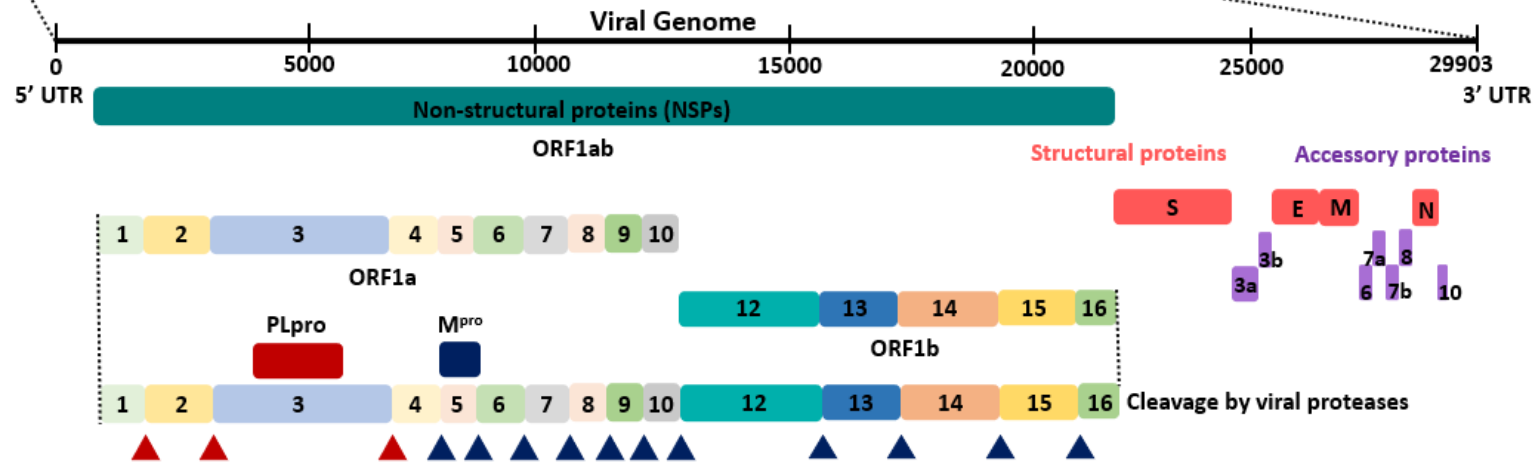
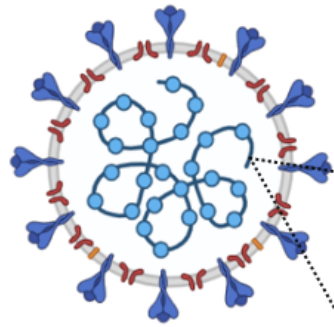
Andre Schutzer Godoy, PhD

X-ray crystallography, cryo-EM, fragment screening, virology

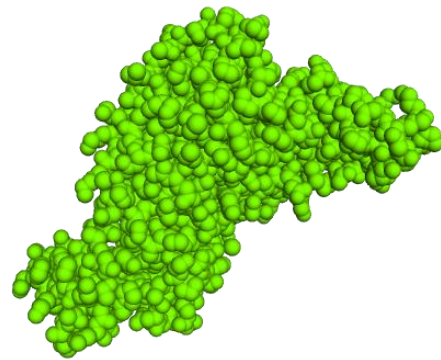
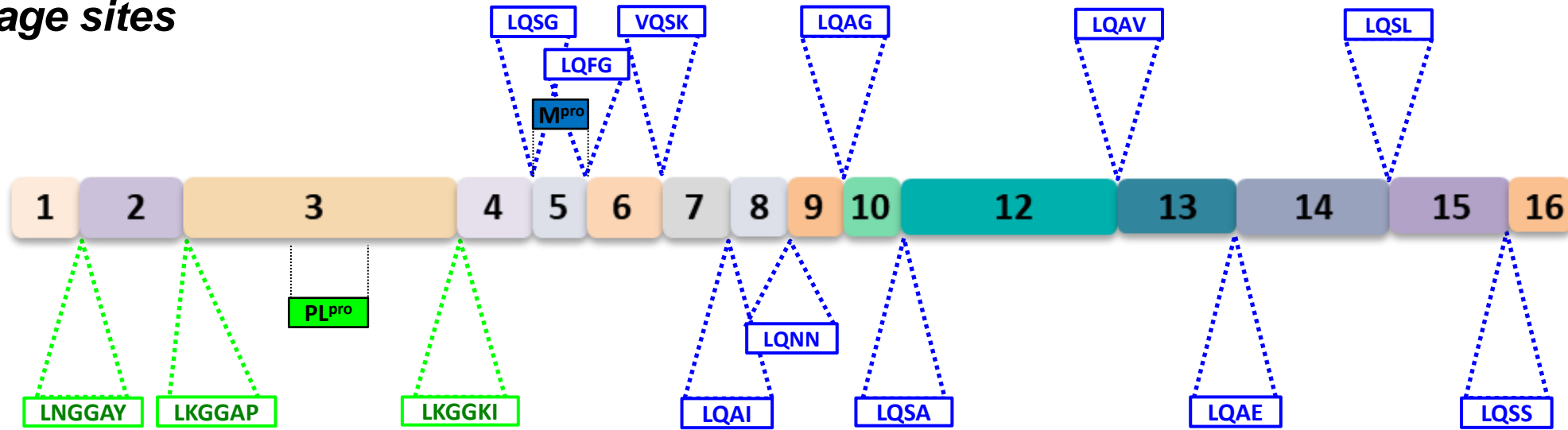
University of Sao Paulo - Brazil
AI-driven Structure-enabled Antiviral Platform
NIH AviDD U19 Center

andregodoy@ifsc.usp.br, andre.schgodoy@gmail.com

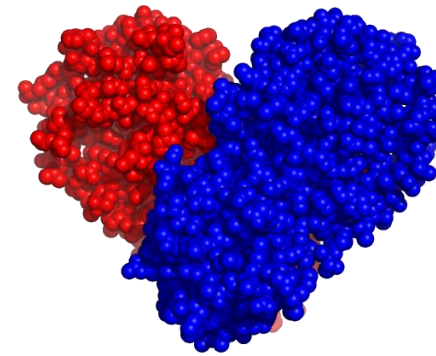
SARS-CoV-2 Genome Organization



Cleavage sites

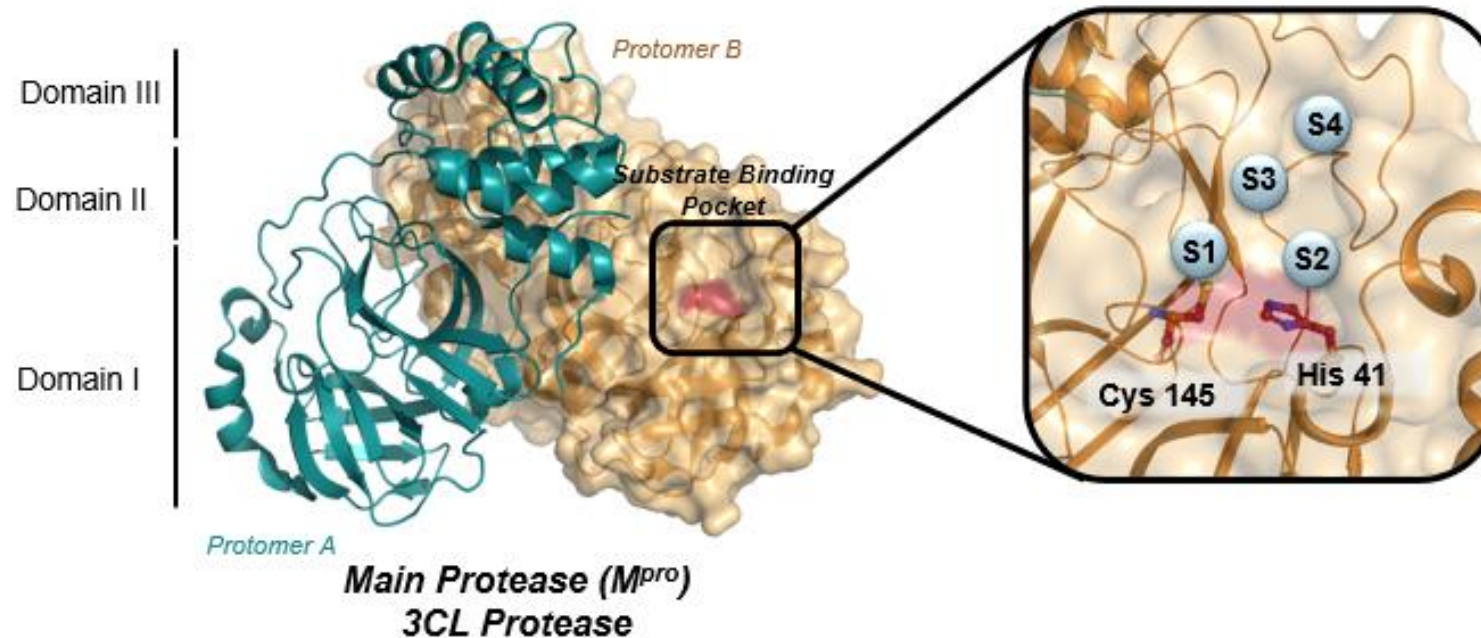


Papain-like protease

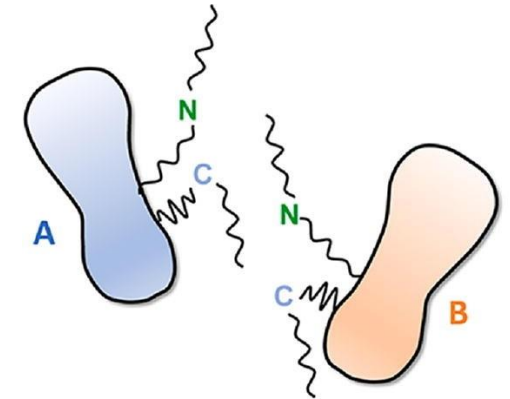


*Main protease
3CL protease*

- Cysteine protease with dimeric structure
- Responsible for the cleavage of 11 sites of the polyprotein, including its own N and C-terminal
- Over 600 structures deposited on PDB (March 2023)
- Key target for antiviral development
- Lack of information about its self-maturation process

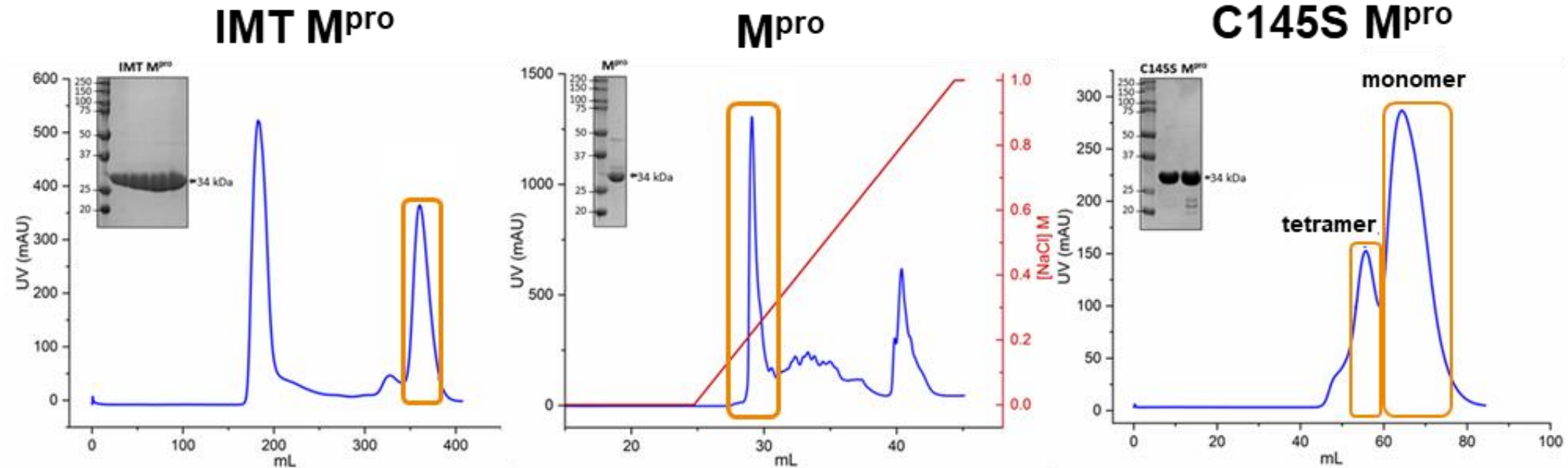
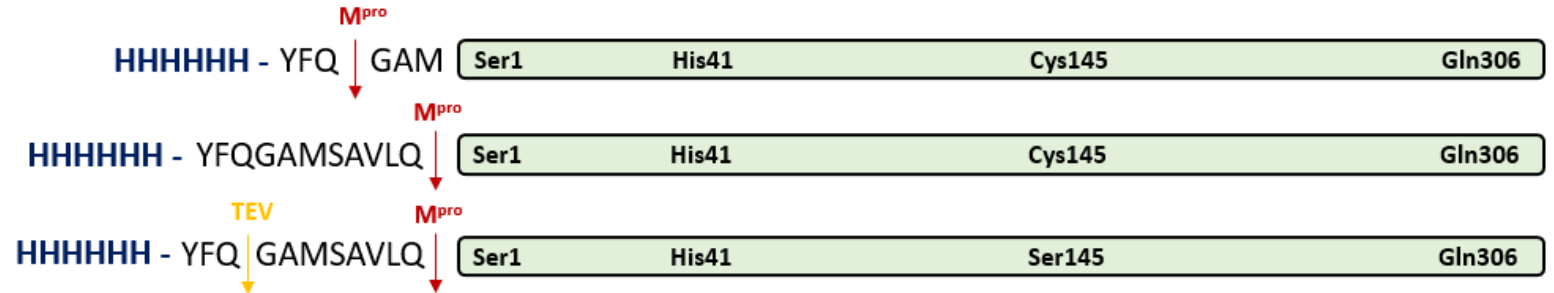


- If the protein is active only as a dimer, how copies of immature M^{pro} , still as part of the entire viral polypeptide, get together to conduct its self-cleavage ?
- The N and C-terminus processing occurs within a dimer (*cis*-cleavage) or between two distinct dimers (*trans*-cleavage)
- What are there conformational changes involved ?
- Can NEW CAVITIES be identified in the immature M^{pro} , that could be explored for early inhibition of the protease activity ?



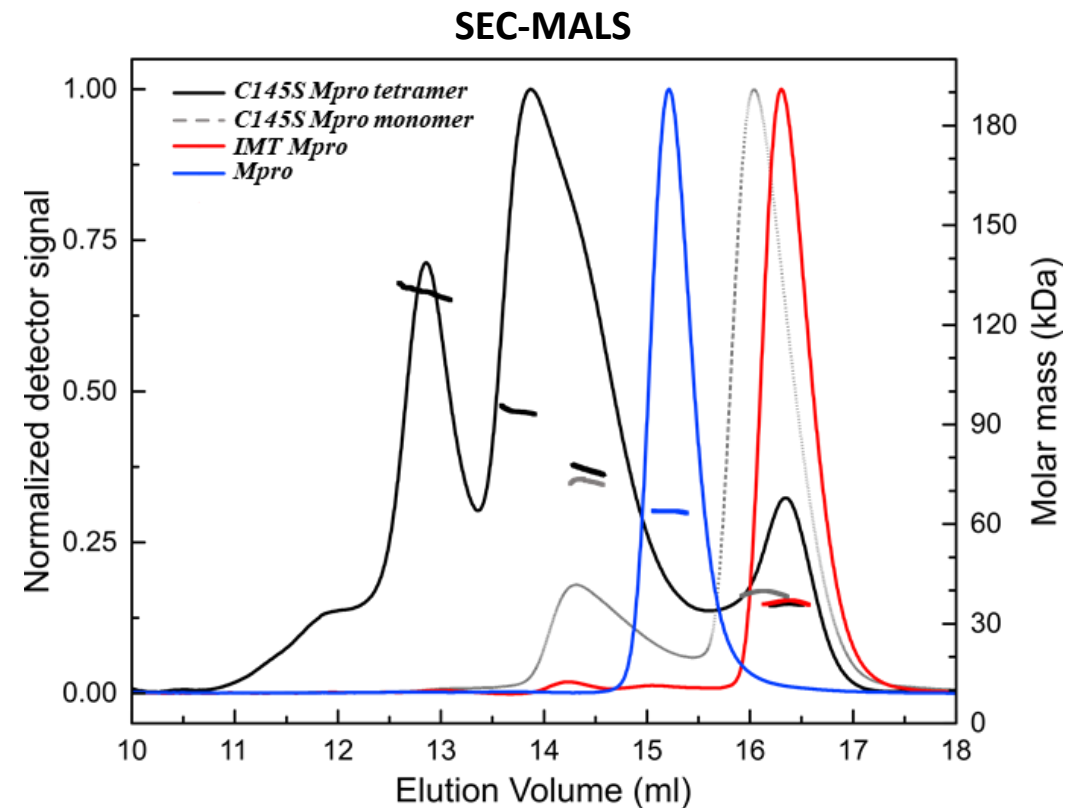
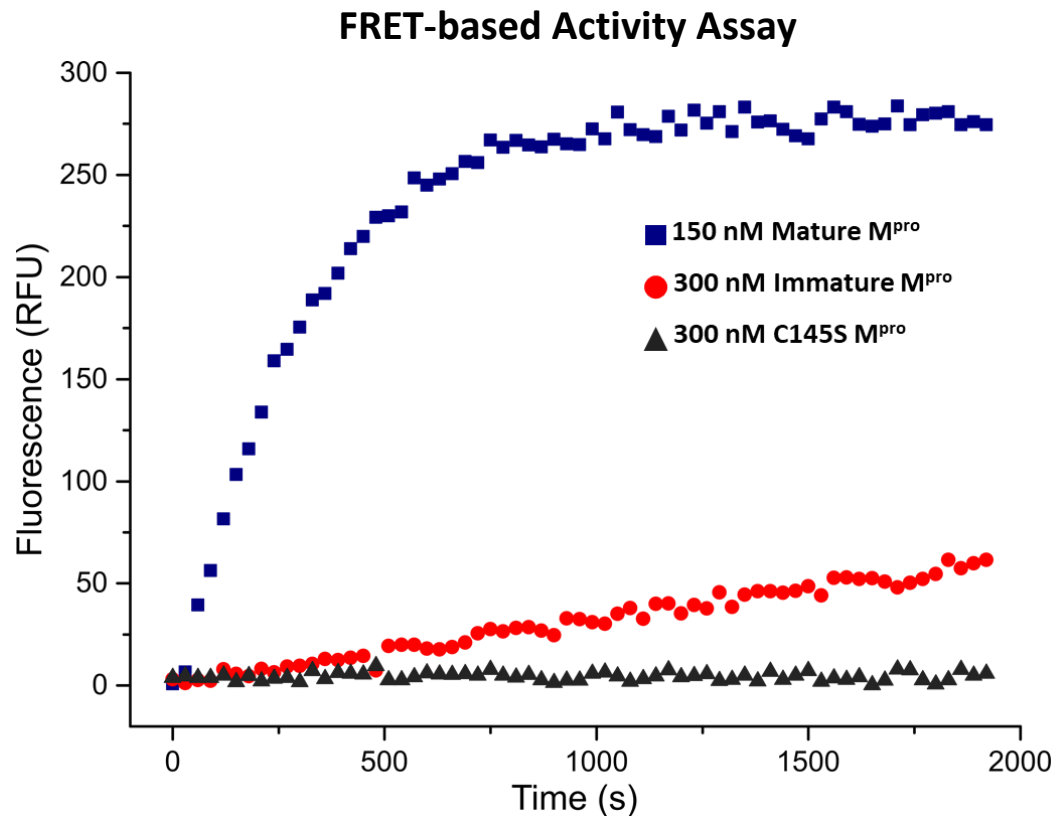
Immature M^{pro}
protomers

- Immature form **IMT M^{pro}**
- Mature or native **M^{pro}**
- C145S mutant **C145S M^{pro}**



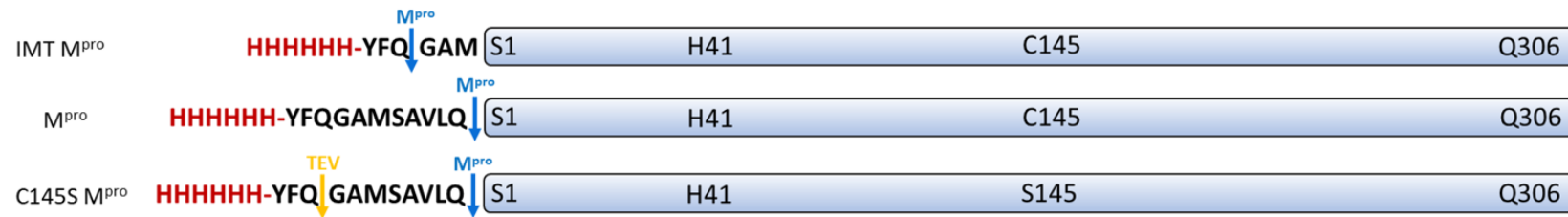
In-solution characterization:

- IMT-Mpro has reduced activity and is monomeric in solution
- C145S Mpro is inactive and a mixture of oligomeric states ranging from monomers to tetramers

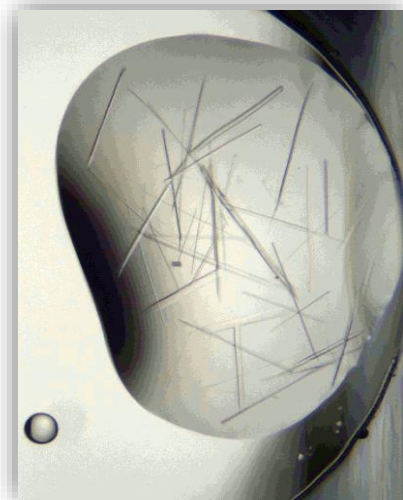


SARS-CoV-2 Main-Protease Constructs:

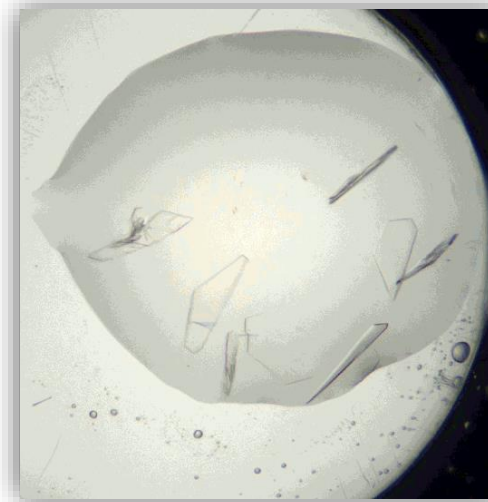
- *Immature form*
- *Mature or native*
- *C145S mutant*



IMT M^{pro}



M^{pro}

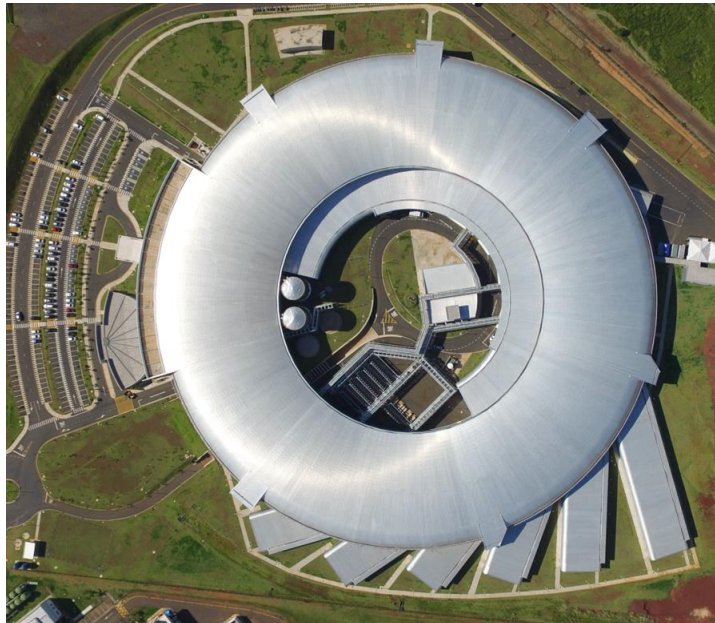
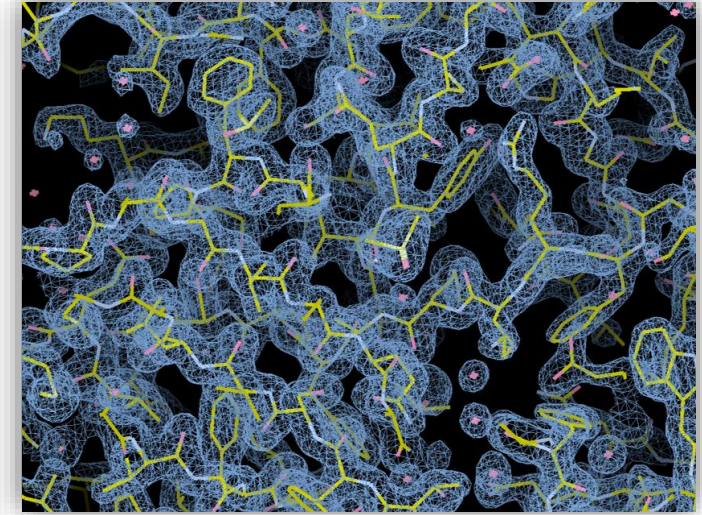


C145S M^{pro}



1st SIRIUS External users experiment

SIRIUS, Campinas – SP - September 2020



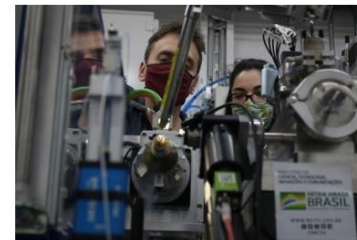
Primeiro experimento realizado no Sirius busca desenvolver fármaco para COVID-19

20 de outubro de 2020

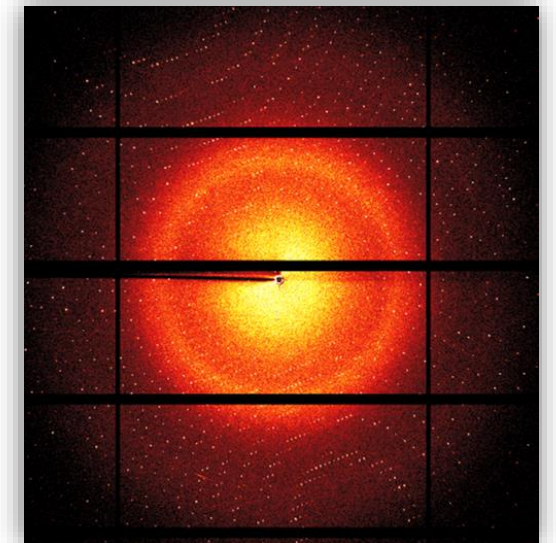


Maria Fernanda Ziegler* | Agência FAPESP – Por meio de um potente feixe de luz síncrotron foi possível determinar, em três dias, a estrutura de mais de 200 cristais de duas proteínas do novo coronavírus (SARS-CoV-2).

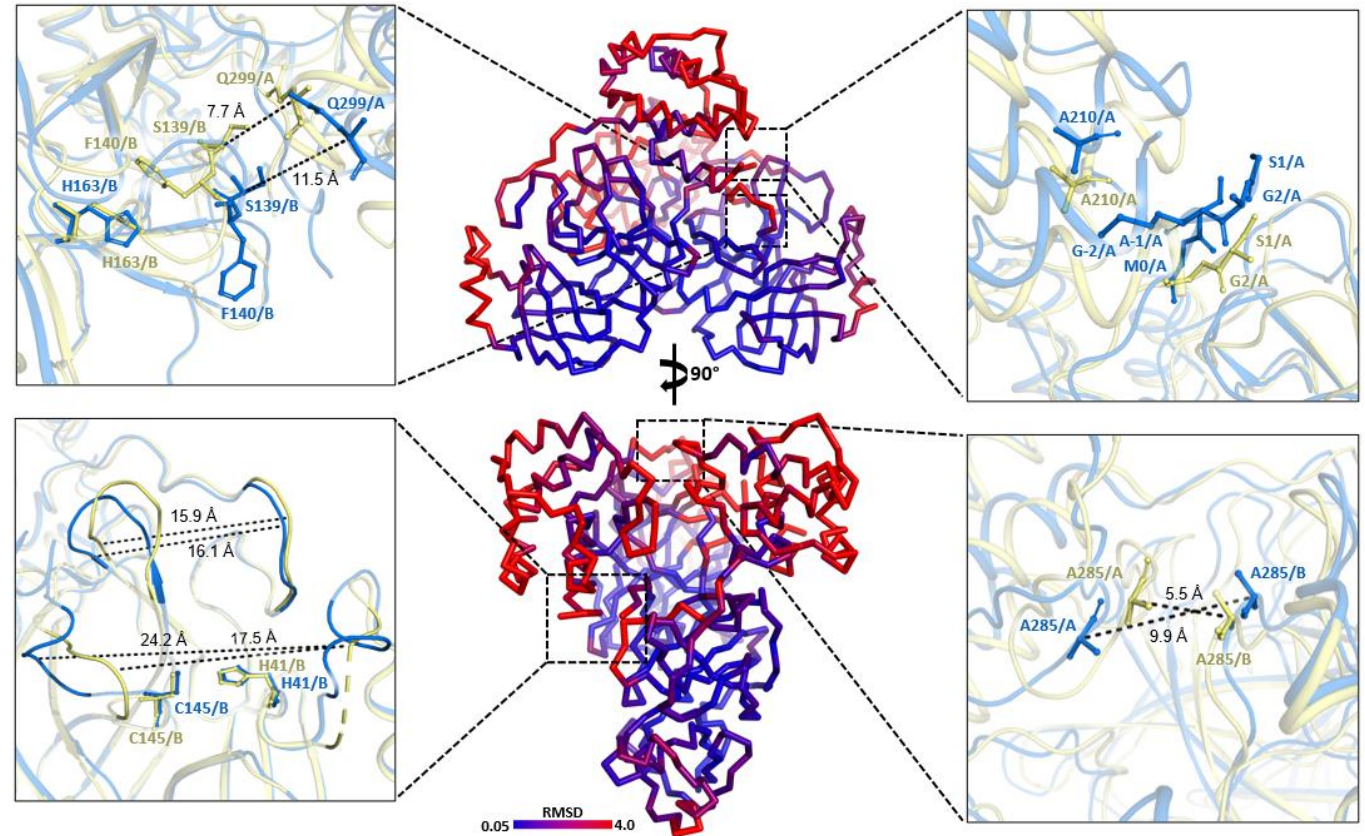
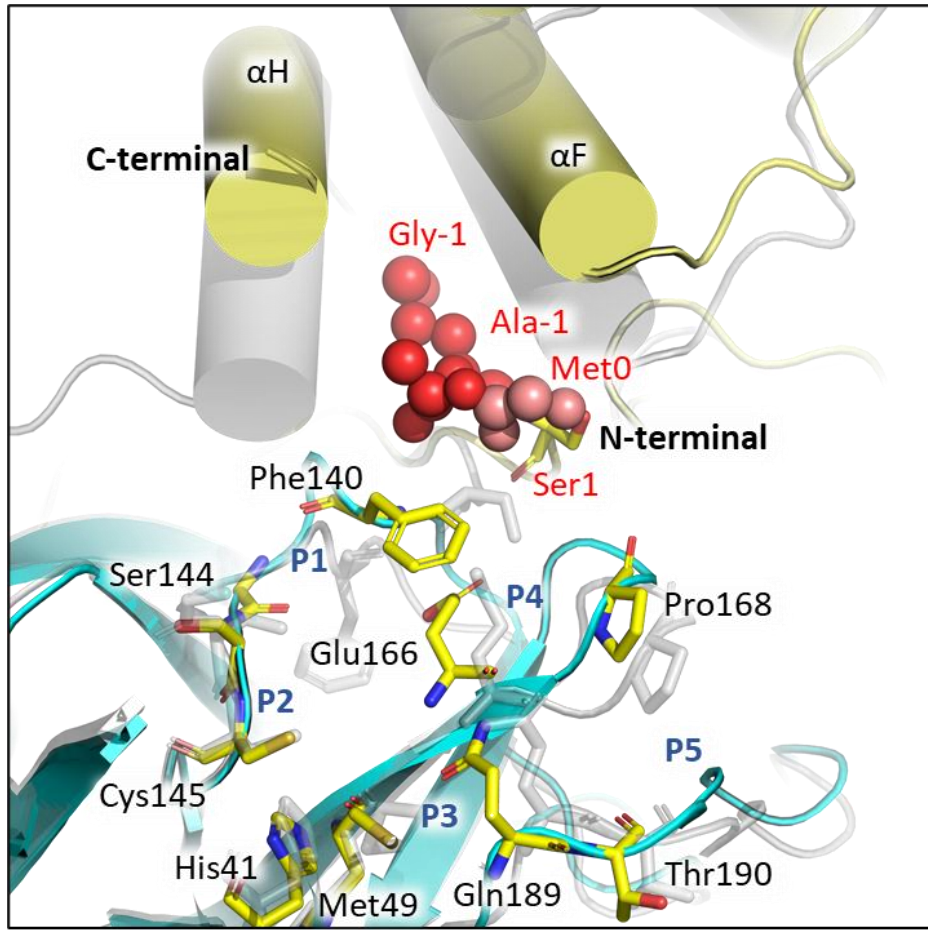
A investigação realizada por pesquisadores do Instituto de Física de São Carlos da Universidade de São Paulo (IF-USP) tem importância não só pela temática – essencial para o desenvolvimento de um possível fármaco contra a COVID-19 –, mas também pelo seu caráter de ineditismo.



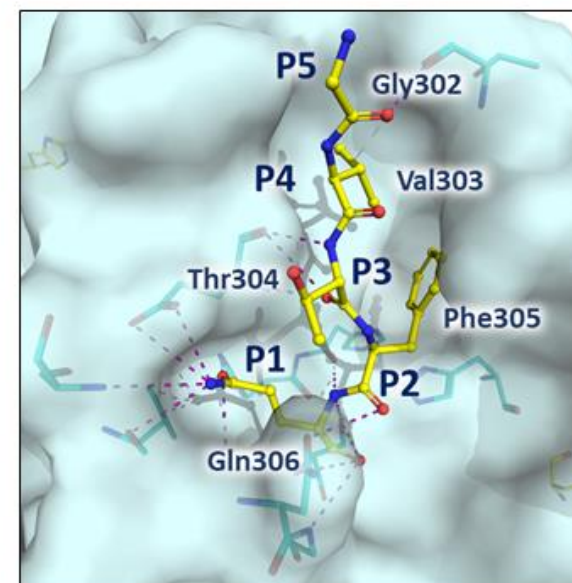
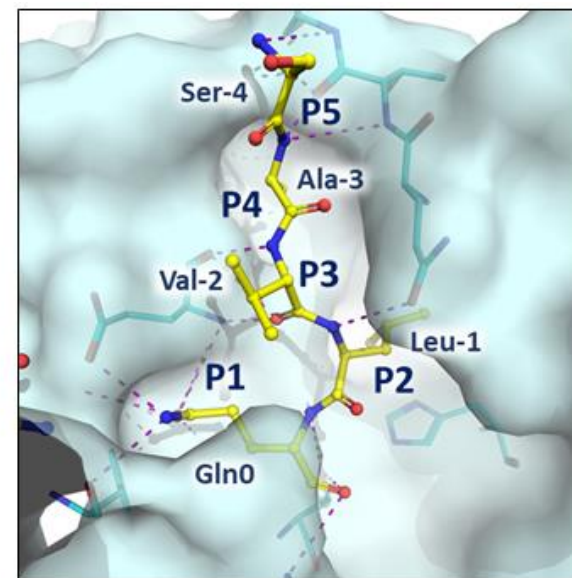
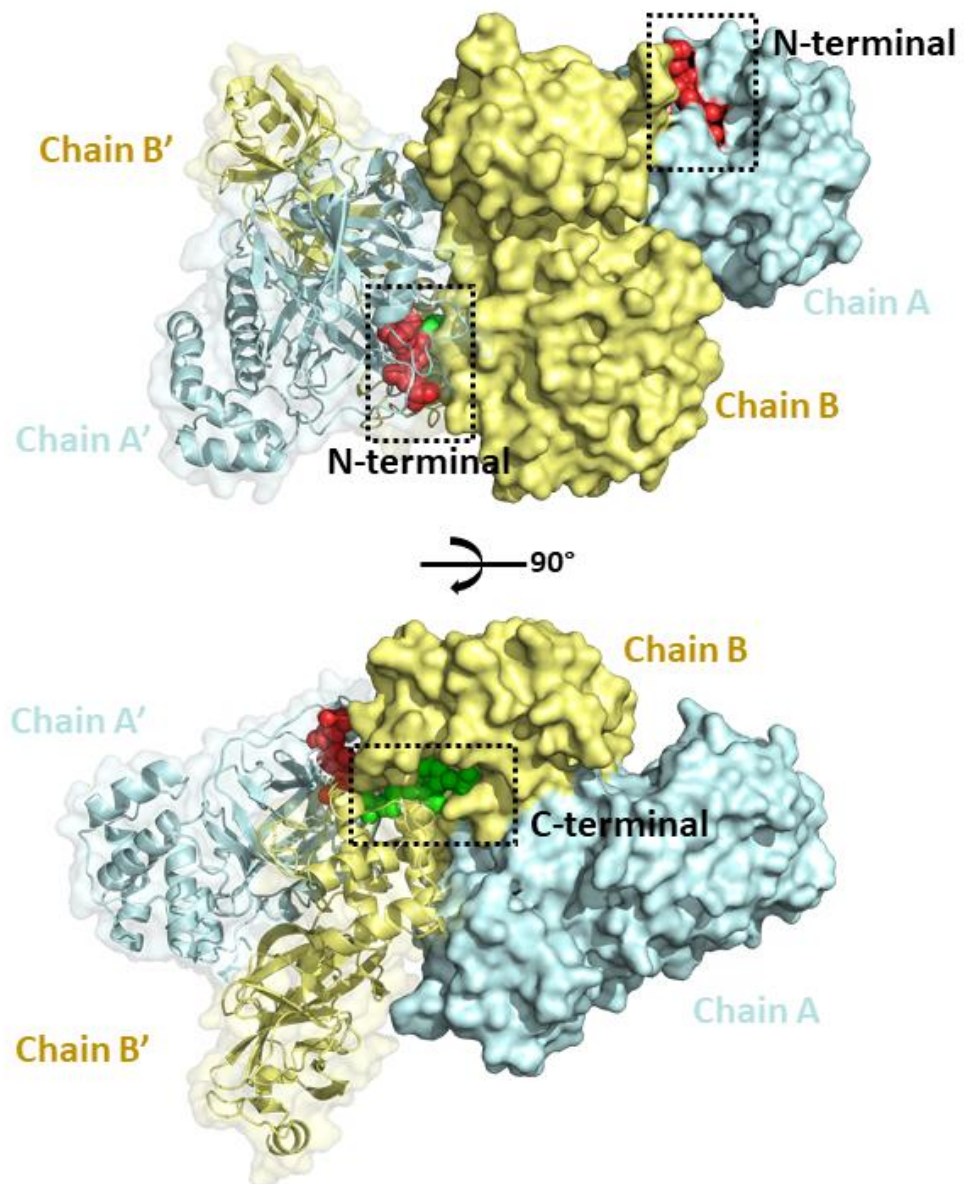
Em sua estreia, linha de cristalografia de proteínas analisou mais de 200 cristais de duas proteínas do novo coronavírus, expostos a pequenas moléculas que são partes de fármacos conhecidos. Esperativa é que, ao identificar essas estruturas, seja possível detectar substâncias que se encaixem perfeitamente nas proteínas, bloqueando sua ação no vírus (André Godoy e Aline Nakamura posicionam cristal de proteína de SARS-CoV-2 para análise no Sirius; foto: CNPEM/divulgação)



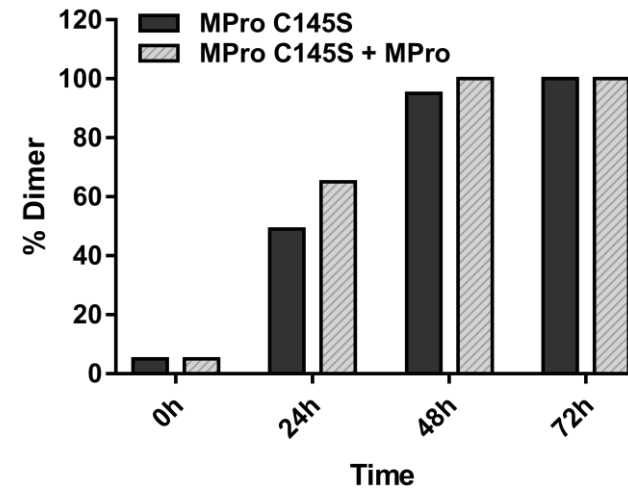
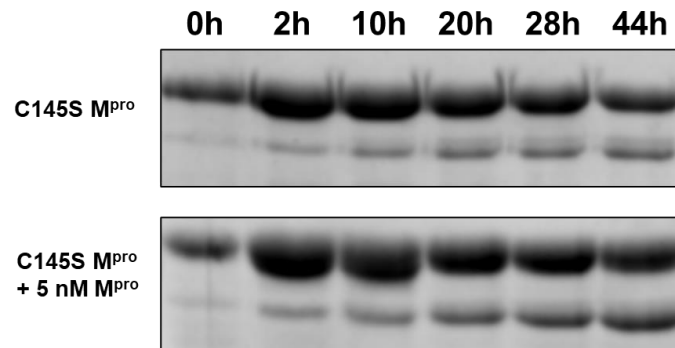
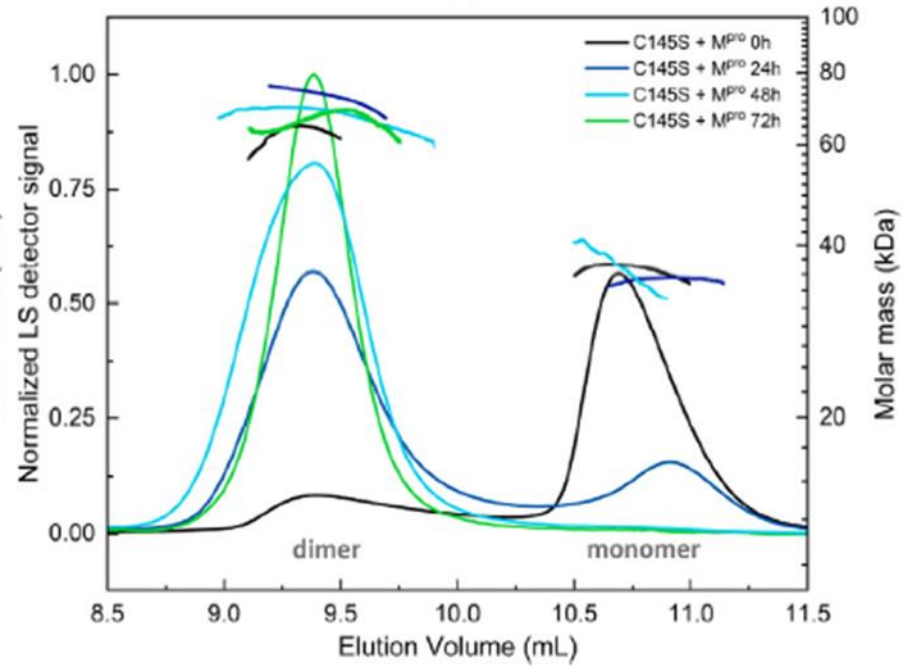
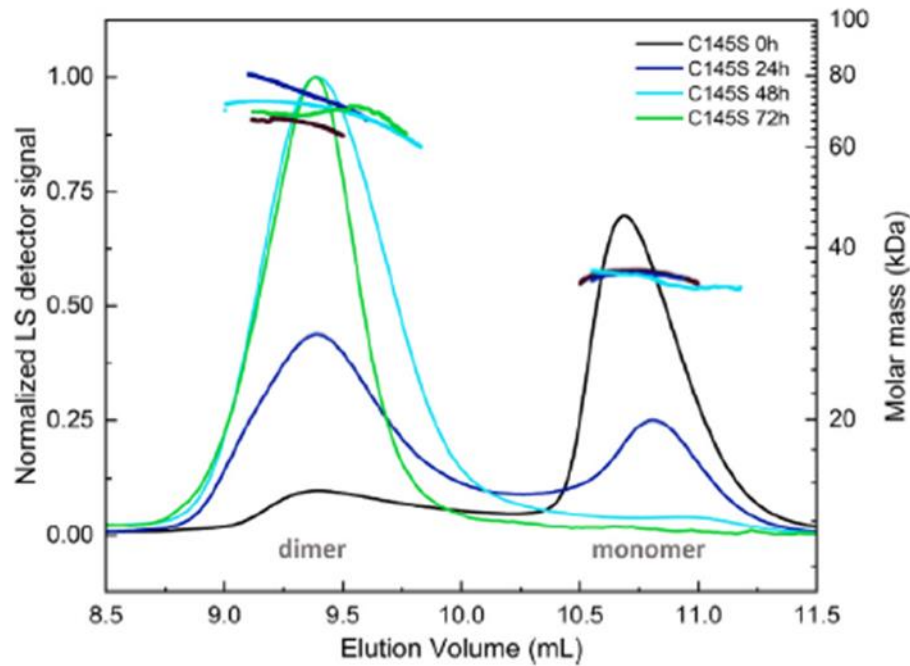
Crystal structure of IMT-M^{pro}



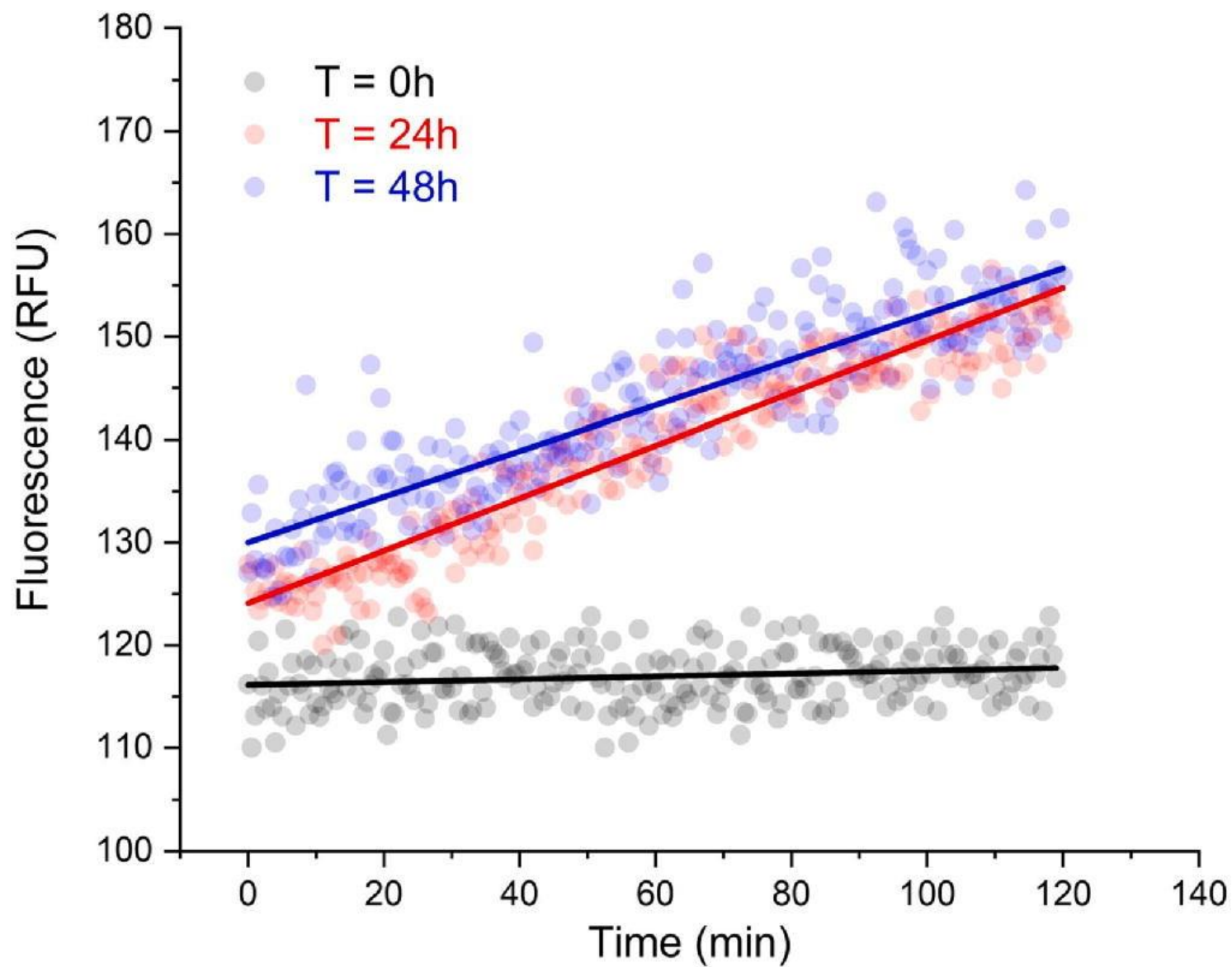
Crystal structure of C145S M^{pro}



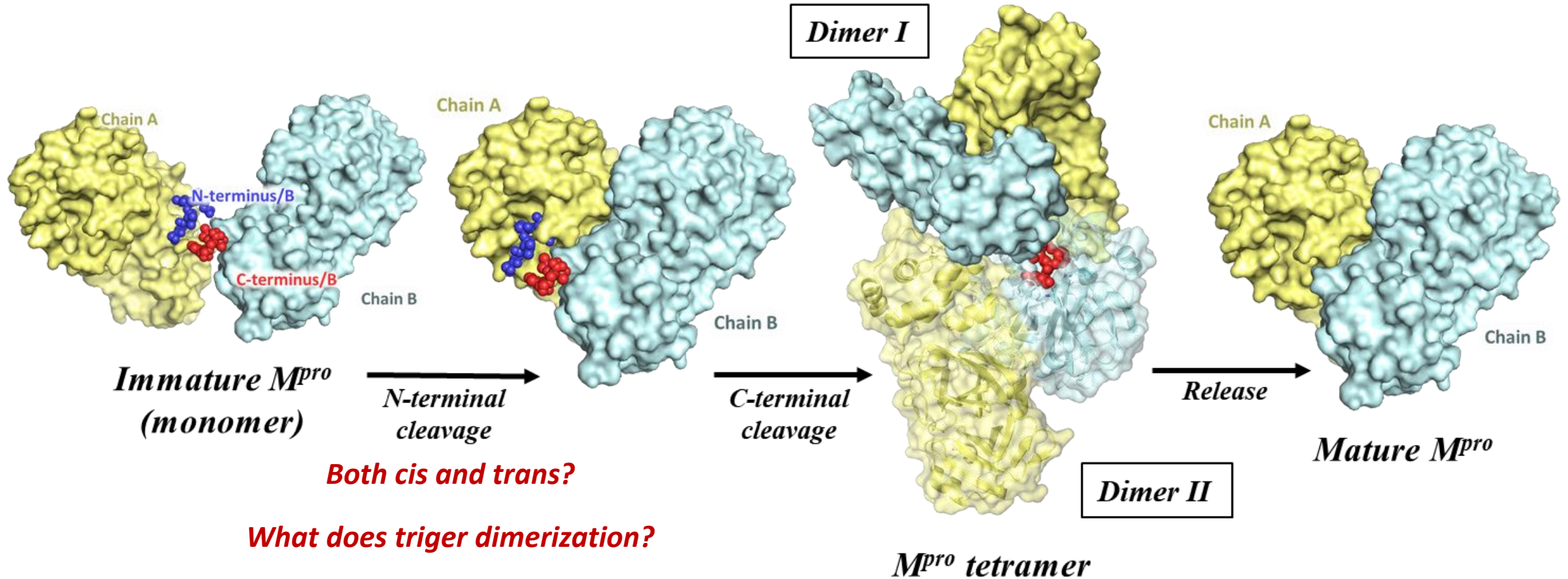
C145S M^{Pro} self-cleavage analysis



C145S M^{pro} self-cleavage analysis

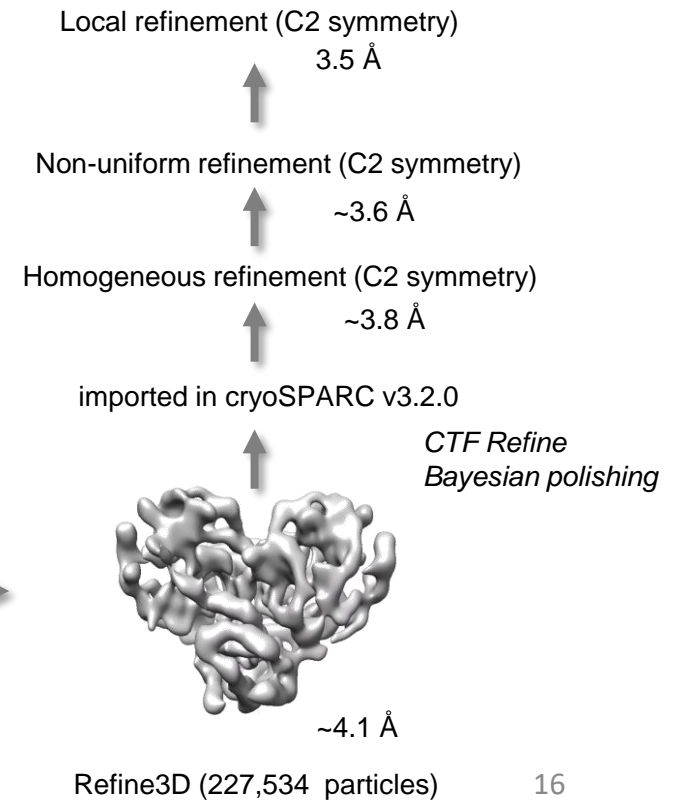
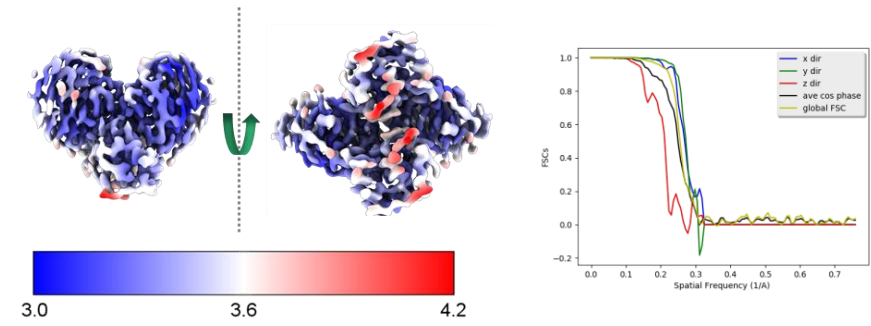
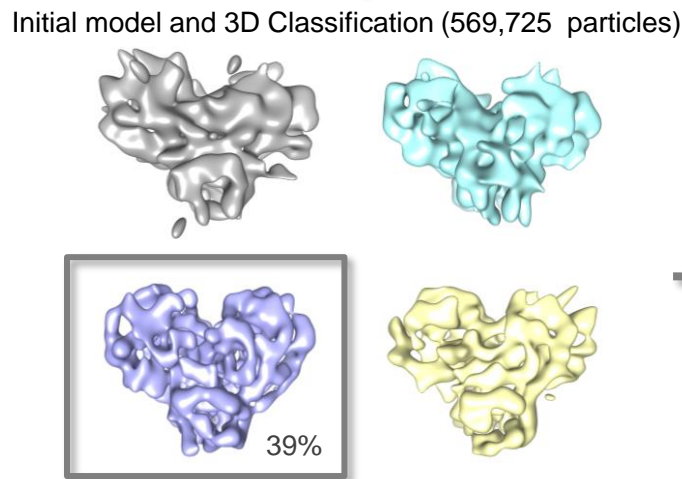
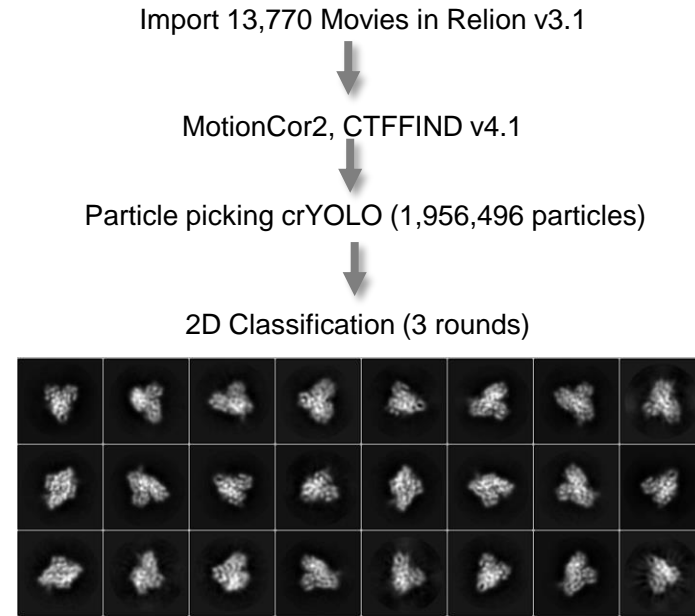
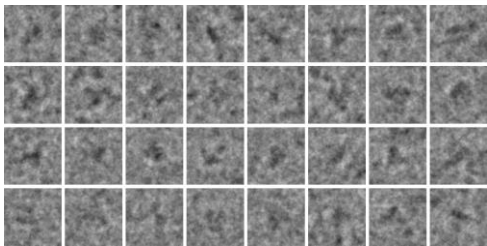
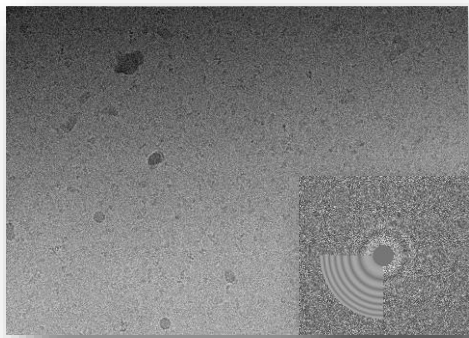
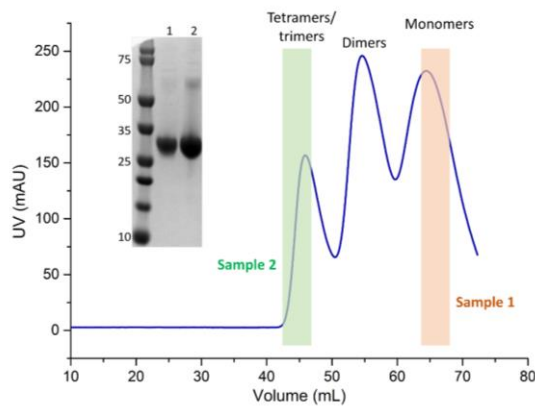


Model for M^{pro} maturation process

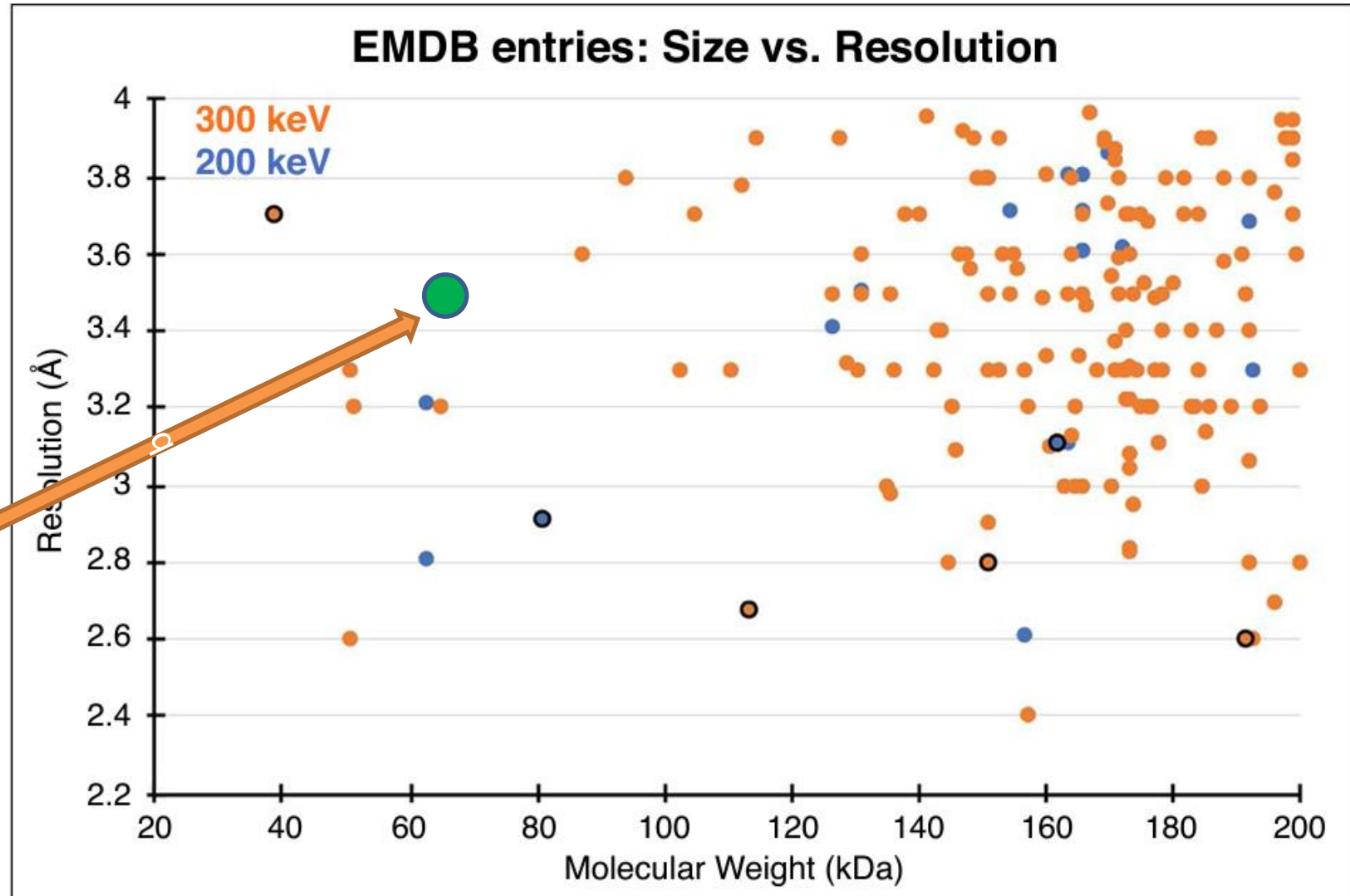


A Crystallographic Snapshot of SARS-CoV-2 Main Protease Maturation Process

Cryo-EM structure of C145S M^{pro}

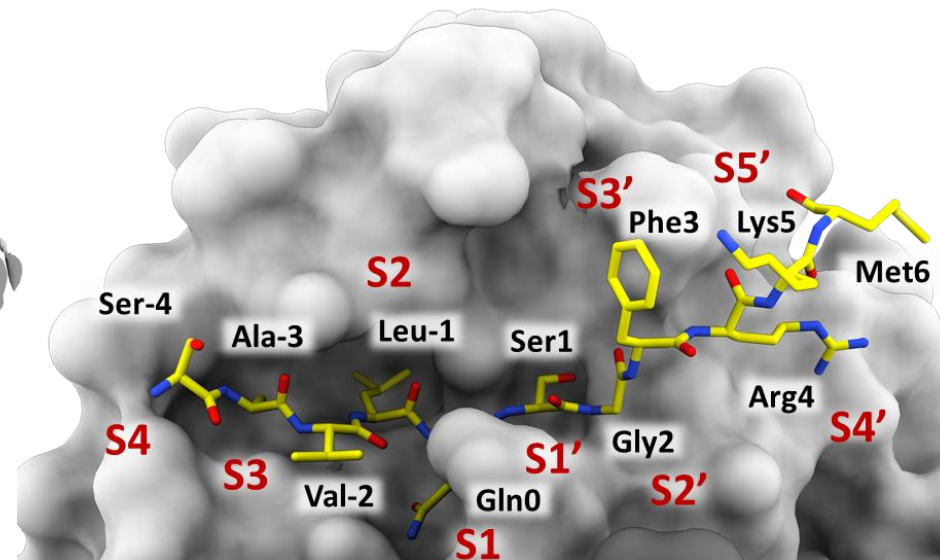
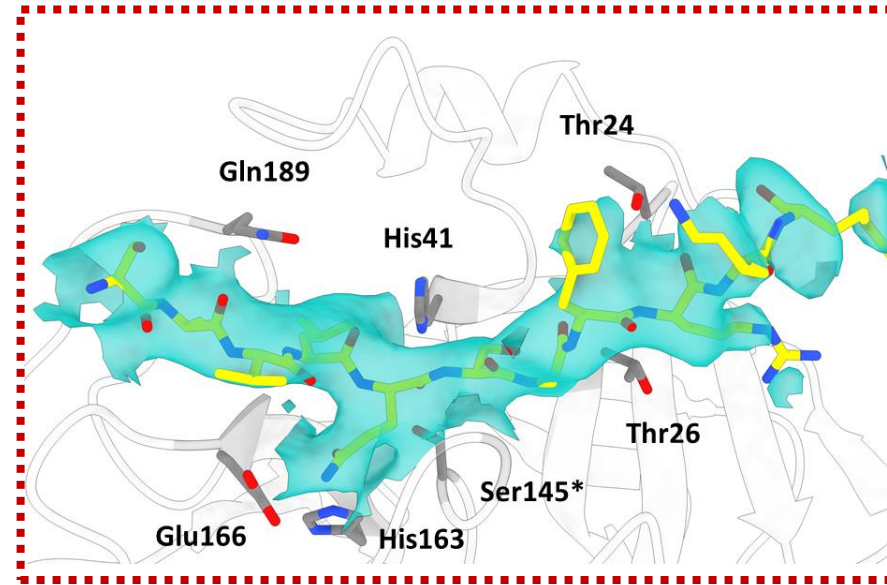
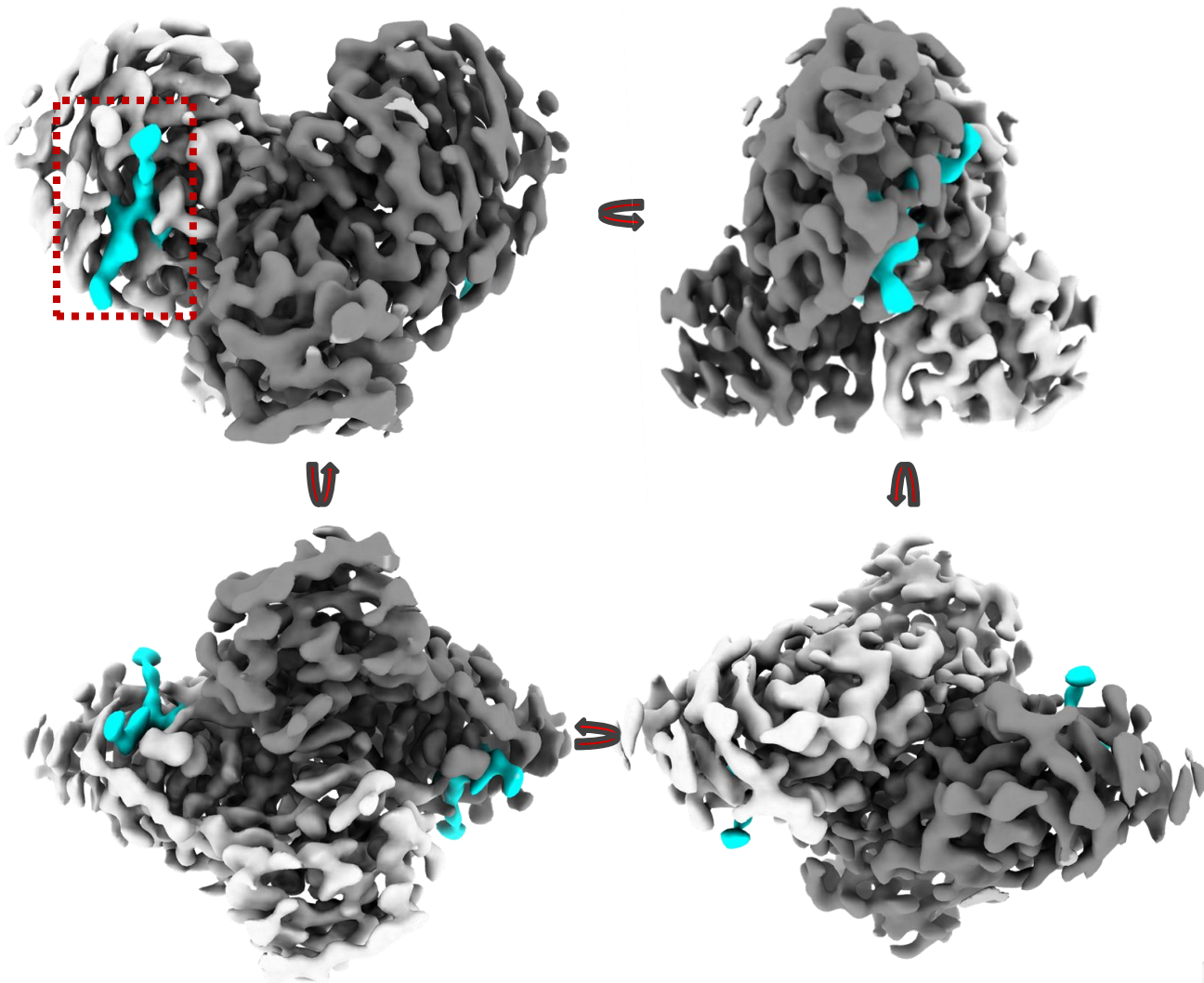


Cryo-EM of M^{pro} - Only 68 kDa at 3.5 Å !!

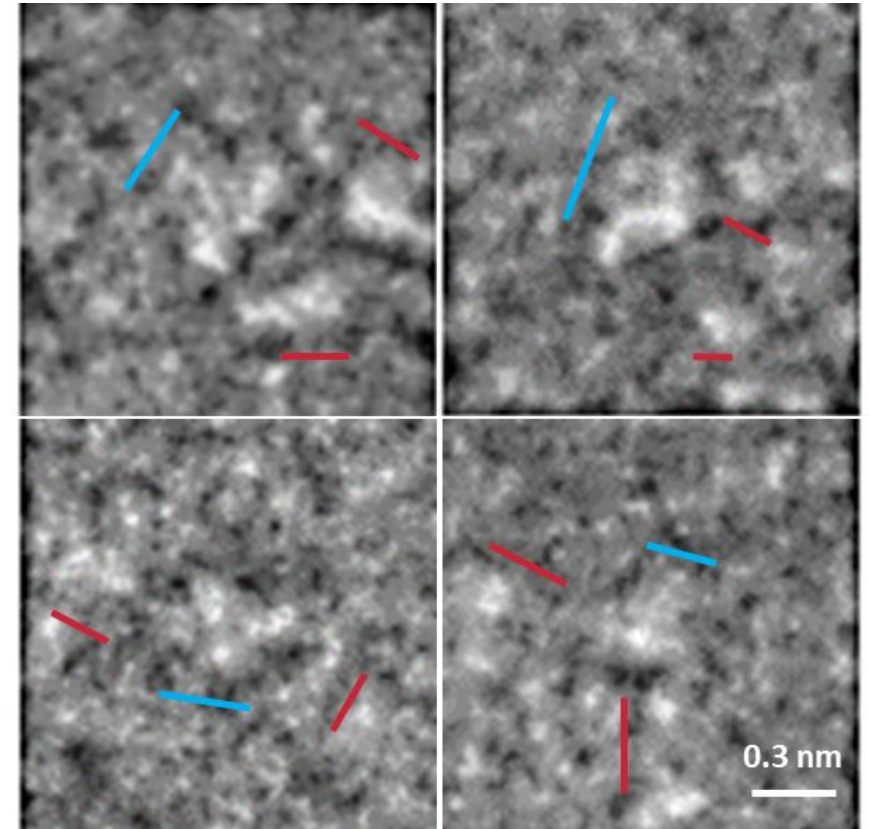
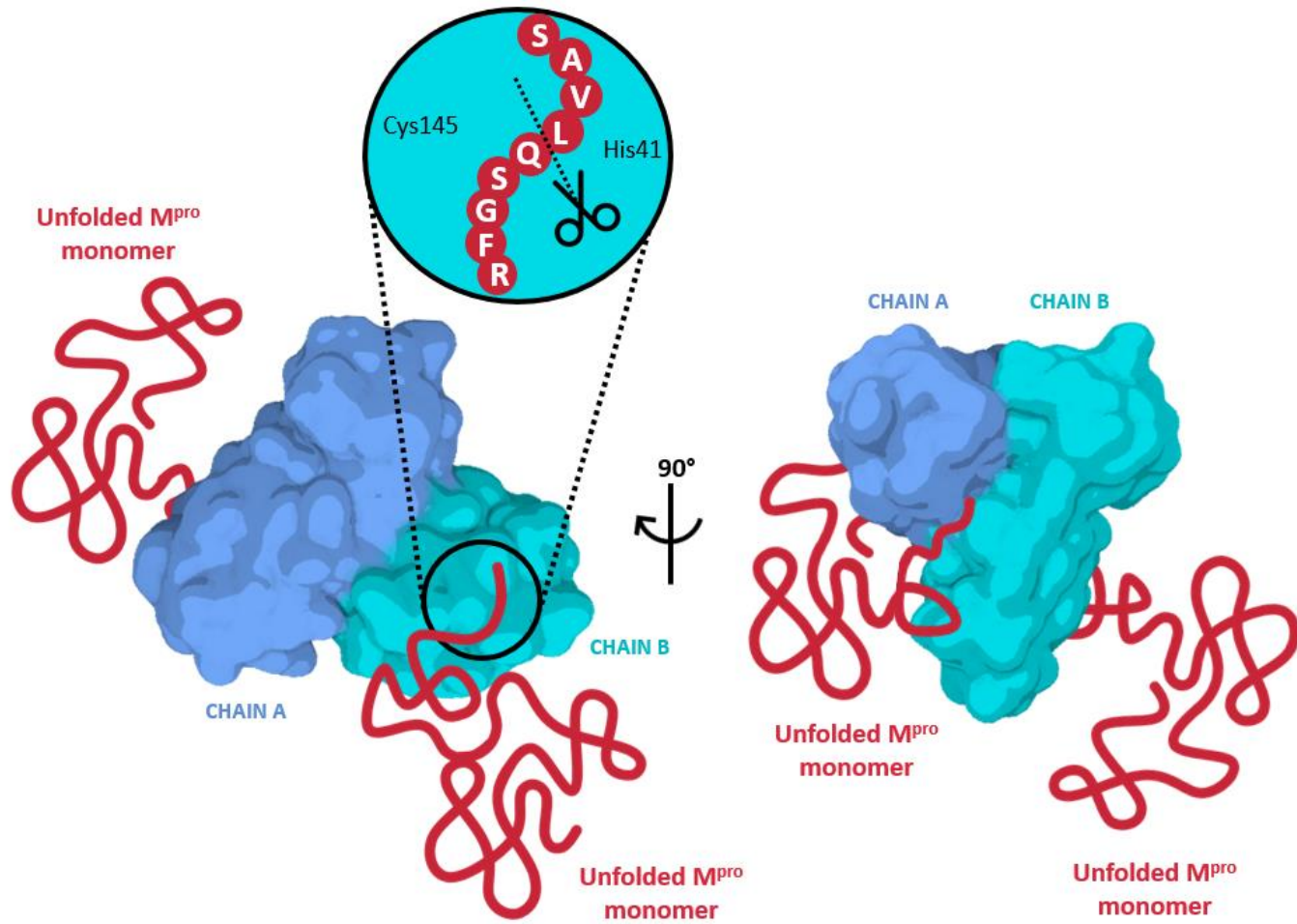


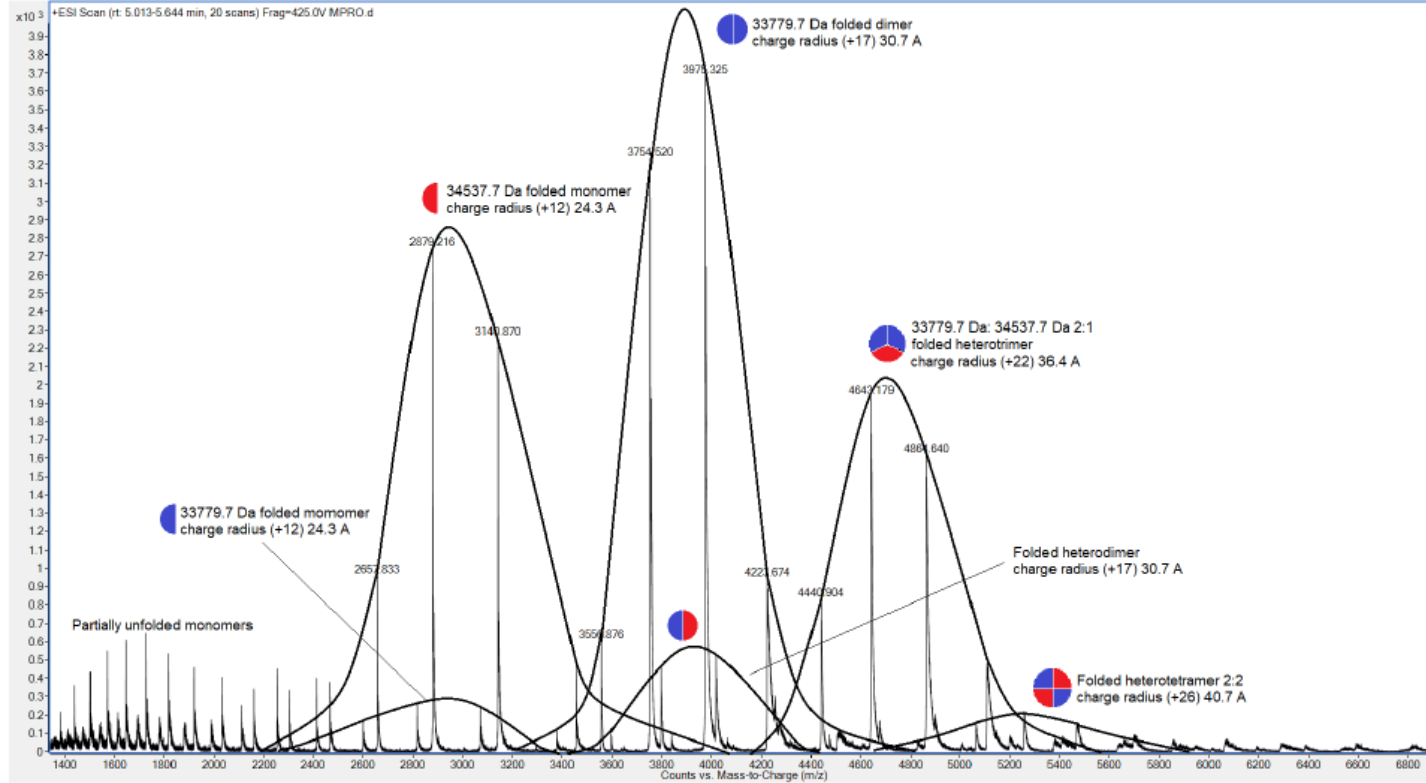
SARS-CoV-2 main protease@CIBFAR

Cryo-EM structure of C145S M^{pro}

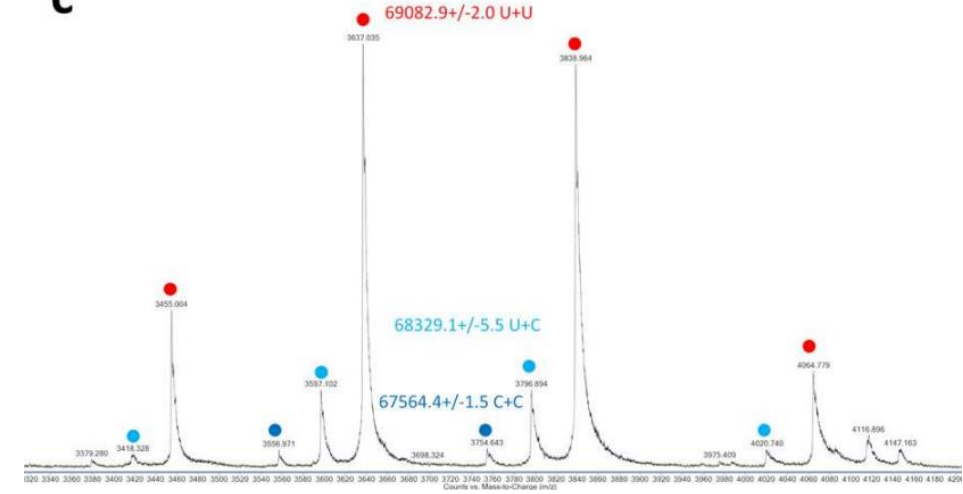


Cryo-EM structure of C145S M^{pro}

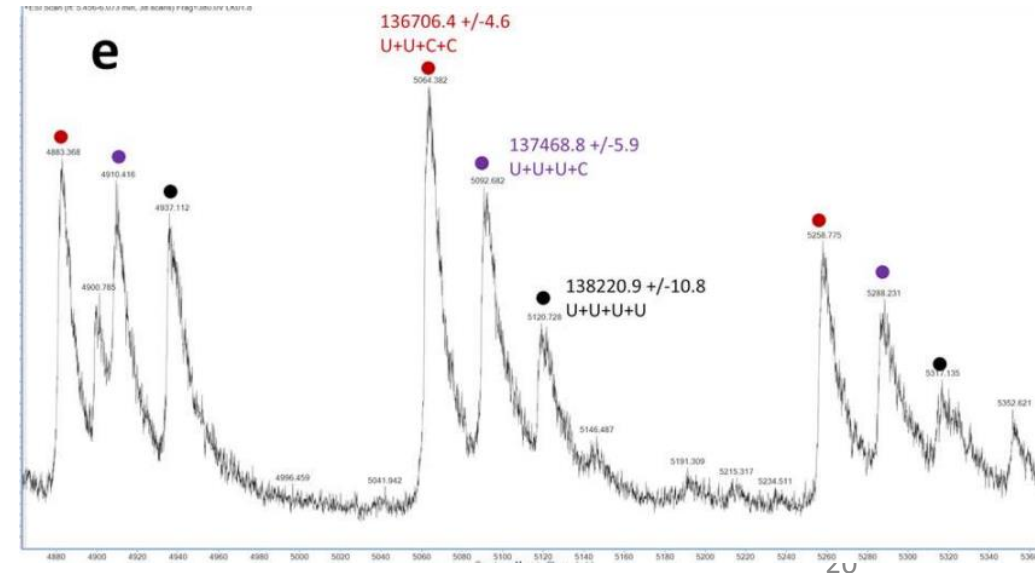


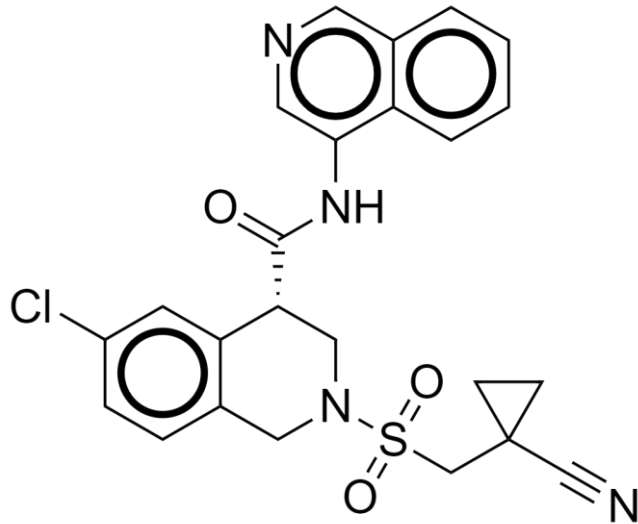


c



e



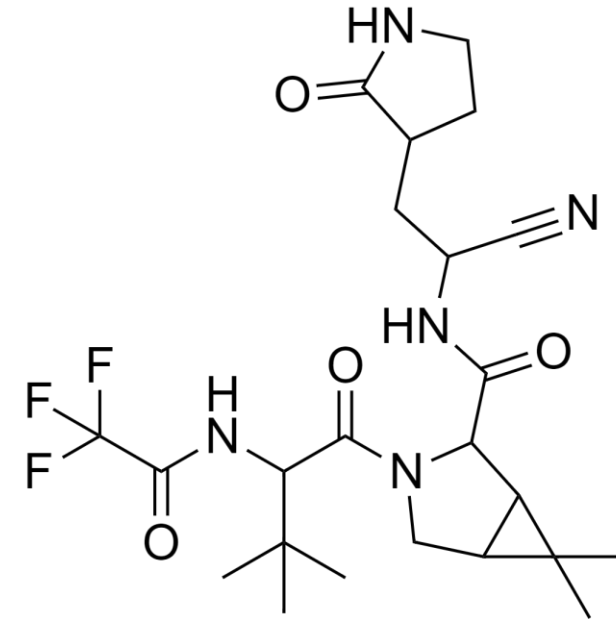


MAT-POS-e194df51-1

Developed by The Moonshot Consortium

pIC₅₀ 7.5

Non-covalent binding mode



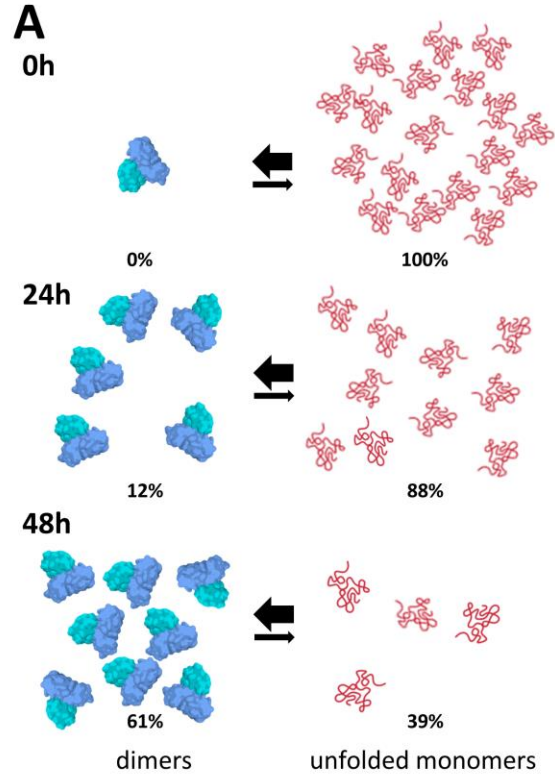
Nirmatrelvir

Developed by Pfizer

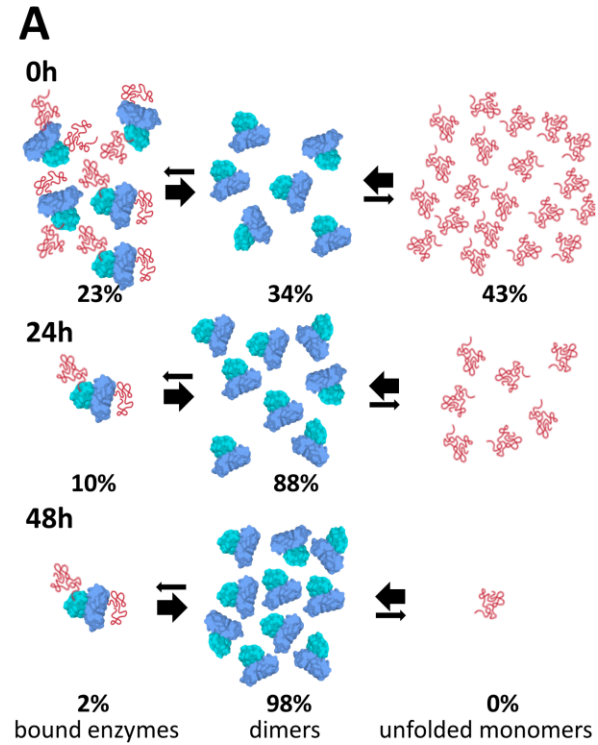
pIC₅₀ 7.7

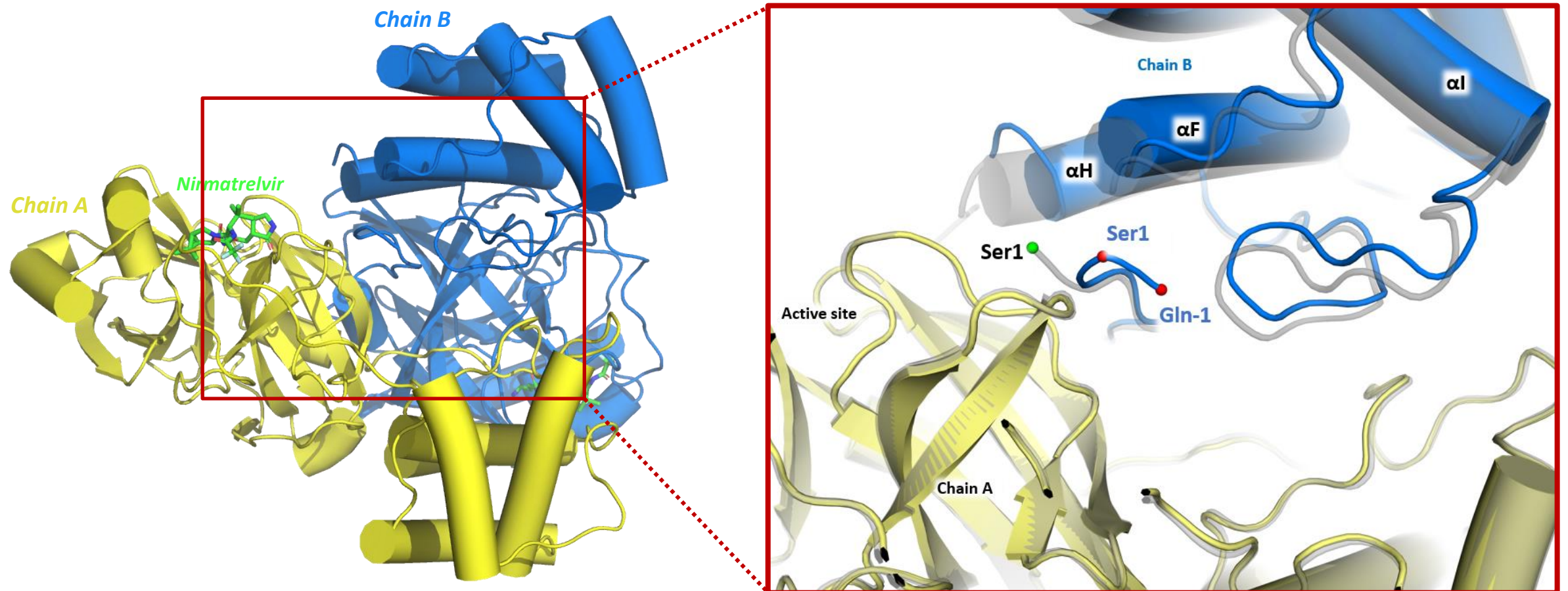
Covalent binding mode

C145S M^{pro} monomers

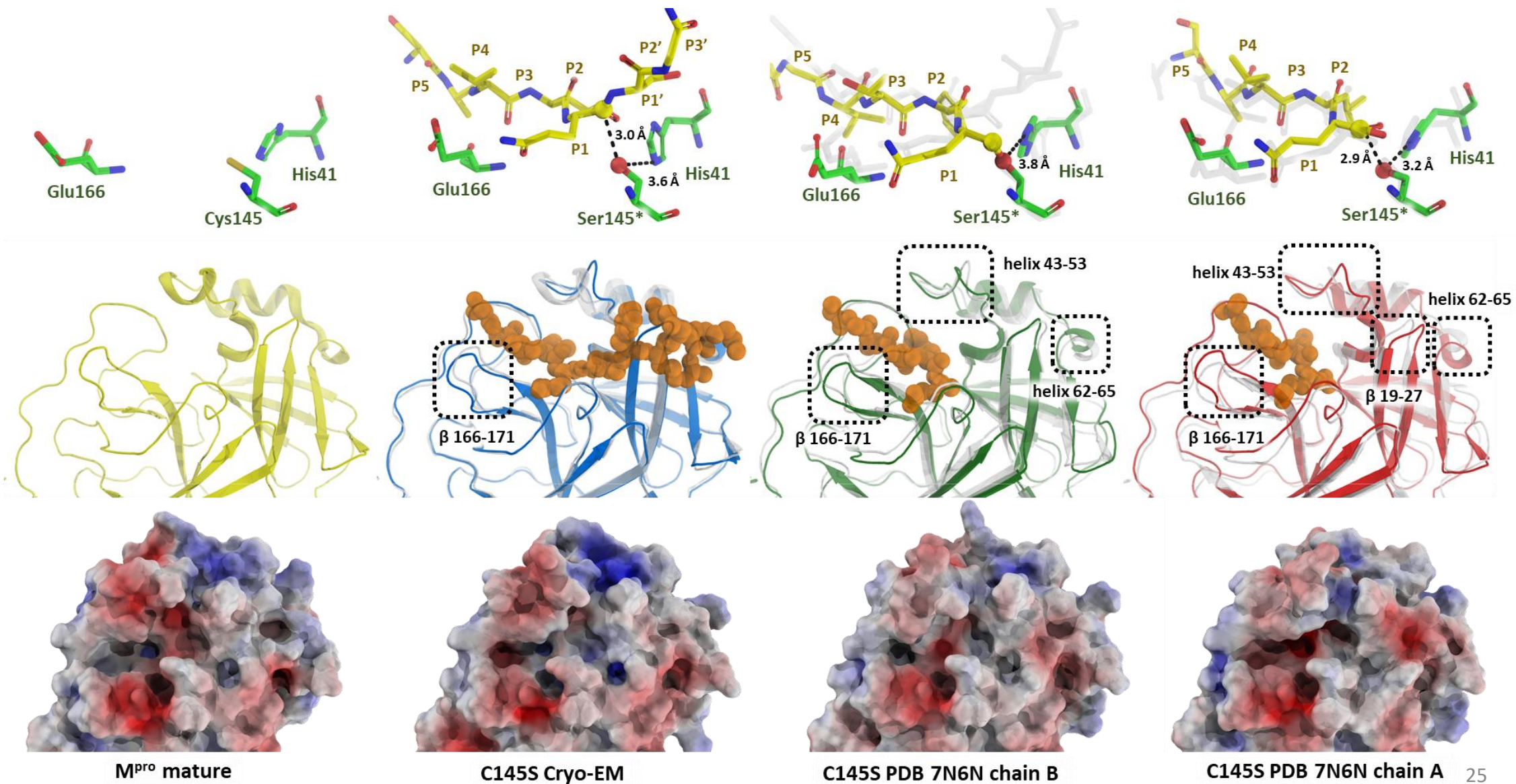


C145S M^{pro} tetramers

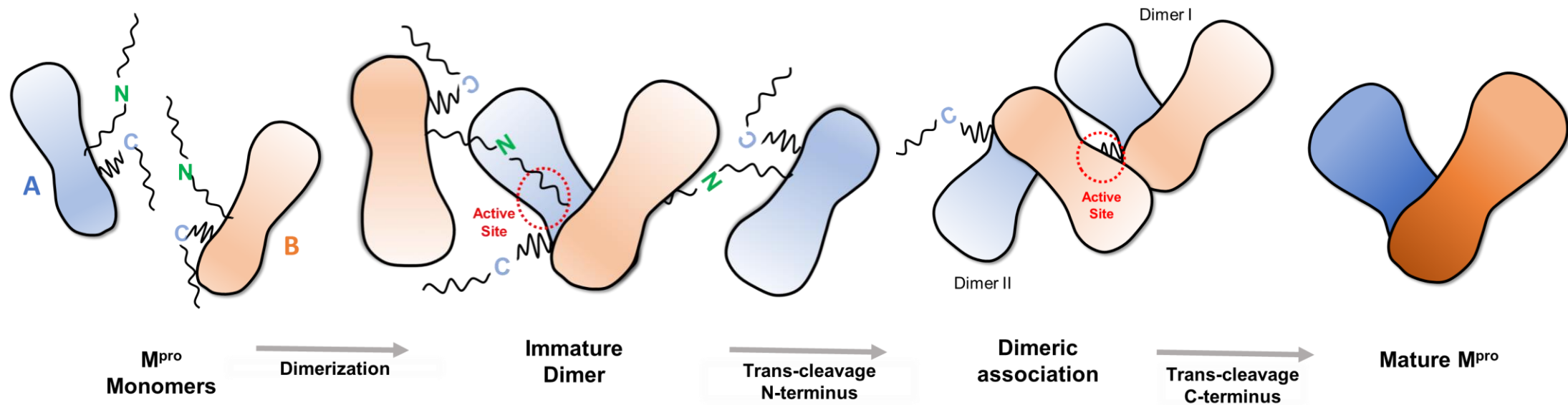




Overview of M^{Pro} cleavage process



- N-terminal cleavage is **NOT** critical for dimerization
- Dimerization is induced by covalent linkage
- Multiple oligomeric states can co-exist and act both cis and trans during maturation
- Structural information can guide the development of a new generation of M^{pro} inhibitors targeting intermediate steps of the maturation process










An in-solution snapshot of SARS-COV-2 main protease maturation process and inhibition

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 Check for updates

Gabriela Dias Noske^{1,6}, Yun Song ^{2,6}, Rafaela Sachetto Fernandes¹, Rod Chalk³, Haitem Elmassoudi³, Lizbé Koekemoer³, C. David Owen², Tarick J. El-Baba^{4,5}, Carol V. Robinson ^{4,5}, The COVID Moonshot Consortium^{*}, Glaucius Oliva ¹ & Andre Schutzer Godoy¹ 

The main protease from SARS-CoV-2 (M^{pro}) is responsible for cleavage of the viral polyprotein. M^{pro} self-processing is called maturation, and it is crucial for enzyme dimerization and activity. Here we use C145S M^{pro} to study the structure and dynamics of N-terminal cleavage in solution. Native mass spectroscopy analysis shows that mixed oligomeric states are composed of cleaved and uncleaved particles, indicating that N-terminal processing is not critical for dimerization. A 3.5 Å cryo-EM structure provides details of M^{pro} N-terminal cleavage outside the constraints of crystal environment. We show that different classes of inhibitors shift the balance between oligomeric states. While non-covalent inhibitor MAT-POS-e194df51-1 prevents dimerization, the covalent inhibitor nirmatrelvir induces the conversion of monomers into dimers, even with intact N-termini. Our data indicates that the M^{pro} dimerization is triggered by induced fit due to covalent linkage during substrate processing rather than the N-terminal processing.