

## **A vision for safer food contact materials: public health concerns as drivers for improved testing**

Jane Muncke<sup>a\*</sup>, Anna-Maria Andersson<sup>b</sup>, Thomas Backhaus<sup>c</sup>, Scott M. Belcher<sup>d</sup>, Justin M. Boucher<sup>a</sup>, Bethanie Carney Almroth<sup>c</sup>, Terrence J. Collins<sup>e</sup>, Birgit Geueke<sup>a</sup>, Ksenia J. Groh<sup>f</sup>, Jerrold J. Heindel<sup>g</sup>, Frank A. von Hippel<sup>h</sup>, Juliette Legler<sup>i</sup>, Maricel V. Maffini<sup>j</sup>, Olwenn V. Martin<sup>k</sup>, John Peterson Myers<sup>e,l</sup>, Angel Nadal<sup>m</sup>, Cristina Nerin<sup>n</sup>, Ana M. Soto<sup>o</sup>, Leonardo Trasande<sup>p</sup>, Laura N. Vandenberg<sup>q</sup>, Martin Wagner<sup>r</sup>, Lisa Zimmermann<sup>a</sup>, R. Thomas Zoeller<sup>q</sup> and Martin Scheringer<sup>s\*</sup>

*<sup>a</sup>Food Packaging Forum Foundation, Zurich, Switzerland; <sup>b</sup>Dept. of Growth and Reproduction, Rigshospitalet and Centre for Research and Research Training in Male Reproduction and Child Health (EDMaRC), Copenhagen University Hospital – Rigshospitalet, Copenhagen, Denmark, Copenhagen, Denmark; <sup>c</sup>Dept of Biological and Environmental Sciences, University of Gothenburg, Sweden; <sup>d</sup>Dept. of Biological Sciences, North Carolina State University, Raleigh, NC, USA; <sup>e</sup>Dept. of Chemistry, Carnegie Mellon University, PA, USA; <sup>f</sup>Department of Environmental Toxicology, Eawag, Swiss Federal Institute of Aquatic Science and Technology, Dübendorf, Switzerland; <sup>g</sup>Healthy Environment and Endocrine Disruptor Strategies, Commonweal, Durham, NC, USA; <sup>h</sup>Mel & Enid Zuckerman College of Public Health, University of Arizona, AZ, USA; <sup>i</sup>Dept. of Population Health Sciences, Faculty of Veterinary Medicine, University of Utrecht, Netherlands; <sup>j</sup>Independent consultant, Frederick, MD, USA; <sup>k</sup>Plastic Waste Innovation Hub, Department of Arts and Science, University College London, England, UK; <sup>l</sup>Environmental Health Sciences, Charlottesville, VA, USA; <sup>m</sup>IDiBE and CIBERDEM, Miguel Hernández University of Elche, Alicante, Spain; <sup>n</sup>Dept. of Analytical Chemistry, I3A, University of Zaragoza, Zaragoza, Spain; <sup>o</sup>Department of Immunology, Tufts University School of Medicine, Boston, MA, USA and Centre Cavailles, Ecole Normale Supérieure, Paris, France; <sup>p</sup>College of Global Public Health and Grossman School of Medicine and Wagner School of Public Service, New York University, New York, NY, USA; <sup>q</sup>Department of Environmental Health Sciences, School of Public Health & Health Sciences, University of Massachusetts Amherst, Amherst, MA, USA; <sup>r</sup>Dept. of Biology, Faculty of Natural Sciences, Norwegian University of Science and Technology, Trondheim, Norway; <sup>s</sup>Environmental Chemistry and Modelling, RECETOX, Masaryk University, Brno, Czech Republic and Department of Environmental System Sciences, ETH Zurich, Switzerland*

\*corresponding authors: [jane.muncke@fp-forum.org](mailto:jane.muncke@fp-forum.org); [scheringer@usys.ethz.ch](mailto:scheringer@usys.ethz.ch)

## **Abbreviations**

AOP Adverse Outcome Pathway

BPA bisphenol A

CVD Cardiovascular Disease

FCC Food Contact Chemical

NCD Non-Communicable Disease

NIAS Non-Intentionally Added Substance

PFAS Per- and polyfluoroalkyl Substances

PFOA perfluorooctanoic acid

SCOD Six Clusters of Disease

*manuscript under review*

## **A vision for safer food contact materials: public health concerns as drivers for improved testing**

Food contact materials and articles are ubiquitous in today's globalized food system. Chemicals migrate from food contact materials into foodstuffs, but current regulatory requirements do not sufficiently protect public health from hazardous food contact chemicals (FCCs) because only individual substances used to make food contact materials are tested and mostly only for genotoxicity while endocrine disruption and other hazard properties are disregarded. Indeed, food contact materials are a known source of a wide range of hazardous chemicals, and they likely contribute to highly prevalent non-communicable diseases. Food contact materials can also include non-intentionally added substances (NIAS), which often are unknown and therefore not subject to risk assessment. To address these important shortcomings, we outline how the safety of food contact materials may be improved by (1) testing the overall migrate, including (unknown) NIAS, and (2) expanding toxicological testing beyond genotoxicity to multiple endpoints associated with non-communicable diseases relevant to human health. To identify mechanistic endpoints for testing, we group chronic health outcomes associated with chemical exposure into Six Clusters of Disease (SCOD) and we propose that finished food contact materials should be tested for their impacts on these SCOD. Future research should focus on development of robust, relevant and sensitive *in vitro* assays based on mechanistic information linked to the SCOD, e.g., through Adverse Outcome Pathways (AOPs) or Key Characteristics of Toxicants. Implementing this vision will improve prevention of chronic diseases that are associated with hazardous chemical exposures, including from food contact materials.

**Keywords:** food packaging; risk assessment; chronic disease; chemical safety

## 1. Introduction

In today's globalized food system, food contact materials and articles such as food packaging, tableware, and food processing equipment are ubiquitous, especially those made of plastic (1, 2). This increases exposures to food contact chemicals (FCCs) migrating from food contact materials (3-5). This widespread, continuous exposure to a wide range of synthetic chemicals requires a more stringent safety assessment of food contact materials than the current approaches used in both low- and high-income countries (6-10).

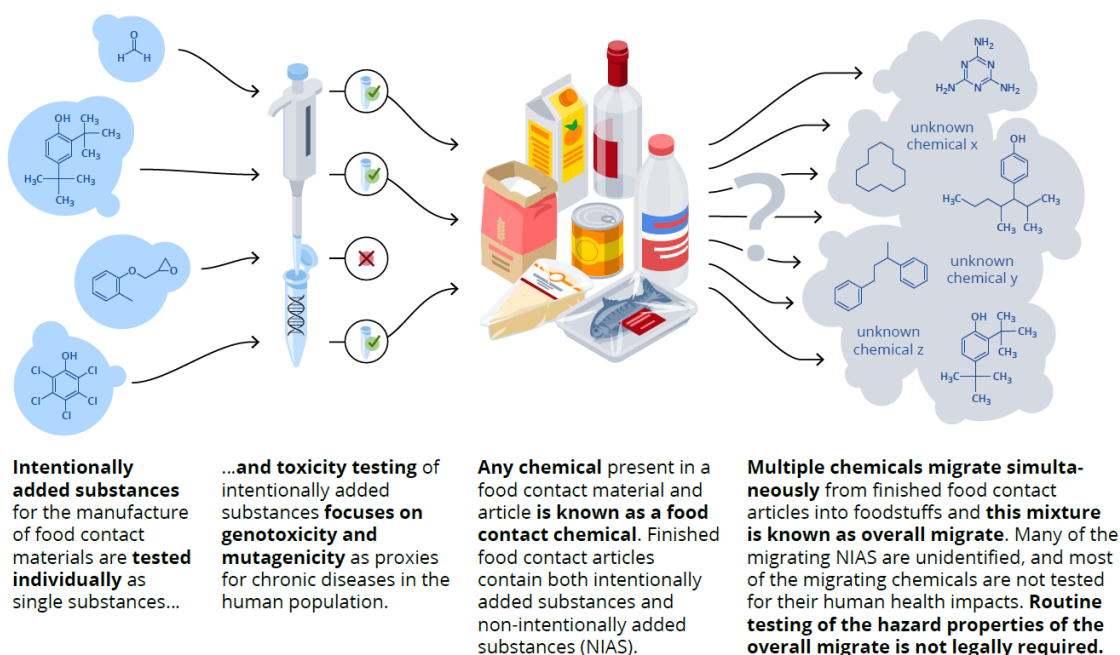
Food contact materials have been studied for over 50 years and are a known source of chemicals that migrate into foodstuffs (11-19). Numerous FCCs, either intentionally used in the manufacture of food contact materials or non-intentionally added substances (NIAS) that are present in the finished food contact material/article and that migrate into foodstuffs (5, 20, 21), are known to be hazardous and implicated with adverse human health impacts (22-29).

However, the current approach to chemical risk assessment for food contact materials is largely focused on assessing genotoxicity of single substances used to manufacture food contact materials and therefore fails to account for other highly relevant mechanisms of toxicity that are of equal concern as genotoxicity (10) and, what is more, the current approach does not assess NIAS that also migrate from food contact materials (Fig. 1) (17, 30). Addressing both issues is feasible in a cost-efficient way and necessary to protect public health.

Indeed, non-cancer non-communicable diseases (NCDs) of increasing prevalence in the global human population have been associated with several widely used FCCs, such as bisphenol A (BPA), bisphenol F, perchlorate, and di(2-ethyl hexyl) phthalate (DEHP), to name a few (Table 1). Given that humans are in daily contact with food contact materials, those materials are likely a relevant exposure source of hazardous chemicals that contribute to various NCDs globally.

In this article, we outline an improved assessment scheme for hazard identification of FCCs that captures all exposure-relevant chemicals including (unknown) NIAS, and we present a vision for assessing the safety of food contact materials that addresses biological effects linked to the most prevalent NCDs (31, 32). These include heart disease, stroke, cancer, diabetes, reproductive disorders, and several neurological conditions. We provide guidance on research and policy actions that should be developed to protect the public from avoidable chronic chemical exposures originating from food contact materials and articles.

## Current chemical risk assessment for food contact chemicals



**Figure 1.** Chemical risk assessment for food contact chemicals (FCCs): current practice. The current approach for assessing the safety of FCCs focuses on testing single substances that are intentionally used to make food contact materials. The toxicological focus is on mutagenicity and genotoxicity, therefore only carcinogenicity is currently determined as a human health relevant endpoint for predicting chronic disease. However, many more chemicals can migrate simultaneously from the finished food contact material, including unidentified compounds that are non-intentionally added substances (NIAS). The migrating mixture is known as the overall migrate, and it can also exert adverse effects (mixture toxicity). Currently, the assessment of overall migrate mixture toxicity is not legally required. Illustrator: Michael Stünzi.

## 2. Problem set-up: Shortcomings of the current approach

### 2.1 Non-communicable diseases are increasingly prevalent and associated with chemical exposures

NCDs are a significant contributor to global mortality (33). However, the impact of NCDs is far greater than mortality alone, especially in low- and middle-income countries. Both mortality and morbidity of selected NCDs have increased substantially over the last 30 years. Premature deaths (<70 years) are primarily associated with cardiovascular disease (17.7 million deaths per year, accounting for 45% of all NCD deaths), cancer (8.8 million deaths per year, 22% of all NCD deaths), chronic respiratory disease (3.9 million deaths per year, 10% of all NCD deaths) and diabetes (1.6 million deaths per year, 4% of all NCD deaths) (33). Expressed in Disability-Adjusted Life Years, cardiovascular diseases have increased by a factor of 1.4 from 1990 to 2017, neoplasms by a factor of 1.5, and diabetes, urogenital, blood and endocrine diseases by a factor of 1.6 (from 1990 to 2016) (34) (Figure S1, Supplemental Material). Furthermore, among reproductive-age women and men, infertility is now the most prevalent chronic disease (35). Importantly, NCDs incur significant human suffering in addition to their estimated economic costs (36-40), which further stresses

the need for urgent action towards prevention of morbidities associated with NCDs.

Chemical exposures are an important contributor to NCDs. Several well-studied types of chemicals such as toxic metals, halogenated aromatics, and some pesticides (41, 42), as well as some members of the endocrine disrupting compounds (43-47) are associated with NCDs such as brain-related disorders, cancers, metabolic disorders and cardiovascular disease. Specific FCCs such as BPA and several members of the ortho-phthalates group are associated with NCDs such as heart disease, diabetes, and some forms of cancer (48, 49) (Table 1). Further, the effects of chemical exposures on risk of NCDs are complex and multifaceted, with some outcomes occurring across generations through transgenerational inheritance (47, 50, 51). It is also clear that these effects are not limited to laboratory animals, as mixtures of chemicals including FCCs have been associated with adverse health outcomes in prenatally exposed humans (46, 49, 52-55).

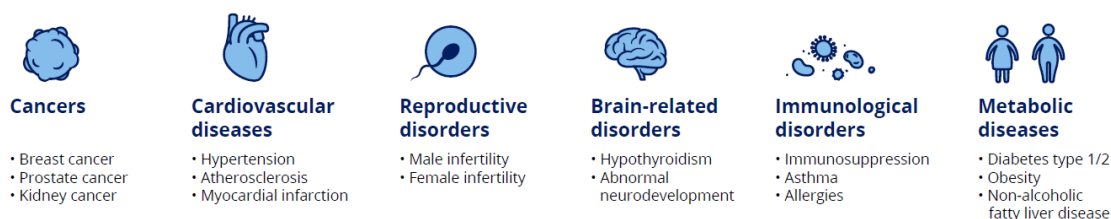
**Table 1.** Food contact chemicals (FCCs) associated with non-communicable diseases (NCDs) from each of the Six Clusters of Disease (SCOD) (non-exhaustive and non-systematic overview). Identification of FCCs was based on the Food Contact Chemicals database (FCCdb) (24) and the database on migrating and extractable food contact chemicals (FCCmigex) (17). This overview is not a complete list of FCCs that are associated with adverse health outcomes. Systematic reviews are indicated with\*. Cancer agents are classified by cancer site (125).

Disease Cluster	Example disease	Associated FCC exposure	References
<b>Cancers</b>	Testicular cancer	PFOA	(229, 230)
	Kidney cancer	PFOA	(229, 231)
	Breast cancer	PFOA	(232)
		Ortho-phthalates	(232)
<b>Cardiovascular diseases</b>	Cardiovascular diseases: including myocardial infarction, arrhythmias, dilated cardiomyopathy, atherosclerosis, and hypertension	BPA	(233-236)
		Ortho-phthalates	(237)
<b>Brain-related disorders</b>	Hypothyroid	BPA	(238)
		Ortho-phthalates	(239)
		Perchlorate	(239)
		PFAS	(240)
	Abnormal neurodevelopment	Ortho-phthalates: DEHP, DBP, BBP and DEP	(241)
	Attention Deficit Hyperactivity Disorder/behavior	Lead, BPA, ortho-phthalates	(242-244)
	Lower Intelligence Quotient	Endocrine disrupting chemical (EDC) mixture (Ortho-phthalates)	(46, 245)
Language delay	EDC mixture	(55)	
<b>Metabolic and endocrine diseases</b>	Type-1 diabetes	BPA, Ortho-phthalates, PFAS	(246)

	Type-2 diabetes	BPA	(247-249)
		PFOA	(250)
	Pre-diabetes and diabetes	Ortho-phthalates	(241, 251, 252)
	Obesity (BMI, waist circumference)	BPA	(237, 253, 254)
			(255, 256)
	Childhood Obesity	PFAS	
		BPA	(257)
			(258)
	Gestational diabetes	Ortho-phthalates	
		Antimony	(259)
			(260)
	Non-alcoholic fatty liver disease	Ortho-phthalates	
		EDC mixture	(261)
			(262)
		PFAS	
<b>Immunological disorders</b>	Immunosuppression	PFAS: PFOS and PFOA	(263)
	Childhood asthma	Ortho-phthalates: DEHP and BBzP	(241)
	Kidney damage	Melamine	(264)
<b>Reproductive disorders</b>	Male infertility	BPA	(265)
		Dibutyl phthalate	(266)
	Semen quality	Ortho-phthalates: DBP, BBP, DEHP, and DINP	(241, 251, 267)
	Female infertility (reduced follicular count)	DEHP	(268)

NCDs that are increasingly prevalent in the human population and that are associated with hazardous chemical exposures can be grouped into disease clusters. On this basis, we developed the novel concept of Six Clusters of Disease (SCOD) (Figure 2). The six clusters are cancers, cardiovascular diseases, reproductive disorders, brain-related disorders, immunological disorders, and metabolic diseases. The SCOD concept provides a rationale for systematically assessing the safety of chemicals in food contact materials, with a focus on the prevention of chemical-associated, highly prevalent, and severe NCDs. As such, the SCOD concept expands current efforts for chemical risk assessment of FCCs.

## Six Clusters of Disease



**Figure 2.** The novel Six Clusters of Disease (SCOD) concept comprises non-communicable diseases (NCDs) that are highly prevalent in the global human population, of increasing concern, and associated with hazardous chemical exposures that can be clustered by disease type. They include cancers, cardiovascular diseases, reproductive disorders, brain disorders, immunological disorders, and metabolic diseases. The SCOD are of major concern for public health and require novel approaches for prevention, namely the identification of chemical contributors. Chemical risk assessment of food contact chemicals (FCCs) should determine contributions to diseases of public health concern. Preventing exposure to chemicals in food contact materials that contribute to NCDs is critical for successful primary prevention strategies. Illustrator: Michael Stünzi.

## 2.2 Current risk assessment of food contact chemicals is not sufficiently protective of human health

The universe of known FCCs comprises at least 14,153 substances, and for at least 1,518 FCCs empirical evidence for migration from food contact articles and materials is publicly available (17). Evidence of human exposure exists for hundreds of these chemicals (4, 55-66). At least 388 FCCs in use today are known to be carcinogenic, mutagenic or toxic to reproduction, possess endocrine disrupting properties, or have other properties of concern such as persistence (22).

Currently, in the United States (US), Canada, the European Union (EU), China and other countries, chemical risk assessment is required for all migrating substances (Figure 1). In practice, however, it is predominantly the intentionally used substances that are assessed for their risk to human health. Humans are exposed to many more FCCs that are non-intentionally added to the finished food contact material or foodstuff. These NIAS include impurities of the starting substances, reaction by-products, or degradation products of starting substances (like additives) (5, 67-69). NIAS most often are unidentified, they are common in food contact materials with high chemical complexity, and they are likely to be biologically active (70). Under the current



chemicals risk assessment paradigm for food contact materials, where a chemical's identity must be known, unidentified FCCs cannot be assessed, although, for example, the EU plastic food contact regulation requires the risk assessment of NIAS (71), and also US FDA's Food Contact Notification has information requirements on impurities and reaction by-products (72).

A second problem is the lack of testing of substances present in the finished food contact material. Several approaches have been developed to approximate the health risks of unknown NIAS (73-79), but these approaches contain substantial uncertainties related to hazard estimation, chemical identification, and quantification (80, 81) because they are based on assumptions that cannot be entirely supported by empirical evidence. For example, generic thresholds for chronic exposures to nongenotoxic carcinogens were derived from testing chemicals at maximum tolerable doses (MTD) and at 1/2 MTD, but it depends on the exact mechanism by which a chemical exerts its toxicity whether a low-dose extrapolation from MTD dosing is appropriate or not (82, 83).

Finally, because some laws prohibit the use of chemicals that cause cancer in humans or animals, testing methods currently focus on genotoxicity as a proxy for predicting cancer risk (10, 84). This focus on genotoxic effects is at the expense of other hazards, including outcomes relevant to other chronic NCDs. Thus, there is a need for novel and more robust approaches to more fully evaluate all the relevant hazards to human health associated with FCCs.

### **3. Our vision: to make safer food contact materials**

#### ***3.1 Assessing toxicological effects relevant to the Six Clusters of Disease***

Chronic exposure to hazardous chemicals is a known modifiable risk factor for cancer and reducing exposure to hazardous or untested chemicals from consumer products, including food contact materials, is a recommended preventive measure (85). It is reasonable to assume that the same holds true for other NCDs that are associated with chemical exposures, especially for endocrine disrupting chemicals (Table 1). Indeed, exposure reductions can lower the incidence of disease (86), for example for neurodevelopmental disorders (87), obesity (88) or male reproductive disorders (89).

NCDs that are increasingly prevalent in the human population and that are associated with hazardous chemical exposures can be grouped into disease clusters. On this basis, we have developed the novel concept of SCOD (Fig. 2). The SCOD concept emerged from discussions with the Food Packaging Forum's Scientific Advisory Board (SAB) during several meetings between 2016 and 2022. The SCOD concept provides for the first time a rationale for systematically assessing the safety of chemicals in food contact materials, with a focus on the prevention of chemical-associated, highly prevalent and severe NCDs. As such, the SCOD concept expands current efforts for chemical risk assessment of FCCs beyond cancers induced via a genotoxic mechanism. For each disease cluster within the SCOD, many widely used FCCs have been associated with relevant diseases in both epidemiology and animal studies (Table 1). For some, mechanistic evidence strengthens these associations. It is also this mechanistic evidence that provides opportunities to use *in silico* and *in vitro* assays to better map toxicity profiles of individual FCCs in finished food contact materials,

before they are placed on the market, as well as mixtures, extracts and migrates from food contact materials and articles. The SCOD provides organizing principles for such an approach.

### ***3.2 Assessing real-life chemical exposures: testing overall migrate from food contact materials***

All FCCs that are relevant for human exposure should be tested, in other words, FCCs used in the manufacturing of food contact materials should be tested as single substances, and the real-life mixture of all migrating FCCs, the *overall migrate*, should also be tested. In addition, the overall migrate should be subjected to non-targeted chemical analyses that are aimed to identify its chemical composition, including NIAS (90). This combined testing and chemical identification approach could inform the development of safer food contact materials by selecting less hazardous ingredients and developing manufacturing processes that generate fewer and less biologically active NIAS. Such an approach would be aligned with the proposed Safe and sustainable by Design criteria included in the EU's Chemicals Strategy for Sustainability (91).

The already available as well as emerging *in vitro* assays provide an opportunity to identify hazardous properties of single substances and of the overall migrate. *In vitro* test systems are small-scale, often single-cell or small organism systems, for example human cancer cell lines, bacteria, and fungi (e.g. yeast). Other high-throughput screening assays utilize embryos and larvae from vertebrates such as zebrafish (*Danio rerio*) or African clawed frog (*Xenopus laevis*). These assays can be performed efficiently both in terms of time and cost and are usually based on mechanistic pathways (92, 93).

Test batteries, where several relevant assays are combined simultaneously, can also be operated as high-throughput screening methods such as those developed in Tox21 and ToxCast (94-96), which demonstrate the feasibility of this approach. In this way, diverse information about the interaction properties of a single chemical with different biological systems can be generated efficiently, and with lower cost, compared to whole-animal testing used in traditional toxicology.

These assessments should be guided by the SCOD concept. However, gaps exist in the current understanding of molecular pathways related to the SCOD, and these *in vitro* assays remain insufficient to identify the full panoply of potential hazards, especially those mediated by endocrine mechanisms. *In vitro* assays included in high-throughput test batteries need to be appropriate for predicting relevant human health outcomes; should be demonstrated to be reproducible, sufficiently specific and sensitive; and must be executed transparently (97, 98). Because of the limited *in vitro* assays for known pathways and mechanisms of action associated with endocrine disruption and other complex biological cascades, animal testing needs to continue, but at a reduced level than in the past. For example, no current *in vitro* approaches would have revealed what is now known to be a feature of some chemical exposures, e.g., transgenerational epigenetic inheritance (99). Acknowledging these and other gaps, the European Commission is funding EURION, a program to develop new testing and screening methods (including many *in vitro* approaches) for identifying endocrine disrupting chemicals (100).

### ***3.3 Shifting from the status quo to a more comprehensive approach to testing***

Within the SCOD, increasingly available mechanistic information enables an understanding of how chemicals contribute to highly prevalent NCDs. Two emerging frameworks are being implemented to describe how chemicals affect complex diseases and to provide a more uniform approach to evaluating mechanistic evidence: the key characteristics concept, and adverse outcome pathways (AOPs). Both offer opportunities to shift from the status quo, modernize hazard assessments, and develop suitable *in vitro* assays.

#### ***3.3.1 The Key Characteristics concept: modernizing chemical hazard assessments***

The key characteristics concept makes use of information about the properties of hazardous chemicals that have empirical evidence linking them causally to relevant apical (disease) endpoints (101). The underlying premise is that chemicals that cause the same disease outcomes in whole organisms share molecular properties (i.e., key characteristics) that are relevant for their hazardous properties. The key characteristics for different disease outcomes are hence defined using empirical evidence for well-characterized chemicals, combined from epidemiological, *in vivo* and mechanistic studies. These disease-specific key characteristics can then be used to develop mechanistic *in vitro* assays to screen chemicals for their propensity to contribute to different disease clusters and thereby reduce the need for *in vivo* experiments while still decreasing scientific uncertainty normally associated with *in vitro* data.

The key characteristics were first developed for carcinogens, drawing from existing mechanistic information from thoroughly assessed chemicals that are known to be carcinogenic in humans (101-105). Additional key characteristics of other disease-causing chemicals have also been described, such as for hepatotoxicants (106), endocrine disrupting chemicals (107), female reproductive toxicants (108), male reproductive toxicants (109), cardiovascular toxicants (110), and immunotoxicants (111). For metabolic toxicants and neurotoxicants, work to describe key characteristics is ongoing. Taken together, the key characteristics approach provides an excellent starting point for the mechanistic understanding of how certain chemicals are associated with NCDs, such as those covered in the SCOD.

#### ***3.3.2 Using other mechanistic information to develop suitable in vitro assays***

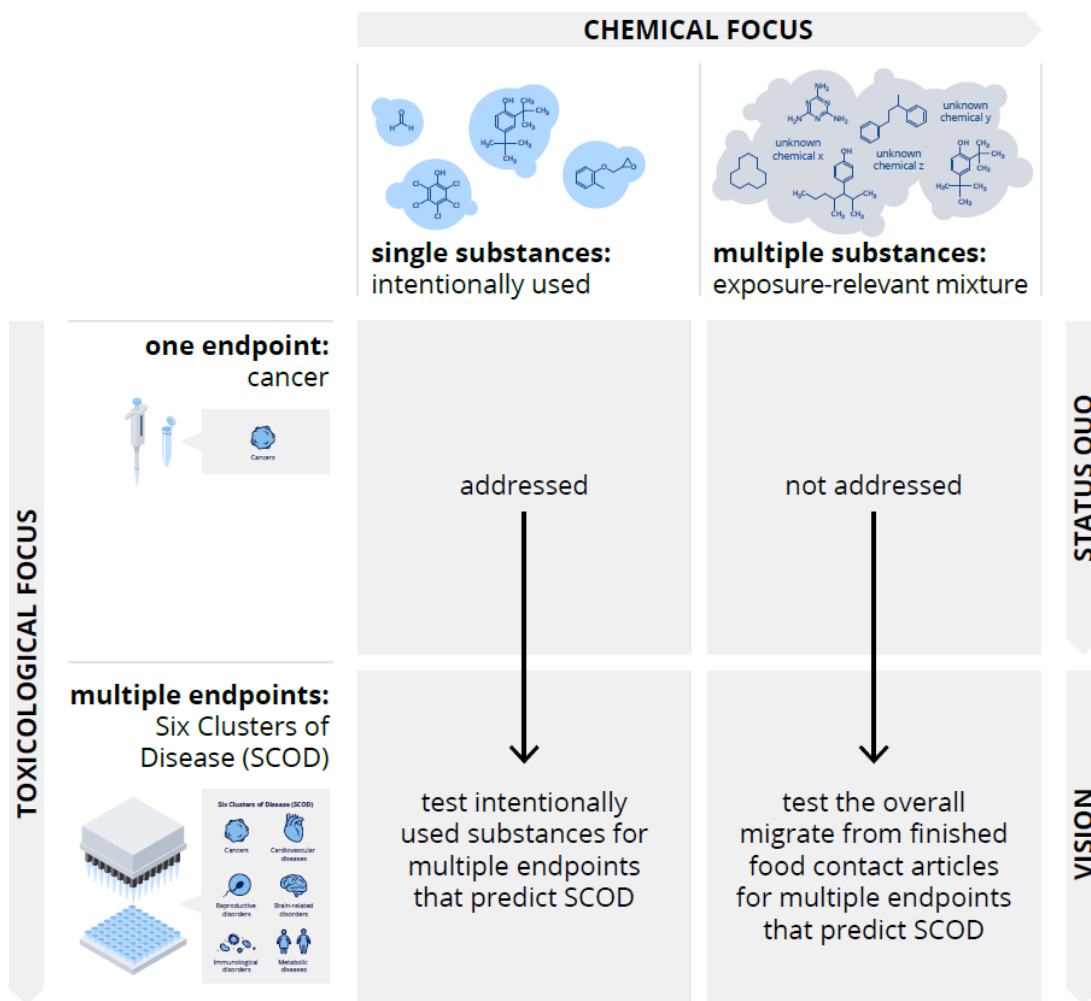
In addition to the key characteristics, further important mechanistic understanding is becoming available and can be useful to inform development of dedicated *in vitro* screening assays for hazard assessments of FCCs. Chemicals exert toxic effects by combinations of many different molecular-level events. These mechanistic events leading to apical endpoints of toxicity can be organized in an AOP (112). Several AOPs relevant to NCDs in the SCOD have been proposed, such as estrogen receptor activation leading to breast cancer (113) and the upregulation of thyroid hormone catabolism (via activation of hepatic nuclear receptors) leading to subsequent adverse neurodevelopmental outcomes in mammals, specifically the loss of cochlear function (114). Thus, AOPs are an emerging approach to organize mechanistic information so that molecular or cellular-level targets can be identified for developing *in vitro* assays that are relevant to the SCOD.

#### ***3.3.3 The novel approach: A vision for safer food contact materials***

Based on the presumption that mechanistic *in vitro* testing of chemicals supports the prevention of NCDs within the SCOD, we propose a novel approach for testing FCCs that

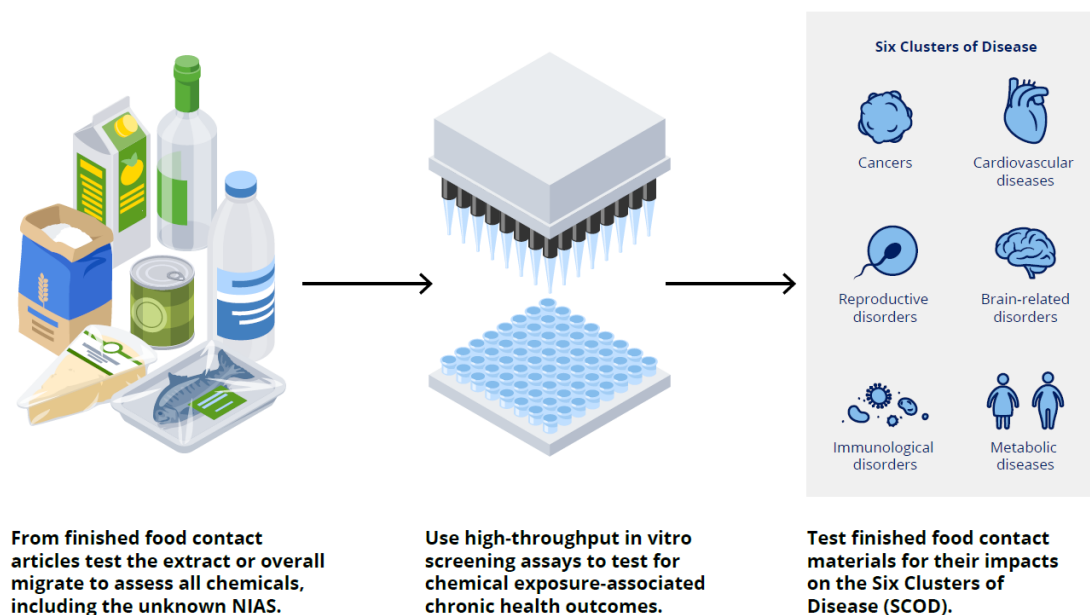
- (1) covers individual FCCs as well as real-life mixtures, migrating (or extractable) from finished food contact materials, including all known and unknown NIAS,
- (2) assesses the health impacts of FCCs and real-life mixtures with respect to the most prevalent NCDs in the human population, and
- (3) evaluates effects that are upstream from the disease, relying on mechanistic information and *in vitro* screening approaches (wherever possible) to accurately predict health effects induced by FCCs and migrates.

This shift from current practice to the proposed approach is summarized in Fig. 3, and a detailed overview is provided in Fig. 4. Our approach overcomes the most challenging shortcomings of the current testing paradigm of chemical hazard assessment of food contact materials, fully recognizing that to assess all adverse effects of chemicals on biological systems, adequate *in vivo* testing is required, where additional aspects would be addressed such as metabolic activation, unknown modes of action leading to apical endpoints, and transgenerational effects. However, we also realize that such extensive, multigeneration *in vivo* testing may not always be feasible for various reasons, including ethical and practical ones. Therefore, we propose this vision to improve FCC testing from the currently too limited scope towards a much more comprehensive yet feasible approach that holds promise for better protection of public health.



**Figure 3.** Schematic overview of the current vs. proposed approach to food contact chemical (FCC) testing. Currently, single substances intentionally used to make food contact materials are tested for genotoxicity using *in vitro* assays. The proposed new approach focuses on testing the overall migrate (i.e., the human exposure-relevant mixture of all migrating FCCs) for its potential to contribute to the Six Clusters of Disease (SCOD). Notably, single substances used to make food contact materials would also be tested individually for the SCOD-relevant endpoints and, if found to have biological activity, excluded from use in the manufacturing of food contact materials. Illustrator: Michael Stünzi.

## Testing finished food contact materials for their impacts on the Six Clusters of Disease (SCOD).



**Figure 4.** The vision for a novel approach to safety assessment of food contact materials and articles. Finished food contact materials and articles are tested for their real-life mixture of all migrating chemicals (the overall migrate), using *in vitro* screening assays as well as non-targeted chemical analyses. The screening assays are mechanism-based and identify the key characteristics, key initiating events, or other mechanisms of action of the overall migrate. Screening assays are selected around the Six Clusters of Disease (SCOD) concept. In addition, intentionally added substances used for the manufacture of food contact materials are also tested as individual substances prior to their authorization, and the overall migrate is chemically characterized using non-targeted approaches. Illustrator: Michael Stünzi.

### 4. Implementing the vision: assessing impacts of FCCs and relevant mixtures on human health outcomes in the SCOD using mechanistic approaches

Here we review the mechanistic basis for each of the disease clusters included in the SCOD, and selectively highlight available *in vitro* testing methods. Importantly, some available assays cover key characteristics that are relevant for several disease clusters.

This vision for expanded hazard assessment of food contact materials is based on the finding that for each of the disease clusters included in the SCOD, some mechanistic understanding is available for the way that chemicals cause disease (Table 2).

#### 4.1 Cancer

As defined by Willis,

A neoplasm is an abnormal mass of tissue, the growth of which exceeds and is

uncoordinated with that of the normal tissues and persists in the same excessive manner after cessation of the stimulus which evoked the change (115).

Regarding cancer causation, the somatic mutation theory posits that cancer is a cellular disease caused by mutations of genes that disrupt the control of cell proliferation. Yet, substantive contradictions exist between this theory and empirical evidence (116), which inspired competing theories consider cancer as a problem of tissue organization akin to organogenesis (117-119). Importantly, not all carcinogens are mutagens (120) and, thus, carcinogenicity cannot be equated with genotoxicity. Yet, because legal requirements restrict the use of cancer-causing agents in food contact materials, testing of FCCs has focused on genotoxicity as a proxy to identify carcinogenic substances.

Both carcinogens and mutagens are found in food contact materials including 1) formaldehyde, a known human carcinogen (121), which migrates from various plastics including melamine-formaldehyde plastics used as tableware for children, and polyethylene terephthalate plastic (PET) (122, 123); 2) antimony trioxide, which “is reasonably anticipated to be a human carcinogen” (124) and “probably carcinogenic to humans” (125), and it is used in the manufacture of PET, where antimony is found to migrate into soft drinks (123, 126); and 3) per- and polyfluoroalkyl substances (PFAS) are widely used in the manufacture of food contact materials as processing aids in plastic and paper food contact material production (127, 128), and perfluorooctanoic acid has limited evidence for testicular and kidney cancers in humans (129).

The key characteristics for carcinogens reveal that these chemicals can be mutagens, but that there are numerous other common features for these agents as well (101-105). Guyton and Schubauer-Berigan (2021) recommended the use of *in vitro* assays based on the key characteristics to identify carcinogens in high-throughput screening (105). Further, Rider et al. (2021) proposed methods to use the key characteristics to test chemical mixtures and their propensity to affect cancer development including in mixtures of chemicals with different key characteristics of carcinogens (130). Approaches such as these will provide important information for testing mixtures such as the overall migrate from finished food contact materials.

Methods for evaluating genotoxicity are readily available, validated, and trusted. Chemicals are considered genotoxic if they damage the structure, information content, or segregation of DNA, with mutagenicity (i.e. changes to the nucleotide sequence) being a sub-type of genotoxicity (131).

These methods include:

- **Mutagenicity:** The Ames test, based on bacterial reverse mutagenicity, is the most employed test for mutagenicity (Organisation for Economic Co-operation and Development (OECD) test guideline (TG) 471). A mammalian cell (mouse lymphoma) gene mutation test (OECD TG 490) is also available (132)
- **Chromosomal aberration:** Cultured mammalian cells are assessed for the presence of chromatid-type and chromosome-type aberrations during metaphase (OECD TG 473)
- **Micronucleus:** Micronuclei represent chromosomal damage (chromosome fragments or whole chromosomes) that have been transmitted to daughter cells. Micronuclei can be assessed *in vitro* by using mammalian cells (OECD TG 487)

or *in vivo* with erythrocytes collected from bone marrow or peripheral blood (OECD TG 874)

These methods are recommended or required for assessing intentionally used FCCs (72, 133). Several other *in vitro* assays for assessing the genotoxic potential of FCCs are also available (134). However, these strategies have not kept pace with discoveries in cancer biology (135). Currently, no *in vitro* assays are available that capture features of carcinogenicity beyond genotoxicity, but research is underway to address this technical gap (136). On the other hand, the causal role of the microenvironment in carcinogenicity, as put forward by tissue-based theories on carcinogenicity (118), is not captured by such *in vitro* assays, because the reciprocal interactions between stroma and parenchyma during development, regeneration, and remodeling are not being considered (137). Although *in vivo* assays involving mammals are available, traditional 2-year rodent carcinogenicity studies (OECD TG 451), either alone or in combination with chronic toxicity studies, are rarely performed for FCCs.

#### **4.2 Cardiovascular diseases**

Cardiovascular diseases (CVDs) are a group of disorders arising due to dysfunction of the heart and blood vessels. The most recognized forms of CVD, coronary heart disease and cerebrovascular disease, result in damage to tissues caused by limited or complete loss of blood supply (138).

FCCs including several phthalates and bisphenols contribute to the causation of CVDs, independent of obesity and diabetes (110). Bisphenols can disrupt calcium signalling in myocardium and vasculature; and phthalates and bisphenols are oxidant stressors that accelerate coronary and other arterial inflammation (110). In the US alone, 100,000 premature deaths from CVD among 55–64-year-olds each year are attributed to exposure to one phthalate, DEHP (139). Other FCCs, such as antimony, may also impair cardiovascular function and accelerate CVDs (140).

Lind et al. (2021) compiled the key characteristics of cardiovascular toxicants and provided a comprehensive overview of robust and sensitive *in vitro*, *ex vivo* and *in vivo* assays that are available for measuring dysregulation of  $\text{Ca}^{2+}$  ion homeostasis and resulting arrhythmogenic activities of chemicals. For example, the increased risk for CVDs associated with higher exposures to BPA is mechanistically associated with  $\text{Ca}^{2+}$  release and reuptake resulting in proarrhythmic delays after depolarizations in isolated cardiomyocytes. BPA promotes  $\text{Ca}^{2+}$ -mediated arrhythmias *ex vivo* in the whole heart of rats and mice (141). However, this is only one of many possible mechanisms for inducing CVDs, and further assay development is required.

Although several FCCs have been associated with CVDs, cardiovascular toxicity is generally not evaluated for FCCs, whether they are intentionally used to make food contact materials or NIAS present in finished food contact materials. This is in part due to a reliance on *in vivo* guideline testing of general toxicity for chemicals migrating at very high levels and limited to assessment of neoplastic and non-neoplastic cardiac lesions in rodent models, which can be confounded by a high incidence of background pathology in many of the rodent strains used for toxicity testing (142). However, these are insensitive apical endpoints that only identify highly cardiotoxic chemicals that result in robust pathology but miss subtle molecular effects (143, 144).



We recommend that comprehensive testing for all new chemicals include *in vitro* and *in silico* testing harmonized with the Comprehensive *in vitro* Proarrhythmia Assay approach (145, 146).

### **4.3 Brain-based disorders**

Disrupted neurodevelopment can have numerous consequences including a lower intelligence quotient, delayed language acquisition, ADHD, and autism (53, 55, 147). Because the role of thyroid hormone in brain development is well established, hypothyroidism, especially during early development, is also a condition of concern upstream of neurodevelopmental disorders. Neurotoxicity can also result from impaired neuronal function due to a variety of factors, such as neuronal misplacement during development, altered synapses, hypomyelin, or degeneration. Other neurodegenerative conditions that typically arise later in life include Parkinson's disease, Alzheimer's disease, and other forms of dementia.

The role of FCCs in the causation of many brain-based disorders is well established, with substantial contribution to the burden of disease for both neurodevelopmental and neurodegenerative disorders (37). For example, FCCs that interfere with thyroid hormone systems or sex steroids (e.g., phthalates and perchlorate) can affect brain development as well as cognitive function in adults (87, 148). The vulnerability of the developing brain and the lack of systematic assessment of neurodevelopmental toxicity for FCCs raises serious concerns (149). At present, the key characteristics of neurotoxicants remain undescribed, but relevant work is ongoing.

In addition to assays covering interference with the thyroid and sex steroid axes, *in vitro* testing of neurotoxicants requires sophisticated and reliable models due to the complexity of the brain. Neuronal cell lines, primary central nervous system cells, transformed neuronal precursors and stem cell derived progenitor cells are used in neurotoxicity assays (150) to evaluate endpoints including migration, synapsis formation, network activity and differentiation. Although single-cell cultures are informative, multi-cell type and three-dimensional models utilizing microfluidics more adequately represent the diversity and spatial properties of the brain (151-154), but high throughput versions of these methods are not yet available, and thus their use in evaluating FCCs has been limited. Additional *in vitro* assays for chemical screening of neurotoxicants are under development in EU-funded research programs (155) and research is ongoing to develop further *in vitro* assays targeting the thyroid system (156). Recently, the establishment of a human cell-based *in vitro* battery has been reported; it combines 10 assays selected to cover major key events in the relevant AOPs (157) and was shown to provide 82% sensitivity in that it was able to identify 24 out of 28 known neurotoxicants (158).

New low- and medium-throughput screening assays have been developed. For example, the nematode is a promising model for evaluating known neurodevelopmental toxicants and could be expanded to profiling chemicals with unknown neurotoxicity (159, 160). Spontaneous movements (161), number and location of neurons (162), and behavioral effects (163) are some of the neurological endpoints measured in zebrafish. Validated high-throughput screening assays using African clawed frog tadpoles are also available (OECD TG 248).

*In vivo* testing in rodents can be used to assess different functional aspects of neurotoxicity including impacts on cognition, learning and memory; and anxiety-like, depressive-like and reproductive behaviors. OECD developmental neurotoxicity (OECD TG 426) and extended one-generation reproductive toxicity assays (OECD TG 443) include optional measurements of learning and memory, motor and sensory function, motor activity, and auditory startle. Neurodegeneration is not covered because animals are not kept until the end of their lifetime (164).

#### **4.4 Obesity and Metabolic diseases**

Metabolic diseases, including obesity, involve the many tissues that comprise the metabolic system (165). These include adipose tissue, skeletal muscle, pancreas, liver, gastrointestinal tract, bone, and brain. Type-2 diabetes, an important metabolic disease with increasing prevalence in human populations, occurs due to systemic insulin resistance, often with an increasing production of insulin by the pancreas. Type-1 diabetes occurs due to a progressive loss of  $\beta$ -cell insulin secretion. Non-alcoholic fatty liver disease is another metabolic disease with increasing prevalence in human populations.

While poor diet and insufficient physical activity are considered the chief drivers of the obesity and diabetes twin pandemics, chemical exposures (for example, to phthalates, bisphenols, parabens, PFAS, etc.) can disrupt the balance between energy expenditure and energy intake (166). A large comprehensive review of metabolic disrupting chemicals, including those that can induce obesity (obesogens), provides strong evidence that numerous FCCs are associated with type-2 diabetes, obesity, and fatty liver disease (167). The key characteristics of metabolic disruptors and obesogens are being compiled. Rusyn et al. (2021) have described the key characteristics of acute and chronic human hepatotoxicants and note that only one of 12 key characteristics are specific to liver tissue (KC9: causing cholestasis) (106).

The simplest assays to identify an obesity hazard are those that measure the effect of chemical exposures on the development of adipocytes (168-170). Primary preadipocyte cultures, or mesenchymal stem cell assays, use animal or human cells to assess proliferation and differentiation into adipocytes (169, 171-176), and a recent study found that around one third of tested food contact articles contained metabolic disrupting chemicals (177). Recently, spheroid adipocyte models have been developed that improve the efficiency and speed of differentiation (178) and can be used for a more comprehensive understanding of adipocyte physiology than monolayer cultures. Other non-adipocyte cell lines, when well characterized, are also useful for mechanistic studies (97, 179). In addition to adipocyte differentiation, several other mechanisms are implicated with metabolic disease causations, for example the disruption of energy homeostasis at the level of the hypothalamus and brain. Therefore, *in vitro* assays that examine effects on hypothalamic neurons are useful (180, 181).

No assays have been developed to identify metabolic disruptors acting as diabetogens. Ongoing projects are developing assays to measure  $\beta$ -cell function and survival (182-184) using rodent  $\beta$ -cell lines (INS-1E and MIN-6) and a human  $\beta$ -cell line (ENDOC- $\beta$ H1). Assays of insulin function on the human liver cell line HepaRG, the skeletal muscle cell line C2C12, and adipocytes are also under investigation (183). One well established system of assays employing both *in vitro* and *in vivo* methods has

been used to explore the relationship between BPA and type-2 diabetes (185).

The most used assay to screen chemicals for effects on the liver uses the HepaRG cell line. This cell line can be customized with different expression levels of various drug metabolizing enzymes (186). Other 2D and 3D *in vitro* approaches use primary hepatocytes (187) and other liver models (188) to screen for effects on liver outcomes.

#### **4.5 Immunological disorders**

The immune system is an intricate network of many different, highly specialized cells interacting with each other and with the nervous and endocrine systems (189). Disorders of the immune system include autoimmune disorders such as multiple sclerosis, Graves' and Hashimoto's diseases, lupus, Celiac's, Addison's, and rheumatoid arthritis, among others. Other diseases including type-1 diabetes and asthma have an important immune component. Therefore, assays for immunotoxicity need to capture a multitude of potential effects, including immunosuppression, immunostimulation, hypersensitivity reactions, mechanisms of autoimmunity, and developmental immunotoxicity, e.g., delayed immunotoxic responses to toxic influences (190).

The human immune system is highly effective, but also sensitive to synthetic chemical insults during development and adult life. Effects of chemicals on the immune system are less well understood in humans than other disease endpoints, but emerging evidence implicates PFAS exposure in reducing immune response to vaccines and increasing susceptibility to infections in early life (191). Other FCCs including bisphenols and phthalates increase the risk of atopy and asthma (192-194), and infections in early life (195).

The key characteristics of immunotoxicants have been described (111). This offers a starting point for development of suitable *in vitro* assays for testing FCCs for immunotoxicity. Due to the complexity of the immune system components and responses, a comprehensive battery of *in vitro* assays covering all relevant aspects of immunotoxicity has not been established. However, several *in vitro* assays, dealing for example with direct immunosuppression, allergic hypersensitivity, or autoimmunity, are being developed to detect a range of immunotoxicants (196-199) and these assays could be used to screen FCCs (200).

#### **4.6 Reproductive disorders**

In industrialized countries, male reproductive health has declined over the past decades, including a 50-60% decrease in sperm counts since 1973 (201, 202) and an increase in testicular cancer (203). Female fertility is also affected, as are maternal health and pregnancy outcomes, and conditions such as polycystic ovary syndrome (PCOS), endometriosis, and premature ovarian failure (204).

The sperm count decrease is associated with chemical exposures (to, e.g. phthalates), especially during fetal development (205). Strong evidence from animal experiments support this interpretation (43, 206-208). FCC exposures are also associated with PCOS (209), and other aspects of reproductive toxicity (210, 211). These adverse outcomes have even been found for FCCs promoted as safer alternatives

to hazardous chemicals such as the plasticizer 1,2-cyclohexane dicarboxylic acid diisononyl ester (tradename Hexamoll DINCH) (212), which is used as a replacement for DEHP and other phthalates. Several FCCs such as BPA have been studied for mechanistic-level impacts on female fertility, including oogenesis, folliculogenesis, and altered expression of gonadotropin and gonadotropin hormone-releasing hormone receptors (213). The key characteristics of male (109) and female reproductive toxicants (108) have been described. Development and function of the reproductive system is fundamentally dependent on sex hormone action. Thus, the key characteristics of endocrine disrupting chemicals (114) are also relevant to the study of chemicals that affect reproductive outcomes. However, a systematic overview of available *in vitro* assays for hazard identification of endocrine disrupting chemicals that affect male and female fertility is unavailable.

*In vitro* assays that identify chemical interference with sex hormone production and signalling have been validated (OECD TG 493, 455, 458, 456). These include assays based on nuclear receptor activation and steroid hormone synthesis. The bovine oocyte maturation assay (ECVAM TM 2010-05) is also a reproduction-relevant *in vitro* assay. A good correlation between *in vitro* results and *in vivo* observations has been established for female fertility endpoints (214, 215). Validated *in vivo* assays exist to evaluate reproductive toxicity for impacts on both male and female fertility (OECD TG 443), but these may not be sufficiently sensitive or comprehensive.

**Table 2.** Examples of food contact chemicals (FCCs) that are associated with diseases from the Six Clusters of Disease (SCOD) by mechanisms from *in vitro* and/or *in vivo* studies. Not a complete list: Select references only.

<b>Disease Cluster</b>	<b>Food Contact Chemical</b>	<b>Reference</b>
<b>Cancers</b>	<b>Melamine</b> (CAS 108-78-1)	(269)
	<b>Formaldehyde</b> (CAS 50-00-0)	(121)
	<b>Benzidine</b> (CAS 92-87-5)	(270)
	<b>4,4'-Diamino-3,3' - Dichlorodiphenylmethane (MOCA)</b> (CAS 101-14-4)	(121)
	<b>Antimony trioxide</b> (CAS 1309-64-4)	(271)
	<b>Perfluorooctanoic acid (PFOA)</b> (CAS 335-67-1)	(272-274)
	<b>Di (2-ethylhexyl) phthalate (DEHP)</b> (CAS 117-81-7)	(275, 276)
	<b>Bisphenol A (BPA)</b> (CAS 80-05-7)	(277-280)
<b>Cardiovascular diseases</b>	<b>Bisphenol A (BPA)</b> (CAS 80-05-7)	(143, 281-285)
	<b>Triclosan</b> (CAS 3380-34-5)	(284)
	<b>Tributyltin chloride</b> (CAS 1461-22-9)	(284)
	<b>Diethanolamine</b> (CAS 111-42-2)	(286)
	<b>DEHP</b>	(287)
<b>Brain-related</b>	<b>Perchlorate</b> (CAS 14797-73-0)	(288)

<b>disorders</b>	<b>Ortho-phthalates</b>	(289)	
	<b>BPA</b>	(290, 291)	
	<b>Bisphenol S (BPS) (CAS 80-09-1)</b>	(290, 292)	
<b>Metabolic diseases</b>	<b>BPA</b>	(293-297)	
	<b>Bisphenol A diglycidyl ether (BADGE) (CAS 1675-54-3)</b>	(298)	
	<b>Organotins</b>	(299)	
	<b>Perchlorate</b>	(300)	
	<b>Perfluorooctanesulfonic acid (PFOS) (CAS 1763-23-1)</b>	(301, 302)	
	<b>Bisphenol F (BPF) (CAS 620-92-8)</b>	(303)	
	<b>BPS</b>	(303)	
	<b>2,4,7,9-tetramethyl-5-decyne-4,7-diol (TMDD; Surfynol) (CAS 126-86-3)</b>	(304-306)	
	<b>DEHP</b>	(297, 307)	
	<b>Immunological disorders</b>	<b>Melamine</b>	(269)
<b>BPA</b>		(290)	
<b>BPF</b>		(290)	
<b>BPS</b>		(290, 308)	
<b>2,4-di-tert-butylphenol (CAS 96-76-4)</b>		(309)	
<b>DEHP</b>		(308, 310)	
<b>Reproductive disorders</b>		<b>BPA</b>	(311-314)
		<b>BADGE</b>	(298, 306)
	<b>BPS</b>	(315)	
	<b>DEHP</b>	(287, 312)	

## 5. What is needed to implement the vision for safer food contact materials?

To achieve our vision, we propose a multi-pronged approach that is grounded in the SCOD concept, which includes many of the most prevalent NCDs of high relevance to human health. We identified three components needed to realize this vision: analytical methods and testing strategies, data integration and interpretation, and science to inform decision making.

### 5.1 Analytical methods and testing strategies

In Section 4 we list several available and emerging assays used in the identification of hazard for each of the SCOD. However much more is needed, especially high-throughput non-animal and low-medium throughput assays with non-mammalian models. These assays would overcome challenges with cost, time, and scientific relevance as the selection of suitable *in vitro* assays would be based on robust mechanistic evidence from key characteristics and AOPs. Identification of the key characteristics for brain disorders and metabolic diseases will form the basis for identification and/or development of relevant *in vitro* assays to identify hazardous chemicals related to these clusters. For *in vitro* testing based on mechanistic pathways to succeed, additional dedicated expertise and financial support are needed to identify assays that would address relevant key characteristics. This work is ongoing and the website [keycharacteristics.org](http://keycharacteristics.org) collates all available information and publications in this area (216).

Another important aspect of testing is the development and validation of methods that reflect real-world chemical exposures from food contact materials. Migration testing protocols exist but ongoing research efforts need to be expanded and validated to ensure minimal loss of potentially hazardous chemicals during sample preparation (e.g. by using polar and apolar food simulants and by capturing not only non-volatile compounds, but also those that are semi-volatile and volatile).

Lastly, a battery of screening assays addressing the SCOD needs to be defined and validated. This step will need the contribution of experts in each field to ensure that the selected endpoints are reliable and result in high confidence.

### 5.2 Data interpretation and integration

Methods must be developed to interpret and corroborate *in vitro* test results. Individual assays should be integrated into an overall high-level / aggregated scheme (e.g. using visualization approaches such as ToxPi (217, 218)). Also, non-targeted chemical analysis needs advancing to allow for better identification of currently unknown compounds, especially when present at low concentrations. One way to improve the latter is to create comprehensive and open mass spectrometry libraries of FCCs, including NIAS. Ideally, an open-access repository of information about food contact material manufacturing processes and the major FCCs associated with specific materials should be generated. Confidential business information poses a critical obstacle, as the full disclosure of the chemical composition of food contact materials is commonly not available. Accordingly, a mechanism needs to be developed that enables such an FCC library without infringing on intellectual property rights.

### 5.3 Science for decision making

The results of testing single chemical or overall migrate from a food contact material using a battery of assays for each of the SCOD would need to be interpreted and integrated with available evidence to reach a conclusion within a regulatory context. A framework, similar to that available for read-across (219, 220), should be developed to effectively utilize results and support conclusions that are actionable for policy makers and regulatory enforcement. The experience gained from development of effect-based trigger values for water quality assessment in Europe could be highly informative (221, 222). Here, effect-based trigger values have been developed as a means to interpret the results of *in vitro* assays through linking the existing water quality guideline values to observed levels of bioactivity elicited by a reference chemical. Then, if a test chemical or mixture causes an activity above the trigger value set for a specific assay, it is highlighted for a follow-up assessment, such as calculation of concentration factors and *in vitro* to *in vivo* extrapolation (223-225). In theory, effect-based trigger values for food contact materials could be developed following the same principle, e.g. by matching effect concentrations in relevant bioassays with existing specific migration limits for FCCs of concern, and possibly factoring in additional exposure-related parameters. This approach appears highly promising, since it has been demonstrated that derivation of effect-based trigger values greatly facilitates regulatory and practical uptake of *in vitro* methods into specific assessment pipelines (222).

## 6. Conclusion

The novel approach we present here is in line with the goals laid out in the EU's Chemicals Strategy for Sustainability (91), the EU Farm to Fork Strategy (226), and the European Parliament's report on food contact materials (227), which emphasize the need for revising food contact material regulation in Europe to adequately reflect recent scientific understanding and improve compliance. Further, this work adds to previous publications on policies and methods related to the risk assessment of food contact chemicals and materials (10, 22, 30).

We think that our vision to create safer food contact materials by linking hazard identification more directly to human health has the potential to spur innovation in assay development and testing, and ultimately, for safer materials as such. Additionally, new findings on the key characteristics for the NCDs included in the SCOD, as well as mechanistic understanding derived from AOP research, will support the development of new assays.

Awareness of adverse health effects of synthetic chemicals is increasing globally, and the need is obvious for significant and urgent improvements in the ways that risks are assessed and managed for FCCs (228).

## Acknowledgements

We are grateful to Michele La Merrill for constructive comments on this manuscript.

## **Declarations**

### *Competing Interests*

The authors have no competing interests to declare. For the sake of transparency, the authors list their relationships with various research funders and not-for-profit organizations in the following. As researchers employed by the Food Packaging Forum Foundation (FPF) (JMB, BG, JM, LZ) or working pro bono as members of the Foundation's board (TB, JPM, MS) and its Scientific Advisory Board (SAB) (AMA, TJC, KJG, JJH, MVM, OVM, AN, CN, AMS, LT, MW, RTZ), we are reporting that the FPF receives donations from diverse companies that may be affected by the research reported in the enclosed paper. FPF funders have no influence on any of the work at FPF and were not involved in any way in the preparation of this manuscript. TB declares that he serves as the board member of the International Panel on Chemical Pollution (IPCP), the Swedish Toxicological Council and the EU Commission's Committee on Health, Environmental and Emerging Risks (SCHEER). All those activities are pro bono and have no bearing on the content of the manuscript. None of the aforementioned organizations have had any impact on the content of the manuscript. TJC declares that he is the creator-founder of Sudoc, LLC, which deploys TAML catalysts for many applications and has potential for remediating FCCs in water. JL reports that she receives funding for another research project (ZonMw/Health-Holland Microplastics and Health project MOMENTUM 458001101) of which some partners may be affected by the research reported here. MVM is a paid consultant to the FPF. OVM is one of the representatives of the European Parliament on the European Chemical Agency's Management Board. JPM is co-founder and board member of Sudoc and he declares to have given all his shares to an irrevocable grantor trust so that he will not benefit financially if the company is successful. AN declares to have received travel reimbursement from universities, NGOs and scientific societies, to speak about endocrine-disrupting chemicals. LNV has received travel reimbursements from universities, governments, NGOs, and industry. She has received research funding from the US National Institutes of Health, the University of Massachusetts Amherst, and NGOs including the Cornell Douglas Foundation, the Allen Family Foundation, and the Great Neck Breast Cancer Coalition. She is a scientific advisor to Sudoc LLC. The FPF foundation board, whose members have no connection with any of the FPF's funders and receive no remuneration for their work, is legally obliged to guarantee that the work of the FPF is in no way influenced by the interests or views of the funders.

### *Authors' contributions*

This manuscript was initiated by the FPF's SAB and guests participating in SAB meetings in 2017, 2018, 2019, 2020, 2021; AMA, MVM, JM, JPM, RTZ and MS were responsible for preparing an outline and a first version; JM, LVM, MVM and MS edited the final draft, and all authors contributed to the various intermediate versions, wrote separate sections of the manuscript, and approved the final version.

### *Funding*

This work was supported by the FPF's own resources. FPF is a charitable foundation and it funded four meetings of its SAB with external scientific experts as guests (SMB,



BCA, FAvH, JL, LNV) during which this manuscript was prepared. Funding for travel and accommodation was provided for three meetings and two additional meetings were held online. Neither FPF's SAB members nor guest participants were reimbursed for their contributions to this manuscript. FPF's funding sources are declared on its website (<https://www.foodpackagingforum.org/about-us/funding>). MS acknowledges funding by the CETOCOEN PLUS project (CZ.02.1.01/0.0/0.0/15\_003/0000469), the project CETOCOEN EXCELLENCE (CZ.02.1.01/0.0/0.0/17\_043/0009632), and RECETOX RI (LM2018121) financed by the Czech Ministry of Education, Youth and Sports.

#### *Availability of data and materials*

Not applicable.

#### **References**

1. Chakori S, Aziz AA, Smith C, Dargusch P. Untangling the underlying drivers of the use of single-use food packaging. *Ecological Economics*. 2021;185:107063.
2. Poças MFF, Oliveira JC, Pinto HJ, Zacarias ME, Hogg T. Characterization of patterns of food packaging usage in Portuguese homes. *Food Addit Contam, Part A* 2009;26(9):1314-24.
3. Biryol D, Nicolas CI, Wambaugh J, Phillips K, Isaacs K. High-throughput dietary exposure predictions for chemical migrants from food contact substances for use in chemical prioritization. *Environ Int*. 2017;108:185-94.
4. Koch HM, Calafat AM. Human body burdens of chemicals used in plastic manufacture. *Philos Trans R Soc Lond B Biol Sci*. 2009;364(1526):2063-78.
5. Qian S, Ji H, Wu X, Li N, Yang Y, Bu J, et al. Detection and quantification analysis of chemical migrants in plastic food contact products. *PloS one*. 2018;13(12):e0208467.
6. Neltner TG, Alger HM, Leonard JE, Maffini MV. Data Gaps in Toxicity Testing of Chemicals Allowed in Food in the United States. *Reproductive Toxicology*. 2013(0).
7. Maffini MV, Alger HM, Olson ED, Neltner TG. Looking Back to Look Forward: A Review of FDA's Food Additives Safety Assessment. *Comprehensive reviews in food science and food safety*. 2013;12(4).
8. Alger HM, Maffini MV, Kulkarni NR, Bongard ED, Neltner T. Perspectives on How FDA Assesses Exposure to Food Additives When Evaluating Their Safety: Workshop Proceedings. *Comprehensive Reviews in Food Science and Food Safety*. 2013;12(1):90-119.
9. Grob K, Biedermann M, Scherbaum E, Roth M, Rieger K. Food contamination with organic materials in perspective: packaging materials as the largest and least controlled source? A view focusing on the European situation. *Crit Rev Food Sci Nutr*. 2006;46(7):529-35.
10. Muncke J, Backhaus T, Geueke B, Maffini MV, Martin OV, Myers JP, et al. Scientific challenges in the risk assessment of food contact materials. *Environ Health Perspect*. 2017;125(9):095001.
11. Castle L, Mayo A, Crews C, Gilbert J. Migration of Poly(ethylene terephthalate) (PET) Oligomers from PET Plastics into Foods during Microwave and Conventional Cooking and into Bottled Beverages. *J Food Prot*. 1989;52(5):337-42.
12. Bradley EL, Driffield M, Harmer N, Oldring PKT, Castle L. Identification of potential migrants in epoxy phenolic can coatings. *Int J Polym Anal Charact* 2008;13(3):200-23.

13. Dionisi G, Oldring PK. Estimates of per capita exposure to substances migrating from canned foods and beverages. *Food Addit Contam.* 2002;19(9):891-903.
14. Jickells SM, Gancedo P, Nerin C, Castle L, Gilbert J. Migration of styrene monomer from thermoset polyester cookware into foods during high temperature applications. *Food Addit Contam.* 1993;10(5):567-73.
15. Sanchis Y, Yusà V, Coscollà C. Analytical strategies for organic food packaging contaminants. *Journal of Chromatography A.* 2017;1490:22-46.
16. Nerin C, Asensio E. Migration of organic compounds from a multilayer plastic-paper material intended for food packaging. *Analytical and Bioanalytical Chemistry.* 2007;389(2):589-96.
17. Geueke B, Groh KJ, Maffini MV, Martin OV, Boucher JM, Chiang Y-T, et al. Systematic evidence on migrating and extractable food contact chemicals: Most chemicals detected in food contact materials are not listed for use. *Critical Reviews in Food Science and Nutrition.* 2022:1-11.
18. Tsochatzis ED, Lopes JA, Gika H, Dalsgaard TK, Theodoridis G. A fast SALTE GC-MS/MS multi-analyte method for the determination of 75 food packaging substances in food simulants. *Food Chemistry.* 2021;361:129998.
19. Oldring PKT, Castle L, O'Mahony C, Dixon J. Estimates of dietary exposure to bisphenol A (BPA) from light metal packaging using food consumption and packaging usage data: a refined deterministic approach and a fully probabilistic (FACET) approach. *Food Additives & Contaminants: Part A.* 2014;31(3):466-89.
20. Nerin C, Alfaro P, Aznar M, Domeño C. The challenge of identifying non-intentionally added substances from food packaging materials: A review. *Analytica Chimica Acta.* 2013;775(2 May 2013):14-24.
21. Tisler S, Christensen JH. Non-target screening for the identification of migrating compounds from reusable plastic bottles into drinking water. *Journal of Hazardous Materials.* 2022;429:128331.
22. Zimmermann L, Scheringer M, Geueke B, Boucher JM, Parkinson LV, Groh KJ, et al. Implementing the EU Chemicals Strategy for Sustainability: The case of Food Contact Chemicals of Concern. *Journal of Hazardous Materials.* 2022:129167.
23. Groh KJ, Geueke B, Martin O, Maffini M, Muncke J. Overview of intentionally used food contact chemicals and their hazards. *Environment international.* 2021;150:106225.
24. Groh KJ, Geueke B, Martin O, Maffini M, Muncke J. Overview of intentionally used food contact chemicals and their hazards. *Environment international.* 2020:106225.
25. Van Bossuyt M, Van Hoeck E, Vanhaecke T, Rogiers V, Mertens B. Prioritizing Substances of Genotoxic Concern for In-Depth Safety Evaluation Using Non-Animal Approaches: The Example of Food Contact Materials. *Altex-Alternatives to Animal Experimentation.* 2019;36(2):215-30.
26. Van Bossuyt M, Van Hoeck E, Vanhaecke T, Rogiers V, Mertens B. Printed paper and board food contact materials as a potential source of food contamination. *Regulatory Toxicology and Pharmacology.* 2016;81:10-9.
27. Souton E, Severin I, Le Hegarat L, Hogeveen K, Aljawish A, Fessard V, et al. Genotoxic effects of food contact recycled paperboard extracts on two human hepatic cell lines. *Food Additives & Contaminants: Part A.* 2017:1-12.
28. Bengtstrom L, Rosenmai AK, Trier X, Jensen LK, Granby K, Vinggaard AM, et al. Non-targeted screening for contaminants in paper and board food-contact materials using effect-directed analysis and accurate mass spectrometry. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2016;33(6):1080-93.

29. Symeonides C, Brunner M, Mulders Y, Toshniwal P, Cantrell M, Mofflin L, et al. Buy-now-pay-later: Hazards to human and planetary health from plastics production, use and waste. *Journal of Paediatrics and Child Health*. 2021;57(11):1795-804.
30. Muncke J, Andersson A-M, Backhaus T, Boucher JM, Carney Almroth B, Castillo Castillo A, et al. Impacts of food contact chemicals on human health: a consensus statement. *Environmental Health*. 2020;19(1):25.
31. Muncke J. Tackling the toxics in plastics packaging. *PLOS Biology*. 2021;19(3):e3000961.
32. Zare Jeddi M, Hopf NB, Viegas S, Price AB, Paini A, van Thriel C, et al. Towards a systematic use of effect biomarkers in population and occupational biomonitoring. *Environment international*. 2021;146:106257.
33. WHO. Noncommunicable diseases. Fact sheet. 2018:<http://www.who.int/mediacentre/factsheets/fs355/en/>.
34. Roser M, Ritchie H, Spooner F. Burden of disease <https://ourworldindata.org/burden-of-disease>: OurWorldInData.org; 2021 [Available from: <https://ourworldindata.org/burden-of-disease>].
35. WHO. Infertility: WHO; 2020 [Available from: <https://www.who.int/news-room/factsheets/detail/infertility>].
36. Kassotis CD, Vandenberg LN, Demeneix BA, Porta M, Slama R, Trasande L. Endocrine-disrupting chemicals: economic, regulatory, and policy implications. *The Lancet Diabetes & Endocrinology*. 2020;8(8):719-30.
37. Attina TM, Hauser R, Sathyanarayana S, Hunt PA, Bourguignon J-P, Myers JP, et al. Exposure to endocrine-disrupting chemicals in the USA: a population-based disease burden and cost analysis. *Lancet Diabetes Endo*. 2016;4(12):996-1003.
38. Trasande L, Zoeller RT, Hass U, Kortenkamp A, Grandjean P, Myers JP, et al. Estimating burden and disease costs of exposure to endocrine-disrupting chemicals in the European union. *J Clin Endocrinol Metab*. 2015;100(4):1245-55.
39. Trasande L, Zoeller RT, Hass U, Kortenkamp A, Grandjean P, Myers JP, et al. Burden of disease and costs of exposure to endocrine disrupting chemicals in the European Union: an updated analysis. *Andrology*. 2016;4(4):565-72.
40. Obsekov V, Kahn LG, Trasande L. Leveraging Systematic Reviews to Explore Disease Burden and Costs of Per- and Polyfluoroalkyl Substance Exposures in the United States. *Exposure and Health*. 2022.
41. Bergman A, Andersson A-M, Becher G, van den Berg M, Blumberg B, Bjerregaard P, et al. Science and policy on endocrine disruptors must not be mixed: a reply to a "common sense" intervention by toxicology journal editors. *Environmental Health*. 2013;12(1):69.
42. Rojas-Rueda D, Morales-Zamora E, Alsufyani WA, Herbst CH, AlBalawi SM, Alsukait R, et al. Environmental Risk Factors and Health: An Umbrella Review of Meta-Analyses. *International Journal of Environmental Research and Public Health*. 2021;18(2):704.
43. Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, Prins GS, et al. EDC-2: The Endocrine Society's Second Scientific Statement on Endocrine-Disrupting Chemicals. *Endocrine Reviews*. 2015;36(6):E1-E150.
44. Goralczyk K. A Review of the Impact of Selected Anthropogenic Chemicals from the Group of Endocrine Disruptors on Human Health. *Toxics* [Internet]. 2021; 9(7).

45. Demeneix BA, Slama R. Endocrine Disruptors: From the scientific evidence to human health protection. European Parliament; 2019. Contract No.: [http://www.europarl.europa.eu/thinktank/fr/document.html?reference=IPOL\\_STU\(2019\)608866](http://www.europarl.europa.eu/thinktank/fr/document.html?reference=IPOL_STU(2019)608866) . Accessed 7 June 2019.
46. Tanner EM, Hallerbäck MU, Wikström S, Lindh C, Kiviranta H, Gennings C, et al. Early prenatal exposure to suspected endocrine disruptor mixtures is associated with lower IQ at age seven. *Environment international*. 2020;134:105185.
47. Chamorro-Garcia R, Diaz-Castillo C, Shoucri BM, Käch H, Leavitt R, Shioda T, et al. Ancestral perinatal obesogen exposure results in a transgenerational thrifty phenotype in mice. *Nat Commun*. 2017;8(1):2012.
48. Martínez-Ibarra A, Martínez-Razo LD, MacDonald-Ramos K, Morales-Pacheco M, Vázquez-Martínez ER, López-López M, et al. Multisystemic alterations in humans induced by bisphenol A and phthalates: Experimental, epidemiological and clinical studies reveal the need to change health policies. *Environmental Pollution*. 2021;271:116380.
49. Svensson K, Tanner E, Gennings C, Lindh C, Kiviranta H, Wikström S, et al. Prenatal exposures to mixtures of endocrine disrupting chemicals and children's weight trajectory up to age 5.5 in the SELMA study. *Scientific Reports*. 2021;11(1):11036.
50. Walker VR, Boyles AL, Pelch KE, Holmgren SD, Shapiro AJ, Blystone CR, et al. Human and animal evidence of potential transgenerational inheritance of health effects: An evidence map and state-of-the-science evaluation. *Environment international*. 2018;115:48-69.
51. Feil R, Fraga MF. Epigenetics and the environment: emerging patterns and implications. *Nature Reviews Genetics*. 2012;13(2):97-109.
52. Kortenkamp A, Faust M. Regulate to reduce chemical mixture risk. *Science*. 2018;361(6399):224-6.
53. Bornehag C-G, Engdahl E, Unenge Hallerbäck M, Wikström S, Lindh C, Rüegg J, et al. Prenatal exposure to bisphenols and cognitive function in children at 7 years of age in the Swedish SELMA study. *Environment international*. 2021;150:106433.
54. Bornehag C-G, Kitraki E, Stamatakis A, Panagiotidou E, Rudén C, Shu H, et al. A Novel Approach to Chemical Mixture Risk Assessment—Linking Data from Population-Based Epidemiology and Experimental Animal Tests. *Risk Analysis*. 2019;39(10):2259-71.
55. Caporale N, Leemans M, Birgersson L, Germain P-L, Cheroni C, Borbély G, et al. From cohorts to molecules: Adverse impacts of endocrine disrupting mixtures. *Science*. 2022;375(6582):eabe8244.
56. Barr DB, Silva MJ, Kato K, Reidy JA, Malek NA, Hurtz D, et al. Assessing human exposure to phthalates using monoesters and their oxidized metabolites as biomarkers. *Environ Health Perspect*. 2003;111(9):1148-51.
57. Calafat A, Kuklennyik Z, Reidy JA, Caudill SP, Ekong J, Needham LL. Urinary Concentrations of Bisphenol A and 4-Nonylphenol in a Human Reference Population. *Environ Health Perspect*. 2005;113(4):391-5.
58. Silva MJ, Slakman AR, Reidy JA, Preau JL, Jr., Herbert AR, Samandar E, et al. Analysis of human urine for fifteen phthalate metabolites using automated solid-phase extraction. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2004;805(1):161-7.
59. Correia-Sá L, Schütze A, Norberto S, Calhau C, Domingues VF, Koch HM. Exposure of Portuguese children to the novel non-phthalate plasticizer di-(iso-nonyl)-cyclohexane-1,2-dicarboxylate (DINCH). *Environment international*. 2017;102:79-86.

60. Cortéjade A, Buleté A, Prouteau L, Chatti S, Cren C, Vulliet E. Development and optimisation of home-made stir bar sorptive extraction for analysis of plastic additives: application in human urine. *Analytical Methods*. 2017;9(23):3549-60.
61. Pouech C, Kiss A, Lafay F, Léonard D, Wiest L, Cren-Olivé C, et al. Human exposure assessment to a large set of polymer additives through the analysis of urine by solid phase extraction followed by ultra high performance liquid chromatography coupled to tandem mass spectrometry. *J Chromatogr A*. 2015;1423(Supplement C):111-23.
62. Rudel RA, Gray JM, Engel CL, Rawsthorne TW, Dodson RE, Ackerman JM, et al. Food Packaging and Bisphenol A and Bis(2-Ethyhexyl) Phthalate Exposure: Findings from a Dietary Intervention. *Environ Health Perspect*. 2011;119(7):914-20.
63. Susmann HP, Schaider LA, Rodgers KM, Rudel R. Dietary Habits Related to Food Packaging and Population Exposure to PFASs. *Environmental health perspectives*. 2019;127(10):10.
64. Isaacs KK, Wall JT, Williams AR, Hobbie KA, Sobus JR, Ulrich E, et al. A harmonized chemical monitoring database for support of exposure assessments. *Sci Data*. 2022;9(1):314.
65. Domínguez-Romero E, Komprdová K, Kalina J, Bessems J, Karakitsios S, Sarigiannis DA, et al. Time-trends in human urinary concentrations of phthalates and substitutes DEHT and DINCH in Asian and North American countries (2009–2019). *Journal of exposure science & environmental epidemiology*. 2022.
66. Bil W, Govarts E, Zeilmaker MJ, Woutersen M, Bessems J, Ma Y, et al. Approaches to mixture risk assessment of PFASs in the European population based on human hazard and biomonitoring data. *International Journal of Hygiene and Environmental Health*. 2023;247:114071.
67. Horodytska O, Cabanes A, Fullana A. Non-intentionally added substances (NIAS) in recycled plastics. *Chemosphere*. 2020;251:126373.
68. Bradley E, Coulier L. An investigation into the reaction and breakdown products from starting substances used to produce food contact plastics. Report. London: Central Science Laboratory; 2007 August 2007. Report No.: FD 07/01 Contract No.: FD07/01.
69. Bauer A, Jesús F, Gómez Ramos MJ, Lozano A, Fernández-Alba AR. Identification of unexpected chemical contaminants in baby food coming from plastic packaging migration by high resolution accurate mass spectrometry. *Food Chemistry*. 2019;295:274-88.
70. Geueke B. Non-intentionally added substances (NIAS). Food Packaging Forum Foundation; 2018 June 2018.
71. COMMISSION REGULATION (EU) No 10/2011 of 14 January 2011 on plastic materials and articles intended to come into contact with food, (2011).
72. FDA. Guidance for Industry: Preparation of Premarket Submissions for Food Contact Substances: Chemistry Recommendations. 2007 [Available from: <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/guidance-industry-preparation-premarket-submissions-food-contact-substances-chemistry>].
73. Koster S, Bani-Estivals M, Bonuomo M, Bradley E, Chagnon M, Garcia M, et al. Guidance on best practices on the risk assessment of non-intentionally added substances (NIAS) in food contact materials and articles. ILSI Europe; 2015.
74. Koster S, Rennen M, Leeman W, Houben G, Muilwijk B, van Acker F, et al. A novel safety assessment strategy for non-intentionally added substances (NIAS) in carton food contact materials. *Food Additives & Contaminants: Part A*. 2013;31(3):422-43.

75. Pieke EN, Granby K, Trier X, Smedsgaard J. A framework to estimate concentrations of potentially unknown substances by semi-quantification in liquid chromatography electrospray ionization mass spectrometry. *Analytica Chimica Acta*. 2017;975:30-41.
76. Taylor RB, Sapozhnikova Y. Assessing Chemical Migration from Plastic Food Packaging into Food Simulant by Gas and Liquid Chromatography with High-Resolution Mass Spectrometry. *Journal of Agricultural and Food Chemistry*. 2022;70(16):4805-16.
77. Leeman W, Krul L. Non-intentionally added substances in food contact materials: how to ensure consumer safety. *Current Opinion in Food Science*. 2015;6:33-7.
78. Omer E, Bichon E, Hutinet S, Royer A-L, Monteau F, Germon H, et al. Toward the characterisation of non-intentionally added substances migrating from polyester-polyurethane lacquers by comprehensive gas chromatography-mass spectrometry technologies. *Journal of Chromatography A*. 2019;1601:327-34.
79. Sapozhnikova Y, Nuñez A, Johnston J. Screening of chemicals migrating from plastic food contact materials for oven and microwave applications by liquid and gas chromatography - Orbitrap mass spectrometry. *Journal of Chromatography A*. 2021;1651:462261.
80. Bschor K. Risk, Uncertainty and Precaution in Science: The case of the Threshold of Toxicological Concern Approach in Food Toxicology. *J Sci Eng Ethics*. accepted.
81. Van Bossuyt M, Van Hoeck E, Vanhaecke T, Rogiers V, Mertens B. Safeguarding human health using in silico tools? *Archives of Toxicology*. 2017;91(7):2705-6.
82. Bailey GS, Reddy AP, Pereira CB, Hartig U, Baird W, Spitsbergen JM, et al. Nonlinear Cancer Response at Ultralow Dose: A 40800-Animal ED001 Tumor and Biomarker Study. *Chemical Research in Toxicology*. 2009;22(7):1264-76.
83. Williams DE, Orner G, Willard KD, Tilton S, Hendricks JD, Pereira C, et al. Rainbow Trout (*Oncorhynchus mykiss*) and Ultra-Low Dose Cancer Studies. *Comparative Biochemistry and Physiology*. 2009;149:175-81.
84. Neltner TG, Alger HM, Leonard JE, Maffini MV. Data gaps in toxicity testing of chemicals allowed in food in the United States. *Reproductive Toxicology*. 2013;42:85-94.
85. Madia F, Worth A, Whelan M, Corvi R. Carcinogenicity assessment: Addressing the challenges of cancer and chemicals in the environment. *Environment international*. 2019;128:417-29.
86. Scholz S, Brack W, Escher BI, Hackermüller J, Liess M, von Bergen M, et al. The EU chemicals strategy for sustainability: an opportunity to develop new approaches for hazard and risk assessment. *Archives of Toxicology*. 2022;96(8):2381-6.
87. Bennett D, Bellinger DC, Birnbaum LS, Bradman A, Chen A, Cory-Slechta DA, et al. Project TENDR: Targeting Environmental Neuro-Developmental Risks The TENDR Consensus Statement. *Environmental health perspectives*. 2016;124(7):A118-A22.
88. Mohanto NC, Ito Y, Kato S, Kamijima M. Life-Time Environmental Chemical Exposure and Obesity: Review of Epidemiological Studies Using Human Biomonitoring Methods. *Frontiers in Endocrinology*. 2021;12.
89. Foresta C, Tescari S, Di Nisio A. Impact of perfluorochemicals on human health and reproduction: a male's perspective. *J Endocrinol Invest*. 2018;41(6):639-45.
90. Nerín C, Bourdoux S, Faust B, Gude T, Lesueur C, Simat T, et al. Guidance in selecting analytical techniques for identification and quantification of non-intentionally added substances (NIAS) in food contact materials (FCMS). *Food Additives & Contaminants: Part A*. 2022;39(3):620-43.

91. EU. Chemicals Strategy for Sustainability. Towards a Toxic-Free Environment. European Commission; 2020.
92. Groh KJ, Muncke J. In Vitro Toxicity Testing of Food Contact Materials: State-of-the-Art and Future Challenges. *Comprehensive Reviews in Food Science and Food Safety*. 2017;16(5):1123–50.
93. Severin I, Souton E, Dahbi L, Chagnon MC. Use of bioassays to assess hazard of food contact material extracts: State of the art. *Food and Chemical Toxicology*. 2017;105:429-47.
94. Richard AM, Judson RS, Houck KA, Grulke CM, Volarath P, Thillainadarajah I, et al. ToxCast Chemical Landscape: Paving the Road to 21st Century Toxicology. *Chem Res Toxicol*. 2016;29(8):1225-51.
95. Tice RR, Austin CP, Kavlock RJ, Bucher JR. Improving the human hazard characterization of chemicals: a Tox21 update. *Environ Health Perspect*. 2013;121(7):756-65.
96. Filer DL, Hoffman K, Sargis RM, Trasande L, Kassotis CD. On the Utility of ToxCast-Based Predictive Models to Evaluate Potential Metabolic Disruption by Environmental Chemicals. *Environ Health Perspect*. 2022;130(5):57005.
97. Janesick AS, Dimastrogiovanni G, Vanek L, Boulos C, Chamorro-García R, Tang W, et al. On the Utility of ToxCast™ and ToxPi as Methods for Identifying New Obesogens. *Environ Health Perspect*. 2016;124(8):1214-26.
98. Schug TT, Abagyan R, Blumberg B, Collins TJ, Crews D, DeFur PL, et al. Designing endocrine disruption out of the next generation of chemicals. *Green Chemistry*. 2013.
99. Fitz-James MH, Cavalli G. Molecular mechanisms of transgenerational epigenetic inheritance. *Nature Reviews Genetics*. 2022;23(6):325-41.
100. Street ME, Audouze K, Legler J, Sone H, Palanza P. Endocrine Disrupting Chemicals: Current Understanding, New Testing Strategies and Future Research Needs. *International Journal of Molecular Sciences*. 2021;22(2):933.
101. Smith MT, Guyton KZ, Gibbons CF, Fritz JM, Portier CJ, Rusyn I, et al. Key Characteristics of Carcinogens as a Basis for Organizing Data on Mechanisms of Carcinogenesis. *Environ Health Perspect*. 2016;124(6):713-21.
102. Guyton KZ, Rusyn I, Chiu WA, Corpet DE, van den Berg M, Ross MK, et al. Application of the key characteristics of carcinogens in cancer hazard identification. *Carcinogenesis*. 2018;39(4):614-22.
103. Krewski D, Bird M, Al-Zoughool M, Birkett N, Billard M, Milton B, et al. Key characteristics of 86 agents known to cause cancer in humans. *Journal of Toxicology and Environmental Health, Part B*. 2019;22(7-8):244-63.
104. Al-Zoughool M, Bird M, Rice J, Baan RA, Billard M, Birkett N, et al. Development of a database on key characteristics of human carcinogens. *Journal of Toxicology and Environmental Health, Part B*. 2019;22(7-8):264-87.
105. Guyton KZ, Schubauer-Berigan MK. Invited Perspective: Prioritizing Chemical Testing and Evaluation Using Validated in Vitro Assays Relevant to Key Characteristics. *Environmental health perspectives*. 2021;129(7):071303.
106. Rusyn I, Arzuaga X, Cattley RC, Corton JC, Ferguson SS, Godoy P, et al. Key Characteristics of Human Hepatotoxicants as a Basis for Identification and Characterization of the Causes of Liver Toxicity. *Hepatology*. 2021;74(6):3486-96.

107. La Merrill MA, Vandenberg LN, Smith MT, Goodson W, Browne P, Patisaul HB, et al. Consensus on the key characteristics of endocrine-disrupting chemicals as a basis for hazard identification. *Nature Reviews Endocrinology*. 2020;16(1):45-57.
108. Luderer U, Eskenazi B, Hauser R, Korach KS, McHale CM, Moran F, et al. Proposed Key Characteristics of Female Reproductive Toxicants as an Approach for Organizing and Evaluating Mechanistic Data in Hazard Assessment. *Environmental health perspectives*. 2019;127(7):075001.
109. Arzuaga X, Smith MT, Gibbons CF, Skakkebaek NE, Yost EE, Beverly BEJ, et al. Proposed Key Characteristics of Male Reproductive Toxicants as an Approach for Organizing and Evaluating Mechanistic Evidence in Human Health Hazard Assessments. *Environ Health Perspect*. 2019;127(6):65001.
110. Lind L, Araujo JA, Barchowsky A, Belcher S, Berridge BR, Chiamvimonvat N, et al. Key Characteristics of Cardiovascular Toxicants. *Environmental health perspectives*. 2021;129(9):095001.
111. Germolec DR, Lebec H, Anderson SE, Burleson GR, Cardenas A, Corsini E, et al. Consensus on the Key Characteristics of Immunotoxic Agents as a Basis for Hazard Identification. *Environ Health Perspect*. 2022;130(10):105001.
112. Ankley GT, Bennett RS, Erickson RJ, Hoff DJ, Hornung MW, Johnson RD, et al. Adverse Outcome Pathways: A Conceptual Framework To Support Ecotoxicology Research And Risk Assessment. . *Environmental Toxicology and Chemistry*. 2010;29(3):730-41.
113. Coumoul X, Barouki R, Koual M, Audouze K, Tomkiewicz C. Activation of the AhR leading to breast cancer. *AOP 439 2022* [Available from: <https://aopwiki.org/aops/439>].
114. Friedman KP, Crofton K, Gilbert M. Upregulation of Thyroid Hormone Catabolism via Activation of Hepatic Nuclear Receptors, and Subsequent Adverse Neurodevelopmental Outcomes in Mammals. *AOP 8 2022* [Available from: <https://aopwiki.org/aops/8>].
115. Willis R. *Pathology of tumors*. London, England: Butterworth & Co Ltd; 1948.
116. Naxerova K. Mutation fingerprints encode cellular histories. *Nature*. 2021;597:334-6.
117. Sonnenschein C, Soto AM. Over a century of cancer research: Inconvenient truths and promising leads. *PLOS Biology*. 2020;18(4):e3000670.
118. Maffini MV, Soto AM, Calabro JM, Ucci AA, Sonnenschein C. The stroma as a crucial target in rat mammary gland carcinogenesis. *Journal of Cell Science*. 2004;117(8):1495-502.
119. L R-J, MJ B. Breast cancer by proxy: can the microenvironment be both the cause and consequence? . *Trends Mol Med*. 2009;15(1):5-13.
120. Keri RA, Ho S-M, Hunt PA, Knudsen KE, Soto AM, Prins GS. An Evaluation of Evidence for the Carcinogenic Activity of Bisphenol A. *Reprod Tox*. 2007;24(2):240-52.
121. IARC. *Chemical Agents and Related Occupations*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans Volume 100F. Lyon, France: International Agency for Research on Cancer; 2012.
122. Kim HS, Lee YJ, Koo YJ, Pack EC, Lim KM, Choi DW. Migration of monomers, plastic additives, and non-intentionally added substances from food utensils made of melamine–formaldehyde resin following ultraviolet sterilization. *Food Control*. 2021;125:107981.
123. Bach C, Dauchy X, Severin I, Munoz JF, Etienne S, Chagnon MC. Effect of temperature on the release of intentionally and non-intentionally added substances from polyethylene terephthalate (PET) bottles into water: chemical analysis and potential toxicity. *Food Chem*. 2013;139(1-4):672-80.



124. NTP. RoC Review of Antimony Trioxide: U.S. Department of Health and Human Services; 2021 [Available from: <https://ntp.niehs.nih.gov/whatwestudy/assessments/cancer/completed/antimonyt/index.html>].
125. IARC. Agents classified by the IARC monographs, volumes 1–129. <https://monographs.iarc.who.int/agents-classified-by-the-iarc/>; 2022.
126. Westerhoff P, Prapaipong P, Shock E, Hillaireau A. Antimony leaching from polyethylene terephthalate (PET) plastic used for bottled drinking water. *Water Res.* 2008;42(3):551-6.
127. Trier X, Granby K, Christensen J. Polyfluorinated surfactants (PFS) in paper and board coatings for food packaging. *Environmental Science and Pollution Research.* 2011:1-13.
128. Minet L, Wang Z, Shalin A, Bruton TA, Blum A, Peaslee GF, et al. Use and release of per- and polyfluoroalkyl substances (PFASs) in consumer food packaging in U.S. and Canada. *Environmental Science: Processes & Impacts.* 2022;24(11):2032-42.
129. Benbrahim-Tallaa L, Lauby-Secretan B, Loomis D, Guyton KZ, Grosse Y, El Ghissassi F, et al. Carcinogenicity of perfluorooctanoic acid, tetrafluoroethylene, dichloromethane, 1,2-dichloropropane, and 1,3-propane sultone. *The Lancet Oncology.* 2014;15(9):924-5.
130. Rider CV, McHale CM, Webster TF, Lowe L, Goodson WH, Merrill MAL, et al. Using the Key Characteristics of Carcinogens to Develop Research on Chemical Mixtures and Cancer. *Environmental health perspectives.* 2021;129(3):035003.
131. OECD. Guidance Document on Revisions to OECD Genetic Toxicology Test Guidelines. 2015.
132. OECD. OECD Test Guidelines for Chemicals 2022 [Available from: <https://www.oecd.org/chemicalsafety/testing/oecdguidelinesforthetestingofchemicals.htm>].
133. EFSA. Guidance document on the submission of a dossier on a substance to be used in Food Contact Materials for evaluation by EFSA by the Panel on additives, flavourings, processing aids and materials in contact with food (AFC). *EFSA Journal.* 2008.
134. Pinter E, Rainer B, Czerny T, Riegel E, Schilter B, Marin-Kuan M, et al. Evaluation of the Suitability of Mammalian In Vitro Assays to Assess the Genotoxic Potential of Food Contact Materials. *Foods.* 2020;9(2).
135. Chiara F, Indraccolo S, Trevisan A. Filling the gap between risk assessment and molecular determinants of tumor onset. *Carcinogenesis.* 2020;42(4):507-16.
136. Hwang SH, Yeom H, Han BI, Ham BJ, Lee YM, Han MR, et al. Predicting Carcinogenic Mechanisms of Non-Genotoxic Carcinogens via Combined Analysis of Global DNA Methylation and In Vitro Cell Transformation. *Int J Mol Sci.* 2020;21(15).
137. Soto AM, Brisken C, Schaeberle CM, Sonnenschein C. Does cancer start in the womb? Altered mammary gland development and predisposition to breast cancer due to in utero exposure to endocrine disruptors. *J Mammary Gland Biol Neoplasia.* 2013;18:199-208.
138. WHO. Cardiovascular diseases (CVDs) 2021 [Available from: [https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-\(cvds\)](https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds))].
139. Wen Z-J, Wang Z-Y, Zhang Y-F. Adverse cardiovascular effects and potential molecular mechanisms of DEHP and its metabolites—A review. *Science of The Total Environment.* 2022;847:157443.
140. El-Kersh K, Danielle Hopkins C, Wu X, Rai SN, Cai L, Huang J. Plasma level of antimony correlates with pulmonary arterial hypertension severity. *Current Research in Toxicology.* 2022;3:100080.

141. Yan S, Chen Y, Dong M, Song W, Belcher SM, Wang H-S. Bisphenol A and 17 $\beta$ -Estradiol Promote Arrhythmia in the Female Heart via Alteration of Calcium Handling. *PLoS one*. 2011;6(9):e25455.
142. Gear R, Kendzioriski JA, Belcher SM. Effects of bisphenol A on incidence and severity of cardiac lesions in the NCTR-Sprague-Dawley rat: A CLARITY-BPA study. *Toxicol Lett*. 2017;275:123-35.
143. Gao X, Wang HS. Impact of bisphenol a on the cardiovascular system - epidemiological and experimental evidence and molecular mechanisms. *Int J Environ Res Public Health*. 2014;11(8):8399-413.
144. Jokinen MP, Lieuallen WG, Boyle MC, Johnson CL, Malarkey DE, Nyska A. Morphologic Aspects of Rodent Cardiotoxicity in a Retrospective Evaluation of National Toxicology Program Studies. *Toxicologic Pathology*. 2011;39(5):850-60.
145. FDA. Impact Story: Improved Assessment of Cardiotoxic Risk in Drug Candidates: The Comprehensive in vitro Proarrhythmia Assay 2022 [Available from: <https://www.fda.gov/drugs/regulatory-science-action/impact-story-improved-assessment-cardiotoxic-risk-drug-candidates-comprehensive-vitro-proarrhythmia>].
146. CIPA. CIPA Initiative 2019 [Available from: <https://cipaproject.org/>].
147. Kim JI, Lee YA, Shin CH, Hong Y-C, Kim B-N, Lim Y-H. Association of bisphenol A, bisphenol F, and bisphenol S with ADHD symptoms in children. *Environment international*. 2022;161:107093.
148. Grandjean P, Landrigan PJ. Developmental neurotoxicity of industrial chemicals. *Lancet*. 2006;368(9553):2167-78.
149. Maffini MV, Trasande L, Neltner TG. Perchlorate and Diet: Human Exposures, Risks, and Mitigation Strategies. *Curr Environ Health Rep*. 2016;3(2):107-17.
150. Arshajyothirmayi VA, Gulia KK. 26 - Neurotoxicity assays. In: Mohanan PV, editor. *Biomedical Product and Materials Evaluation*: Woodhead Publishing; 2022. p. 703-23.
151. Caffrey TM, Button EB, Robert J. Toward three-dimensional in vitro models to study neurovascular unit functions in health and disease. *Neural Regen Res*. 2021;16(11):2132-40.
152. Kilic O, Pamies D, Lavell E, Schiapparelli P, Feng Y, Hartung T, et al. Brain-on-a-chip model enables analysis of human neuronal differentiation and chemotaxis. *Lab Chip*. 2016;16(21):4152-62.
153. Maoz BM. Brain-on-a-Chip: Characterizing the next generation of advanced in vitro platforms for modeling the central nervous system. *APL Bioengineering*. 2021;5(3):030902.
154. Park SE, Ahn J, Jeong H-E, Youn I, Huh D, Chung S. A three-dimensional in vitro model of the peripheral nervous system. *NPG Asia Materials*. 2021;13(1):2.
155. Cediell-Ulloa A, Lupu DL, Johansson Y, Hinojosa M, Özel F, Rüegg J. Impact of endocrine disrupting chemicals on neurodevelopment: the need for better testing strategies for endocrine disruption-induced developmental neurotoxicity. *Expert Review of Endocrinology & Metabolism*. 2022;17(2):131-41.
156. Kortenkamp A, Axelstad M, Baig AH, Bergman Å, Bornehag C-G, Ceniñ P, et al. Removing Critical Gaps in Chemical Test Methods by Developing New Assays for the Identification of Thyroid Hormone System-Disrupting Chemicals—The ATHENA Project. *International Journal of Molecular Sciences*. 2020;21(9):3123.

157. Sachana M, Willett C, Pistollato F, Bal-Price A. The potential of mechanistic information organised within the AOP framework to increase regulatory uptake of the developmental neurotoxicity (DNT) in vitro battery of assays. *Reprod Toxicol.* 2021;103:159-70.
158. Blum J, Masjosthusmann S, Bartmann K, Bendt F, Dolde X, Dönmez A, et al. Establishment of a human cell-based in vitro battery to assess developmental neurotoxicity hazard of chemicals. *Chemosphere.* 2022;311(Pt 2):137035.
159. Ruszkiewicz JA, Pinkas A, Miah MR, Weitz RL, Lawes MJA, Akinyemi AJ, et al. *C. elegans* as a model in developmental neurotoxicology. *Toxicology and Applied Pharmacology.* 2018;354:126-35.
160. Hunt PR, Olejnik N, Bailey KD, Vaught CA, Sprando RL. *C. elegans* Development and Activity Test detects mammalian developmental neurotoxins. *Food and Chemical Toxicology.* 2018;121:583-92.
161. Parg C, Roy NM, Ton C, Lin Y, McGrath P. Neurotoxicity assessment using zebrafish. *Journal of Pharmacological and Toxicological Methods.* 2007;55(1):103-12.
162. Rericha Y, Truong L, Leong C, Cao D, Field JA, Tanguay RL. Dietary Perfluorohexanoic Acid (PFHxA) Exposures in Juvenile Zebrafish Produce Subtle Behavioral Effects across Generations. *Toxics.* 2022;10(7):372.
163. Fitzgerald JA, Könemann S, Krümpelmann L, Županič A, vom Berg C. Approaches to Test the Neurotoxicity of Environmental Contaminants in the Zebrafish Model: From Behavior to Molecular Mechanisms. *Environmental Toxicology and Chemistry.* 2021;40(4):989-1006.
164. Huff J, Jacobson MF, Davis DL. The Limits of Two-Year Bioassay Exposure Regimens for Identifying Chemical Carcinogens. *Environ Health Perspect.* 2008;116(11):1439-42.
165. Mohajer N, Du CY, Checkcinco C, Blumberg B. Obesogens: How They Are Identified and Molecular Mechanisms Underlying Their Action. *Front Endocrinol (Lausanne).* 2021;12:780888.
166. Heindel JJ, Howard S, Agay-Shay K, Arrebola JP, Audouze K, Babin PJ, et al. Obesity II: Establishing causal links between chemical exposures and obesity. *Biochem Pharmacol.* 2022;199:115015.
167. Heindel JJ. History of the Obesogen Field: Looking Back to Look Forward. *Front Endocrinol (Lausanne).* 2019;10:14.
168. Kassotis CD, Vom Saal FS, Babin PJ, Lagadic-Gossmann D, Le Mentec H, Blumberg B, et al. Obesity III: Obesogen assays: Limitations, strengths, and new directions. *Biochem Pharmacol.* 2022;199:115014.
169. Kassotis CD, Stapleton HM. Endocrine-Mediated Mechanisms of Metabolic Disruption and New Approaches to Examine the Public Health Threat. *Front Endocrinol (Lausanne).* 2019;10:39.
170. Seo Y, Shin TH, Kim HS. Current Strategies to Enhance Adipose Stem Cell Function: An Update. *Int J Mol Sci.* 2019;20(15).
171. Desai M, Ferrini MG, Jellyman JK, Han G, Ross MG. In vivo and in vitro bisphenol A exposure effects on adiposity. *J Dev Orig Health Dis.* 2018;9(6):678-87.
172. Shoucri BM, Hung VT, Chamorro-García R, Shioda T, Blumberg B. Retinoid X Receptor Activation During Adipogenesis of Female Mesenchymal Stem Cells Programs a Dysfunctional Adipocyte. *Endocrinology.* 2018;159(8):2863-83.
173. Chamorro-Garcia R, Blumberg B. Current Research Approaches and Challenges in the Obesogen Field. *Front Endocrinol (Lausanne).* 2019;10:167.

174. Lane JM, Doyle JR, Fortin JP, Kopin AS, Ordovás JM. Development of an OP9 derived cell line as a robust model to rapidly study adipocyte differentiation. *PLoS one*. 2014;9(11):e112123.
175. Tang QQ, Otto TC, Lane MD. Commitment of C3H10T1/2 pluripotent stem cells to the adipocyte lineage. *Proc Natl Acad Sci U S A*. 2004;101(26):9607-11.
176. Pillai HK, Fang M, Beglov D, Kozakov D, Vajda S, Stapleton HM, et al. Ligand Binding and Activation of PPAR $\gamma$  by Firemaster® 550: Effects on Adipogenesis and Osteogenesis *in Vitro*. *Environmental health perspectives*. 2014;122(11):1225-32.
177. Völker J, Ashcroft F, Vedøy Å, Zimmermann L, Wagner M. Adipogenic Activity of Chemicals Used in Plastic Consumer Products. *Environmental Science & Technology*. 2022;56(4):2487-96.
178. Turner PA, Gurumurthy B, Bailey JL, Elks CM, Janorkar AV. Adipogenic Differentiation of Human Adipose-Derived Stem Cells Grown as Spheroids. *Process Biochem*. 2017;59:312-20.
179. Auerbach S, Filer D, Reif D, Walker V, Holloway AC, Schlezinger J, et al. Prioritizing Environmental Chemicals for Obesity and Diabetes Outcomes Research: A Screening Approach Using ToxCast™ High-Throughput Data. *Environ Health Perspect*. 2016;124(8):1141-54.
180. Ye W, Ramos EH, Wong BC, Belsham DD. Beneficial Effects of Metformin and/or Salicylate on Palmitate- or TNF $\alpha$ -Induced Neuroinflammatory Marker and Neuropeptide Gene Regulation in Immortalized NPY/AgRP Neurons. *PLoS one*. 2016;11(11):e0166973.
181. Loganathan N, Salehi A, Chalmers JA, Belsham DD. Bisphenol A Alters Bmal1, Per2, and Rev-Erba mRNA and Requires Bmal1 to Increase Neuropeptide Y Expression in Hypothalamic Neurons. *Endocrinology*. 2018;160(1):181-92.
182. Audouze K, Sarigiannis D, Alonso-Magdalena P, Brochot C, Casas M, Vrijheid M, et al. Integrative Strategy of Testing Systems for Identification of Endocrine Disruptors Inducing Metabolic Disorders—An Introduction to the OBERON Project. *International Journal of Molecular Sciences*. 2020;21(8):2988.
183. Legler J, Zalko D, Jourdan F, Jacobs M, Fromenty B, Balaguer P, et al. The GOLIATH Project: Towards an Internationally Harmonised Approach for Testing Metabolism Disrupting Compounds. *International Journal of Molecular Sciences*. 2020;21(10):3480.
184. Küblbeck J, Vuorio T, Niskanen J, Fortino V, Braeuning A, Abass K, et al. The EDCMET Project: Metabolic Effects of Endocrine Disruptors. *International Journal of Molecular Sciences*. 2020;21(8):3021.
185. Dos Santos RS, Medina-Gali RM, Babiloni-Chust I, Marroqui L, Nadal A. In Vitro Assays to Identify Metabolism-Disrupting Chemicals with Diabetogenic Activity in a Human Pancreatic  $\beta$ -Cell Model. *International Journal of Molecular Sciences*. 2022;23(9):5040.
186. Tolosa L, Jiménez N, Pérez G, Castell JV, Gómez-Lechón MJ, Donato MT. Customised in vitro model to detect human metabolism-dependent idiosyncratic drug-induced liver injury. *Archives of Toxicology*. 2018;92(1):383-99.
187. Kim YE, Jeon HJ, Kim D, Lee SY, Kim KY, Hong J, et al. Quantitative Proteomic Analysis of 2D and 3D Cultured Colorectal Cancer Cells: Profiling of Tankyrase Inhibitor XAV939-Induced Proteome. *Scientific Reports*. 2018;8(1):13255.
188. Zeilinger K, Freyer N, Damm G, Seehofer D, Knöspel F. Cell sources for in vitro human liver cell culture models. *Exp Biol Med (Maywood)*. 2016;241(15):1684-98.
189. Nicholson LB. The immune system. *Essays Biochem*. 2016;60(3):275-301.

190. Germolec D, Luebke R, Rooney A, Shipkowski K, Vandebriel R, van Loveren H. Immunotoxicology: A brief history, current status and strategies for future immunotoxicity assessment. *Curr Opin Toxicol*. 2017;5:55-9.
191. Grandjean P, Heilmann C, Weihe P, Nielsen F, Mogensen UB, Timmermann A, et al. Estimated exposures to perfluorinated compounds in infancy predict attenuated vaccine antibody concentrations at age 5-years. *Journal of Immunotoxicology*. 2017;14(1):188-95.
192. Xie M-Y, Ni H, Zhao D-S, Wen L-Y, Li K-S, Yang H-H, et al. Exposure to bisphenol A and the development of asthma: A systematic review of cohort studies. *Reproductive Toxicology*. 2016;65:224-9.
193. Wang JJ, Chen C-Y, Bornehag C-G. Bisphenol A exposure may increase the risk of development of atopic disorders in children. *International Journal of Hygiene and Environmental Health*. 2016;219(3):311-6.
194. Kim EH, Jeon BH, Kim J, Kim YM, Han Y, Ahn K, et al. Exposure to phthalates and bisphenol A are associated with atopic dermatitis symptoms in children: a time-series analysis. *Environ Health*. 2017;16(1):24.
195. Gascon M, Casas M, Morales E, Valvi D, Ballesteros-Gómez A, Luque N, et al. Prenatal exposure to bisphenol A and phthalates and childhood respiratory tract infections and allergy. *Journal of Allergy and Clinical Immunology*. 2015;135(2):370-8.e7.
196. Corsini E, Roggen EL. Overview of in vitro assessment of immunotoxicity. *Current Opinion in Toxicology*. 2017;5:13-8.
197. Luebke R. Immunotoxicant screening and prioritization in the twenty-first century. *Toxicol Pathol*. 2012;40(2):294-9.
198. Deprouw C, Courties A, Fini JB, Clerget-Froidevaux MS, Demeneix B, Berenbaum F, et al. Pollutants: a candidate as a new risk factor for osteoarthritis-results from a systematic literature review. *RMD Open*. 2022;8(2).
199. Naidenko OV, Andrews DQ, Temkin AM, Stoiber T, Uche UI, Evans S, et al. Investigating Molecular Mechanisms of Immunotoxicity and the Utility of ToxCast for Immunotoxicity Screening of Chemicals Added to Food. *International Journal of Environmental Research and Public Health*. 2021;18(7):3332.
200. Ogungbesan A, Neal-Kluever A, Rice P. Exploring the use of current immunological assays for the developmental immunotoxicity assessment of food contact materials. *Food and Chemical Toxicology*. 2019;133:110801.
201. Levine H, Jørgensen N, Martino-Andrade A, Mendiola J, Weksler-Derri D, Mindlis I, et al. Temporal trends in sperm count: a systematic review and meta-regression analysis. *Hum Reprod Update*. 2017;23(6):646-59.
202. Skakkebaek NE, Lindahl-Jacobsen R, Levine H, Andersson A-M, Jørgensen N, Main KM, et al. Environmental factors in declining human fertility. *Nature Reviews Endocrinology*. 2022;18(3):139-57.
203. Znaor A, Skakkebaek NE, Rajpert-De Meyts E, Kuliš T, Laversanne M, Gurney J, et al. Global patterns in testicular cancer incidence and mortality in 2020. *International Journal of Cancer*. 2022;151(5):692-8.
204. Boomsma CM, Eijkemans MJ, Hughes EG, Visser GH, Fauser BC, Macklon NS. A meta-analysis of pregnancy outcomes in women with polycystic ovary syndrome. *Hum Reprod Update*. 2006;12(6):673-83.

205. Skakkebaek NE, Rajpert-De Meyts E, Main KM. Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects: Opinion. *Human Reproduction*. 2001;16(5):972-8.
206. Skakkebaek NE, Rajpert-De Meyts E, Buck Louis GM, Toppari J, Andersson AM, Eisenberg ML, et al. Male Reproductive Disorders and Fertility Trends: Influences of Environment and Genetic Susceptibility. *Physiol Rev*. 2016;96(1):55-97.
207. Jorgensen A, Svingen T, Miles H, Chetty T, Stukenborg JB, Mitchell RT. Environmental Impacts on Male Reproductive Development: Lessons from Experimental Models. *Hormone Research in Paediatrics*. 2021.
208. Kortenkamp A. Which chemicals should be grouped together for mixture risk assessments of male reproductive disorders? *Mol Cell Endocrinol*. 2020;499:110581.
209. Soave I, Occhiali T, Assorgi C, Marci R, Caserta D. Environmental toxin exposure in polycystic ovary syndrome women and possible ovarian neoplastic repercussion. *Curr Med Res Opin*. 2020;36(4):693-703.
210. Nerín C, Su QZ, Vera P, Mendoza N, Ausejo R. Influence of nonylphenol from multilayer plastic films on artificial insemination of sows. *Anal Bioanal Chem*. 2020;412(24):6519-28.
211. Garcia-Calvo E, Machuca A, Nerín C, Rosales-Conrado N, Anunciação DS, Luque-Garcia JL. Integration of untargeted and targeted mass spectrometry-based metabolomics provides novel insights into the potential toxicity associated to surfynol. *Food Chem Toxicol*. 2020;146:111849.
212. Boisvert A, Jones S, Issop L, Erythropel HC, Papadopoulos V, Culty M. In vitro functional screening as a means to identify new plasticizers devoid of reproductive toxicity. *Environmental Research*. 2016;150:496-512.
213. Vessa B, Perlman B, McGovern PG, Morelli SS. Endocrine disruptors and female fertility: a review of pesticide and plasticizer effects. *F S Rep*. 2022;3(2):86-90.
214. Corton JC, Liu J, Kleinstreuer N, Gwinn MR, Ryan N. Towards replacement of animal tests with in vitro assays: a gene expression biomarker predicts in vitro and in vivo estrogen receptor activity. *Chem Biol Interact*. 2022;363:109995.
215. Pinto CL, Markey K, Dix D, Browne P. Identification of candidate reference chemicals for in vitro steroidogenesis assays. *Toxicol In Vitro*. 2018;47:103-19.
216. Key Characteristics. Key Characteristics. Identifying the Key Characteristics of Hazardous Chemicals and Other Exposures: A Collaborative Approach 2022 [Available from: <https://keycharacteristics.org/>].
217. Rajkumar A, Luu T, Beal MA, Barton-Maclaren TS, Hales BF, Robaire B. Phthalates and alternative plasticizers differentially affect phenotypic parameters in gonadal somatic and germ cell lines†. *Biology of Reproduction*. 2022;106(3):613-27.
218. Marvel SW, To K, Grimm FA, Wright FA, Rusyn I, Reif DM. ToxPi Graphical User Interface 2.0: Dynamic exploration, visualization, and sharing of integrated data models. *BMC Bioinformatics*. 2018;19(1):80.
219. Lizarraga LE, Suter GW, Lambert JC, Patlewicz G, Zhao JQ, Dean JL, et al. Advancing the science of a read-across framework for evaluation of data-poor chemicals incorporating systematic and new approach methods. *Regul Toxicol Pharmacol*. 2023;137:105293.
220. Agency EC. Read-Across Assessment Framework (RAAF). European Chemicals Agency; 2017.

221. Escher BI, Ait-Aïssa S, Behnisch PA, Brack W, Brion F, Brouwer A, et al. Effect-based trigger values for in vitro and in vivo bioassays performed on surface water extracts supporting the environmental quality standards (EQS) of the European Water Framework Directive. *Science of The Total Environment*. 2018;628-629:748-65.
222. Neale PA, Escher BI, de Baat ML, Enault J, Leusch FDL. Effect-Based Trigger Values Are Essential for the Uptake of Effect-Based Methods in Water Safety Planning. *Environ Toxicol Chem*. 2023;42(3):714-26.
223. Escher BI, Neale PA. Effect-Based Trigger Values for Mixtures of Chemicals in Surface Water Detected with In Vitro Bioassays. *Environ Toxicol Chem*. 2021;40(2):487-99.
224. Robitaille J, Denslow ND, Escher BI, Kurita-Oyamada HG, Marlatt V, Martyniuk CJ, et al. Towards regulation of Endocrine Disrupting chemicals (EDCs) in water resources using bioassays - A guide to developing a testing strategy. *Environ Res*. 2022;205:112483.
225. Escher BI, Neale PA, Leusch FD. Effect-based trigger values for in vitro bioassays: Reading across from existing water quality guideline values. *Water Res*. 2015;81:137-48.
226. EU. A Farm to Fork Strategy for a fair, healthy and environmentally-friendly food system. European Commission; 2020.
227. European Parliament. Implementation of the Food Contact Materials Regulation. European Parliament resolution of 6 October 2016 on the implementation of the Food Contact Materials Regulation (EC) No 1935/2004 (2015/2259(INI)). 2016. Contract No.: P8\_TA(2016)0384.
228. Fenner K, Scherlinger M. The Need for Chemical Simplification As a Logical Consequence of Ever-Increasing Chemical Pollution. *Environ Sci Technol*. 2021;55(21):14470-2.
229. IARC. Some Chemicals Used as Solvents and in Polymer Manufacture. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans Volume 110. Lyon, France; 2016.
230. ATSDR. Toxicological Profile for Perfluoroalkyls 2021 [Available from: <https://wwwn.cdc.gov/TSP/ToxProfiles/ToxProfiles.aspx?id=1117&tid=237>].
231. Melnick RL. Is peroxisome proliferation an obligatory precursor step in the carcinogenicity of di(2-ethylhexyl)phthalate (DEHP)? *Environ Health Perspect*. 2001;109(5):437-42.
232. Wan MLY, Co VA, El-Nezami H. Endocrine disrupting chemicals and breast cancer: a systematic review of epidemiological studies. *Crit Rev Food Sci Nutr*. 2022;62(24):6549-76.
233. Moon S, Yu SH, Lee CB, Park YJ, Yoo HJ, Kim DS. Effects of bisphenol A on cardiovascular disease: An epidemiological study using National Health and Nutrition Examination Survey 2003–2016 and meta-analysis. *Science of The Total Environment*. 2021;763:142941.
234. Zhang Y-F, Shan C, Wang Y, Qian L-L, Jia D-D, Zhang Y-F, et al. Cardiovascular toxicity and mechanism of bisphenol A and emerging risk of bisphenol S. *Science of The Total Environment*. 2020;723:137952.
235. Wehbe Z, Nasser SA, El-Yazbi A, Nasreddine S, Eid AH. Estrogen and Bisphenol A in Hypertension. *Current Hypertension Reports*. 2020;22(3):23.
236. Ramadan M, Cooper B, Posnack NG. Bisphenols and phthalates: Plastic chemical exposures can contribute to adverse cardiovascular health outcomes. *Birth Defects Research*. 2020;112(17):1362-85.
237. Fu X, Xu J, Zhang R, Yu J. The association between environmental endocrine disruptors and cardiovascular diseases: A systematic review and meta-analysis. *Environmental Research*. 2020;187:109464.

238. Rebolledo-Solleiro D, Flores LYC, Solleiro-Villavicencio H. Impact of BPA on behavior, neurodevelopment and neurodegeneration. *FBL*. 2021;26(2):363-400.
239. Radke EG, Braun JM, Nachman RM, Cooper GS. Phthalate exposure and neurodevelopment: A systematic review and meta-analysis of human epidemiological evidence. *Environment international*. 2020;137:105408.
240. Piekarski DJ, Diaz KR, McNERney MW. Perfluoroalkyl chemicals in neurological health and disease: Human concerns and animal models. *NeuroToxicology*. 2020;77:155-68.
241. Eales J, Bethel A, Galloway T, Hopkinson P, Morrissey K, Short RE, et al. Human health impacts of exposure to phthalate plasticizers: An overview of reviews. *Environment international*. 2022;158:106903.
242. Moore S, Paalanen L, Melymuk L, Katsonouri A, Kolossa-Gehring M, Tolonen H. The Association between ADHD and Environmental Chemicals—A Scoping Review. *International Journal of Environmental Research and Public Health*. 2022;19(5):2849.
243. Li N, Papandonatos GD, Calafat AM, Yolton K, Lanphear BP, Chen A, et al. Gestational and childhood exposure to phthalates and child behavior. *Environ Int*. 2020;144:106036.
244. Park S, Lee JM, Kim JW, Cheong JH, Yun HJ, Hong YC, et al. Association between phthalates and externalizing behaviors and cortical thickness in children with attention deficit hyperactivity disorder. *Psychol Med*. 2015;45(8):1601-12.
245. van den Dries MA, Guxens M, Spaan S, Ferguson KK, Philips E, Santos S, et al. Phthalate and Bisphenol Exposure during Pregnancy and Offspring Nonverbal IQ. *Environ Health Perspect*. 2020;128(7):77009.
246. Predieri B, Bruzzi P, Bigi E, Ciancia S, Madeo SF, Lucaccioni L, et al. Endocrine Disrupting Chemicals and Type 1 Diabetes. *International Journal of Molecular Sciences*. 2020;21(8):2937.
247. Wang B, Li M, Zhao Z, Lu J, Chen Y, Xu Y, et al. Urinary bisphenol A concentration and glucose homeostasis in non-diabetic adults: a repeated-measures, longitudinal study. *Diabetologia*. 2019;62(9):1591-600.
248. Rancière F, Lyons JG, Loh VHY, Botton J, Galloway T, Wang T, et al. Bisphenol A and the risk of cardiometabolic disorders: a systematic review with meta-analysis of the epidemiological evidence. *Environmental Health*. 2015;14(1):1-23.
249. Akash MSH, Sabir S, Rehman K. Bisphenol A-induced metabolic disorders: From exposure to mechanism of action. *Environmental Toxicology and Pharmacology*. 2020;77:103373.
250. He X, Liu Y, Xu B, Gu L, Tang W. PFOA is associated with diabetes and metabolic alteration in US men: National Health and Nutrition Examination Survey 2003-2012. *Sci Total Environ*. 2018;625:566-74.
251. Radke EG, Braun JM, Meeker JD, Cooper GS. Phthalate exposure and male reproductive outcomes: A systematic review of the human epidemiological evidence. *Environment international*. 2018;121:764-93.
252. Dales RE, Kauri LM, Cakmak S. The associations between phthalate exposure and insulin resistance,  $\beta$ -cell function and blood glucose control in a population-based sample. *Science of The Total Environment*. 2018;612:1287-92.
253. Pérez-Bermejo M, Mas-Pérez I, Murillo-Llorente MT. The Role of the Bisphenol A in Diabetes and Obesity. *Biomedicines*. 2021;9(6).



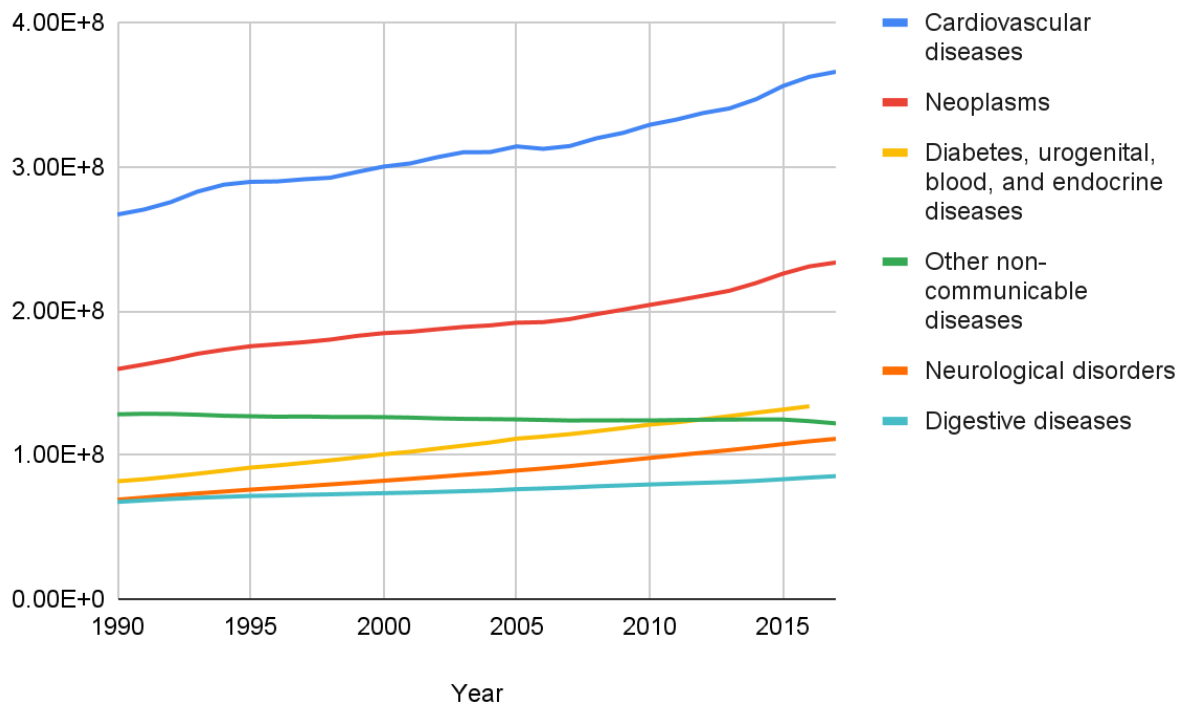
254. Wu W, Li M, Liu A, Wu C, Li D, Deng Q, et al. Bisphenol A and the Risk of Obesity a Systematic Review With Meta-Analysis of the Epidemiological Evidence. *Dose Response*. 2020;18(2):1559325820916949.
255. Liu P, Yang F, Wang Y, Yuan Z. Perfluorooctanoic Acid (PFOA) Exposure in Early Life Increases Risk of Childhood Adiposity: A Meta-Analysis of Prospective Cohort Studies. *Int J Environ Res Public Health*. 2018;15(10).
256. Geiger SD, Yao P, Vaughn MG, Qian Z. PFAS exposure and overweight/obesity among children in a nationally representative sample. *Chemosphere*. 2021;268:128852.
257. Ribeiro CM, Beserra BTS, Silva NG, Lima CL, Rocha PRS, Coelho MS, et al. Exposure to endocrine-disrupting chemicals and anthropometric measures of obesity: a systematic review and meta-analysis. *BMJ Open*. 2020;10(6):e033509.
258. Buckley JP, Engel SM, Braun JM, Whyatt RM, Daniels JL, Mendez MA, et al. Prenatal Phthalate Exposures and Body Mass Index Among 4- to 7-Year-old Children: A Pooled Analysis. *Epidemiology*. 2016;27(3):449-58.
259. Zhang Q, Li X, Liu X, Dong M, Xiao J, Wang J, et al. Association between maternal antimony exposure and risk of gestational diabetes mellitus: A birth cohort study. *Chemosphere*. 2020;246:125732.
260. Shaffer RM, Ferguson KK, Sheppard L, James-Todd T, Butts S, Chandrasekaran S, et al. Maternal urinary phthalate metabolites in relation to gestational diabetes and glucose intolerance during pregnancy. *Environ Int*. 2019;123:588-96.
261. Midya V, Colicino E, Conti DV, Berhane K, Garcia E, Stratakis N, et al. Association of Prenatal Exposure to Endocrine-Disrupting Chemicals With Liver Injury in Children. *JAMA Network Open*. 2022;5(7):e2220176-e.
262. Stratakis N, D VC, Jin R, Margetaki K, Valvi D, Siskos AP, et al. Prenatal Exposure to Perfluoroalkyl Substances Associated With Increased Susceptibility to Liver Injury in Children. *Hepatology*. 2020;72(5):1758-70.
263. DeWitt JC, Blossom SJ, Schaidler LA. Exposure to per-fluoroalkyl and polyfluoroalkyl substances leads to immunotoxicity: epidemiological and toxicological evidence. *Journal of exposure science & environmental epidemiology*. 2019;29(2):148-56.
264. Hsieh TJ, Hsieh PC, Tsai YH, Wu CF, Liu CC, Lin MY, et al. Melamine induces human renal proximal tubular cell injury via transforming growth factor- $\beta$  and oxidative stress. *Toxicol Sci*. 2012;130(1):17-32.
265. Sharma A, Mollier J, Brocklesby RWK, Caves C, Jayasena CN, Minhas S. Endocrine-disrupting chemicals and male reproductive health. *Reproductive Medicine and Biology*. 2020;19(3):243-53.
266. Estill M, Hauser R, Nassan FL, Moss A, Krawetz SA. The effects of di-butyl phthalate exposure from medications on human sperm RNA among men. *Scientific Reports*. 2019;9(1):12397.
267. Thurston SW, Mendiola J, Bellamy AR, Levine H, Wang C, Sparks A, et al. Phthalate exposure and semen quality in fertile US men. *Andrology*. 2016;4(4):632-8.
268. Messerlian C, Souter I, Gaskins AJ, Williams PL, Ford JB, Chiu YH, et al. Urinary phthalate metabolites and ovarian reserve among women seeking infertility care. *Hum Reprod*. 2016;31(1):75-83.
269. IARC. Some Chemicals That Cause Tumours of the Urinary Tract in Rodents. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans Volume 119. Lyon, France; 2019.
270. IARC. Some Aromatic Amines, Organic Dyes, and Related Exposures. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans Volume 99. Lyon, France: IARC; 2010.

271. NTP. Report on Carcinogens. Monograph on Antimony Trioxide. U.S. Department of Health and Human Services; 2018 October 2018.
272. Temkin AM, Hocevar BA, Andrews DQ, Naidenko OV, Kamendulis LM. Application of the Key Characteristics of Carcinogens to Per and Polyfluoroalkyl Substances. *International Journal of Environmental Research and Public Health*. 2020;17(5):1668.
273. Pierozan P, Jernerén F, Karlsson O. Perfluorooctanoic acid (PFOA) exposure promotes proliferation, migration and invasion potential in human breast epithelial cells. *Arch Toxicol*. 2018;92(5):1729-39.
274. Charazac A, Hinault C, Dolfi B, Hautier S, Decondé Le Butor C, Bost F, et al. Low Doses of PFOA Promote Prostate and Breast Cancer Cells Growth through Different Pathways. *Int J Mol Sci*. 2022;23(14).
275. Hager E, Chen J, Zhao L. Minireview: Parabens Exposure and Breast Cancer. *Int J Environ Res Public Health*. 2022;19(3).
276. IARC. Some Chemicals Present in Industrial and Consumer Products, Food and Drinking-water. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans Volume 101. Lyon, France: IARC; 2012.
277. Sang C, Song Y, Jin TW, Zhang S, Fu L, Zhao Y, et al. Bisphenol A induces ovarian cancer cell proliferation and metastasis through estrogen receptor- $\alpha$  pathways. *Environ Sci Pollut Res Int*. 2021;28(27):36060-8.
278. Jun JH, Oh JE, Shim JK, Kwak YL, Cho JS. Effects of bisphenol A on the proliferation, migration, and tumor growth of colon cancer cells: In vitro and in vivo evaluation with mechanistic insights related to ERK and 5-HT3. *Food Chem Toxicol*. 2021;158:112662.
279. Dhimolea E, Wadia PR, Murray TJ, Settles ML, Treitman JD, Sonnenschein C, et al. Prenatal exposure to BPA alters the epigenome of the rat mammary gland and increases the propensity to neoplastic development. *PloS one*. 2014;9(7):e99800.
280. Prins GS, Hu WY, Shi GB, Hu DP, Majumdar S, Li G, et al. Bisphenol A promotes human prostate stem-progenitor cell self-renewal and increases in vivo carcinogenesis in human prostate epithelium. *Endocrinology*. 2014;155(3):805-17.
281. Pant J, Ranjan P, Deshpande SB. Bisphenol A decreases atrial contractility involving NO-dependent G-cyclase signaling pathway. *J Appl Toxicol*. 2011;31(7):698-702.
282. Kofron CM, Kim TY, Munarin F, Soepriatna AH, Kant RJ, Mende U, et al. A predictive in vitro risk assessment platform for pro-arrhythmic toxicity using human 3D cardiac microtissues. *Scientific Reports*. 2021;11(1):10228.
283. Hyun SA, Lee CY, Ko MY, Chon SH, Kim YJ, Seo JW, et al. Cardiac toxicity from bisphenol A exposure in human-induced pluripotent stem cell-derived cardiomyocytes. *Toxicol Appl Pharmacol*. 2021;428:115696.
284. Krishna S, Berridge B, Kleinstreuer N. High-Throughput Screening to Identify Chemical Cardiotoxic Potential. *Chem Res Toxicol*. 2021;34(2):566-83.
285. Cooper BL, Posnack NG. Characteristics of Bisphenol Cardiotoxicity: Impaired Excitability, Contractility, and Relaxation. *Cardiovasc Toxicol*. 2022;22(3):273-80.
286. Jokinen MP, Lieuallen WG, Johnson CL, Dunnick J, Nyska A. Characterization of spontaneous and chemically induced cardiac lesions in rodent model systems: the national toxicology program experience. *Cardiovasc Toxicol*. 2005;5(2):227-44.

287. Mariana M, Feiteiro J, Verde I, Cairrao E. The effects of phthalates in the cardiovascular and reproductive systems: A review. *Environment international*. 2016;94:758-76.
288. Kirk AB. Environmental perchlorate: Why it matters. *Analytica Chimica Acta*. 2006;567(1):4-12.
289. Hliseníková H, Petrovičová I, Kolena B, Šidlovská M, Sirotkin A. Effects and mechanisms of phthalates' action on neurological processes and neural health: a literature review. *Pharmacol Rep*. 2021;73(2):386-404.
290. McDonough CM, Xu HS, Guo TL. Toxicity of bisphenol analogues on the reproductive, nervous, and immune systems, and their relationships to gut microbiome and metabolism: insights from a multi-species comparison. *Critical Reviews in Toxicology*. 2021;51(4):283-300.
291. Wang Z, Alderman MH, Asgari C, Taylor HS. Fetal Bisphenol-A Induced Changes in Murine Behavior and Brain Gene Expression Persisted in Adult-aged Offspring. *Endocrinology*. 2020;161(12).
292. Naderi M, Kwong RWM. A comprehensive review of the neurobehavioral effects of bisphenol S and the mechanisms of action: New insights from in vitro and in vivo models. *Environment international*. 2020;145:106078.
293. Villar-Pazos S, Martinez-Pinna J, Castellano-Muñoz M, Alonso-Magdalena P, Marroqui L, Quesada I, et al. Molecular mechanisms involved in the non-monotonic effect of bisphenol-a on  $Ca^{2+}$  entry in mouse pancreatic  $\beta$ -cells. *Sci Rep*. 2017;7(1):11770.
294. Martinez-Pinna J, Marroqui L, Hmadcha A, Lopez-Beas J, Soriano S, Villar-Pazos S, et al. Oestrogen receptor  $\beta$  mediates the actions of bisphenol-A on ion channel expression in mouse pancreatic beta cells. *Diabetologia*. 2019;62(9):1667-80.
295. Wassenaar PNH, Trasande L, Legler J. Systematic Review and Meta-Analysis of Early-Life Exposure to Bisphenol A and Obesity-Related Outcomes in Rodents. *Environ Health Perspect*. 2017;125(10):106001.
296. Desai M, Ferrini MG, Han G, Jellyman JK, Ross MG. In vivo maternal and in vitro BPA exposure effects on hypothalamic neurogenesis and appetite regulators. *Environ Res*. 2018;164:45-52.
297. Manikkam M, Tracey R, Guerrero-Bosagna C, Skinner MK. Plastics derived endocrine disruptors (BPA, DEHP and DBP) induce epigenetic transgenerational inheritance of obesity, reproductive disease and sperm epimutations. *PloS one*. 2013;8(1):e55387.
298. Wang D, Zhao H, Fei X, Synder SA, Fang M, Liu M. A comprehensive review on the analytical method, occurrence, transformation and toxicity of a reactive pollutant: BADGE. *Environment international*. 2021;155:106701.
299. Rotenberg Iu S, Mazaev VT, Shlepnina TG. Peculiarities of alkyl tin effects on respiration and oxidative phosphorylation of rat liver mitochondria. *Ukr Biokhim Zh (1978)*. 1978;50(6):695-700.
300. Larsson-Nyrén G, Sehlin J, Rorsman P, Renström E. Perchlorate stimulates insulin secretion by shifting the gating of L-type  $Ca^{2+}$  currents in mouse pancreatic B-cells towards negative potentials. *Pflugers Arch*. 2001;441(5):587-95.
301. Qin W-P, Cao L-Y, Li C-H, Guo L-H, Colbourne J, Ren X-M. Perfluoroalkyl Substances Stimulate Insulin Secretion by Islet  $\beta$  Cells via G Protein-Coupled Receptor 40. *Environmental Science & Technology*. 2020;54(6):3428-36.
302. Sant KE, Jacobs HM, Borofski KA, Moss JB, Timme-Laragy AR. Embryonic exposures to perfluorooctanesulfonic acid (PFOS) disrupt pancreatic organogenesis in the zebrafish, *Danio rerio*. *Environ Pollut*. 2017;220(Pt B):807-17.

303. Marroqui L, Martinez-Pinna J, Castellano-Muñoz M, Dos Santos RS, Medina-Gali RM, Soriano S, et al. Bisphenol-S and Bisphenol-F alter mouse pancreatic  $\beta$ -cell ion channel expression and activity and insulin release through an estrogen receptor ER $\beta$  mediated pathway. *Chemosphere*. 2021;265:129051.
304. Garcia-Calvo E, Machuca A, Nerín C, Rosales-Conrado N, Anunciação DS, Luque-Garcia JL. Integration of untargeted and targeted mass spectrometry-based metabolomics provides novel insights into the potential toxicity associated to surfynol. *Food and Chemical Toxicology*. 2020;146:111849.
305. Nerin C, Canellas E, Vera P, Garcia-Calvo E, Luque-Garcia JL, Cámara C, et al. A common surfactant used in food packaging found to be toxic for reproduction in mammals. *Food and Chemical Toxicology*. 2018;113:115-24.
306. Nerin C, Ubeda JL, Alfaro P, Dahmani Y, Aznar M, Canellas E, et al. Compounds from multilayer plastic bags cause reproductive failures in artificial insemination. *Sci Rep*. 2014;4.
307. Li J, Zheng L, Wang X, Yao K, Shi L, Sun X, et al. Taurine protects INS-1 cells from apoptosis induced by Di(2-ethylhexyl) phthalate via reducing oxidative stress and autophagy. *Toxicol Mech Methods*. 2019;29(6):445-56.
308. Nowak K, Jabłońska E, Ratajczak-Wrona W. Immunomodulatory effects of synthetic endocrine disrupting chemicals on the development and functions of human immune cells. *Environment international*. 2019;125:350-64.
309. Liu W, Zhang J, Liang X, Wang Y, Liu R, Zhang R, et al. Environmental concentrations of 2, 4-DTBP cause immunotoxicity in zebrafish (*Danio rerio*) and may elicit ecological risk to wildlife. *Chemosphere*. 2022;308:136465.
310. Hessel EVS, Tonk ECM, Bos PMJ, van Loveren H, Piersma AH. Developmental immunotoxicity of chemicals in rodents and its possible regulatory impact. *Critical Reviews in Toxicology*. 2015;45(1):68-82.
311. Liu Q. Effects of Environmental Endocrine-Disrupting Chemicals on Female Reproductive Health. In: Zhang H, Yan J, editors. *Environment and Female Reproductive Health*. Singapore: Springer Singapore; 2021. p. 205-29.
312. Vessa B, Perlman B, McGovern PG, Morelli SS. Endocrine disruptors and female fertility: a review of pesticide and plasticizer effects. *F&S Reports*. 2022;3(2):86-90.
313. Wang W, Hafner KS, Flaws JA. In utero bisphenol A exposure disrupts germ cell nest breakdown and reduces fertility with age in the mouse. *Toxicol Appl Pharmacol*. 2014;276(2):157-64.
314. Mahalingam S, Ther L, Gao L, Wang W, Ziv-Gal A, Flaws JA. The effects of in utero bisphenol A exposure on ovarian follicle numbers and steroidogenesis in the F1 and F2 generations of mice. *Reprod Toxicol*. 2017;74:150-7.
315. Desmarchais A, Tétéau O, Papillier P, Jaubert M, Druart X, Binet A, et al. Bisphenol S Impaired In Vitro Ovine Early Developmental Oocyte Competence. *International Journal of Molecular Sciences*. 2020;21(4):1238.

## Supplemental Material



**Figure S1:** Disability-Adjusted Life Years (DALYs) of worldwide selected non-communicable diseases in both sexes and all age groups, 1990 - 2017 (Diabetes, urogenital, blood, and endocrine diseases: data 1990-2016). Data: Global Burden of Disease 2021.