



Data life cycle for Life Science

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SZN Seminar, Naples, 2023-02-09

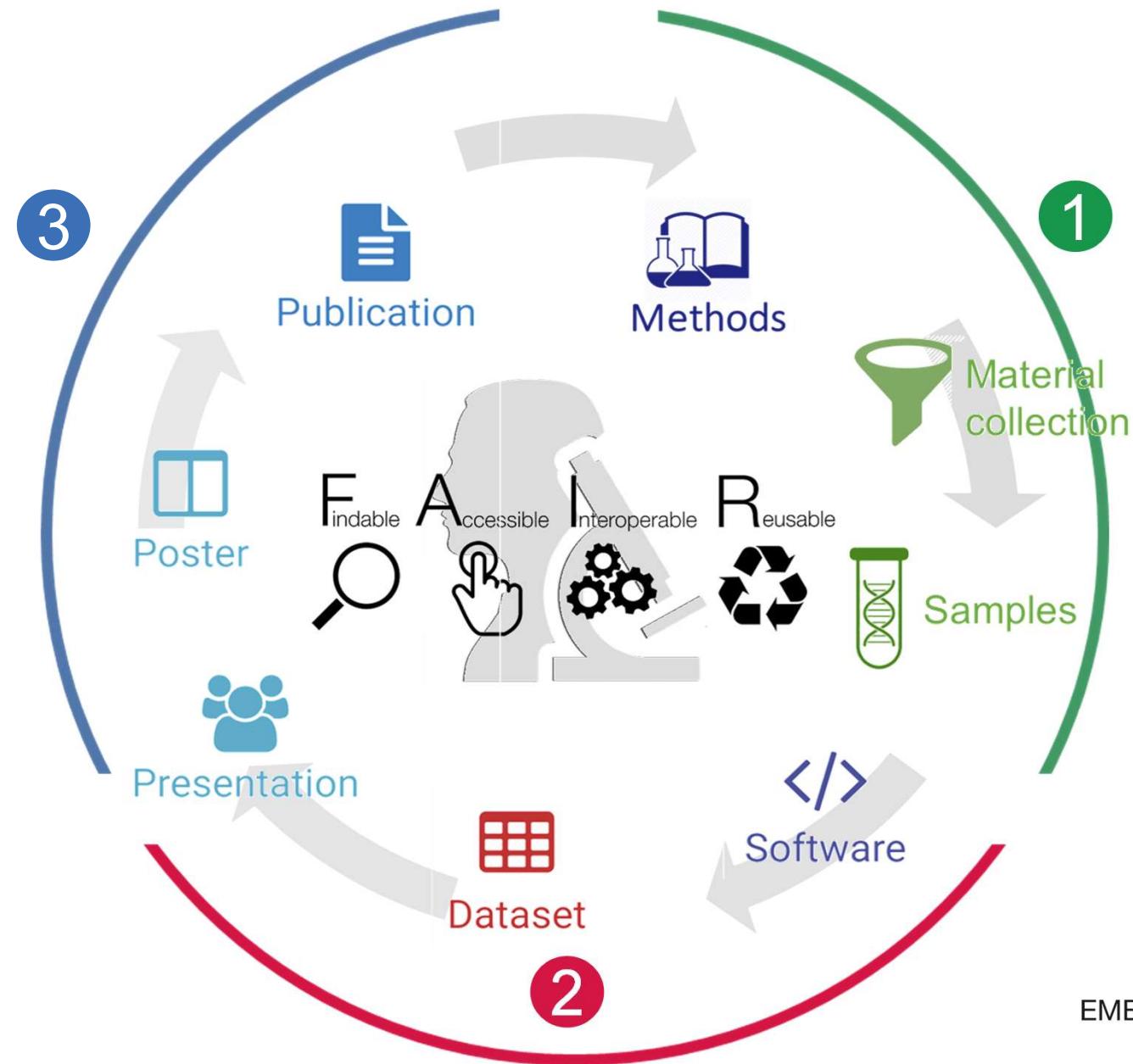


Data life cycle

Sampling
Best Practices

Data Sharing
Best Practices

Publication
Best Practices



Sampling best practices

Exemplar initiatives - developing and experimenting with best practices

- Tara Oceans
- MicroB3
- EMO-BON
- AtlantECO



System of protocols

International System of Units (SI)

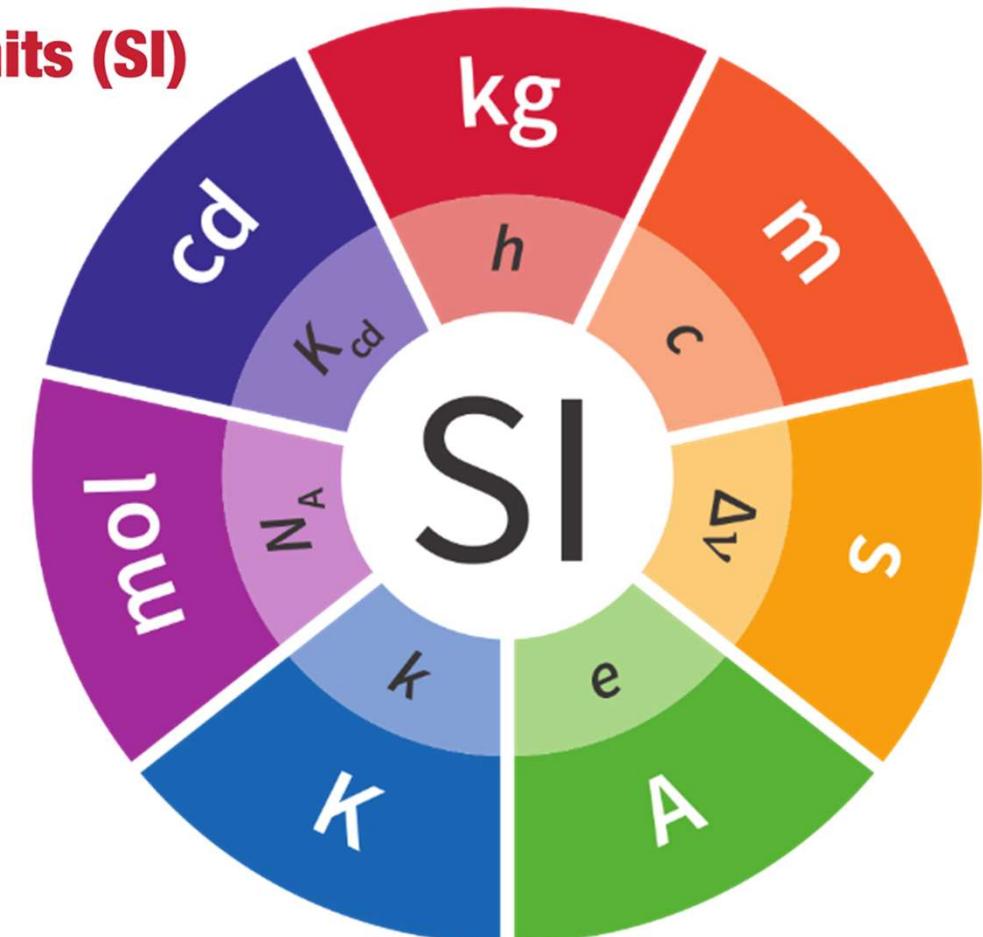
SI Base Units

Base Quantity	Name	Symbol
Length	meter	m
Mass	kilogram	kg
Time	second	s
Electric current	ampere	A
Thermodynamic temperature	kelvin	K
Amount of substance	mole	mol
Luminous intensity	candela	cd

SI Derived Units

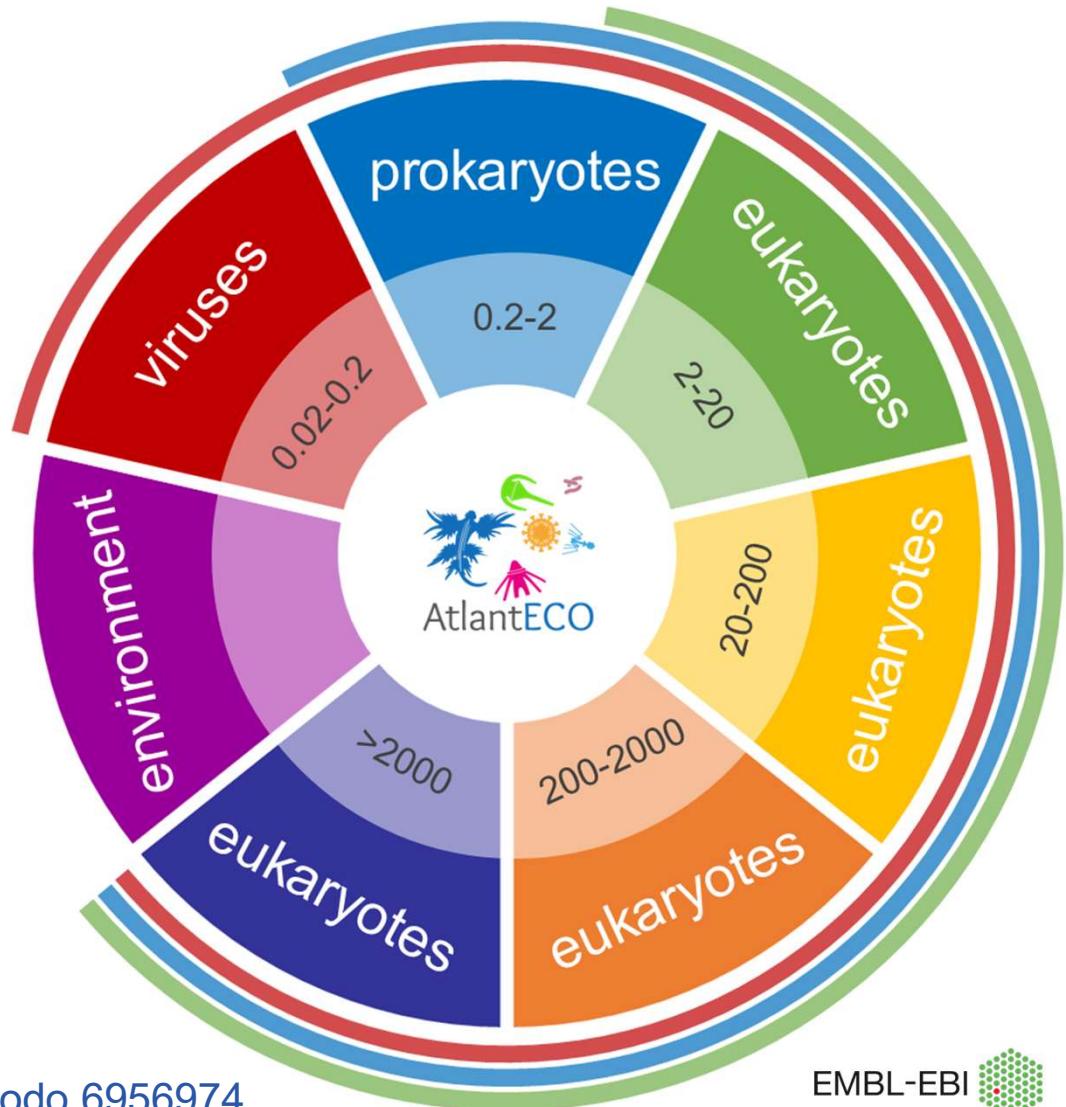
Derived Quantity	Name	Symbol	Equivalent SI units
Frequency	hertz	Hz	s^{-1}
Force	newton	N	$m \cdot kg \cdot s^{-2}$
Pressure	pascal	Pa	N/m^2
Energy	joule	J	$N \cdot m$
Power	watt	W	J/s
Electric charge	coulomb	C	$s \cdot A$
Electric potential	volt	V	W/A
Electric resistance	ohm	Ω	V/A
Celsius temperature	degree Celsius	$^{\circ}C$	K^*

*Unit degree Celsius is equal in magnitude to unit kelvin.



System of protocols

- Size
- Taxonomy
- Across size-fractions
- Genomics
- Transcriptomics
- Proteomics
- Metabolomics
- Phenomics

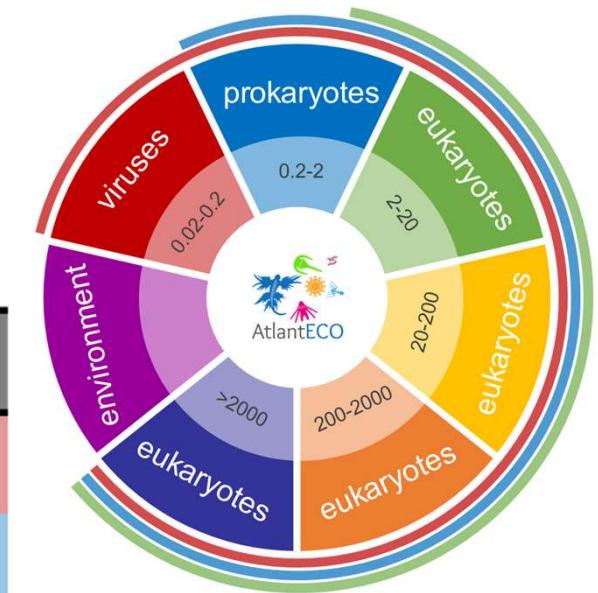


<https://doi.org/10.5281/zenodo.6956974>

System of protocols

Base phenomics protocols

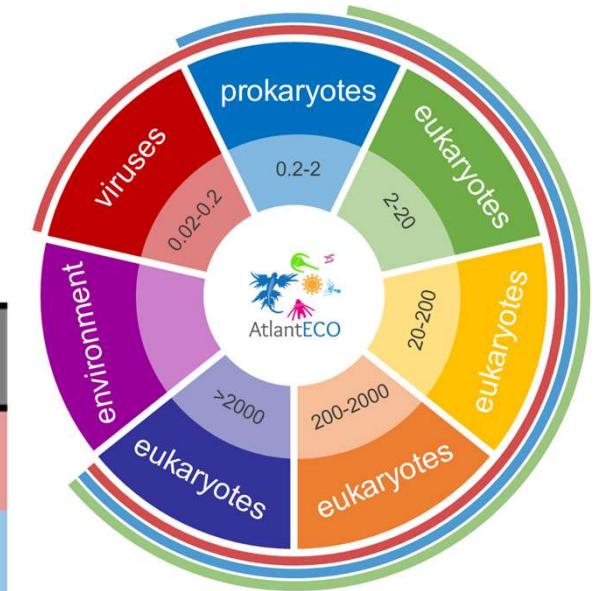
Base Protocol	Target Size fraction	Target Analysis	Target Volume (L)	Target Concentration	Preservation
I002	0.02-0.2	Flow cytometry	10^{-3}		LN_2 or -80°C
I02	0.2-2	Flow cytometry	10^{-3}		LN_2 or -80°C
I2	2-20	Fluorescence microscopy	10^2		$+4^\circ\text{C}$
I20	20-200	Flow imaging microscopy	$10^2\text{-}10^4$		live
I200	200-2000	Flatbed scan imaging	$10^3\text{-}10^5$		formaldehyde
I2000	>2000	Flatbed scan imaging	$10^3\text{-}10^5$		formaldehyde
environment	multiple	multiple	multiple		



System of protocols

Base genomics protocols

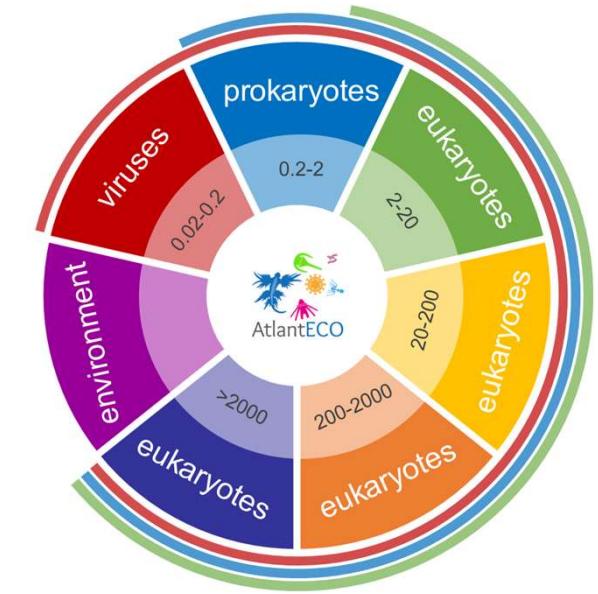
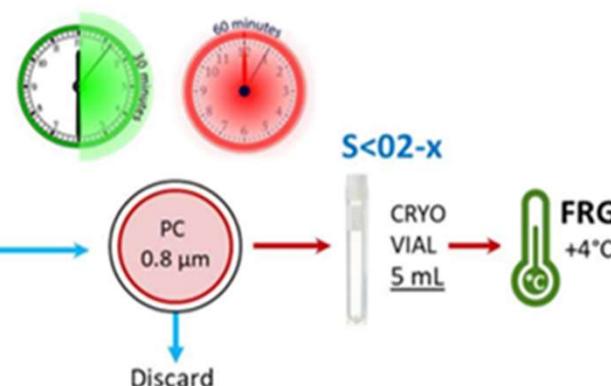
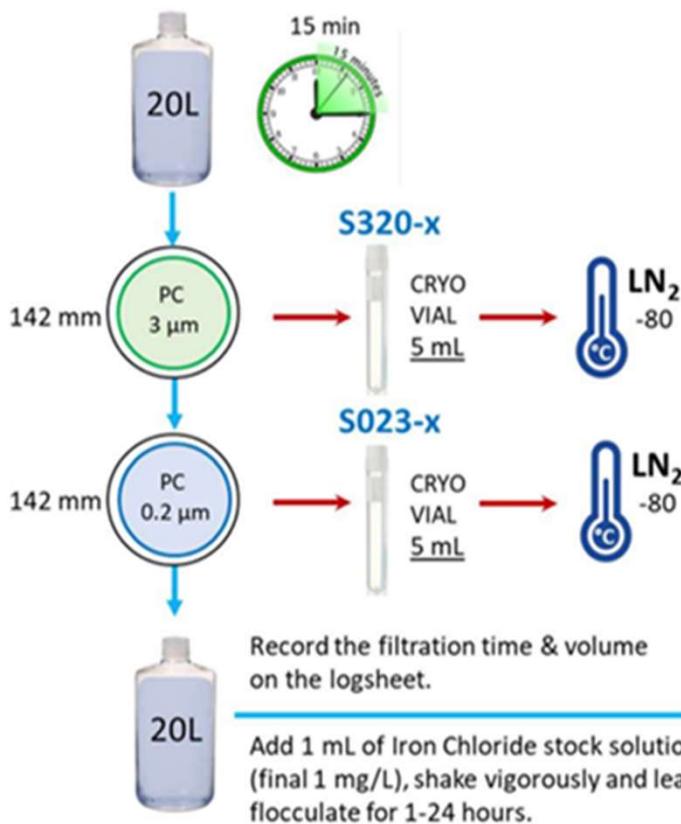
Base Protocol	Target Size fraction	Target Analysis	Target Volume (L)	Target Time (min)	Preservation
S002	0.02-0.2	MetaG, MetaT	20	Flocculated 4-24h	+4°C
S02	0.2-2	MetaB, MetaG, MetaT	20	<15	LN ₂ or -80°C
S2	2-20	MetaB, MetaG, MetaT	20	<15	LN ₂ or -80°C
S20	20-200	MetaB, MetaG, MetaT	10 ² -10 ⁴	<15	LN ₂ or -80°C
S200	200-2000	MetaB, MetaG, MetaT	10 ³ -10 ⁵	<15	LN ₂ or -80°C
S2000	>2000	MetaB, MetaG, MetaT	10 ³ -10 ⁵	<15	LN ₂ or -80°C
eDNA	>0.2	MetaB	2		LN ₂ or -80°C



Derived Protocols

- Size fractions
- Filtration volume
- Filtration time
- Preservation method

System of protocols



Derived Protocols

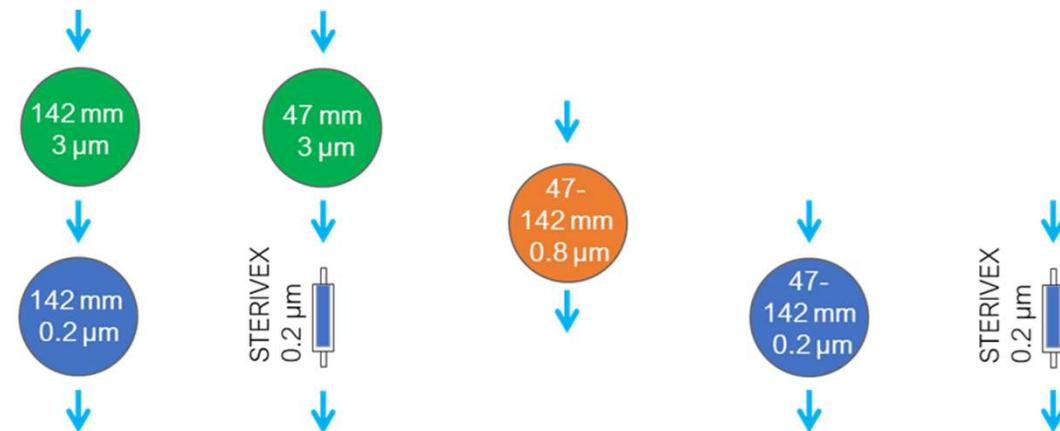
- Size fractions
- Filtration volume
- Filtration time
- Preservation method

Community Survey

All Atlantic Ocean
Microbiome Sampling



Ocean Sampling Day



METHODS	A	B	C	D	E
Filters	2x 142 mm membranes	47 mm membr. + sterivex	47 or 142 mm membrane	47 or 142 mm membrane	sterivex
Vol. in 15 min.	10-20 L	2-5 L	1-10L	1-10L	1-10 L
Pump system	large peristaltic	small peristaltic	various	various	various
ADOPTED BY	A	B	C	D	E
Ocean Sampling Day			(✓)		✓
Bio-GO-SHIP*					✓
Tara Oceans	✓		(✓)	(✓)	
EMO-BON**	✓				

*Bio-GO-SHIP Linking marine biodiversity and biogeochemistry (<https://biogoship.org/>)

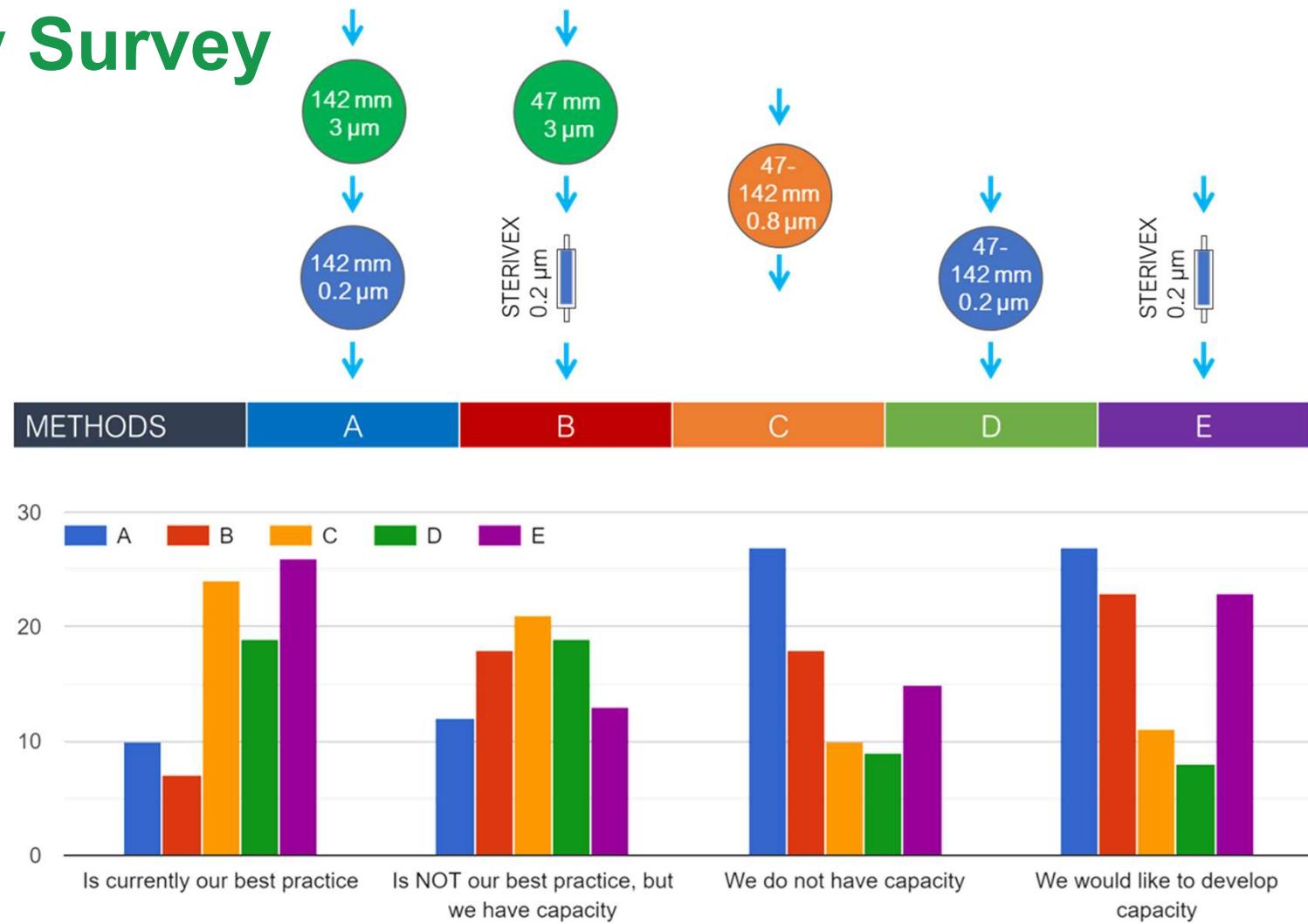
**European Marine Omics Biodiversity Observatory Network (<https://www.embrc.eu/emo-bon>)

Community Survey

All Atlantic Ocean
Microbiome Sampling



Ocean Sampling Day



Collecting Metadata – unique identifiers



Sample provenance metadata

Write in left column of the logsheet:

1. Sampling depth (m)
2. Replicate # or Control #

Write on the large barcode sticker:

1. Station ID (e.g. 021)
2. Sampling depth (e.g. 200 m)
3. Protocol label (e.g. S320)
4. Replicate # or Control # (e.g R1)

Fix the large barcode sticker on the sample container

Fix the corresponding small barcode sticker on the logsheet, in the appropriate protocol column



Collecting Metadata – provenance & context

Fondation **tara**ocean explore and share LOG_W-LAB-142 Mission Microbiomes C1

LOG_SAMPLES	YYYY	MM	DD	#	#	#	
LOG_SAMPLES	2022	04	09	_STATION-	1 0 2	_W-LAB-142	
OPERATOR(S)	Cora						
Depth	Replicate	S320 Cryo-5mL LN2 #1	S023 Cryo-5mL LN2 #1	Filtration Volume (Litres)	Filtration Duration (minutes)	S<02 Cryo-5mL FRG +4°C	Filtration Volume (Litres)
Z00	R01	7908620	7908621	[] 20L [] 50L 18.5 L	[] 15' [] 60' min.	7908634	[] 10L [] 20L L
Z00	R02	###-Z00 S320-2	###-Z00 S023-2	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z00 S<02-2	[] 10L [] 20L L
Z00	R03	###-Z00 S320-3	###-Z00 S023-3	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z00 S<02-3	[] 10L [] 20L L
Z00	R04	###-Z00 S320-4	Collect filtrate for <3 µm protocols	[] 20L [] 50L L	[] 15' [] 60' min.		
Z02	m	###-Z02 S320	###-Z02 S023	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z02 S<02	[] 10L [] 20L L
Z04	60 m	7918723	7918724	[] 20L [] 50L L	[] 15' [] 60' 14 min.	7918725	[] 10L [] 20L L
Z06	m	###-Z06 S320	###-Z06 S023	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z06 S<02	[] 10L [] 20L L
Depth	Replicate	P320 Cryo-5mL LN2 #1	P023 Cryo-5mL LN2 #1	Filtration Volume (Litres)	Filtration Duration (minutes)		
Z00	0 m	7908635	7908636	[] 20L [] 50L 37 L	[] 15' [] 60' min.		
Z02	m	###-Z02 P320	###-Z02 P023	[] 20L [] 50L L	[] 15' [] 60' min.		
Z04	60 m	7918726	7918727	[] 20L [] 50L 18 L	[] 15' [] 60' min.		
Z06	m	###-Z06 P320	###-Z06 P023	[] 20L [] 50L L	[] 15' [] 60' min.		

AtlantECO

LOG_W-LAB-142_recto_V1



Collecting Metadata – structured checklists

Fondation **tara** océan explore and share LOG_W-LAB-142 Mission **Microbiomes** C1

<input type="checkbox"/>	YYYY	MM	DD	#	#	#
LOG_SAMPLES	2021	04	09	_STATION-	1 0 2	_W-LAB-142
OPERATOR(S)	Gta					
Depth Replicate	(S320 Cryo-5mL LN2 #1)	(S023 Cryo-5mL LN2 #1)	Filtration Volume (Litres)	Filtration Duration (minutes)	(S<02 Cryo-5mL FRG +4°C)	Filtration Volume (Litres)
Z00 R01 0 m			[] 20L [] 50L 18.5 L	[] 15' [] 60' min.		[] 10L [] 20L L
Z00 R02 m	###-Z00 S320-2	###-Z00 S023-2	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z00 S<02-2	[] 10L [] 20L L
Z00 R03 m	###-Z00 S320-3	###-Z00 S023-3	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z00 S<02-3	[] 10L [] 20L L
Z00 R04 m	###-Z00 S320-4	Collect filtrate for <3 µm protocols	[] 20L [] 50L L	[] 15' [] 60' min.		
Z02 m	###-Z02 S320	###-Z02 S023	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z02 S<02	[] 10L [] 20L L
Z04 60 m			[] 20L [] 50L L	[] 15' [] 60' 14 min.		[] 10L [] 20L L
Z06 m	###-Z06 S320	###-Z06 S023	[] 20L [] 50L L	[] 15' [] 60' min.	###-Z06 S<02	[] 10L [] 20L L
Depth Replicate	(P320 Cryo-5mL LN2 #1)	(P023 Cryo-5mL LN2 #1)	Filtration Volume (Litres)	Filtration Duration (minutes)		
Z00 0 m			[] 20L [] 50L 37 L	[] 15' [] 60' min.		
Z02 m	###-Z02 P320	###-Z02 P023	[] 20L [] 50L L	[] 15' [] 60' min.		
Z04 60 m			[] 20L [] 50L L	[] 15' [] 60' 18 min.		
Z06 m	###-Z06 P320	###-Z06 P023	[] 20L [] 50L L	[] 15' [] 60' min.		

AtlantECO

LOG_W-LAB-142_recto_V1

attribute	Format / units	comment
sample id	SAMEA0000000	
sample label	Alpha-numeric, e.g. "project_date-time_station_environment_size-fraction_method"	human readable and meaningful label
sampling design, label(s)	alpha-numeric	campaign, station, site, transect, etc.
sampling device	alpha-numeric	device name & specifications
operator	alpha-numeric	initials or full name
sampling date and time	yyyy-mm-dd T hh:mm	in UTC
latitude	N/S dd.dddd or N/S dd mm.mmm	ultimately in decimal degree N
longitude	E/W ddd.ddddd or E/S ddd mm.mmm	ultimately in decimal degree E
elevation, depth below soil surface	cm	ultimately in metre
elevation, depth below sediment surface	cm	ultimately in metre
elevation, depth below water surface	m	
elevation, altitude above sea level	m	
methodological details	alpha-numeric	e.g. processing time & volume

Data sharing best practices

Permanent archives selected for the different data types:

- BioSamples for metadata (<https://www.ebi.ac.uk/biosamples/>)
- ENA for genomics data (<http://www.ebi.ac.uk/ena>)
- MGnify for metagenomic data (<https://www.ebi.ac.uk/metagenomics/>)
- PRIDE for proteomics data (<https://www.ebi.ac.uk/pride/>)
- Metabolights for metabolomics data (<https://www.ebi.ac.uk/metabolights/>)
- BioImage Archive for imaging data (<https://www.ebi.ac.uk/bioimage-archive/>)



Data workflow

Environmental
EMODnet, Copernicus, etc.

primary data
curation & archiving



Provenance & Envir. Context
BioSamples

Genomics
ENA, Mgnify & Ensembl

Proteomics
PRIDE

Metabolomics
Metabolights

Imaging
BiolImage archive & EcoTaxa



annotated data
analysis & archiving



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EMODnet



annotated data
analysis & archiving



Harmonised context

- Georeferences
- Taxonomy
- Marine domain
- Longhurst Bioge. Provinces
- IHO Sea Areas
- Oceanographic Products
- Astronomical Products
- Countries & Territories
- Exclusive Economic Zones
- Nagoya requirements
- ABS regulations

Data workflow

Environmental
EMODnet, Copernicus, etc.

Provenance & Envir. Context
BioSamples

Genomics
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Proteomics
PRIDE

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Metabolights

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BiolImage archive & EcoTaxa

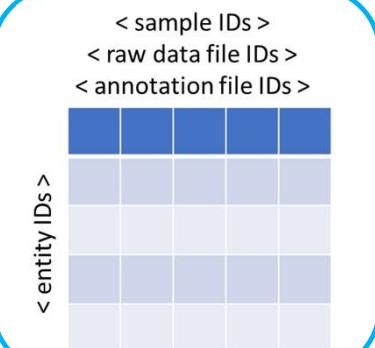
primary data
curation & archiving

annotated data
analysis & archiving

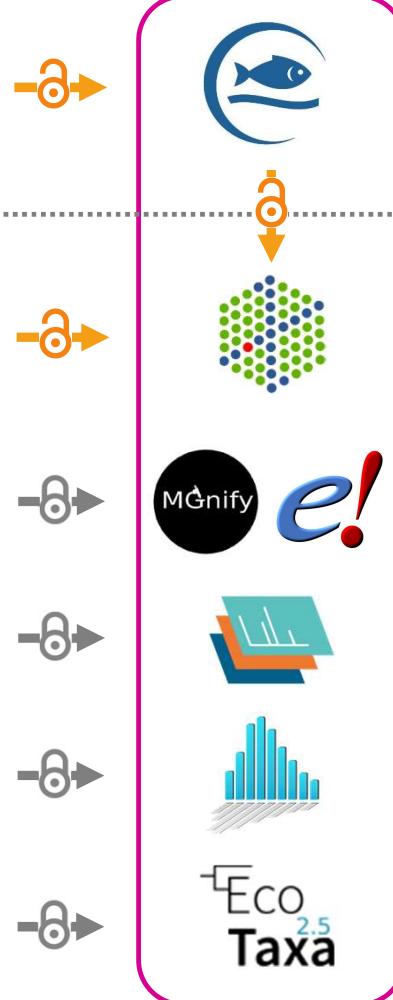
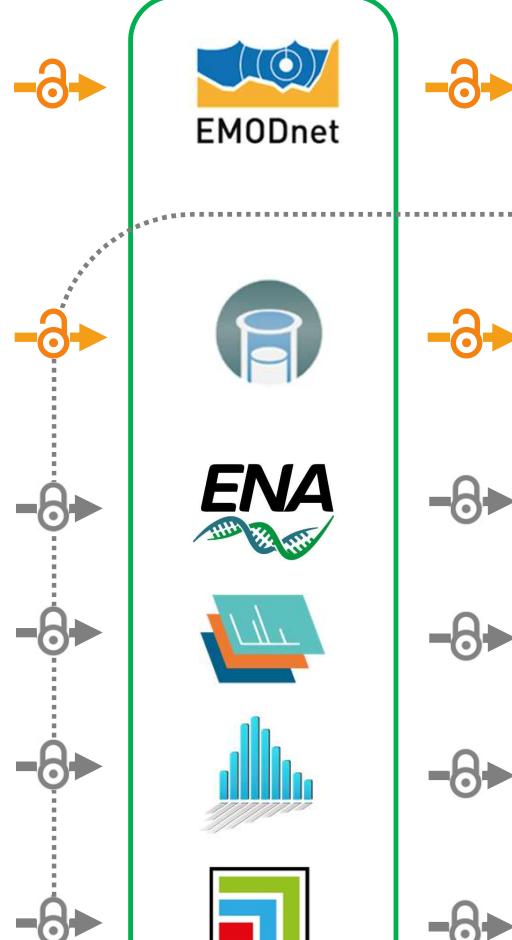
DATA HUB



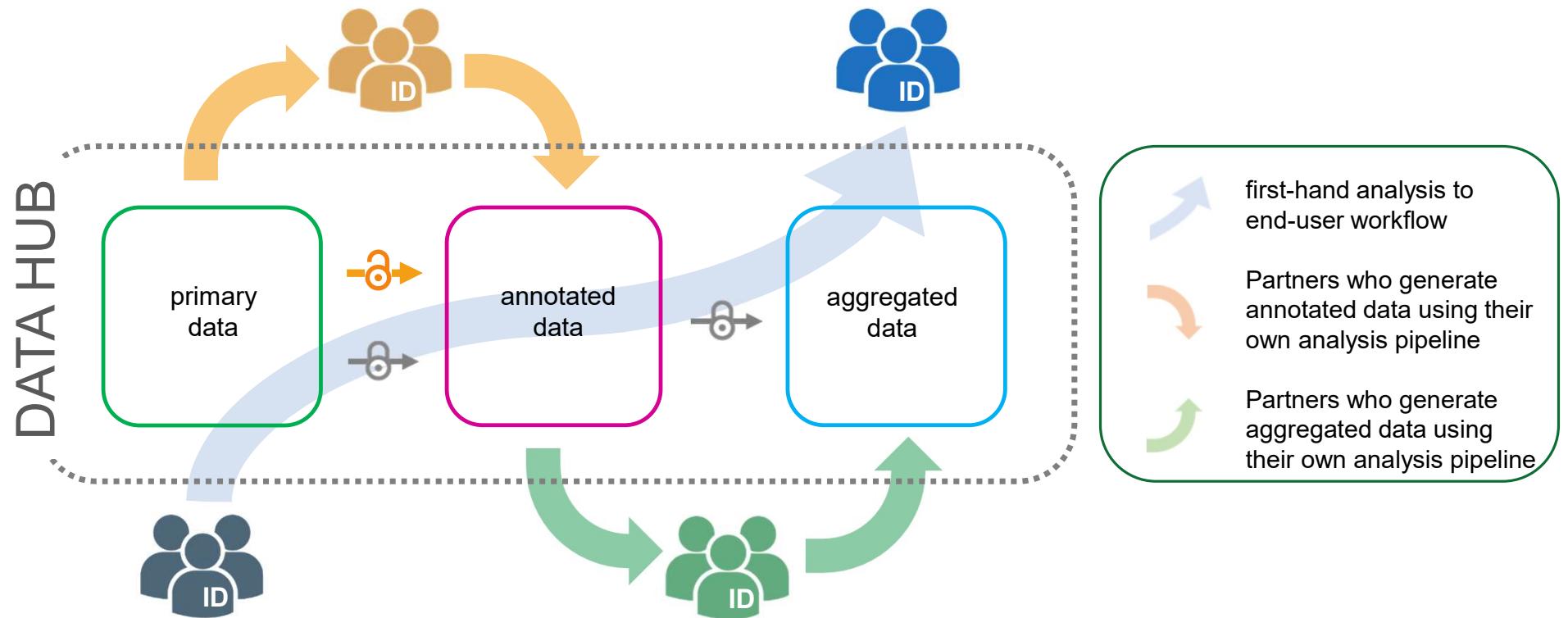
aggregated data
analysis & archiving



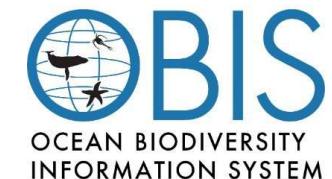
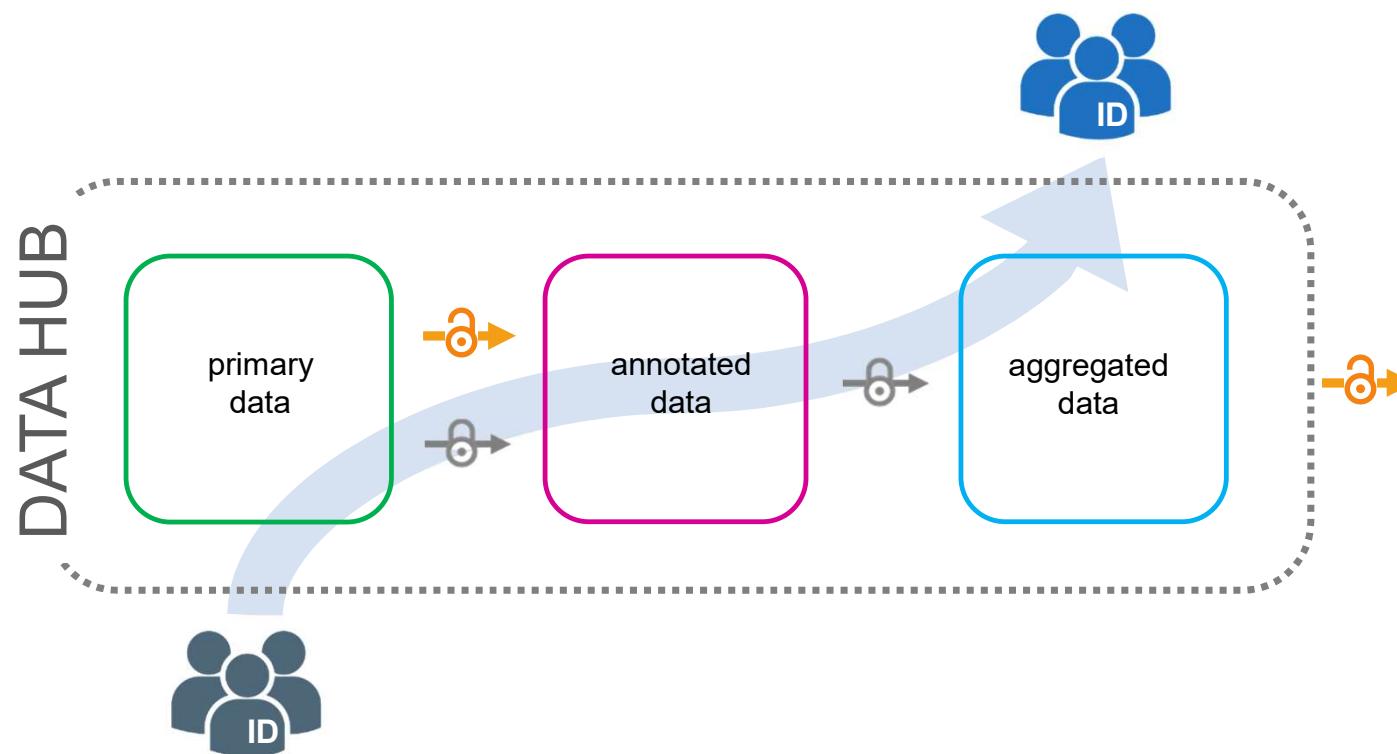
EMBL-EBI



Data Hub workflow



Data Hub workflow

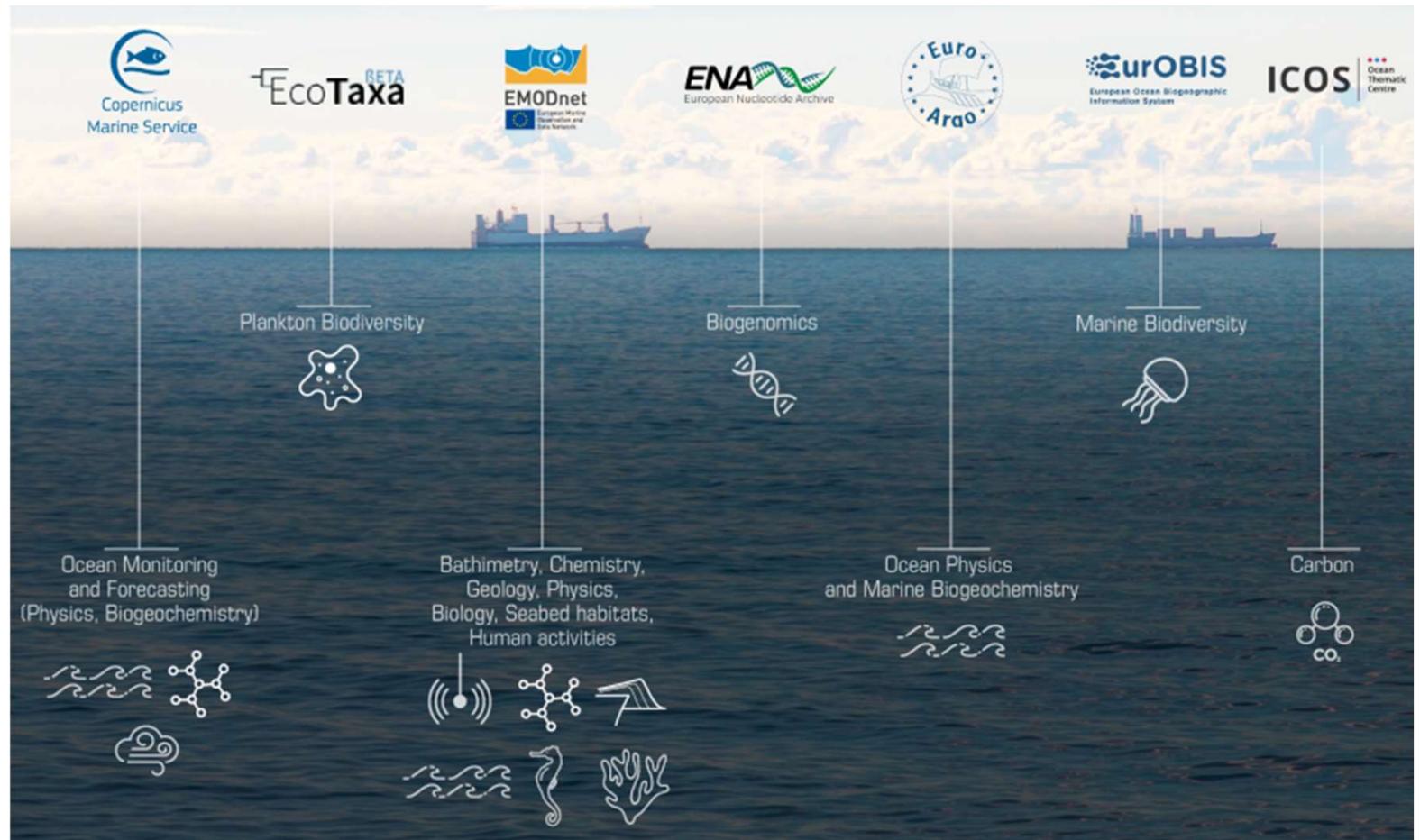




<https://blue-cloud.org/>

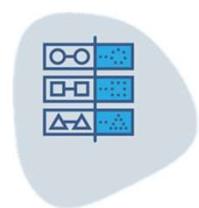


Discovery & Access

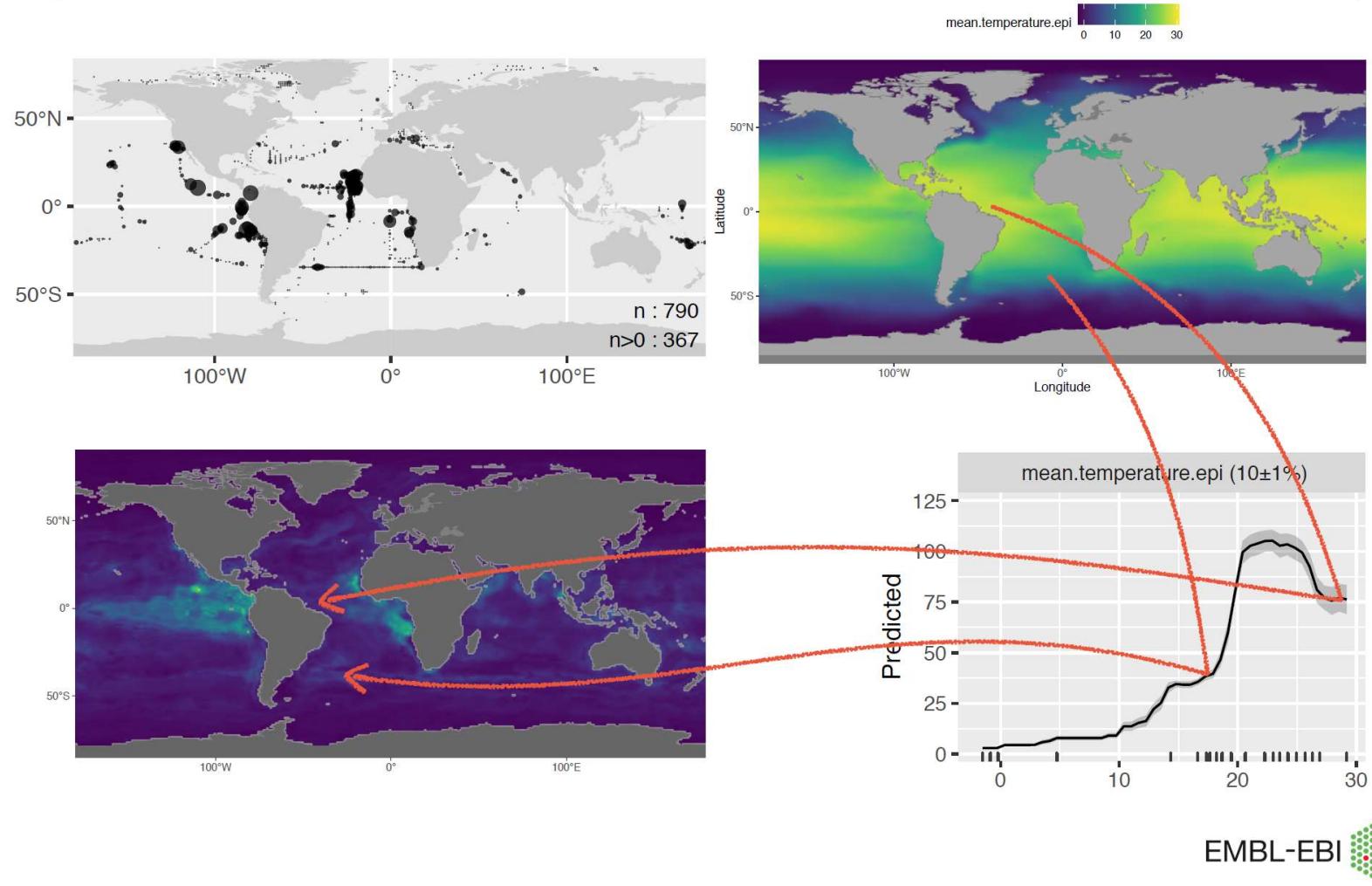




Discovery & Access



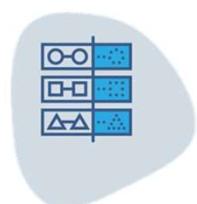
Virtual Labs &
Workbenches



<https://blue-cloud.org/>



Discovery & Access



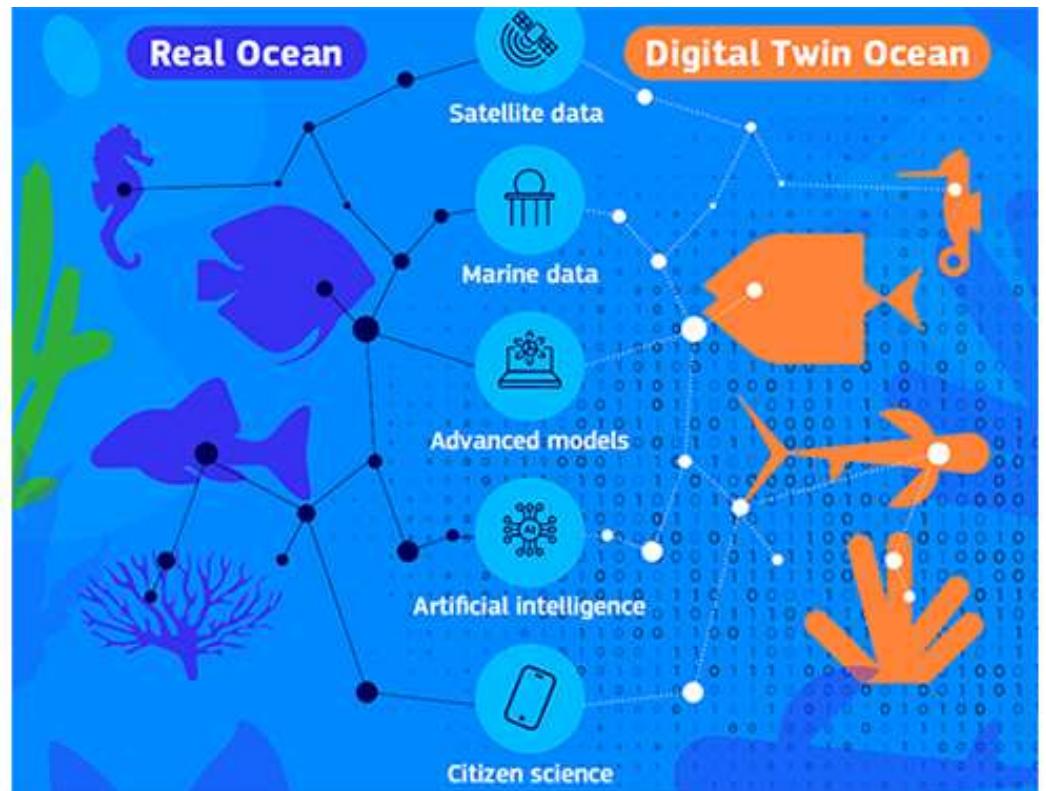
Virtual Environment



Publishing Services



EMODnet



Publication best practices

Exemplar initiatives

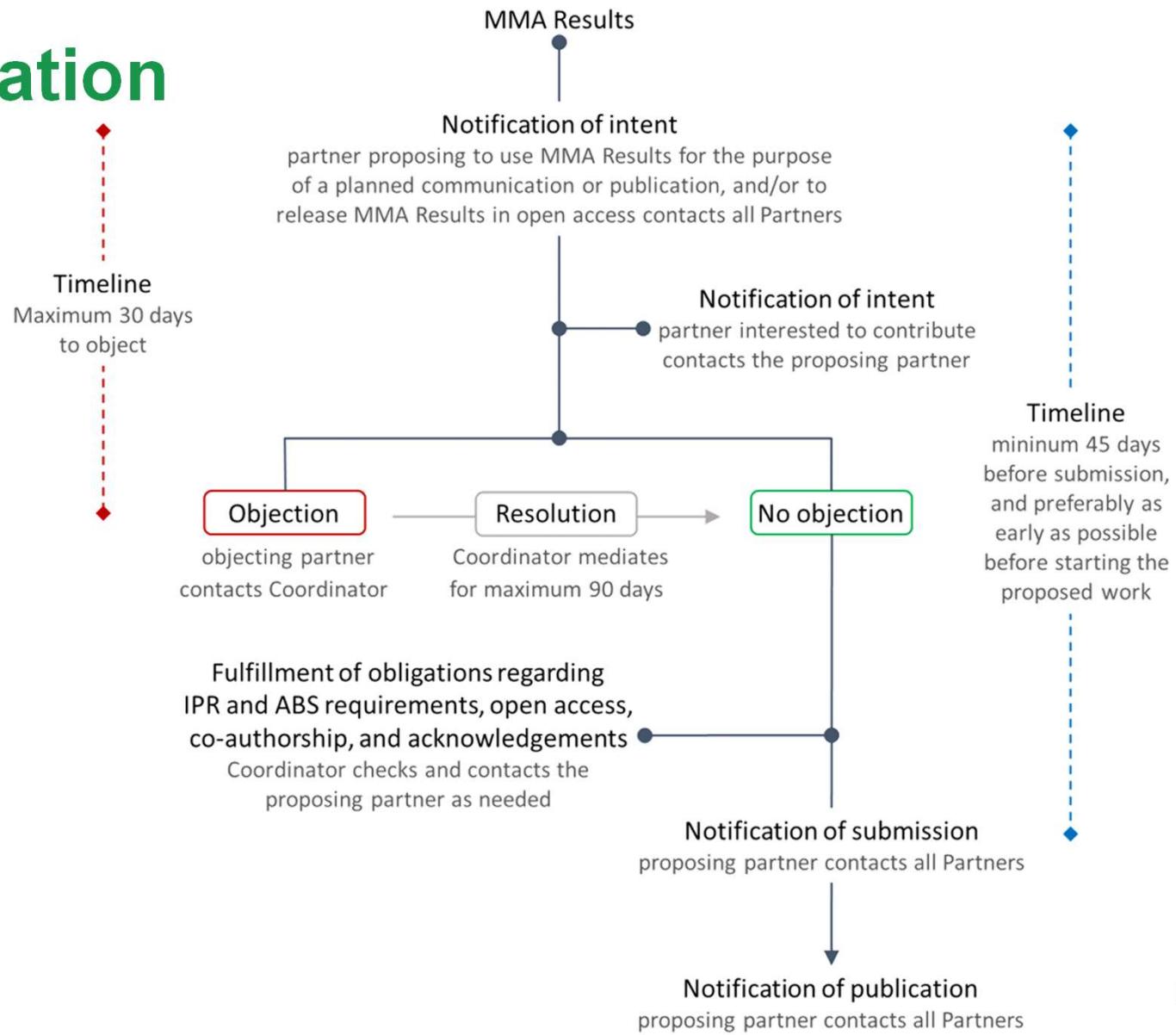
- Tara Oceans
- Tara Pacific
- AtlantECO

Best Practices

- Jointly owned results
- Early notification of intent to use “jointly owned results”
- Co-authorship
- Open Access publication



Early notification



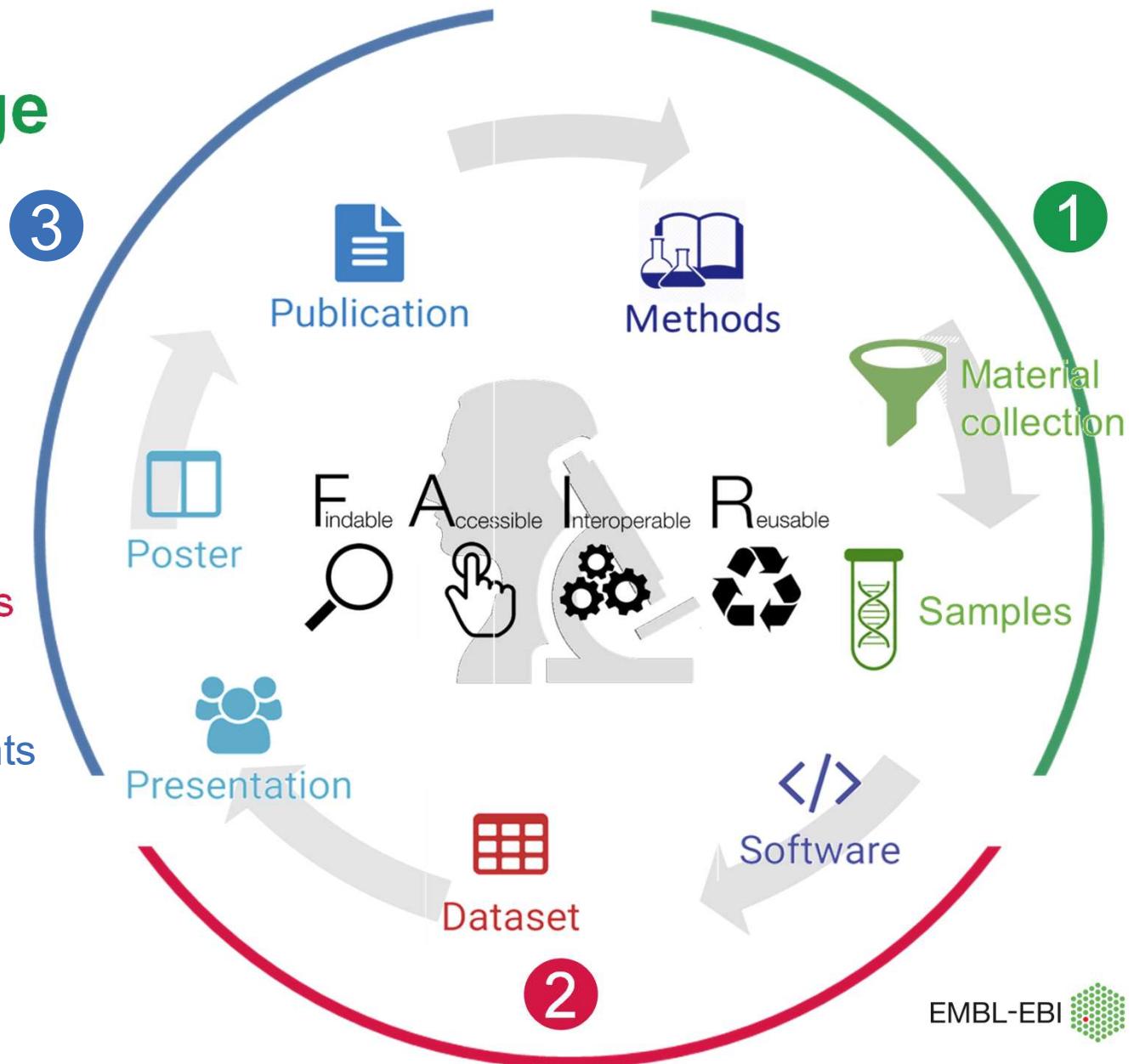
Take home message

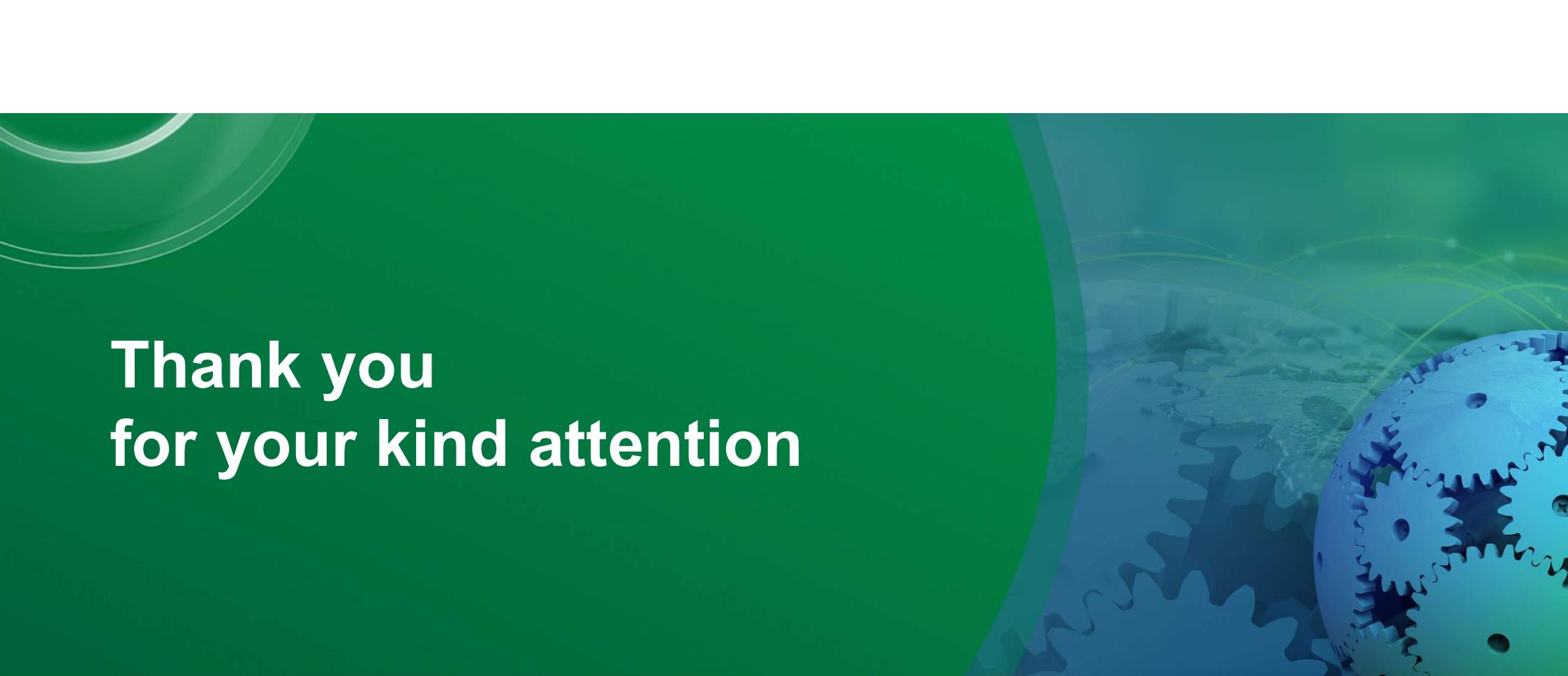
Use standard methods
ADAPT
Collect structured metadata

Deposit metadata & data
EARLY
Progressive closed > open access

Communicate your research intents
EARLY
Seek collaboration

Publish in open access





Thank you for your kind attention

Stéphane PESANT

Senior marine biocurator

pesant@ebi.ac.uk

