

References to known species are restricted to the original descriptions and new combinations. For full references see Monné (2022a, b, c) and Tavakilian & Chevillotte (2022).

The acronyms used in the text are as follows:

MEFEIS: Museu de Entomologia da Faculdade de Engenharia de Ilha Solteira, Universidade Estadual Paulista, Ilha Solteira, São Paulo, Brazil;

MNRJ: Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil;

MPUJ: Museo Javeriano de Historia Natural Lorenzo Uribe, S.J., Pontificia Universidad Javeriana, Bogotá, Colombia;

MZSP: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil;

UNAB: Museo Entomológico, Facultad de Ciencias Agrarias, Universidad Nacional de Colombia, Bogotá, Colombia.

RESULTS

CERAMBYCINAE Latreille, 1802

EBURIINI Blanchard, 1845

Eburodacrys pilicornis Fisher, 1944

(Fig. 1)

Eburodacrys pilicornis Fisher, 1944: 5.

Redescription: Female from Brazil (Figs. 1A-1E): Head capsule dark orangish brown, irregularly lighter ventrally; ventral mouthparts brownish; antennae orangish brown; basal $\frac{2}{3}$ of mandibles dark orangish brown, more brownish on margins, and apical third blackish. Prothorax mostly yellowish brown, lighter on anterior third and central area of prosternum and prosternal process, dark orangish brown on sides of posterior $\frac{2}{3}$ of prosternum and sides of prosternal process, dark brown on anterolateral gibbositities of pronotum, reddish brown on posterocentral gibbosity, anterocentral and posterocentral regions of pronotum, and brownish on center of sides of prothorax. Ventral surface of meso- and metathorax dark orangish brown laterally, mostly yellowish brown centrally. Elytra mostly orangish brown on anterior $\frac{2}{3}$, pale on posterior third; with three sub-elliptical eburneous maculae dorsally, one centrally on base, one slightly oblique centrally, another smaller on posterior third, located more laterally; eburneous maculae partially surrounded with narrow reddish brown band; each elytron with five pale yellow costae, one between scutellum and basal eburneous maculae, gradually inclined toward suture, disappearing after middle, one from apex of basal eburneous macula to apex, interrupted by central eburneous macula, one from base to near apex, interrupted by last eburneous macula, one laterally, from humerus to near apex, fused with apex of outermost dorsal band, another on epipleural margin, from base to apex. Femora orangish brown, except brownish apex and blackish spine of meso- and

metafemora; tibiae orangish brown; tarsi dark reddish brown. Ventrites dark orangish brown laterally, mostly yellowish brown centrally.

Head: Frons somewhat finely rugose-punctate, except smooth anterocentral region; with short, sparse, decumbent white setae, absent on smooth area. Area between antennal tubercles coarsely, sparsely punctate; with short, sparse, decumbent white setae; central area between antennal tubercles and middle of upper eye lobes smooth; area between antennal tubercles and anterior margin of upper eye lobes, coarsely, sparsely punctate laterally; remaining surface of vertex finely, somewhat abundantly punctate, punctures denser, coarser on sides after eyes; vertex with short, decumbent, somewhat sparse white setae, absent on smooth area, sparser on posterocentral region. Area behind eyes somewhat coarsely, densely punctate, transversely striate behind lower eye lobe, except smooth area close to superior region of lower eye lobe; with short, sparse yellowish-white setae behind upper eye lobe, setae distinctly shorter than on vertex, almost glabrous on remaining surface, except short yellowish setae close to inferior margin of lower eye lobe. Genae minutely, abundantly punctate, except smooth apex; with yellowish-white pubescence not obscuring integument, except glabrous smooth area. Area between antenna and eye with dense grayish-white pubescence, gradually whiter toward lower eye lobe. Wide central area of postclypeus with somewhat long white setae directed forward; sides glabrous. Labrum with long yellowish-brown setae posteriorly, glabrous anteriorly. Ventral surface of head smooth, glabrous on posterior half, somewhat finely and abundantly punctate, with long, erect white setae on anterior half, setae shorter, denser, decumbent close to eyes. Antennal tubercles not spiniform apically. Distance between upper eye lobes 0.30 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.57 times distance between outer margins of eyes. Antennae 1.5 times elytral length, reaching elytral apex at apex of antennomere X. Scape moderately narrow, slightly widened toward apex; dorsal sulcus slightly marked; coarsely, abundantly punctate laterally and anterodorsal third, punctures sparser dorsally toward smooth apex, almost smooth ventrally; with short, sparse white setae on punctate region, and long, erect yellowish setae interspersed laterally, especially on posterior third. Pedicel finely punctate on basal half of dorsal and lateral surfaces, and posterior half of ventral surface, smooth remaining surface; with short, sparse, decumbent white setae dorsally and laterally, absent ventrally, and long, erect yellowish setae interspersed on posterior half of sides and ventral surface. Antennomere III finely, sparsely punctate, punctures denser on posterior half of outer side; longitudinally sulcate dorsally and ventrally; with minute, sparse white setae dorsally, short, decumbent, abundant white setae on basal half of outer side, shorter, yellowish on posterior half of outer side, long, decumbent yellowish setae ventrally, and distinct long,



Figure 1. *Eburdacrys pilicornis* Fisher, 1944. (A-E) Female from Brazil: (A) Dorsal habitus; (B) Ventral habitus (C) Lateral habitus; (D) Head, frontal view; (E) Elytral apex. (F-H) Female from Colombia (Cundinamarca): (F) Head, frontal view; (G) Dorsal habitus; (H) Lateral habitus.

erect, abundant yellowish setae on outer margin of ventral surface. Antennomeres IV-V longitudinally carinate dorsally, longitudinally sulcate ventrally; with minute, sparse white setae dorsally, minute, abundant yellowish setae on outer surface, setae on remaining surface as on III. Antennomeres VI-XI with short, abundant yellowish-white pubescence, long, erect, yellowish setae on outer margin of ventral surface, erect setae gradually shorter and sparser toward XI; antennomere XI with distinct constriction after middle. Antennomeres III-X with a few long, erect yellowish setae on dorsal apex. Antennal formula based on length of antennomere III: scape = 0.69; pedicel = 0.12; IV = 0.84; V = 0.84; VI = 0.78; VII = 0.74; VIII = 0.62; IX = 0.55; X = 0.48; XI = 0.64.

Thorax: Prothorax wider than long (including lateral tubercles); lateral tubercles conical, moderately large, located centrally. Pronotum with three distinct tubercles, one on each side of anterior half, slightly elongated, with rounded apex, another on center of posterior half, distinctly less elevated than anterolateral ones; coarsely, somewhat abundantly punctate, punctures denser, forming slightly rugose area close to posterior margin, except smooth tubercles; with short, decumbent, sparse white setae, more abundant on sides of posterior half, absent on tubercles and almost absent centrally; with a few long, erect whitish setae interspersed. Sides of prothorax coarsely, somewhat rugose-punctate; with moderately abundant short white setae, longer than on pronotum, and long, erect setae of same color interspersed. Posterior half of prosternum with sculpturing and setae as on sides of prothorax; anterior half slightly striate-punctate (striae more distinct close to posterior half), and with short, both decumbent and erect white setae. Prosternal process with abundant white setae not obscuring integument; narrowest area 0.3 times procoxal width. Sides of ventral surface of meso- and metathorax with abundant, decumbent white setae not obscuring integument; central area with sparse, decumbent white setae, except glabrous are close to metathoracic disc. Scutellum with somewhat sparse, decumbent white setae. **Elytra:** Coarsely, shallowly, abundant punctate on anterior third and sides of central third, distinctly finer, sparser on remaining surface, especially close to apex; apex almost obliquely truncate; with short, sparse white setae, absent on eburneous maculae, and long, erect, sparse setae of same color interspersed, almost absent on eburneous maculae. **Legs:** Profemora with a few long, erect yellowish setae dorsally and laterally, short, decumbent, sparse white setae ventrally, more abundant basally, and long, erect setae of same color interspersed basally; meso- and metafemora with short, decumbent, sparse whitish setae, and moderately long, bristly yellowish-brown setae interspersed, more abundant ventrally; inner spine of meso- and metatibiae long. Tibiae gradually widened toward apex, more distinctly in metatibiae; with short, sparse yellowish-brown setae, more abundant on posterior third of ventral surface, and long, erect setae of same color interspersed.

Dimensions (mm): Total length, 17.95; prothoracic length, 3.25; anterior prothoracic width, 3.15; posterior prothoracic width, 3.45; maximum prothoracic width, 4.15; humeral width, 4.70; elytral length, 12.10.

Male: Similar to females. Differs by longer antennae (only one male without antennomeres IX-XI measured), 2.0 times elytral length (from base of scape to apex of antennomere VIII), reaching elytral apex at posterior fifth of antennomere VI, and by scape wider and more distinctly sulcate dorsally.

Material examined: VENEZUELA, *Distrito Capital (new state record)*: Caracas, 1 ♀, 03.VI.1959, Bordon leg. (MZSP). *Aragua*: El Limon, 450 m, 1 ♀, 31.V.1964, C.J. Rosales leg. (MZSP); Cagua, 1 ♂, 20.XI.1960, Bordon leg. (MZSP). COLOMBIA, *Cundinamarca (new department record)*: Guaduas, Rancho las Vegas, 05°03'42"N, 74°35'53"W, 1,000 m, 1 ♀, 09-16.V.2016, V. Sinyaev & C. Pinilla leg. (MZSP). *Bolívar*: San Jacinto (San Cristóbal, 80 m, 09°53'16.48"N, 75°94'44.09"W), 1 ♀, 02-05.XII.2019, J.P. Botero, A.F. García, D. Ahumada & H. Vides, leg. (MPUJ). *Santander*: Barichara, 06°38'20"N, 73°13'38"W, 1,294 m, 30.III.1999, 1 ♂, J. Afanador & J. Sánchez leg. (UNAB). BRAZIL (**new country record**), *Mato Grosso do Sul*: Selvíria, UNESP Farm, hand collected, 1 ♀, 06.IV.2019, J. Milan leg. (MEFEIS).

Remarks: Currently, *E. pilicornis* is known from Nicaragua, Costa Rica, Panama, Colombia (Huila, Santander, Bolívar), and Venezuela (Monagas, Anzoátegui, Aragua, Miranda, Guárico) (Monné, 2022a; Tavakilian & Chevillotte, 2022).

All specimens from Venezuela and Colombia that were examined have the apex of the antennal tubercles distinctly acute (somewhat spiniform). The specimen from Brazil has the apex of the antennal tubercles flattened. However, as no other morphological differences were found, we believe that this is just an extreme variation. In the same way, the specimen from Brazil has a very distinct longitudinal pale-yellow band on the curvature of the elytra (Fig. 1C). The specimens from Venezuela and the two males from Colombia (Figs. 1F-1H) do not even have a trace of that band. However, the other male from Colombia has that band slightly distinct. Therefore, we believe that the presence of that band is another feature variable in the species.

HEXOPLONINI Martins, 2006
***Gnomidolon cruciferum* (Gounelle, 1909)**
(Fig. 2A)

Hexoplon cruciferum Gounelle, 1909: 659.

Gnomidolon cruciferum; Martins, 1967: 202.

Remarks: *Gnomidolon cruciferum* was described from Brazil (Goiás). Currently it is also known from Peru, Bolivia, and the Brazilian states of Paraíba and Bahia (Monné, 2022a; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Minas Gerais (**new state record**): Nova Porteirinha, Khaya Woods, *Khaya grandifoliola* stand planted Feb/2009, 15°41'04.15"S, 43°17'41.63"W, ethanol-baited FIT, 1 ♀, 21.XI.2018, L.G.A. Oliveira leg. (MEFEIS).

TROPIDINI Martins & Galileo, 2007

Note: Although the current catalogs and checklists of Cerambycidae report the name of the tribe as Neobidionini Monné, 2012 (e.g., Monné, 2022a; Tavakilian & Chevillotte, 2022), the correct is Tropidini, as indicated by Bouchard & Bousquet (2020).

COMPSINA Martins & Galileo, 2007

Microibidion bimaculatum

Mehl, Galileo, Martins & Santos-Silva, 2015
(Fig. 2B)

Microibidion bimaculatum Mehl, Galileo, Martins & Santos-Silva, 2015: 38.

Remarks: *Microibidion bimaculatum* was described from Paraguay. Currently it is also known from Bolivia (Monné, 2022a; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL (**new country record**), Minas Gerais: São Roque de Minas, Florestas da Canastra, Fazenda Taquaril, *Khaya grandifoliola* stand planted Feb/2010, ethanol-baited FIT, 20°06'30.31"S, 46°27'03.61"W, 1 ♀, 28.IX.2018, L.S. Covre leg. (MEFEIS).

PIEZOCERINI Lacordaire, 1868

Piezocera flavipennis (Zajciw, 1970)
(Fig. 2C)

Pyrgotes flavipennis Zajciw, 1970: 593.
Piezocera flavipennis; Martins, 1976: 272.

Remarks: According to Martins (1976) on *Piezocera flavipennis* (translated): "Absence of a demarcated anterior pronotal sulcus separates *flavipennis* from the unicolorous species examined above, where this sulcus is visible," and in the key, "Pronotum without trace of anterior longitudinal sulcus ...," leading to *P. flavipennis*. Still according to Martins (1976), he examined the holotype. The holotype belonged to the MNRJ and was destroyed in a fire. Posteriorly, Martins (2003) repeated the same information about the pronotal shape of *Piezocera flavipennis*. However, this information does not correspond to the shape of the anterior area of the pronotum of this species. Zajciw (1970) reported (translated): "Prothorax narrow ... in the middle of the anterior area with a longitudinal groove resembling that of *Piezocera nodicollis* Melz., 1934 ..." In fact, it is possible to see the distinct sulcus

in the photograph of the holotype taken by Steven W. Lingafelter (Fig. 11).

Piezocera monochroa Bates, 1885 (Figs. 2D-2E)

Piezocera monochroa Bates, 1885: 258.
Piezocera serraticollis Linell, 1897: 394. **Syn. nov.**

Remarks: Comparing a paralectotype male of *P. monochroa* (Figs. 2D-2E) with specimens identified as *P. serraticollis*, from Mexico and the USA (Figs. 2F-2G), we could not find any morphological difference. Furthermore, the current geographical distribution of both species is overlapped in the Mexican state of Chiapas, reinforcing the indication that the two names correspond to the same species.

Martins (1976) reported about *P. serraticollis* (translated): "The redescription given below is restricted to topotypic specimens. Other specimens, in some cases intermediate between *serraticollis* and *monochroa*, are discussed in the item variations. Only the examination of very abundant material from Mexico and Central America will be able to elucidate the status of this species;" "Three specimens from Almolonga, Veracruz, Mexico (BMNH, MNHN) were included by Bates (1885: 258) in the type series of *Piezocera monochroa*; it seems to me that they actually belong to *serraticollis*. They are different from topotypic specimens: elytra apex with less prolonged lateral projection; erect setae on dorsal surface almost the same length as the scape." This later affirmation contradicts one of the differences between *P. serraticollis* and *P. monochroa* pointed out by him: erect dorsal setae shorter than length of the scape in *P. serraticollis*, and longer than the scape in *P. monochroa*. Furthermore, the elytral apex is variable in other species of the genus, which makes it evident that the same also occurs in *P. monochroa*. To reinforce that the differences pointed out by Martins (1976) are just specific variations, it is interesting to translate another part of his comments on variations in *P. serraticollis*: "Specimen from Salina Cruz, Oaxaca, Mexico: erect dorsal setae very elongated; spicule of the prothorax distinct; erect setae very abundant on entire body..." The number of erect setae was a feature used by him to separate *P. serraticollis* from *P. monochroa* in the key: erect setae more abundant in the latter than in the former. Another feature used in the key by Martins (1976) was the sexual punctuation on the abdomen: present in males of *P. serraticollis*; absent in males of *P. monochroa*. However, in the redescription of *P. monochroa*, he reported that perhaps they are absent in males of this species. Therefore, based on this evidence of intraspecific morphological variation, *P. serraticollis* is synonymized with *P. monochroa*.

Material examined: 1 ♀, without labels. UNITED STATES OF AMERICA, Texas: Brownsville, 1 ♀, VI.1901, no collector indicated (MZSP). MEXICO, no further data, 1 ♀, no date indicated, J. Flohr leg. (MZSP). GUATEMALA, Baja Verapaz: San Gerónimo, paralectotype male, Champion (MZSP).

Key to species of *Piezocera* Audinet-Serville, 1834 (adapted from Martins, 2003)

Note: The features used in the key to separate the species were verified only in *P. flavipennis*, *P. rufula*, and *P. monochroa*, in order to correctly include them in the key. It will be necessary to do a revision of the genus to verify the features in all species.

1. Pronotum without sulcus on anterocentral region 2
- Pronotum with sulcus on anterocentral region 3
- 2(1). Meso- and metafemoral clubs abruptly widened from peduncle. French Guiana, Brazil (Amazonas, Rondônia) *P. aenea* (Bates, 1867)
- Meso- and metafemoral clubs gradually widened from peduncle. United States of America (Texas), Mexico (Veracruz, Hidalgo, Chiapas, Oaxaca, Quintana Roo), Guatemala, Honduras, Nicaragua, Costa Rica *P. monochroa* Bates, 1885
- 3(1). Elytra unicolorous, from brown to orangish-red, without longitudinal bands 4
- Elytra bicolorous, with longitudinal dark or green band 9
- 4(3). Antennomere III not expanded on outer apex 5
- Antennomere III expanded on outer apex 6
- 5(4). Margins of anterocentral sulcus of pronotum elevated, distant from each other and sloped toward anterior margin; central tubercle of pronotum not microsculptured, shiny. Brazil (Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul), Paraguay *P. nodicollis* Melzer, 1934
- Margins of anterocentral sulcus of pronotum less elevated, closer to each other and reaching anterior margin; central tubercle of pronotum microsculptured. Trinidad and Tobago *P. rufula* Martins & Galileo, 2010
- 6(4). Elytral apex obliquely truncate or slightly emarginate and unarmed; antennomere III distinctly expanded toward outer apex. Bolivia, Brazil (Mato Grosso do Sul, São Paulo) *P. costula* Martins, 1976 (part)
- Elytral apex with spiniform projection or with distinct spine on outer apex; antennomere III somewhat expanded toward outer apex 7
- 7(6). Pronotum mostly shiny. Peru, Bolivia, Brazil (Pará) *P. flavipennis* (Zajciw, 1970)
- Pronotum opaque 8
- 8(7). Area between upper eye lobes with two brushes of setae; anterocentral region of pronotum microsculptured as remaining surface of pronotum; prothorax with distinct spiniform lateral tubercle. Brazil (Alagoas, Sergipe, Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro) *P. araujosilvai* Melzer, 1935
- Area between upper eye lobes without brushes of setae; anterocentral region of pronotum shiny; prothorax without lateral spine. Colombia (Cauca, Valle del Cauca) *P. silvia* Galileo & Martins, 2000
- 9(3). Antennae of males with sexual setae; elytral setae short, not arranged in rows; antennomere III with spicule on inner apex and slightly expanded toward outer apex; antennomeres IV-V with spicule on inner apex; elytral apex concave, with short spine on outer apex. Venezuela (Distrito Federal, Miranda) .. *P. advena* Martins, 1976
- Antennae of males without sexual setae; elytral setae long, arranged in five longitudinal rows; antennomere III expanded on outer apex, without spicule on inner apex; elytral apex with different shape 10
- 10(9). Antennomere III projected on both apices; pronotum densely microsculptured; metatibiae projected on outer apex 11
- Antennomere III not projected on inner apex; pronotum sparsely microsculptured 12
- 11(10). Elytral apex distinctly projected on outer angle; dark longitudinal band on elytra well-marked; metatibiae with acute projection on outer apex. Brazil (Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul), Paraguay *P. bivittata* Audinet-Serville, 1834
- Elytral apex obliquely truncate; dark longitudinal band on elytra slightly contrasting; metatibiae without apical projection. Bolivia, Brazil (Mato Grosso do Sul, São Paulo) *P. costula* Martins, 1976 (part)
- 12(10). Upper eye lobes with three rows of ommatidia; pronotum with granules, without basal gibbositities; abdomen of males without sexual punctures; tarsal claws reduced in length. Venezuela (Miranda) *P. gratiosa* Lameere, 1893
- Upper eye lobes with four rows of ommatidia; abdomen of males with sexual punctures; tarsal claws normal. Bolivia, French Guiana, Brazil (Pará, Paraíba, Bahia, Mato Grosso) *P. ataxia* Martins, 1976

TILLOMORPHINI Lacordaire, 1868

Tilloglomus spectabile Martins, 1975

(Fig. 2H)

Tilloglomus spectabile Martins, 1975: 19.

Tilloglomus spectabilis; Monné, 1993: 76 (cat.).

Remarks: *Tilloglomus spectabile* was described from the Brazilian state of Santa Catarina. Monné (2022a) and Tavakilian & Chevillotte (2022) listed the species as known only from this state. However, Saldanha et al. (2021) listed the species from the Brazilian state of Rio Grande do Sul.

Monné (1993) listed the species as *Tilloglomus spectabilis*, a condition kept until today. However, as indicated in the original description, the correct is *T. spectabile*, because “glomus” (Latin) is neuter gender.

Material examined: BRAZIL, Paraná (new state record):

Colombo, EMBRAPA Florestas campus, mixed ombrophilous forest (Araucaria forest) fragment, 25°19'47.9"S, 49°09'40.2"W, ethanol-baited FIT, 1 ♀, 03.VIII.2018, D.L. Queiroz leg. (MEFEIS). Minas Gerais (new state record): Gonçalves (Cantagalo), 1 ♂, no date indicated, E.P. Teixeira leg. (MZSP 52182).

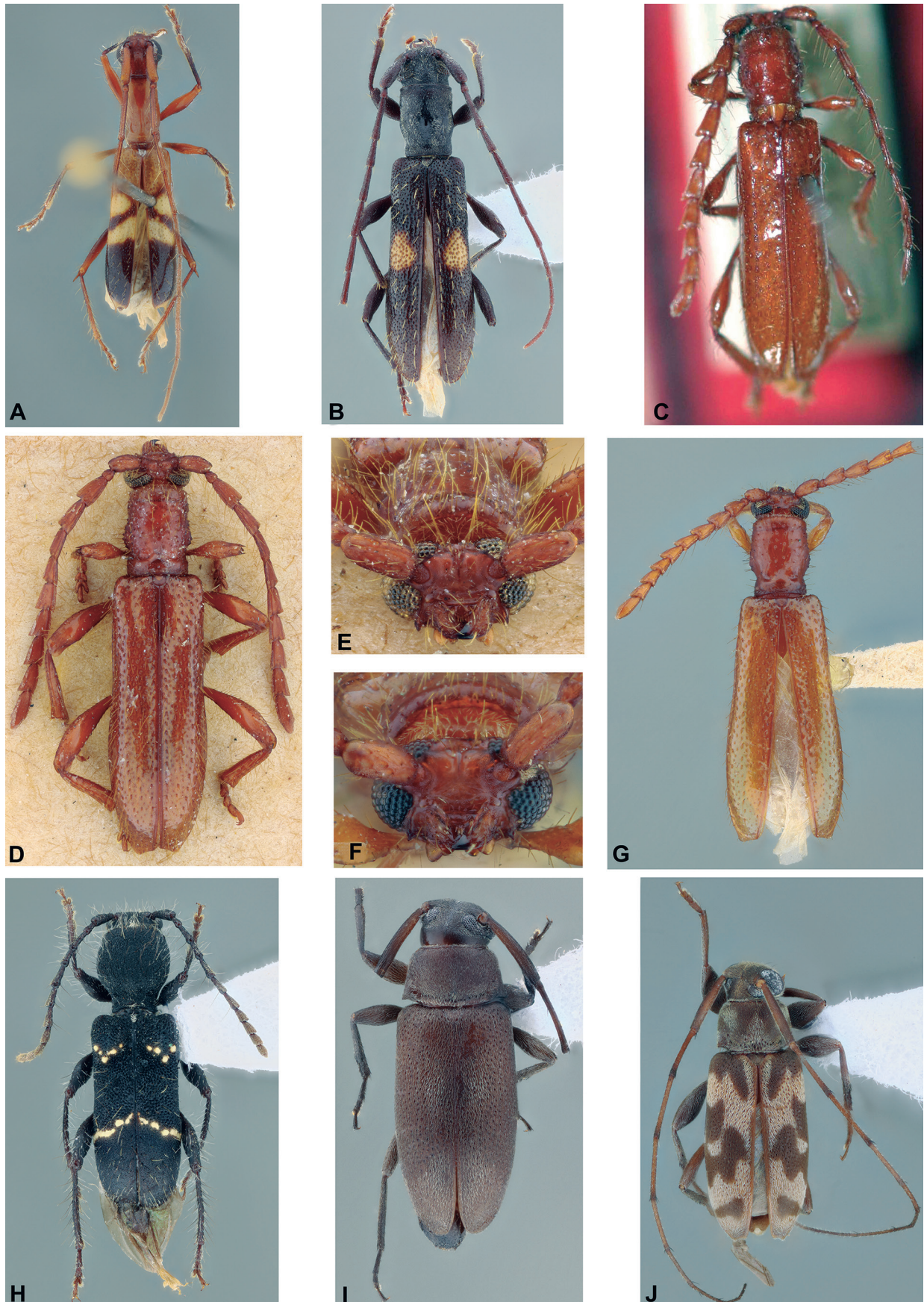


Figure 2. (A) *Gnomidolon cruciferum* (Gounelle, 1909), female from Brazil (Minas Gerais), dorsal habitus. (B) *Microibidion bimaculatum* Mehl et al., 2015, female from Brazil (Minas Gerais), dorsal habitus. (C) *Pyrgotes flavipennis* Zajciw, 1970, holotype, dorsal habitus, by Steven W. Lingafelter. (D-E) *Piezocera monochroa* Bates, 1885, paralectotype male from Guatemala: (D) Dorsal habitus; (E) Head, frontal view. (F-G) *Piezocera monochroa* female from the USA (Texas): (F) Head, frontal view; (G) Dorsal habitus. (H) *Tilloglomus spectabile* Martins, 1975, female from Brazil (Paraná), dorsal habitus. (I) *Lepturges (Lepturges) centralis* Monné, 1978, female from Brazil (Mato Grosso do Sul), dorsal habitus. (J) *Lepturges (Lepturges) mattogrossis* Gilmour, 1962, male from Brazil (Mato Grosso do Sul), dorsal habitus.

LAMIINAE Latreille, 1825
ACANTHOCININI Blanchard, 1845

Leptostylus perniciosus Monné & Hoffmann, 1981
(Fig. 4A)

Leptostylus perniciosus Monné & Hoffmann, 1981: 259.

Remarks: *Leptostylus perniciosus* was described from Suriname, Bolivia, Brazil (Roraima, Amazonas, Pará, Paraíba, Bahia, Minas Gerais, Espírito Santo, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul), Argentina, and Paraguay. Currently, it is also known from French Guiana and the Brazilian states of Sergipe and Maranhão (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Ceará (new state record): Redenção, Fazenda Experimental Piroás, UNILAB, caatinga fragment, 04°09'18.06"S, 38°47'50.17"W, ethanol-baited FIT, 1 ♀, 13.X.2016, J.G.L. Moraes leg. (MEFEIS); 1 ♂, 10.XI.2016, J.G.L. Moraes leg. (MEFEIS).

Lepturges (Lepturges) centralis Monné, 1978
(Fig. 2I)

Lepturges centralis Monné, 1978: 3.

Remarks: *Lepturges (Lepturges) centralis* was described from Brazil (Minas Gerais). Currently, it is also known from the Brazilian state of Maranhão (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Mato Grosso do Sul (new state record): Selvíria, UNESP Farm, cerradão fragment in advanced stage of regeneration, ethanol-baited FIT, 20°23'02.25"S, 51°24'45.65"W, 1 ♀, 20.IV.2021, G.C. Pinheiro leg. (MEFEIS).

Lepturges (Lepturges) mattogrossis Gilmour, 1962
(Fig. 2J)

Lepturges mattogrossis Gilmour, 1962a: 555.

Remarks: *Lepturges (Lepturges) mattogrossis* was described from the Brazilian state of Mato Grosso. Currently, it is also known from the Brazilian states of Bahia and São Paulo (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Mato Grosso do Sul (new state record): Inocência, Fazenda Laguna, *Khaya senegalensis* stand planted Mar/2016, ethanol-baited FIT, 19°43'19.9"S, 51°59'21.4"W, 1 ♂, 05.V.2018, M.B.C. Ramos leg. (MZSP 52183, formerly MEFEIS). Goiás (new state record): Niquelândia, Fazenda Rio do Peixe, *Khaya grandiloliola* stand planted Dec/2014, ethanol-baited FIT, 14°25'05.41"S, 48°46'59.40"W, 1 ♂, 19.X.2018, L.M. Santos leg. (MEFEIS).

Lepturges (Lepturges) luanae sp. nov.
(Fig. 3)

Description: Holotype female: Integument mostly brown, with irregular dark-brown or dark reddish-brown areas; anterior area of anteclypeus yellowish-brown; labrum brownish basally, gradually pale yellowish brown toward anterior margin; maxillary palpomeres I-III brown, and IV yellowish brown; labial palpomeres I-II dark yellowish brown, III light yellowish brown; antennomeres III-VI dark brown on apex; antennomeres VII-X dark brown; antennomere XI dark brown with reddish-brown apex. Elytra gradually lighter toward apex. Tibiae gradually dark brown toward apex; tarsomeres mostly dark brown. Ventrites 1-4 dark brown laterally, mostly reddish brown centrally; 5 mostly dark brown.

Head (Figs. 3A, 3D): Frons finely, abundantly punctate; with abundant yellowish-white pubescence not obscuring integument (pubescence whiter depending on light intensity), a short, erect setae of same color interspersed laterally; with a few long, erect dark-brown setae close to eyes. Vertex and area behind upper eye lobes with sculpturing and pubescence as on frons, except smooth and glabrous median groove; with a few long, erect dark-brown setae close to eyes. Area behind lower eye lobes almost smooth; with dense yellowish-white pubescent band close to eye, glabrous on remaining surface. Antennal tubercles with sculpturing and pubescence as on frons, except almost glabrous area of dorsal base. Genae finely, somewhat rugose-punctate, except smooth apex; with moderately sparse yellowish-white pubescence, except glabrous smooth area; with a few long, erect dark brown setae toward ventral surface. Gula mentum slightly transversely striate, glabrous, except narrow, depressed, finely punctate, with yellowish-white pubescence not obscuring integument anterior area. Postclypeus with abundant, bristly yellowish-white pubescence not obscuring integument, and long, erect dark-brown setae interspersed. Labrum with somewhat sparse yellowish-white pubescence on posterior half, almost glabrous on anterior half; with long, erect dark brown setae interspersed on posterior half. Distance between upper eye lobes 0.25 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.50 times distance between outer margins of eyes. Antennae 3.1 times elytral length, reaching elytral apex at middle of antennomere VI. Scape, pedicel, and flagellomeres with whitish pubescence not obscuring integument; scape with a few long, erect blackish setae on ventral apex; pedicel with long, erect, thick black setae ventrally; antennomeres III-VII with long, erect, thick, sparse black setae ventrally, sparser from IV; antennomeres VI-X with a few long, erect, thick black setae dorsally; antennomeres III-X with a few long, erect, black setae apically; antennomeres IV-XI with short, erect yellowish setae interspersed dorsally and laterally. Antennal formula based on length of antennomere III: scape = 1.38; pedicel = 0.14; IV = 1.14; V = 1.14; VI = 1.19; VII = 1.14; VIII = 1.14; IX = 1.19; X = 1.38; XI = 2.14.

Thorax (Figs. 3A, 3C): Prothorax distinctly wider than long; sides gradually widened toward lateral tubercles, which are located on posterolateral angles. Pronotum very finely, densely punctate, except arched row of coarse and deep punctures near posterior margin; with abundant, dense yellowish-white pubescence, forming wide longitudinal band centrally and slightly distinct longitudinal band on each side, both from base to apex, and distinctly shorter, somewhat yellowish pubescence between longitudinal bands. Sides of prothorax very finely, abundantly punctate; with abundant yellowish-white pubescence not obscuring integument (pubescence whiter depending on light intensity). Prosternum with dense yellowish-white pubescence laterally, pubescence whiter, distinctly sparser centrally. Prosternal process with whitish pubescence not obscuring integument; strongly narrowed centrally, with narrowest area 0.1 times procoxal width. Ventral surface of meso- and metathorax with abundant whitish pubescence not obscuring integument. Scutellum with abundant yellowish-white pubescence partially obscuring integument.

Elytra (Figs. 3A, 3C): Coarsely, abundantly punctate on anterior third, punctures gradually finer and sparser toward apex; each elytron with five longitudinal white pubescent bands dorsally, one on anterior third close to suture, one laterally on anterior third, one on middle, longest, located more closer to suture than side, one laterally from just before middle to posterior quarter, another on posterior third close to suture; remaining surface with abundant yellowish-white pubescence not obscuring integument; apex truncate. **Legs (Figs. 3B):** Femora with abundant whitish pubescence partially obscuring integument. Tibiae with abundant whitish pubescence not obscuring integument dorsally, pubescence gradually dark yellowish brown toward ventral surface, except posterior half with bristly, dark brown pubescence; with short, erect, thick, black setae interspersed on meso- and metatibiae. Metatarsomere I 1.4 times longer than II-III together.

Abdomen (Figs. 3B): Ventrites with abundant whitish pubescence not obscuring integument, denser laterally on ventrites 1-4. Apex of ventrite 5 concave.



Figure 3. *Lepturges (Lepturges) luanae*, holotype female: (A) Dorsal habitus; (B) Ventral habitus; (C) Lateral habitus; (D) Head, frontal view.

Dimensions (mm): Total length, 5.30; prothoracic length, 0.80; anterior prothoracic width, 1.10; posterior and maximum prothoracic width, 1.55; humeral width, 1.80; elytral length, 3.80.

Type material: Holotype female from BRAZIL, Goiás: Niquelândia, Fazenda Rio do Peixe, *Khaya grandiloliola* stand planted Dec/2012, ethanol-baited FIT, 14°24'51.39"S, 48°46'52.67"W, 11.XI.2018, L.M. Santos leg. (MZSP 52178, formerly MEFEIS).

Etymology: The name is given in honor of Luana Souza Covre, for her outstanding contributions in the survey of Brazilian Cerambycidae.

Remarks: *Lepturges (Lepturges) luanae* sp. nov. is similar to *L. (L.) funereus* Monné, 1976 (see photographs on Bezark, 2022), but differs as follows: lateral tubercles of the prothorax located on posterolateral angles; elytra with five longitudinal whitish pubescent bands, with similar width; elytral apex truncate. In *L. (L.) funereus*, lateral tubercles of the prothorax not located on posterolateral angles; elytra with seven pubescent maculae with different shape, size, and width; elytral apex rounded. The new species differs from *L. (L.) fisheri* Melzer, 1928 (see photographs on Bezark, 2022), by the distance between upper eye lobes shorter than twice maximum diameter of the scape (equal to twice maximum diameter of the scape in *L. (L.) fisheri*), wider upper eye lobes (narrower in *L. (L.) fisheri*), elytra with five longitudinal whitish pubescent bands dorsally (six with different shape and position in *L. (L.) fisheri*), and elytra not obliquely truncate at apex (obliquely truncate, forming acute outer angle in *L. (L.) fisheri*).

***Urgleptes villiersi* Gilmour, 1962
(Fig. 4B)**

Urgleptes villiersi Gilmour, 1962b: 21.

Remarks: *Urgleptes villiersi* was described from Brazil without any further details. Currently, it is known from Brazil (Minas Gerais, Rio de Janeiro, Paraná) and Paraguay (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Santa Catarina (new state record): Água Doce, Sítio Boa Vista, 27°00'29.8"S 51°30'58.0"W, mixed ombrophilous forest fragment, ethanol-baited FIT, 1 ♂, 31.XII.2011, E. Foelkel leg. (MEFEIS).

**ACROCININI Swainson, 1840
Oreodera bituberculata Bates, 1861
(Fig. 4C)**

Oreodera bituberculata Bates, 1861: 51.

Remarks: *Oreodera bituberculata* was described based on syntypes from Brazil (Pará, Amazonas) and French Guiana. Currently, it is known from Colombia, Ecuador, French Guiana, Guyana, Bolivia, Brazil (Roraima, Amazonas, Pará, Rondônia, Mato Grosso, Goiás, Maranhão) (Monné, 2022b; Tavakilian & Chevillotte, 2022). According to Bates (1861), he "took this species at Ega [now Tefé, Brazilian state of Amazonas] and on the banks of the Tapajós." Tapajós River rises in the Brazilian state of Mato Grosso and flows into the Amazonas River, still in the Brazilian state of Pará. Although Henry W. Bates explored and sailed along the Tapajós River, he did not reach the region that today comprises the state of Mato Grosso (Bates, 1863).

Material examined (only new record listed): BRAZIL, Acre (new state record): Rio Branco, Parque Zoobotânico da Universidade Federal do Acre, bamboo-suppressed open ombrophilous forest (open terra firme forest), ethanol-baited FIT, 09°56'57.93"S, 67°52'12.29"W, 1 ♂, 01.XII.2019, F.W.S. Silva leg. (MEFEIS).

**APOMECCYNINI Thomson, 1860
Rosalba smaragdina (Breuning, 1940)
(Fig. 4D)**

Aletretia smaragdina Breuning, 1940b: 153.
Rosalba smaragdina; Breuning, 1960: 174 (cat.).

Remarks: *Rosalba smaragdina* was described from Brazil (Rio de Janeiro). Currently, it is known from Brazil (Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Santa Catarina) (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Paraná (new state record): Curitiba, Parque Barreirinha, secondary mixed ombrophilous forest in intermediary stage of regeneration, ethanol-baited FIT, 25°25'29.56"S, 49°18'37.43"W, 1 ♀, 03.IV.2019, A.S. Santos leg. (MEFEIS).

**COLOBOTHEINI Thomson, 1860
Colobothea rubroornata Zajciw, 1962
(Fig. 4E)**

Remarks: *Colobothea rubroornata* was described from Brazil (Espírito Santo, Rio de Janeiro, Paraná, Santa Catarina) and Paraguay. Currently, it is also known from Bolivia and the Brazilian states of Minas Gerais and São Paulo (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, Bahia (new state record): Barra do Choça, Fazenda do Miro, montane seasonal semideciduous forest fragment, ethanol-baited FIT, 14°52'33.59"S, 40°41'45.25"W, 1 ♂, XI.2017, R.C.S. Lima leg. (MEFEIS).

COMPOSOMATINI THOMSON, 1857
***Aerenea subimpetiginosa* Breuning, 1948**
(Fig. 4G)

Aerenea subimpetiginosa Breuning, 1948: 29.

Remarks: *Aerenea subimpetiginosa* was described from the Brazilian state of Pernambuco. Currently, it is also known from the Brazilian state of Rio Grande do Norte (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined: BRAZIL, *Minas Gerais (new state record)*: Janaúba, Bico da Pedra, 675 m, submontante deciduous seasonal forest, ethanol-baited FIT, 15°50'13.64"S, 43°16'03.59"W, 1 ♀, 22.V.2020, C.A.R. Matrangolo leg. (MZSP 52179, formerly MEFEIS).

DESMIPHORINI Thomson, 1860
***Cicuiara nitidula* (Bates, 1866)**
(Fig. 4F)

Exocentrus nitidulus Bates, 1866a: 191.
Cosmotomidius ? nitidulus; Gilmour, 1965: 594 (cat.).
Cicuiara nitidula; Galileo & Martins, 1996: 875.

Remarks: *Cicuiara nitidula* was described from Brazil (Pará). Currently, it is also known from Bolivia and the Brazilian state of Mato Grosso do Sul (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, *Goiás (new state record)*: Niquelândia, Fazenda Rio do Peixe, *Khaya grandiloliola* stand planted Dec/2014, ethanol-baited FIT, 14°24'51.39"S, 48°46'52.67"W, 2 ♀♀s, 19.X.2018, L.M. Santos leg. (MEFEIS); *Khaya grandiloliola* stand planted Oct/2013, ethanol-baited FIT, 14°25'51.26"S, 48°46'56.01"W, 1 ♀, 19.X.2018, L.M. Santos leg. (MEFEIS).

***Desmiphora (Desmiphora) crocata* Melzer, 1935**
(Fig. 4H)

Desmiphora crocata Melzer, 1935: 186.
Desmiphora (Desmiphora) crocata; Breuning, 1963: 512 (cat.).

Remarks: *Desmiphora (Desmiphora) crocata* from the Brazilian state of Goiás. According to Tavakilian & Chevillotte (2022), this species occurs in the Brazilian states of Maranhão, Goiás, and Minas Gerais. Monné (2022b) also listed the Brazilian state of São Paulo. We do not know who previously listed this last Brazilian state.

Material examined (only new record listed): BRAZIL, *Mato Grosso do Sul (new state record)*: Selvíria, UNESP Farm, cerradão fragment in advanced stage of regeneration, ethanol-baited FIT, 20°22'57.67"S, 51°24'41.76"W, 1 ♀, 30.X.2014, F.L.L. Leonel leg. (MEFEIS).

***Estola acricula* Bates, 1866**
(Fig. 4I)

Estola acricula Bates, 1866b: 293.
Estola multilineata Breuning, 1940a: 62.

Remarks: *Estola acricula* and *Estola multilineata* were both described from Brazil (Rio de Janeiro). Breuning (1949) synonymized *E. multilineata* with *E. acricula*. Tavakilian & Chevillotte (2022) listed only the Brazilian state of Rio de Janeiro as a place where this species occurs. However, Monné (2022b) also listed the Brazilian states of São Paulo and Santa Catarina. We do not know who previously listed these last two Brazilian states, although the new state record included herein makes it very probable that it really occurs in both states.

Material examined (only new record listed): BRAZIL, *Paraná (new state record)*: Curitiba, Parque Barreirinha, secondary mixed ombrophilous forest in intermediary stage of regeneration, 25°21'40.06"S, 49°15'32.83"W, ethanol-baited FIT, 1 ♀, 03.IV.2020, A.S. Santos leg. (MEFEIS).

FORSTERIINI Tippmann, 1960
***Gisostola bahiensis* Martins & Galileo, 1988**
(Fig. 5A)

Gisostola bahiensis Martins & Galileo, 1988: 220.

Remarks: *Gisostola bahiensis* was described and remains known only from the Brazilian state of Bahia (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, *Minas Gerais (new state record)*: Janaúba, Bico da Pedra, 15°50'07.62"S, 43°16'05.03"W, submontante deciduous seasonal forest, ethanol-baited FIT, 1 ♀, 22.V.2020, C.A.R. Matrangolo leg. (MZSP 52181, formerly MEFEIS).

ONCIDERINI Thomson, 1860
***Hypsioma chapadensis* Dillon & Dillon, 1945**
(Fig. 5B)

Hypsioma chapadensis Dillon & Dillon, 1945: 22.

Remarks: *Hypsioma chapadensis* was described from and remains known from Brazil (Mato Grosso) and Paraguay (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, *Minas Gerais (new state record)*: Janaúba, *Khaya grandifoliola* stand planted May/2012, 15°46'20.97"S, 43°18'42.98"W, ethanol-baited FIT, 1 ♂, 21.VI.2019, L.G.A. Oliveira leg. (MEFEIS). *São Paulo (new state record)*: Santana da Ponte Pensa, Chácara São Judas Tadeu, riparian forest fragment, 20°13'19.1"S, 50°48'15.6"W, ethanol-baited FIT, 1 ♂, 15.XII.2012, J.C.P. Silva leg. (MEFEIS).

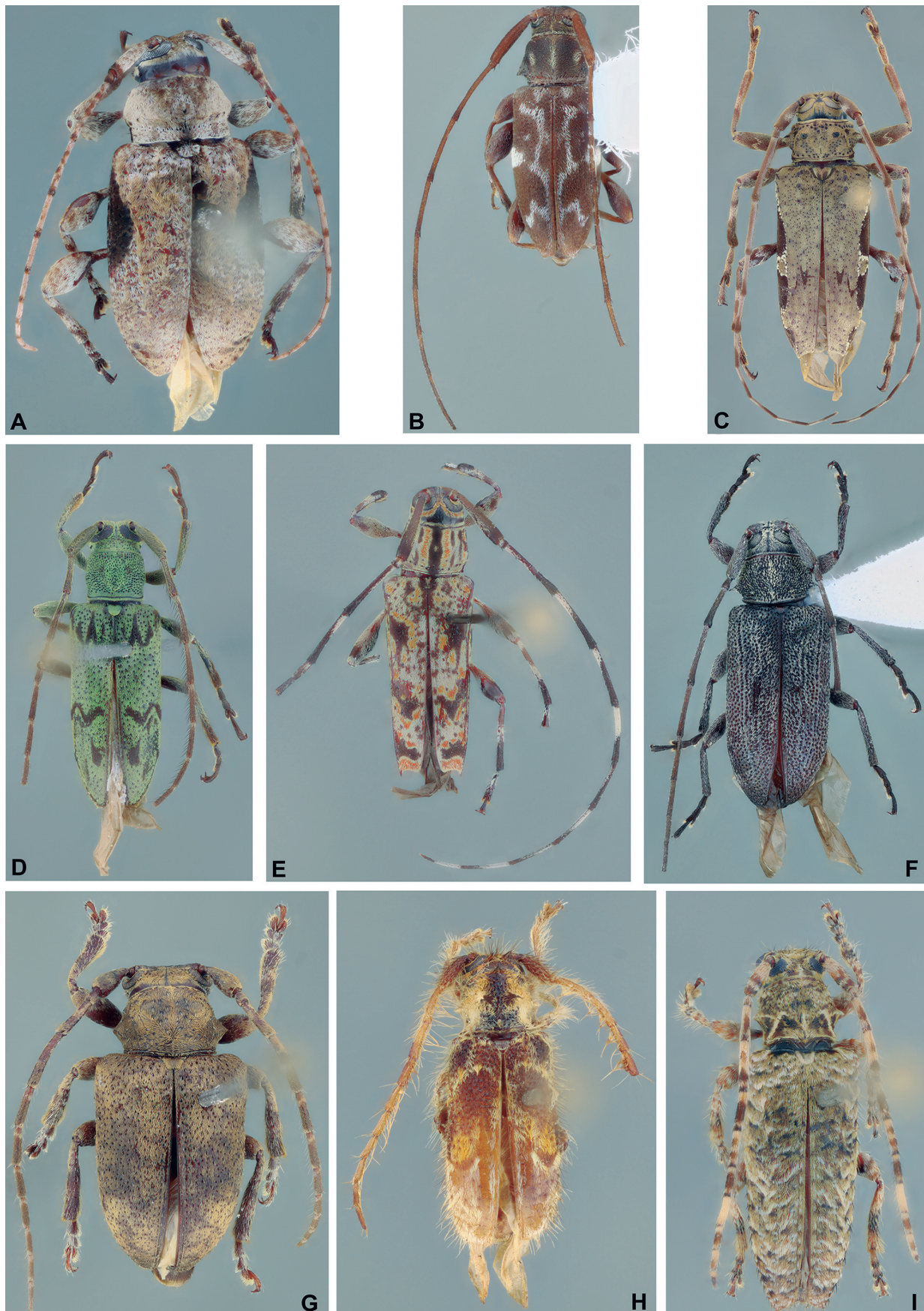


Figure 4. Dorsal habitus. (A) *Leptostylus perniciosus* Monné & Hoffmann, 1981, male from Brazil (Ceará). (B) *Urgleptes villiersi* Gilmour, 1962, male from Brazil (Santa Catarina). (C) *Oreodera bituberculata* Bates, 1861, male from Brazil (Acre). (D) *Rosalba smaragdina* (Breuning, 1940), female from Brazil (Paraná). (E) *Colobothea rubroornata* Zajciw, 1962, male from Brazil (Bahia). (F) *Ciciuara nitidula* (Bates, 1866), female from Brazil (Goiás). (G) *Aerenea subimpetiginosa* Breuning, 1948, female from Brazil (Minas Gerais). (H) *Desmiphora* (*Desmiphora*) *crocata* Melzer, 1935, female from Brazil (Mato Grosso do Sul). (I) *Estola acricula* Bates, 1866, female from Brazil (Paraná).

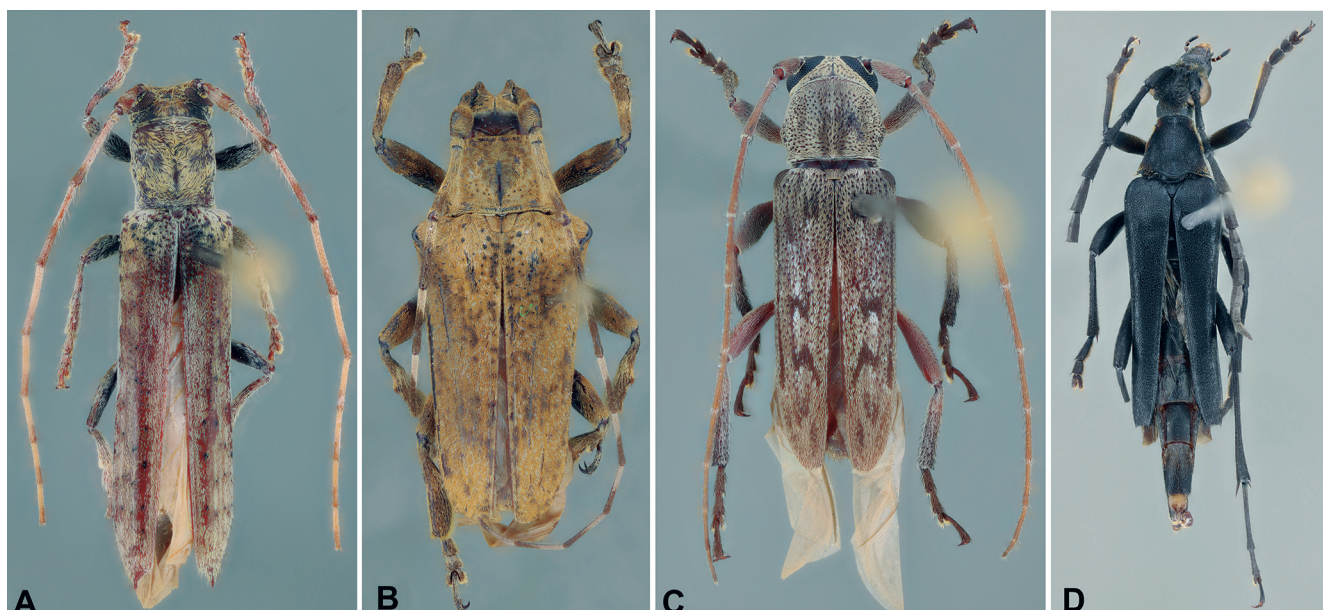


Figure 5. Dorsal habitus. (A) *Gisostola bahiensis* Martins & Galileo, 1988, female from Brazil (Minas Gerais). (B) *Hypsioma chapadensis* Dillon & Dillon, 1945, male from Brazil (Minas Gerais). (C) *Lypsimena fuscata* Haldeman, 1847, male from Brazil (Mato Grosso do Sul). (D) *Strangalia flavocincta* (Thomson, 1861), male from Brazil (Rio Grande do Sul).

POGONOCERINI Mulsant, 1839
***Lypsimena fuscata* Haldeman, 1847**
(Fig. 5C)

Lypsimena fuscata Haldeman, 1847: 54.
Alloeoscelis leptis Bates, 1885: 358.
Lypsimena californica Horn, 1885: 194.
Lypsimena brasiliensis Aurivillius, 1922: 164.
Estoloderces navarroi Melzer, 1928: 148.

Remarks: *Lypsimena fuscata* has a large geographical distribution. Currently it is known from the United States of America (New York, North Carolina, Florida, Georgia, Texas, Alabama, California), Mexico (Jalisco, Morelos), Honduras, Guatemala, Nicaragua, Costa Rica, Panama, Cuba, Jamaica, Puerto Rico, Bahamas, Cayman Islands, Hispaniola, Colombia, Venezuela, French Guiana, Brazil (Roraima, Maranhão, Piauí, Rio Grande do Norte, Paraíba, Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul), Argentina (Salta, Tucumán, Chaco, Corrientes), Bolivia (Santa Cruz), Paraguay, Uruguay (Monné, 2022b; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, *Mato Grosso do Sul (new state record):* Inocência, Fazenda Laguna, *Khaya senegalensis* stand planted Mar/2016, 19°43'19.9"S, 21°59'21.4"W, ethanol-baited FIT, 1 ♂, 04.VIII.2018, L.S. Covre leg. (MEFEIS).

LEPTURINAE Latreille, 1802
LEPTURINI Latreille, 1802
***Strangalia flavocincta* (Thomson, 1861)**
(Fig. 5D)

Ophistomis flavocinctus Thomson, 1861: 155.

Ophistomis tristis Melzer, 1922: 5.
Ophistomis latifasciata Melzer, 1926: 9.
Strangalia flavocincta; Linsley & Chemsak, 1971: 24.

Remarks: *Strangalia flavocincta* is known from Bolivia (Cochabamba, Santa Cruz) and Brazil (Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina) (Monné, 2022c; Tavakilian & Chevillotte, 2022).

Material examined (only new record listed): BRAZIL, *Rio Grande do Sul (new state record):* Barra do Ribeiro, CMPC Celulose Riograndense – Horto Florestal Barba Negra, *Eucalyptus saligna* stand planted May/2013, 30°22'56.46"S, 51°08'06.15"W, ethanol-baited FIT, 1 ♂, 19.XI.2018, A.A. Melo leg. ((MEFEIS).

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