

RESEARCHA RTICLE

Effect of dietary Aqua fix on qualitative protein profile in skeletal muscles of *Labeorohita*

Ankamma, N.¹ and VivekaVardhani, V.²

^{1,2}Department of Zoology and Aquaculture AcharyaNagarjuna University, Nagarjunanagar-522 510 (A.P.) India

Email: :vadlamudi_vv@yahoo.co.in

ABSTRACT

The effect of Aqua Fix (immunomodulator) on the qualitative protein index of skeletal muscles of *Labeorohita* was investigated. Test fish (twenty five fish of 6 months old, 12-15 g.wt; 25 fish of 9 months old, 47-50 g.wt.) were fed with Aqua Fix @ 50 mg / 100 g of feed for 4 days (group A, B). Another 2 groups (a, b) (25 nos. in each group) of fish of same age and weight were fed with normal diet and kept as control for comparison. Significant alterations were found in the molecular weight of skeletal muscle protein in the test and control fish through SDS-PAGE analysis on day 1, 4, 7, 15 and 30 of experiment. Qualitative analysis depicted varied number of low and high molecular weight protein bands in the skeletal muscles of *L. rohita* of experimental and controls.

Key words: Dietary Aqua Fix, Skeletal muscles, Protein, L. rohita

INTRODUCTION

Aquaculture is one of the fastest food producing sector and infectious diseases are causing heavy loss to the fish culturists. *Aeromonashydrophila*, an opportunistic pathogen is wide spread globally and infects both freshwater and warm water fish. This bacterium causes haemorrhagic septicemia, ulcers, abscesses, exopthalmia and abdominal distension (Austin and Austin, 1987; Chowdhury, 1998; Rajeswariet al., 2005). Recent knowledge on immunonutrition studies reveal that the growth, disease resistance, and non-specific and specific immunity of the fish may be raised by the use of some nutrients (Priyaet al., 2004; Kumar et al.,

HowtoSiteThisArticle:

Ankamma, N and VivekaVardhani, V(2017).Effect of dietary Aqua fix on qualitative protein profile in skeletal muscles of *Labeorohita*.Biolife. 5(2), pp 279-283.

DOI: 10.5281/zenodo.7364624 Received: 8 April 2017; Accepted; 27 May 2017; Available online : 5 June 2017

2005). Use of antibiotics and chemotherapeutics to control fish diseases has the risk of bioaccumulation and development of resistant pathogens. Under such

conditions, feeding cultivable fish through dietary immunostimulantscannot be overlooked.

Various herbal extracts such as Aloe vera (Kim et al., 1999), Ocimum sanctum (Logambalet al., 2000), Zingiberofficinale(Dugenciet 2003). al., Achyranthesaspera (Vasudeva and Chakrabarti, 2005a, b) and Solanumtrilobatum(Divyagnaneswariet al., 2007) have been reported to enhance immunity in fish Cynodondactylon (L.) and Coriandrumsativumextract mixed diet enhanced disease resistance and production of specific antibodies in Catlacatla (Xavier Innocent, et al., 2011; Kaleswaranet al., 2012) against aeromoniasis. Administration of glucan and Aeglemarmelos enhanced survival and immunity in Cyprinuscarpio challenged with A. hydrophila (Selvarajet al., 2005; Pratheepaet al., 2010 and Jasmin Gold, V and VivekaVardhani, V, 2016). Administration of microbial levan showed enhanced haemtological and non-specific immunological changes in Labeorohita (Gupta et al., 2008). The structure and molecular weight of muscle protein in fish be altered due to the stress caused by various physiologic factors, environment, seasons of the year, starvation, breeding season and migration (Gomez et al., 2000; Ladratet al., 2000; Delbare-Ladratet al., 2006). Sultana et al., (2016) assessed the effect of different feed on the quantitative and qualitative changes in the protein content of C. mrigala, C. catla and L. rohita. Keeping in view the importance of the dietary immunostimulants and economic importance of the Indian major carps, the present study is aimed to assess the efficacy of Aqua Fix (as an immunostimulant) on the qualitative changes in the content of skeletal muscle protein of *L. rohita*.

MATERIALS AND METHODS

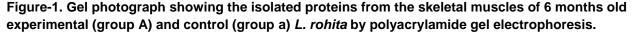
Six months and nine months old experimental fish (12-15 g.wt. and 47-50 g.wt.) were collected from Singh ponds, Kuchipudi village, Guntur District, Andhra Pradesh, India and allowed to acclimatize to laboratory conditions for one week. Four groups (A, B, a, b) of fish were maintained; two experimental (group A, B) and two control groups (group a, b) of 25 fish in each group. Experimental group of fish (A, 6 months old; B, 9 months old) were fed with Agua Fix @ 50 mg/100 g of feed for 4 days and control groups (a, 6 months old; b, 9 months old) with normal diet. Five fish from groups A and B were necropsied on day 1, 4, 7, 15 and 30 after treating with immunostimulant diet. Control fish (from groups a and b) were also necropsied on the same designated days. Pieces of skeletal muscle tissue were removed from both the experimental and control groups of fish and qualitative analysis of proteins was performed by SDS-PAGE using the Discontinuous Buffer system of Laemmli (1970).

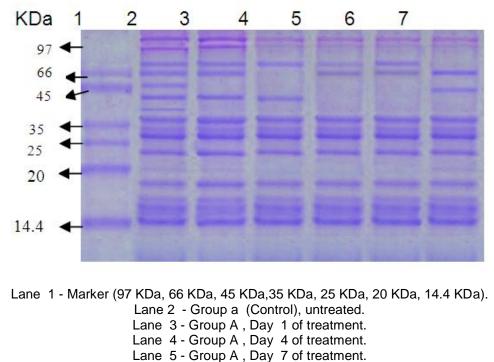
RESULTSAND DISCUSSION

L. rohita of 6 months old fish fed with diet supplemented with Aqua Fix (group A) and controls (group a) showed a number of protein bands ranging from 97 to 14.4 KDa and a protein band ~ 45 KDa disappeared on day 1 of experiment (Fig-1, Table-1).

On day 4 and 7, protein band ~ 66 KDa disappeared; protein band ~ 45 KDa (found on day 1 and 4) and ~ 40 KDa (found on day 4) were absent on day 7. No marked changes were found in the isolated proteins on day 15 except the absence of a protein band ~ 40 KDa and presence of a protein band ~ 66 KDa. On day 30 of treatment the isolated proteins have a molecular weight ranging between 35 to 14.4 KDa; a protein band ~ 82 KDa disappeared and other protein band ~ 45 KDa appeared.

Control (group b) and experimental (group B, fed with Aqua Fix for 4 days @ 50 mg/100 g. of feed) 9 months old L. rohita showed a series of several protein bands ranging ~ 97 to 14.4 KDa (Fig-2, Table-2). In the muscle samples of experimental fish (group B), 10 protein bands were recorded on day 1 of experiment. The isolated protein bands have molecular weight as ~ 97, 90, 66, 41.3 35, 25, 20, 18, 16, 14.4 KDa. Compared with controls, 2 protein bands ~ 64.4 KDa and 41 KDa disappeared and protein bands ~ 35 to 14.4 KDa were present both in groups B and b. On day 4, experimental fish showed an additional protein band ~ 40 KDa and disappearance of ~ 64.4 KDa and 41 KDa protein bands; no marked change was found in protein bands of 35 to 14.4 KDa. Two protein bands 66 and 45 KDa appeared distinctly as in controls on day 7 of treatment in fish of group B. The additional protein band appeared on day 4 (~ 40 KDa mol. wt.) retained, a protein band ~ 19.1 KDa disappeared and protein bands between ~ 35





Lane 6 - Group A , Day 15 of treatment. Lane 7 - Group A , Day 30 of treatment.

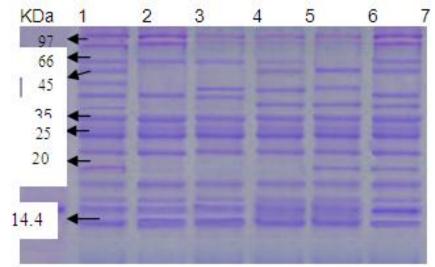
S.No.	Marker Mol.wt. KDa	Control group a	Group A					
			Day 1	Day 4	Day 7	Day 15	Day 30	
1	97.0	180.181	180.181	176.329	180.181	176.329	180.181	
2	66.0	115.209	143.525	149.697	143.525	143.525	142.525	
3	45.0	85.941	89.257	72.420	87.028	90.399	67.889	
4	35.0	67.889	59.621	35.497	68.683	68.683	45.512	
5	25.0	51.162	36.592	28.714	29.192	30.495	28.184	
6	20.0	35.899	28.407	25.941	25.941	28.870	25.723	
7	14.4	30.680	25.941	23.337	24.334	25.831	24.463	
8		29.030	23.337	21.533	22.184	23.513	21.989	
9		25.831	21.417	20.107	19.889	22.159	20.649	
10		24.420	19.916	19.648	19.653	20.090	20.073	
11		21.574	19.672	19.427	19.484	19.657	19.616	
12		20.515	19.484	19.345	19.349	19.512	19.464	
13		19.930	19.290			19.343	19.266	
14		19.667						
15		19.505						
16		19.340						

Table-1: Characterization of isolated protein on the basis of their molecular weight in skeletal muscles of experimental (group A) and control (group a) 6 months old *L. rohita.*

to 14.4 KDa remained stable on day 7 of treatment. Fish of group B showed distinct protein bands on day 15; a protein band ~ 66 KDa disappeared and that of ~ 19.1 KDa reappeared and no marked alteration was found in protein bands ~ 35 KDa to 14.4. No marked changes were found in the arrangement of protein bands on day 30 of treatment in group B in comparison with controls (group b) except the disappearance of a protein band \sim 41.4 KDa.

In the present investigation, the impact of Aqua Fix on the qualitative profile of muscle protein in 6 and 9 months old *L. rohita* is confirmed by the presence of

Figure-2.Gel photograph showing the isolated proteins from the skeletal muscles of 9 months old experimental (group B) and control (group b) *L. rohita* by polyacrylamide gel electorphoresis..



Lane 1 - Marker (97 KDa, 66 KDa, 45 KDa, 35 KDa, 25 KDa, 20 KDa, 14.4 KDa). Lane 2 - Group a (Control), untreated. Lane 3 - Group A , Day 1 of treatment. Lane 4 - Group A , Day 4 of treatment. Lane 5 - Group A , Day 7 of treatment. Lane 6 - Group A , Day 15 of treatment. Lane 7 - Group A , Day 30 of treatment.

S.No.	Marker Mol.wt. KDa	Control group b	Group B					
			Day 1	Day 4	Day 7	Day 15	Day 30	
1	97.0	147.715	176.897	156.283	165.433	160.783	126.28	
2	66.0	106.609	129,781	52,950	119.623	102.963	95.322	
3	45.0	53.999	67.115	33.271	66.128	49.387	54.35	
4	35.0	41.375	28.556	27.762	49.694	34.627	47.05	
5	25.0	33.143	24.600	24.501	33.400	27.118	30.082	
6	20.0	28.111	23.477	23.277	26.072	24.469	26.54	
7	14.4	26.327	22.630	22.893	24.501	23.519	24.284	
8		24.438	22.204	22.240	23.477	22.583	23.22	
9		23.258	21.697	21.692	22.572	22.042	22.57	
10		22.550	21.618	21.609	22.240	21.783	22.00	
11		22.162	21.587	21.598	21.701	21.703	21.77	
12		21.839	21.548	21.548	21.606	21.621	21.65	
13		21.694			21.580	21.598	21.61	
14		21.622			21.575	21.577	21.59	
15		21.585					21.57	
16		21.569						
17								

Table-2: Characterization of isolated protein on the basis of their molecular weight in skeletal muscles of experimental (group B) and control (group b) 9 months old *L. rohita.*

varied number of isolated proteins of different molecular weights. These results are in agreement with the findings of Li et al., (2000) and Islam (2006) who reported the effect of dietary protein on growth of Ictaluruspunctatus and expression of muscle protein in pikeperches, Stizostedionlucioperca and S. volgensa. Singh et al., (2005) and Tarakalakshmi Y and VivekaVardhani V (2015)also determined the importance of protein for optimum growth in L. rohita. In the present study, high molecular weight (97 KDa) protein bands were isolated and characterized by SDS-PAGE in 6 and 9 month old muscle proteins. Mitsuhashiet al., (2002) and Fock and Hinssen (2002) also characterized several proteins as an integral parts of skeletal and cardiac muscles in fish. The present findings are further supported by the findings of Okagakiet al., (2005) and Montowska and Pospiech (2007) who found protein bands ranging between 16 and 26 KDa in skeletal muscles of species of carp. In 6 and 9 month old L. rohita fed with a diet supplemented with immunostimulant, variable number of protein bands were observed in skeletal muscles on day 1, 4, 7, 15 and 30 of treatment. Two bands were shared between 6 months (group A) and 9 months (group B) old L. rohita on day 1 (24.4, 21.5; 24.6, 21.5 KDa), 4 (23.3, 21.5; 23.2, 21.6 KDa), 7 (24.3 22.1; 24.5 22.5 KDa) 15 (23.5, 22.1; 23.5, 22.5 KDa) and 30 (24.4, 21.9; 24.2 21.7 KDa). These findings are in agreement with that of Mathew and Prakash (2006) who reported 23 and 22 KDa proteins from Sardinellalongiceps. The present study on immunomodulatory effect of Aqua Fix on the concentrations of skeletal muscle proteins of L. rohita are supported by the findings of Salim (2006) and Sultana et al., (2016) and Madhuri, D and VivekaVardhani, V (2015).who reported increased muscle protein content with higher dietary protein in carps.

Acknowledgement:

One of the authors (Ankamma, N.) is thankful to UGC for providing financial support in the form of FDP.

Conflict of Interests

Authors declare that there is no conflict of interests regarding the publication of this paper.

References

- [1]. **Austin, B. and Austin, D.A. (1987**). Bacterial Fish Pathogens: Disease in Farmed and Wild Fish. Ellis Horwood Ltd., West Sussex, England, pp.13-350.
- [2]. Chowdhury, M.B.R. (1998). Involvement of *Aeromonads*and *Pseudomonas* in disease of farmed fish in Bangladesh. Fish Pathol. 33: 247-254.
- [3]. Delbare-Ladrat, C., Cheret, R., Taylor, R. and Verrez-Bagins, V. (2006). Trends in postmortem aging in fish: understanding of proteolysis and disorganisation of the myofibrillar structure. Critical Rev. in Food. Sci. Nutr. 46: 409-421.
- [4]. Divyagnaneswari, M., Christybapita, D. and Michael, R.D. (2007). Enhancement of non-specific immunity and disease resistance in Oreochromismossambicusby Solonumtrilobatumleaf fractions. Fish & Shellfish Immunol. 23: 249-259.

- [5]. Dugenci, S.K., Arda, N. and Candan, A. (2003). Some medicinal plants as immunostimulant for fish. J. Ethnopharmacol. 88: 99-106.
- [6]. Fock, U. and Hinssen, H. (2002). Nebulin is a thin filament protein of the cardiac muscle of the agnathans. J. Muscle Research and Cell Motility. 23(3): 205-213.
- [7]. Gomez, G., M.C., Montero, P., Hurtado, O. and Borderias, A. (2000). Biological characteristics affect the quality of farmed Atlantic salmon and smoked muscle. J. Food Sci. 65(1): 53-60.
- [8]. Gupta, S.K., Pal, A.K., Sahu, N.P., Dalvi, R., Kumar, V. and Mukherjee, S.C. (2008). Microbial levan in the diet of *Labeorohita*Hamilton juveniles: effect on nonspecific immunity and histopathological changes after challenge with *Aeromonashydrophila*. J. Fish Dis. 31: 649-657.
- [9]. Islam, A. (2006). Muscle protein expression of pikeperches (*Stizostedionlucioperca*and *S. volgensa*). Integ. Zool., 2: 96-103.
- [10]. Jasmin Gold, V and Viveka Vardhani, V (2016). Protein level in the heart of mice during immunostimulation and hepatitis infection. Biolife, 4(2), pp 271-274. doi:10.17812/blj.2016.4210
- [11]. Kaleswaran, B., Ilavenil, S. and Ravikumar, S. (2012). Changes in biochemical, histological and specific immune parameters in *Catlacatla*(Ham.) by *Cynodondactylon* (L.). J. King Saud Univ. Sci. 24: 139-152.
- [12]. Kim, K.H., Hwang, Y.J. and Bai, S.C. (1999). Resistance to *Vibrio alginolyticus*in juvenile rockfish (*Sebastesschlegeli*) fed diets containing different doses of aloe. Aquaculture 180, 13-21.
- [13]. Kumar, S., Sahu, N.P., Pal, A.K., Choudhury, D., Yengkokpam, S. and Mukherjee, S.C. (2005). Effect of dietary carbohydrate on haematology, respiratory burst activity and histological changes in *L. rohita*juveniles. Fish & Shellfish Immunol. 19: 331-344.
- [14]. Ladrat, C., Chaplet, M., Verrez-Bagnis, V. Noel. and J. Fleurence, J. (2000). Neutral calciumactivated proteases from European sea bass (*Dicentrachuslabrax* L.) muscle: Polymorphism and Biochem. Stud. Comp. Biochem. Physiol. 125B: 83-95.
- [15]. Laemmli, U.K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature, 227: 680-685.
- [16]. Li, M.H., Bosworth, B.G. and Robinsson, E.H. (2000). Effect of dietary protein concentration on growth and processing yield of channel catfish (*Ictaluruspunctatus*). J. World Aquaculture. Soc. 31: 592-598.
- [17]. Logambal, S.M., Venkatalakshmi, S. and Michael, R.D. (2000).Immuno-stimulatory effect of leaf extract of Ocimum sanctum Linn. inOreochromismosambicus(Peters). Hydrobiologia, 430: 113-120.
- [18]. Madhuri, D and Viveka Vardhani, V (2015). GST level in the abdominal muscles of mice treated with Immunex DS and Gene Vac B vaccine. Biolife. 3(1) 26-30.

- [19]. Mathew, S. and Prakash, V. (2006). Effect of calcium salts on the properties of proteins from oil sardine (*Sardinellalongiceps*) during frozen storage. J. Food Sci., 71: 178-183.
- [20]. Mitsuhashi, T., Kasai, M. and Hatae, K. (2002). Detection of giant myofibrillar proteins connection and nebulin in fish meat by electrophoresis in 3-5 gradient sodium dodecyl sulfate polyacrylamide slab gels. J. Agric. Food Chem., 50(26): 7499-7503.
- [21]. Sateesh Pujari, & Estari Mamidala. (2015). Antidiabetic activity of Physagulin-F isolated from Physalis angulata fruits. The American Journal of Science and Medical Research, 1(2), 53–60. https://doi.org/10.5281/zenodo.7352308
- [22]. Montowska, M. and Pospiech, E. (2007). Species identification of meat by electrophoretic methods. Acta Sci. Pol. Technol. Aliment., 6(1): 5-16.
- [23]. Okagaki, T., Takami, M., Hosokawa, K., Yano, M., Fujime, S.H. and Ooi, A. (2005). Biochemical properties of ordinary and dark muscle myosine from carp skeletal muscle. J. Biochem., 138: 255-262.
- [24]. Pratheepa, V., Ramesh, S. and Sukumaran, N. (2010).Immunomodulatory effect of Aeglemarmelos leaf extract on freshwater fish Cyprinuscarpio infected by bacterial pathogen Aeromonashydrophila. Pharmaceutical Biol. 48(11): 1224-1239.
- [25]. Priya, K., Mukherjee, S.C., Pal, A.K. and Sahu, N. (2004). Effects of dietary lipids on histological changes in hepatic tissues of *Catlacatla* fingerlings. Indian J. Vet. Pathol. 28: 121-124.
- [26]. Rajeswari, S., Shome, B.R., Mazumder, Y., Das, A., Kumar, A., Rahman, H. and Bujarbaruah, K.M. (2005). Abdominal dropsy disease in major craps of Meghalaya: isolation and characterisation of *Aeromonashydrophila*. Curr. Sci. 88: 1897-1900.
- [27]. Salim, M. (2006). Role of fish as food to human nutrition. International Conference on "Solving Problems of Freshwater Fish Farming in Pakistan", p.20, November 27-28, 2006, UVASS, Lahore, Pakistan.
- [28]. Selvaraj, V., Sampath, K. and Sekar, V. (2005). Administration of yeast glucan enhances survival and some non-specific and specific immune parameters in carp (*Cyprinuscarpio*) infected with *Aeromonashydrophila*. Fish & Shellfish Immnol. 19: 293-306.
- [29]. Singh, P.K., Gaur, S.R., Barik, P., Shukla, S.S. and Singh, S. (2005). Effect of protein levels on growth and digestibility in the Indian major carp, *Labeorohita*(Hamilton) using slaughter house waste as the protein source, Int. J. Agri. Biol., 7: 939-941.
- [30]. Sultana, S., Zahra, A., Sultana, T., Al-Ghanim, K.A. and Mahboob S. (2016). Effect of different artificial feeds formulated from local ingredients on the meat quality of Indian major carps. The J. Animal and Plant Sci. 26(4): 1140-1145.
- [31]. Tarakalakshmi Y and Viveka Vardhani V (2015). SDS PAGE Protein profile in small intestine of mice during ancylostomiasis. Biolife, 3(2), pp 524-527.doi:10.17812/blj2015.32.25
- [32]. Vasudeva, R.Y. and Chakrabarti, R. (2005a).Stimulation of immunity in Indian major carp

*Catlacatla*with herbal feed ingredients.Fish & Shellfish Immunol. 18: 327-334.

- [33]. Vasudeva, R.Y and Chakrabarti, R. (2005b). Dietary incorporation of *Achyranthesaspera*seed influences the immunity of common carp *Cyprinuscarpio*. Indian J. Anim. Sci. 75: 1097-1102.
- [34]. Xavier Innocent, B., Syed Ali Fathima, M. and Siva Rajani, S. (2011). Immune response of *Catlacatla*fed with an oral immunostimulant *Plumbagorosea*and post-challenged with *Aeromonashydrophila*. Inter. J. Applied Biol. and Pharmaceutical Technol. 2(4): 447-454.