

<https://doi.org/10.25221/fee.367.4>

<http://urn:lsid:zoobank.org:pub:4180A9D7-27F0-4A25-B6D2-D85705EA743E>

**MORPHOLOGICAL REDESCRIPTION AND DNA BARCODING OF  
*KALUGINIA LEBETIFORMIS* MAKARCHENKO, 1987 (DIPTERA:  
CHIRONOMIDAE, DIAMESINAE) FROM SOUTH KOREA**

**E. A. Makarchenko<sup>1\*</sup>, A. A. Semenchenko<sup>2)</sup>, H. Kang<sup>3)</sup>, Y. J. Bae<sup>3)</sup>**

1) Federal Scientific Center of the East Asia Terrestrial Biodiversity, Far East Branch of the Russian Academy of Sciences, Vladivostok 690022, Russia. \*Corresponding author, E-mail: [makarchenko@biosoil.ru](mailto:makarchenko@biosoil.ru)

2) Far Eastern Federal University, Vladivostok 690950, Russia.

3) Laboratory of Biodiversity and Ecology, College of Life Sciences Korea University, Life Science (WEST) Bldg 145 Anam-ro, Seongbuk-gu, Seoul 136-713, Korea.

**Summary.** Illustrated redescription of adult male as well as the results of DNA barcoding of *Kaluginia lebetiformis* Makarchenko in comparison with known species of the tribe Boreoheptagyini from South Korea are provided. The species-specificity of *K. lebetiformis* COI sequences is analyzed and the sequences are presented as diagnostic characters – molecular markers. Intergenous K2P distance between three genera and species of the tribe Boreoheptagyini – *K. lebetiformis*, *Boreoheptagyia* sp. and *Shilovia rara* Makarchenko ranged from 0.105–0.143 (mean 0.124) and these values are sufficient to maintain the genus level. DNA barcodes of *K. lebetiformis* was uploaded to GenBank.

**Key words:** Diptera, Chironomidae, Diamesinae, *Kaluginia*, taxonomy, DNA barcoding, South Korea.

**Е. А. Мака́рченко, А. А. Семенченко, Х. Кан, Ю. Дж. Бэ. Морфологическое переписание и ДНК баркодинг *Kaluginia lebetiformis* Makarchenko, 1987 (Diptera: Chironomidae, Diamesinae) из Южной Кореи // Дальневосточный энтомолог. 2018. N 367. С. 26-32.**

**Резюме.** По материалу из Южной Кореи приведены иллюстрированное переписание имаго самца и результаты анализа фрагмента гена цитохромоксидазы субъединицы I (658 пн.) мтДНК хирономиды *Kaluginia lebetiformis* Makarchenko из подсемейства Diamesinae. Полученные последовательности для этого вида были депонированы в геномном банке. Генетические дистанции (K2P) между родами трёх представителей трибы Boreoheptagyini – *K. lebetiformis*, *Boreoheptagyia* sp. и *Shilovia rara* Makarchenko находятся в пределах 0,105–0,143 (в среднем 0,124), что достаточно для поддержания родового статуса *Kaluginia* Makarchenko.

**INTRODUCTION**

The genus *Kaluginia* Makarchenko, 1987 was established with the description of *K. lebetiformis* Makarchenko, 1987 from the south part of Sakhalin Island (Makarchenko, 1987).

Later one male of this species was found in Khasansk District of Primorye Territory (Makarchenko *et al.*, 2017). Males of this monotypic genus have antenna with 7 flagellomeres and reduced plume, pedicel with setae; eye bare, sometimes slightly pubescence, not extended dorsomedially; anteprenotal lobes narrowly joined medially, lateral anteprenotals occupy basal 2/3 of lobe, median anteprenotals absent; acrostichals present, beginning near anteprenotum and reach middle mesonotum, dorsocentrals erect in 1–2 rows, supraalars present; costa not extending,  $R_{2+3}$  reduced and visible only in basal part, FCu distal to MCu,  $R_{4+5}$  with setae, squama fringed; legs speckled; anal point like small rounded protuberance, sternapodeme high, almost trapezoidal, gonocoxite with basal lobe and inferior volsella, gonostylus broad and scoop-shaped, with 5–9 megasetae along inner margin.

Some years ago additional imaginal material was collected by Dr. Y.J. Bae and PhD student H. Kang in South Korea. It allows us to make a more detailed description of the male imago, as well as to conduct a DNA analysis of this species. Below we redescribe adult male of *K. lebetiformis* from the South Korea as well as give the results of DNA barcoding in comparison with the closely related members of tribe Boreoheptagiini, namely *Boreoheptagyia* Brundin and *Shilovia* Makarchenko. The DNA barcode corresponding to the 650-bp fragment of the mitochondrial gene cytochrome c oxidase I (COI) has been identified as the core of a global bio-identification system at the species level (Hebert *et al.*, 2003) and has proved to be useful in non-biting midges (Montagna *et al.*, 2016; Ekrem *et al.*, 2010; Carew *et al.*, 2005). K2P genetic distances been used to establish specific independence of the redescribed species.

## MATERIAL AND METHODS

The material was preserved in 96% ethanol for DNA-analysis and in 70% ethanol for further study of morphology, and slide-mounted following the methods by Makarchenko (1985). The terminology follows Sæther (1980). All material is deposited in the Laboratory of Freshwater Hydrobiology of the Federal Scientific Center of the East Asia Terrestrial Biodiversity, Far East Branch of the Russian Academy of Sciences, Vladivostok.

Genomic DNA was extracted with using the Invitrogen PureLink Genomic DNA Mini Kit in compliance with the manufacturer's protocols. Approximately 650 base pairs of the COI were amplified from diluted genomic DNA by polymerase chain reaction (PCR) in a total volume of 10  $\mu$ l with 5  $\mu$ l of Go Taq Green Master Mix (Promega Corp, Madison, WI, USA), 0.5  $\mu$ l of each primer (100 ng/ $\mu$ l), 3  $\mu$ l of nuclease-free water and 1  $\mu$ l of total DNA. The PCR thermal regime consisted of one cycle of 1 min at 94 °C; five cycles of 1 min at 94 °C, 1.5 min at 45 °C and 1.5 min at 72 °C; 35 cycles of 1 min at 94 °C, 1.5 min at 50 °C and 1 min at 72 °C; and a final cycle of 5 min at 72 °C. The primers COIF-ALT (5'-ACAAA TCAYAARGAYATYGG-3') and COIR-ALT (5'-TTCAGGRTGNCCRAARAA CA-3'), which were obtained from Mikkelsen *et al.* (2006), were used. All PCR products were verified using electrophoresis on a 1,5% TBE agarose by visualizing on GelDoc XR+ imaging systems (BioRad), only positive PCR products were purified for cycle sequencing using Exonuclease I (ExoI) and Thermosensitive Alkaline Phosphatase (FastAP) by ThermoFisher Scientific. The PCR products were bidirectionally sequenced using the BigDye Terminator v3.1 cycle kit and run on an ABI 3130xl DNA analyzer (Applied Biosystems). Sequences were aligned and manually edited in MEGA 7 (Kumar *et al.*, 2016). Based on the Kimura-2-Parameter (K2P) model were calculated interspecific and intragenus genetic distances using MEGA 7. Two COI sequences of *K. lebetiformis* have been deposited in GenBank (MH547037–MH547038).

## MORPHOLOGICAL REDESCRIPTION

### *Kaluginia lebetiformis* Makarchenko, 1987

Figs 1–10

*Kaluginia lebetiformis* Makarchenko, 1987: 786, 2006: 266; Oliver, 1989: 134; Ashe & Connor, 2009: 291.

**MATERIAL EXAMINED.** **South Korea:** 2♂, Gyeonggi-do, Gapyeong-gun, Gapyeong-cheon, Bukhan River, Han River basin, 15. IV 2016 (Light trap), 37°58' N, 36.4° 127°26' 35.5" E, leg. Y. Bae; 1♂, the same data except 22.V 2014 (Light trap), leg. H. Kang. **Russia:** South Sakhalin, 1♂ (holotype), Belaya River, Sokol Village, Dolinsk District, 29.VI 1985, leg. S. Bestalannaya, E. Makarchenko.

**REDESCRIPTION.** *Male imago* (n=3). Total length 2.3–2.6 mm. Total length/wing length 1.03–1.10. Total coloration brown to dark-brown; antepronotum light yellow; methonotum yellowish, with brown stripes; legs spotted: basal 2/3 of femur yellowish, distal 1/3 brown; tibia in basal and apical parts brown and yellowish in middle part; basal 2/3 of  $ta_1$  yellowish and distal 1/3 brown;  $ta_2$ – $ta_5$  brown (Fig. 1).

*Head.* Temporal setae (from one side) including 3–4 frontals, 7 orbitals and 7–10 verticals. Clypeus with 17–26 setae. Palpomere length ( $\mu\text{m}$ ): 28–48, 60–64, 104–120, 132–140, 212–220. Head width/palpal length 0.92–1.0. Antenna with 7 flagellomeres and reduced plume of setae 48–68  $\mu\text{m}$  long (Fig. 2). Flagellomeres length ( $\mu\text{m}$ ): 52–56, 24, 28, 24–28, 28–32, 28–32, 68–84; terminal flagellomere with 2 subapical setae 60–76  $\mu\text{m}$  long; scape with 3 setae 28–36  $\mu\text{m}$  long. AR 0.34–0.46.

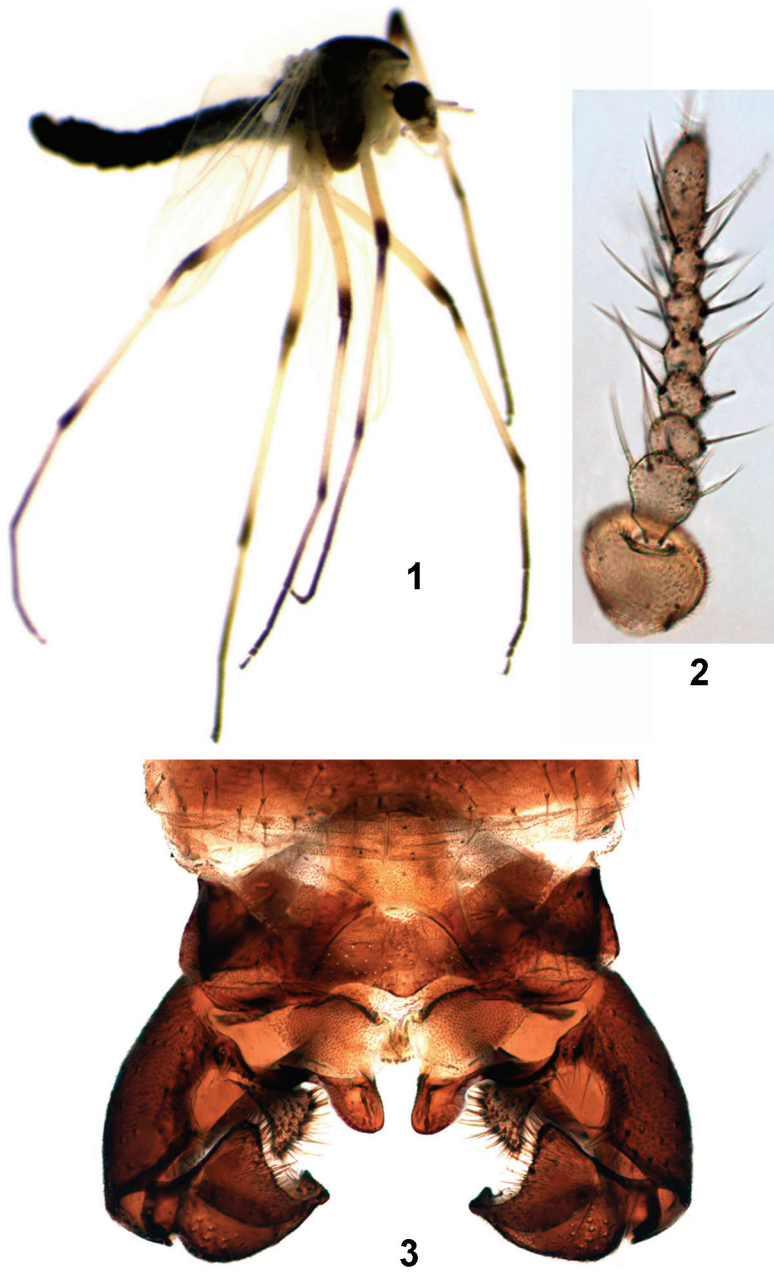
*Thorax.* Antepronotum with 18–20 lateral setae. Acrostichals 25 (48–56  $\mu\text{m}$ ), dorsocentrals 21–27 (72–100  $\mu\text{m}$ ), prealars 16, supraalars 1, scutellars *ac* 50.

*Wing.* Length 2.08–2.52 mm, width 0.68–0.76 mm. Anal lobe well developed; squama with 19–31 setae (64–80  $\mu\text{m}$ ) in 1–2 rows. R and  $R_1$  with 55–58 setae (32–36  $\mu\text{m}$ ),  $R_{4+5}$  with 24–29 setae (28–40  $\mu\text{m}$ ) in subapical part. Wing of one male with 3 short setae on  $M_{3+4}$ .

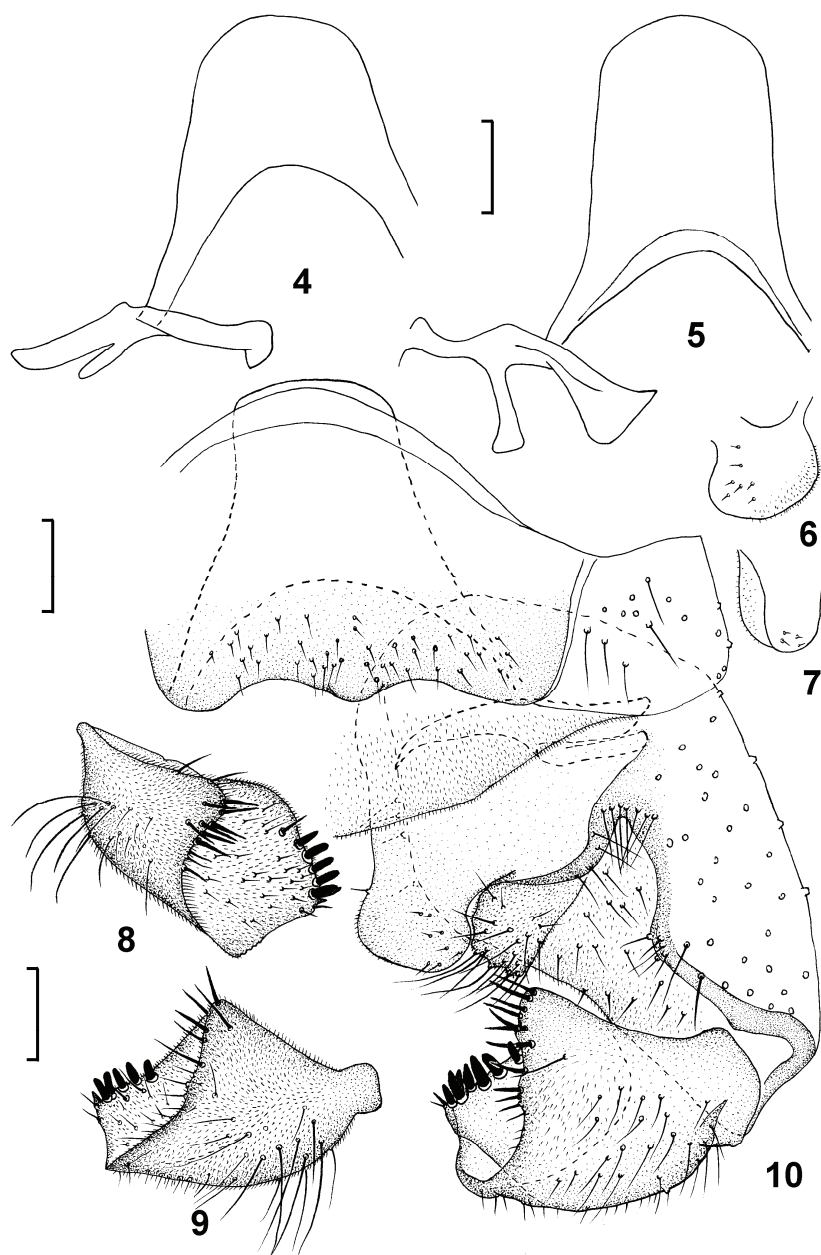
*Legs.*  $BR_1$  2.0;  $BR_2$  1.8;  $BR_3$  1.9. Spur of front tibia 44  $\mu\text{m}$  long. Spurs of middle tibia 28–52 and 44–53  $\mu\text{m}$  long. Spurs of hind tibia 68–72  $\mu\text{m}$  and 44–52  $\mu\text{m}$  long. Hind tibial comb with 12–14 spine-like setae. Middle legs with 4 pseudospurs 36–40  $\mu\text{m}$  long on  $ta_1$ ; hind legs with 7 pseudospurs on  $ta_1$ . Claw with 4 denticles apically. Length ( $\mu\text{m}$ ) and proportions of legs segments are as follow:

| P              | fe    | ti    | $ta_1$ | $ta_2$ | $ta_3$ | $ta_4$ | $ta_5$ | LR    | BV    | SV    |
|----------------|-------|-------|--------|--------|--------|--------|--------|-------|-------|-------|
| P <sub>1</sub> | 918–  | 1132– | 722–   | 394–   | 197–   | 66–    | 98–    | 0.64– | 3.67– | 2.76– |
|                | 1115  | 1328  | 886    | 459    | 213    | 74     | 115    | 0.67  | 3.87  | 2.84  |
| P <sub>2</sub> | 984–  | 1082– | 574–   | 312–   | 189–   | 115–   | 115    | 0.53– | 3.61– | 3.31– |
|                | 1197  | 1246  | 738    | 394    | 213    | 74     |        | 0.59  | 4.0   | 3.60  |
| P <sub>3</sub> | 1000– | 1230– | 705–   | 394–   | 213–   | 66–    | 98–    | 0.57– | 3.69– | 3.0–  |
|                | 1214  | 1443  | 886    | 508    | 246    | 74     | 131    | 0.61  | 3.81  | 3.16  |

*Hypopygium* (Figs 3–10). Tergite IX with “anal point” which like small rounded protuberance, and with 29–42 setae, 12–20  $\mu\text{m}$  long; laterosternite IX with 15–20 setae (from one side), 44–48  $\mu\text{m}$  long. Transverse sternapodeme high, almost trapezoidal, 128–132  $\mu\text{m}$  long of specimens from South Korea, 92  $\mu\text{m}$  long of specimen from South Sakhalin, and 148–152  $\mu\text{m}$  wide in basal part, 88–100  $\mu\text{m}$  wide in subapical part, with rounded apex (Figs 4–5). Gonocoxite 308–310  $\mu\text{m}$  long. Basal lobe of gonocoxite is various shapes and depends on its position; in the inner half with microtrichia, along the outer margin with short setae (Figs 3,



Figs 1–3. Adult male of *Kaluginia lebetiformis* Makarchenko from South Korea. 1 – total view of male, from one side; 2 – antenna; 3 – total view of hypopygium, from above.



Figs 4–10. Hypopygium of *Kaluginia lebetiformis* Makarchenko from South Sakhalin (4) and South Korea (5–10). 4–5 – transverse sternapodeme and aedeagal lobe; 6–7 – basal lobe of gonocoxite; 8–9 – gonostylus; 10 – total view of hypopygium, from above. Scale bars = 50  $\mu\text{m}$ .

6–7, 10); inferior volsella as in Figs 3, 10, covered by setae 40–60 µm long. Gonostylus 156 µm long, scoop-shaped; inner lobe along the margin with 5–9 megasetae 12–16 µm long and one subterminal tooth; outer lobe widely triangular, inner margin of which with strong setae 20–24 µm long, outer half with thinner and longer setae 56–72 µm long (Figs 3, 8–10). HR 1.97.

REMARKS. Additional material from South Korea allowed a more detailed study of the internal structure of the male hypopygium, namely transverse sternapodeme, which in complex with aedeagal lobe, basal lobe of gonocoxite and inferior volsella is typical for Boreoheptagyini. Males from both populations are close related by all features but specimens from South Korea have more a long transverse sternapodeme (128–132 µm) than specimen from South Sakhalin (92 µm).

DISTRIBUTION. East Palaearctic continental-insularis species known from Sakhalin Island and Primorye Territory in Russia and from South Korea.

### RESULTS OF DNA BARCODING

The final alignment of the COI gene yielded 658 bp for 2 samples of *K. lebetiformis* that were 2 haplotypes. The nucleotide composition of the studied sequences of *K. lebetiformis* COI gene fragments deviated from an equilibrium one, comprising 26.4 % of A, 40.0 % of T, 17.0 % of C, and 16.5 % of G. Total intraspecific sequence divergence was 0.0061, which is based on four nucleotide substitutions. All the substitutions were synonymous and observed only in the third codon positions. Three substitutions were transitions and one transversion. Average intergenus K2P distance between *K. lebetiformis* and other genera of the tribe Boreoheptagyini (Diamesinae) showed the following results: *Boreoheptagyia* sp. (KY640386) – 0.143, *Shilovia rara* Makarchenko (KY640384–KY640385) – 0.124. High differences between the groups can argue genus independence of the *Kaluginia* (Ekrem *et al.*, 2007).

### ACKNOWLEDGMENTS

This work partly was supported by Russian Science Foundation (project No. 14-50-00034).

### REFERENCES

- Ashe, P. & O'Connor, J. P. 2009. *A World Catalogue of Chironomidae (Diptera). Part 1. Buchonommyiinae, Chilenomeiinae, Podonominae, Aphroteniinae, Tanypodinae, Usambaromyiinae, Diamesinae, Prodiamesinae and Telmatogetoninae*. Irish Biogeographical Society & National Museum of Ireland, Dublin. 445 pp.
- Carew, M.E., Pettigrove, V. & Hoffmann, A.A. 2005. The utility of DNA markers in classical taxonomy: using cytochrome oxidase I markers to differentiate Australian *Cladopelma* (Diptera: Chironomidae) midges. *Annals of the Entomological Society of America*, 98: 587–594. DOI: [https://doi.org/10.1603/0013-8746\(2005\)098\[0587:TUODMI\]2.0.CO;2](https://doi.org/10.1603/0013-8746(2005)098[0587:TUODMI]2.0.CO;2)
- Ekrem, T., Stur, E. & Hebert, P.D.N. 2010. Females do count: Documenting Chironomidae (Diptera) species diversity using DNA barcoding. *Organisms Diversity & Evolution*, 10: 397–408. DOI: <https://doi.org/10.1007/s13127-010-0034-y>
- Ekrem, T., Willassen, E. & Stur, E. 2007. A comprehensive DNA sequence library is essential for identification with DNA barcodes. *Molecular Phylogenetics and Evolution*. 43: 530–542. DOI: <http://dx.doi.org/10.1016/j.ympev.2006.11.021>

- Kumar, S., Stecher, G. & Tamura, K. 2016. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33(7): 1870–1874. DOI: <https://doi.org/10.1093/molbev/msw054>
- Makarchenko, E.A. 1985. *Chironomids of the Soviet Far East. Subfamilies Podonominae, Diamesinae and Prodiamesinae (Diptera, Chironomidae)*. DVNC AN SSSR Press, Vladivostok. 208 pp. [In Russian]
- Makarchenko, E.A. 1987. *Kaluginia lebetiformis* gen. et sp. n. (Diptera, Chironomidae, Diamesinae) from southern Sakhalin. *Zoological Journal*, 66: 784–786. [in Russian]
- Makarchenko, E.A. 2006. 3. Subfamily Diamesinae. Pp. 253–276, 468–480, 607–621. In: Lelei, A.S. (Ed.). *Key-book of insects of the Russian Far East. Vol. 6. Diptera and Siphonaptera. Part 4*. Dalnauka, Vladivostok. [In Russian]
- Makarchenko, E.A. & Makarchenko, M.A. 2017. Fauna and distribution of the Podonominae, Diamesinae, Prodiamesinae and Orthoclaadiinae (Diptera, Chironomidae) of the Russian Far East and bordering territory. *Vladimir Ya. Levanidov's Biennial Memorial Meetings. Vol. 7*. FSCEATB FEB RAS, Vladivostok. 127–142.
- Montagna, M., Mereghetti, V., Lencioni, V. & Rossaro, B. 2016. Integrated Taxonomy and DNA Barcoding of Alpine Midges (Diptera: Chironomidae). *PLoS ONE*, 11(3): e0149673. DOI: <https://doi.org/10.1371/journal.pone.0149673>
- Oliver, D.R. 1989. 7. The adult males of Diamesinae (Diptera, Chironomidae) of the Holarctic region. - Keys and diagnoses. *Entomologica Scandinavica*, Suppl. 34: 129–154.
- Sæther, O.A. 1980. Glossary of chironomid morphology terminology (Diptera, Chironomidae). *Entomologica Scandinavica*, Suppl. 14: 1–51.

---

© **Far Eastern entomologist (Far East. entomol.)** Journal published since October 1994.

Editor-in-Chief: S.Yu. Storozhenko

Editorial Board: A.S. Lelej, S.A. Belokobylskij, M.G. Ponomarenko, E.A. Beljaev, V.A. Mutin, E.A. Makarchenko, A.V. Gorochoy, T.M. Tiunova, M.Yu. Proshchalykin, S.A. Shabalin

Address: Federal Scientific Center of the East Asia Terrestrial Biodiversity (former Institute of Biology and Soil Science), Far East Branch of the Russian Academy of Sciences, 690022, Vladivostok-22, Russia.

E-mail: [storozhenko@biosoil.ru](mailto:storozhenko@biosoil.ru)

web-site: <http://www.biosoil.ru/fee>