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RESEARCH ARTICLE

QUANTITATIVE ESTIMATION OF SOME ANTIOXIDANTS AND ANTI NUTRITIONAL CONTENT OF *GLINUSOPPOSITIFOLIUS*

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Glinusoppositifolius, Totalphenol, Totalsaponin, Steriodalsaponin, Cardiacglycosides

Abstract

Glinusoppositifolius(L.)Aug.DC (Molluginaceae), known as gimashak and it contains linear toovate,opposite leaves and greenish flowers. (1).This study was performed to measure theantioxidants and anti nutritional contents of the aqueous paste of *Glinusoppositifolius*(ediblepart). Some of these phytochemicals are heat labile, and reduced after cooking .So all parameterswerealsomeasuredafterheatreatment.Resultshowedadifferencebetweentherawsampleandcookedsampleofthese allparameters.Studyfoundthatafterboilingtherewas17.74%,8.36%,8.8%,0.44%,60%,11.11%,52.17%,1.90%18.33%,4.34%and5.71%reductionoftot alphenol,flavonoid,DPPH(IC50),FRAPassay,totalalkaloid,oxalate,phytat e,tannin,totalsaponin,steriodalsaponinandcardiac glycosidesrespectively.

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Introduction:-

Gimashakisconsumed asleafyvegetablesby localpeopleofalltropicalcountry (2).Thisbitterleafyvegetableisknownas‘GimaShak’inWestBengal, Assam&Bangladesh(3). *Glinusoppositifolius*areused for treatingjoint pain, inflammation, diarrhea ,Intestinalparasites,feverboils andskindisorders (4).Leaves of the *Glinusoppositifolius*(Linn)contain spergulagenic, spergulagenin A and a tri hydroxy ketone (5).A bioactive pecticpolysaccharideisolatedfrom*G. oppositifolius*is foundtopossessimmunomodulationproperty (6).*Glinusoppositifolius*issishowntoexhibitantioxidant(7),hepatoprotective(8)antidiabetic(9) andantihyperlipidemic(10)activity.

TaxonomicalClassification:(11)

1. Kingdom–Plantae
2. Division–Magnoliophyta
3. Class–Magnoliopsid
4. Sub-class–Caryophyllideae
5. Order–Caryophyllales
6. Family–Molluginaceae
7. ScientificName–*Glinusoppositifolius*(L.)Aug.DC.
8. Genus–Glinus
9. Species–oppositifolius
10. Synonyms–MollugoSpergulaL.

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It was identified and classified by Botanical Survey Of India, Kolkata, West Bengal
Identification number: CNH/Tech.II/2019/38.

Aims & Objectives:-

Glinus oppositifolius is a widely grown plant and available at a very low price. So poor people can consume this leafy vegetable in their diet. Micronutrient deficiencies are a major health problem in our country among low income groups due to lack of food availability and poor purchasing capacity. Previous research found that *Glinus oppositifolius* contains a good amount of vitamins and minerals (12). Antioxidants, vitamins and minerals prevent potential damage caused by reactive oxygen species to the cellular tissues and modulate immune function in our body. Whereas antinutrients are responsible for deleterious effects related to the absorption of micronutrients and macronutrients. So the objective of this study was to quantify of antioxidants and antinutritional factors of *Glinus oppositifolius* in the form normally consumed by humans in the cooked form..

Materials and methods:-

- Collection of Sample: *Glinus oppositifolius* was collected from Sovabazar market, Kolkata, West Bengal, India.
- Preparation of Sample:

Preparation of raw sample: Fresh *gimashak* was collected from Sovabazar market, cleaned and washed. Then the usual standard procedure was followed for the estimation of antioxidants and anti-nutritional factors.

Preparation of cooked sample: Fresh harvested *gimashak* (*Glinus oppositifolius*) was collected, cleaned and washed. Estimated amount of *gimashak* was weighed then boiled with measured amount of fresh drinking water for 30 minutes. Extraction procedure was done according to the guideline of the methods. For estimation of all phytochemicals and anti-nutritional factors, both raw and cooked sample was used.

With this raw and cooked sample all parameters were observed by the following methods with few modifications:

1. Estimation of Total phenol content (Barman K, 2004 method). (13)
2. Estimation of Flavonoid content (Chang et al, 2002 method). (14)
3. Estimation of DPPH Antioxidant assay (Mensor et al, 2001 method). (15)
4. Estimation of FRAP Assay (Benzie et al, 1996 method) (16)
5. Estimation of Total alkaloid (Harbone JB, 1973 method) (17)
6. Estimation of oxalate (Onyema et al, 2016) (18)
7. Estimation of phytate (Ifemeje et al, 2014 method) (19)
8. Estimation of tannin (Saxena et al, 2013 method) (20)
9. Estimation of total saponin (Pasaribu et al, 2014 method) (21)
10. Estimation of steroidalsaponin (Singh et al, 2015 method) (22)
11. Estimation of cardiac glycosides (Onyema et al, 2016) (18)

Results:-

Table 4.1:- Quantitative Estimation Result of Antioxidant and Antinutritional Parameters.

Sln o	Parameters	Results	Percentage of Reduction After Cooking (%)	
		Raw Sample (value ± SE)	Cooked Sample (value ± SE)	
1.	Total phenol content	102 ± 0.4 mg GAE/100 gm	83.9 ± 0.88 mg GAE/100 gm	17.74
2.	Flavonoid	98 ± 1.1 mg Quercetin equivalent/100 gm	89.8 ± 0.83 mg Quercetin equivalent/100 gm	8.36
3.	DPPH Antioxidant (IC ₅₀)	10.2 ± 0.26 µg/ml	9.03 ± 0.03 µg/ml	8.8
4.	FRAP Assay	2.26 ± 0.1 µM	2.25 ± 0.04 µM	0.44
5.	Total alkaloid	2 ± 0.03 gm/100 gm	0.8 ± 0.01 gm/100 gm	60
6.	Oxalate	0.8 ± 0.03 gm/100 gm	0.72 ± 0.01 gm/100 gm	11.11
7.	Phytate	23 ± 0.53 mg/100 gm	11 ± 0.27 mg/100 gm	52.17

8.	Tannin	42±0.58mg/100gm	41.2±0.63/100gm	1.90
9.	Totalsaponin	1.2±0.31gm/100gm	0.98±0.02/100gm	18.33
10.	Steroidal saponin	0.23±0.003gm/100gm	0.22±0.01/100gm	4.34
11.	Cardiac glycosides	1.4±0.04gm/100gm	1.32±0.28gm/100gm	5.71

GAE:Gallicacidequivalent

StatisticalAnalysis:

Correlation coefficient was calculated as per Pearson's Coefficient.

Table 4. 2:- Correlation Coefficient of Antioxidant Activities of Total Phenolic Content(TPC), Flavonoid,DPPH radicals scavenging activity and FRAP Assay(cooked sample)

Correlation coefficient	TPC (Total Phenol Content)	Flavonoid	DPPH radicals scavenging activity	FRAP Assay
TPC		-0.70(very weak)	-0.87(very weak)	0.56(moderate)
Flavonoid			0.27(weak)	-0.98(very weak)
DPPH radicals scavenging activity				-0.09(very weak)

Table 4.3:- Correlation Coefficient of Antioxidant Parameters and Anti nutritional Parameters of Cooked Sample.

Correlation coefficient	Total Phenol	Flavonoid	DPPH	FRAP Assay
Total alkaloid	-0.94(very weak)	0.43(moderate)	0.98(very strong)	-0.26(very weak)
Oxalate	0.86(very strong)	0.96(very strong)	-0.51(very weak)	0.9(very strong)
Phytate	-0.87(very weak)	0.96(very strong)	0.53(moderate)	-0.88(very weak)
Tannin	0.69(strong)	-0.99(very weak)	-0.25(very weak)	0.98(very strong)
Totalsaponin	0.09(very weak)	-0.76(very weak)	0.4(moderate)	0.87(very strong)
Steroidal saponin	0.89(very strong)	-0.95(very weak)	-0.56(very weak)	0.87(very strong)
Cardiac Glycosides	-0.33(very weak)	0.9(very strong)	0.15(very weak)	-0.96(very weak)

Standard range: 0.00-0.19=very weak, 0.20-0.39=weak, 0.40-0.59=moderate, 0.60-0.79=strong, 0.80-1=very strong.

Discussion:-

Ali et al. claimed that natural antioxidants mainly present in the form of phenolic compounds such as flavonoids and phenolic acids from the plants.(23) *Glinus oppositifolius* contains a better amount of antioxidants than other Indian vegetables. It contains 102 mg GAE /100gm total phenol and 98mg quercetin/100gm whereas amaranth contains 24.76mg GAE/gm and 8.13 mg CAE/gm respectively. (24) The antioxidant activity was also determined as radical scavenging activity as DPPH assay and ability to reduce Fe^{3+} - Fe^{2+} (FRAP). Total phenol content of cooked sample and FRAP showed a good correlation. It was found that after boiling there was 17.74%, 8.36%, 8.8%, 0.44% reduction of total phenol, flavonoid, DPPH (IC50), FRAP. Heat may disrupt the hydroxyl group structure of total phenol which is mainly responsible for phenolic antioxidant properties. (25) Increased surface area of tissues in contact with cooking water as well as high temperature was likely to have caused disruption of cell walls and breakdown of phenolic compounds.(26) Flavonoids are commonly present in edible fruits and vegetables. It is a heat labile compound, so the heat exposure during cooking can influence their content in vegetables. Thermal treatment can affect both the extractability and bio accessibility of phytochemicals because of the destruction of the cell wall in plant material. (27) Study found the highest correlation between (0.98) tannin and FRAP

assay and total alkaloid and DPPH (0.98) assay. Oxalate showed very strong correlation with TPC, Flavonoid and FRAP assay. A very strong relationship was found between tannin and FRAP assay, total saponin and FRAP assay. Steroidal saponin revealed a very strong relationship with total phenol and FRAP assay whereas cardiac glycosides showed a very strong relationship with flavonoid. The correlation discrepancies found in literature are explained on the basis of differences in the interpretation of the results by individual methods. Antioxidant activity of a substance can vary from method to method depending on factors such as oxidation state, antioxidant solubility, and medium of pH..(28) Study showed, 60%, 11.11%, 52.17%, 1.90% 18.33%, 4.34% and 5.71% reduction of total alkaloid, oxalate, phytate, tannin, total saponin, steroidalsaponin and cardiac glycosides respectively after cooking. Total alkaloid and phytate content showed a significant reduction whereas oxalate and total saponin showed moderate reduction. Tannin, steroidalsaponin, cardiac glycosides showed a minimal reduction. From this experiment it may be concluded that this sample contains heat sensitive alkaloid and phytate. From this study it may be concluded that consumption of cooked *Glinus oppositifolius* is good for us because of these high antioxidant content. Though heat treatment reduces the antioxidant content but simultaneously heat can also reduce the antinutritional factors. Reductions of these antinutritional factors are necessary as these factors can cause deleterious effects on health.

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Conflictsofinterest:

There are no conflicts of interest.

References:-

1. N. Hoque, M.Zafar Imam, S. Akter, Md. E.H. Mazumder, S.M. Raquibul Hasan, J. Ahmed and Md.S. Rana. Antioxidant and Antihyperglycaemic Activity Of Methanolic Extracts of *Glinus oppositifolius* leaves. Journal of Applied Pharmaceutical Science. 2011;01(07):50-53.
2. C.Panda,U.Mishra,S.Mahapatra and G.Panigrahi. Free Radical Scavenging Activity and Phenolic Content Estimation of *Glinus oppositifolius* & *Sesbania grandiflora*. International Journal Of Pharmacy. 2013;4:722-727
3. S.K.Sahu,D.Das,N.K.Tripathy .S.C.Dinda and H.K.Sundeep .Evaluation of Hypoglycaemic Activity of *Mollugopentaphylla* and *Glinus oppositifolius* .Rasayan.J.Chem. 2012;5(1):57-62
4. D.Diallo,B.Hveem MA.Mahmoud,G.Berge,BS.Paulsen and A.Maiga. An Ethnobotanical Survey of Herbal Drugs of Gourma District ,Mali. Pharmaceutical Biology. 1999;37(1):80-91.
5. A.Ghani,2nd Edn, Medicinal Plants Of Bangladesh with chemical constituents and uses .The Asiatic Society of Bangladesh,2003.
6. K.T.Inngjerdingen,S.C.Debes,M.Inngjerdingen,S.Hokpusta,S.E.Hardling and B.Rolsted . Bioactive Pectic Polysaccharide From *Glinus oppositifolius*(L) Aug .DC., a Malian Medicinal Plant Isolation and Partial Characterization. J.Ethnopharmacol.2005;101:204-214
7. K.Asokkumar,M.Umamaheswari,A.T.Sivashanmugam,V.Subhadra Devi,N.Subhashini and T.K.Ravi. Free Radicals scavenging *Glinus oppositifolius*(carpetweed) Using Different In Vitro Systems. Pharmaceutical Biology,2009;47(6):47 4-482.
8. M.Mazumder,U.K.Halder,P.K.Manikandan L,Senthilkumar GP and Kandar.C.G.Hepatoprotective Activity Of Methanol Extract of *Glinus oppositifolius* & *Trianthemadecandra* Against Paracetamol Induced Liver Damage. Advances In Traditional Medicine.2007;7(1):74-78.
9. S.Pattanayak and K.Padmalatha. Anti diabetic Activity Of Parts of *Glinus oppositifolius*L Against Glucose overloaded and Streptozotocin Induced Diabetes In Albinorats. International Journal Of Natural Products and Marine Biology.2015;1(1):29-34.
10. GM.Behera, S.Kumar, B.N.Malay Baidya and G.Panigrahi. Anti Hyperlipidaemic and Antioxidant Activity of *Glinus oppositifolius*(L.) Aug ,DC. Pharmacology online.2010;20:915-936.
11. Cronquist A,2nd Edn, The Evaluation and Classification Of Flowering Plants. Boston,1988.
12. S.P.Choudhury and K.L.Khaled. Quantitative Estimation of Some Essential Vitamins and Minerals of *Glinus oppositifolius*. International Journal of Research and Analytical Reviews.2021;8(2):458-461.
13. K.Barman. Biodegradation Of Tanniniferous Feeds And Their Influence On Nutrient Utilization and Productivity Of The Dairy Animals. Ph.D thesis, Submitted to NDRI,Karnal.

14. C.Chang ,M.Yang,H.WenandJJ.Chern.EstimatioinOfTotalFlavonoidContentByTwoComplementaryColorimetricMethods. FoodDrugAnalysis.2002;10:178-182.
15. MensorLL,M.FS,LeitaoGG,ReisAS,SantosTC,CoubeCSand LeitaoSG.,ScreeningOfBrazilianPlantExtractsForAnti-oxidantActivityByTheUseOfDPPHFreeRadicalMethod.PhytotherResearch.2001;15:127-130.
16. Benzei,F.F&Strain,J.J,Methods Of Biochemical Analysis, A book by(NIN,Hyderabad.1999),pp-31-34.
17. HarborneJB.PhytochemicalMethods: A Guide to Modern Techniques of Plant Analysis, Edn3.(Chapman And Hall London,1998)p. 302.
18. C.T.Oneyema,C.E.Ofor,V.C.Okudo and Ogubuaga.Phytochemical and Antimicrobial Analysis of Banana Pseudo Stem(*Musa acuminate*). British Journal Of Pharmaceutical Research.2016;10(1):1-9.
19. J.C.Igemeje,C.Egbuna,J.O.Eziokwudiso and F.C.Ezebuo.Determination of The Antinutrient Composition of *Ocium gratissimum*,*Corchorus olitorius*,*Murraykoenigii* and *Cucurbita maxima*. International Journal of Innovation and Scientific Research2014;3(2):127-133.
20. V.Saxena, G. Mishra, A. Saxena, Kamlesh KR. Vishwakarma..A Comparative Study On Quantitative Estimation Of Tannins In *Terminalia chebula*, *Terminalia bellerica*, *Terminalia arjuna*, *Saraca indica* using spectrophotometer. Asian Journal Of Pharmaceutical and Clinical Research.2013;6(3):148-149.
21. T.Pasaribu,D.A.Astuti,E.Wina,Sumiati and A.Setyono .Saponin Content of *Sapindusrarak* Pericarp Affected by Particle Size and Type of Solvent, Its Biological Activity on Eimeria tenella oocytes. International Journal of Poultry Science.2014;13(6):347-352.
22. R.Singh and V.d.Mendulkar: *Abutilon indicum*(Linn) sweet leaves, A Natural Source of Saponin: A Spectrophotometric Assay. International Journal of PharmTech Research.2015,8(4):725-729.
23. G.Ali and G.Neda.Flavanoids and Phenolic Acids : Role and Biochemical Activity In Plants and Human. Journal of Medicinal Plants Research.2011;5(31):6697-6703.
24. J.XK.P.Sanchez,C.R.Moreno,M.A.G.Favela, JM.Carrillo, EOC.Rodriguez, AV.Ortiz and R.G Darado. Increasing The Antioxidant Activity, Total Phenolic Acid , Flavonoids Contents By Optimizing The Germination Conditions of Amaranth Seeds. Plant Foods for Human Nutrition .2014;69(3):196-202.
25. Lo Scalzo R,Genna A, Branca F, Chedin M and Chassaigne H. Anthocyanin Composition of Cauliflower (*Brassica oleracea L.var.botrytis*) and Cabbage (*Brassica oleracea L.var.capitata*) and Its Stability In Relation To Thermal Treatments. Food Chem.2008;107:136-144.
26. M.Palermo,N.Pellegrini and V.Fogliano.The Effect Of Cooking On The Phytochemicals Content of Vegetables. J Sci Food Agriculture.2014;94:1057-1070.
27. M.Hilemori,Koh E and AE.Mitchell. Influence of Cooking on Anthocyanins In Black Rice (*Oryza sativa L.japonica* var. SBR). J Agric Food Chem.2009;57:1908-1914.
28. HA .Moharram and MM Yousef : Methods of Determining the Antioxidant Activity- A Review . Alexandria Journal of Food Science and Technology2014.11(1):31-42.