

ISSN NO: 2231-6876



INDO AMERICAN JOURNAL OF PHARMACEUTICAL RESEARCH

ZIZIPHUS JUJUBA FRUIT WITH PROMISING ANTICANCER ACTIVITIES

Dr. Dhanapal Venkatachalam

Sree Sastha Pharmacy College, Chennai-600123.

ARTICLE INFO

Article history

Received 10/08/2022 Available online 03/09/02022

Keywords

Anticancer Activity, Bioactive Compounds, Chemoresistance, Ziziphus Jujube Mill.

ABSTRACT

Ziziphus jujube Mill. (Z. jujuba) is a traditional herb with a long history of use for nutrition and the treatment of a broad spectrum of diseases. It grows mostly in South and East Asia particularly in India, as well as in Australia and Europe. Mounting evidence shows the health benefits of Z. jujuba, including anticancer, anti-inflammation, anti-obesity, antioxidant, and hepato-protective properties, which are due to its bioactive compounds. Chemotherapy, such as with cis-diamminedichloroplatinium (CDDP, cisplatin) and its derivatives, is widely used in cancer treatment. It is an effective treatment for human cancers, including ovarian cancer; however, drug resistance is a major obstacle to successful treatment. A better understanding of the mechanisms and strategies for overcoming chemo resistance can greatly improve therapeutic outcomes for patients. In this review article, the bioactive compounds present in Z. Jujube are explained. The high prevalence of many different cancers worldwide has recently attracted the attention of many researchers. This is why our research group focused on studying the anticancer activity of Z. Jujube as well as its impact on chemo resistance both in vivo and in vitro. We hope that these studies can lead to a promising future for cancer patients.

Corresponding author

Dr Dhanapal Venkatachalam

Principal
Sree Sastha College of Pharmacy
Chennai –Bangalore High Way
Chembarambakkam
Chennai – 600123.
vddpaul@gmail.com
+ 919443952113

Please cite this article in press as **Dr Dhanapal Venkatachalam** et al. Ziziphus Jujuba Fruit with Promising Anticancer Activities. Indo American Journal of Pharmaceutical Research. 2022:12(08).

INTRODUCTIONThe practice of application of medicinal plants for relief from many illnesses dates back to ancient times. ¹ *Ziziphus jujuba* Mill., (*Z. jujuba*) or the jujube, a herbal plant used in traditional medicine, belongs to the Rhamnaceae family and is one of the most important *Ziziphus* species. ² In terms of geographical distribution, *Z. jujuba* is widely located in the tropical and subtropical regions of Asia and America as well as in the Mediterranean regions. ³ The mature fruit of *Z. jujuba* are red to purplish black in color, resembling small dates, and in China are known as the date or red date. The dried fruit of *Z. jujuba* is known in Persian cuisine as "annab." Jujube trees grow in the arid and semiarid zones of Iran, especially in Birjand, South Khorasan province ⁴ Because of the plant's extensive utilization in food and pharmaceutical industries, production has increased sharply over the past decade. China has been the site of 90% of *Z. jujuba* production in the world, as it is indigenous to China with a history of over 4,000 years ⁵⁻⁶



Fig 1: Fruits of Zizyphus jujube.

Different parts of *Z. jujuba* can be used for the curing of many kinds of illness including diabetes, diarrhoea, skin infections, liver complaints, urinary disorders, obesity, fever, pharyngitis, bronchitis, anaemia, insomnia, cancer, and also for blood purification and tonification of the gastrointestinal tract⁷⁻⁹ Cancer continues to be and is increasingly a serious health problem, and one of the leading causes of death in the world. Aging and the growth of the world population, changes in lifestyle, and the adoption of cancer-causing behaviours are some of the reasons for this prevalence. According to cancer statistics in 2013, stomach and liver cancers are the most common in Asia and both of them are associated with high mortality rates, while bladder cancer is the most common in the USA. Colorectal and breast cancers have high incidence rates in all countries. Pancreatic cancer is also listed among the top five cancers in all countries except for China and Brazil. Some fruits and vegetables have protective effects on organs that are damaged by anticancer drugs. For example, whole fruit extract of pomegranate can reduce the myocardial toxicity induced by doxorubicin, which is an anthracycline antibiotic with broad antitumor spectrum and has been used against a wide variety of hematopoietic malignancies and solid tumors. Also, Hassanpour *et al.* showed that the juice of *Lagenariasiceraria* (Molina) Standley (bottle gourd) had the same cardioprotective effect, which was less than that of the whole fruit extract of pomegranate

The same researchers attributed the cardioprotective effect of pomegranate and bottle gourd to the preservation of endogenous antioxidants and reduction of lipid peroxidation. The high antioxidant contents of these fruits can neutralize the free radicals generated by doxorubicin, causing an unusual and often irreversible cardiomyopathy.

NUTRIENTS

Several studies in recent years have shown the important roles of bioactive compounds and natural substances derived from plants in the discovery of new drugs, particularly anticancer drugs. ¹⁵ Dried pulp of *Z. jujuba* is a source of essential unsaturated fatty acids. The main fatty acids in the jujube are oleic, linoleic (omega-6), palmitic, and palmitoleic acids. ¹⁶ Jujube fruits contain various types of amino acids and proteins; in the maturity stage when the Jujube fruits are harvested; the protein and free asparagines content are influenced potently. Combined free amino acids show a 3.3-fold increase from S1 to S5 stages of ripeness and a decrease from S6 to S8 (S1-S8 are the eight stages of maturation of jujube extracts). Additionally, free asparagines decrease during the last three stages of maturity, while their peak level is at S5¹⁷ Dietary fiber and fructose contents of the jujube fruit play a role in the regulation of blood sugar levels by slowing digestion ¹⁸ The major sugars found in the jujube fruit are glucose, fructose, sucrose, rhamnose, and sorbitol. The jujube fruit is also abundant in vitamin C, which is one of the watersoluble antioxidants ¹⁹ The postharvest sorting process is important for increasing the economic benefits and dietary values of the jujube fruit, especially vitamin C content protection during storage and marketing ²⁰

Moreover, the jujube is enriched, though to a lesser extent, with other vitamins including thiamin, riboflavin, niacin, vitamin B_6 , and vitamin A. Jujube fruit is also considered a good source of minerals such as magnesium, phosphorus, potassium, sodium, and zinc

BIOACTIVE COMPOUNDS

Various studies have shown that the jujube fruit contains many bioactive compounds, including triterpenic acids, flavonoids, cerebrosides, phenolic acids, α -tocopherol, β -carotene, and polysaccharides. Each constituent of the jujube presents some health benefits, thus making it a healthy food choice²¹ Jujube fruit has more total phenolic compounds compared to other common fruits that exhibit antioxidant activities, such as cherry, apple, persimmon, or red grape. ²² Flavonoids, phenolic acids, tannins, stilbenes, and lignans are derivatives of phenolic compounds²³

ANTICANCER ACTIVITY OF Z. JUJUBA

Cancer is considered to be one of the most common diseases, causing death worldwide, and is a serious public health problem. According to the World Health Organization, 10 million new cases of cancer are diagnosed by physicians each year and based on statistical trends, this figure is estimated to double by 2020. ²⁴ Although chemotherapy has many side effects, it remains a standard method for the treatment of cancer patients. Due to their projected low costs, fewer side effects, and low toxicity compared to the standard treatment, the development of new agents such as medicinal herbs with anticancer effects can herald a promising future in cancer treatment²⁵⁻²⁶ Triterpenic acids are in the form of free acids or glycones such as saponins, which have multiple biological effects including anti-inflammatory, ²⁷ antimicrobial 28 hepatoprotective ²⁹ and antioxidant³⁰ effects. In recent years, the anticarcinogenic and antitumor activities of triterpenic acids have made them attractive in the fields of scientific research and healthcare products³¹]Guo et al. in 2009 identified 10 triterpenic acids, that is, ceanothic, alphitolic, zizyberanal, zizyberanalic, epiceanothic, ceanothenic, betulinic (BA), oleanolic (OA), ursonic, and zizyberenalic acids, and two triterpenes, that is, ziziberanalic acid and ursolic acid (UA), in the dried jujube fruit³² Among all the compounds found in the dried jujube fruit, a few have cytotoxic effects: BA, OA, and UA. Their chemical structures have been shown in Figure- 2³³⁻³⁴ Defects in apoptosis or programmed cell death play a role in various physiological and pathological processes, which are characteristically linked to the multistep process of tumor genesis. Thus, one of the most important mechanisms of the anticancer properties of bioactive compounds is the modulation of the apoptotic process³⁵

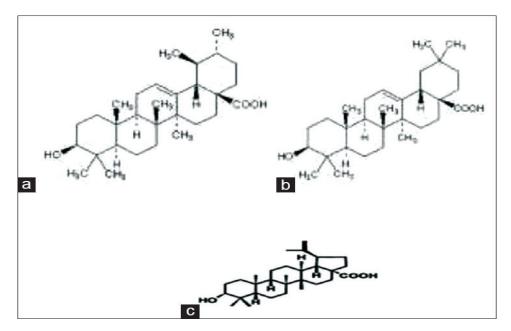


Fig 2: The chemical structure of Ursolic acid (UA) (a) Oleanolic acid (OA), (b) Betulinic acid (BA), (c) Bioactive compounds present in *Z. jujuba* with cytotoxic effects.

URSOLIC ACID (UA)

In a study carried out by Kim in 2000, it has been shown that UA exerts proapoptotic activity and decreases cell viability in a concentration- and time-dependent fashion on HepG2 human hepatoblastoma cells. UA at the dose of 30 µm induced DNA fragmentation and the activation of caspase-3³⁶ In another study in 2010, UA effects were investigated on mammary tumor cells both *in vivo* and *in vitro*. Female ovariectomized C57BL/6 mice as a model of the postmenopausal state received for 8 weeks either control diet [following the American Institute of Nutrition (AIN) formula, AIN-93G] or diet supplemented with UA at three different concentrations: 0.05%, 0.10%, and 0.25%. After 3 weeks on the diet, the mice were injected with syngeneic MMTV-Wnt-1 mammary tumor cells.

In addition, the growth-inhibitory effects of UA on WA4 mammary tumor cells in vitro were studied. UA was effective in inhibiting WA4 cell proliferation at 25 µm and 50 µm, which supported the antitumor activity in the mouse model of postmenopausal breast cancer. All doses of UA in Ki-67 immuno-histo-chemical staining showed inhibition of proliferation in vivo, while at the concentration of 0.10%, it had the greatest effect in decreasing the final tumor size. It seems that the modulation of phosphatidylinositol 3 kinase (PI3K/AKt) and mitogen-activated protein kinase (MAPK) cellular signaling pathways involved in UA affects cell survival and apoptosis³⁷ UA and OA are the two triterpene acids that have similar chemical structures but different positions on a methyl group in the E loop³⁸ UA and OA were studied by Li *et al.* in 2002 and they demonstrated marked antitumor effects on the human colon carcinoma cell line HC-T15³⁹ Colorectal cancer, with its high incidence in Australia, New Zealand, and East Asia, is recognized to be the fourth-most common cancer in the world.⁴⁰ The study revealed that the concentration of 60 µmol/L of UA or OA as treatment for the duration of 24-72 h could increase the number of dead cells and cell fragmentations more than the 30 µmol/L concentration. Cell cycle analysis by flow cytometry showed that UA 30 µmol/L and OA 60 µmol/L for the duration of 36 h and 72 h, respectively, caused the accumulation of HCT-15 cells in the G0/G1 phases and concomitant cell decrease in the S phase. Additionally, Shyu et al. in 2010 demonstrated the inhibitory effects of OA and UA on human hepatocellular carcinoma (HCC) HuH-7 cell population growth with (IC₅₀) 100 μm and 75 μm, respectively. HCC is a form of cancer that involves the predominant cell type of the liver, hepatocytes; it is the fifth-most common cancer and the third-most frequent cancer causing death worldwide. 41 OA and UA increased the permeability of the transition pore in the mitochondria that in turn cause the rapid release of caspase activators such as cytochrome c into the cytoplasm and the subsequent activation of caspase-9 and caspase-3, followed by the cleavage of poly (ADP-ribose) polymerase (PARP) and the induction of apoptosis in HuH-7 cells. Several other studies also showed that UA and OA induce apoptosis in HL-60 leukemia cells, B16F10 melanoma cells, MCF-7 breast cancer cells, and DU-145 prostate cancer cells. 42-45

BETULINIC ACID (BA)

BA, a pentacyclictriterpene and lipophilic compound, has selectively demonstrated cytotoxic effects in both in vitro and xenograft mouse models of human melanoma and neuroectodermaltumors (neuroblastoma, glioblastoma, medulloblastoma), both of which arise from the neural crest. BA acts through caspase activation in neuroectodermal cells. Melanoma, a malignant neoplasm of melanocytes, according to surveillance in epidemiology, is the sixth-most common fatal malignancy in the USA. 46 In the study carried out by Selzer et al., BA induced apoptosis in human melanoma cells, and the effects of BA in combination with irradiation were clearly additive, thus possibly differing in their mode of action. The growth-inhibitory effect of BA has been reported to be more pronounced on human melanoma cell lines than on normal human melanocytes. Interestingly, in spite of the induction of apoptosis, BA induced the expression of MC1-1 (a member of BCL-2 protein family showing antiapoptotic activity). Additionally, in another study, BA showed synergistic cytotoxic effects on melanoma cells in combination with vincristine⁴⁷ Furthermore, the antitumor activity of BA has been reported in the broad spectrum of cancer cell lines, including leukemia, colon, lung, prostate, head and neck, and cervical 48-50 In the study carried out by Damle et al. in 2013, BA showed cytotoxic activity on MCF-7 breast adenocarcinoma xenografts in athymic nude mice. Also, BA in MCF-7 cells in vitro after injections in the concentrations of 50 mg/kg and 100 mg/kg body weight in mice decreased tumor size by 52% and 77%, respectively. This antitumor activity possibly resulted from the decreased angiogenesis, proliferation, and invasion in the BA-treated animals⁵¹ Breast cancer is a major public health problem and is considered to be the most common cancer in women worldwide, accounting for 23% of all cancers in women.⁵² Cytotoxic effects of the water extract of the jujube in vitro were shown to occur on the Jurkatleukemia cell line. Furthermore, HeLa and HEP-2 cells from the epithelioid cervix and human larynx carcinomas were affected by the dialyzed extract of jujube. However, it seems that the lack of DNA fragmentation by the induction of apoptosis in these two latter cell lines indicate that other mechanisms may contribute to cell death.

In an *in vivo* model of metastatic melanoma, the concomitant use of BA and vincristine augmented suppression of the experimental lung metastasis of melanoma cells in mice, compared to the animals treated with vincristine alone. Of course, there were no systemic toxicities or weight loss in BA-treated mice even at a high systemic dose. One of the most striking features of BA and its derivatives is their different effects on cancer cells and healthy cells; they are less toxic to cells of healthy tissues. Mechanisms of BA action include the induction of apoptosis via mitochondrial pathway and the loss of mitochondrial membrane potential without any effect on the caspase inhibitor. There are two major signaling pathways for apoptotic cell death: The extrinsic or receptor pathway and the intrinsic or mitochondrial pathway⁵³ A schematic presentation of the major signaling pathways for apoptotic cell death and bioactive compounds present in the jujube, inducing apoptosis through several mechanisms for the anticancer activities, is presented in Figure 3⁵⁴ In another mechanism, BA can trigger the production of reactive oxygen species (ROS), which in turn can activate nuclear factor kappa-B (NF-κB), an inflammatory signaling pathway in a variety of tumor cell lines, and the inhibition of BA-induced NF-κB activation can attenuate BA-induced apoptosis⁵⁵

Fig 3: Schematic presentation of major signaling pathways for apoptotic cell deathand bioactive compounds available in Z. jujuba, which induce apoptosis through several mechanisms for anticancer activities. UA = Ursolic acid, OA = Oleanolic acid, BA = Betulinic acid, DPP = deproteinized polysaccharide.

With regard to yet another mechanism, the Bcl-2 family of proteins include both antiapoptotic members, such as Bcl-2, Bcl-X₁, and Mcl-1, and proapoptotic ones, such as Bax, Bak, Bad, and BH3 domain⁵⁶ BA has been reported in neuroblastoma, glioblastoma, and melanoma cells, upregulating the proapoptotic Bcl-2 family protein Bax. The expression levels of antiapoptotic Bcl-2 remained unchanged in neuroblastoma and squamous cell carcinoma cells but increase in glioblastomacells⁵⁷ BA can also inhibit aminopeptidase N, an enzyme involved in the regulation of angiogenesis that is overexpressed in several cancers⁵⁸ Jujube extract is used in the treatment of breast cancer in traditional Chinese medicine; the bioactive compounds of triterpenic acids, such as ZE2 and ZE4, are present in the extract. These compounds are effective in inhibiting cell growth and inducing cell apoptosis, as has been shown in in vitro studies on the MCF-7 (estrogen receptor alpha-positive) and SKBR3 (estrogen receptor alpha-negative) breast cancer cell lines, by DNA fragmentation and terminal deoxynucleotidyltransferase-mediated deoxyuridine triphosphate nick endlabeling (TUNEL) staining.[59]In another study, deproteinized polysaccharide (DPP) obtained from the water extract of jujube revealed an antiproliferative effect in vitro on melanoma cells in a dose- and time-dependent fashion. Also, the cell cycle assay showed melanoma cells to be arrested in the G2/M phases. Additionally, there was a generation of apoptotic bodies accompanied by an increase in caspase-3 and caspase-9 activities⁵⁹ Finally, in a study in 2012, normal liver (Chang) and lung (Hel 299) cells and HeLa cervical cancer cells, A549 lung cancer cells, and U937 lymphoma cancer cells were treated with jujube extracts in different (eight; S1-S8) stages of maturation, at four concentrations (1 µg/ml, 10 µg/ml, 50 µg/ml, and 100 µg/ml). HeLa cervical cancer cells were inhibited by the extract derived at all growth stages dose-dependently, whereas the inhibition of Hel 299 normal lung and A549 lung cancer cells decreased with fruit maturation in correlation with the flavonoid content. Therefore, fruit maturity may have an influence on both the nutritional and bioactive components of the jujube and, consequently, on anticancer activity.

CANCER AND CHEMORESISTANCE

Although chemotherapy is an effective treatment for some human cancers, chemoresistance remains a major hurdle for successful clinical treatment. Chemoresistance is a multifactorial difficulty and has been traditionally thought to arise from altered drug transportation, modified drug targets, gene expression (e.g. multidrug resistance gene), and reduced drug-induced macromolecular damage or increased DNA repair efficiency⁶⁰ Recent evidence has suggested that the inability of the cells to undergo apoptosis is one of the key determinants for chemoresistance⁶¹ Dysregulation of antiapoptotic [e.g. Akt, X-linked IAP (Xiap), and FLICE-like inhibitory protein (FLIP)⁶²⁻⁶⁵ and proapoptotic (e.g. Fas, caspases, and p53⁶⁶⁻⁶⁷ pathways have been found in chemoresistant cancer cells.In these contexts, it has been shown that FLIP is a determinant of OVCA (ovarian cancer) chemoresistance and *cis*-diamminedichloroplatinium (CDDP, cisplatin) decreases FLIPL and FLIPS (two FLIP isoforms) contents in chemosensitive OVCA cells but not in their resistant counterparts.[64] We have also tested the hypothesis that the inability of CDDP decreases FLIP content and induces apoptosis in the cisplatin-sensitive cells but not in the -resistant counterpart; the overexpression of FLIP by cDNA transfection is effective in attenuating CDDP-induced apoptosis in chemosensitive cells; and FLIP siRNA expression facilitates apoptotic cell death in the chemoresistant counterpart induced by CDDP.

These findings suggest that the downregulation of FLIP may increase the sensitivity of chemoresistant cells to CDDP and may be a potential therapeutic strategy for CDDP-resistant OVCA associated with FLIP overexpression. The tumor suppressor protein p53 is a transcription factor regulating the cell cycle, DNA repair, and apoptosis, and is rapidly upregulated by DNA-damaging agents including CDDP. ⁶⁸ It is maintained at low levels by its negative regulator, mouse double minute 2 homolog (MDM2), which ubiquitinates p53, targeting it for proteasomal degradation. ⁶⁹ TP53 mutations are frequently observed in human OVCA cells ⁷⁰ and are associated with decreased chemo responsiveness. In this regard, we also investigated the involvement of Itch, an E3-ligase protein, and p53 in CDDP-induced FLIP downregulation. We showed that CDDP enhances FLIP-p53-Itch interaction and induces FLIP ubiquitination and degradation in a p53- and Itch-dependent fashion. These results suggest that the modulation of FLIP content may be an effective strategy to overcome chemoresistance in OVCA⁷¹ Akt/PKB (protein kinase B) promotes survival and malignant transformation in different cancers ⁷²⁻⁷³ It is a determinant of CDDP resistance, and activation of the PI-3K/Akt pathway increases FLIP mRNA and/or protein expression in human cancer cells ⁷⁴⁻⁷⁷ We have also demonstrated that p53 facilitates FLIP-Itch interactions and FLIP ubiquitination and degradation in chemosensitive cells. However, in chemoresistant cells, Akt inhibits FLIP-p53 interaction, FLIP ubiquitination, and apoptosis, suggesting that Akt modulation may be an effective means to overcome chemoresistance in human OVCA⁷⁸

Angiogenesis, the growth of new capillaries from preexisting ones, is a regulated process which is modulated by numerous angiogenic and antiangiogenic factors. Since cancer cells require to blood vessel supply for growth and metastasis therefore, tumor angiogenesis is an important factor in the progression of cancer. Nowadays, despite advances in the therapeutic modalities treatment, results are disappointing in cancer patients hence; identification of antiangiogenic molecules including bioactive compounds present in medicinal herbs such as z. jujuba can be considered as an adjuvant strategy beside standard methods for cancer treatment⁷⁹⁻⁸¹]

It has also been shown that gelsolin (GSN) plays a key role in the regulation of gynecological and Head–Neck cancer chemo resistance, it may be an appropriate therapeutic target in chemo resistant cancers. 82-83 It has been shown that *Z. jujuba* shows cytotoxic activity; however, if and how it regulates CDDP-induced apoptosis and chemo sensitivity is still unclear. Understanding the mechanism by which *Z. jujuba* shows its cytotoxic activity can improve therapeutic strategies for future cancer treatment.

CONCLUSION

Phytochemical data combined with information about biological activity confirm that jujube fruits are rich in bioactive compounds that can benefit human health. This review article shows that among the bioactive compounds present in jujube fruit, triterpenic acids and polysaccharides have antiproliferative and anticancer effects on various cancer cell lines. It seems that the induction of apoptosis is one of the main mechanisms for the anticancer activities of the jujube fruit, due to the presence of the bioactive compounds. However, the biological effects of other identified and unidentified compounds in these fruits should be also investigated in areas where there are suitable climatic conditions for the cultivation of these plants.

Conflict of Interest:

None of the authors have any conflicting interest with the submission of the manuscript.

REFERENCES

- 1. Sampson W. Studying herbal remedies. N Engl J Med. 2005;353:337
- 2. Huang YL, Yen GC, Sheu F, Chau CF. Effects of water-soluble carbohydrate concentrate from Chinese jujube on different intestinal and fecal indices. *J Agric FoodChem.* 2008;56:1734–9.
- 3. Plastina P, Bonofiglio D, Vizza D, Fazio A, Rovito D, Giordano C, et al. Identification of bioactive constituents of *Ziziphus jujube* fruit extracts exerting antiproliferative and apoptotic effects in human breast cancer cells. *J Ethnopharmacol.* 2012;140:325–32.
- 4. Vahedi F, FathiNajafi M, Bozari K. Evaluation of inhibitory effect and apoptosis induction of *Zyzyphus Jujube* on tumor cell lines, an *in vitro* preliminary study. *Cytotechnology*. 2008; 56:105–11.
- 5. Li J, Chen Y, Ding S, Zhang L. Isolation and Analysis of a Novel Proteoglycan from Zizyphus jujuba cv. Jinsixiaozao. *J Food Drug Anal.* 2007;15:271–7.
- 6. Shen X, Tang Y, Yang R, Yu L, Fang T, Duan JA. The protective effect of *Zizyphus jujube* fruit on carbon tetrachloride-induced hepatic injury in mice by anti-oxidative activities. *J Ethnopharmacol*. 2009;122:555–60.
- 7. Pawlowska AM, Camangi F, Bader A, Braca A. Flavonoids of *Zizyphus jujuba* L. and *Zizyphusspina-christi* (L.) Willd (Rhamnaceae) fruits. *Food Chem.* 2009;112:858–62.
- 8. Li J, Ding S, Ding X. Optimization of the ultrasonically assisted extraction of polysaccharides from *Zizyphus jujuba* cv. *jinsixiaozao*. *J Food Eng*. 2007; b80:176–83.
- 9. Ghaly IS, Said A, Abdel-Wahhab MA. Zizyphus jujuba and Origanummajorana extracts protect against hydroquinone-induced clastogenicity. *Environ ToxicolPharmacol*. 2008;25:10–9.
- 10. Saika K, Sobue T. Cancer statistics in the world. GanTo Kagaku Ryoho. 2013;40:2475–80.
- 11. Ogura M. Adriamycin (doxorubicin) GanTo Kagaku Ryoho. 2001;28:1331–8.
- 12. HassanpourFard M, Ghule AE, Bodhankar SL, Dikshit M. Cardioprotective effect of whole fruit extract of pomegranate on doxorubicin-induced toxicity in rat. *Pharm Biol.* 2011; 49:377–82.
- 13. HassanpourFard M, Naseh G, Bodhankar SL, Dikshit M. Cardioprotective effect of *Lagenariasiceraria* (Molina) standley (Cucurbitaceae) fruit juice on doxorubicin induced cardiotoxicity in rats. *Am J PharmacolToxicol*. 2010; 5:103–8.

- 14. HassanpourFard M, Bodhankar SL, Dikshit M. Cardioprotective activity of fruit of *Lagenariasiceraria* (Molina) Standley on Doxorubicin induced cardiotoxicity in rats. *Int J Pharmacol.* 2008; 6:466–71.
- Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981-2002. J Nat Prod. 2003; 66:1022-37
- 16. San B, Yildirim AN. Phenolic, alpha-tocopherol, beta-carotene and fatty acid composition of four promising jujube (*Ziziphus jujuba* Miller) selections. *J Food Compost Anal.* 2010; 23:706–10
- 17. Choi SH, Ahn JB, Kim HJ, Im NK, Kozukue N, Levin CE, et al. Changes in free amino acid, protein, and flavonoid content in jujube (*Ziziphus jujube*) fruit during eight stages of growth and antioxidative and cancer cell inhibitory effects by extracts. *J Agric Food Chem.* 2012; 60:10245–55.
- 18. Gusakova SD, Sagdullaev SS, Aripov KN, Basher KH, Kurkcuoglu M, Demirci B. Isomers of plamitoleic acid in lipids and volatile substances from the fruits of *Ziziphus jujuba*. *Chem Nat Compd.* 1999; 35:401–3
- 19. Li J, Fan L, Ding S, Ding X. Nutritional composition of five cultivars of Chinese jujube. Food Chem. 2007; 103:454-60.
- 20. Barrett DM, Lloyd B. Advanced preservation methods and nutrient retention in fruits and vegetables. *J Sci Food Agric*. 2012; 92:7–22
- 21. Gao QH, Wu CS, Wang M. The jujube (*Ziziphus jujuba* Mill.) fruit: A review of current knowledge of fruit composition and health benefits. *J Agric Food Chem.* 2013; 61:3351–63.
- 22. Carlsen MH, Halvorsen BL, Holte K, Bøhn SK, Dragland S, Sampson L, et al. The total antioxidant content of more than 3100 foods, beverages, spices, herbs and supplements used worldwide. *Nutr J.* 2010;9:3
- 23. D'Archivio M, Filesi C, Di Benedetto R, Gargiulo R, Giovannini C, Masella R. Polyphenols, dietary sources and bioavailability. *Ann 1st Super Sanita*. 2007;43:348–61. [
- 24. Mignogna M, Fedele S, Lo Russo L. The World Cancer Report and the burden of oral cancer. *Eur J Cancer Prev.* 2004;13:139–42.
- 25. Mans DR, da Rocha AB, Schwartsmann G. Anti-cancer drug discovery and development in Brazil: Targeted plant collection as a rational strategy to acquire candidate anti-cancer compounds. *Oncologist*. 2000:185–98.
- 26. Bown D. London: Ball Pub; 1995. Encyclopedia of Herbs and their Uses; pp. 240-1
- 27. Fan YM, Xu LZ, Gao J, Wang Y, Tang XH, Zhao XN, et al. Phytochemical and antiinflammatory studies on Terminalia catappa. *Fitoterapia*. 2004;75:253–60.
- 28. Gbaguidi F, Accrombessi G, Moudachirou M, Quetin-Leclercq J. HPLC quantification of two isomeric triterpenic acids isolated from Mitracarpusscaber and antimicrobial activity on Dermatophiluscongolensis. *J Pharm Biomed Anal.* 2005;39:990–5.
- 29. Liu Y, Hartley DP, Liu J. Protection against carbon tetrachloride hepatotoxicity by oleanolic acid is not mediated through metallothionein. *ToxicolLett.* 1998;95:77–85.
- 30. Kalogeropoulos N, Chiou A, Ioannou M, Karathanos VT, Hassapidou M, Andrikopoulos NK. Nutritional evaluation and bioactive microconstituents (phytosterols, tocopherols, polyphenols, triterpenic acids) in cooked dry legumes usually consumed in the Mediterranean countries. *Food Chem.* 2010;121:682–90.
- 31. Romero C, García A, Medina E, Ruíz-Méndez V, Castro A, Brenes M. Triterpenic acids in table olives. *Food Chem.* 2010;118:670–4.
- 32. Guo S, Duan JA, Tang Y, Su S, Shang E, Ni S, et al. High-performance liquid chromatography--Two wavelength detection of triterpenoid acids from the fruits of *Ziziphus jujuba* containing various cultivars in different regions and classification using chemometric analysis. *J Pharm Biomed Anal.* 2009;49:1296–302
- 33. Shyu MH, Kao TC, Yen GC. Oleanolic acid and ursolic acid induce apoptosis in HuH7 human hepatocellular carcinoma cells through a mitochondrial-dependent pathway and downregulation of XIAP. *J Agric Food Chem.* 2010;58:6110–8.
- 34. Pisha E, Chai H, Lee IS, Chagwedera TE, Farnsworth NR, Cordell GA, et al. Discovery of betulinic acid as a selective inhibitor of human melanoma that functions by induction of apoptosis. *Nat Med.* 1995;1:1046–51.
- 35. Fulda S. Betulinic acid: A natural product with anticancer activity. MolNutr Food Res. 2009;53:140-6.
- 36. Kim DK, Baek JH, Kang CM, Yoo MA, Sung JW, Chung HY, et al. Apoptotic activity of ursolic acid may correlate with the inhibition of initiation of DNA replication. *Int J Cancer*. 2000;87:629–36.
- 37. De Angel RE, Smith SM, Glickman RD, Perkins SN, Hursting SD. Antitumor effects of ursolic acid in a mouse model of postmenopausal breast cancer. *Nutr Cancer*. 2010;62:1074–86.
- 38. Kowalski R. Studies of selected plant raw materials as alternative sources of triterpenes of oleanolic and ursolic acid types. *J Agric Food Chem.* 2007;55:656–62
- 39. Li J, Guo WJ, Yang QY. Effects of ursolic acid and oleanolic acid on human colon carcinoma cell line HCT15. World J Gastroenterol. 2002;8:493–5
- 40. Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer*. 2010;127:2893–917.
- 41. Levrero M. Viral hepatitis and liver cancer: The case of hepatitis C. Oncogene. 2006;25:3834–47]
- 42. Zhang P, Li H, Chen D, Ni J, Kang Y, Wang S. Oleanolic acid induces apoptosis in human leukemia cells through caspase activation and poly (ADP-ribose) polymerase cleavage. *ActaBiochimBiophys Sin (Shanghai)* 2007;39:803–9.
- 43. Manu KA, Kuttan G. Ursolic acid induces apoptosis by activating p53 and caspase-3 gene expressions and suppressing NF-kappaB mediated activation of bcl-2 in B16F-10 melanoma cells. *IntImmunopharmacol.* 2008;8:974–81
- 44. Kassi E, Sourlingas TG, Spiliotaki M, Papoutsi Z, Pratsinis H, Aligiannis N, et al. Ursolic acid triggers apoptosis and Bcl-2 downregulation in MCF-7 breast cancer cells. *Cancer Invest.* 2009;27:723–33.

- 45. Zhang YX, Kong CZ, Wang HQ, Wang LH, Xu CL, Sun YH. Phosphorylation of Bcl-2 and activation of caspase-3 via the c-Jun N-terminal kinase pathway in ursolic acid-induced DU145 cells apoptosis. *Biochimie*. 2009;91:1173–9.
- 46. Riker AI, Zea N, Trinh T. The epidemiology, prevention, and detection of melanoma. Ochsner J. 2010;10:56–65.
- 47. Selzer E, Pimentel E, Wacheck V, Schlegel W, Pehamberger H, Jansen B, et al. Effects of betulinic acid alone and in combination with irradiation in human melanoma cells. *J Invest Dermatol*. 2000;114:935–40.
- 48. Thurnher D, Turhani D, Pelzmann M, Wannemacher B, Knerer B, Formanek M, et al. Betulinic acid: A new cytotoxic compound against malignant head and neck cancer cells. *Head Neck*. 2003;25:732–40.
- 49. Chintharlapalli S, Papineni S, Ramaiah SK, Safe S. Betulinic acid inhibits prostate cancer growth through inhibition of specificity protein transcription factors. *Cancer Res.* 2007;67:2816–23
- 50. Zuco V, Supino R, Righetti SC, Cleris L, Marchesi E, Gambacorti-Passerini C, et al. Selective cytotoxicity of betulinic acid on tumor cell lines, but not on normal cells. *Cancer Lett.* 2002;175:17–25.
- 51. Damle AA, Pawar YP, Narkar AA. Anticancer activity of betulinic acid on MCF-7 tumors in nude mice. *Indian J Exp Biol.* 2013;51:485–91.
- 52. Moodi M, Norozi E, Rezaeiyan M, Mostafavi F, Sharifirad G. Comparing the effectiveness of three educational interventions based on health belief model on mammography screening behaviors in women above 40 years in Isfahan, Iran. *J Health Syst Res.* 2013;9:385–94.
- 53. Mullauer FB, Kessler JH, Medema JP. Betulinic acid, a natural compound with potent anticancer effects. *Anticancer Drugs*. 2010;21:215–27.
- 54. de Vries EG, Gietema JA, de Jong S. Tumor necrosis factor-related apoptosis-inducing ligand pathway and its therapeutic implications. *Clin Cancer Res.* 2006;12:2390–3.
- 55. Takada Y, Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B alpha kinase and p65 phosphorylation: Abrogation of cyclooxygenase-2 and matrix metalloprotease-9. *J Immunol.* 2003;171:3278–86.
- 56. Adams JM, Cory S. The Bcl-2 apoptotic switch in cancer development and therapy. Oncogene. 2007;26:1324–37
- 57. Selzer E, Thallinger C, Hoeller C, Oberkleiner P, Wacheck V, Pehamberger H, et al. Betulinic acid-induced Mcl-1 expression in human melanoma-mode of action and functional significance. *Mol Med.* 2002;8:877–84
- 58. Kwon HJ, Shim JS, Kim JH, Cho HY, Yum YN, Kim SH, et al. Betulinic acid inhibits growth factor-induced *in vitro* angiogenesis via the modulation of mitochondrial function in endothelial cells. *Jpn J Cancer Res.* 2002;93:417–25.
- 59. Hung CF, Hsu BY, Chang SC, Chen BH. Antiproliferation of melanoma cells by polysaccharide isolated from Zizyphus jujuba. *Nutrition*. 2012;28:98–105
- 60. Reed JC, Miyashita T, Takayama S, Wang HG, Sato T, Krajewski S, et al. BCL-2 family proteins: Regulators of cell death involved in the pathogenesis of cancer and resistance to therapy. *J Cell Biochem.* 1996;60:23–32
- 61. Cheng JQ, Jiang X, Fraser M, Li M, Dan HC, Sun M, et al. Role of X-linked inhibitor of apoptosis protein in chemoresistance in ovarian cancer: Possible involvement of the phosphoinositide-3 kinase/Akt pathway. *Drug Resist Updat.* 2002;5:131–46
- 62. Sasaki H, Sheng Y, Kotsuji F, Tsang BK. Down-regulation of X-linked inhibitor of apoptosis protein induces apoptosis in chemoresistant human ovarian cancer cells. *Cancer Res.* 2000;60:5659–66.
- 63. Li J, Feng Q, Kim JM, Schneiderman D, Liston P, Li M, et al. Human ovarian cancer and cisplatin resistance: Possible role of inhibitor of apoptosis proteins. *Endocrinology*. 2001; 142:370–80
- 64. Abedini MR, Qiu Q, Yan X, Tsang BK. Possible role of FLICE-like inhibitory protein (FLIP) in chemoresistant ovarian cancer cells *in vitro*. *Oncogene*. 2004; 23:6997–7004.
- 65. Fraser M, Leung BM, Yan X, Dan HC, Cheng JQ, Tsang BK. p53 is a determinant of X-linked inhibitor of apoptosis protein/Akt-mediated chemoresistance in human ovarian cancer cells. *Cancer Res.* 2003; 63:7081–8.
- 66. Schneiderman D, Kim JM, Senterman M, Tsang BK. Sustained suppression of Fas ligand expression in cisplatin-resistant human ovarian surface epithelial cancer cells. *Apoptosis*. 1999;4:271–81.
- 67. Fraser M, Bai T, Tsang BK. Akt promotes cisplatin resistance in human ovarian cancer cells through inhibition of p53 phosphorylation and nuclear function. *Int J Cancer*. 2008;122:534–46.
- 68. Buttitta F, Marchetti A, Gadducci A, Pellegrini S, Morganti M, Carnicelli V, et al. p53 alterations are predictive of chemoresistance and aggressiveness in ovarian carcinomas: A molecular and immunohistochemical study. *Br J Cancer*. 1997;75:230–5
- 69. Honda R, Tanaka H, Yasuda H. Oncoprotein MDM2 is a ubiquitin ligase E3 for tumor suppressor p53. FEBS Lett. 1997; 420:25–7
- 70. Kmet LM, Cook LS, Magliocco AM. A review of p53 expression and mutation in human benign, low malignant potential, and invasive epithelial ovarian tumors. *Cancer*. 2003;97:389–404.
- 71. Abedini MR, Muller EJ, Brun J, Bergeron R, Gray DA, Tsang BK. Cisplatin induces p53-dependent FLICE-like inhibitory protein ubiquitination in ovarian cancer cells. *Cancer Res.* 2008;68:4511–7.
- 72. Shayesteh L, Lu Y, Kuo WL, Baldocchi R, Godfrey T, Collins C, et al. PIK3CA is implicated as an oncogene in ovarian cancer. *Nat Genet*. 1999;21:99–102.
- 73. Sun M, Wang G, Paciga JE, Feldman RI, Yuan ZQ, Ma XL, et al. AKT1/PKBalpha kinase is frequently elevated in human cancers and its constitutive activation is required for oncogenic transformation in NIH3T3 cells. *Am J Pathol.* 2001; 159:431–7.
- 74. Nam SY, Amoscato AA, Lee YJ. Low glucose-enhanced TRAIL cytotoxicity is mediated through the ceramide-Akt-FLIP pathway. *Oncogene*. 2002; 21:337–46.

- 75. Nam SY, Jung GA, Hur GC, Chung HY, Kim WH, Seol DW, et al. Upregulation of FLIP (S) by Akt, a possible inhibition mechanism of TRAIL-induced apoptosis in human gastric cancers. *Cancer Sci.* 2003; 94:1066–73
- 76. Panka DJ, Mano T, Suhara T, Walsh K, Mier JW. Phosphatidylinositol 3-kinase/Akt activity regulates c-FLIP expression in tumor cells. *J Biol Chem.* 2001; 276:6893–6.
- 77. Suhara T, Mano T, Oliveira BE, Walsh K. Phosphatidylinositol 3-kinase/Aktsignaling controls endothelial cell sensitivity to Fasmediated apoptosis via regulation of FLICE-inhibitory protein (FLIP) *Circ Res.* 2001;89:13–9
- 78. Abedini MR, Muller E, Bergeron R, Gray D, Tsang BK. Akt promotes chemoresistance in human ovarian cancer cells by modulating cisplatin-induced, p53-dependent ubiquitination of FLICE-like inhibitory protein. *Oncogene*. 2009; 29:11–25.
- 79. Tahergorabi Z, Khazaei M. The relationship between inflammatory markers, angiogenesis, and obesity. *ARYA Atheroscler*. 2013; 9:247–53.
- 80. Khazaei M, Kalantari E, Saeidi H, ShabaniKia N, Tahergorabi Z, Rashidi B, et al. Gamma-secretase inhibitor does not modulate angiogenesis in colon adenocarcinoma in obese mice. *BratislLekListy*. 2015; 116:248–51
- 81. Kalantari E, Saeidi H, Kia NS, Tahergorabi Z, Rashidi B, Dana N, et al. Effect of DAPT, a gamma secretase inhibitor, on tumor angiogenesis in control mice. *Adv Biomed Res.* 2013;30:83
- 82. Abedini MR, Wang PW, Huang YF, Cao M, Chou CY, Shieh DB, et al. Cell fate regulation by gelsolin in human gynecologic cancers. *ProcNatlAcadSci U S A*. 2014; 111:14442–7.
- 83. Wang PW, Abedini MR, Yang LX, Ding AA, Figeys D, Chang JY, et al. Gelsolin regulates cisplatin sensitivity in human head-and-neck cancer. *Int J Cancer*. 2014; 135:2760–9.



