

Stronger together



Marius Lazar¹, Lucas Smith¹, Olga Ivanova¹, David Seideman^{1,2}, Marianne Kaufmann³, Sophia Papadopoulou¹, Adrian Tudor¹, Jonathan Feldman¹, Georg Schuster², Riku Honda³, Jacob Adams³, Carolina Diamandis¹

Institutions: 1) LCG Greece Research, 2) Jewish University of Colorado, 3) International H63D Syndrome Research Consortium

Corresponding author: LCG Greece Dr. Carolina Diamandis, 16 Kifissias Avenue, 115 26 Athens, Hellenic Republic

HFE H63D mutation: why are you killing your patients?

ABSTRACT

Evidence-based medicine has shown for many years that homozygous mutations of the HFE gene H63D are by no means negligible. Not only can it cause, usually after a second hit, rather mild classical hemochromatosis, but it can also cause numerous other disorders of iron metabolism, such as hypotransferrinemia, changes in binding capacity, and others. In addition, it may lead - among other symptoms - to damages of the heart and the substantia nigra via a causal relationship that remains to be investigated, most likely via a cascade dysfunction in iron metabolism. The clinical facts are compelling. Any physician who dismisses mutations of the HFE gene H63D as clinically irrelevant risks the health and life of his patient. We report two patients with an HFE H63D mutation who were treated almost too decades too late because of outdated expertise.

INTRODUCTION

Homozygous mutations of the HFE gene H63D have not been taken seriously enough for many decades. This mutation of gene H63D is a Pandora's box. It has been linked to liver disease, bone and joint disease, diabetes mellitus, heart disease, hormonal disorders, porphyria cutanea tarda (PCT), infertility, stroke, severe neurodegenerative disease, cancer, venous peripheral artery disease, hereditary hemochromatosis (after a second hit), and H63D syndrome. In the years since the discovery of HFE and its mutations, researchers have focused their studies primarily on the C282Y mutation because it is particularly common in people with elevated iron levels. About 85% of people with abnormally high iron levels have two copies of C282Y, so this mutation has been studied more intensively. Other mutations, such as S65C or H63D, have not attracted the attention of researchers. The S65C mutation can lead to mild to moderate hepatic (liver) iron overload, especially in combination with other mutations. Increased serum iron indices and iron overload have been observed in C282Y/ S65C compound heterozygotes. In scientific evaluation, H63D stands out as a significant modifier of disease onset, disease progression and even

response to therapy. H63D is associated with arterial rigidity, prooxidation, higher total and low-density lipoprotein cholesterol, acute lymphoblastic leukemia (ALL), decreased sperm production, and higher risk of type II diabetes mellitus, and hereditary hemochromatosis after a second hit. Being a carrier of the H63D hemochromatosis mutation is also a risk factor for earlier onset and longer duration of kidney disease in type II diabetics. The most striking risk associated with H63D is that for neurodegenerative disease. Connor and colleagues were among the first researchers to examine the role of H63D in brain iron accumulation, oxidative stress, and neurotransmitter performance. Connor reported that the HFE variant H63D contributes to many of the processes associated with various types of dementia. These processes include increased cellular iron, oxidative stress (free radical activity), glutamate dyshomeostasis (abnormal balance), and an increase in tau phosphorylation (abnormal levels of tau proteins can lead to dementias such as Alzheimer's disease). As demonstrated by Jacobs, Papadopoulos Kaufmann, and colleagues (2012, 2015, 2017, 2019, 2020, 2021) using solid patient data, the numerous damages in parenchymal tissues, heart, and brain (substantia nigra and basal ganglia) can be explained by insidious non-transferrin-bound iron (NTBI) intoxication as a consequence of chronic transferrin saturation of >50%. This constellation (H63D syndrome) is similar to Wilson's disease, except that NTBI iron, rather than copper, is the culprit here. In addition, the damage caused by H63D syndrome is more widespread in the body, affecting not only the liver but also the heart, brain, and in men, the testes. Synucleinopathies are a major problem of H63D syndrome, but other forms of cognitive decline are also common. Connor states further that HFE H63D cells have been shown to have more oxidative stress, further supporting their role as modifiers of neurodegenerative diseases. He found that patients homozygous for H63D had earlier signs of mild cognitive impairment and earlier onset of dementia disease than patients with normal HFE H63D or H63D heterozygote individuals. Despite this fact, which has been known for 25 years, many clinicians still dismiss homozygous HFE-H63D mutations as irrelevant. Even some of the highest authorities in the field of iron metabolism seem to be trapped in the knowledge of the early 1990s.¹⁻¹⁷

CASES

Patient 1

Male, 46 years, Southern European, with Parkinson's-like neurological symptoms, incipient dementia, loss of IQ by >40%, steatosis hepatitis, regressed testes, cryptic (mainly vesicular) skin lesions, mildly impaired lipid metabolism (with a BMI of 22), and high-grade cardiac conduction abnormalities due to micro-scarring. The patient was unsuccessfully investigated by all means of modern medicine from 1997 to 2018, without any of the total >70 colleagues involved taking the patient's most severely disturbed iron metabolism seriously. The patient's ferritin has always been at the lower range of the norm, while his transferrin is statically 20% below the norm, no matter which laboratory or method is used. As a result, transferrin saturation (TFsat) values of 60-90% are the result. Postprandial no transferrin response, the level of this crucial iron transporter remains static. Due to the consequence, high TFsat values, it must be taken for granted that there is a constant release of NTBI in his organism. All other proteins, etc. are normal. Albumin is high-normal. We interpret this as a reactive attempt by the patient's body to compensate for the lack of transferrin, especially postprandial. In biopsies, no Prussian blue reaction, which was to be expected given his low ferritin and non-stainable NTBI. On skin biopsy, a histopathologist thought he saw what was considered iron in the dermal and sub-dermal layers. We found the patient's substantia nigra full of iron and scars (white in TCS), he has been suffering from non-motor Parkinsonism, movement disorders, tics, most severe narcolepsy with cataplexy, early signs of dementia, and REM sleep disorders. He also suffers from left bundle branch block (LBBB) due to micro-scarring with decreasing left ventricular ejection fraction (Teichhaus method). Sudden episodes of immune system overdrive are observed, resulting in small bullous skin and mucosal symptoms, petechiae, and leaking capillaries (with normal platelet count and other normal test parameters), as well as skin disorders that seasoned dermatologists refer to as "autoimmune" or "toxic." Steatosis hepatitis at a BMI of 23, regressive hyperplasia nodule in the liver with an abundance of calcifications, testicular parenchyma regressive with progressive calcifications (bilateral). Basophilia, eosinophilia, slightly low MCH, MCV with normal Hb. Chelation therapy was not tried due to very low ferritin levels. Administration of transferrin by plasma infusion was discussed but dismissed because of the often overshooting immune responses seen in this

patient; especially since the effect of constant plasma therapy is questionable in the presence of already manifest organ damage. A medically supervised low iron diet showed a drop in TFSat by one third with borderline low ferritin. If the high TFSat values had been considered clinically relevant before the organ damages manifested and the mutation of the HFE gene H63D had not been downplayed, the patient would be, most likely, in a much better clinical condition today. Instead, one organ after the other has been damaged. A low-iron diet currently remains the only causal treatment option in this patient, and the medium- and long-term prognosis is unforeseeable.

Patient 2

Male, 44 years, Western European. Latest since 2005 highly significant elevated ferritin levels with strikingly low transferrin values. The transferrin level is within normal range but insufficiently reactive. No treatment offered by any physician consulted in the United States and Europe for 16 years. In his mid-30s development of metabolic syndrome with hepatic steatosis and short after manifestation of type 2 diabetes. Still no reaction to the abnormal iron metabolism by all treating physicians. Instead, the patient was made responsible for his manifold symptoms due to his moderately elevated BMI. This is nothing but malpractice and victim blaming in a clinical setting. Still no treatment for his highly elevated ferritin for nine more years. Eventually detection of a homozygous HFE gene H63D mutation with the awareness of an endocrinologist and a practice for rare diseases. To date, no other mutations have been detectable. The patient will now be treated for hemochromatosis with abnormally reacting transferrin. One of the leading 'iron experts' in western Europe who ignored the highly abnormal ferritin and only partly reactive transferrin already before is still opposing treatment because one should "disregard the HFE H63D variant which has been well described as non deleterious". Basically, this statement is nothing less than a call for negligent homicide.

DISCUSSION

As physicians specialized in rare diseases, we regularly see patients with complex syndromes consistent with those mentioned before. Just as regularly homozygous mutations of HFE gene H63D are found as *primum movens* (primary cause) of complex metabolic and toxic syndromes. It is also typical for treating colleagues to ignore this finding, as old textbooks (and new ones copy-pasted from old ones) still state that the HFE gene H63D or its homozygous mutation would be clinically irrelevant.

This is false, misleading and potentially deadly misinformation.

The knowledge about the high clinical relevance is neither new nor a fringe topic. HFE H63D is not a strong hemochromatosis gene, however, with a second hit it can easily cause hereditary hemochromatosis. But even more important than this, a homozygous mutation of the HFE gene H63D is, according to overwhelming evidence, responsible for many cases of complex syndromes associated with heterogeneously altered iron metabolism.¹⁻⁶⁸

- Patient 1 has a type of "Iron Morbus Wilson," correctly referred to as "H63D syndrome", due to a persistent NTBI production in his organism. Treatment is mainly symptomatic, causal by a low iron diet under medical supervision. The prognosis is unclear.
- Patient 2, most likely due to a secondary hit (presumably his increased BMI), has developed hemochromatosis with further disturbances in his metabolism. This is due to the homozygous mutation of his HFE gene H63D. No other reason was found, not even remotely. His diabetes mellitus may very well be a consequence of his iron overload, a fact that has been obscured and ignored because of the stigmatization and discrimination against moderately obese patients. His prognosis is slightly better than that of patient 1, as classical hemochromatosis treatment may slow the progression of sequelae in him.

CONCLUSION

Both patients stand as *pars pro toto*. They are not exceptions but two typical representatives of a considerable patient collective. The correct assessment of the clinical relevance of a homozygous mutation of the gene HFE H63D with its heterogeneous clinical picture must finally be taken most seriously also outside the realm of medical research.

Otherwise, we as clinicians will continue to kill our patients based on outdated "knowledge", even though all recent findings on the HFE gene H63D are freely available to everyone.

CONFLICTS OF INTEREST

None declared.

SOURCES

1. Environ Health Perspect. 2010; HFE H63D Polymorphism as a Modifier of the Effect of Cumulative Lead Exposure on Pulse Pressure: the Normative Aging Study. Zhang A, Park SK, Wright RO, Weisskopf MG, Mukherjee B, Nie H, Sparrow D, Hu H. *Pol Arch Med Wewn.* 2010;120(4):127-31.
2. HFE gene mutations in patients with alcoholic liver disease. A prospective study from northwestern Poland. Raszeja- Wyszomirska J, Kurzawski G, Zawada I, Suchy J, Lubinski J, Milkiewicz P. *Respirology.* 2010 Jan;15(1):141-9.
3. Population-based study of cystic fibrosis disease severity and haemochromatosis gene mutations. Prata U, Quinn S, Blizzard LB, Reid DW. *J Exp Clin Cancer Res.* 2010 Mar 2;29:18. Association between C282Y and H63D mutations of the HFE gene with hepatocellular carcinoma in European populations: a meta-analysis. Jin F, Qu LS, Shen XZ. *J Alzheimers Dis.* 2010 Apr;20(1):333-41.
4. Prevalent iron metabolism gene variants associated with increased brain ferritin iron in healthy older men. Bartzokis G, Lu PH, Tishler TA, Peters DG, Kosenko A, Barrall KA, Finn JP, Villablanca P, Laub G, Altshuler LL, Geschwind DH, Mintz J, Neely E, Connor JR. *J Diabetes Complications.* 2010
5. Mutation H63D in the HFE gene confers risk for the development of type 2 diabetes mellitus but not for chronic complications. Colli ML, Gross JL, Canani LH. *Diabetes Care.* 2001 Jul;24(7):1187-91. Role of hemochromatosis C282Y and H63D mutations in HFE gene in development of type 2 diabetes and diabetic nephropathy. *Diabetes Metab.* 2002 Feb;28(1):33-8.
6. Clinical expression and insulin sensitivity in type 2 diabetic patients with heterozygous mutations for haemochromatosis. Van Lerberghe S, Hermans MP, Dahan K, Buyschaert M. *Endocrine.* 2004 Jul;24(2):111-4.
7. The HFE gene is associated to an earlier age of onset and to the presence of diabetic nephropathy in diabetes mellitus type 2. Oliva R, Novials A, Sánchez M, Villa M, Ingelmo M, Recasens M, Ascaso C, Bruguera M, Gomis R. *Gut.* 2002 Nov;51(5):723-30.
8. Mild iron overload in patients carrying the HFE S65C gene mutation: a retrospective study in patients with suspected iron overload and healthy controls. Holmström P, Marmur J, Eggertsen G, Gäfvels M, Stål P. *J Hepatol.* 2002 Apr;36(4):474-9.
9. Frequency of the S65C mutation of HFE and iron overload in 309 subjects heterozygous for C282Y. Wallace DF, Walker AP, Pietrangolo A, Clare M, Bomford AB, Dixon JL, Powell LW, Subramaniam VN, Dooley JS. *Biochim Biophys Acta.* 2010 Apr;1802(4):389-95.
10. Prolyl-peptidyl isomerase, Pin1, phosphorylation is compromised in association with the expression of the HFE polymorphic allele, H63D. Hall EC 2nd, Lee SY, Simmons Z, Neely EB, Nandar W, Connor JR. *Neurobiol Aging.* 2009 Expression of the HFE allelic variant H63D in SH-SY5Y cells affects tau phosphorylation at serine residues. Hall EC 2nd, Lee SY, Mairuae N, Simmons Z, Connor JR.
11. *Neurobiol Aging.* 2009 HFE polymorphisms affect cellular glutamate regulation. Mitchell RM, Lee SY, Simmons Z, Connor JR. *Neurobiol Aging.* 2004 Apr;25(4):465-74.
12. Evaluation of HFE (hemochromatosis) mutations as genetic modifiers in sporadic AD and MCI. Berlin D, Chong G, Chertkow H, Bergman H, Phillips NA, Schipper HM. *Ann Hematol.* 2009
13. An extensive analysis of the hereditary hemochromatosis gene HFE and neighboring histone genes: associations with childhood leukemia. Davis CF, Dorak MT. *Pediatr Blood Cancer.* 2009 Dec 15;53(7):1242-8.
14. Hereditary hemochromatosis gene (HFE) variants are associated with birth weight and childhood leukemia risk. Dorak MT, Mackay RK, Relton CL, Worwood M, Parker L, Hall AG. *Leuk Lymphoma.* 2006 Nov;47(11):2331-4. HFE gene mutations in patients with acute leukemia. Viola A, Pagano L, Laudati D, D'Elia R, D'Amico MR, Ammirabile M, Palmieri S, Prossomariti L, Ferrara F.
15. Pizza F, Antelmi E, Vandi S, Meletti S, Erro R, Baumann CR, Bhatia KP, Dauvilliers Y, Edwards MJ, Iranzo A, Overeem S, Tinazzi M, Liguori R, Plazzi G. The distinguishing motor features of cataplexy: a study from video-recorded attacks. *Sleep.* 2018 May 1;41(5). doi: 10.1093/sleep/zsy026. PMID: 29425380.
16. Moturi S, DeWolfe JL. Isolated Cataplexy. In: *Sleep Review Mag.,* 2010
17. Hartse KM, Zorick FJ, Sicklesteel JM, Roth T. Isolated cataplexy: a familial study. *Henry Ford Hosp Med J.* 1988;36(1):24-7.
18. Morgenthaler TI, Kapur VK, Brown T; Standards of Practice Committee of the American Academy of Sleep Medicine. Practice parameters for the treatment of narcolepsy and other hypersomnias of central origin. *Sleep.* 2007;30:1705-11.
19. Reading P. Cataplexy. *Pract Neurol.* 2019 Feb;19(1):21-27. doi: 10.1136/practneurol-2018-002001. Epub 2018 Oct 24. PMID: 30355740.
20. Matos N, Gaig C, Santamaria J, Iranzo A. Cataplexy causing subdural hematomas. *Sleep Med.* 2017 Feb;30:15-16. doi: 10.1016/j.sleep.2016.01.018. Epub 2016 Mar 7. PMID: 28215239.
21. Adams R, Mayhew IG. Neurologic diseases. *Vet Clin North Am Equine Pract.* 1985 Apr;1(1):209-34. doi: 10.1016/s0749-0739(17)30778-2. PMID: 3000543.
22. Peacock J, Benca RM. Narcolepsy: clinical features, co-morbidities & treatment. *Indian J Med Res.* 2010 Feb;131:338-49. PMID: 20308759.

23. Pillen S, Pizza F, Dhondt K, Scammell TE, Overeem S. Cataplexy and Its Mimics: Clinical Recognition and Management. *Curr Treat Options Neurol.* 2017 Jun;19(6):23. doi: 10.1007/s11940-017-0459-0. PMID: 28478511.
24. Jory C, Oak K, Organ C, Mclean B, Shankar R. Head first - Review of epilepsy head injury risk and protection. *Seizure.* 2019 Oct;71:66-79. doi: 10.1016/j.seizure.2019.06.013. Epub 2019 Jun 12. PMID: 31207395.
25. Treffer E, Fitzgerald SG, Hobson DA, Bursick T, Joseph R. Outcomes of wheelchair systems intervention with residents of long-term care facilities. *Assist Technol.* 2004; 16(1):18-27. PMID: 15357146
26. LaPlante MP. Demographics of wheeled mobility device users. In: *Proceedings of the Conference on Space Requirements for Wheeled Mobility*; 2003 Oct 9-11; Buffalo (NY). Buffalo (NY): University at Buffalo, State University of New York; 2003.
27. Simpson RC et al. How many people would benefit from a smart wheelchair? In: *Journal of Rehabilitation Research & Development.* Volume 45, Number 1, 2008
28. Chang ET, Lin CL, Chen SF, Hsu CY, Shen YC. Risk of bone fractures in patients with narcolepsy: a nationwide population-based cohort study. *Sleep Med.* 2020 Jun;70:55-59. doi: 10.1016/j.sleep.2020.02.015. Epub 2020 Feb 26. PMID: 32197225.
29. Diamandis C, Tudor A: Medical devices that should be prescribed to patients with cataplexy to reduce their risk of injury. *Authorea Publishing*, 2021 doi: 10.22541/au.162187144.47184988/v1
30. Diamandis C, Adams Jacob S, Honda R, et al. Regularly missed symptoms in primary & secondary narcolepsy. *Authorea Publishing*, 2021 doi: 10.22541/au.162134961.11409756/v1
31. Seideman D et al. H63D Syndrome: What we know about it in 2021. *Authorea Publishing*, 2021 doi: 10.22541/au.162191694.46218714/v1
32. Sørensen S et al. H63D Syndrome: Consensus Paper of the International H63D Research Consortium, Oslo 2019
33. Deekollu et al. Seizure-related injuries in a group of young people with epilepsy wearing protective helmets: Incidence, types and circumstances, *Seizure*, Volume 14, Issue 5, 2005, Pages 347-353, ISSN 1059-1311, doi.org/10.1016/j.seizure.2005.04.008.
34. Camfield C, Camfield P. Injuries from seizures are a serious, persistent problem in childhood onset epilepsy: a population-based study. *Seizure.* 2015 Apr;27:80-3. doi: 10.1016/j.seizure.2015.02.031. Epub 2015 Mar 6. PMID: 25891933
35. Wirrell EC. Epilepsy-related injuries. *Epilepsia.* 2006;47 Suppl 1:79-86. doi: 10.1111/j.1528-1167.2006.00666.x. PMID: 17044832.
36. Frey K, Zöllner JP, Knake S, Oganian Y, Kay L, Mahr K, Keil F, Willems LM, Menzler K, Bauer S, Schubert-Bast S, Rosenow F, Strzelczyk A. Risk incidence of fractures and injuries: a multicenter video-EEG study of 626 generalized convulsive seizures. *J Neurol.* 2020 Dec;267(12):3632-3642. doi: 10.1007/s00415-020-10065-5. Epub 2020 Jul 10. PMID: 32651672; PMCID: PMC7674387.
37. Kostas Pantopoulos: Inherited Disorders of Iron Overload. *Front. Nutr.* 5:103. doi: 10.3389/fnut.2018.0010
38. Wint Nandar, James R. Connor: HFE Gene Variants Affect Iron in the Brain. *The Journal of Nutrition*, Volume 141, Issue 4, April 2011
39. Dekker MC, Giesbergen PC, Njajou OT, van Swieten JC, Hofman A, 127. Breteler MM, van Duijn CM. Mutations in the hemochromatosis gene (HFE), Parkinson's disease and parkinsonism. *Neurosci Lett.* 2003;348:117-119
40. Steven M. LeVine, James R. Connor, Hyman M. Schipper: Redoxactive Metals in Neurological Disorders. *New York Academy of Sciences*, 2004.
41. Sareen S. Gropper, Jack L. Smith, Timothy P. Carr: *Advanced Nutrition and Human Metabolism.* Cengage Learning, 7th edition, Boston 2016.
42. Bartzokis G, Lu PH, Tishler TA, Peters DG, Kosenko A, Barrall KA, Finn JP, Villablanca P, Laub G, Altshuler LL, Geschwind DH, Mintz J, Neely E, Connor JR: Prevalent iron metabolism gene variants associated with increased brain ferritin iron in healthy older men. *J Alzheimers Dis.* 2010 Apr;20(1):333-341.
43. Brissot P, Ropert M, Le Lan C, Loreal O. Non-transferrin bound iron: a key role in iron overload and iron toxicity. *BBA Gen Subjects* (2012) 1820:403-10. doi: 10.1016/j.bbagen.2011.07.014
44. Athiyarath R, Arora N, Fuster F, Schwarzenbacher R, Ahmed R, George B, et al. Two novel missense mutations in iron transport protein transferrin causing hypochromic microcytic anaemia and haemosiderosis: molecular characterization and structural implications. *Br J Haematol.* (2013) 163:404- 7. doi: 10.1111/bjh.12487
45. Akbas N, Hochstrasser H, Deplazes J, Tomiuk J, Bauer P, Walter U, Behnke S, Riess O, Berg D.: Screening for mutations of the HFE gene in Parkinson's disease patients with hyperechogenicity of the substantia nigra. *Neurosci Lett.* 2006
46. Borie C, Gasparini F, Verpillat P, Bonnet AM, Agid Y, Hetet G, Brice A, Durr A, Grandchamp B.: Association study between iron-related genes polymorphisms and Parkinson's disease. *J Neurol.* 2002; 249: 801-804.
47. Guerreiro RJ, Bras JM, Santana I, Januario C, Santiago B, 120. Morgadinho AS, Ribeiro MH, Hardy J, Singleton A, et al.: Association of HFE common mutations with Parkinson's disease, Alzheimer's disease and mild cognitive impairment in a Portuguese cohort. *BMC Neurol.* 2006;6:24
48. Fujii H, Takagaki N, Yoh T, Morita A, Ohkawara T, Yamaguchi K, Minami M, Sawa Y, Okanoue T, Ohkawara Y, Itoh Y: Non-prescription supplement-induced hepatitis with hyperferritinemia and mutation (H63D) in the HFE gene. *Hepatol Res.* 2008 Mar;38(3):319-323
49. Castiella, Urreta, Zapata et al.: H63/H63D genotype and the H63D allele are associated in patients with hyperferritinemia to the development of metabolic syndrome. *Eur J Intern Med.* 2019 Nov 30. doi:10.1016/j.ejim.2019.11.021
50. Gkouvatsos K, Papanikolaou G, Pantopoulos K. Regulation of iron transport and the role of transferrin. *Biochim Biophys Acta* (2012) 1820:188-202. doi: 10.1016/j.bbagen.2011.10.013
51. Mitchell RM, Lee SY, Simmons Z, Connor JR: HFE polymorphisms affect cellular glutamate regulation. *Neurobiol Aging.* 2009
52. Wint Nandar, James R. Connor: HFE Gene Variants Affect Iron in the Brain. *The Journal of Nutrition*, Volume 141, Issue 4, April 2011, 729S-739S, doi:10.3945/jn.110.130351
53. Borie C, Gasparini F, Verpillat P, Bonnet AM, Agid Y, Hetet G, Brice A, Durr A, Grandchamp B.: Association study between iron-related genes polymorphisms and Parkinson's disease. *J Neurol.* 2002; 249: 801-804
54. Steven M. LeVine, James R. Connor, Hyman M. Schipper: *Redoxactive Metals in Neurological Disorders.* New York Academy of Sciences, 2004
55. Valenti L, Fracanzani AL, Bugianesi E, Dongiovanni P, Galmozzi E, Vanni E, Canavesi E, Lattuada E, Roviato G, Marches G, Fargion S.: HFE genotype, parenchymal iron accumulation, and liver fibrosis in patients with nonalcoholic fatty liver disease. *Gastroenterology.* 2010 Mar;138(3):905-912.
56. P. Adams, P. Brissot, L. W. Powell: *EASL International Consensus Conference on Haemochromatosis.* *Journal of Hepatology* 2000;33:485-504.

57. de Valk, Addicks, Gosriwatana et al.: Non-transferrin-bound iron is present in serum of hereditary haemochromatosis heterozygotes. *Eur J Clin Invest.* 2000 Mar;30(3):248-51
58. Jakeman A, Thompson T, McHattie J, Lehotay DC: Sensitive method for nontransferrin-bound iron quantification by graphite furnace atomic absorption spectrometry. *Clin Biochem.* 2001 Feb;34(1):43-7
59. Nakamura M, Nishida S, Hayashida K, Ueki Y, Dauvilliers Y, Inoue Y. Differences in brain morphological findings between narcolepsy with and without cataplexy. *PLoS One.* 2013 Nov 28;8(11):e81059. doi: 10.1371/journal.pone.0081059. PMID: 24312261; PMCID: PMC3842956
60. Kumar S, Sagili H. Etiopathogenesis and neurobiology of narcolepsy: a review. *J Clin Diagn Res.* 2014 Feb;8(2):190-5. doi: 10.7860/JCDR/2014/7295.4057. Epub 2013 Dec 27. PMID: 24701532; PMCID: PMC3972560
61. Hauser RA. Levodopa: past, present, and future. *Eur Neurol.* 2009;62(1):1-8. doi: 10.1159/000215875. Epub 2008 Sep 9. PMID: 19407449
62. Jankovic J. Levodopa strengths and weaknesses. *Neurology.* 2002 Feb 26;58(4) doi: 10.1212/wnl.58.suppl_1.s19. PMID: 11909982
63. Black SW, Yamanaka A, Kilduff TS. Challenges in the development of therapeutics for narcolepsy. *Prog Neurobiol.* 2017 May;152:89-113. doi: 10.1016/j.pneurobio.2015.12.002. Epub 2015 Dec 23. PMID: 26721620; PMCID: PMC5114175
64. Aoun F, Slaoui A, Naoum E, Hassan T, Albisinni S, Azzo JM, Kallas-Chemaly A, Assenmacher G, Peltier A, Roumeguère T. Testicular microlithiasis: Systematic review and Clinical guidelines. *Prog Urol.* 2019 Sep;29(10):465-473. doi: 10.1016/j.purol.2019.07.001. Epub 2019 Aug 2. PMID: 31383508
65. Balawender K, Orkisz S, Wisz P. Testicular microlithiasis: what urologists should know. A review of the current literature. *Cent European J Urol.* 2018;71(3):310-314. doi: 10.5173/cej.2018.1728. Epub 2018 Aug 21. PMID: 30386652; PMCID: PMC6202617
66. Brambatti M, Matassini MV, Adler ED, Klingel K, Camici PG, Ammirati E. Eosinophilic Myocarditis: Characteristics, Treatment, and Outcomes. *J Am Coll Cardiol.* 2017 Nov 7;70(19):2363-2375. doi: 10.1016/j.jacc.2017.09.023. PMID: 29096807.
67. Oakley CM, Olsen GJ. Eosinophilia and heart disease. *Br Heart J.* 1977 Mar;39(3):233-7. doi: 10.1136/hrt.39.3.233. PMID: 320988; PMCID: PMC483226.
68. Maia Palhano AC, Kim LJ, Moreira GA, Santos Coelho FM, Tufik S, Levy Andersen M. Narcolepsy, Precocious Puberty and Obesity in the Pediatric Population: a Literature Review. *Pediatr Endocrinol Rev.* 2018 Dec;16(2):266-274. doi: 10.17458/per.vol16.2018.Narcolepsypubertyobesity. PMID: 30556659.

**Non-profit research. Made in the Hellenic Republic (Greece).
Zenodo Publishing is powered by CERN and supported by the European Union**



**Supported by
the European Union**