

- **Title.**

Cost-effective smartphone-based electrochemical instrument for alcohol determination in whole blood samples

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Abstract

The determination of ethanol intoxication in whole blood samples may open the opportunity for a precise and quick point-of-measurement in the ambit of medical emergency or law enforcement. In contrast with traditional techniques based on breath sampling, direct blood measurements present greater immunity to errors specially in case of unconscious or non-collaborative patients. In this context, a portable, sensitive and easy-to-use instrument is highly desirable. In the current work we present a smartphone-based μ Potentiostat which combines a novel circuitual technique for sensor readout digitalization with a reusable lab-on-a-chip (LoC) concept. Such system allows both chronoamperometric and cyclic voltammetry measurements with a reduced number of electronic components on a very compact PCB ($38.5 \times 22.5 \text{ mm}^2$). Power, data-link and user interface are provided in combination with a standard smartphone, enabling cost-effectiveness and reconfigurability without sacrificing precision. The readout platform discussed in this work has been coupled to a LoC for point-of-care combining Pt electrodes microfabricated on silicon substrate for electrochemical measurement and a microfluidic structure of methacrylate for fluid management. Biosensing is enabled by in situ electrodeposition of a calcium alginate hydrogel containing horseradish peroxidase (HPR) and alcohol oxidase (AOx) for selective ethanol detection.

Alginate membrane electrodeposition has been here optimized for rapid generation (2 min) and to retain the cellular fraction, thus allowing the measurement in whole blood samples. The μ Potentiostat features a sensitivity of 36 nA/g L^{-1} to ethanol concentration in blood in the $0\text{--}1.25 \text{ g;L}^{-1}$ range, with a limit of quantification (LoQ) of 4.5 nA , which is a suitable response for discerning the legal, illegal, severely illegal thresholds in a $40 \mu\text{L}$ sample of blood.

Smartphone based electrochemical instrument, Low-cost reusable point-of-measurement, Blood alcohol concentration, Whole blood analysis

1. Introduction

The degree of ethanol intoxication of a person is assessed in a number of circumstances including traffic police checkpoints, emergency departments or emergency quick responses (Kaisdotter Andersson et al., 2015; Sebbane et al., 2012) and is quantified by blood alcohol concentration (BAC). Currently, hand-held breath analyzers (commonly referred to as breathalyzers) are employed either for law enforcement in traffic security or for clinical trial. These devices sample patients' breath alcohol concentration (BrAC) in a rapid and non-invasive way and rely on an established BrAC/BAC ratio (BBR) to finally estimate patients' BAC. Despite the versatility of these devices, their reliability in critical cases such as emergency situations is undermined by: (i) BBR inter-/intra-individual variability, (ii) respiratory impairment or lack of patient collaboration and (iii) presence of residual mouth alcohol. BBR values vary a lot among different countries, depending on the respective legal framework for traffic security and ranging from a 2000 to 2400 (Logan and Osselton, 2011), giving rise to a fervid scientific debate (Hartung et al., 2016; Jones, 2016; Kriikku et al., 2014). Subject will and/or capacity to sustaining a constant breath flow through the instrument is another issue that can only be addressed by the most sophisticated and expensive models of breathalyzers. Finally test results can be false positive due to the presence of alcohol in the mouth, but not absorbed in the bloodstream: use of mouth rinses or even endogenous fermentation in human guts are possible causes of that (Lindberg et al., 2015).

Alternatively, a simple direct BAC measurement on a very small quantity of blood, in the same fashion of common diabetes tests, can overcome all the above mentioned issues. In the era of the maturing Internet of Things, such point-of-care/point-of-measurement instrument should be thought as a low-cost plug-and-play extension to the almost universally diffused smart devices (phones, tablets, convertible laptops). Smartphone-based electrochemical instruments for health monitoring are in vogue among the scientific community in the recent years, promising portable, real-time, cloud-connected and inexpensive measurements capable of basic potentiostatic operation, chronoamperometry (CA), cyclic voltammetry (CV), electrochemical impedance spectroscopy (EIS) (Aggidis et al., 2015; Quesada-González and Merkoçi, 2017; Roda et al., 2016; Zhang and Liu, 2016). Cost is an important parameter of these solutions when penetration in the market is considered in the near future, and it is generally determined by the number of discrete components used to link the transducer electrodes to the computational power of a smartphone: in the current

state of the art, the best cost-effective solution is reported in (Wang et al., 2015) (10\$) while other solutions score above \$20 (Alexander C. Sun, Chengyang Yao, A.G. Venkatesh, 2016; Andres Felipe Diaz Cruz, Nicolas Norena, Ajeet Kaushik, 2014; Carlyn Loncaric, Yiting Tang, Cassie Hob, M. Ash Parameswaran, 2012; Daizong Ji, Lei Liu, Shuang Li, Chen Chen, Yanli Lu, Jiajia Wu, 2017; Eliah Aronoff-Spencer, A.G. Venkatesh, Alex Sun, Howard Brickner, David Looney, 2016; Ioannis Ramfosa, Nikolaos Vassiliadis, Spyridon Blionas, Konstantinos Efstathiou, Alex Fragoso, Ciara K. O'Sullivan, 2013; Lillehoj et al., 2013; Nemiroski et al., 2014; Rowe et al., 2011; Wang et al., 2017; Yan Fan, Juntao Liu, Yang Wang, Jinping Luo, Hui ren Xu, 2017). However the system in (Wang et al., 2015) is based on audio jack connection which, differently from USB connectivity, is not available in most of the last generation smartphones.

Here we present a portable instrument for BAC measurement device employing the recently published transducer structures based on electrodeposited alginate membranes (Márquez et al., 2017) in combination with an original circuit architecture enabling the integration of complex functions over readout signals using very few discrete components. The resulting very low-cost (<10\$) and reusable Lab-On-Chip (LOC) allows BAC assessment in whole blood samples which are emergency point-of-measurements, where rapidity, versatility and simplicity are of paramount importance.

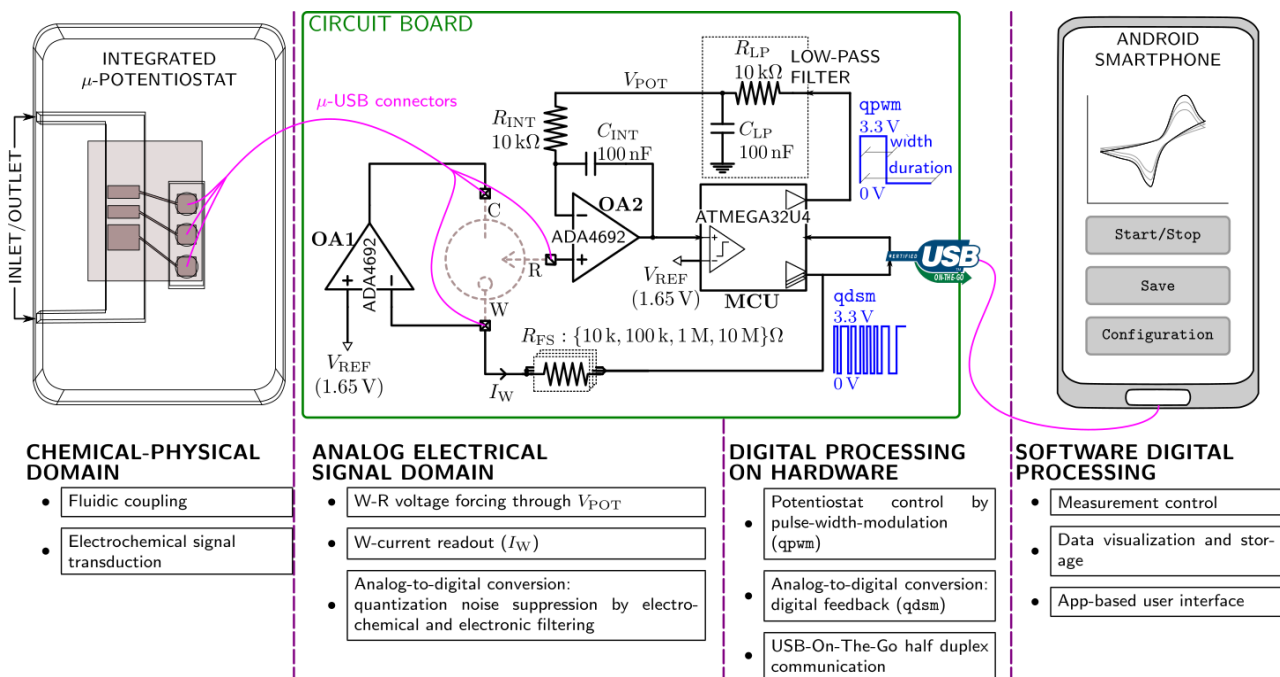


Fig.1. Smartphone based electrochemical instrument for BAC assesment: from sensing to end-user interface.

2. Material and methods

2.1 Architecture of the smartphone-based electrochemical instrument

Figure 1 depicts the system concept and the detailed functions from the electrochemical transduction up to the data capture and visualization on an Android-based smartphone. The extremely compact design of the printed circuit board (PCB) is based on original circuitual topologies which reuses the intrinsic electrochemical time constant and a few electrical components to implement a flexible-yet-high resolution potentiostat (Aymerich et al., 2017; Sutula et al., 2014). Such miniaturized USB-powered PCB is capable to be operate the sensor in potentiostatic, cyclic voltammetry and chronoamperometry mode. In this novel architecture, a number of design techniques combining the electrochemical, analog-to-digital conversion and mixed hardware/software aspects concur to drastically reduce the list of components and thus the cost of the final system.

The working electrode voltage is set to 1.65 V, i.e. at half the available regulated supply available from the USB link, by the local feedback loop built around an operational amplifier (OA1), that also provides the needed current to bias the sensor and to counterbalance the current at working electrode (I_w). This condition is reinforced by using ultra-low input currents operational amplifiers (both OA1 and OA2) which avoids parasitic charge flows, especially at the reference electrode.

Another control loop, namely the delta-sigma loop, is implemented around OA2 and the microcontroller (MCU) providing the analog-to-digital conversion through delta-sigma modulation. A simple way to describe the delta-sigma action is as follows: whenever I_w imbalances the sensor redox current, charge is either accumulated in or depleted from the electrochemical cell. This allows instantaneous small variations of the reference voltage with respect to its DC value, which should be accounted as an instantaneous error. The error is accumulated by the integrator build around OA2 in form of a charge stored in C_{INT} . The voltage output of OA2, holding the information of the error charge, is connected to the MCU comparator input port. Periodically, at a pace provided by the MCU clock, the accumulated error is assessed to be either positive or negative, corresponding respectively to a logical "1" and a logical "0". Consequently, the MCU output digital drivers set the voltage across the R_{FS} either to be negative or positive: this generates an I_w current fed back to the sensor compensating for the accumulated error. At this point, it is important to

notice that the MCU sampling frequency, set to 4 kHz, pushes the error fluctuations far from the sensor intrinsic bandwidth, providing an effective frequency domain decoupling of the control signal with respect to the redox signal. On the other hand, the negative feedback nature of the delta-sigma loop forces the DC component of I_w to follow the redox current generated by the sensor: this process generates an output stream of bit ($qdsm$) that digitally represents the redox current and can readily transmitted through a USB-On-The-Go (USB-OTG) cable to a smartphone for data capture and visualization. The same loop also provide the correct potentiostatic DC voltage since any difference between V_{POT} and V_{RE} is also accumulated into the electronic integrator and then corrected by the delta-sigma loop. Finally, V_{POT} is generated by low-pass filtering a pulse-width modulated digital signal ($qpwm$) also coming from the MCU. Since the low-pass filter extracts the DC value of $qpwm$, by simply modifying the width of the the pulses, V_{POT} can span from 3.3 V and 0 V, which corresponds to a potentiostatic setting point between -1.65V and 1.65 V. Different current full-scales (165 nA to 165 μ A) can be set by parallelizing multiple MCU three-state digital buffers attached to weighted resistors R_{FS} . Both full-scale current and V_{POT} values can be digitally programmed by the MCU accessed by the user trough the intuitive user interface implemented on an Android device.

2.2 Electronic components and software programming

Surface Mounted Devices (SMD) 0805 resistors/capacitors, SMD single chip dual operational amplifiers ADA4692-2, ATmega32U4 AVR MCU solded on a standard double layer PCB (size: 38.5 mm \times 22.5 mm). The micro USB-OTG cable has been obtained from a standard USB cable for smartphones by grounding the fifth pin in the male micro-USB connector.

The Arduino Leonardo firmware has been flashed onto the MCU by using a standard Arduino board as in-system program (ISP), allowing to handle USB connectivity and programming without any external hardware programmer. This allowed to use the user friendly and very popular Arduino development environment.

A very simple android application has been developed in Android Studio 2.3.1. which allows for opening the serial port connection trough the USB cable. The application has two operation modes: CA and CV. In the former mode VPOT is set, while the latter mode is defined by the scan rate and the peak-to-peak VPOT parameters. Full-scale current is chosen for both operations by a drop-down menu to be set between the following values: 165 nA, 1.65 μ A, 16.5 μ A, 165 μ A. The

user starts data acquisition which is recollected in the smartphone memory and visualized by a minimal plot interface.

2.3 Chemicals and reagents

Alginic acid sodium salt, CaCl_2 ($\geq 93\%$), 2-(N-morpholino)ethanesulfonic acid hydrate (MES hydrate, $\geq 99\%$), NaOH ($\geq 98\%$), KOH ($\geq 86\%$), 3,3',5,5'-Tetramethylbenzidine (TMB, $\geq 98\%$), NaCl (ACS reagent, $\geq 99\%$), H_2O_2 (30% in H_2O), HRP (Type VI-A, essentially salt-free lyophilized powder, 250-330 units mg^{-1}) and AOx from *Pichia pastoris* (buffered aqueous solution, 1200 units ml^{-1}) were purchased from Sigma-Aldrich. CaCO_3 ($\geq 98.5\%$), $\text{K}_3[\text{Fe}(\text{CN})_6]$ (ACS reagent, $\geq 99\%$), dimethyl sulphoxide (DMSO, 99.5%), Na_2HPO_4 (ACS reagent, $\geq 99\%$) and ethanol absolute ($\geq 99.5\%$) were purchased from Panreac. KCl ($\geq 99\%$) and KH_2PO_4 (ACS reagent, $\geq 99.5\%$) were purchased from Fluka.

All chemicals were used as received and aqueous solutions were prepared using de-ionized water with a resistivity of $2\text{ M } \Omega\text{ cm}$. Used PBS contains NaCl (8 g L^{-1}), KCl (0.2 g L^{-1}), Na_2HPO_4 (1.42 g L^{-1}) and KH_2PO_4 (0.24 g L^{-1}). TMB 50 mM stock solution was daily prepared using DMSO as solvent. Whole blood was extracted from a volunteer and stored at 4°C until measurement.

2.5 Equipment

For comparison with the fabricated $\mu\text{Potentiostat}$, a $\mu\text{Autolab}$ potentiostat Type-III controlled with Nova 2.0 software was used. The pH of the used buffer was established using a Crison pH & ION-Meter GLP 22+.

2.6 Electrochemical transducer

For the electrochemical part of the biosensor, a three Pt electrodes linear array was used as reference (RE), working (WE) and counter (CE) electrodes respectively. The electrodes were fabricated on a silicon substrate at the CNM-IMB clean room, according to a previously described photolithographic process (Orozco et al., 2007).

The presented electrodes do not precise of electrical bonding as they are linked to adjacent areas of the same material, later coupled by spring loaded connectors.

2.7 Microfluidic LOC

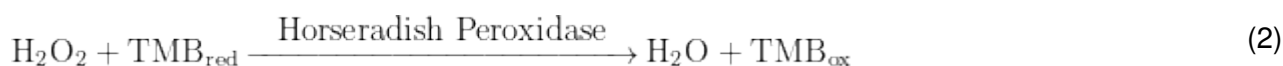
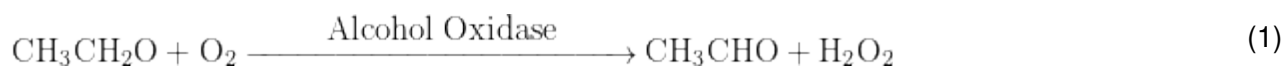
A PMMA-PDMS structure was designed and fabricated in accordance with the silicon substrate. The PMMA and PDMS pieces were designed using CorelDRAW V7 software and then cut with an Epilog Mini 24 CO₂ laser ablation machine. The LOC structure was designed and constructed layer by layer in three blocks: i) a base for the Si substrate positioning, ii) the channel-chamber part and iii) a cover to seal the chamber. The different PMMA pieces were joined using a pressure-sensitive adhesive (PSA) for the definitive unions while the PDMS layers were used between the different PMMA blocks, in order to allow the disassembly of the whole device. Four screws in the corners of the device prevent any leakage between the PMMA and PDMS layers. The inlet and outlet of the channel were glued to a conventional fluidic connector using epoxy resin. Prior to use, the electrodes were cleaned by immersion during 10 min in an aqueous solution containing 0.05 M KOH and 25 % w/v H₂O₂.

2.8 Electrochemical characterization

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2.9 Alginate-based biosensor

As the recognition part of the biosensor, a reversible electrodepositable calcium alginate hydrogel was used for the enzymes retention, following a previously reported protocol (Márquez et al., 2017). The measure cycle consists in three main steps: hydrogel electrodeposition, amperometric detection and hydrogel removal. Briefly, for the electrodeposition, an alginic acid sodium salt was dissolved in warm water with magnetic stirring and then mixed with CaCO₃ and the enzymes. Subsequently, it was injected into the reaction chamber of the device through the channel. In presence of CaCO₃ particles, the application of an anodizing potential (1.5 V vs. RE) promotes water oxidation into O₂ and protons. These protons react with the CaCO₃ particles, promoting their dissociation into Ca²⁺ cations and HCO₃⁻. Finally, the Ca²⁺ cations complete the cross-link reaction among the alginic acid monomers and a gel is formed, attached to the anodized electrode (WE). After the electrodeposition, the chamber is washed with deionized water to remove the excess of precursor and leave only the recently built hydrogel. For the measurement, the sample was introduced in the chamber and a constant potential of 0 V (vs. Pt RE) was applied between the WE and the RE, while the current was measured between the WE and the CE. The registered current is related to the initial amount of ethanol in the medium due to the following cascade reaction:



Where the ethanol is first oxidized by the AOx into acetaldehyde and hydrogen peroxide (1) and this is used by the HRP to oxidize the TMB (2) which is present as a mediator. Finally the oxidized TMB is reduced at the WE at the applied potential. Once the measurement is finished, the chamber and the channel were cleaned using PBS. The phosphate acts as a chelating agent, trapping Ca^{2+} and disaggregating the gel rapidly. The process can then start from the beginning in a second analysis using the same device and only regenerating the membrane.

3. Results

3.1 Electrical tests

Since the proposed μ Potentiostat follows the idea of (Sutula et al., 2014), where the intrinsic sensor time constant participates in the readout process of the transducer current, electrical characterization of the sensor, in the sense of Randles model, is fundamental. To this purpose experimental impedance measurements of the structures of Section 2.6, have been performed using Autolab PGSTAT302N potentiostat coupled with Eco Chemie FRA32M impedance analysis module. For the circuit model in Fig 2(a), the following electrical parameters have been extracted:

$R_S = 52 \pm 2$ ohm, $C_{DL} = 1.392 \pm 0.002$ μ F, $R_{CT} = 58 \pm 2$ kohm, $W = 3 \pm 2$ kohm.

Starting from these values a discrete component RC network, first order matched to the Randles model, has been assembled in order to emulate the sensor behavior and test the μ Potentiostat correct operation ($R_S = 330$ ohm, $C_{DL} = 1.5$ μ F, $R_{CT} = 56$ kohm, $W = 0$). In this first approach the Warburg element has been neglected since its effects are evident only at extremely low frequencies. With such dummy sensor a zero input signal condition can be easily implemented at electrical level which is useful to estimate the instrumentation noise floor. Data acquisition of the *qdsm* signal has been performed in this condition and the resulting power spectrum is shown in Fig. 2(b). As expected, two regions are clearly visible: thermal flat noise dominates up to approximately 80 Hz, while for the higher frequencies the noise profile is dominated by the typical shaped quantization noise due to the delta-sigma modulation. The sensor intrinsic bandwidth can be roughly estimated as $1/(2 \cdot \pi \cdot R_{CT} \cdot C_{DL})$ approximately 1.9 Hz. Considering an observation time of 10 seconds, and a bandwidth of 2 Hz, the residual integrated noise is less than 1 nA root-mean-squared (RMS). It is worth noting that the part of the spectrum exceeding the sensor bandwidth will be numerically filtered out in the software algorithms implemented in the smartphone application.

To further characterize the μ Potentiostat, the sensing structures described in section 2.6 have been submerged in KCl saline solution and have been connected to the PCB to perform chronoamperometric readouts which allowed to determine the limit of detection (LoD) and limit of quantification (LoQ) of 1.35 nA and 4.50 nA, respectively. (AUTOLAB = 2.8 nA; 9.4 nA) Systematic current offset due to deviation of the reference voltage from the nominal value of 1.65 V was estimated to be 3.84 nA.

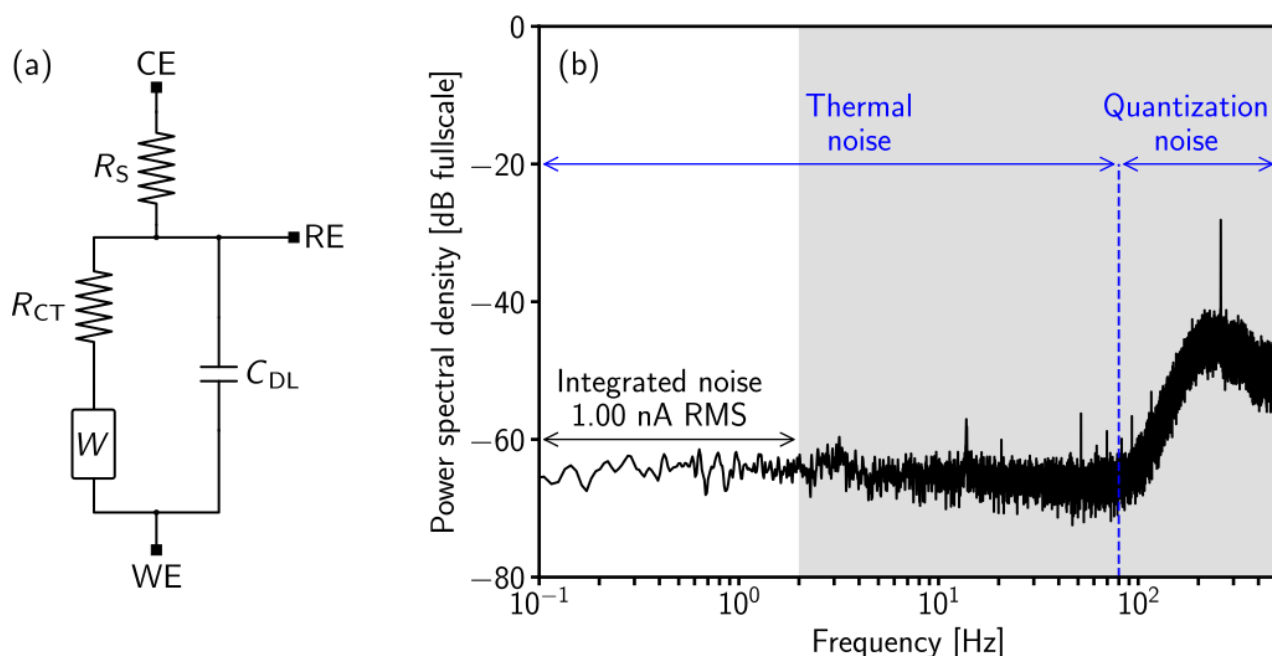


Fig.2. Simplified Randles electrical equivalent circuit (a) and measured electronic readout noise (b).

3.2 Electrochemical test

The μ Potentiostat has been characterized through cyclic voltammetry experiments of the $\text{Fe}^{\text{III}}(\text{CN})_6^{3-}/\text{Fe}^{\text{II}}(\text{CN})_6^{4-}$ couple. Figure 3 shows the results for a 1 mM Ferri-Ferro in 150 mM KCl at different scan rates compared to the benchtop instrument. For this experiment the DAC resistor was fixed to be 330 kohm, in order to provide a nominal current full-scale of 5 μA (both positive and negative). Data was extracted recording the digital signal *qdsm* sampled at 2.5 kS/s. At this point, since *qdsm* contains the high frequency quantization noise, filtering is needed to correctly reconstruct the WE current waveform: ideally an analog bessel filter should be employed to preserve the group delay of the signal, in practice a digital equivalent derived from the analog model through a bilinear transform is sufficient since the bandwidth of interest is much lower than the sampling frequency, this way the high frequency noise evident in Fig.2 is largely suppressed. The tuning parameter of the filter must be adjusted to accordingly since the bandwidth of interests scale proportionally to the scan-rate. It is worth noting that an excessive filtering should be carefully chosen in order to avoid filtering frequency component of the signal of interest. For the third-order digital bessel filter used here, the critical frequency at phase response midpoint has been set to 0.67 Hz, 8.33 Hz, 20.83 Hz for the scan-rates of 2 mV/s, 10 mV/s and 25 mV/s, respectively. The comparison between the μ Potentiostat versus the bench-top cyclic voltammetries are evident in Fig.3(a) and (b), where it can be seen that some high frequency noise is still visible in out system.

A more smooth characteristic can be obtained, without bandwidth impairment, by averaging several cycles.

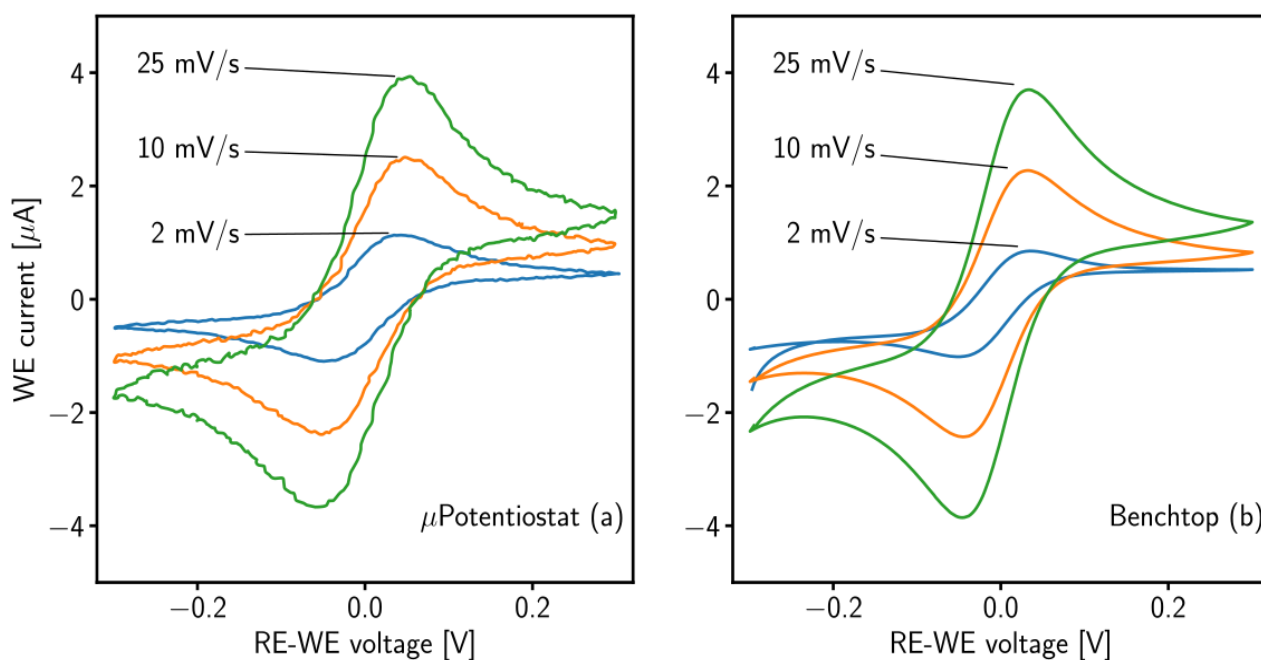


Fig.3. Cyclic voltammeteries of 1 mM Ferri-Ferro in 150 mM KCl at different scan rates: (a) $\mu\text{Potentiostat}$ and (b) bench-top setup described in Section 2.5.

3.3 Alcohol biosensor characterization

The biosensor was optimized in order to obtain a fast response time (2 min after analyte addition), a good sensitivity ($-54 \text{ nA} / \text{g L}^{-1}$) and an accurate linear range for measure (0 to 1 g L^{-1}). For that purpose, the amounts in the precursor of alginic acid (1% w/v), CaCO_3 (0.5 % w/v) and HRP (25 units ml^{-1}) were fixed while the amount of AOX was changed from 0.1 to 0.6 units ml^{-1} , reaching the saturation response of the biosensor in buffer at 0.4 units ml^{-1} [Fig_Optimization] for the maximum ethanol concentration (1 g L^{-1}).

Maintaining these conditions for the membrane electrodeposition, the biosensor was calibrated using buffer solutions with different ethanol concentrations, from 0.25 to 1 g L^{-1} . The buffer solution contained 0.2 M MES-NaOH pH 7, 0.145 M KCl, 0.01 M CaCl_2 and $1.5 \cdot 10^{-3}$ M TMB.

The different solutions were injected in the cell through the channel and then a chronoamperometric measurement was started applying a constant potential of 0 V vs Pt. (RE) [Fig_Calibration_buffer].

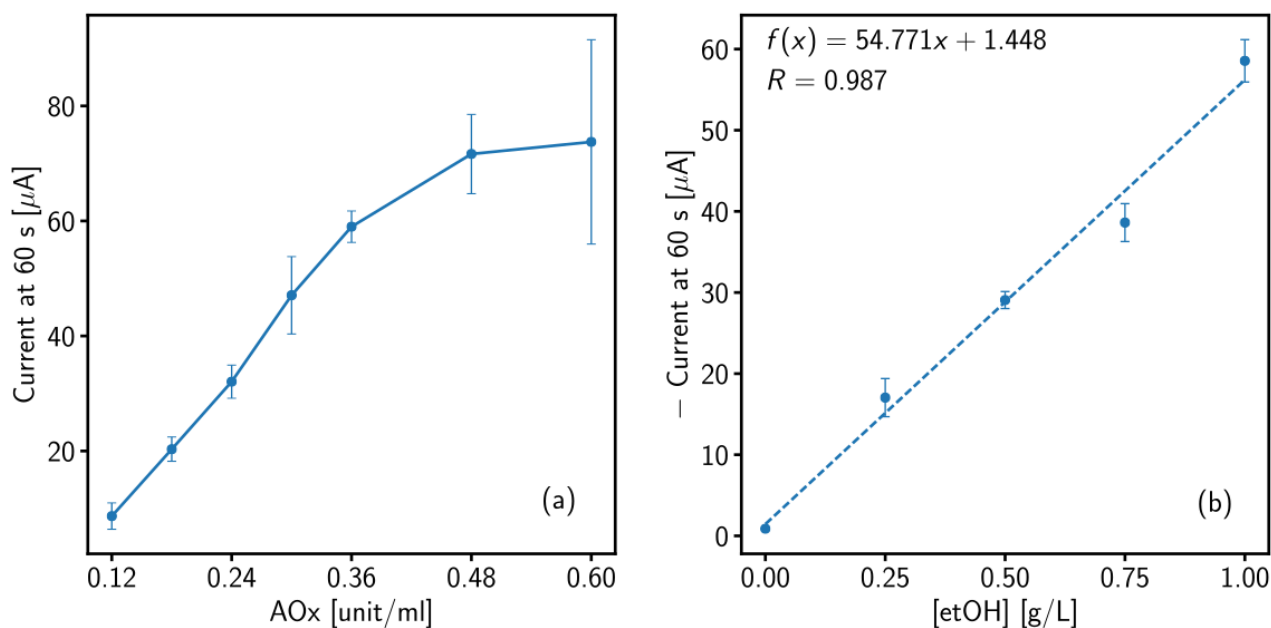


Fig.4. [Fig_Optimization, Fig_Calibration_buffer].

3.4 Alcohol determination in whole blood samples

In the case of real sample, 40 μL of sample were mixed with 2 μL of 50 mM TMB and 8 μL of aqueous ethanol solutions at different concentrations yielding to different final ethanol concentrations. In this case, the sample is not injected through the channel because of its small volume. Therefore, the cover piece is modified with a cavity situated over the WE that can be open for the blood addition. Due to diffusion limitations, 2 minutes were established as incubation time prior to measurement to ensure the stability of the recorded signal. The use of alginate in case of complex samples analysis such as whole blood is essential in order to protect the WE from biofouling and obtain a good response as it has been reported previously (Márquez et al., 2017). The resulting calibration curve [Fig_Calibration_buffer] shows a good correlation between the registered current and the amount of ethanol added to the blood samples, although there is a loss of sensitivity compared to buffer calibrations probably due to matrix effects.

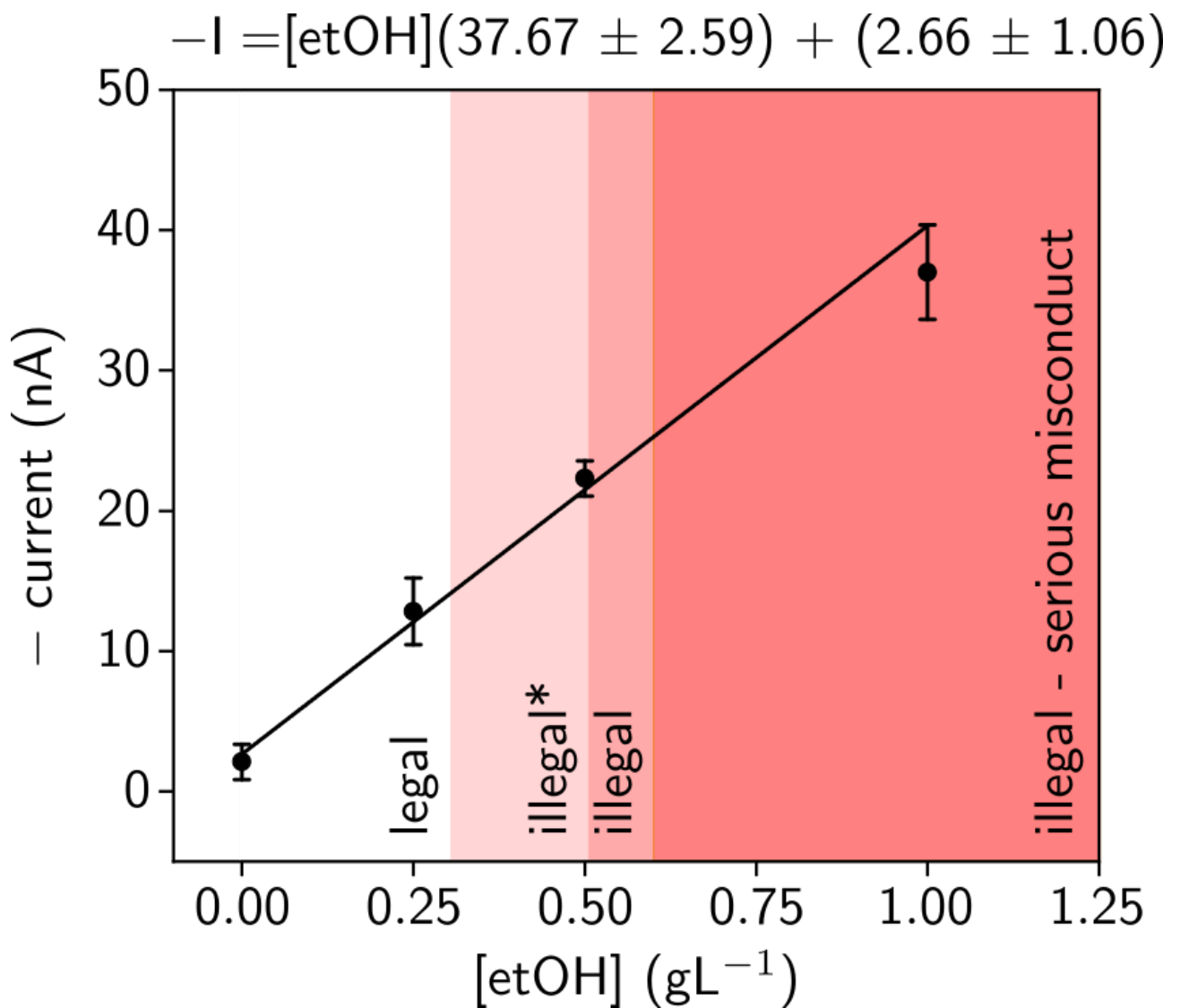


Fig.5. Whole blood measurement spiked with etOH.

4. Conclusions

In conclusion, blood alcohol rate determination in whole blood samples is possible with the portable, miniaturized and lightweight smartphone-based LoC presented in the current study. The system reports on glucose and hydrogen peroxide concentrations, conductivity and ORP as a way to monitor cell metabolism. In addition to the latter, electrochemical analysis is completed with high-resolution imaging by confocal microscopy after labelling with fluorescent dyes. With MRC-5 cells, induction of oxidative stress by hydrogen peroxide produces a dose-dependent reduction of glucose consumption and a proportional increase of DHE emission. ORP is also affected by the reduction in cell metabolism but changes in conductivity due to cell lysis are not observed. For simplicity, integration, automation, compartmentalisation and micro-fluidic control, we envisage the current technology as a promising alternative to traditional methods for drug development in the near future.

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