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Monograph

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Systematic revision of the parasitoid wasp genus *Glyptapanteles* Ashmead (Hymenoptera: Braconidae: Microgastrinae) for Australia results in a ten-fold increase in species

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Abstract. Despite several decades of active research, there are still substantial gaps in the knowledge of parasitoid wasps in Australia, with many families and genera yet to be revised using modern approaches and only a fraction of the estimated fauna currently described. The genus *Glyptapanteles* Ashmead, 1904 is a member of the subfamily Microgastrinae (Hymenoptera: Braconidae) and all species in the subfamily are lepidopteran parasitoids. The genus previously contained only three species known from Australia: *G. deliasa* Austin & Dangerfield, 1992, *G. drioplanetus* Fagan-Jeffries & Austin, 2021 and *G. mnesampela* Austin, 2000. To undertake a revision of this morphologically-conserved group in Australia, we used a combination of molecular (cytochrome oxidase subunit one (*COI*) and *wingless* genes) and minimal morphological data to delimit and describe an additional 31 species: *G. austini* Fagan-Jeffries & Bird sp. nov. and the following 30 species all authored by Fagan-Jeffries, Bird & Austin: *G. albigena* sp. nov., *G. andamookaensis* sp. nov., *G. arcanus* sp. nov., *G. aspersus* sp. nov., *G. austrinus* sp. nov., *G. baylessi* sp. nov., *G. bradfordae* sp. nov., *G. cooperi* sp. nov., *G. doreyi* sp. nov., *G. dowtoni* sp. nov., *G. eburneus* sp. nov., *G. erucadesolator* sp. nov., *G. ferrugineus* sp. nov., *G. foraminous* sp. nov., *G. goodwinnoakes* sp. nov., *G. guzikae* sp. nov., *G. harveyi* sp. nov., *G. kingae*

sp. nov., *G. kittelae* sp. nov., *G. kurandaensis* sp. nov., *G. lambkinae* sp. nov., *G. lessardi* sp. nov., *G. mouldsi* sp. nov., *G. niveus* sp. nov., *G. rixi* sp. nov., *G. rodriguezae* sp. nov., *G. ruhri* sp. nov., *G. sanniopulus* sp. nov., *G. vergrandiacus* sp. nov. and *G. wrightae* sp. nov. We provide a key to species groups and to the species able to be identified on morphological characters alone. Additionally, we provide a brief discussion of the difficulties in describing small, morphologically conserved wasps and the challenges associated with revising the taxonomy of hyperdiverse taxa in the context of the planned mission of Taxonomy Australia to accelerate the documentation of Australia's biodiversity.

Keywords. Microgastrine, species delimitation, DNA barcoding, Australasia.

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Introduction

Accelerating the documentation of Australia's biodiversity, in line with the decadal plan for Taxonomy in Australia and New Zealand (Taxonomy Decadal Plan Working Group 2018), requires the rapid description of new species of hyper-diverse groups of organisms. One such group are the parasitoid wasps, of which only a fraction are formally named (Yeates *et al.* 2003). Parasitoid wasps are important components of nearly all ecosystems, yet only fragmentary work has been completed on the systematics of most families in Australia. The Microgastrinae Foerster, 1962, a subfamily of the Braconidae Latreille, 1829, are thought to number in the tens of thousands of species (Rodriguez *et al.* 2013), with approximately 3000 species in 81 genera currently described worldwide (Fernández-Triana *et al.* 2020). In Australia, 143 species in 22 genera (Fagan-Jeffries & Austin 2020; Fernández-Triana *et al.* 2020) are currently recognised, with 280 Barcode Index Numbers (BINs – operational taxonomic units which closely correspond to species) (Ratnasingham & Hebert 2013) present on the Barcode of Life Database (BOLD) as of 24 July 2021. All microgastrines are parasitoids of lepidopteran larvae, are important in both regulating native caterpillar populations and as current and potential biological control agents of agricultural pests.

The genus *Glyptapanteles* Ashmead, 1904 has only three species described from Australia: *G. deliasa* Austin & Dangerfield, 1992, *G. drioplanetus* Fagan-Jeffries & Austin, 2021 and *G. mnesampela* Austin, 2000. Worldwide, there are 307 species described of an estimated 3000+, with the genus being found in all biogeographical regions (Fernández-Triana *et al.* 2020). The limits of the genus are controversial, with close morphological similarities to *Cotesia* Cameron, 1891, *Distatrix* Mason, 1981, *Lathrapanteles* Williams, 1985, *Protapanteles* Ashmead, 1898 and *Sathon* Mason, 1981 (Arias-Penna *et al.* 2019). In Australia, the genus, as currently recognised, is clearly paraphyletic with respect to *Cotesia* based on sequence data from cytochrome c oxidase subunit 1 (*COI*) and *wingless* genes (Fagan-Jeffries *et al.* 2018; Fagan-Jeffries & Austin 2020). Despite the likelihood that the genus is not monophyletic, *Glyptapanteles* can normally be separated from *Cotesia* using morphology. *Cotesia* has the first metasomal tergite parallel-sided and the second metasomal tergite normally broad and rectangular, only ever slightly triangular, whereas *Glyptapanteles* often has a strongly narrowing first tergite and strongly triangular second tergite.

Of the other genera mentioned above, *Distatrix* has the median area on T2 less well-defined than in *Glyptapanteles* and has not been recorded from the Australasian region; *Lathrapanteles* is not a natural grouping and is not yet recorded from Australia and we see no reason to place more species here until a better classification of the subfamily is established; and *Protapanteles* is not yet recorded from Australia

and has the first metasomal tergite parallel-sided except for a strongly rounded apex (Arias-Penna *et al.* 2019). Whilst *Sathon* in other parts of the world has the fore wing areolet absent (like in *Glyptapanteles*), in Australia all species currently placed in *Sathon* have a large closed areolet and are most likely unrelated to the northern hemisphere species (Austin & Dangerfield 1992; Fagan-Jeffries *et al.* 2019). As such, *Glyptapanteles* in Australia is a relatively easily recognised group of species despite its paraphyletic nature and we feel it is untenable to split the genus into smaller units until a more thorough global phylogenetic study is conducted. Current ongoing projects utilising ultraconserved element bait sets and exon capture next-generation sequencing techniques will hopefully provide better resolution on the relationships and limits of microgastrine genera in the foreseeable future.

Using *COI* and *wingless* DNA barcoding data and key morphological characters, we here describe 31 new species of *Glyptapanteles* from Australia and provide a dichotomous key to species. All new species are authored by Fagan-Jeffries, Bird & Austin other than *Glyptapanteles austini*, which is authored by Fagan-Jeffries & Bird. Each new species description includes DNA barcode information, high-resolution images and a diagnosis against other *Glyptapanteles* described from Australia. Members of the genus from the broader Australasian region (Fiji, Samoa, New Zealand and Papua New Guinea) are listed, illustrated and diagnosed against the newly described Australian species where possible. We focus here on providing descriptions that are useful for end-user identification purposes and are as efficient as possible within the confines of what is generally accepted by the hymenopteran taxonomic community in terms of minimalist descriptions. As morphology is often not particularly useful for species delimitation in this genus, as it is not in many groups of Microgastrinae, we provide morphological diagnoses where possible, but also rely on *COI* and *wingless* sequences to delimit and diagnose species.

This paper includes important material from the ‘Bush Blitz’ surveys of regional Australia, a multi-institutional partnership coordinated by the Federal Government’s Australian Biological Resources Study (ABRS). The ‘Bush Blitz’ program brings together taxonomists, traditional landowners, property managers and park rangers to intensely survey the flora and fauna of reserves across the continent and describe the new species discovered during the process.

Material and methods

As the aim of the Taxonomy Australia decadal plan is to increase the number of species described in Australia in the most efficient and robust way possible, we use molecular information to inform the species prioritised for description in this study. From a large dataset of both published and unpublished *COI* and *wingless* sequences of Australian microgastrines, the *Glyptapanteles* sequences were extracted using a combination of BOLD identifications and the morphological examination of specimens. The previously unpublished DNA sequences used in this study were generated following the high-throughput methods outlined in Fagan-Jeffries *et al.* (2018) and are now uploaded to BOLD.

Placing the Australian *Glyptapanteles* in the global context

All of the available *COI* sequences of specimens identified as *Glyptapanteles* on BOLD as of 5 June 2021 (n = 5289) were aligned using MAFFT ver. 7.308 (Katoh *et al.* 2002; Katoh & Standley 2013). A Maximum Likelihood (ML) analysis was conducted on all of the unique sequences (n = 2558) using IQTREE ver. 1.6.12 (Nguyen *et al.* 2015) with models and partitions selected using the inbuilt ModelFinder (Kalyaanamoorthy *et al.* 2017) with the –TESTMERGE function, with a specimen of *Microplitis demolitor* Wilkinson, 1934 as the outgroup. Ultrafast bootstraps (1000 replicates) (Hoang *et al.* 2018) and SH-aLRT support (1000 replicates) were calculated for the tree. Full sequence details and the alignment are available as supplementary material ([Supp. file 1](#), [Supp. file 2](#)).

Phylogenetics of the Australian *Glyptapanteles*

BOLD was used to collate a dataset of all *Glyptapanteles* from Australia, Fiji and Papua New Guinea (as of the 9 June 2021), with sequences of *Cotesia* from Australia included for the contextual placement of this genus. A concatenated *COI* + *wingless* ML phylogeny of 321 specimens was constructed using IQTREE ver. 1.6.12 (Nguyen *et al.* 2015) with models and partitions selected using the inbuilt ModelFinder (Kalyaanamoorthy *et al.* 2017) with the –TESTMERGE function, with a specimen of *Microplitis demolitor* as the outgroup. Ultrafast bootstraps (1000 replicates) (Hoang *et al.* 2018) and SH-aLRT support (1000 replicates) were calculated for the tree. Full sequence details and the alignment are available as supplementary material ([Supp. file 3](#), [Supp. file 4](#)).

Species delimitation

An integrative approach to species delimitation using a general lineage species concept (de Queiroz 1998) was employed, with species being delimited based on a combination of their *COI* pairwise difference, *wingless* haplotype and distinct morphological characters. All clades with greater than a *COI* 2% pairwise difference (calculated using the percent identity function on the alignment in Geneious ver. 9.1.8) from each other were examined morphologically and most were recognised as new species. Some provisional species were not described, often because they contained only male specimens. Some described species are represented by single specimens and/or contain only damaged or incomplete specimens, but we feel that this approach optimises the number of valid species recognised as part of Australia's named biodiversity and the available molecular data will allow additional specimens to be easily assigned to these species in the future.

We also treat the described species of *Glyptapanteles* from Australia and species from the broader Australasian region, diagnosing them against the new species where possible. Without molecular data for these species to guide the delimitations, diagnosing some of these species was extremely difficult. We feel that with the mission to describe Australia's biodiversity within 25 years and considering the fact that only a very small proportion of the microgastrine species have been described without molecular data, it is better to describe new species even when they cannot be conclusively separated from historically described species of the region using morphology. We only decide to do this where there is a very low probability that species are synonymous (based on knowledge of the general overlap of species distribution between countries or other information presented under each species), acknowledging that it is possible there may be some synonymy required in the future when improved technologies allow the non-destructive DNA extraction from old type material to be more cost-effective and guaranteed of success.

Morphology

Terms for general morphology and wing venation follow Fagan-Jeffries & Austin (2020) and terms for sculpture follow Eady (1968). Nearly all images of specimens and morphological characters were generated using a Visionary Digital BK + imaging system with a Canon EOS 7D 18 megapixel camera, compiled in Zerene Stacker (Zerene Systems LLC), PMax software and edited for clarity in Adobe Photoshop 2021 (Adobe Systems Inc., San Jose, CA, USA).

Measurements of morphological characters were completed using an ocular micrometre on a Leica Mz16 microscope under a 10× or 100× magnification. Characters which we consider particularly important for diagnoses but are prone to variability within a species (colouration, shape of tergites, number of pits in the scutellar sulcus, ocelli measurements, flagellomere length/width ratios, ovipositor length and sculpture – including of the propodeum, anteromesoscutum and tergites) were coded for every available female specimen. Characters that are traditionally included in hymenopteran descriptions but that we consider very prone to inaccurate measurement and therefore not particularly useful for diagnoses between species in the context of the Australian fauna, we measured only on the holotypes (wing vein

ratios, tergite length/width ratios) and note that these characters, unless extremely divergent, should not be used as diagnostic characters in the future for these species.

Abbreviations

The following abbreviations are used throughout the text:

ACT	=	Australian Capital Territory
bp	=	base pairs
NSW	=	New South Wales
NT	=	Northern Territory
OOL	=	ocular-ocellar line
POL	=	posterior interocellar line
POD	=	posterior ocellus diameter (largest measurement if ocellus not round)
QLD	=	Queensland
TAS	=	Tasmania
T1–6	=	first–sixth mediotergites
SA	=	South Australia
S1–6	=	first–sixth sternites
VIC	=	Victoria
WA	=	Western Australia

Institutional abbreviations

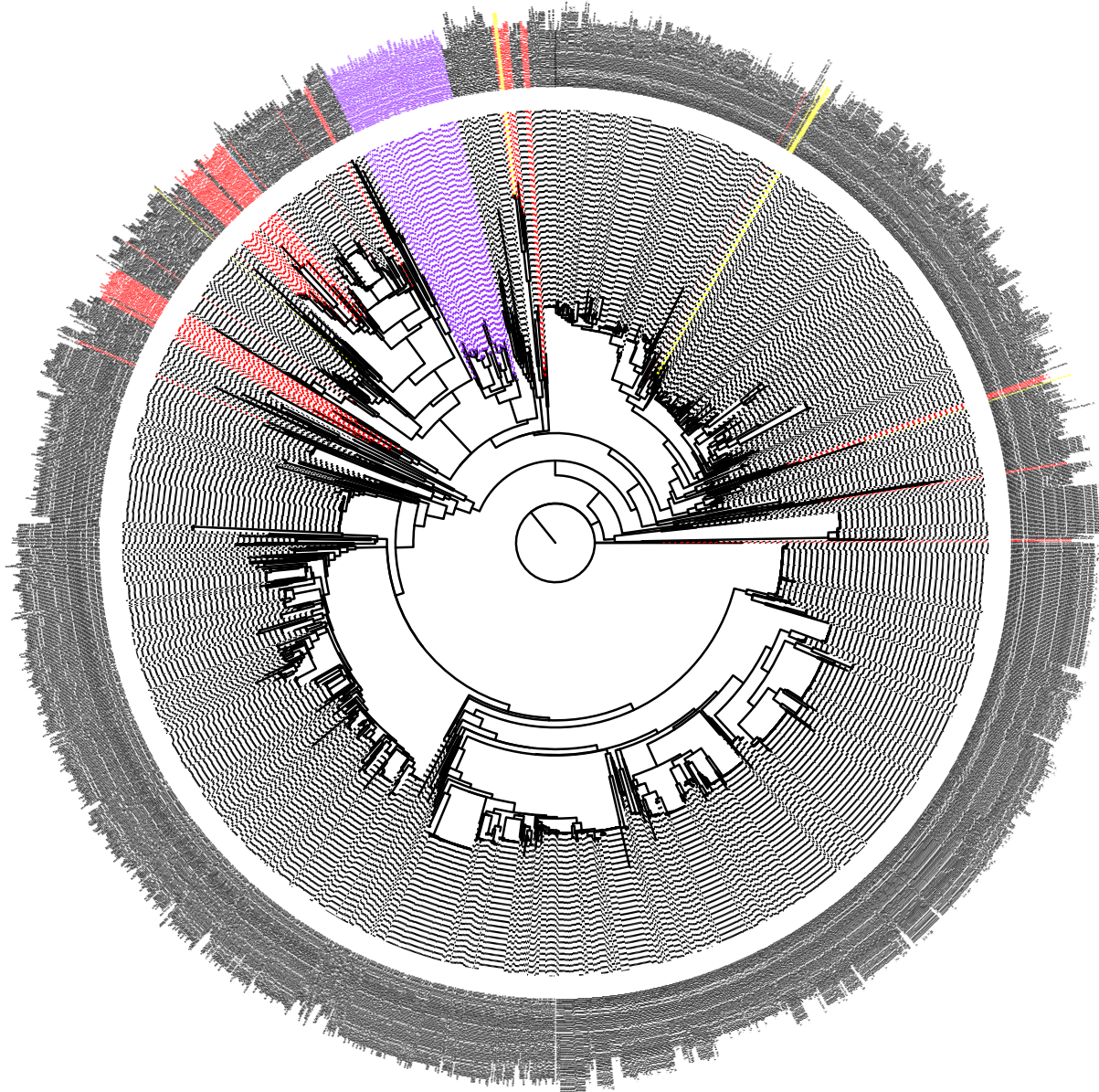
AM	=	Australian Museum, Sydney, Australia
ANIC	=	Australian National Insect Collection, Canberra, Australia
BPBM	=	Bernice P. Bishop Museum, Honolulu, Hawaii, USA
NHMUK	=	Natural History Museum, London, UK
QM	=	Queensland Museum, Brisbane, Australia
SAMA	=	South Australian Museum, Adelaide, Australia
TMAG	=	Tasmanian Museum and Art Gallery, Hobart, Australia
WAM	=	Western Australian Museum, Perth, Australia

Results

Molecular results

A *COI* phylogeny of all sequences identified as *Glyptapanteles* on BOLD suggests that the Australian fauna are not a monophyletic group (Fig. 1). There are several clades that appear to be endemic to Australia or to Australia and Papua New Guinea, whilst there are also several species that are more closely related to specimens from Asia or South America than they are to other Australian lineages. Whilst *COI* alone does not necessarily provide reliable measures of relationships at these deeper levels, this preliminary phylogeny also suggests that the New Zealand fauna is a monophyletic group and not closely related to any sequenced Australian species (Fig. 1). The analysis also confirms that none of the Australian lineages described in this study are identical to previously sequenced specimens found in other regions, decreasing the possibility that the species described in this study overlap with taxa described from elsewhere. There is one Australian sequence sourced from BOLD (ASQAS186-11) that has the collection locality listed only as ‘Australia’, that is identical to sequences from Sweden, but we did not have access to this specimen and did not describe this species.

A *COI* + *wingless* ML analysis of the *Glyptapanteles* available on BOLD from Australia, Papua New Guinea and Fiji indicates there are more than 25 lineages yet to be described (Fig. 2) and highlights the paraphyletic nature of *Glyptapanteles* (as defined in this study for the Australian fauna) in relation to *Cotesia*.



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0.2

Fig. 1. COI ML phylogeny of all sequences identified as *Glyptapanteles* Ashmead, 1904 on BOLD as of 5 June 2021. Australian taxa are highlighted in red, New Zealand taxa in purple, Papua New Guinean taxa in yellow and taxa from Fiji in blue (only one lineage, difficult to discern in circular tree – see expanded phylogeny in [Supp. file 5](#)). Lineages in black are from elsewhere in the world.

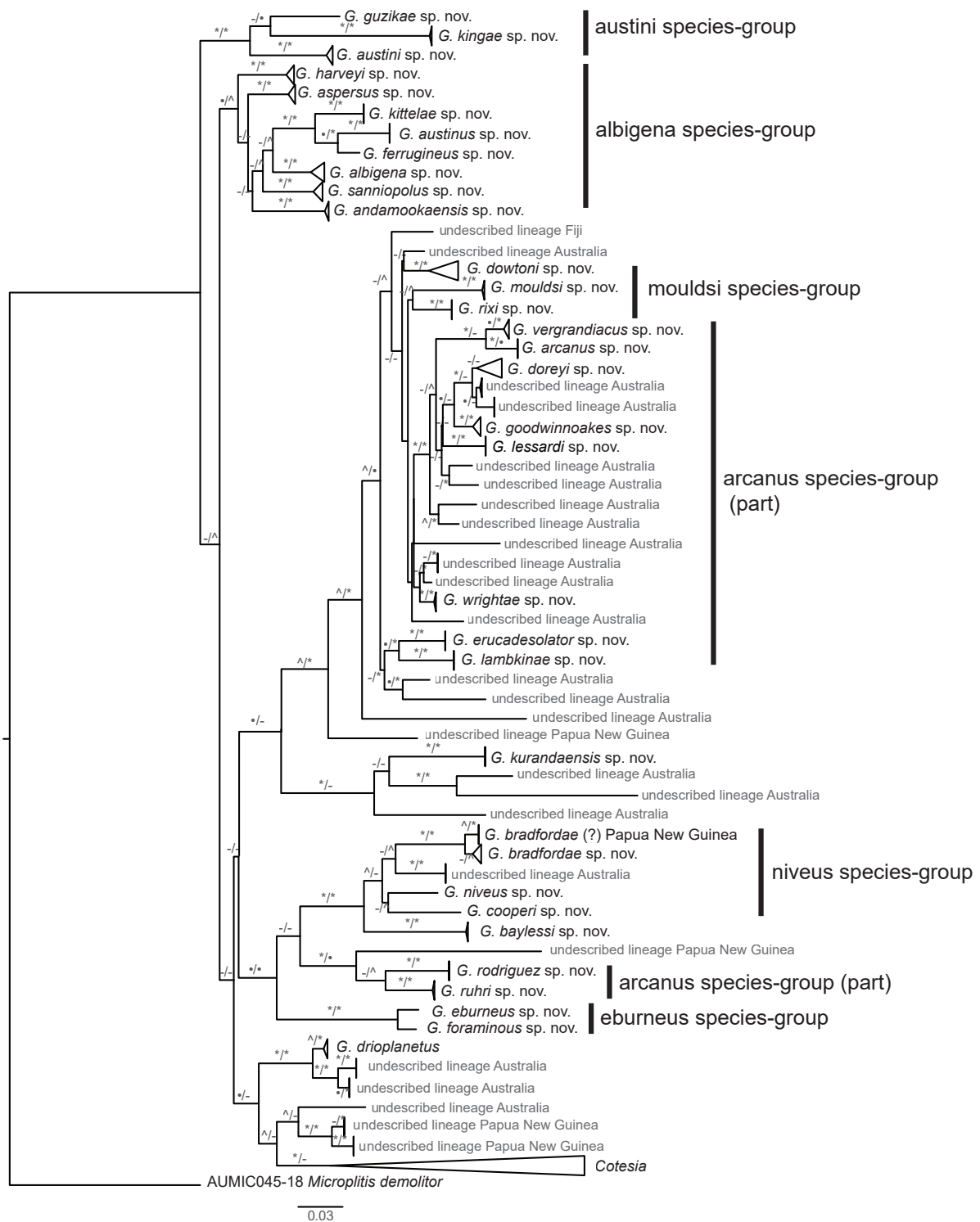


Fig. 2. Maximum likelihood phylogeny constructed using IQ-TREE ver. 1.6.12 of a concatenated *COI* and *wingless* alignment including *Glyptapanteles* Ashmead, 1904 from Australia, Papua New Guinea and Fiji, with specimens of *Cotesia* Cameron, 1891 from Australia included for contextual placement of the genus. Branch support values are given as SH-aLRT support (%) / ultrafast bootstrap support (%), with symbols representing value ranges as follows: * = 96–100; • = 91–95; ^ = 85–90; - = < 85.

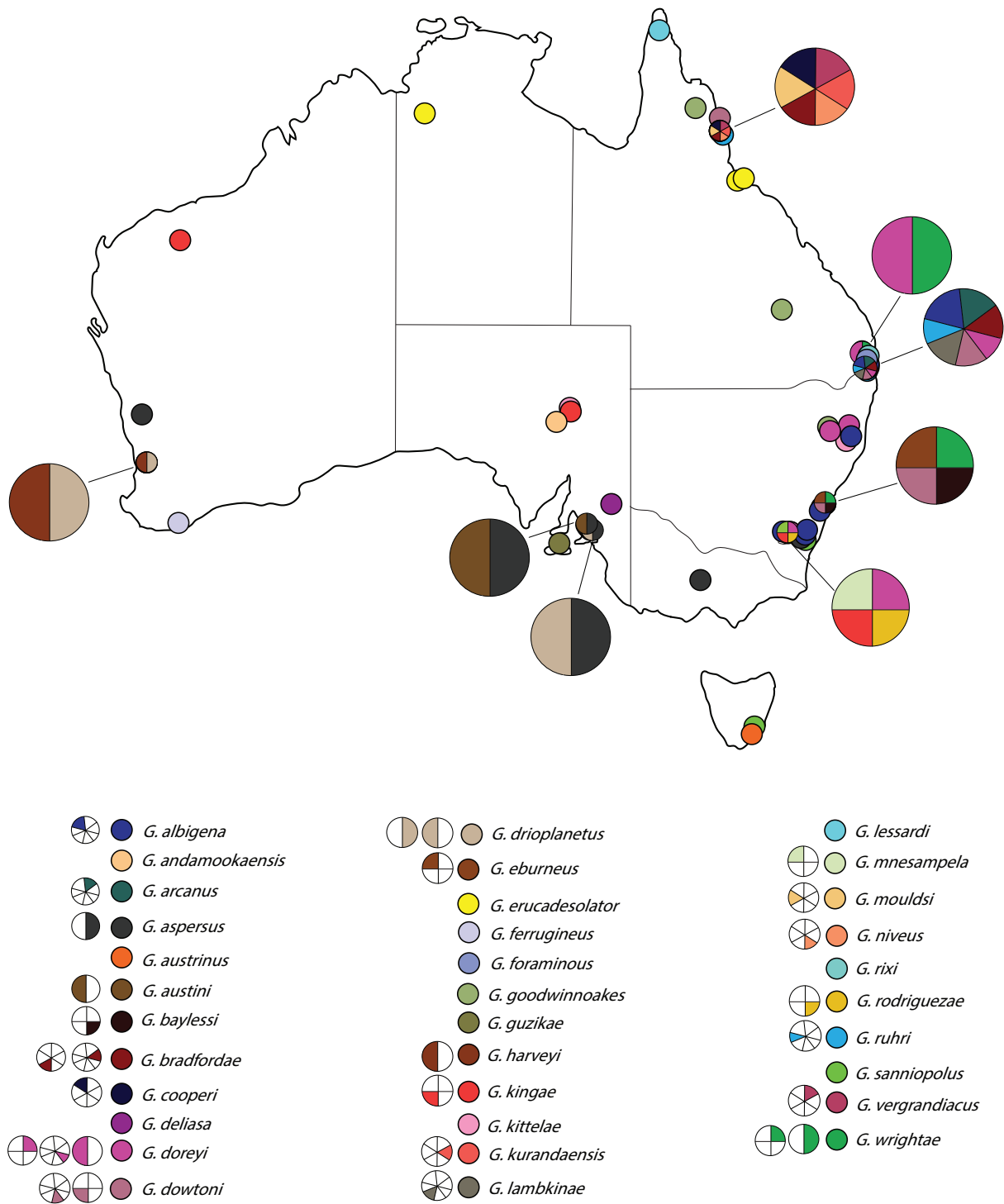


Fig. 3. Known distribution of the described species of *Glyptapanteles* Ashmead, 1904 from Australia, represented by coloured circles or part circles, with each species represented by a different colour (see key to colours below map).

Distribution of the described species of *Glyptapanteles* from Australia

Sampling sites for this study were restricted to where there had been previous collections made into ethanol, allowing specimens to be DNA sequenced. The current known distribution of species of *Glyptapanteles* in Australia is therefore highly biased towards the recent field trips of researchers who deposit bulk samples into accessible state collections, or the field sites utilised by the study team and collaborators. The diversity of species at particular sites is also strongly biased towards collecting effort, with sites where long-term Malaise trapping occurred (e.g., Kuranda and Lamington National Park, both in Queensland; Fig. 3 represented by coloured wheels with at least six species) having a large number of species present. Sixteen of the 34 species of *Glyptapanteles* found in Australia are currently known from only one locality, which is most likely a reflection of limited sampling.

Taxonomic treatment of species

Class Insecta Linnaeus, 1758
Order Hymenoptera Linnaeus, 1758
Family Braconidae Latreille, 1829
Subfamily Microgastrinae Foerster, 1962

Genus *Glyptapanteles* Ashmead, 1904

Glyptapanteles Ashmead, 1904: 147.

Glyptapanteles – Mason 1981: 105. — Austin & Dangerfield 1992: 32. — Arias-Penna *et al.* 2019a: 561. For complete list of earlier synonyms and bibliography, see Shenefelt (1972) and Fernández-Triana *et al.* (2020).

Type species

By monotypy, *Apanteles ashmeadi* Wilkinson, 1928: 84, a replacement name for *Glyptapanteles manilae* Ashmead, 1904.

Diagnosis

The diagnosis by Fernández-Triana *et al.* (2020) works well for the Australian fauna and is as follows: “...fore wing without an areolet; propodeum that is either completely smooth (often) to more or less rugose (more rarely), with a median longitudinal carina that is entirely absent (often), partially defined posteriorly (often) to complete and strong (rarely), or no median carina but instead a series of very short carinae radiating from the nucha (rarely); T1 narrows towards the posterior margin, usually strongly (almost always), or more parallel-sided, or rounded at apex, as in some species of *Protapanteles* (rarely); T2 is almost always subtriangular or trapezoidal (rarely shaped differently); ovipositor and ovipositor sheaths are relatively short (usually) to moderately long (rarely); setae at apex of ovipositor sheaths relatively long (as long or longer than setae on hypopygium)” (Fernández-Triana *et al.* 2020: 36).

Key to the described species groups and to the morphologically distinct species of *Glyptapanteles* from Australia

This key is based on adult females. Due to there likely being many other undescribed species of *Glyptapanteles* in Australia, it should be treated with caution as undescribed species could possibly key to described species within this key. We feel that a morphological key is not particularly useful for this group, other than for separating the species groups which are quite distinct (e.g., the *G. albigena* species group, that has the gena with a pale spot), but we present one here for completeness. Ideally, morphological identifications should be supported with *COI* and *wingless* DNA barcodes. Species that come out together at a couplet are not reliably distinguished using morphology (e.g., couplet 7).

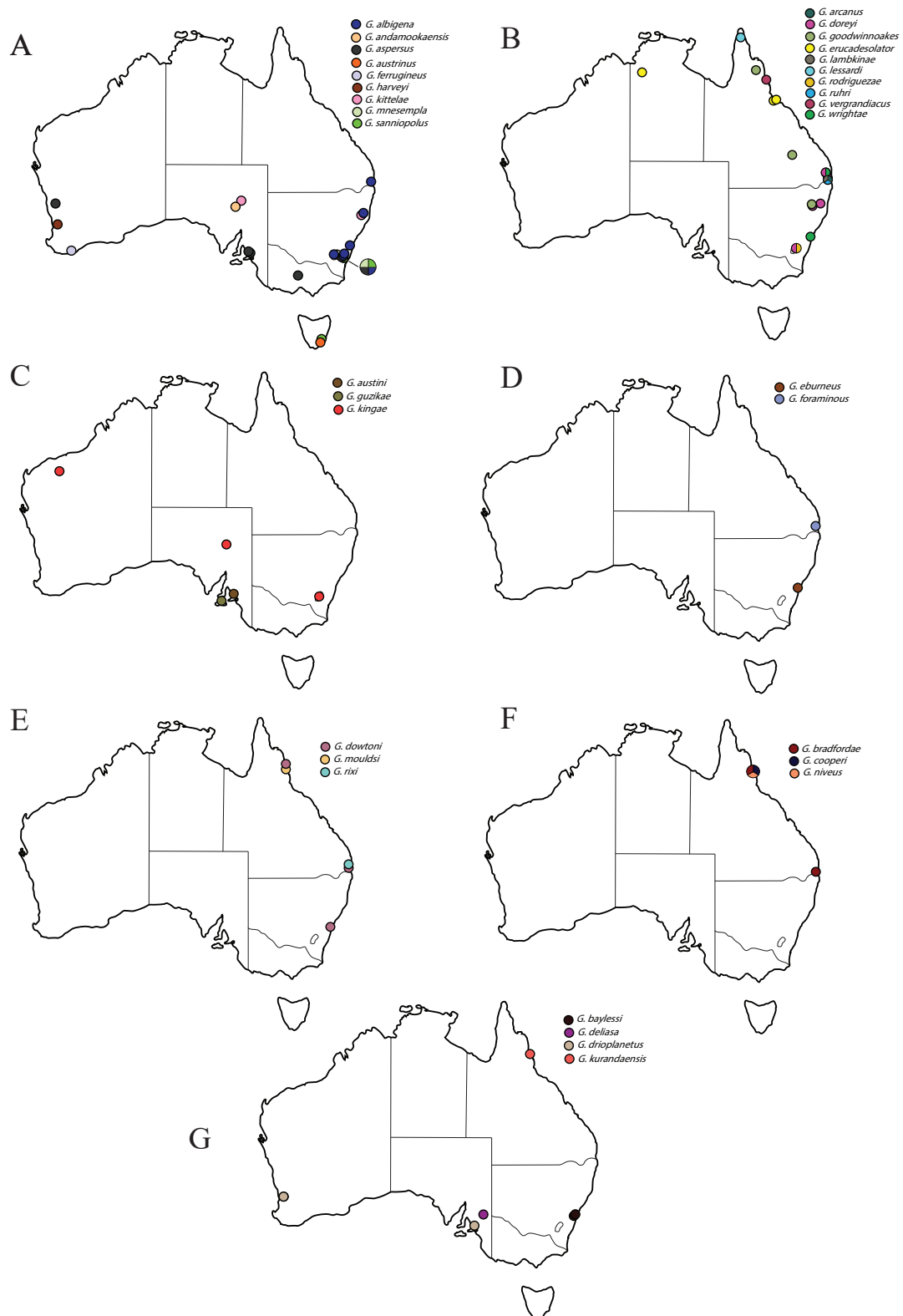


Fig. 4. Distribution of species groups of *Glyptapanteles* Ashmead, 1904 in Australia. **A.** *G. albigena* species group. **B.** *G. arcanus* species group. **C.** *G. austini* species group. **D.** *G. eburneus* species group. **E.** *G. mouldsi* species group. **F.** *G. niveus* species group. **G.** Unplaced species of *Glyptapanteles* in Australia.

1. Gena with a pale spot (*G. albigena* species group) (Fig. 5A-D) 2
 - Gena without a pale spot (Fig. 5E) 8
2. Gena with a large pale spot (Fig. 5A–B) 3
 - Gena with a small pale spot (e.g., Fig. 5C–D) 4
3. Gena with a pale spot measuring at least one third of face height (Fig. 5A)
 - *Glyptapanteles albigena* Fagan-Jeffries, Bird & Austin sp. nov.
 - Gena with a pale spot less than one third of face height (Fig. 5B)
 - *Glyptapanteles sanniolus* Fagan-Jeffries, Bird & Austin sp. nov.
4. T2 pale (e.g., Fig. 6A–E) *G. mnesampela* Austin, 2000
 - T2 dark (e.g., Fig. 6F) 5
5. Propodeum with median carina faintly indicated posteriorly and anteriorly (Fig. 7A); and gena with a barely visible pale spot (Fig. 5D) *Glyptapanteles harveyi* Fagan-Jeffries, Bird & Austin sp. nov.
 - Propodeum with median carina completely absent (e.g., Fig. 7B); gena with a clearly visible pale spot (e.g., Fig. 5C) 6
6. Hind femur mostly dark (sometimes with lighter area proximally) (Fig. 8A)
 - *Glyptapanteles andamookaensis* Fagan-Jeffries, Bird & Austin sp. nov.
 - Hind femur mostly light brown or pale (e.g., Fig. 8B) 7
7. Mesosoma with strong red tinge (Fig. 9A)
 - *Glyptapanteles ferrugineus* Fagan-Jeffries, Bird & Austin sp. nov.
 - Mesosoma dark, no strong red tinge (e.g., Fig. 9B)
 - *Glyptapanteles kittelae* Fagan-Jeffries, Bird & Austin sp. nov.,
 - *Glyptapanteles austrinus* Fagan-Jeffries, Bird & Austin sp. nov. and
 - *Glyptapanteles aspersus* Fagan-Jeffries, Bird & Austin sp. nov.

These species cannot be reliably identified using morphology alone; see under the relevant species for molecular information which can be used to differentiate them

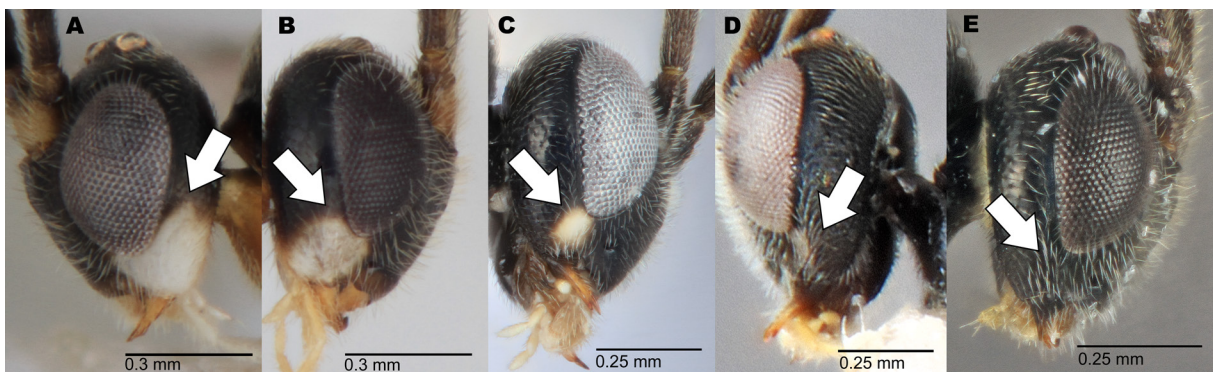


Fig. 5. **A.** *Glyptapanteles albigena* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (ANIC 32 130334), arrow indicating a large pale gena spot. **B.** *G. sanniolus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (ANIC 32 130370), arrow indicating a large pale gena spot. **C.** *G. kittelae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-46156) arrow indicating small (clearly visible) pale gena spot. **D.** *G. harveyi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (WAM E109889), arrow indicating small (faint, barely visible) pale gena spot. **E.** *G. baylessi* Fagan-Jeffries, Bird & Austin sp. nov. paratype, ♀ (AM K.517936), gena without a pale spot.

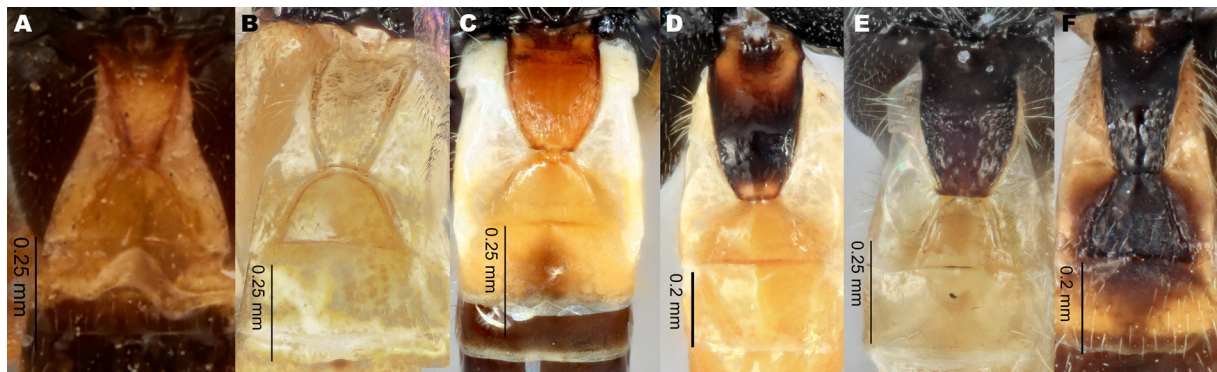


Fig. 6. **A.** *Glyptapanteles mnesampela* Austin, 2000, holotype, ♀ (ANIC 32-141445), T1 and T2 pale. **B.** *G. eburneus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (AM K.517935), T1 and T2 pale. **C.** *G. rixi* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250981), T2 pale, T1 darker than T2. **D.** *G. mouldsi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250978), T1 dark, T2 pale. **E.** *G. dowtoni* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250953), T1 dark, T2 pale. **F.** *G. harveyi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (WAM E109889), T1 dark, T2 dark.

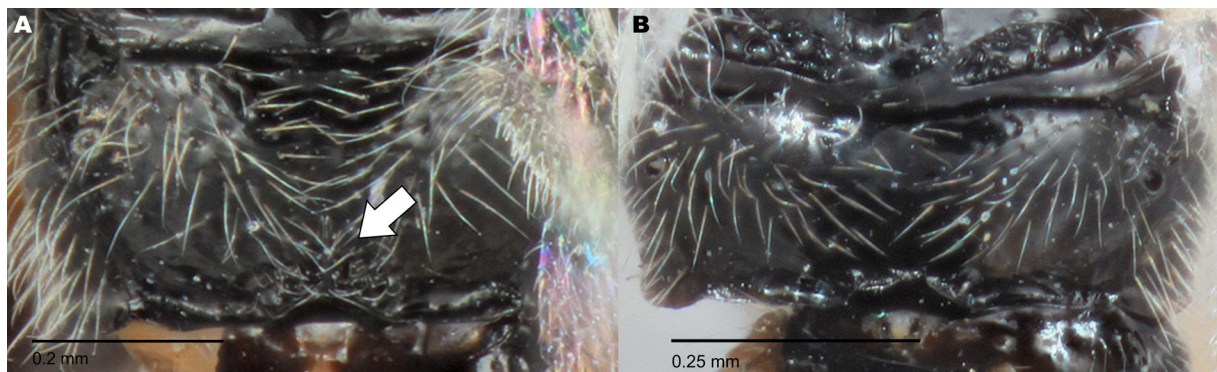


Fig. 7. **A.** *Glyptapanteles harveyi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (WAM E109889), arrow indicating faint median carina at the posterior end of the propodeum. **B.** *G. kittelae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-46156), propodeum with median carina completely absent.

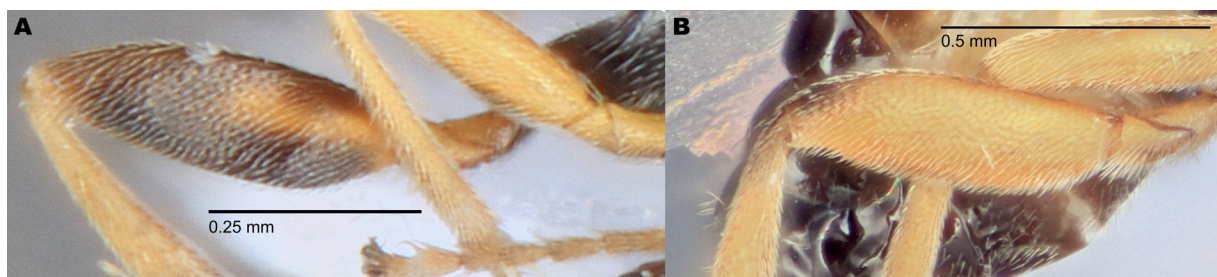


Fig. 8. **A.** *Glyptapanteles andamookaensis* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-035451), hind femur mostly dark. **B.** *G. kittelae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-46156), hind femur mostly light brown.

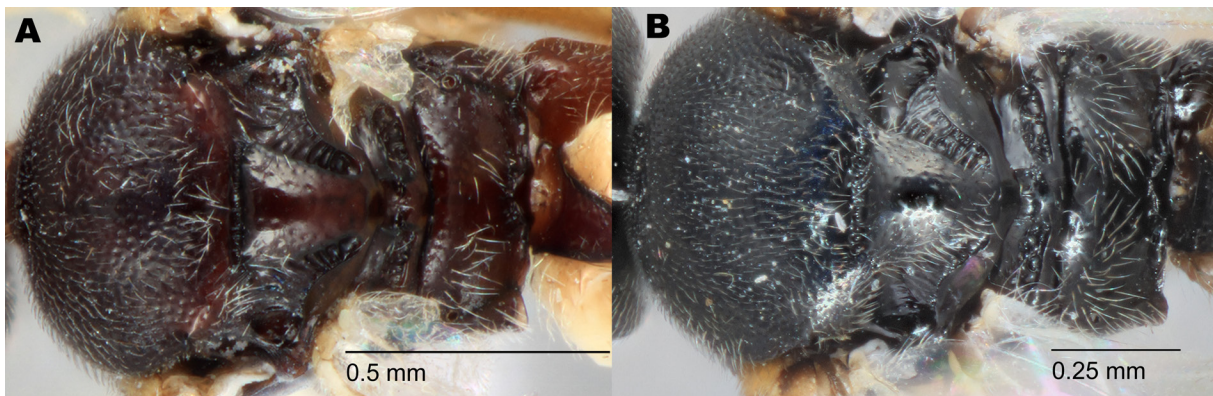


Fig. 9. A. *Glyptapanteles ferrugineus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (ANIC 32 130189), mesosoma with strong red tinge. B. *G. kittelae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-46156), mesosoma dark, no strong red tinge.

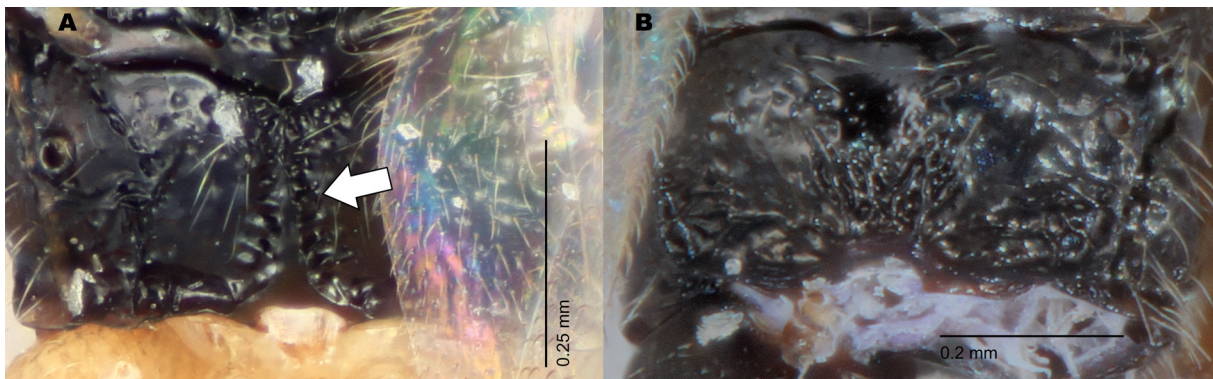


Fig. 10. A. *Glyptapanteles eburneus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (AMK.517935), propodeum with median carina clear and complete (indicated by arrow). B. *G. rixi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250982), propodeum with median carina absent.

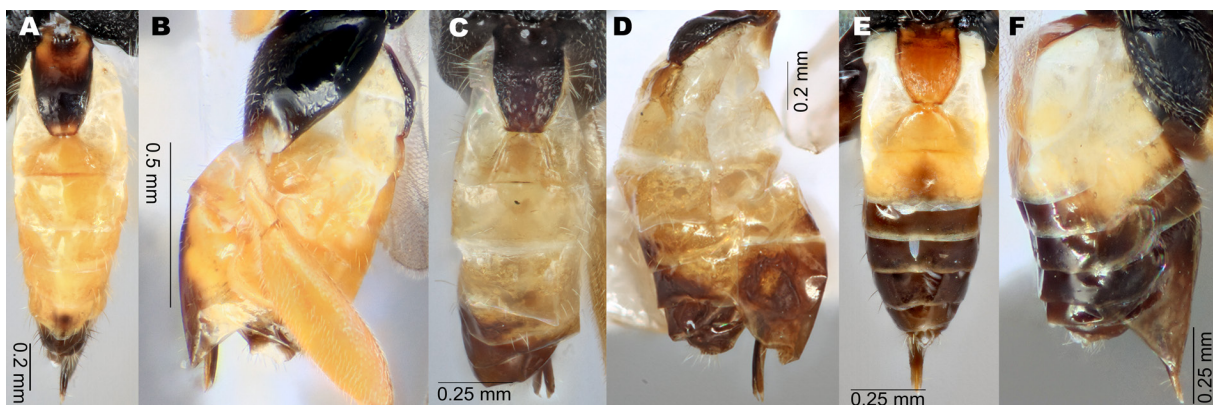


Fig. 11. A–B. *Glyptapanteles mouldsi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250978). A. Dorsal metasoma. B. Lateral metasoma. C–D. *G. dowtoni* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250953). C. Dorsal metasoma. D. Lateral metasoma. E–F. *G. rixi* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250981). E. Dorsal metasoma. F. Lateral metasoma.

8. T2 pale (e.g., Fig. 6A–E) 9
 – T2 dark (e.g., Fig. 6F) 11
9. T1 same colour as T2, extremely pale (Fig. 6B); propodeum with median carina clear and complete; hind coxa pale (*G. eburneus* species group, part)
 *Glyptapanteles eburneus* Fagan-Jeffries, Bird & Austin sp. nov.
 – T1 darker than T2, either dark or orange-brown (e.g., Fig. 6C–D); propodeum with median carina absent; hind coxa dark (*G. mouldsi* species group) 10
10. T1 dark, S3–5 mostly pale or yellow other than hypopygium (which is dark); T3–5 mostly pale or yellow (Fig. 11A–D) *Glyptapanteles mouldsi* Fagan-Jeffries, Bird & Austin sp. nov. and
 *Glyptapanteles dowtoni* Fagan-Jeffries, Bird & Austin sp. nov.
 These species cannot be reliably identified using morphology alone; see under the relevant species for molecular information which can be used to differentiate them
 – T1 orange to light brown, S5–7 dark (including hypopygium); T4–6 completely dark (Fig. 11E–F)
 *Glyptapanteles rixi* Fagan-Jeffries, Bird & Austin sp. nov.
11. T1 pale; hind coxa pale (*G. eburneus* species group, part)
 *Glyptapanteles foraminous* Fagan-Jeffries, Bird & Austin sp. nov.
 – T1 dark; hind coxa dark 12
12. Anteromesoscutum with very sparse deep punctures, always with some smooth areas greater than diameter of punctures; T1 never wedge-shaped, propodeum coarsely rugose; sometimes with slight orange markings on postero-lateral anteromesoscutum (*G. niveus* species group) (e.g., Fig. 12A–B) 13
 – Anteromesoscutum normally with punctures denser than in Fig. 12A–B, smooth areas generally smaller than diameter of punctures; T1 variable, including sometimes wedge-shaped, propodeum variable, coarsely rugose to smooth; sometimes with strong orange markings on postero-lateral anteromesoscutum (e.g., Fig. 12C–D) 14
13. Hind coxa with strong punctures covering most of area; antennal flagellomeres all dark
 *Glyptapanteles bradfordae* Fagan-Jeffries, Bird & Austin sp. nov.
 – Hind coxa with only sparse punctures, particularly smooth anteriorly; antennal flagellomeres sometimes with white distal segments
 *Glyptapanteles cooperi* Fagan-Jeffries, Bird & Austin sp. nov. and
 *Glyptapanteles niveus* Fagan-Jeffries, Bird & Austin sp. nov.
 These species cannot be reliably identified using morphology alone; see under the relevant species for molecular information which can be used to differentiate them
14. Postero-lateral anteromesoscutum with strong orange markings (Fig. 12C) [whole body strongly punctured, propodeum strongly rugose]
 *Glyptapanteles baylessi* Fagan-Jeffries, Bird & Austin sp. nov.
 – Postero-lateral anteromesoscutum without strong orange markings (e.g., Fig. 12D) [sculpturing of propodeum variable, sometimes strongly rugose] 15
15. Ventral side of antennal scape (at least in distal half) darker than or the same colour as flagellomeres (e.g., Fig. 13A) 16
 – Ventral side of antennal scape (at least in distal half) paler than flagellomeres (*G. arcanus* species group) (e.g., Fig. 13B) 20

16. Fore wing veins r and 2RS narrow and long, each significantly longer than vein 2m and smoothly curved (not meeting at sharp angle) (Fig. 14A)
 *Glyptapanteles kurandaensis* Fagan-Jeffries, Bird & Austin sp. nov.
 – Fore wing veins r and 2RS each only slightly longer than vein 2m (Fig. 14B), sometimes meeting at sharp angle 17
17. Tegula dark *G. deliasa* Austin & Dangerfield, 1992
 – Tegula pale 18
18. Maximum height of mesoscutellum lunules less than 0.64× maximum height of lateral face of mesoscutellum (Fig. 15A) [hind femur darkening posteriorly, propodeum smooth]
 *Glyptapanteles drioplanetus* Fagan-Jeffries & Austin, 2021

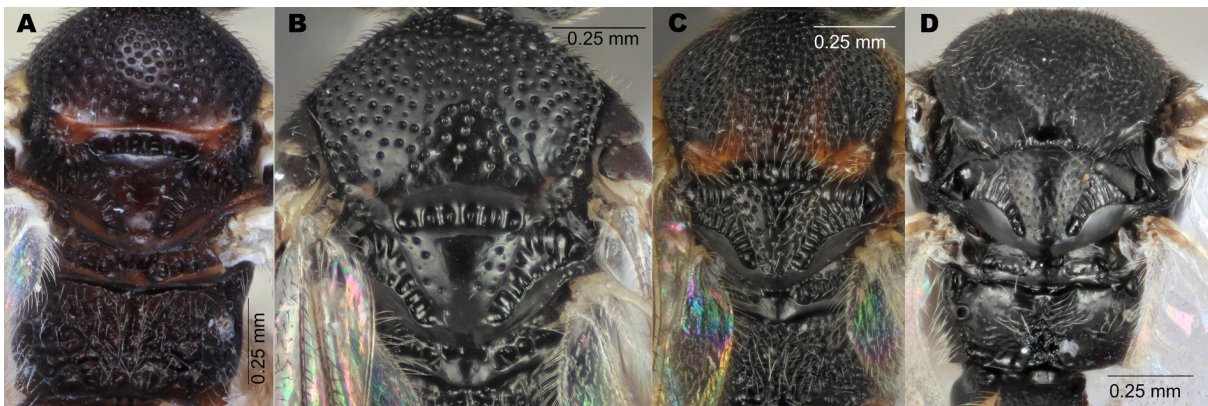


Fig. 12. **A.** *Glyptapanteles bradfordae* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250944), anteromesoscutum with very sparse deep punctures and slight orange markings. **B.** *G. cooperi* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250947), anteromesoscutum with very sparse deep punctures and no orange markings. **C.** *G. baylessi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (AM K.517936), anteromesoscutum with dense punctures, strong orange markings on the postero-lateral corners. **D.** *G. kurandaensis* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250971), with dense shallow punctures, but no strong orange markings on the anteromesoscutum.

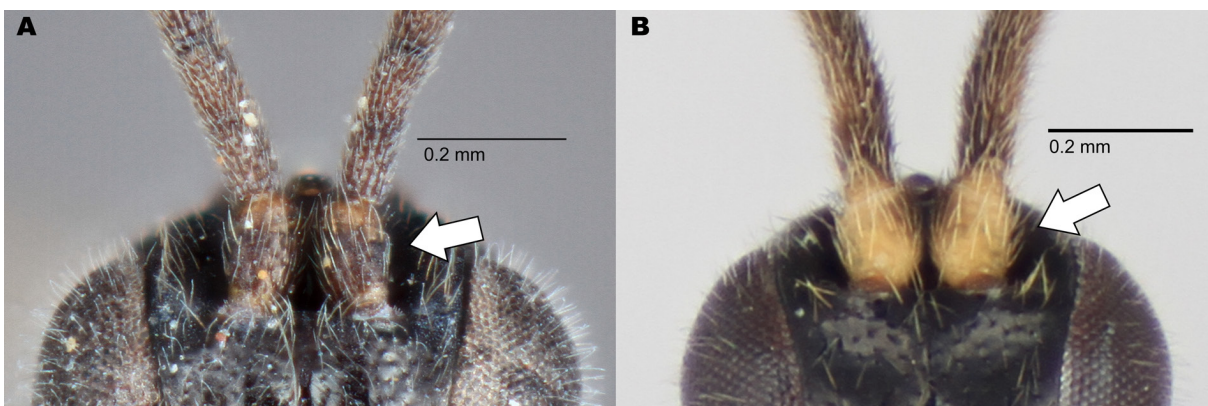


Fig. 13. **A.** *Glyptapanteles deliasa* Austin & Dangerfield, 1992, paratype, ♀ (WINC), ventral half of antennal scape darker than flagellomeres. **B.** *G. ruhri* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250986), ventral half of antennal scape paler than flagellomeres.

- Maximum height of mesoscutellum lunules approximately 0.75× maximum height of lateral face of mesoscutellum (e.g., Fig. 15B) [hind femur completely dark or mostly pale yellow with dark patch posteriorly, propodeum smooth or with some shallow indistinct sculpturing in centre] (*G. austini* species group) 19
- 19. Hind femur mostly pale with dark area posteriorly [propodeum smooth and shiny]
 *Glyptapanteles guzikae* Fagan-Jeffries, Bird & Austin sp. nov.
- Hind femur completely dark [propodeum smooth or with some sculpturing in centre]
 *Glyptapanteles kingae* Fagan-Jeffries, Bird & Austin sp. nov. and
 *Glyptapanteles austini* Fagan-Jeffries & Bird sp. nov.
- These species cannot be reliably identified using morphology alone; see under the relevant species for molecular information which can be used to differentiate them
- 20. Propodeum with coarse rugose sculpturing over most of area 21
- Propodeum smooth, or with rugose sculpturing only in posterior half of centre, anterior half punctured or smooth 22

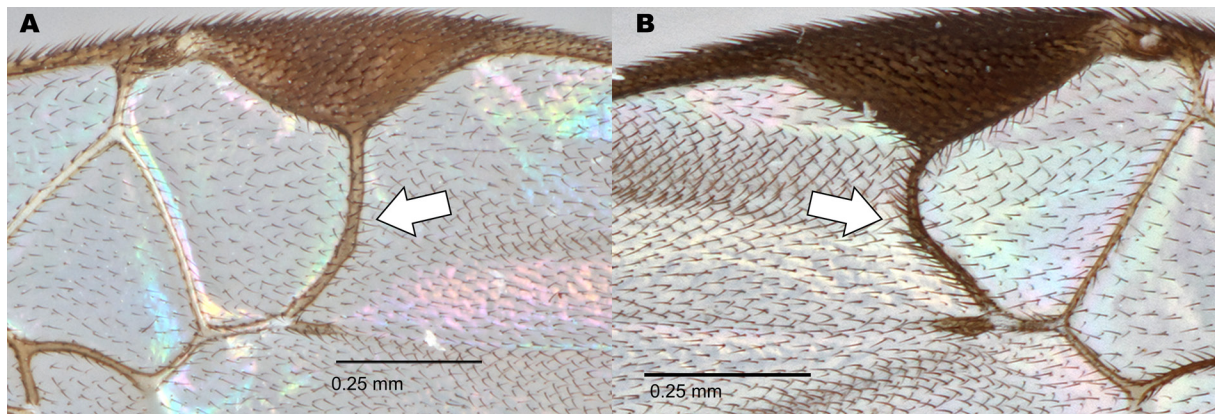


Fig. 14. **A.** *Glyptapanteles kurandaensis* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250969), fore wing, arrow indicating vein r. **B.** *G. kingae* Fagan-Jeffries, Bird & Austin sp. nov., holotype (SAMA 32-46154), arrow indicating vein r.

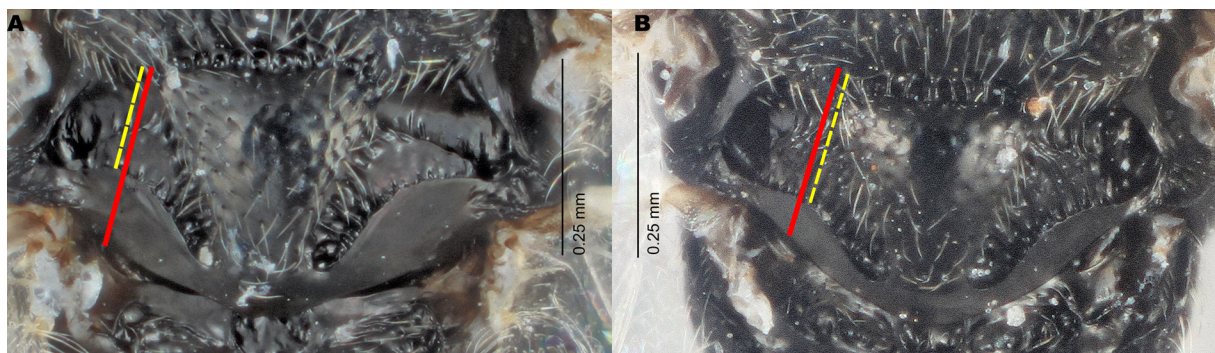


Fig. 15. **A.** *Glyptapanteles drioplanetus* Fagan-Jeffries & Austin, 2021 (SAMA 32-45154). **B.** *G. austini* Fagan-Jeffries & Bird sp. nov., paratype, ♀ (SAMA 32-46151). Solid red line indicating lateral face of the mesoscutellum, yellow dashed line indicating mesoscutellum lunules.

coast of Australia, from the northern regions of QLD through to central NSW and the ACT. One species, *G. erucadesolator* sp. nov., is also known from northern NT (Fig. 4B).

Species in the *G. arcanus* species group can be separated from the other described species of *Glyptapanteles* in Australia by the combination of the following characters: gena without a pale spot, T1 and T2 dark, anteromesoscutum punctures not extremely sparse and deep (i.e., not as in the *niveus* species group), no orange markings on anteromesoscutum postero-laterally, propodeum either strongly sculptured (rugose) or with punctures in the anterior half and shallow rugose sculpturing in the posterior half, ventral side of antennal scape (at least in distal half) paler than flagellomeres, tegula pale.

***Glyptapanteles austini* species group**

The *G. austini* species group contains three species: *G. austini* Fagan-Jeffries & Bird sp. nov., *G. guzikae* Fagan-Jeffries, Bird & Austin sp. nov. and *G. kingae* sp. nov., which are a monophyletic, molecularly distinct lineage (Fig. 2). Two species in this group (*G. guzikae* sp. nov. and *G. austini* sp. nov.) are known from single localities in southern SA, whilst *G. kingae* sp. nov. has a broad distribution and is found in the northern half of WA, central SA and the ACT (Fig. 4C). The group can be separated from the other species of *Glyptapanteles* described from Australia by the following combination of characters: gena without a pale spot, T1–T2 dark, anteromesoscutum all dark and with punctures without smooth areas greater than the diameter of punctures (i.e., anteromesoscutum not as in the *G. niveus* species group (i.e., not as in Fig. 12B)), propodeum not coarsely rugose, T1 only parallel for at most $\frac{2}{3}$ of length before narrowing posteriorly, mesoscutellar disk without dense, strong punctures, scape darker or the same colour as flagellomeres, tegula pale, labrum dark.

***Glyptapanteles eburneus* species group**

The *G. eburneus* species group currently contains only two closely related species, *G. eburneus* Fagan-Jeffries, Bird & Austin sp. nov. and *G. foraminous* Fagan-Jeffries, Bird & Austin sp. nov. which are differentiated from all other species of *Glyptapanteles* in Australia by having the gena without a pale spot, T1 completely pale, hind coxa pale, deep punctures on the anteromesoscutum and a clear median carina on the propodeum. This species group is known from two single localities on the east coast of Australia, in QLD and NSW (Fig. 4D).

***Glyptapanteles mouldsi* species group**

The *G. mouldsi* species group can be distinguished from all of the other *Glyptapanteles* described from Australia by having the gena without a pale spot, T2 pale in colouration, with T1 darker than T2 and no median carina on the propodeum. It contains three newly described species: *G. dowtoni* Fagan-Jeffries, Bird & Austin sp. nov., *G. mouldsi* Fagan-Jeffries, Bird & Austin sp. nov. and *G. rixi* Fagan-Jeffries, Bird & Austin sp. nov. The group is paraphyletic in the current phylogeny (Fig. 2). The *G. mouldsi* species group is currently only known from the east coast of Australia, from northern QLD to central NSW (Fig. 4E).

***Glyptapanteles niveus* species group**

The *G. niveus* species group can be separated from the other described *Glyptapanteles* from Australia by having the gena without a pale spot, T1 and T2 dark, propodeum coarsely rugose and the anteromesoscutum with very sparse deep punctures, always with some smooth areas larger than the diameter of punctures, face strongly punctured and T1 not wedge-shaped. All the described species also have a vividly white fore coxa, mid coxa, hind trochanter and anterior metasomal sternites. The species group currently contains three described species: *G. bradfordae* Fagan-Jeffries, Bird & Austin sp. nov., *G. cooperi* Fagan-Jeffries, Bird & Austin sp. nov. and *G. niveus* Fagan-Jeffries, Bird & Austin sp. nov.

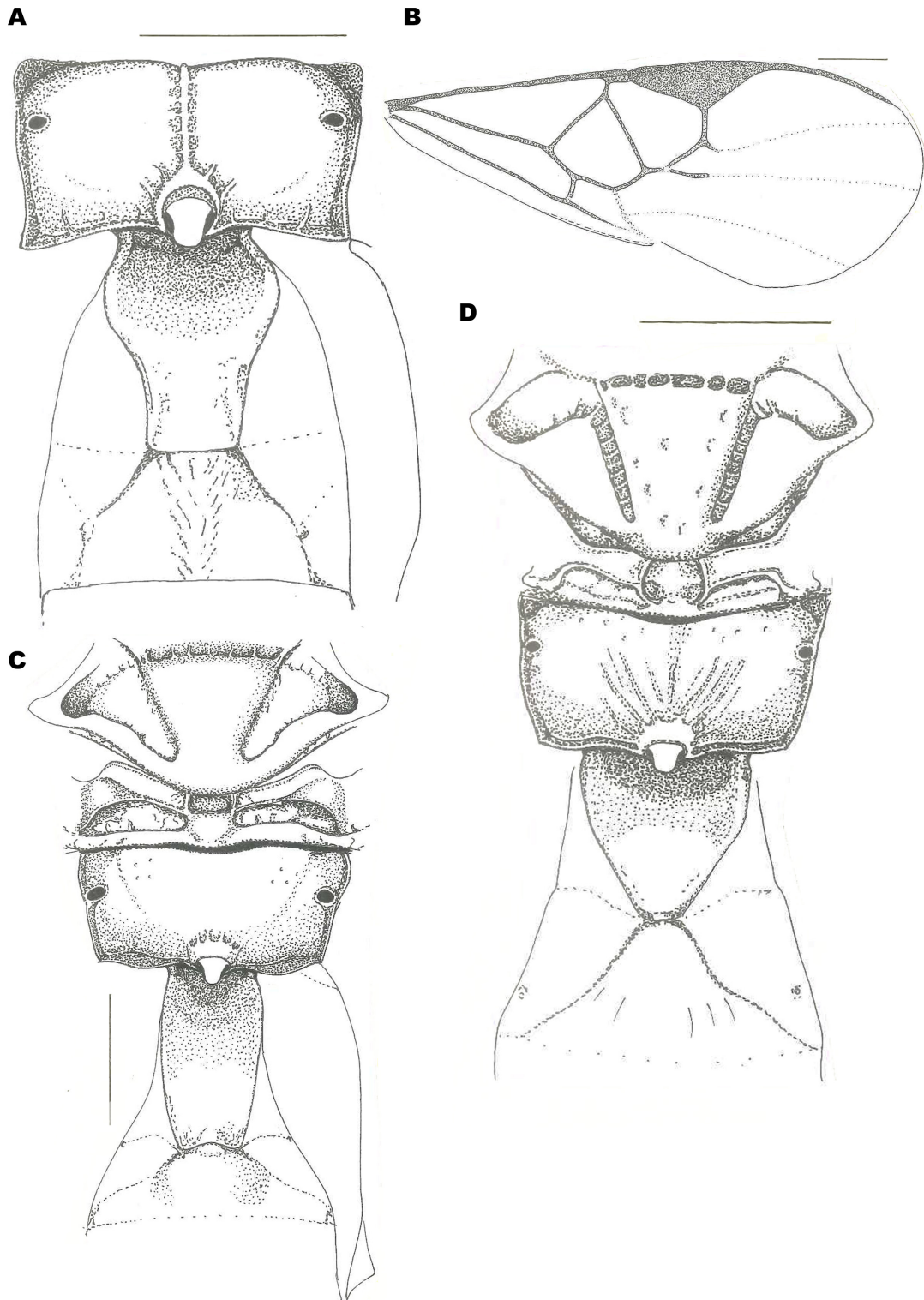


Fig. 16. A–B. *Glyptapanteles afimaluanus* (Fullaway, 1941), holotype, ♀ (BPBM) A. Propodeum and T1–2. B. Fore wing. C. *G. fullawayi* Austin & Dangerfield, 1992, holotype, ♂ (BPBM), dorsal scutellum, propodeum and T1–2. D. *G. operculinae* (Fullaway, 1941), holotype, ♀ (BPBM), dorsal scutellum, propodeum and T1–2. Reproduced from Austin & Dangerfield (1992).

All three species also have larger eyes (and consequently, smaller malar spaces) than most other species of *Glyptapanteles* from Australia. This species group is currently only known from eastern QLD, from Kuranda (northern QLD) and from near Brisbane (southern QLD) (Fig. 4F).

Unplaced species of *Glyptapanteles* from Australasia

There are several species of *Glyptapanteles* from Australasia which we do not place within a species group, either because they do not form a monophyletic group with other species with which they share a clear morphological character, or because they are previously described species without molecular data and/or without clear morphological affinities to any of the assigned species groups. This includes *G. afiamaluanus* (Fullaway, 1941), *G. artonae* (Rohwer, 1926), *G. aucklandensis* (Cameron, 1909), *G. baylessi* Fagan-Jeffries, Bird & Austin sp. nov., *G. deliasa* Austin & Dangerfield, 1992, *G. drioplanetus* Fagan-Jeffries & Austin, 2021, *G. fullawayi* Austin & Dangerfield, 1992, *G. kurandaensis* Fagan-Jeffries, Bird & Austin sp. nov., *G. operculinae* (Fullaway, 1941), *G. phytometrae* (Wilkinson, 1928) and *G. taylori* (Wilkinson, 1928). Of these unplaced species, only *G. baylessi* sp. nov., *G. deliasa*, *G. drioplanetus* and *G. kurandaensis* sp. nov. are found in Australia.

Species treatments

***Glyptapanteles afiamaluanus* (Fullaway, 1941)**

Fig. 16A–B

Diagnosis

Glyptapanteles afiamaluanus is easily separated from any of the described species of *Glyptapanteles* in the Australasian region by the distinctive shape of T1, which broadens significantly in the anterior half and then narrows posteriorly and the strong median carina on the propodeum.

Material examined

No specimens examined, illustrations in Austin & Dangerfield (1992: figs 49–50) used as reference (Fig. 16A–B).

Remarks

Host unknown. This species is very distinctive amongst the Australasian fauna and the genus placement is very tentative.

Distribution

Samoa (Upolu Is).

***Glyptapanteles albigena* Fagan-Jeffries, Bird & Austin sp. nov.**

[urn:lsid:zoobank.org:act:B83B4BAE-3154-4717-A00E-EB6FC9EFF9CD](https://zoobank.org/act:B83B4BAE-3154-4717-A00E-EB6FC9EFF9CD)

Figs 5A, 17

Diagnosis

Glyptapanteles albigena sp. nov. is in the *G. albigena* species group and can be separated from other members of the group by the gena having a very large pale spot, covering at least a third of the gena height.

Etymology

The species epithet ‘*albigena*’ is from the Latin adjective ‘album’ (meaning ‘white’) and refers to the pale spot on the gena that is characteristic of this species and the broader species group.

Material examined**Holotype**

AUSTRALIA • ♀; New South Wales, Werrikimbe National Park, Plateau Beach Cpg; 31.1807, 152.3266; 1055 m a.s.l.; 17–20 Jan. 2018; K.M. Bayless leg.; sweep ferns along trail; Extraction1110, BOLD: AUGLY048-21; ANIC 32 130334.

Paratypes

AUSTRALIA – **Australian Capital Territory** • 1 ♂; Brindabella, Blundells Creek; -35.416667, 148.833333; 8 Jan. 2018; K.M. Bayless leg.; sweep; Extraction1185, BOLD: AUGLY070-21; AM K.379877. – **New South Wales** • 1 ♂; same collection data as for holotype; Extraction1177, BOLD: AUGLY068-21; ANIC 32 130335 • 1 ♂; same collection data as for holotype; Extraction1178, BOLD: AUGLY069-21; ANIC 32 130336 • 1 ♀ (ethanol); Monga National Park, near Penance Grove Walk; -35.597372, 149.912126; 9 Nov. 2019–16 Jan. 2020; K.M. Bayless and J.G. Lumbers leg.; Malaise trap over stream, trap destroyed by bushfire; 30 Dec. 2019; partially dried out; Extraction1664, BOLD: AUGLY126-21; ANIC 32 130337 • 1 ♂ (ethanol); Budawang National Park, Mt Budawang Road; -35.4563, 149.9955; 750 m a.s.l.; 16 Jan.–10 Feb. 2019; K.M. Bayless and X. Li leg.; Malaise over Feagans Creek; Extraction1634, BOLD: AUGLY120-21; ANIC 32 130338 • 1 ♀; Barren Grounds NR, 21 m NE of Barren Ground Rd; -34.66971, 150.71167; 1–6 Feb. 2020; K.M. Bayless and J.G. Lumbers leg.; Malaise trap, wet sclerophyll forest; Extraction872, BOLD: AUGLY013-21; ANIC 32 130339 • 1 ♀; same collection data as for preceding; Extraction1136, BOLD: AUGLY061-21; ANIC 32 130340 • 1 ♂; same collection data as for preceding; Extraction871, BOLD: AUGLY012-21; ANIC 32 130341 • 1 ♂; same collection data as for preceding; Extraction1133, BOLD: AUGLY060-21; ANIC 32 130342 • 1 ♂ (ethanol); same collection data as for preceding; 23–29 Jan. 2020; Extraction1607, BOLD: AUGLY114-21; ANIC 32 130343 • 1 ♂; same collection data as preceding; Extraction1609, BOLD: AUGLY115-21; ANIC 32 130344 • 1 ♂ (ethanol); same collection data as for preceding; Extraction1617, BOLD: AUGLY117-21; ANIC 32 130345. – **Queensland** • 1 ♀; Lamington National Park; -28.259, 153.162; 1142 m a.s.l.; 11–21 Mar. 2007; C Lambkin and N. Starick leg.; IBISCA Plot # IQ-1100-B rainforest Malaise trap; Extraction609, BOLD: AUMIC394-18; QM T208405.

Description**Female**

COLOURATION. Gena with a pale spot; labrum pale; scape colour in ventral half paler than flagellomeres at proximal end; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark, dark with significant orange patches on posterolateral corners or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark or dark reddish-brown; T2 sclerotised area dark or dark reddish-brown; T2 lateral area dark, extends past indentation, but then pale; T3 dark, mostly pale with darker patch in centre or mostly dark with paler lateral areas; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.3 mm; fore wing length 2.4 mm; antennal length similar to body length.

HEAD. Antennal flagellomere 14 length/width 1.71–2.16; antennal flagellomere 2 length/width 3.00–3.71; OOD/POD 1.67–1.71; IOD/POD 1.33–1.57.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size. Often smoother in posterior centre

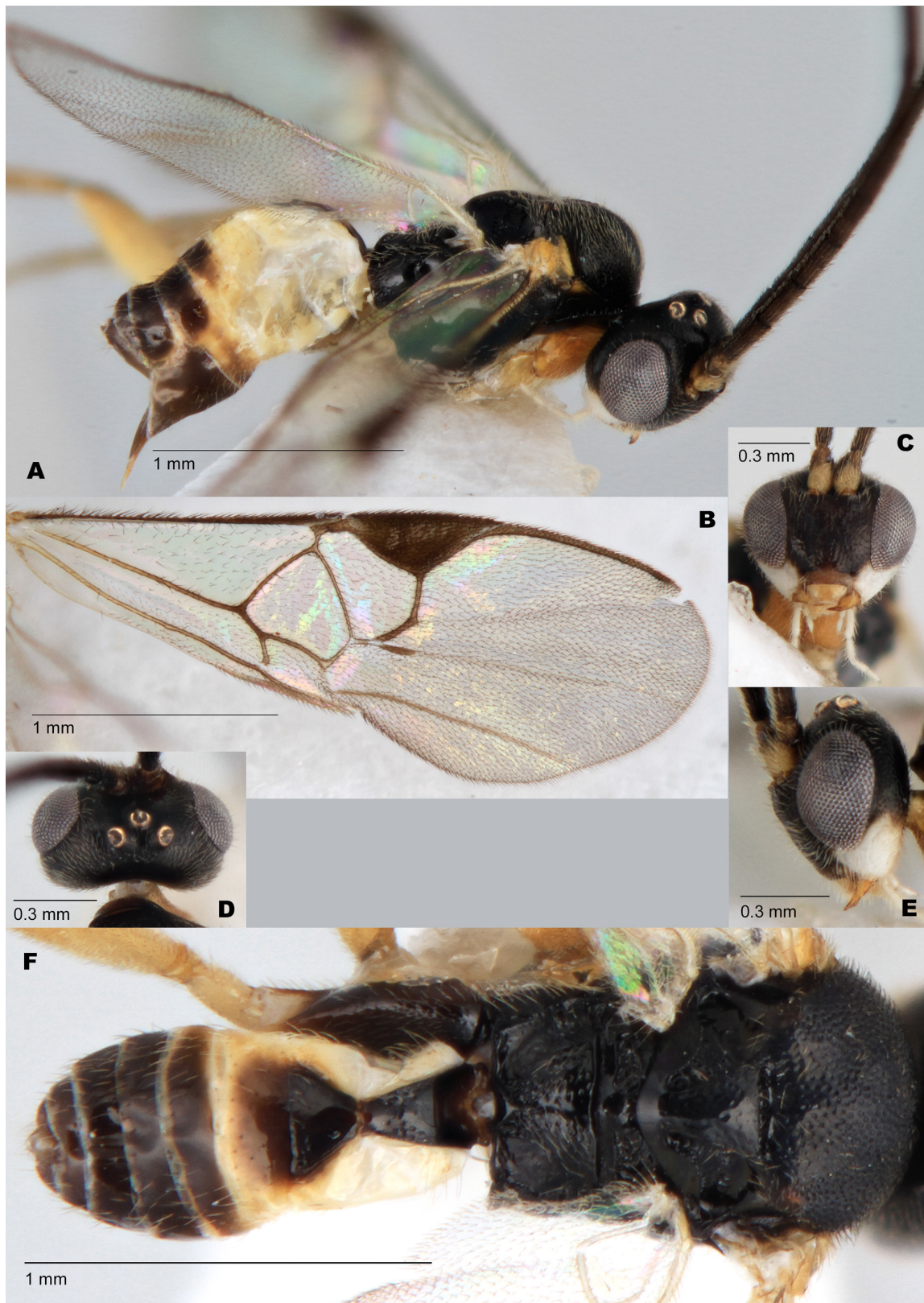


Fig. 17. *Glyptapanteles albigena* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (ANIC 32 130334). **A.** Lateral habitus. **B.** Fore wing. **C.** Anterior head. **D.** Dorsal head. **E.** Lateral head. **F.** Dorsal habitus.

or shallow punctures, space between punctures generally smaller than diameter of punctures; scutellar disk sculpturing with shallow or very shallow punctures scattered over most of area; 6–8 pits in the scutellar sulcus; propodeum with median carina faintly indicated for all of length or sometimes only in posterior half; rest of propodeum either smooth with some punctures in anterior half, or smooth with some shallow rugosity, or smooth with both punctures and some shallow rugosity.

WINGS. Pterostigma length 0.57 mm; pterostigma width 0.23 mm; r 0.16 mm; 2RS 0.15 mm; 2m 0.14 mm; (RS+M)b 0.09 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 smooth and shiny, some shallow scattered punctures on lateral edges or smooth and shiny, some shallow scattered punctures on lateral edges and posterior half; T1 length 0.36 mm; T1 width at posterior edge 0.07 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.15 mm; T2 width at posterior edge 0.26 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, but antenna slightly longer than body.

Remarks

Glyptapanteles albigena sp. nov. constitutes BIN: BOLD:ADL3908 and is 5.48% (p-dist.) divergent from the closet BIN in the database (BOLD:ABA6208; *Glyptapanteles sannipolus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 6.3% different from the most similar *COI* sequence of an Australian specimen (HYAT425-11 and HYAT424-11; *Glyptapanteles sannipolus* sp. nov.). All specimens able to be sequenced for the *wingless* gene share a unique barcode that differs by a minimum of 1 bp from all other species with available sequence data.

Distribution

This species is known from the ACT and NSW and from southern QLD.

Glyptapanteles andamookaensis Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:B0900129-30DE-49B5-9853-D09AD31602AC](https://zoobank.org/act:B0900129-30DE-49B5-9853-D09AD31602AC)

Figs 8A, 18

Diagnosis

Glyptapanteles andamookaensis sp. nov. is in the *G. albigena* species group and can be separated from the other members of the species group by the pale spot on the gena being small but clearly visible, T2 dark, propodeum with the median carina completely absent and hind femur mostly dark (sometimes with a lighter area anteriorly).

Etymology

Glyptapanteles andamookaensis sp. nov. is named after the collection locality.

Material examined

Holotype

AUSTRALIA • ♀; South Australia, Andamooka Station; -30.8198802, 137.1783585 to -30.6998403, 137.1574435; 31 Aug. 2016; R. Leijs leg.; Vehicle net, Bush Blitz Lake Torrens; Extraction260, BOLD: AUMIC130-18; SAMA 32-035451.

Paratype

AUSTRALIA • 1 ♀; same collection data as for holotype; Extraction565, BOLD: AUMIC357-18; SAMA 32-035452.

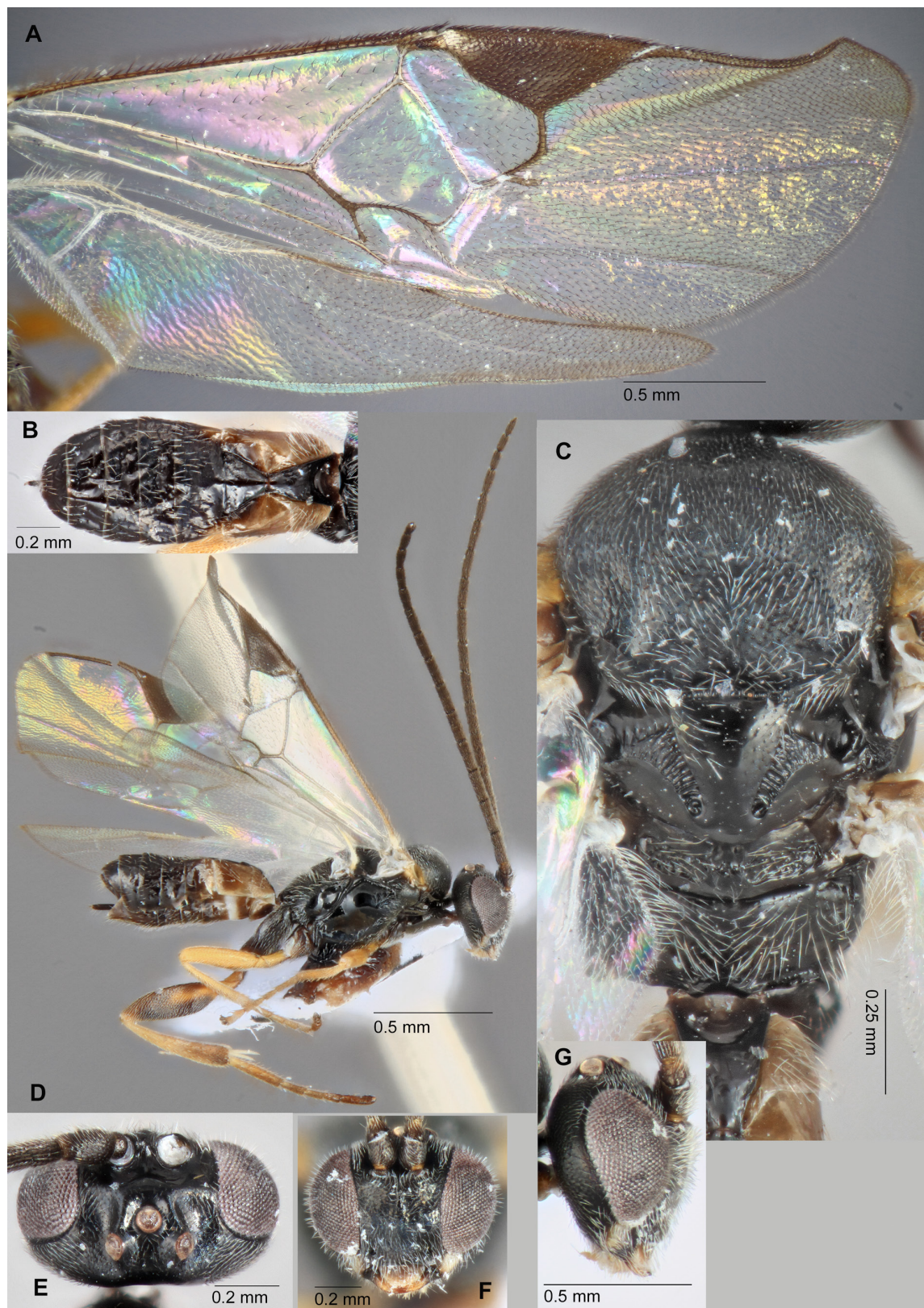


Fig. 18. *Glyptapanteles andamookaensis* Fagan-Jeffries, Bird & Austin sp. nov. **A–C, F–G.** Holotype, ♀ (SAMA 32-035451). **D–E.** Paratype, ♀ (SAMA 32-035452). **A.** Fore wing. **B.** Dorsal metasoma. **C.** Dorsal mesosoma. **D.** Lateral habitus. **E.** Dorsal head. **F.** Anterior head. **G.** Lateral head.

Description

Female

COLOURATION. Gena with a pale spot; labrum reddish-brown; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur dark reddish-brown; fore tibia orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark, dark area extending past indentation to non-sclerotised area, but T2 lateral area then pale; T3 dark; T4+ dark.

HOLOTYPE BODY MEASUREMENTS. Body length 2.7 mm; fore wing length 2.7 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 2.00; antennal flagellomere 2 length/width 3.57; OOD/POD 1.25–1.38; IOD/POD 1.88.

MESOSOMA. Anteromesoscutum sculpturing with only very shallow punctures, reasonably smooth; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; 10–11 pits in scutellar sulcus; propodeum with median carina absent, only very shallow punctures associated with setae.

WINGS. Pterostigma length 0.71 mm; pterostigma width 0.29 mm; r 0.18 mm; 2RS 0.15 mm; 2m 0.11 mm; (RS+M)b 0.13 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 smooth and shiny; T1 length 0.41 mm; T1 width at posterior edge 0.08 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.17 mm; T2 width at posterior edge 0.28 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles andamookaensis sp. nov. constitutes BIN: BOLD:ADL4705 and is 6.41% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL3094; *Glyptapanteles aspersus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 6.5% different from the most similar *COI* sequence from an Australian specimen (AUMIC374-18; *Glyptapanteles aspersus* sp. nov.). The two available specimens, both sequenced for the *wingless* gene, share a unique barcode, that differs by a minimum of 1 bp from all other species with available sequence data.

Distribution

This species is known only from Andamooka Station in the central arid region of SA.

Glyptapanteles arcanus Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:FC2D5B3D-1AD2-478C-8920-8706F1BD594F](https://zoobank.org/act:FC2D5B3D-1AD2-478C-8920-8706F1BD594F)

Fig. 19

Diagnosis

Glyptapanteles arcanus sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the species group (other than from *G. vergrandiacus* sp. nov.) by the propodeum with having

coarser sculpturing than all but *G. lambkinae* sp. nov. and *G. erucadesolator* sp. nov. and with the T1 less strongly sculptured than in *G. lambkinae* sp. nov. and *G. erucadesolator* sp. nov. It is noted, however, that all these species (along with *G. goodwinnoakes* sp. nov., which has T1 smooth) are morphologically very similar and identifications should be made by comparison of DNA barcodes. There was not a morphological character found to easily differentiate *G. arcanus* sp. nov. from *G. vergrandiacus* sp. nov., which are sister lineages in the current phylogeny (Fig. 2), but these two species differ in the *wingless* barcode by a single base pair and can also be separated by the *COI* barcodes, which have a >3% divergence.

Etymology

The species epithet ‘*arcanus*’ is a Latin adjective meaning ‘mysterious’ and refers to the cryptic nature of this species’ morphology, which has few distinctive features.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Lamington National Park; -28.207, 153.137; 560 m a.s.l.; 9–19 Mar. 2007; C. Lambkin and N. Starick leg.; IBISCA Plot # IQ-500-A rainforest Malaise trap, bulk vial 22028; Extraction1462, BOLD: AUGLY082-21; QM T250938.

Paratype

AUSTRALIA • 1 ♀; Queensland, Lamington National Park; -28.155, 153.139; 282 m a.s.l.; 13–23 Jan. 2007; C. Lambkin and N. Starick leg.; IBISCA Plot # IQ-300-B rainforest Malaise trap; Extraction641, BOLD: AUMIC423-18; QM T208391.

Description

Female

COLOURATION. Gena without a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres all black/dark brown; tegula light brown; wing veins uniformly reddish-brown; anteromesoscutum dark; scutellar disk and metanotum dark with red tinge; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur light brown; hind femur dark reddish-brown or orange to light brown; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark reddish-brown, dark area extending past indentation to non-sclerotised area, but T2 lateral area then pale; T3 mostly dark with paler lateral areas; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.3 mm; fore wing length 2.3 mm; antennal length similar to body length.

HEAD. Antennal flagellomere 14 length/width 1.75–2.16; antennal flagellomere 2 length/width 3.57–3.71; OOD/POD 1.86–2.17; IOD/POD 1.14–1.50.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size, often smoother in posterior centre; scutellar disk sculpturing with shallow punctures scattered over most of area; 8–9 pits in scutellar sulcus. Propodeum with median carina absent, propodeum coarsely rugose.

WINGS. Pterostigma length 0.59 mm; pterostigma width 0.12 mm; r 0.17 mm; 2RS 0.12 mm; 2m 0.1 mm; (RS+M)b 0.08 mm.

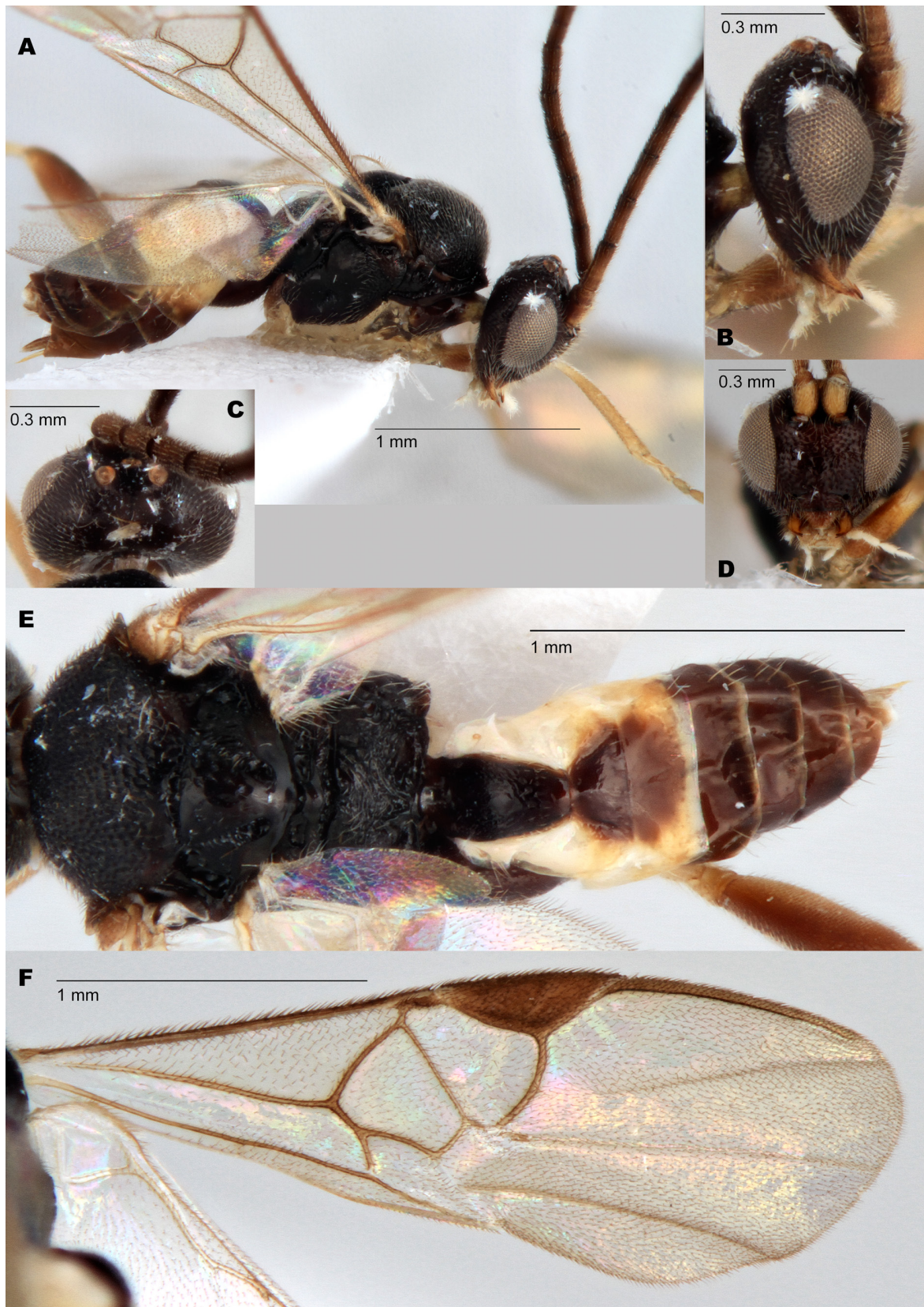


Fig. 19. *Glyptapanteles arcanus* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T208391). A. Lateral habitus. B. Lateral head. C. Dorsal head. D. Anterior head. E. Dorsal habitus. F. Fore wing.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly or broadest at centre of length, narrowing both posteriorly and anteriorly from mid-point; T1 smooth and shiny, some shallow scattered punctures on lateral edges and posterior half or indistinct sculpturing for most of length; T1 length 0.44 mm; T1 width at posterior edge 0.1 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.12 mm; T2 width at posterior edge 0.28 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

The *wingless* barcode for this species is shared with *G. doreyi* sp. nov. (one haplotype, as there are two *wingless* haplotypes present within *G. doreyi* sp. nov.) and *G. goodwinnoakes* sp. nov. and differs by 1 bp from the barcodes of *G. vergrandiacus* sp. nov., *G. doreyi* sp. nov. (second haplotype), and *G. lessardi* sp. nov. This group of species are very closely related and would benefit from future, more detailed taxonomic work to ensure that they are not simply divergent populations of the same species. For this species hypothesis, we have made the decision to split it into different species based on the *COI* barcode sequences being >3% divergent; however, we note that this is a hypothesis and is open to change with future work.

Glyptapanteles arcanus sp. nov. constitutes BIN BOLD:ADL5262 and is 3.67% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL3012; *Glyptapanteles vergrandiacus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 3.5% different from the most similar *COI* sequence from an Australian specimen (AUGLY102-21; *Glyptapanteles vergrandiacus* sp. nov.).

Distribution

This species is known only from Lamington National Park in southern QLD.

Glyptapanteles artonae (Rohwer, 1926)

Diagnosis

As Austin & Dangerfield (1992) state that this species is unlikely to actually occur in the Australasian region and we were unable to find morphological characters that definitively separate this species from those in Australia, we do not diagnose it. However, the extremely dark femur of this species would distinguish it from most new species described in our study. There is a very low risk of this species being conspecific with any of the newly described species from Australia, due to it being unlikely to actually occur in the Australasian region.

Material examined

Holotype

MALAYSIA • ♀; “Kuala Lumpur F.M.S.”, “From B.A.R. Gater”, “Pars. On *Artona catoxantha*”; USNM 40097. Only images examined (available at the NMNH type database: <http://n2t.net/ark:/65665/30bd958a4-dc6a-446d-8279-4168f0a8e0cc>).

Remarks

See Shenefelt (1972) for a compilation of extra-limital host records (some potentially incorrect).

Distribution (in the Australasian region)

Fiji (also peninsular Malaysia and Java). Fullaway (1957) is the only record of this species occurring in Fiji. However, Austin & Dangerfield (1992) were unable to find any such material in world collections and stated that this locality record may be based on a misidentification and the species may not occur in the Australasian region.

Glyptapanteles aspersus Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:956B5E0B-3F2A-408A-AAAE-263C197BC3DE](https://zoobank.org/act:956B5E0B-3F2A-408A-AAAE-263C197BC3DE)

Fig. 20

Diagnosis

Glyptapanteles aspersus sp. nov. is in the *G. albigena* species group and can be separated from most members of the species group by the pale spot on the gena being small but clearly visible, T2 dark, the media carina completely absent and the hind femur light brown to pale.

With the limited specimens available, there was not a morphological character found that easily differentiates *G. aspersus* sp. nov. from *G. kittelae* sp. nov. or *G. austrinus* sp. nov. The COI divergence between *G. aspersus* sp. nov. and *G. austrinus* sp. nov. is > 7%, whilst the divergence between *G. aspersus* sp. nov. and *G. kittelae* sp. nov. is also > 7%, which are considered large divergences at the species level for this group of wasps. The *wingless* sequences of *G. aspersus* sp. nov. and *G. kittelae* sp. nov. are very distinct, differing by 7 bp; however, there are no *wingless* sequences available for *G. austrinus* sp. nov.

Etymology

The species epithet '*aspersus*' is an adjective, from the Latin for 'scattered' or 'sprinkled' and refers to the widely dispersed distribution of this species.

Material examined**Holotype**

AUSTRALIA • ♀; Western Australia, Watheroo National Park, Jingemia Caves; -30.2542, 115.999; 273 m a.s.l.; 17 Sep.–7 Nov. 2003; C. Lambkin, N. Starick and J. Recsei leg.; Malaise closed heath; Extraction684, BOLD: AUMIC450-18; WAM E109885 (previously ANIC 32 130212).

Paratypes

AUSTRALIA – **South Australia** • 1 ♀; Belair National Park Gate 9; -35.009, 138.654; 25. Nov–1 Dec. 2007; J.T. Jennings leg.; Malaise trap; Extraction59, BOLD: AUMIC374-18; WAM E109886 • 1 ♀; Bibaringa, Wistow; -35.112, 138.887; Jan.–Feb. 2008; A. Austin leg.; Malaise trap; Extraction73, BOLD: AUMIC482-18; SAMA 32-45045. – **Victoria** • 1 ♂; Mt Macedon; -37.40348, 144.57237; 529 m a.s.l.; 14 Dec. 2019; J.B. Dorey leg.; general sweep of grass and flowering weeds in open area in schlerophyll, many gums flowering, not overly visited, sunny and overcast ~18°C; Extraction1153, BOLD: AUGLY064-21; SAMA 32-45046.

Description**Female**

COLOURATION. Gena with a pale spot; labrum pale or reddish-brown; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally or uniformly reddish-brown; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa dark; fore femur pale yellow or orange to light brown; mid femur pale yellow or orange to light brown; hind femur orange to light brown or dark; fore tibia pale yellow or orange to light

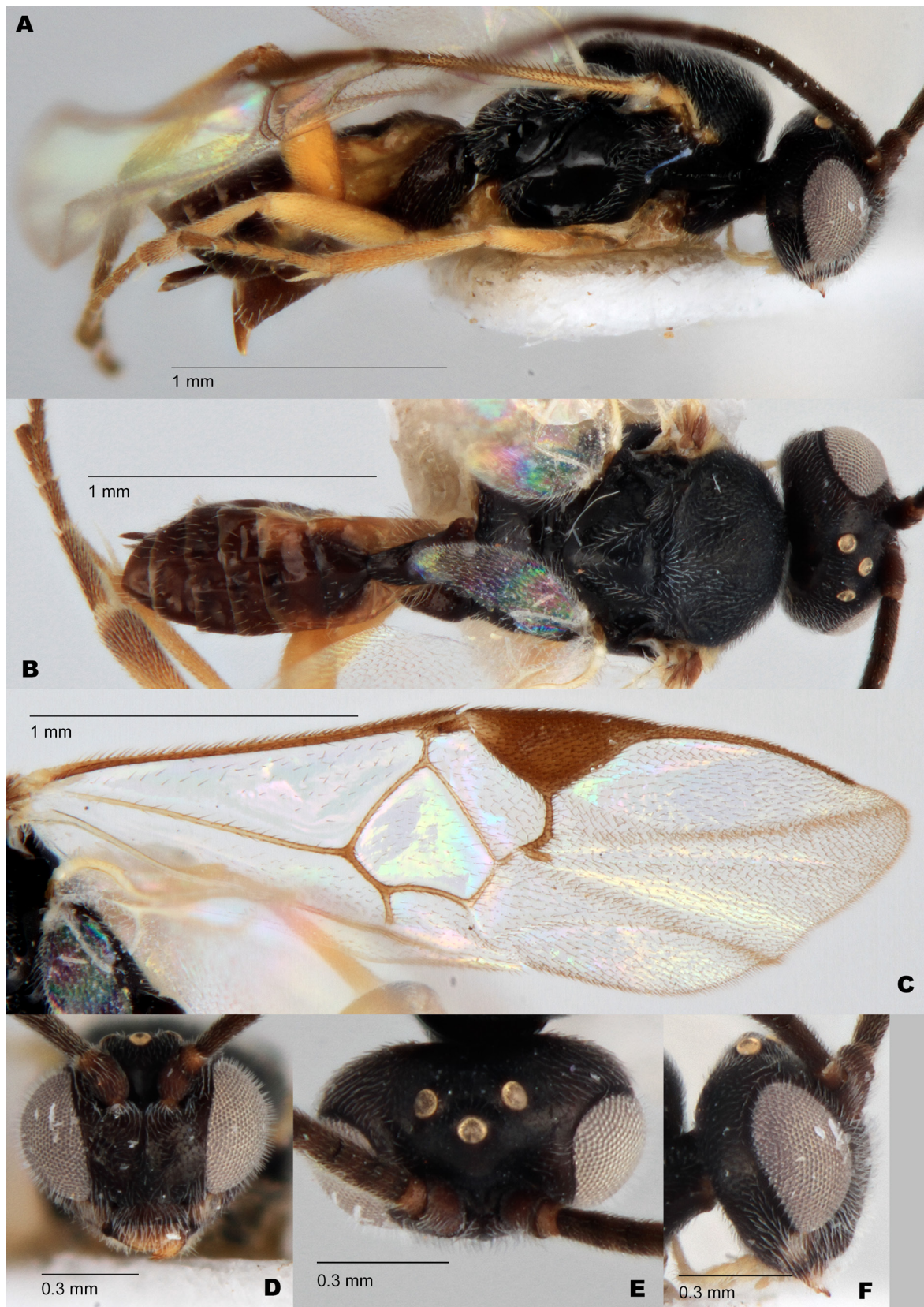


Fig. 20. *Glyptapanteles aspersus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (WAM 32 130212). A. Lateral habitus. B. Dorsal habitus. C. Fore wing. D. Anterior head. E. Dorsal head. F. Lateral head.

brown; mid tibia pale yellow, light brown or orange to light brown; hind tibia darkening posteriorly; hind basitarsus light brown or dark; T1 dark; T2 sclerotised area dark or dark reddish-brown, dark area extending past indentation to non-sclerotised area, but T2 lateral area then pale; T3 dark or mostly dark with paler lateral areas; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.2 mm; fore wing length 2.2 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 14 length/width 2.00–2.20; antennal flagellomere 2 length/width 3.00–4.66; OOD/POD 1.71–2.00; IOD/POD 2.00.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures; scutellar disk sculpturing with very shallow punctures scattered over most of area; 7–9 pits in scutellar sulcus; propodeum with median carina absent, very smooth and shiny, with either only very shallow punctures associated with setae or punctures over most of area and some shallow rugosity in posterior corners.

WINGS. Pterostigma length 0.54 mm; pterostigma width 0.22 mm; r 0.14 mm; 2RS 0.07 mm; 2m 0.08 mm; (RS+M)b 0.1 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 smooth and shiny, some shallow scattered punctures on lateral edges; T1 length 0.38 mm; T1 width at posterior edge 0.08 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.13 mm; T2 width at posterior edge 0.27 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, but antenna slightly longer, six pits in scutellar sulcus.

Remarks

Glyptapanteles aspersus sp. nov. constitutes BIN: BOLD:ADL3094 and is 5.71% (p-dist.) divergent from the closet BIN in the database (BOLD:ACL9711; *Glyptapanteles* sp. from New Zealand).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 6.6% different from the most similar *COI* sequence from an Australian specimen (AUMIC021-18; *Glyptapanteles harveyi* sp. nov.). The holotype was able to be sequenced for the *wingless* gene, which differs by a minimum of 7 bp from all other species with available sequence data.

Distribution

This species has a wide distribution, from VIC through to SA across to western WA.

Glyptapanteles aucklandensis (Cameron, 1909)

Fig. 21

Diagnosis

Glyptapanteles aucklandensis was described from a single male specimen that is not in excellent condition; however, it can be separated from many of the Australian species as follows: from the *G. albigena* species group by the absence of a pale gena spot, from the *mouldsi* and *eburneus* species groups by having T1–2 dark and from the *niveus* species group by having a relatively smooth anteromesoscutum. With only the one male type available for examination, we cannot conclusively differentiate *G. aucklandensis* from other Australian species, although we note that it has a large number of pits in the scutellar sulcus



Fig. 21. *Glyptapanteles aucklandensis* holotype, ♂ (NHMUK 3.c998). **A.** Fore wing. **B.** Lateral head and mesosoma. **C.** Dorsal head and mesosoma. **D.** Propodeum and T1–2. Images © The Trustees of the Natural History Museum, London (CC-BY 4.0).

(>16) which, if consistent among the rest of the individuals in the species, would separate it easily. There is very little overlap between the Australian and New Zealand microgastrine fauna, with the *Glyptapanteles* from New Zealand on BOLD forming a distinct clade (Fig. 1), so we feel there is little risk that this species is conspecific with any of the newly described ones from Australia.

Material examined

Holotype

NEW ZEALAND • ♂; Auckland; “P. Cameron Coll 191?-110”; NHMUK 3.c998 (only images examined).

Remarks

Host unknown.

Distribution

New Zealand (Auckland Is).

Glyptapanteles austini Fagan-Jeffries & Bird sp. nov.

[urn:lsid:zoobank.org:act:7DAEC136-31EE-4EC2-B197-66F9B686ACEE](https://zoobank.org/act:7DAEC136-31EE-4EC2-B197-66F9B686ACEE)

Figs 15B, 22

Diagnosis

Despite the large molecular divergence in both the *COI* and *wingless* genes (see Remarks section), the only morphological character found to differentiate *G. austini* sp. nov. from *G. guzikae* sp. nov. is the femur being completely dark (*G. guzikae* sp. nov. with the femur pale with a dark area posteriorly). *Glyptapanteles austini* sp. nov. can be separated from *G. kingae* sp. nov. as follows: *G. kingae* sp. nov. has the propodeum with stronger sculpturing in the centre, whilst *G. austini* sp. nov. has the propodeum smoother in the centre. However, we note that there are limited specimens available and the variability of propodeal sculpturing could broaden as more populations of these species are identified and we recommend confirming identifications with DNA barcodes.

Etymology

Named for Professor Andrew Austin, who has championed the documentation of Microgastrinae in Australia and has made a significant impact on the field of systematics of parasitic Hymenoptera, along with providing over a decade of mentorship and training to EPF-J.

Material examined

Holotype

AUSTRALIA • ♀; South Australia, Mt Barker Summit; -35.0661, 138.923; 20 Mar.–3 Apr. 2016; A. Austin and E. Fagan-Jeffries leg.; Malaise trap; Extraction343, BOLD: AUMIC198-18; SAMA 32-45047.

Paratypes

AUSTRALIA – **South Australia** • 1 ♀; same collection data as for holotype; 3–22 Apr. 2016; Extraction340, BOLD: AUMIC196-18; SAMA 32-45048 • 1 ♀ (ethanol); same collection data as for preceding; Extraction210, BOLD: AUMIC095-18; SAMA 32-45049 • 1 ♀ (ethanol); Belair National Park Gate 11; -35.009, 138.654; 11–24 Nov. 2007; J.T. Jennings leg.; Malaise trap; Extraction355, BOLD: AUMIC208-18; SAMA 32-45050 • 1 ♀; same collection data as for preceding; 25 Nov.–1 Dec. 2007; Extraction367, BOLD: AUMIC219-18; SAMA 32-46151 • 1 ♀; Millbrook Reservoir Gate 28, 25 km NE Adelaide; -34.8078, 138.827; 12 Sep. 2007; L. Farrington leg.; Malaise trap; Extraction469, BOLD: AUMIC291-18; SAMA 32-46152.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale or reddish-brown; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur orange to light brown; mid femur dark proximally and lightening distally; hind femur dark; fore tibia orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus dark reddish-brown; T1 dark; T2 sclerotised area dark or dark reddish-brown; T2 lateral area much paler; T3 mostly pale with darker patch in centre or mostly dark with paler lateral areas; T4+ dark.

BODY MEASUREMENTS. Body length 2.1 mm; fore wing length 2.0 mm; antennal length slightly shorter than body length.

HEAD. Face with fine punctures associated with setae; antennal flagellomere 14 length/width 1.83–2.50; antennal flagellomere 2 length/width 2.57–3.85; OOD/POD 1.83–2.00; IOD/POD 1.43–2.17.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures, slightly denser and deeper punctures anteriorly, smoother in posterior centre; scutellar disk sculpturing with only very shallow punctures; 7–9 pits in scutellar sulcus; propodeum with median carina absent, scattered punctures associated with setae in posterior half, rest of propodeum smooth and shiny.

WINGS. Pterostigma length 0.52 mm; pterostigma width 0.18 mm; r 0.12 mm; 2RS 0.11 mm; 2m 0.06 mm; (RS+M)b 0.09 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, or lateral edges parallel for anterior $\frac{1}{4}$ of length, then gently narrowing posteriorly, lateral edges either straight or with slight curve; T1 mostly smooth, some punctures in posterior half; T1 length 0.3 mm; T1 width at posterior edge 0.09 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.11 mm; T2 width at posterior edge 0.19 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles austini sp. nov. constitutes BIN BOLD:ADL2587 and is 7.69% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL2952; *Glyptapanteles guzikae* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 7.9% different from the most similar *COI* sequence from an Australian specimen (AUMIC524-18; *Glyptapanteles guzikae* sp. nov.). All five paratype specimens were sequenced for the *wingless* gene, which is identical amongst all specimens (although there is a single ambiguity in one sequence). The *wingless* sequences differ by a minimum of 16 bp from all other species with available sequence data.

Distribution

This species is currently known from SA, from Adelaide and close surrounds.

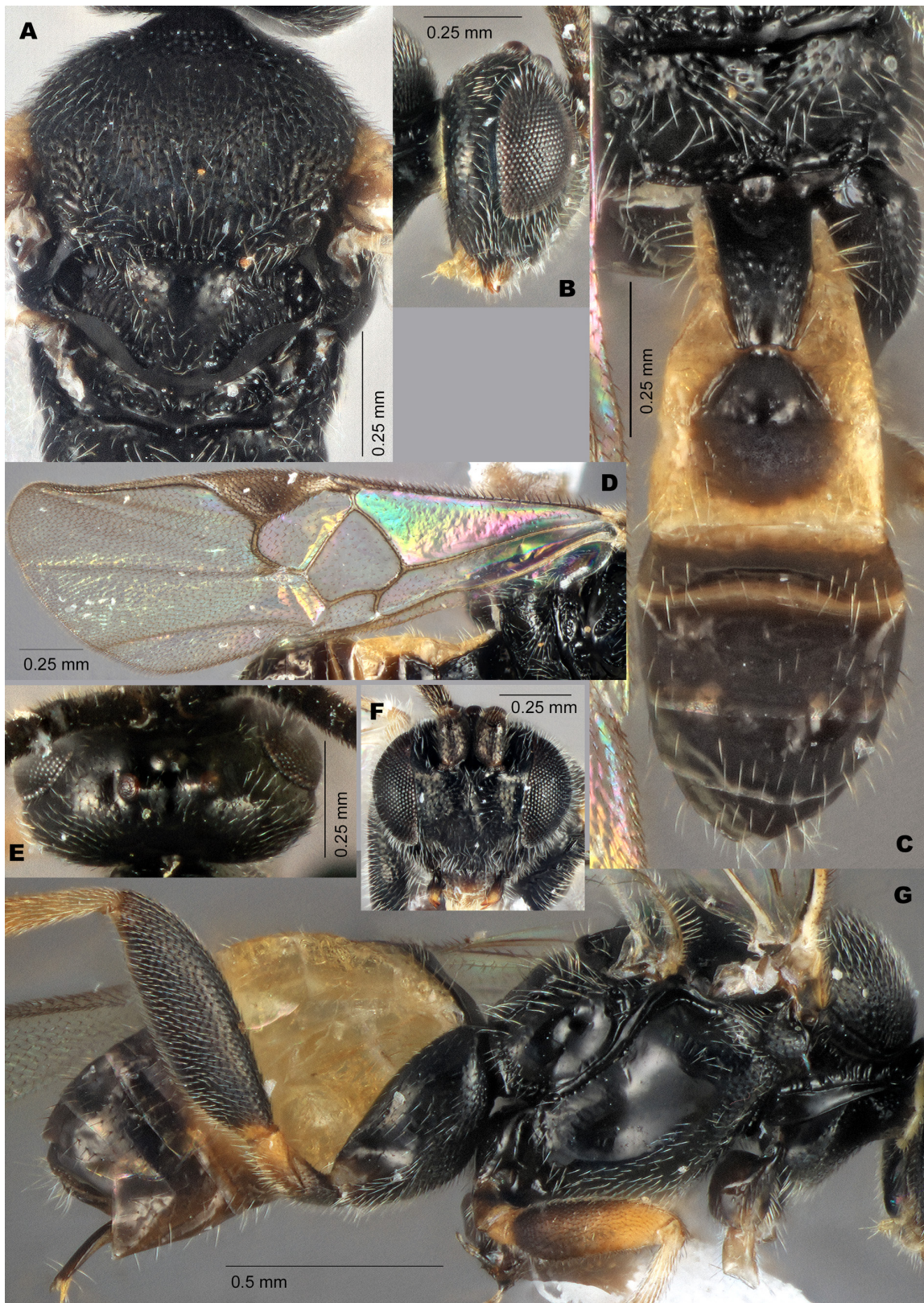


Fig. 22. *Glyptapanteles austini* Fagan-Jeffries & Bird sp. nov. **A, D–F.** Paratype, ♀ (SAMA 32-46151). **B–C, G.** Holotype, ♀ (SAMA 32-45047). **A.** Dorsal mesosoma. **B.** Lateral head. **C.** Dorsal metasoma. **D.** Fore wing. **E.** Dorsal head. **F.** Anterior head. **G.** Lateral habitus.

Glyptapanteles austrinus Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:28E8E832-7CC3-40A0-A29D-D7D799318C95](https://doi.org/10.28881/zoobank.org/act:28E8E832-7CC3-40A0-A29D-D7D799318C95)

Fig. 23

Diagnosis

Glyptapanteles austrinus sp. nov. is in the *G. albigena* species group and can be separated from most of the other members of the species group by the pale spot on the gena being small but clearly visible, T2 dark, the media carina completely absent, and the hind femur light brown to pale.

With the limited specimens available, there was not a morphological character found that easily differentiates *G. austrinus* sp. nov. from *G. kittelae* sp. nov. or *G. aspersus* sp. nov. The *COI* divergence between *G. austrinus* sp. nov. and *G. kittelae* sp. nov. is > 6%, whilst the divergence between *G. austrinus* sp. nov. and *G. aspersus* sp. nov. is > 8%, both of which are considerably large divergences at the species level for this subfamily of wasps. There are no available *wingless* sequences for *G. austrinus* sp. nov.

Etymology

The Latin adjective ‘*austrinus*’ means ‘southern’ and refers to the collection of this species near the southern tip of Australia, in Hobart, TAS.

Material examined

Holotype

AUSTRALIA • ♀; Tasmania, Hobart, Kingston Beach; -42.986, 147.317; 25–30 Apr. 2010; B. Ward leg.; BIOUG00995-H02, BOLD: HYAT371-11; TMAG F121367.

Paratype

AUSTRALIA • 1 ♀; same collection data as for holotype; BIOUG00996-B04, BOLD: HYAT396-11; TMAG F121368.

Description

Female

COLOURATION. Gena with a pale spot; labrum pale; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow or dark; mid coxa pale yellow or dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark reddish-brown; T2 sclerotised area dark reddish-brown; T2 lateral area same colour as sclerotised area, or only slightly paler or dark portion extends past indentation, but then pale; T3 uniformly brown; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.3 mm; fore wing length 2.4 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 14 length/width 1.22–1.50; antennal flagellomere 2 length/width 3.00–4.00; OOD/POD 2.00–2.17; IOD/POD 1.83.

MESOSOMA. Anteromesoscutum sculpturing with only very shallow punctures, reasonably smooth; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; 8–11 pits in scutellar sulcus; propodeum with median carina absent, only very shallow punctures associated with setae.

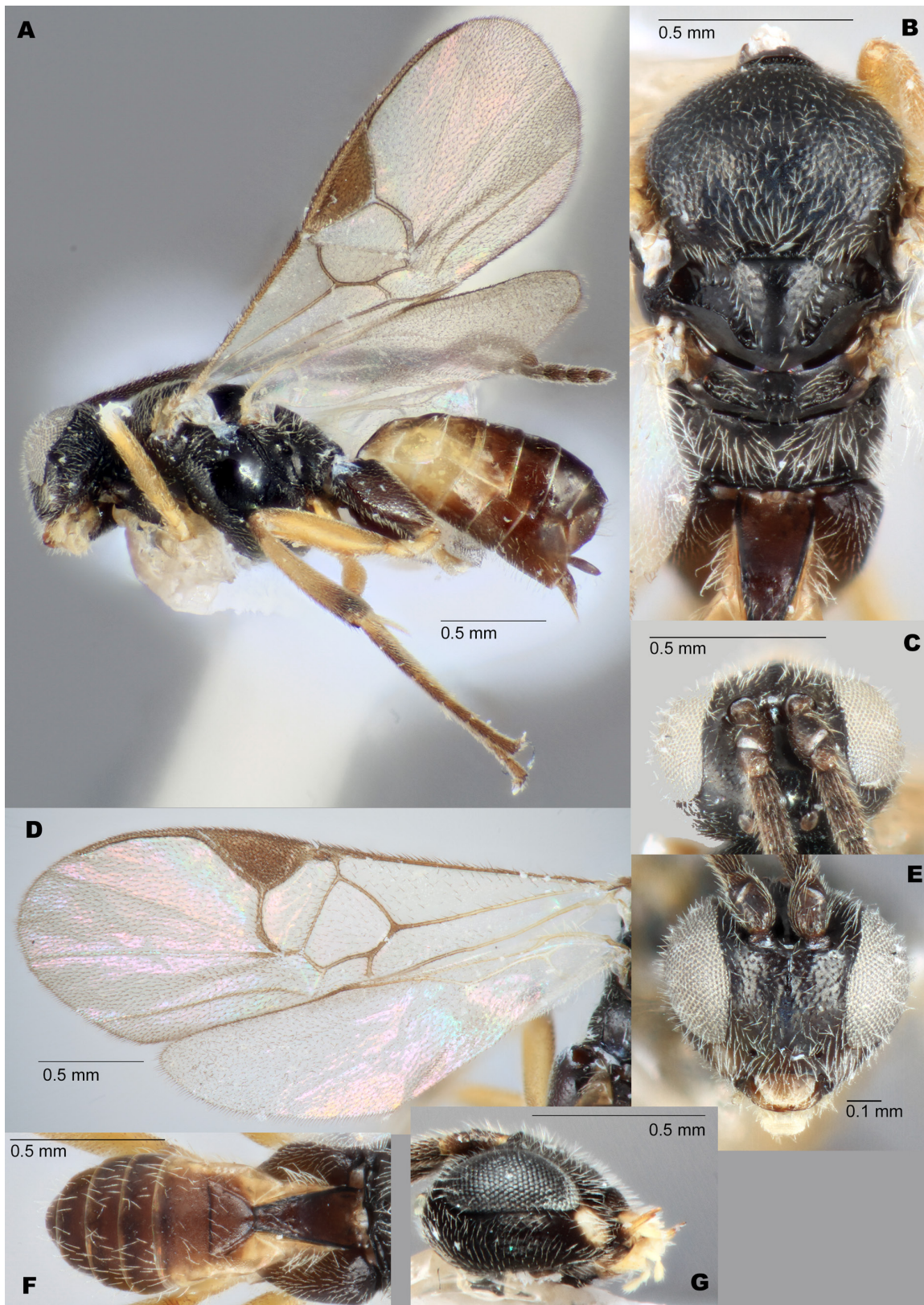


Fig. 23. *Glyptapanteles austrinus* Fagan-Jeffries, Bird & Austin sp. nov. **A, C–E.** Holotype, ♀ (TMAG F121367). **B, F–G.** Paratype, ♀ (TMAG F121368). **A.** Lateral habitus. **B.** Dorsal mesosoma. **C.** Dorsal head. **D.** Fore wing. **E.** Anterior head. **F.** Dorsal metasoma. **G.** Lateral head.

WINGS. Pterostigma length 0.57 mm; pterostigma width 0.27 mm; r 0.17 mm; 2RS 0.11 mm; 2m 0.12 mm; (RS+M)b 0.07 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 mostly smooth, some punctures in posterior half or smooth and shiny; T1 length 0.34 mm; T1 width at posterior edge 0.07 mm; T2 an isosceles trapezoid, lateral edges straight or with curved lateral and anterior edges, becoming arch- or semicircle-shaped; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.26 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles austrinus sp. nov. constitutes BIN BOLD:ABA6213 and is 4.17% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL4089; *Glyptapanteles ferrugineus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 4.1% different from the most similar *COI* sequence from an Australian specimen (AUMIC042-18; *Glyptapanteles ferrugineus* sp. nov.). The holotype was able to be sequenced for the *wingless* gene, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

This species is known only from Hobart in southern TAS.

Glyptapanteles baylessi Fagan-Jeffries, Bird & Austin sp. nov.

[urn:lsid:zoobank.org:act:D0EE87F6-0435-407D-AE00-11BBCAF4A184](https://zoobank.org/act:D0EE87F6-0435-407D-AE00-11BBCAF4A184)

Figs 5E, 12C, 24

Diagnosis

Glyptapanteles baylessi sp. nov. is closely related to the *G. niveus* species group, but can be differentiated by the deep punctures on the anteromesoscutum being denser (much less smooth space between punctures). *Glyptapanteles baylessi* sp. nov. has strong orange markings on the posterior anteromesoscutum that are significantly larger than in any of the other described species of *Glyptapanteles* in Australia.

Etymology

This species is named for Dr Keith Bayless, who collected the type series and also contributed a significant amount of critical additional material used in this study. We thank him for his continual willingness to share Malaise trap samples and his collegiate and open approach to taxonomic science.

Material examined

Holotype

AUSTRALIA • ♀; New South Wales, Royal National Park, near Waterfall Couranga Track near Hacking River; -34.1486, 151.0221; 40 m a.s.l.; 20 Jan.–6 Feb. 2020; K.M. Bayless and J.G. Lumbers leg.; 6 m Malaise trap over Waterfall Creek; Extraction852, BOLD: AUGLY004-21; ANIC 32 130346.

Paratypes

AUSTRALIA – New South Wales • 1 ♂; same collection data as for holotype; Extraction859, BOLD: AUGLY007-21; ANIC 32 130347 • 1 ♂; same collection data as for preceding; Extraction937, BOLD: AUGLY022-21; ANIC 32 130348 • 1 ♂; same collection data as for preceding; Extraction941, BOLD: AUGLY023-21; ANIC 32 130349 • 1 ♂; same collection data as for preceding; Extraction1117, BOLD:

AUGLY052-21; ANIC 32 130350 • 1 ♂; same collection data as for preceding; Extraction1114, BOLD: AUGLY049-21; ANIC 32 130351 • 1 ♂ (ethanol); same collection data as for preceding; Extraction1118, BOLD: AUGLY053-21; ANIC 32 130352 • 1 ♀; same collection data as for preceding; Extraction861, BOLD: AUGLY009-21; ANIC 32 130353 • 1 ♀; same collection data as for preceding; Extraction932, BOLD: AUGLY020-21; ANIC 32 130354 • 1 ♀; same collection data as for preceding; Extraction1024, BOLD: AUGLY037-21; ANIC 32 130355 • 1 ♀; same collection data as for preceding; Extraction1029, BOLD: AUGLY038-21; ANIC 32 130356 • 1 ♀ (ethanol); same collection data as for preceding; Extraction1116, BOLD: AUGLY051-21; ANIC 32 130357 • 1 ♀ (ethanol); same collection data as for preceding; Extraction1123, BOLD: AUGLY056-21; ANIC 32 130358 • 1 ♀ (ethanol); same collection data as for preceding; Extraction1127, BOLD: AUGLY058-21; ANIC 32 130359 • 1 ♀ (ethanol); same collection data as for preceding; Extraction1130, BOLD: AUGLY059-21; ANIC 32 130360 • 1 ♀; Mt Keira via Wollongong; -34.4035, 150.857; 1 Mar. 2004; A.D. Austin and M. Dowton leg.; Extraction43, BOLD: AUMIC264-18; AM K.517936.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with significant orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow or white; mid coxa pale yellow or white; hind coxa dark or dark lightening distally; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow or orange to light brown; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown, dark reddish-brown or dark; T1 dark; T2 sclerotised area dark; T2 lateral area same colour as sclerotised area, or only slightly paler or dark extends past indentation, but then pale; T3 uniformly brown; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.4 mm; fore wing length 2.4 mm; antennal length slightly longer than body length.

HEAD. Face densely sculptured, punctate reticulate; antennal flagellomere 14 length/width 1.63–2.25; antennal flagellomere 2 length/width 3.5–3.62; OOD/POD 1.13–1.71; IOD/POD 1.29–1.45.

MESOSOMA. Anteromesoscutum sculpturing with deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size; scutellar disk sculpturing with deep, irregularly spaced punctures; 6–8 pits in scutellar sulcus; propodeum with median carina difficult to discern amongst sculpturing, propodeum strongly rugose, median carina absent and propodeum strongly rugose, or median carina very faintly indicated at posterior and anterior edges, propodeum strongly rugose.

WINGS. Pterostigma length 0.64 mm; pterostigma width 0.18 mm; r 0.13 mm; 2RS 0.11 mm; 2m 0.11 mm; (RS+M)b 0.08 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{4}$ of length, then gently narrowing posteriorly, lateral edges often with slight curve; T1 smooth in anterior half, punctate reticulate in posterior half or smooth in anterior half, punctate reticulate to rugose in posterior half; T1 length 0.4 mm; T1 width at posterior edge 0.19 mm; T2 width greater than length, rectangular with rounded anterior corners; T2 with some shallow indistinct sculpturing or rugose; T2 length 0.13 mm; T2 width at posterior edge 0.36 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, 6–9 pits in scutellar sulcus.

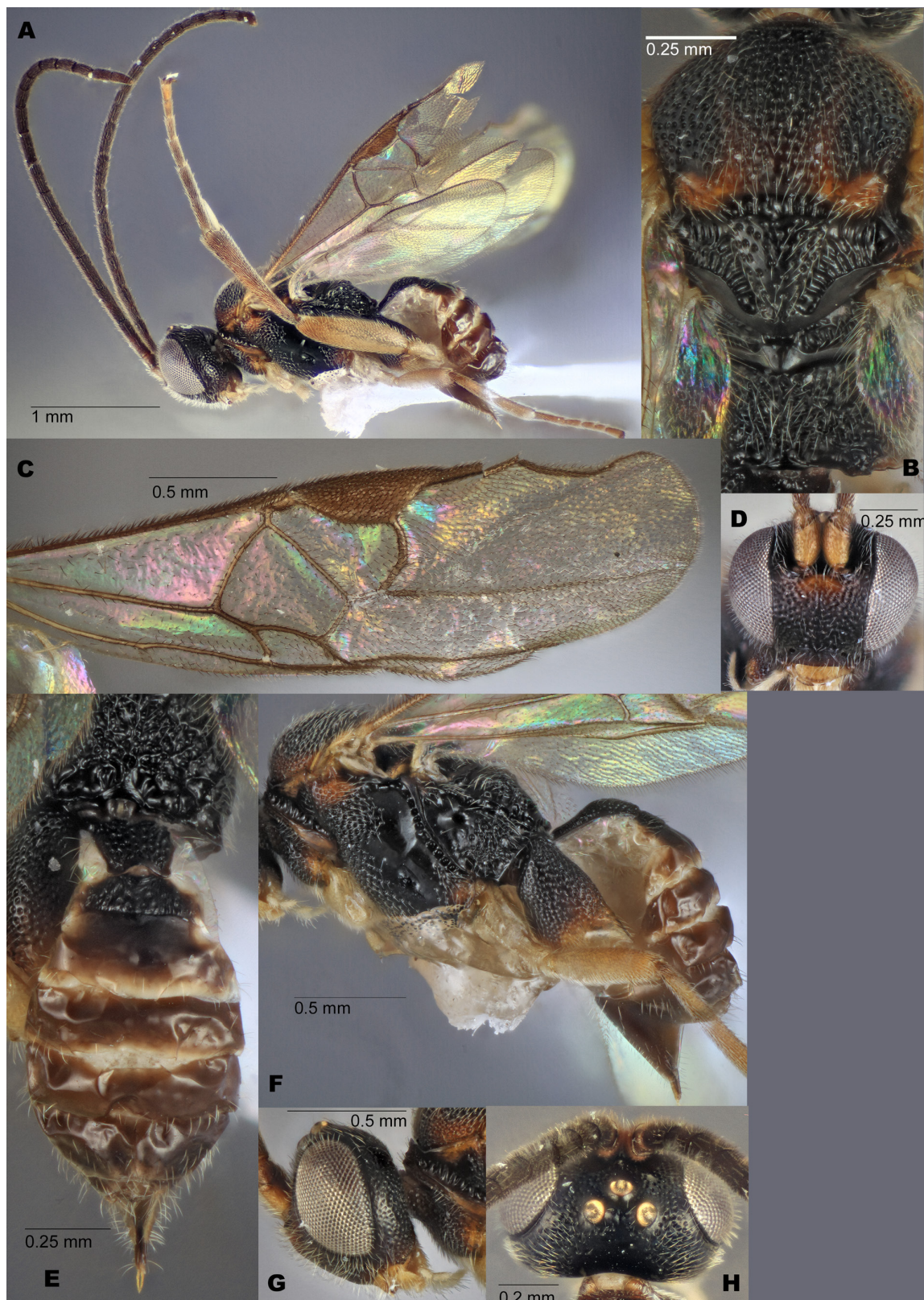


Fig. 24. *Glyptapanteles baylessi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (AM K.517936). A. Lateral habitus. B. Dorsal mesosoma. C. Fore wing. D. Anterior head. E. Dorsal metasoma. F. Lateral body. G. Lateral head. H. Dorsal head.

Remarks

Glyptapanteles baylessi sp. nov. constitutes BIN BOLD:ADL4626 and is 7.98% (p-dist.) divergent from the closet BIN in the database (BOLD:ADD5918, a BIN consisting of one specimen from Papua New Guinea).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 8.3% different from the most similar *COI* sequence from an Australian specimen (AUGLY096-21; *Glyptapanteles niveus* sp. nov.). Sixteen specimens of the type series were sequenced for the *wingless* gene, which is identical across all specimens and differs by a minimum of 6 bp from all other species with available sequence data.

Distribution

This species is currently known from national parks south of Sydney, NSW.

Glyptapanteles bradfordae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:6C1A43F3-D6F0-4FD3-A100-F83B4EB894E8](https://zoobank.org/act:6C1A43F3-D6F0-4FD3-A100-F83B4EB894E8)
 Figs 12A, 25

Diagnosis

Glyptapanteles bradfordae sp. nov. is in the *G. niveus* species group and can be differentiated from *G. niveus* sp. nov. by the absence of white distal flagellomeres, a pale labrum and denser punctures on the hind coxa. *Glyptapanteles bradfordae* sp. nov. can be separated from *G. cooperi* sp. nov. by having denser punctures on the hind coxa, particularly in the dorsal area.

Etymology

This species is named for Dr Tessa Bradford, Facility Manager of the South Australian Regional Facility for Molecular Ecology and Evolution, without whom the DNA sequencing of many of these specimens would not have occurred. The authors thank Tessa for her continual advice, support and training in the molecular labs!

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Kuranda; -16.8135, 145.6430586; 317 m a.s.l.; 14 Dec. 2019–6 Jan. 2020; M.S. Moulds leg.; Malaise Trap EFJ2020MT11; Extraction1096, BOLD: AUGLY046-21; QM T250939.

Paratypes

AUSTRALIA • 1 ♂ (ethanol); same collection data as for holotype; 19 May–8 Jul. 2017; Extraction1295, BOLD: AUGLY078-21; QM T250940 • 1 ♂; same collection data as for holotype; Extraction1297, BOLD: AUGLY079-21; QM T250941 • 1 ♂ (ethanol); same collection data as for holotype; 12 Feb.–6 Apr. 2020; EFJ2020MT36; Extraction1524, BOLD: AUGLY094-21; QM T250942 • 1 ♂ (ethanol); same collection data as for holotype; 1–21 Jan. 2017; Extraction292, BOLD: AUMIC157-18; QM T250943 • 1 ♀; same collection data as for holotype; Extraction290, BOLD: AUMIC155-18; QM T250944 • 1 ♀; same collection data as for holotype; 12 Feb.–6 Apr. 2020; Extraction1544, BOLD: AUGLY100-21; QM T250945 • 1 ♂; Lamington National Park; -28.207, 153.137; 560 m a.s.l.; 9–19 Mar. 2007; C.7 Lambkin and N. Starick leg.; IBISCA Plot # IQ-500-A rainforest Malaise trap, bulk vial 22028; Extraction1461, BOLD: AUGLY081-21; QM T250946 • 1 ♀; same collection data as for preceding; Extraction633, BOLD: AUMIC417-18; QM T208409.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale; scape colour in ventral half uniformly paler than flagellomeres or the same colour or darker than flagellomeres; flagellomeres all black/dark brown or uniformly reddish-brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with significant orange patches on posterolateral corners; scutellar disk and metanotum dark or dark with red tinge; propodeum dark; fore coxa white; mid coxa white; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark or dark reddish-brown; T2 sclerotised area dark or dark reddish-brown; T2 lateral area much paler; T3 dark or mostly dark with paler lateral areas; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 1.8 mm; fore wing length 2.3 mm; antennal length slightly longer than body length.

HEAD. Face densely sculptured, punctate reticulate; antennal flagellomere 14 length/width 1.85–2.00; antennal flagellomere 2 length/width 3.63–4.50; OOD/POD 1.33–1.71; IOD/POD 1.33–1.71. Eyes particularly large, with a smaller malar space than most species of *Glyptapanteles*.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures, slightly denser and deeper punctures anteriorly, smoother in posterior centre or very sparse, deep punctures; scutellar disk sculpturing smooth in centre, with some deep punctures on lateral edges; six pits in scutellar sulcus; propodeum with median carina very faintly indicated at either anterior or both posterior and anterior edges, propodeum strongly rugose.

WINGS. Pterostigma length 0.57 mm; pterostigma width 0.18 mm; r 0.2 mm; 2RS 0.1 mm; 2m 0.12 mm; (RS+M)b 0.09 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly or broadest at centre of length, narrowing both posteriorly and anteriorly from mid-point; T1 smooth in anterior half, punctate reticulate to rugose in posterior half; T1 length 0.38 mm; T1 width at posterior edge 0.13 mm; T2 an isosceles trapezoid, lateral edges straight or almost square, lateral edges almost parallel, only broadening posteriorly very slightly; T2 with some shallow indistinct sculpturing; T2 length 0.12 mm; T2 width at posterior edge 0.22 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, 6–7 pits in scutellar sulcus.

Remarks

Glyptapanteles bradfordae sp. nov. constitutes BIN BOLD:AEI7940 and is 2.01% (p-dist.) divergent from the closet BIN in the database (BOLD:ADD8337, with three specimens from Papua New Guinea, see below).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 7.6% different from the most similar *COI* sequence from an Australian specimen (GMAQJ131-17; an undescribed lineage, with two specimens). Eight specimens were able to be sequenced for the *wingless* gene, which all share a unique barcode that differs by a minimum of 3 bp from all other species with available sequence data.

There are three additional specimens sequenced for *COI* on BOLD which fall within 2% pairwise divergence and are in the closely related BIN BOLD:ADD8337. These specimens (GMNGH687-16,

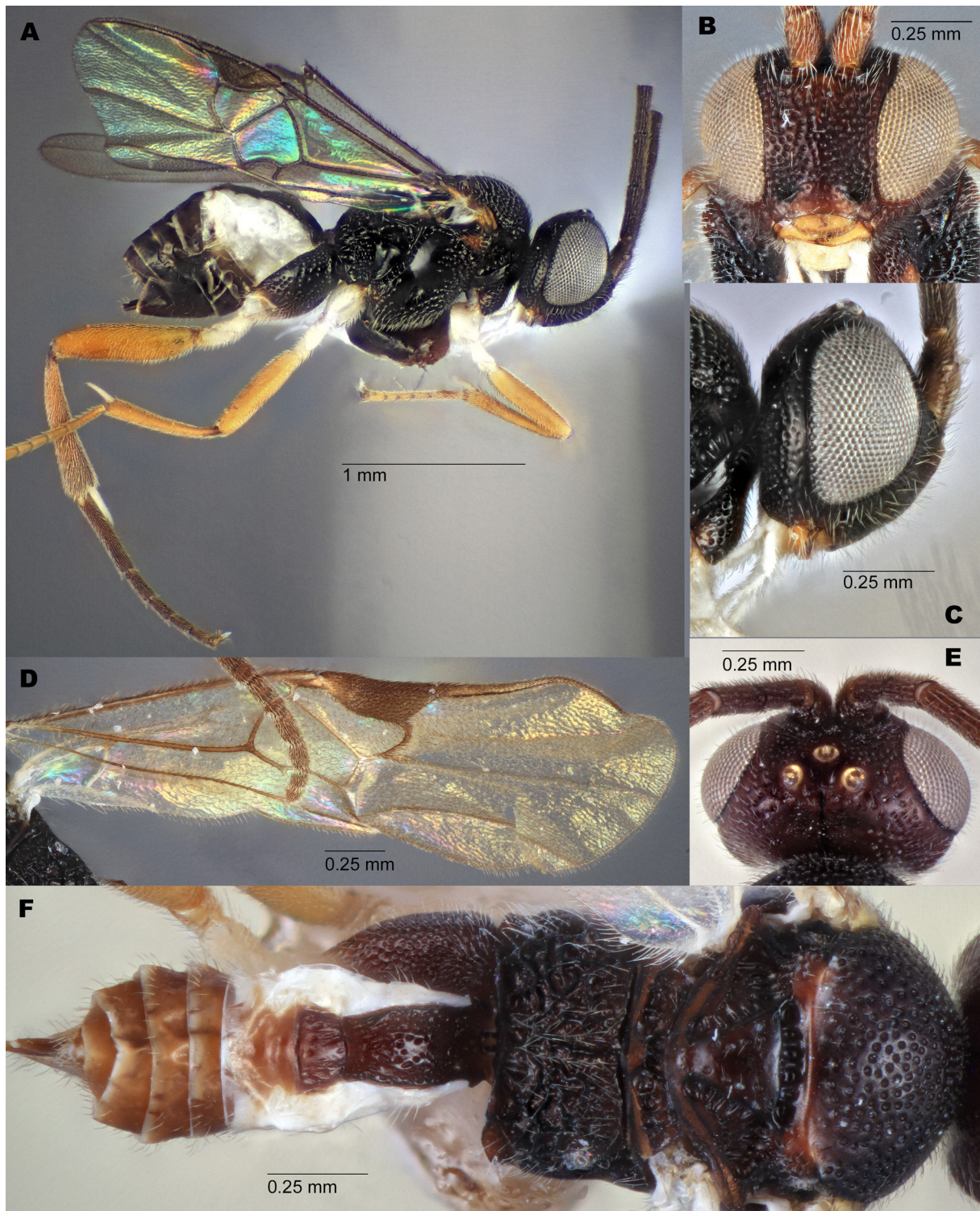


Fig. 25. *Glyptapanteles bradfordae* Fagan-Jeffries, Bird & Austin sp. nov., paratypes, ♀. **A, C–D, F.** QM T250944. **B, E.** QM T208409. **A.** Lateral habitus. **B.** Anterior head. **C.** Lateral head. **D.** Fore wing. **E.** Dorsal head. **F.** Dorsal habitus.

GMNGJ510-16 and GMNGS061-16) were all collected in Papua New Guinea, Madang, by Pagi Toko and are most likely the same species. Specimens were not examined, but a dorsal habitus image on BOLD (<http://www.boldsystems.org/pics/GMNGS/BIOUG30657-C01%2B1487790120.jpg>) supports this suggestion.

Distribution

This species is currently known from the east coast of QLD, from both Lamington National Park in the south and Kuranda in the north. It is also likely found in Papua New Guinea (based on the presence of closely related *COI* barcodes on BOLD).

Glyptapanteles cooperi Fagan-Jeffries, Bird & Austin sp. nov.
urn:lsid:zoobank.org:act:8E2A7ED6-8383-43C1-B1DA-2712F7B6529C
Figs 12B, 26

Diagnosis

Glyptapanteles cooperi sp. nov. is in the *G. niveus* species group and can be separated from *G. bradfordae* sp. nov. by the hind coxa of *G. cooperi* sp. nov. only being sparsely punctured, particularly on the dorsal area and from *G. niveus* sp. nov. by the labrum being pale in colour (labrum dark in *G. niveus* sp. nov.).

Etymology

Named for Professor Steven Cooper, who has provided extensive mentoring to EPF-J during her PhD.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Kuranda; -16.8154, 145.643; 317 m a.s.l.; 16 Mar.–12 Apr. 2017; M.S. Moulds leg.; Malaise Trap; Extraction548, BOLD: AUMIC340-18; QM T250947.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale; scape colour in ventral half paler than flagellomeres at proximal end; flagellomeres all black/dark brown; tegula dark; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa white; mid coxa white; hind coxa dark; hind femur orange to light brown; hind tibia light brown; hind basitarsus light brown; T1 dark; T2 sclerotised area dark; T2 lateral area same colour as sclerotised area, or only slightly paler; T3 mostly dark with paler lateral areas; T4+ dark.

HOLOTYPE BODY MEASUREMENTS. Body length 2.3 mm; fore wing length 2.2 mm.

HEAD. Face densely sculptured, punctate reticulate; antennal flagellomere 2 length/width 3.63; OOD/POD 1.50; IOD/POD 1.50.

MESOSOMA. Anteromesoscutum with very sparse, deep punctures; scutellar disk smooth in centre, some deep punctures on lateral edges; six pits in scutellar sulcus; propodeum with median carina present and complete, rest of propodeum strongly rugose.

WINGS. Pterostigma length 0.58 mm; pterostigma width 0.17 mm; r 0.15 mm; 2RS 0.1 mm; 2m 0.11 mm; (RS+M)b 0.08 mm.

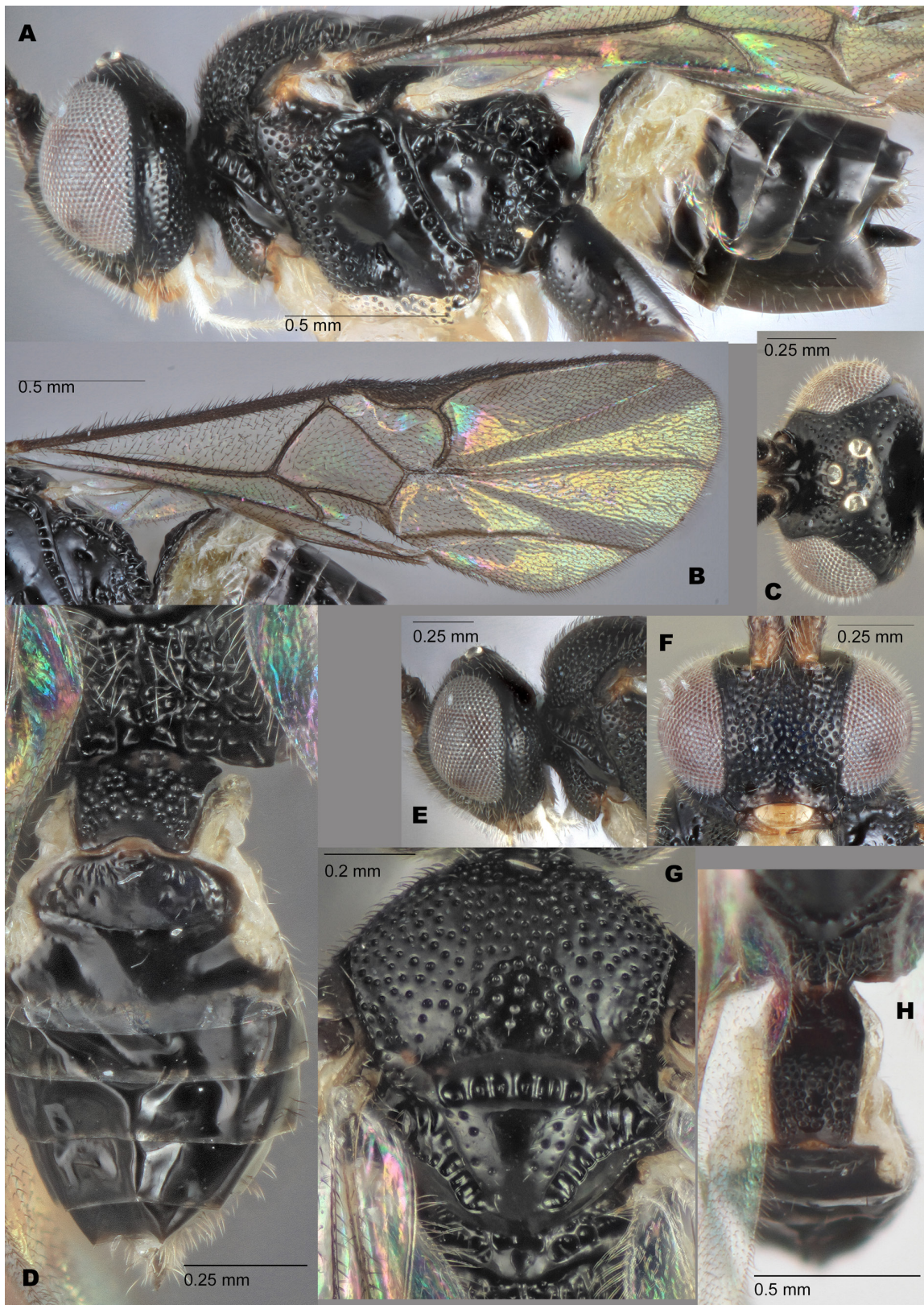


Fig. 26. *Glyptapanteles cooperi* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250947). A. Lateral body. B. Fore wing. C. Dorsal head. D. Dorsal propodeum and metasoma. E. Lateral head. F. Anterior head. G. Dorsal mesosoma. H. Dorsal metasoma.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{4}$ of length, then gently narrowing posteriorly, lateral edges often with slight curve; T1 smooth in anterior half, punctate reticulate in posterior half; T1 length 0.5 mm; T1 width at posterior edge 0.21 mm; T2 width greater than length, rectangular with rounded anterior corners; T2 smooth and shiny; T2 length 0.15 mm; T2 width at posterior edge 0.36 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles cooperi sp. nov. constitutes BIN: BOLD:ADL4822 and is 6.98% (p-dist.) divergent from the closet BIN in the database (BOLD:ADG5990, an undescribed lineage from Australia, with two specimens).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 6.8% different from the most similar *COI* sequence from an Australian specimen (GMQQW352-18; an undescribed lineage, with two specimens). The type specimen was able to be sequenced for the *wingless* gene, which differs by a minimum of 4 bp from all other species with available sequence data.

Distribution

This species is currently only represented by a single specimen from Kuranda, northern QLD.

Glyptapanteles deliasa Austin & Dangerfield, 1992
Figs 13A, 27

Diagnosis

Glyptapanteles deliasa can be separated from the other described species of *Glyptapanteles* in Australia by the combination of the following characters: gena without a pale spot, T1 and T2 dark, anteromesoscutum very smooth, with only shallow and regular punctures, no orange markings on the postero-lateral anteromesoscutum, propodeum very smooth with only shallow punctures and the median carina faintly indicated, T1 parallel-sided for most of length, only narrowing/curving inwards right at boundary with T3, ventral side of antennal scape (at least in distal half) darker than flagellomeres, tegula dark.

Material examined

Paratypes

AUSTRALIA • 2 ♀♀; South Australia, Waikerie; 22 Jun. 1988; M.D. Moore leg.; ex *Delias aganippe* (Donovan) (Pieridae); WINC.

Remarks

There is a possibility that this species is conspecific with *Cotesia deliadis* (Bingham, 1906), as it is morphologically very similar and parasitises the same host. However, several attempts to extract DNA from paratype specimens to either confirm or refute this hypothesis have been unsuccessful. As such, we leave it within the genus *Glyptapanteles* until next-generation sequencing techniques can be used on the type series.

Distribution

Known from Australia, currently only from SA.

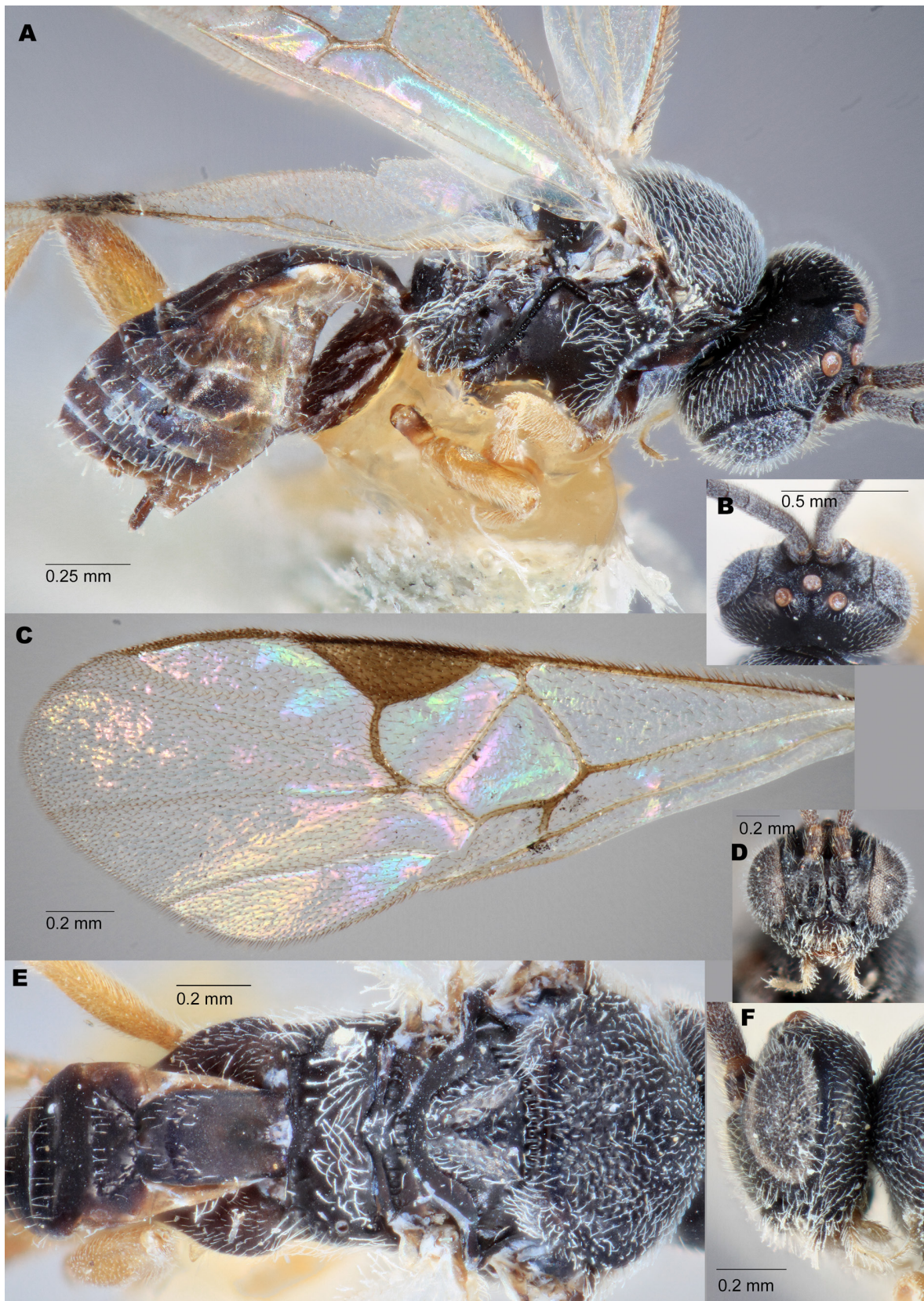


Fig. 27. *Glyptapanteles deliasa* Austin & Dangerfield, 1992, paratype, ♀ (WINC). **A.** Lateral habitus. **B.** Dorsal head. **C.** Fore wing. **D.** Anterior head. **E.** Dorsal habitus. **F.** Lateral head.

Glyptapanteles doreyi Fagan-Jeffries, Bird & Austin sp. nov.
urn:lsid:zoobank.org:act:34246DE7-8966-4FBC-8D6A-ECC5DADF79BC

Figs 28–29

Diagnosis

Glyptapanteles doreyi sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the species group as follows:

Glyptapanteles doreyi sp. nov. can be separated from *G. rodriguezae* sp. nov. and *G. ruhri* sp. nov. by T1 being smooth and shiny, not having punctures that cover at least a third of the area of the posterior half of the tergite.

Glyptapanteles doreyi sp. nov. can be separated from *G. goodwinnoakes* sp. nov., *G. erucadesolator* sp. nov., *G. lambkinae* sp. nov., *G. arcanus* sp. nov. and *G. vergrandiacus* sp. nov. by the propodeum being less coarsely and less consistently, rugose sculptured across the anterior half of the propodeum. *Glyptapanteles doreyi* sp. nov. has the propodeum with shallow or strong punctures in the anterior half, the posterior half smooth or with shallow or strong rugose sculpturing and sometimes with a smooth area in the centre.

Glyptapanteles doreyi sp. nov. can be separated from *G. wrightae* sp. nov. and *G. lessardi* sp. nov. by the tegula being dark in colouration, the same colour or only slightly lighter than the mesosoma (the tegula in *G. wrightae* sp. nov. and *G. lessardi* sp. nov. is pale) the hind femur mostly dark (pale or light brown in *G. wrightae* sp. nov. and *G. lessardi* sp. nov.) and the indentation in the centre of the mesopleuron being strongly canaliculate. The indentation on the mesopleuron of *G. wrightae* sp. nov. and *G. lessardi* sp. nov. is smoother, not strongly canaliculate.

Etymology

Named for James Dorey, who collected the holotype specimen. EPF-J would like to acknowledge James' collegiate sharing of specimens and his contribution to a successful field trip in northern QLD in 2019 that led to the collection of many important microgastrine specimens.

Material examined

Holotype

AUSTRALIA • ♀; New South Wales, Cockburn River Camp; -31.05222, 151.14411; 461 m a.s.l.; 10 Dec. 2019; J.B. Dorey leg.; 19JDEC100, general sweep over *Brachychiton* flowering species in sclerophyll forest along dry creek bed, sunny and warm ~33°C; Extraction1067, BOLD: AUGLY041-21; AM K.517929.

Paratypes

AUSTRALIA – **Australian Capital Territory** • 1 ♀; CSIRO Black Mountain off Frith Rd; -35.268, 149.1107; 610 m a.s.l.; 9–26 Oct. 2019; K.M. Bayless leg.; dry sclerophyll forest Malaise trap; Extraction1660, BOLD: AUGLY125-21; ANIC 32 130318 • 1 ♀; same collection data as for preceding; Extraction1645, BOLD: AUGLY122-21; ANIC 32 130319 • 1 ♀; same collection data as for preceding; Extraction1646, BOLD: AUGLY123-21; ANIC 32 130320 • 1 ♀ (ethanol); CSIRO Black Mountain, close to Botanic Garden fence; -35.273611, 149.110556; 538 m a.s.l.; 30 Oct.–6 Nov. 2017; T. Pleines and J. Rodriguez leg.; Malaise; Extraction959, BOLD: AUGLY027-21; ANIC 32 130321 • 1 ♀; same collection data as for preceding; 23–30 Oct. 2017; Extraction974, BOLD: AUGLY028-21; ANIC 32 130322 • 1 ♀ (ethanol); CSIRO Black Mountain; -35.2744, 149.1115; 6 Dec. 2017–5 Jan. 2018; J. Rodriguez, C. Waichert, K.M. Bayless and T. Pleines leg.; Malaise 2, green dry wash; Extraction976, BOLD: AUGLY029-21; ANIC 32 130323 • 1 ♀ (ethanol); same collection data as for preceding; Extraction978, BOLD: AUGLY030-21; ANIC 32 130324 • 1 ♀; same collection data as for preceding;

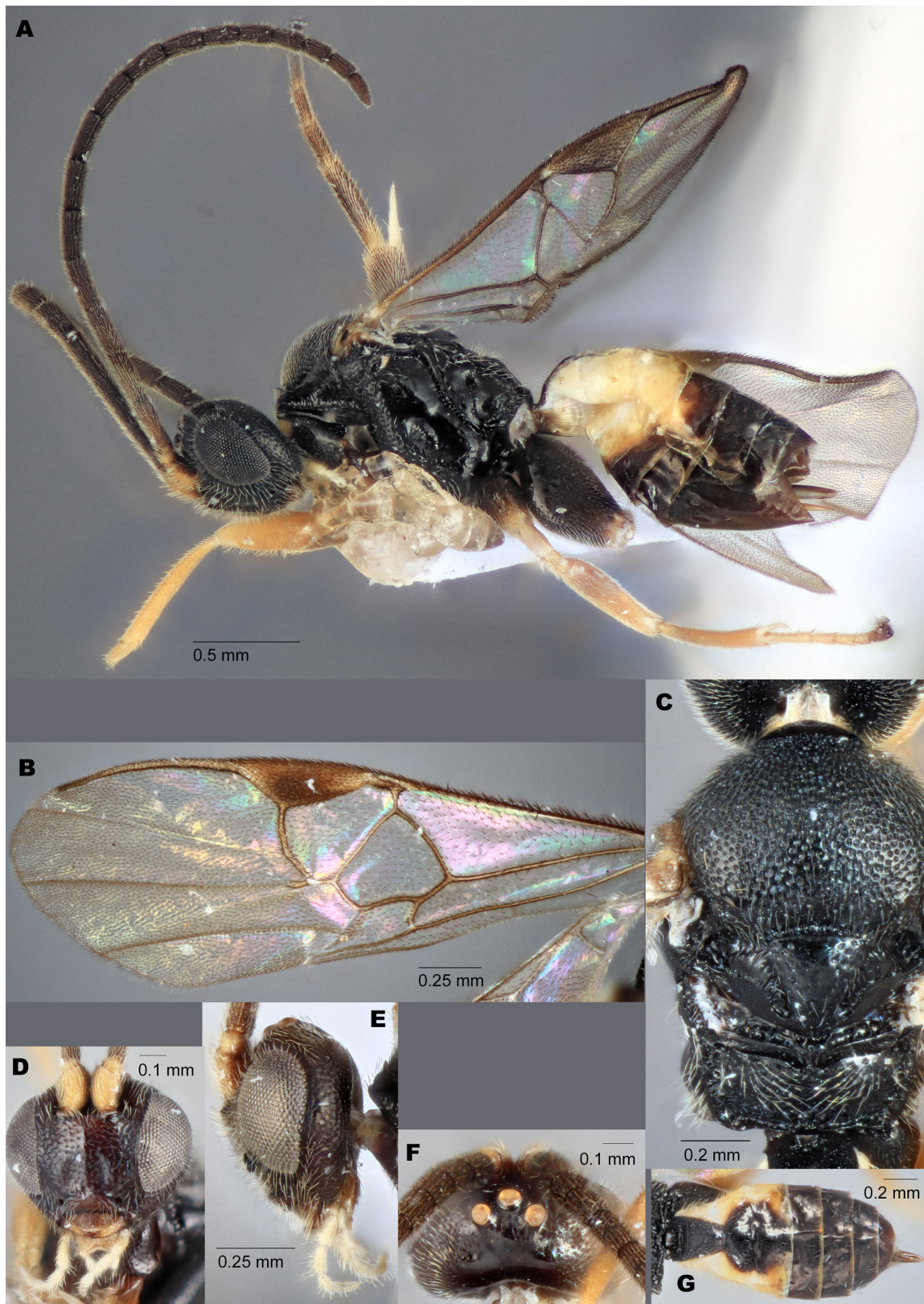


Fig. 28. *Glyptapanteles doreyi* Fagan-Jeffries, Bird & Austin sp. nov., paratypes, ♀, 'clade A'. A, F–G. AUMIC412-18. B–E. AUMIC389-18. A. Lateral habitus. B. Fore wing. C. Dorsal mesosoma. D. Anterior head. E. Lateral head. F. Dorsal head. G. Dorsal metasoma.

Extraction980, BOLD: AUGLY032-21; ANIC 32 130325 • 1 ♀ (ethanol); same collection data as for preceding; Extraction981, BOLD: AUGLY033-21; ANIC 32 130326 • 1 ♀ (ethanol); same collection data as for preceding; Extraction982, BOLD: AUGLY034-21; ANIC 32 130327 • 1 ♀ (ethanol); same collection data as for preceding; Extraction983, BOLD: AUGLY035-21; ANIC 32 130328 • 1 ♀ (ethanol); same collection data as for preceding; 6–20 Jan. 2020; Extraction1658, BOLD: AUGLY124-21; ANIC 32 130329 • 1 ♀; CSIRO property; -35.275, 149.111; 588 m a.s.l.; 7 Sep. 2011; P. Hebert leg.; Malaise; BIOUG02156-H03, BOLD: MCCA225-12; ANIC 32 130330. – **New South Wales** • 1 ♀ (ethanol); same collection data as for holotype; Extraction1086, BOLD: AUGLY043-21; AM K.383784 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1156, BOLD: AUGLY066-21; AM K.383785 • 1 ♂; same collection data as for holotype; Extraction1068, BOLD: AUGLY042-21; AM K.517930 • 1 ♂; same collection data as for holotype; Extraction1087, BOLD: AUGLY044-21; AM K.517931 • 1 ♂; same collection data as for holotype; Extraction1094, BOLD: AUGLY045-21; AM K.517932 • 1 ♂; same collection data as for holotype; Extraction1155, BOLD: AUGLY065-21; AM K.517933 • 1 ♂ (ethanol); same collection data as for holotype; Extraction1157, BOLD: AUGLY067-21; AM K.383786 • 1 ♀; Oxley Wild Rivers National Park, East Kunderang Track; -30.818056, 152.135; 7 Nov. 2015; D.M. Bray leg.; blue pan trap; Extraction1248, BOLD: AUGLY073-21; AM K.517937. – **Queensland** • 1 ♀; Lamington National Park; -28.142, 153.133; 248 m a.s.l.; 8–18 Apr. 2007; C. Lambkin and N. Starick leg.; IBISCA Plot # IQ-300-D rainforest Malaise trap; Extraction605, BOLD: AUMIC390-18; QM T208398 • 1 ♀; same collection data as for preceding; Extraction604, BOLD: AUMIC389-18; QM T208397 • 1 ♀; same collection data as for preceding; Extraction193, BOLD: AUMIC080-18; QM T208393 • 1 ♀ (ethanol); same collection data as for preceding; Extraction632, BOLD: AUMIC416-18; QM T208395 • 1 ♀ (ethanol); same collection data as for preceding; 23 Sep.–5 Oct. 2014; Extraction623, BOLD: AUMIC409-18; QM T208394 • 1 ♀; same collection data as for preceding; 5–22 Oct. 2014; Extraction627, BOLD: AUMIC412-18; QM T208396.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark or reddish-brown; scape colour in ventral half uniformly paler than flagellomeres or paler than flagellomeres at proximal end; flagellomeres all black/dark brown; tegula normally dark (dark in holotype, sometimes paler in paratypes); wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow or light brown; hind femur dark reddish-brown, orange to light brown or dark; fore tibia pale yellow; mid tibia pale yellow or light brown; hind tibia darkening posteriorly; hind basitarsus light brown or dark reddish-brown; T1 dark; T2 sclerotised area dark; T2 lateral area same colour as sclerotised area, or only slightly paler or dark extends past indentation, but then pale; T3 dark, mostly dark with paler lateral areas or uniformly brown; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.4 mm; fore wing length 2.3 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 14 length/width 1.25–2.00; antennal flagellomere 2 length/width 2.00–4.20; OOD/POD 1.57–2.50; IOD/POD 1.29–1.83.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures generally smaller than diameter of punctures; scutellar disk sculpturing with only very shallow punctures; 8–12 pits in scutellar sulcus; propodeum with median carina absent, shallow or strong punctures in anterior half, occasionally with small areas of shallow rugosity, posterior half of propodeum smooth or with shallow or strong rugose sculpturing, propodeum sometimes with smooth area in centre.

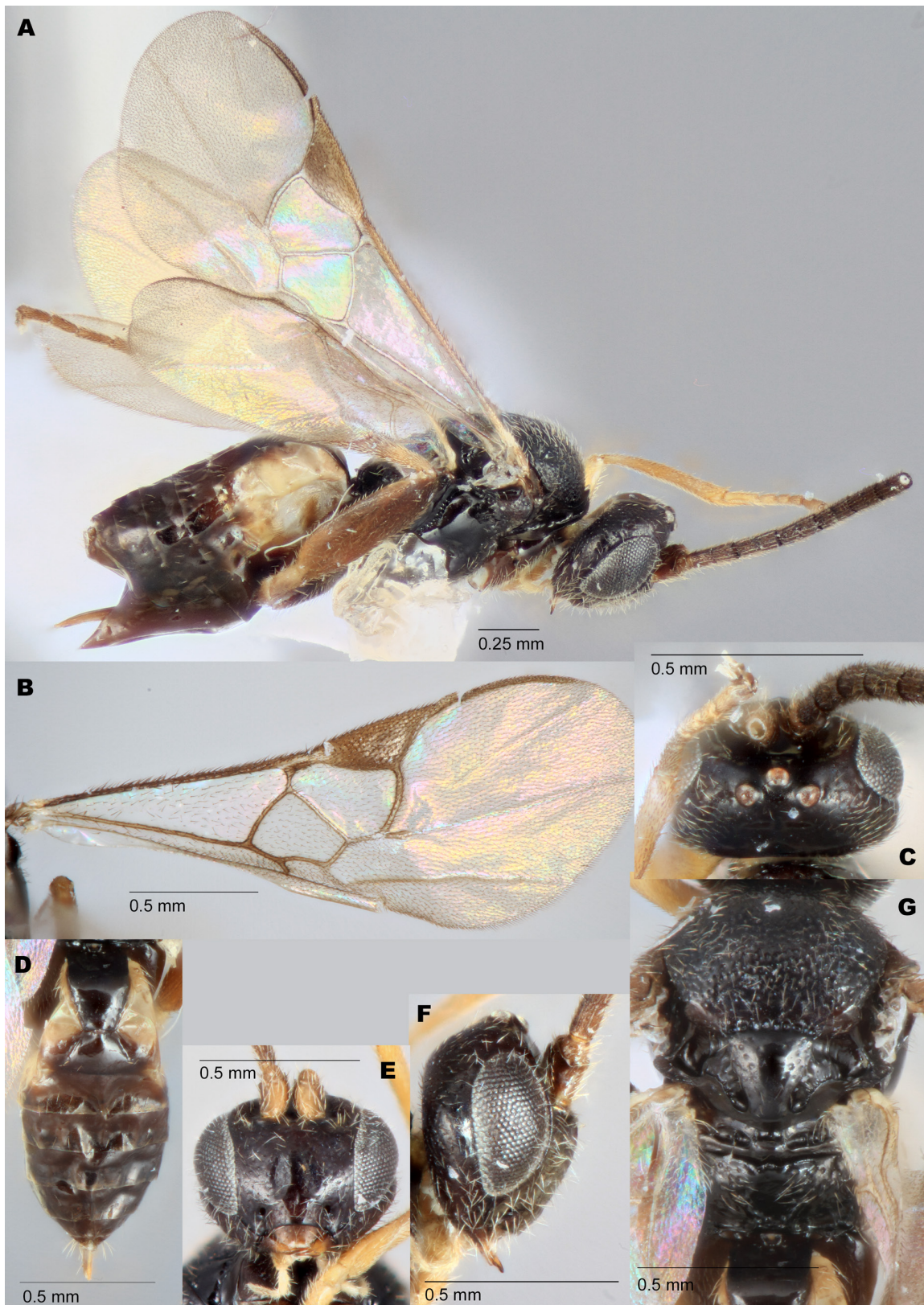


Fig. 29. *Glyptapanteles doreyi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (ANIC 32 130330), 'clade B'. **A.** Lateral habitus. **B.** Fore wing. **C.** Dorsal head. **D.** Dorsal metasoma. **E.** Anterior head. **F.** Lateral head. **G.** Dorsal mesosoma.

WINGS. Pterostigma length 0.53 mm; pterostigma width 0.17 mm; r 0.15 mm; 2RS 0.14 mm; 2m 0.09 mm; (RS+M)b 0.08 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 sometimes shiny, smooth, or mostly smooth with some punctures in posterior half, or smooth in anterior half with indistinct sculpturing in posterior half; T1 length 0.34 mm; T1 width at posterior edge 0.13 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.31 mm; ovipositor slightly protruding from end of metasoma.

Male

As female but with slightly longer antennae.

Remarks

Glyptapanteles doreyi sp. nov. has two *wingless* haplotypes (differing by a single base pair); however, as the *COI* divergence between specimens with the two different haplotypes is not consistent (i.e., one specimen has a *COI* sequence that is more closely related to specimens with the alternative *wingless* haplotype than to others) and the *COI* divergence among all specimens is not above 2%, we consider all specimens to be one species. One of the *wingless* haplotypes is shared with the species *G. arcanus* sp. nov. and *G. goodwinnoakes* sp. nov. and both barcodes only differ by 1 bp from the species *G. vergrandiacus* sp. nov. and *G. lessardi* sp. nov. This group of species is very closely related and would benefit from future, more detailed taxonomic work to ensure that they are not simply divergent populations of the same species. For this species hypothesis, we have made the decision to split this large clade into different species based on the *COI* barcode sequences being >3% divergent; however, we note that this is a hypothesis and is open to change with future work.

In the IQ-TREE analysis of the concatenated *COI* and *wingless* alignment, there are two clades within *G. doreyi* sp. nov. that differ, in their *COI* sequences, between 2.0% and 2.5%. The first clade ('clade A') contains seven sequences (AUMIC416-18, AUMIC409-18, AUMIC080-18, AUGLY044-21, AUMIC412-18, AUMIC390-18, AUMIC389-18). Specimen AUGLY044-21 has the same *wingless* haplotype as the specimens in the second clade within *G. doreyi* sp. nov. ('clade B') whilst the other six specimens share a *wingless* haplotype that differs by one base pair from that of AUGLY044-21. We have made the conservative decision to combine these two clades into a single species due to the *COI* divergence being borderline of what is often used in Microgastrinae, lack of clear morphological differences and the *wingless* haplotype of AUGLY044-21 being identical to that of the alternate clade; however, we provide images of a representative of both clades and note that future evidence (particularly host data) may end up splitting this species further.

Glyptapanteles doreyi sp. nov. constitutes BIN BOLD:ABY0421 (clade B) and BOLD:AEC8663 (clade A). Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 2.6% different from the most similar *COI* sequence from an Australian specimen (MCCAA7675-20; an undescribed lineage).

Distribution

This species is currently known from eastern NSW and from the ACT.

Glyptapanteles dowtoni Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:35AA0388-8874-46DB-8614-DF3CA15601B6](https://zoobank.org/act:35AA0388-8874-46DB-8614-DF3CA15601B6)
Figs 6E, 11C–D, 30

Diagnosis

Glyptapanteles dowtoni sp. nov. is in the *G. mouldsi* species group and can be separated from *G. mouldsi* sp. nov. by having S5+ dark and T4–6 with at least some dark areas, or completely dark.

Glyptapanteles mouldsi sp. nov. has the metasomal sternites all pale yellow other than the hypopygium, which is dark and T4–5 all pale/yellow. With the limited specimens available, there was not a morphological character found that easily differentiates *G. dowtoni* sp. nov. from *G. rixi* sp. nov. The *wingless* sequences of the three species differ by at least 6 bp and the *COI* differs by at least 4.3%.

Etymology

This species is named for Professor Mark Dowton, who collected one of the paratypes, along with many other specimens, whilst kindly running a Malaise trap for this and other microgastrine projects at Mt Keira, NSW.

Material examined

Holotype

AUSTRALIA • ♀; New South Wales, Royal National Park, near Waterfall Couranga Track near Hacking River; -34.1486, 151.0221; 40 m a.s.l.; 20 Jan.–6 Feb. 2020; K.M. Bayless and J.G. Lumbers leg.; 6 m Malaise trap over Waterfall Creek; Extraction1031, BOLD: AUGLY039-21; ANIC 32 130361.

Paratypes

AUSTRALIA – **New South Wales** • 1 ♀ (ethanol); same collection data as for holotype; Extraction944, BOLD: AUGLY024-21; ANIC 32 130362 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1019, BOLD: AUGLY036-21; ANIC 32 130363 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1121, BOLD: AUGLY054-21; ANIC 32 130364 • 1 ♀; same collection data as for holotype; Extraction1126, BOLD: AUGLY057-21; ANIC 32 130365 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1270, BOLD: AUGLY074-21; ANIC 32 130366 • 1 ♀ (ethanol); same collection data as for holotype; Extraction860, BOLD: AUGLY008-21; ANIC 32 130367 • 1 ♂ (ethanol); same collection data as for holotype; Extraction935, BOLD: AUGLY021-21; ANIC 32 130368 • 1 ♂; same collection data as for holotype; Extraction946, BOLD: AUGLY025-21; ANIC 32 130369 • 1 ♀; Mt Keira; -34.416, 150.882; 5–20 Mar. 2020; M. Dowton leg.; Malaise trap, EFJ2020MT29; Extraction878, BOLD: AUGLY014-21; AM K.517934. – **Queensland** • 1 ♀; Lamington National Park; -28.155, 153.139; 282 m a.s.l.; 13–23 Jan. 2007; C. Lambkin and N. Starick leg.; IBISCAPlot # IQ-300-B rainforest Malaise trap, bulk vial 22139; Extraction1465, BOLD: AUGLY083-21; QM T250948 • 1 ♀ (ethanol); same collection data as for preceding; Extraction1466, BOLD: AUGLY084-21; QM T250949 • 1 ♀; same collection data as for preceding; Extraction1468, BOLD: AUGLY086-21; QM T250950 • 1 ♀; same collection data as for preceding; Extraction1469, BOLD: AUGLY087-21; QM T250951 • 1 ♀ (ethanol); same collection data as for preceding; Extraction1470, BOLD: AUGLY088-21; QM T250952 • 1 ♀; same collection data as for preceding; Extraction600, BOLD: AUMIC386-18; QM T208403 • 1 ♀; same collection data as for preceding; Extraction602, BOLD: AUMIC388-18; QM T208404 • 1 ♀; Cape Tribulation, Cow Bay, Daintree Discovery Centre; -16.2382, 145.427; 86 m a.s.l.; 22 Aug. 2015; C.J. Bennett leg.; BIOUG44012-D01, BOLD: GMQQY084-18; QM T250953.

Description

Female

COLOURATION. Gena without a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres darkening distally; tegula dark or pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark reddish-brown, pale or orange to light brown; T2 lateral area same colour as sclerotised area, or only slightly paler or dark extends past indentation, but then pale; T3 mostly pale with darker patch in centre, uniformly brown or pale; T4+ reddish-brown, or T4 pale or pale with dark patch in centre; T5–7 darkening posteriorly.

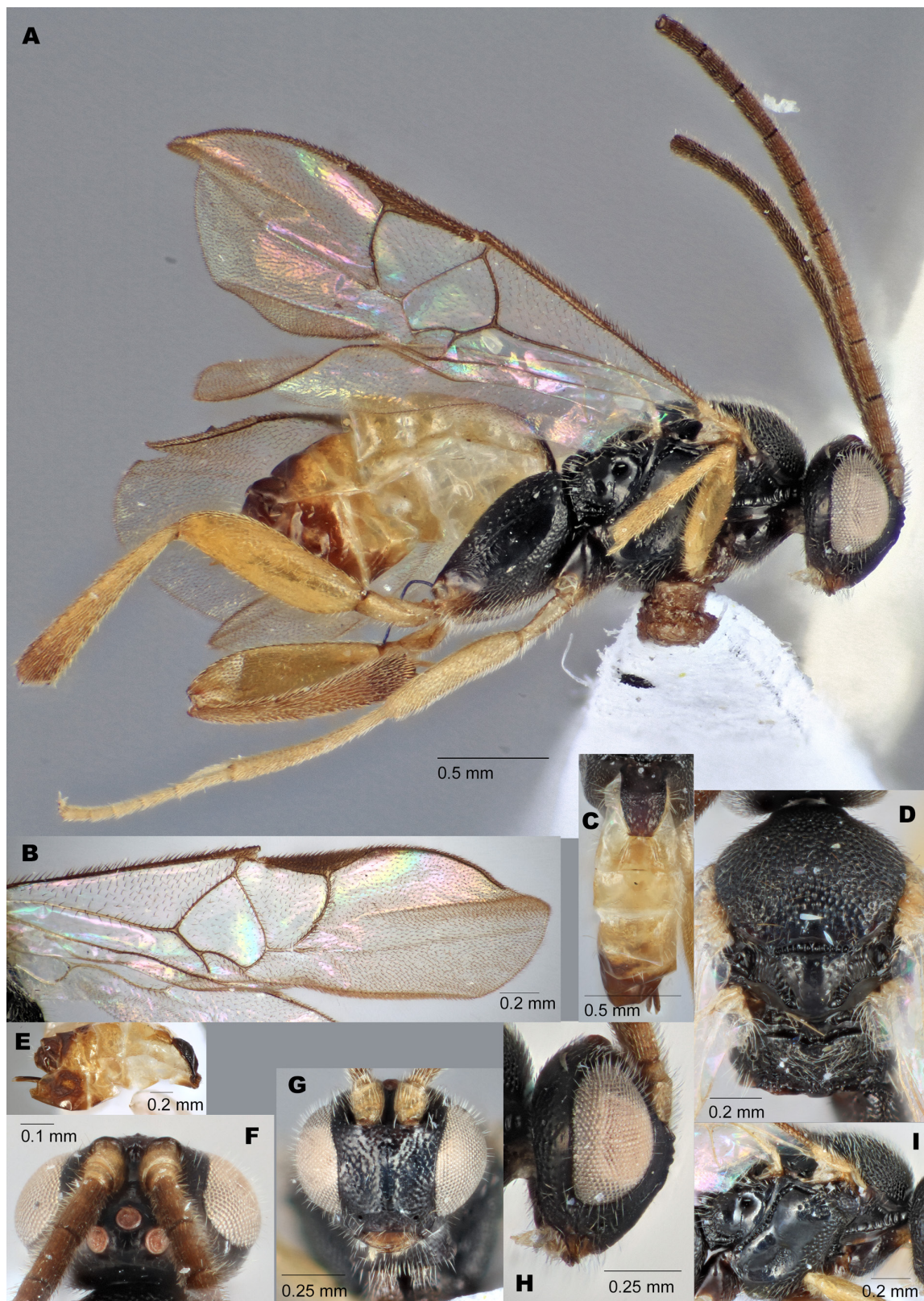


Fig. 30. *Glyptapanteles dowtoni* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250953). A. Lateral habitus. B. Fore wing. C. Dorsal metasoma. D. Dorsal mesosoma. E. Lateral metasoma. F. Dorsal head. G. Anterior head. H. Lateral head. I. Lateral mesosoma.

HOLOTYPE BODY MEASUREMENTS. Body length 2.3 mm; fore wing length 2.3 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 2.00–2.16; antennal flagellomere 2 length/width 2.55–3.83; OOD/POD 1.33–2.17; IOD/POD 1.11–1.67.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size, often smoother in posterior centre; scutellar disk sculpturing with deep, sparse, irregularly spaced punctures, more common on anterior lateral edges, or with sparse punctures scattered over most of area; 9–12 pits in scutellar sulcus; propodeum with median carina absent, propodeum strongly sculptured, punctate reticulate to rugose, sometimes with smooth shining areas in posterior third.

WINGS. Pterostigma length 0.55 mm; pterostigma width 0.17 mm; r 0.18 mm; 2RS 0.15 mm; 2m 0.07 mm; (RS+M)b 0.11 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly or with lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly; T1 smooth with shallow sculpturing along lateral edges or smooth and shiny, some shallow scattered punctures on lateral edges and posterior half; T1 length 0.38 mm; T1 width at posterior edge 0.11 mm; T2 an isosceles trapezoid, lateral edges straight or with curved lateral and anterior edges, becoming arch- or semicircle-shaped; T2 smooth, sometimes shiny, some indistinct sculpturing on postero-lateral corners; T2 length 0.14 mm; T2 width at posterior edge 0.19 mm; ovipositor slightly protruding from end of metasoma.

Male

As female.

Remarks

Glyptapanteles dowtoni sp. nov. constitutes BIN: BOLD:ADL2613 and is 1.61% (p-dist.) divergent from the closest BIN in the database (BOLD:AAH1085). BOLD:AAH1085 has two sequences within it (one of which is public: ASPNI556-09 from Papua New Guinea). This sequence was not included in the original analysis as it is not identified to genus level in BOLD. With the low p-dist. between these two BINs, we consider it possible that they may be the same species, which would mean that *G. dowtoni* sp. nov. is also present in Papua New Guinea. Morphological examination should be completed and a second gene should be sequenced to confirm.

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 2.8% different from the most similar *COI* sequence from an Australian specimen (AUGLY141-21; an undescribed lineage, with a single specimen). Fourteen specimens were able to be sequenced for the *wingless* gene and share a unique barcode that differs by a minimum of 3 bp from all other species with available sequence data.

Distribution

This species is currently known from southern QLD and eastern NSW.

Glyptapanteles drioplanetus Fagan-Jeffries & Austin, 2021

Figs 15A, 31

Diagnosis

Glyptapanteles drioplanetus can be separated from the other described species of *Glyptapanteles* in Australia by the combination of the following characters: gena without a pale spot, T1 and T2 dark,

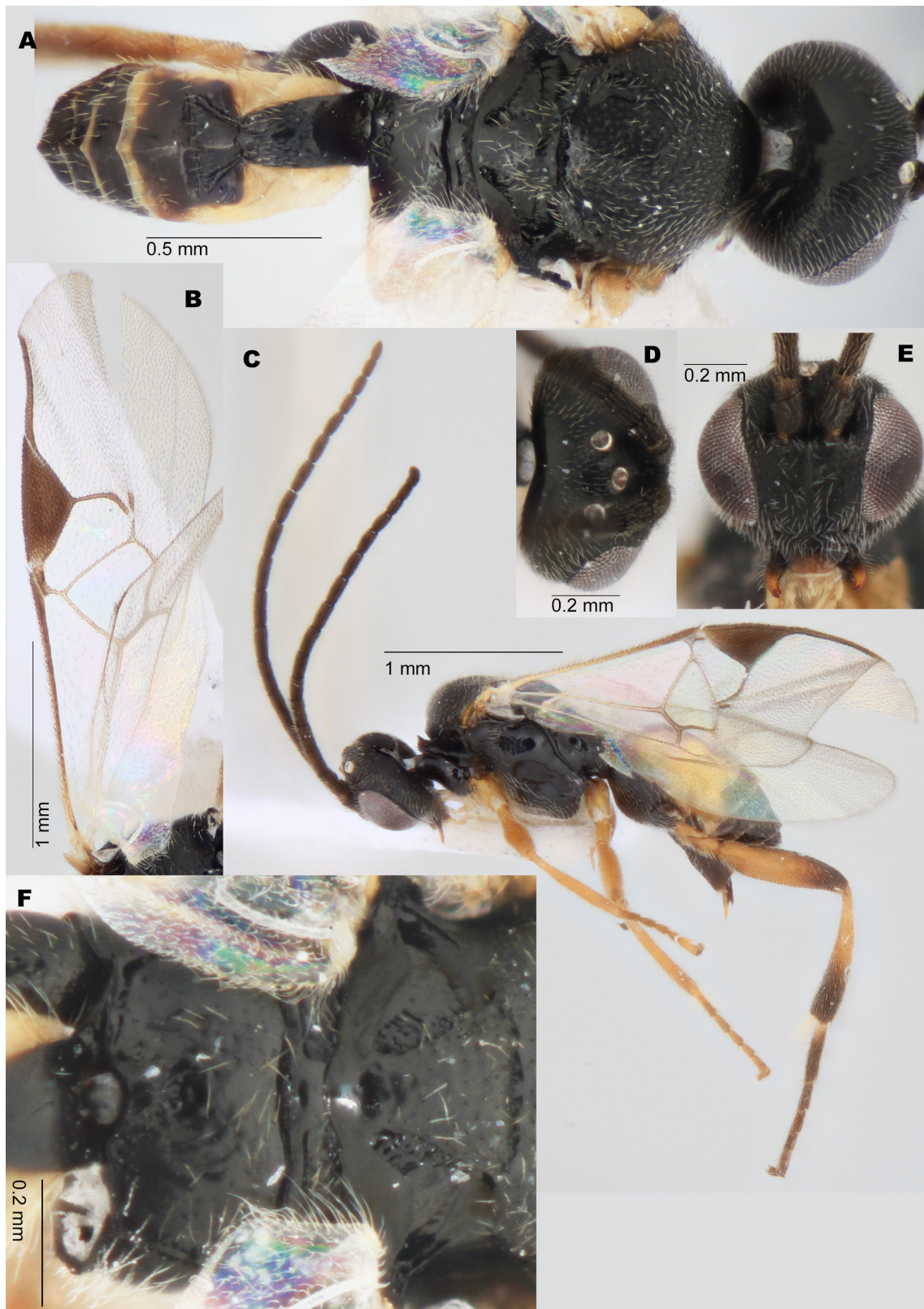


Fig. 31. *Glyptapanteles drioplanetus* Fagan-Jeffries & Austin, 2021, holotype, ♀ (WAM WAME10965). A. Dorsal habitus. B. Fore wing. C. Lateral habitus. D. Dorsal head. E. Anterior head. F. Scutellar disk and propodeum. Images from Fagan-Jeffries & Austin (2021: fig. 5).

anteromesoscutum very smooth, with only shallow and regular punctures, no orange markings on the postero-lateral anteromesoscutum, propodeum very smooth with only shallow punctures and the median carina entirely absent, T1 narrowing slightly for most of length, distal ventral half of antennal scape darker than flagellomeres, hind femur pale anteriorly, darkening posteriorly, tegula pale, labrum pale.

Material examined

Holotype

AUSTRALIA • ♀; South Australia, Macclesfield Primary School; -35.17084, 138.84025; 14 Oct.–6 Nov. 2019; E. Fagan-Jeffries and Macclesfield PS yr. 5–7 class leg.; Malaise Trap EFJ2020MT9; Extraction1033, BOLD: AUMIC549-20; SAMA 32-45153.

Paratypes

AUSTRALIA – **South Australia** • 1 ♀; Douglas Scrub; -35.1844, 138.6; 27–28 Sep. 2010; G.S. Taylor and S. Mantel leg.; Malaise Trap 2010 001; Extraction79, BOLD: AUMIC507-18; SAMA 32-45154 • 1 ♀; Gleneagle State Forest; -32.2711, 116.163; 29 Nov. 2005; M.S. Harvey leg.; Malaise trap; Extraction117, BOLD: AUMIC019-18; WAM WAME10965.

Remarks

This species was recently described and we therefore only provide a diagnosis here. *Glyptapanteles drioplanetus* constitutes BIN: BOLD:ADL3660 and is 2.56% (p-dist.) from the nearest neighbour (BOLD:AEI2300; an undescribed lineage from Australia, with two specimens).

Glyptapanteles eburneus Fagan-Jeffries, Bird & Austin sp. nov.

[urn:lsid:zoobank.org:act:814EBC38-DD57-4503-87B3-01C47A2A517D](https://doi.org/10.12101/zoobank.org/act:814EBC38-DD57-4503-87B3-01C47A2A517D)

Figs 6B, 10A, 32

Diagnosis

Glyptapanteles eburneus sp. nov. is in the *G. eburneus* species group and can be separated from *G. foraminous* sp. nov. by having T2 pale, the same colour as T1 (*G. foraminous* sp. nov. has T2 dark). Additionally, the *wingless* sequences of the two holotype specimens are distinct and differ by 5 bp.

Etymology

The species epithet ‘*eburneus*’ is a Latin adjective for ‘ivory’ and refers to the pale colouration of T1–2.

Material examined

Holotype

AUSTRALIA • ♀; New South Wales, Royal National Park. south end, Lady Carrington Drive Rainforest; -34.1482, 151.031; 24 Nov. 1996; A.D. Austin leg.; Extraction439, BOLD: AUMIC274-18; AM K.517935.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres darkening distally; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with significant orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa pale yellow; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light



Fig. 32. *Glyptapanteles eburneus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (AM K.517935). A. Lateral body. B. Dorsal mesosoma. C. Dorsal head. D. Anterior head. E. Lateral head. F. Dorsal metasoma. G. Lateral habitus.

brown; T1 pale; T2 sclerotised area pale; T2 lateral area same colour as sclerotised area, or only slightly paler; T3 pale; T4 pale with dark patch in centre, then darkening posteriorly on T5–7.

HOLOTYPE BODY MEASUREMENTS. Body length 2.7 mm; fore wing length 2.7 mm; antennal length slightly longer than body length.

HEAD. Face densely sculptured, punctate reticulate; antennal flagellomere 14 length/width 2.16; antennal flagellomere 2 length/width 3.62; OOD/POD 1.75; IOD/POD 1.75.

MESOSOMA. Anteromesoscutum sculpturing with deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size; scutellar disk sculpturing with deep irregularly spaced punctures; nine pits in scutellar sulcus; propodeum with median carina present and complete, rest of propodeum mostly smooth.

WINGS. Pterostigma length 0.62 mm; pterostigma width 0.23 mm; r 0.18 mm; 2RS 0.16 mm; 2m 0.10 mm; (RS+M)b 0.10 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 mostly smooth, some punctures in posterior half; T1 length 0.4 mm; T1 width at posterior edge 0.12 mm; T2 with curved lateral and anterior edges, becoming arch- or semicircle-shaped; T2 smooth and shiny; T2 length 0.17 mm; T2 width at posterior edge 0.3 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles eburneus sp. nov. constitutes BIN: BOLD:ADL5650 and is 2.4% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL3303; *Glyptapanteles foraminous* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 2.3% different from the most similar *COI* sequence from an Australian specimen (AUMIC069-18; *Glyptapanteles foraminous* sp. nov.). The type specimen was able to be sequenced for the *wingless* gene, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

This species is currently represented by a single specimen from Royal National Park, just south of Sydney.

Glyptapanteles erucadesolator Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:22704EB8-67CA-41FF-BACE-0B9CD656788C](https://doi.org/10.21203/rs.3.rs-22704EB8-v1)

Fig. 33

Diagnosis

Glyptapanteles erucadesolator sp. nov. is in the *G. arcanus* species group and can be separated from other members of the species group, other than from *G. lambkinae* sp. nov., by having the propodeum with very coarse and strong rugose sculpting and T1 with strong sculpturing. *Glyptapanteles arcanus* sp. nov., *G. vergrandiacus* sp. nov. and *G. goodwinnoakes* sp. nov. also have coarse sculpturing on the propodeum (although to a lesser degree), but T1 is either smooth or with clear punctures (but not rugose sculpturing). It is noted, however, that all these species are morphologically very similar and

identifications should be made with DNA barcodes. There was not a morphological character found to easily differentiate *Glyptapanteles erucadesolator* sp. nov. from *G. lambkinae* sp. nov., which are sister lineages in the current phylogeny (Fig. 2); however, as the species differ by 2 bp in the *wingless* sequence and there is >6% divergence in the *COI* barcode, we feel confident that these are different species.

Etymology

The species epithet '*erucadesolator*' is derived from the Latin words 'eruca' ('caterpillar') and 'desolator' ('destroyer') and refers to the lepidopteran parasitoid habit of this subfamily. It is a noun in apposition.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Hermit Park; -19.2828, 146.801; 10 m a.s.l.; 7 Jul. 2008; G. Cocks leg.; Malaise trap; gvc9348-1L, BOLD: HYQT050-08; QM T250954.

Paratypes

AUSTRALIA – **Northern Territory** • 1 ♀; Gregory National Park, Station Creek, 0.2 km NNW of Bullita Camp Ground; -16.1117, 130.423; 12 Jan. 2001; M.E. Irwin, F.D. Parker and C. Lambkin leg.; Extraction160, BOLD: AUMIC054-18; ANIC 32 130199. – **Queensland** • 1 ♂; Hervey Range; -19.3812, 146.449; 380 m a.s.l.; 22 Aug. 2017; G. Cocks leg.; Malaise trap; gvcT10741, BOLD: GCQT2624-18; QM T250955.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark; scape colour in ventral half uniformly paler than flagellomeres or the same colour or darker than flagellomeres; flagellomeres darkening distally; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur orange to light brown; mid femur orange to light brown; hind femur orange to light brown; fore tibia orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark or dark reddish-brown; T2 lateral area dark extends past indentation, but then pale; T3 mostly dark with paler lateral areas or uniformly brown; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.4 mm; fore wing length 2.5 mm.

HEAD. Antennal flagellomere 14 length/width 2.66; antennal flagellomere 2 length/width 2.85; OOD/POD 2.50–2.57; IOD/POD 1.57–2.17.

MESOSOMA. Anteromesoscutum sculpturing with densely scattered deep and shallow punctures of irregular size; scutellar disk sculpturing with deep irregularly spaced punctures; 8–12 pits in scutellar sulcus; propodeum with median carina absent, propodeum strongly sculptured, punctate reticulate to rugose, with smooth shining areas in posterior third.

WINGS. Pterostigma length 0.54 mm; pterostigma width 0.2 mm; r 0.17 mm; 2RS 0.16 mm; 2m 0.09 mm; (RS+M)b 0.12 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly, or lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly; T1 mostly smooth, some shallow rugose

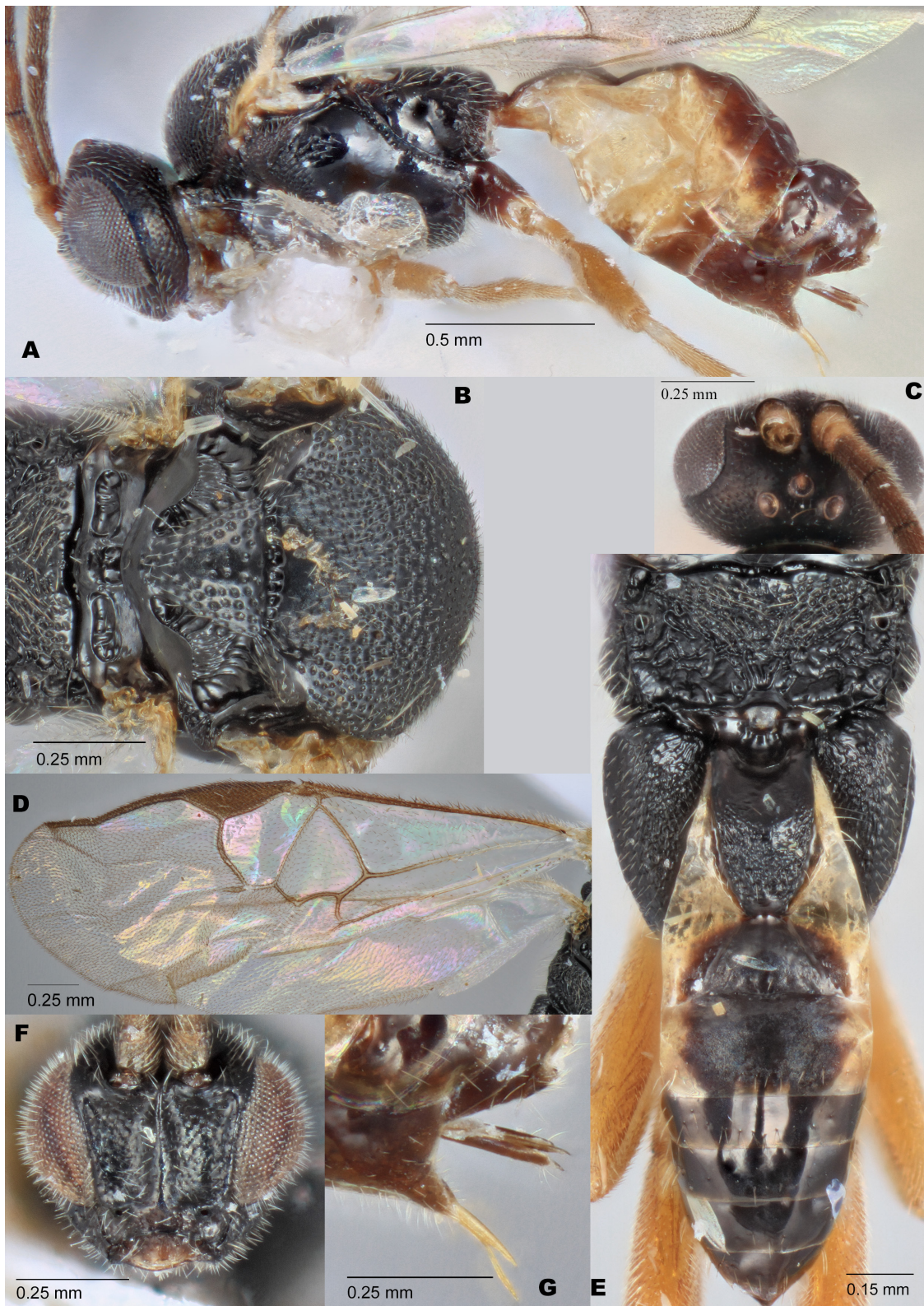


Fig. 33. *Glyptapanteles erucadesolator* Fagan-Jeffries, Bird & Austin sp. nov. **A, C, F.** Paratype, ♀ (ANIC 32 130199). **B, D–E, G.** Holotype, ♀ (QM T250954). **A.** Lateral habitus. **B.** Dorsal mesosoma. **C.** Dorsal head. **D.** Fore wing. **E.** Dorsal propodeum and metasoma. **F.** Anterior head. **G.** Ovipositor sheaths.

sculpturing in posterior half; T1 length 0.37 mm; T1 width at posterior edge 0.08 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny, some shallow punctures along posterior edge; T2 length 0.16 mm; T2 width at posterior edge 0.27 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, antennal flagellomeres slightly longer.

Remarks

Glyptapanteles erucadesolator sp. nov. constitutes BIN BOLD:AAG2095 and is 5.13% (p-dist.) divergent from the closet BIN in the database (BOLD:AEI7392; an undescribed lineage, from Australia).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 5.9% different from the most similar *COI* sequence from an Australian specimen (AUGLY160-21; an undescribed lineage, with one specimen). One of the paratype specimens (AUMIC054-18) was sequenced for the *wingless* gene, which differs by a minimum of 2 bp from all other species with available sequence data.

Distribution

This species is currently known from northern Australia: from Gregory National Park in the NT and northern QLD.

Glyptapanteles ferrugineus Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:FF8A3C0E-096B-40AE-ADA3-33E4069EF15E](https://zoobank.org/act:FF8A3C0E-096B-40AE-ADA3-33E4069EF15E)

Figs 9A, 34

Diagnosis

Glyptapanteles ferrugineus sp. nov. is in the *G. albigena* species group and can be separated from the other members of the species group by the pale spot on the gena being small but clearly visible, T2 dark, propodeal median carina completely absent, hind femur light brown to pale and mesosoma with a strong dark red tinge.

Etymology

The species epithet, '*ferrugineus*', is a Latin adjective for 'rust-coloured' and refers to the red tinge for the mesosoma.

Material examined

Holotype

AUSTRALIA • ♀; Western Australia, 23 km NNW of Albany; -34.8532, 117.809; 16 Nov. 2000; T. Simmul and S. Cunningham leg.; Malaise in remnant vegetation; Extraction144, BOLD: AUMIC042-18; ANIC 32 130189.

Description

Female

COLOURATION. Gena with a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres uniformly reddish-brown; tegula pale; wing veins uniformly pale; anteromesoscutum dark with red tinge; scutellar disk and metanotum dark with red tinge; propodeum dark with red tinge; fore coxa pale yellow; mid coxa pale yellow; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind basitarsus light brown; T1 dark reddish-brown; T2 sclerotised area dark reddish-brown; T2 lateral area dark, extends past indentation, but then pale; T3 mostly dark with paler lateral areas; T4+ reddish-brown.

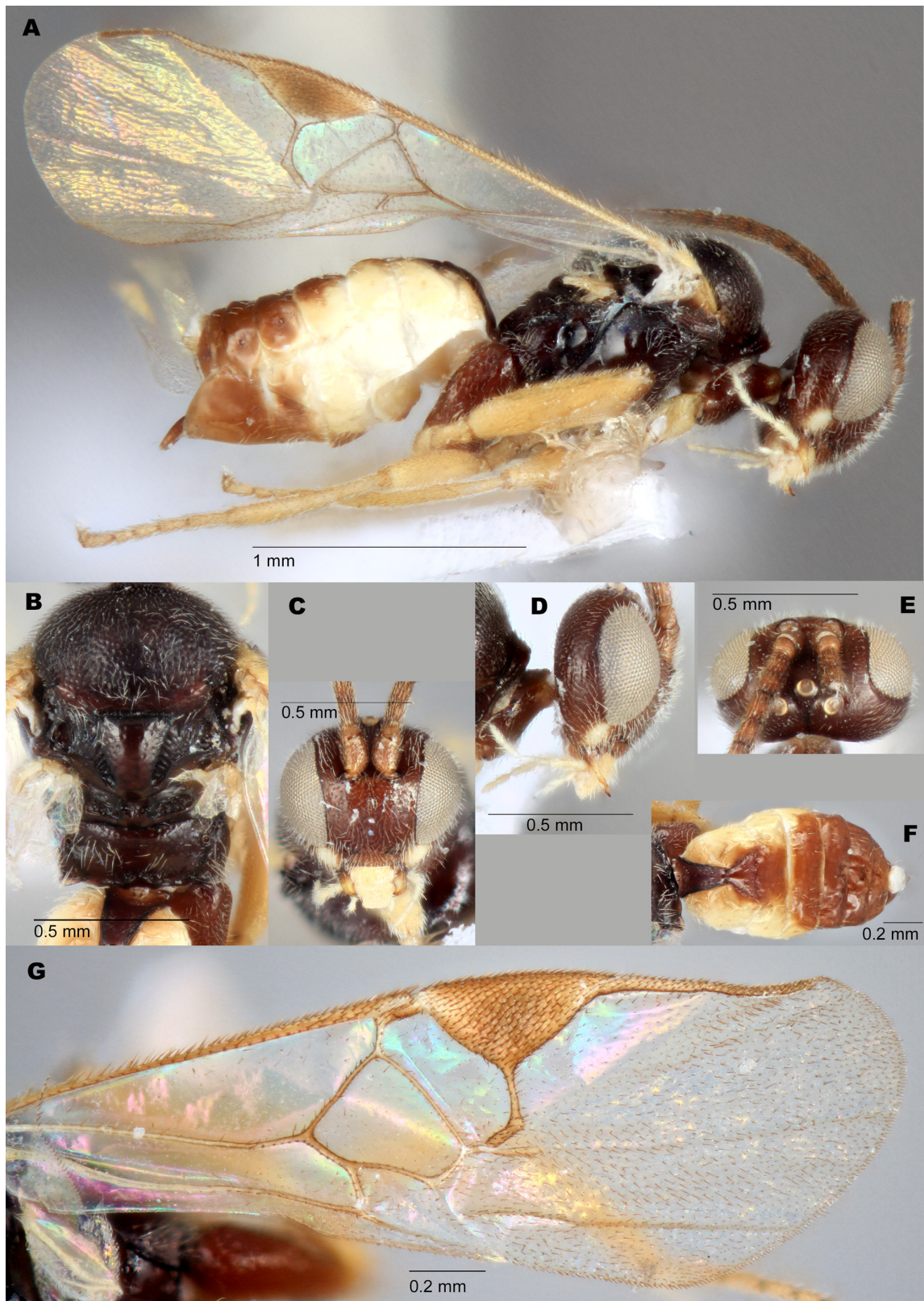


Fig. 34. *Glyptapanteles ferrugineus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (ANIC: 32 130189). A. Lateral habitus. B. Dorsal mesosoma. C. Anterior head. D. Lateral head. E. Dorsal head. F. Dorsal metasoma. G. Fore wing.

HOLOTYPE BODY MEASUREMENTS. Body length 2.2 mm; fore wing length 2.1 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 14 length/width 1.66; antennal flagellomere 2 length/width 3.33; OOD/POD 1.71; IOD/POD 1.57.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures; scutellar disk sculpturing with only very shallow punctures; eight pits in scutellar sulcus; propodeum with median carina absent, very smooth and shiny, only very shallow punctures associated with setae.

WINGS. Pterostigma length 0.48 mm; pterostigma width 0.22 mm; r 0.12 mm; 2RS 0.1 mm; 2m 0.13 mm; (RS+M)b 0.07 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 smooth and shiny; T1 length 0.31 mm; T1 width at posterior edge 0.07 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.13 mm; T2 width at posterior edge 0.21 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles ferrugineus sp. nov. constitutes BIN: BOLD:ADL4089 and is 4.17% (p-dist.) divergent from the closet BIN in the database (BOLD:ABA6213; *Glyptapanteles austrinus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 4.0% different from the most similar *COI* sequence from an Australian specimen (HYAT371-11; *Glyptapanteles austrinus* sp. nov.). The holotype was able to be sequenced for the *wingless* gene, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

This species is known only from Albany in southern WA.

Glyptapanteles foraminous Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:F8AD515F-46A6-4AA9-996B-D05FB4650188](https://zoobank.org/act:F8AD515F-46A6-4AA9-996B-D05FB4650188)

Fig. 35

Diagnosis

Glyptapanteles foraminous sp. nov. is in the *G. eburneus* species group and can be separated from *G. eburneus* sp. nov. by having T2 dark; *G. eburneus* sp. nov. has T2 pale. Additionally, the *wingless* sequences of the two holotype specimens are distinct and differ by 5 bp.

Etymology

The species epithet ‘*foraminous*’ is a Latin adjective meaning ‘full of holes’ and refers to the highly sculptured mesosoma with dense indentations.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Cainbale Quarry, “OF”; -28.145, 153.113; 3–19 Feb. 2009; F. Turco leg.; Extraction182, BOLD: AUMIC069-18; QM T208408.

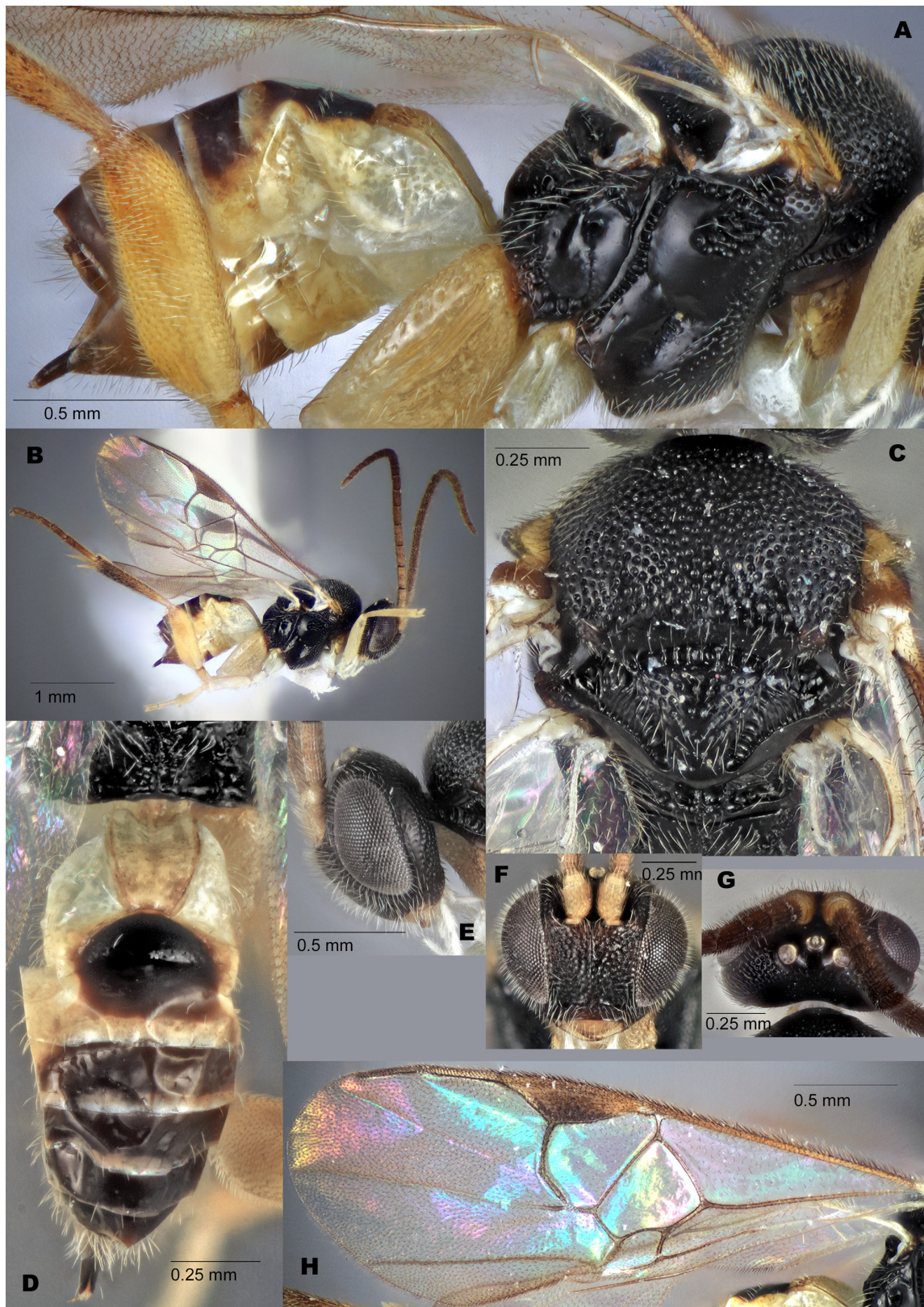


Fig. 35. *Glyptapanteles foraminous* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T208408). A. Lateral mesosoma. B. Lateral habitus. C. Dorsal mesosoma. D. Dorsal metasoma. E. Lateral head. F. Anterior head. G. Dorsal head. H. Fore wing.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres uniformly reddish-brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa white; mid coxa white; hind coxa pale yellow; mid femur pale yellow; hind femur pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 pale; T2 sclerotised area dark; T2 lateral area much paler; T3 mostly dark with paler lateral areas; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.0 mm; fore wing length 2.0 mm; antennal length slightly longer than body length.

HEAD. Face densely sculptured, punctate reticulate; antennal flagellomere 14 length/width 2.71; antennal flagellomere 2 length/width 3.57; OOD/POD 1.75; IOD/POD 1.88.

MESOSOMA. Anteromesoscutum sculpturing with deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size; scutellar disk sculpturing with deep irregularly spaced punctures; 10 pits in scutellar sulcus; propodeum with median carina present and complete, rest of propodeum mostly smooth.

WINGS. Pterostigma length 0.62 mm; pterostigma width 0.21 mm; r 0.22 mm; 2RS 0.19 mm; 2m 0.09 mm; (RS+M)b 0.11 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 mostly smooth, some punctures in posterior half; T1 length 0.5 mm; T1 width at posterior edge 0.1 mm; T2 with curved lateral and anterior edges, becoming arch- or semicircle-shaped; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.33 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles foraminous sp. nov. constitutes BIN: BOLD:ADL3303 and is 2.32% (p-dist.) divergent from the closet BIN in the database (BOLD:ADZ3782, a BIN that has not been released publicly).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 2.3% different from the most similar *COI* sequence from an Australian specimen (AUMIC274-18; *Glyptapanteles eburneus* sp. nov.). The type specimen was able to be sequenced for the *wingless* gene, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

This species is currently only represented by a single specimen from Cainbale Quarry in southern QLD.

Glyptapanteles fullawayi Austin & Dangerfield, 1992

Fig. 16C

Diagnosis

The combination of a smooth propodeum, T1–2 dark, T1 only narrowing slightly posteriorly and gena without a pale spot, allow *G. fullawayi* to be separated from the *G. albigena*, *G. mouldsi*, *G. eburneus*, *G. niveus* and *G. arcanus* species groups.

Glyptapanteles fullawayi is morphologically similar (at least using the available illustrations) to the *G. austini* species group and to *Glyptapanteles drioplanetus*. *Glyptapanteles fullawayi* appears to have only 6–7 large pits in the scutellar sulcus (differing from the *G. austini* species group and from *G. drioplanetus*, which all have at least 8); however, this character is prone to variability. Despite this, we feel that it is not very likely that this species is conspecific with any of the newly described species due to the distribution being restricted to Western Samoa.

Material examined

No specimens examined, illustrations and diagnosis in Austin & Dangerfield (1992) used as reference (Fig. 16).

Distribution

Western Samoa.

Glyptapanteles goodwinnoakes Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:C39D8F6A-C62D-4D6C-AC74-BBEBAED50ED5](https://zoobank.org/urn:lsid:zoobank.org:act:C39D8F6A-C62D-4D6C-AC74-BBEBAED50ED5)

Fig. 36

Diagnosis

Glyptapanteles goodwinnoakes sp. nov. can be separated from the other members of the *G. arcanus* species group by having the propodeum with generally coarser sculpturing than most species other than *G. erucadesolator* sp. nov., *G. lambkinae* sp. nov., *G. arcanus* sp. nov. and *G. vergrandiacus* sp. nov. and with T1 smooth (in all four of the previously mentioned species, T1 is either rugose or at least clearly punctured in the posterior half). It is noted, however, that all these species are morphologically very similar; limited specimens were available for examination and identifications should be made or corroborated with DNA barcodes.

Etymology

This species is named for Alison Goodwin and Roger Noakes, who ran a Malaise trap on their property near Bendemeer, NSW, from which a paratype of the species originates. It is a noun in apposition.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Lonesome National Park; -25.495, 148.812; 585 m a.s.l.; 3–26 Nov. 2010; C. Lambkin *et al.* leg.; near lookout (LNP4m), closed *Eucalyptus* woodland on rocky ridge, Malaise trap; Extraction647, BOLD: AUMIC425-18; QM T208399.

Paratypes

AUSTRALIA - **New South Wales** • 1 ♀; Bendemeer; -30.819, 151.142; 840 m a.s.l.; 9–23 Feb. 2020; A. Goodwin and R. Noakes leg.; Malaise trap, remnant bushland on cattle-grazed property; Extraction923, BOLD: AUGLY018-21; QM T250956. – **Queensland** • 1 ♀; West Quinkan, Bush Blitz Site F3; -15.812550, 144.41974; 495 m a.s.l.; 6–15 Mar. 2017; B. Baehr leg.; open woodland; Extraction895, BOLD: AUGLY017-21; QM T250957.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres all black/dark brown; tegula light brown; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark, or dark with very slight

orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur orange to light brown; hind femur dark reddish-brown or orange to light brown; fore tibia pale yellow; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus dark reddish-brown; T1 dark; T2 sclerotised area dark; T2 lateral area same colour as sclerotised area, or only slightly paler or dark extends past indentation, but then pale; T3 dark; T4+ dark.

HOLOTYPE BODY MEASUREMENTS. Body length 2.2 mm; fore wing length 2.1 mm; antennal length similar to body length.

HEAD. Antennal flagellomere 14 length/width 1.50–1.66; antennal flagellomere 2 length/width 2.62–3.50; OOD/POD 1.71–2.00; IOD/POD 1.43–1.67.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size. Often smoother in posterior centre; scutellar disk sculpturing with shallow punctures scattered over most of area; 8–10 pits in scutellar sulcus; propodeum with median carina absent and propodeum strongly rugose, coarse sculpturing over most of propodeum, sometimes sculpturing clearly deeper in a ‘v’ shape with smooth anterior centre area.

WINGS. Pterostigma length 0.46 mm; pterostigma width 0.15 mm; r 0.17 mm; 2RS 0.12 mm; 2m 0.07 mm; (RS+M)b 0.08 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 smooth and shiny, a few scattered punctures; T1 length 0.34 mm; T1 width at posterior edge 0.11 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.17 mm; T2 width at posterior edge 0.12 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

The *wingless* barcode for this species is shared with *G. doreyi* sp. nov. and *G. arcanus* sp. nov. and differs by 1 bp from the barcodes of *G. vergrandiacus* sp. nov., *G. doreyi* sp. nov. (second haplotype) and *G. lessardi* sp. nov. This group of species is very closely related and would benefit from future, more detailed taxonomic work to ensure that they are not simply divergent populations of the same species. For this species hypothesis, we have made the decision to split them into different species based on the *COI* barcode sequences being >3% divergent; however, we note that this is a hypothesis and is open to change with future work.

Glyptapanteles goodwinnoakes sp. nov. constitutes BIN BOLD:ADL3801 and is 3.69% (p-dist.) divergent from the closet BIN in the database (BOLD:AEI4383; an undescribed lineage, from Australia).

Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 6.1% different from the most similar *COI* sequence from an Australian specimen (AUGLY141-21; an undescribed lineage, with a single specimen).

Distribution

This species has a very disjunct distribution, found in northern QLD and in central NSW. This likely reflects poor sampling rather than the true species distribution.

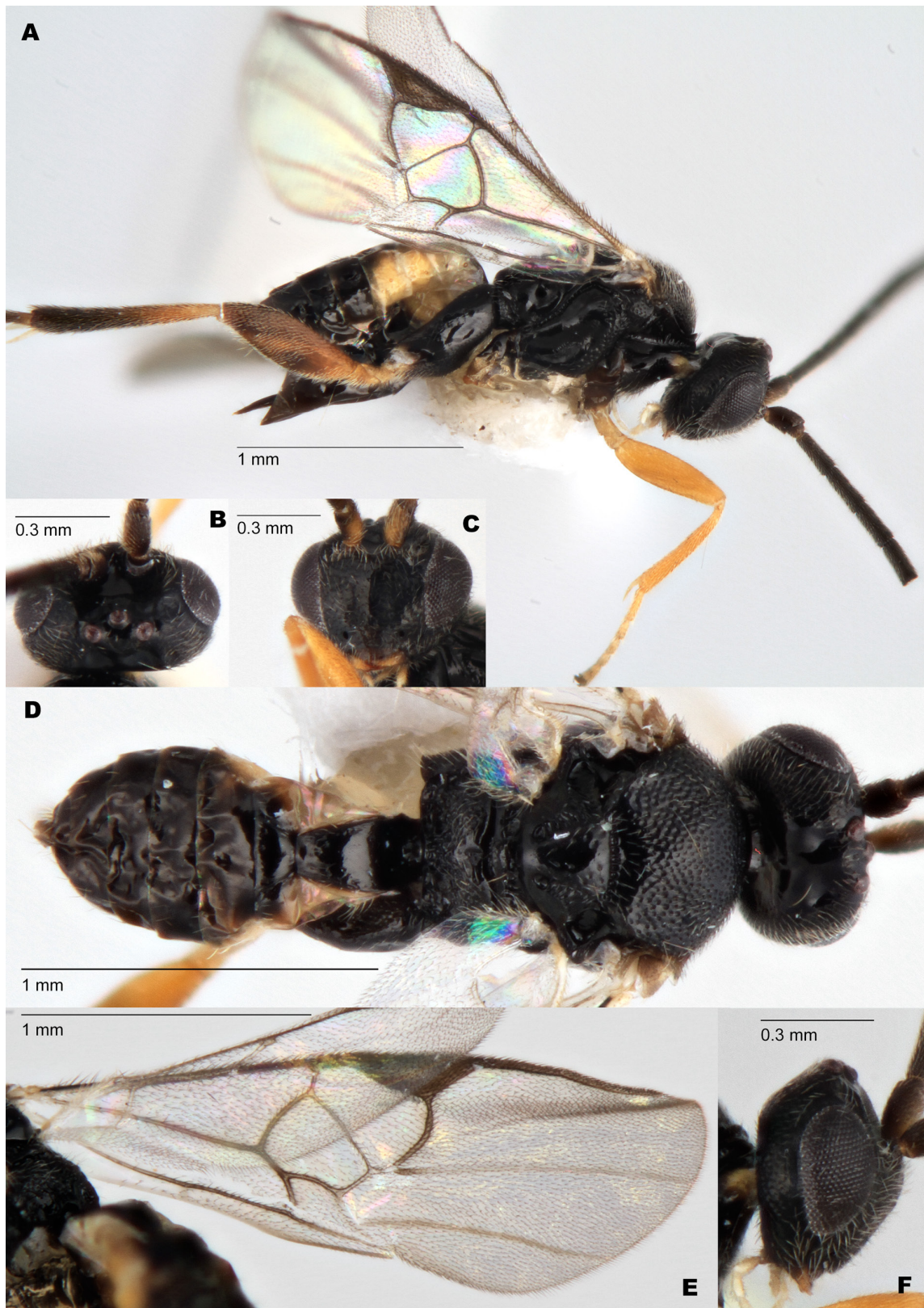


Fig. 36. *Glyptapanteles goodwinnoakes* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250956). A. Lateral habitus. B. Dorsal head. C. Anterior head. D. Dorsal habitus. E. Fore wing. F. Lateral head.

Glyptapanteles guzikae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:A1FFF18B-61D9-4839-B54B-E936F756125D](https://zoobank.org/urn:lsid:zoobank.org:act:A1FFF18B-61D9-4839-B54B-E936F756125D)

Fig. 37

Diagnosis

Despite the large molecular divergence in both the *COI* and *wingless* genes (see remarks section), the only morphological character found to differentiate *G. guzikae* sp. nov. from *G. kingae* sp. nov. and *G. austini* sp. nov. is the femur being pale with a dark area posteriorly, rather than the femur being all dark as in *G. kingae* sp. nov. and *G. austini* sp. nov. However, we note that this subtle colour character may not be robust when further specimens are added to the dataset.

Etymology

Named for Dr Michelle Guzik, who has provided many years of mentorship and advice to EPF-J.

Material examined

Holotype

AUSTRALIA • ♀; South Australia, Kangaroo Island; -35.7533, 137.321; 17–24 Mar. 2011; G. Taylor, E. Kinnaird and R. Kittel leg.; Malaise trap MT3; Extraction97, BOLD: AUMIC524-18; SAMA 32-46153.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa dark; mid femur orange to light brown; hind femur orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark; T2 lateral area same colour as sclerotised area, or only slightly paler; T3 mostly pale with darker patch in centre; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.1 mm; fore wing length 2.3 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 2.00; antennal flagellomere 2 length/width 2.57; OOD/POD 2.17; IOD/POD 1.83.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size, smoother in posterior centre; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; nine pits in scutellar sulcus; propodeum with median carina absent, very smooth and shiny, only very shallow punctures associated with setae.

WINGS. Pterostigma length 0.51 mm; pterostigma width 0.19 mm; r 0.13 mm; 2RS 0.14 mm; 2m 0.07 mm; (RS+M)b 0.08 mm.

METASOMA. T1 lateral edges parallel for entirety of length, posterior corners sometimes rounded at boundary with T2; T1 mostly smooth, some punctures in posterior half or smooth in anterior half, indistinct sculpturing in posterior half; T1 length 0.34 mm; T1 width at posterior edge 0.08 mm; T2

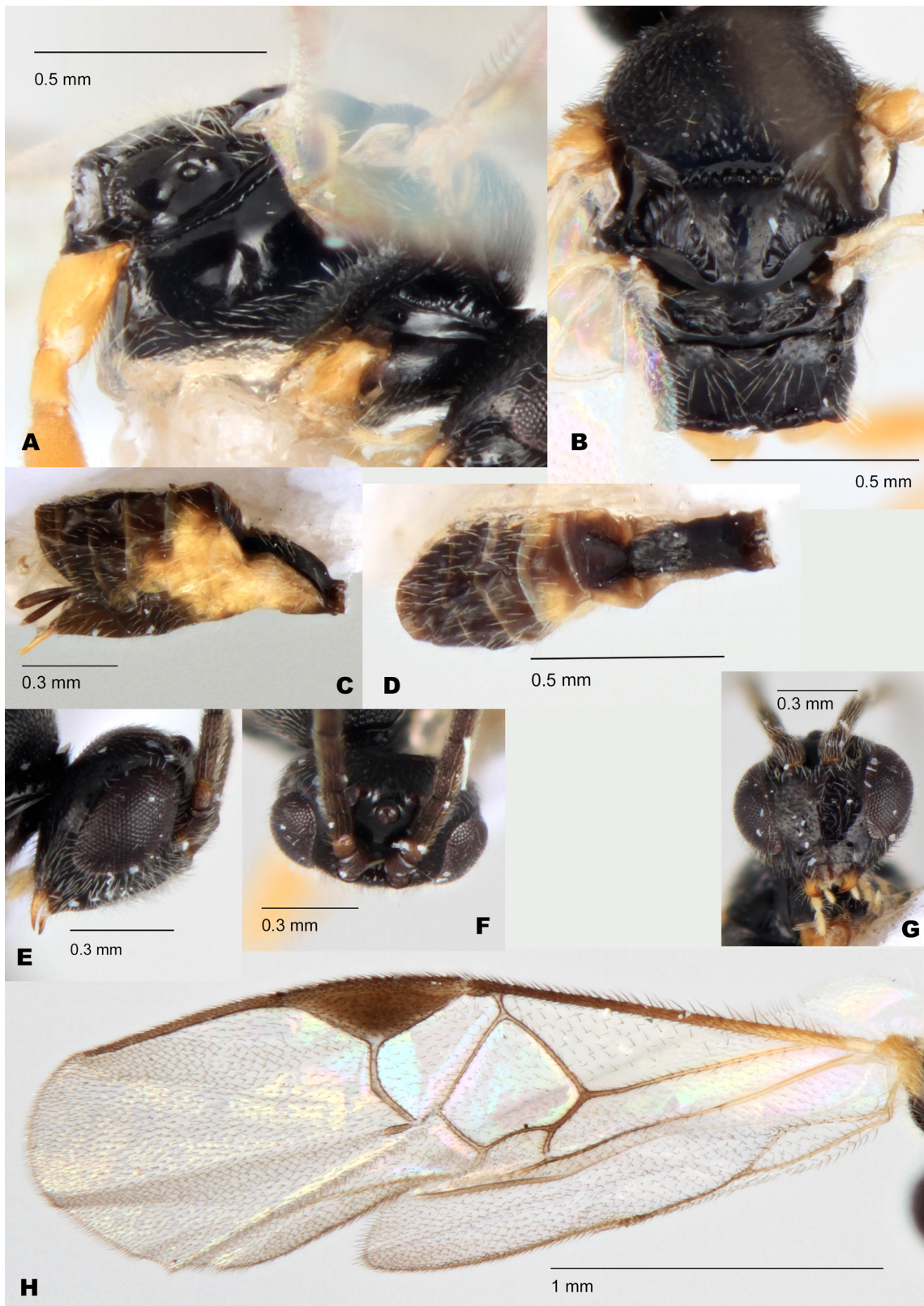


Fig. 37. *Glyptapanteles guzikae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-46153). A. Lateral mesosoma. B. Dorsal mesosoma. C. Lateral metasoma. D. Dorsal metasoma. E. Lateral head. F. Dorsal head. G. Anterior head. H. Fore wing.

an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.12 mm; T2 width at posterior edge 0.14 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles guzikae sp. nov. constitutes BIN BOLD:ADL2952 and is 6.86% (p-dist.) divergent from the closet BIN in the database (BOLD:ACL9923; an undescribed lineage, from New Zealand).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 7.7% different from the most similar *COI* sequence from an Australian specimen (AUMIC291-18; *Glyptapanteles austini* sp. nov.). The type specimen was sequenced for the *wingless* gene and has a unique *wingless* barcode which differs by a minimum of 15 bp from all other species with available sequence data.

Distribution

This species is known from Kangaroo Island, off the coast of SA.

Glyptapanteles harveyi Fagan-Jeffries, Bird & Austin sp. nov.

[urn:lsid:zoobank.org:act:2217AE78-E98A-4283-97B4-033E2175309C](https://zoobank.org/act:2217AE78-E98A-4283-97B4-033E2175309C)

Figs 5D, 6F, 7A, 38

Diagnosis

Glyptapanteles harveyi sp. nov. is in the *G. albigena* species group and can be separated all other members of the species group by having the pale spot on the gena barely visible, T2 dark and the propodeum with the median carina faintly indicated at anterior and posterior ends.

Etymology

Named for Dr Mark Harvey, who collected the type material and has generously run Malaise traps to collect microgastrine material, along with providing advice for many years on the International Code of Zoological Nomenclature to EPF-J. We thank Mark for his dedication to robust taxonomic science and his support of early career researchers.

Material examined

Holotype

AUSTRALIA • ♀; Western Australia, Gleneagle State Forest; -32.2711, 116.163; 8–9 Oct. 2005; M.S. Harvey leg.; Malaise trap; Extraction119, BOLD: AUMIC021-18; WAM E109888.

Paratype

AUSTRALIA • 1 ♀; same collection data as for holotype; Extraction413, BOLD: AUMIC246-18; WAM E109889.

Description

Female

COLOURATION. Gena with a pale spot, but only weakly visible; labrum reddish-brown; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur orange to light brown; mid femur orange to light brown; hind femur orange to light

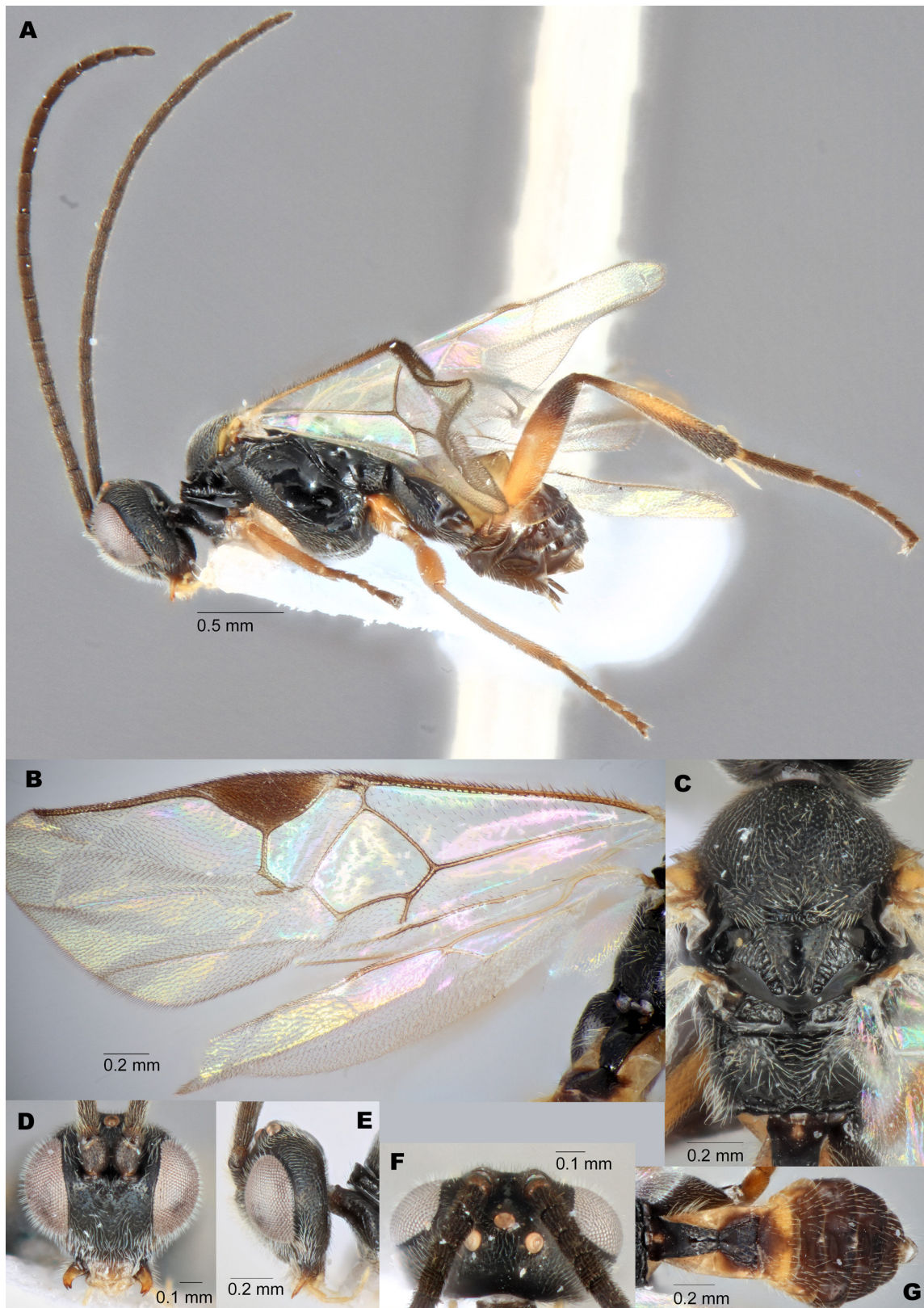


Fig. 38. *Glyptapanteles harveyi* Fagan-Jeffries, Bird & Austin sp. nov. **A–B, D.** Holotype, ♀ (WAM E109888). **C, E–G.** Paratype, ♀ (WAM E109889). **A.** Lateral habitus. **B.** Fore wing. **C.** Dorsal mesosoma. **D.** Anterior head. **E.** Lateral head. **F.** Dorsal head. **G.** Dorsal metasoma.

brown; fore tibia orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus dark reddish-brown; T1 dark; T2 sclerotised area dark; T2 lateral area dark extending past indentation, but then pale; T3 mostly pale with darker patch in centre or uniformly brown; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.5 mm; fore wing length 2.6 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 2.16–2.4; antennal flagellomere 2 length/width 3.28–3.57; OOD/POD 1.86; IOD/POD 1.71–1.86.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures; scutellar disk sculpturing with only very shallow punctures; seven pits in scutellar sulcus; propodeum with median carina very faintly indicated in posterior half, anterior half with some punctures, postero-lateral corners smooth and shiny.

WINGS. Pterostigma length 0.63 mm; pterostigma width 0.26 mm; r 0.14 mm; 2RS 0.1 mm; 2m 0.11 mm; (RS+M)b 0.1 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 mostly smooth, some punctures in posterior half; T1 length 0.37 mm; T1 width at posterior edge 0.07 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth, some very shallow punctures in posterior half; T2 length 0.16 mm; T2 width at posterior edge 0.25 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles harveyi sp. nov. constitutes BIN: BOLD:ADL3315 and is 5.33% (p-dist.) divergent from the closet BIN in the database (BOLD:ACL9711; *Glyptapanteles* sp. from New Zealand).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 6.2% different from the most similar *COI* sequence from an Australian specimen (AUMIC482-18; *Glyptapanteles aspersus* sp. nov.). The two specimens were sequenced for the *wingless* gene and share a unique barcode, which differs by a minimum of 3 bp from all other species with available sequence data.

Distribution

This species is currently known from Gleneagle State Forest near Perth, WA.

Glyptapanteles kingae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:9FC2A388-3A78-47E5-99BE-789CD56291CE](https://zoobank.org/act:9FC2A388-3A78-47E5-99BE-789CD56291CE)
Figs 14B, 39

Diagnosis

Glyptapanteles kingae sp. nov. is in the *G. austini* species group. Despite the large molecular divergence in both the *COI* and *wingless* genes (see Remarks section), the only morphological character found to differentiate *G. kingae* sp. nov. from *G. guzikae* sp. nov. is the femur being completely dark (*G. guzikae* sp. nov. has the femur pale with a dark area posteriorly). *Glyptapanteles kingae* sp. nov. can be separated from *G. austini* sp. nov. as follows: *G. kingae* sp. nov. has the propodeum with stronger sculpturing in

the centre, whilst *G. austini* sp. nov. has the propodeum smoother in the centre. However, we note that there are limited specimens available and the variability of propodeal sculpturing could broaden as more populations of these species are identified and we recommend confirming identifications with DNA barcodes.

Etymology

This species is named for Dr Rachael King, who first introduced the concepts of taxonomy and systematics to EPF-J during an honours project on amphipods in the Pilbara. Fittingly, this species is the only member of *Glyptapanteles* currently described from the Pilbara region.

Material examined

Holotype

AUSTRALIA • ♀; South Australia, Witchelina Stn; -30.1211, 137.923; 19 Oct. 2010; F. Colombo leg.; Bush Blitz Svy FC055 sweeping; Extraction508, BOLD: AUMIC309-18; SAMA 32-46154.

Paratypes

AUSTRALIA – **Australian Capital Territory** • 1 ♀; Black Mountain Res, W side; -35.2764, 149.0897; 610 m a.s.l.; 31 Mar. 2020; K.M. Bayless leg.; sweep over grass, rocks and *Callistemon*; Extraction1591, BOLD: AUGLY113-21; ANIC 32 130331. – **South Australia** • 1 ♀; Witchelina Stn; -30.1853, 137.977; 18–22 Oct. 2010; S. Mantel, F. Colombo and R. Kittel leg.; Bush Blitz Svy, Malaise 2 in dry creek bed [with] *Eremophila freelingi* and *Acacia tetragonophila*; Extraction22, BOLD: AUMIC101-18; SAMA 32-46155. – **Western Australia** • 1 ♂; Western Pilbara, Hamersley Stn, Nanutarra-Wittenoom Rd., approx. 13 km NE of Railway Rd. crossing, near fence line; -22.435556, 117.832222; 22–27 Sep. 2005; Conservation Volunteers Australia leg.; Extraction1354, BOLD: AUGLY080-21; AM K.247607.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark; scape colour in ventral half same colour as flagellomeres; flagellomeres all dark; tegula pale; wing veins uniformly dark, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur orange to light brown; mid femur orange to light brown; hind femur dark; fore tibia orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus dark; T1 dark; T2 sclerotised area black; T2 lateral area same colour as sclerotised area, or only slightly paler, or dark extends past indentation, but then pale; T3 dark; T4+ dark.

BODY MEASUREMENTS. Body length 2.0 mm; fore wing length 2.2 mm; antennal length slightly longer than body length.

HEAD. Face with fine punctures associated with setae; flagellomere 14 length/width 1.43–1.83; flagellomere 2 length/width 2.86–3.80; OOD/POD 1.86–2.00; IOD/POD 1.43.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures, slightly denser and deeper punctures anteriorly, smoother in posterior centre; scutellar disk sculpturing with only very shallow punctures; 8–11 pits in scutellar sulcus; propodeum with median carina absent and indistinct sculpturing present in centre, with rest of propodeum smooth, or propodeum with median carina present in posterior third, with indistinct sculpturing present in centre, with rest of propodeum smooth.

WINGS. Pterostigma length 0.5 mm; pterostigma width 0.17 mm; r 0.13 mm; 2RS 0.14 mm; 2m 0.07 mm; (RS+M)b 0.08 mm.

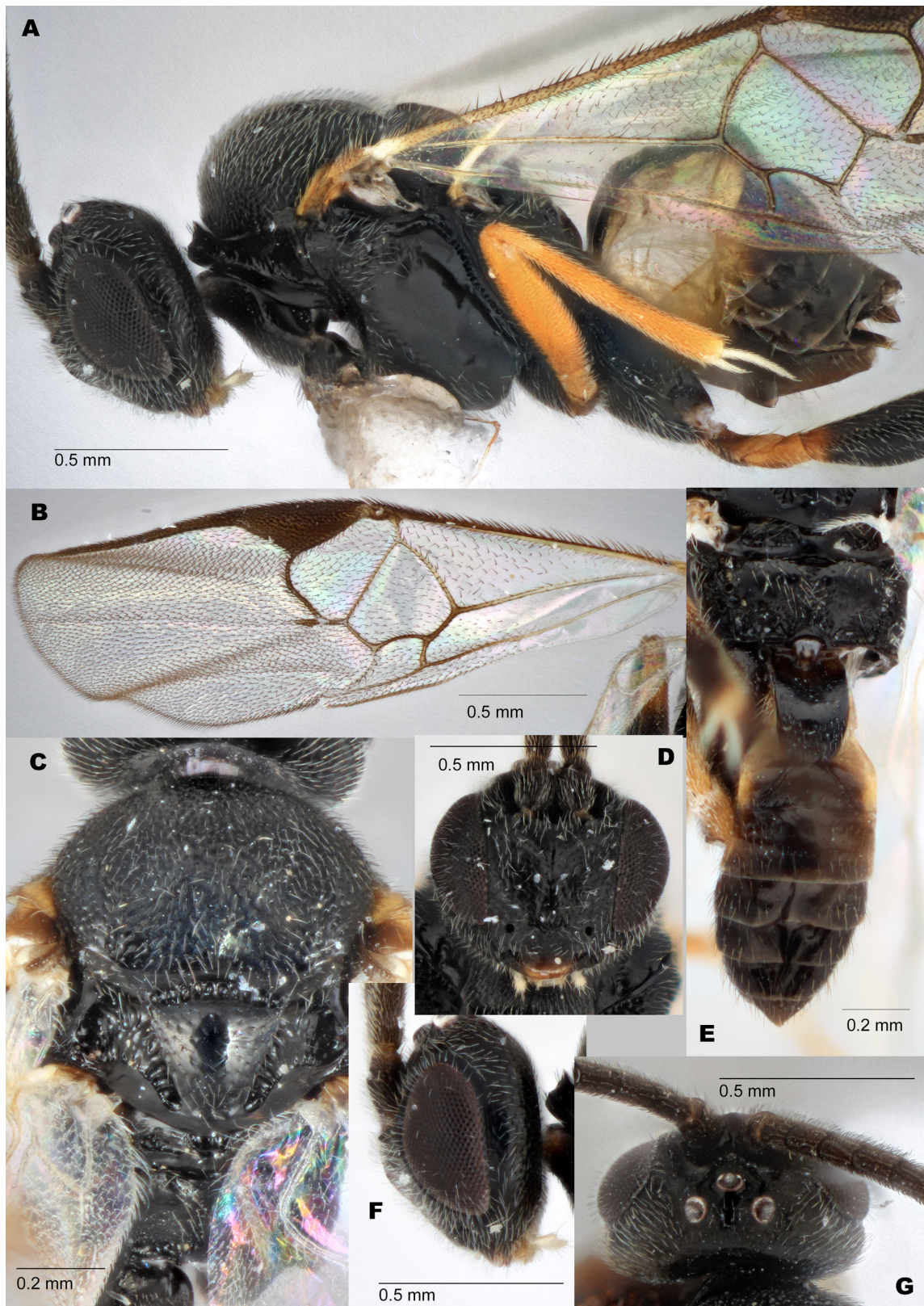


Fig. 39. *Glyptapanteles kingae* Fagan-Jeffries, Bird & Austin sp. nov. A, C–D, F. Paratype, ♀ (SAMA 32-46155). B, E, G. Holotype, ♀ (SAMA 32-46154) A. Lateral mesosoma. B. Fore wing. C. Dorsal mesosoma. D. Anterior head. E. Dorsal metasoma. F. Lateral head. G. Dorsal head.

METASOMA. T1 broadest at centre of length, narrowing both posteriorly and anteriorly from mid-point; T1 smooth; T1 length 0.31 mm; T1 width at posterior edge 0.11 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.23 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, eight pits in scutellar sulcus.

Remarks

Glyptapanteles kingae sp. nov. constitutes BIN BOLD:ADL5629 and is 9.94% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL2952; *Glyptapanteles guzikae* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 10.1% different from the most similar *COI* sequence from an Australian specimen (AUMIC524-18; *Glyptapanteles guzikae* sp. nov.). All four specimens of the type series were sequenced for the *wingless* gene, which is identical amongst all specimens (although there is a single ambiguity in the WA specimen sequence). The *wingless* sequences differ by a minimum of 19 bp from all other species with available sequence data.

Distribution

This species has a wide distribution, from the Pilbara in WA, through to the arid region of SA and in the ACT at Black Mountain Reserve.

Glyptapanteles kittelae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:97C25961-9321-4C68-83FD-3563AADE8B60](https://zoobank.org/act:97C25961-9321-4C68-83FD-3563AADE8B60)
 Figs 5C, 7B, 8B, 9B, 40

Diagnosis

Glyptapanteles kittelae sp. nov. is in the *G. albigena* species group and can be separated from most of the other members of the species group by the pale spot on the gena being small but clearly visible, T2 dark, the media carina completely absent, and the hind femur light brown to pale.

With the limited specimens available, there was not a morphological character found that easily differentiates *G. kittelae* sp. nov. from *G. austrinus* sp. nov. or *G. aspersus* sp. nov. The *COI* divergence between *G. kittelae* sp. nov. and *G. austrinus* sp. nov. is > 6%, whilst the divergence between *G. kittelae* sp. nov. and *G. aspersus* sp. nov. is > 7%. The *wingless* sequences of *G. kittelae* sp. nov. and *G. aspersus* sp. nov. are very distinct, differing by 7 bp; however, there are no *wingless* sequences available for *G. austrinus* sp. nov.

Etymology

This species is named for Dr Rebecca Kittel, who collected the holotype and many other important and useful Hymenoptera, including many specimens of microgastrines, during her time studying Australian Cheloninae.

Material examined

Holotype

AUSTRALIA • ♀; South Australia, Witchelina Stn; -30.0186, 137.901; 23 Oct. 2010; R. Kittel leg.; Bush Blitz Svy RK091 sweeping *Acacia victoriae*; Extraction557, BOLD: AUMIC348-18; SAMA 32-46156.

Paratype

AUSTRALIA • 1 ♀; New South Wales, track off Oxley Highway, 15 km from Gingers Creek, roadside; -31.3655, 152.045278; 1116 m a.s.l.; 15 Dec. 2008; K. Harvey and K. Umbers; reared from flowers of *Senecio minimus* 1 Apr. 2009; Extraction1313, BOLD: AUGLY152-21; AM K379892.

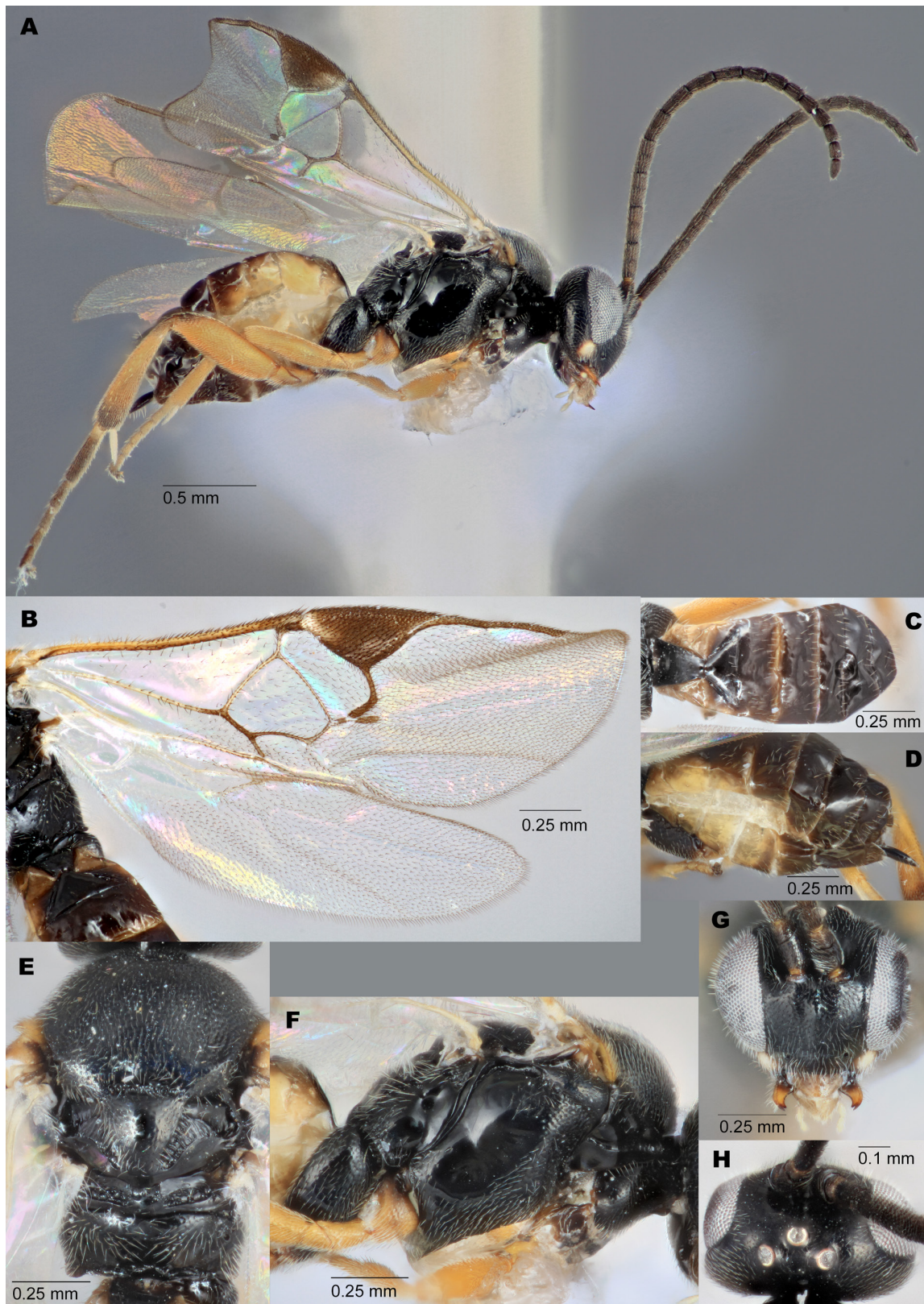


Fig. 40. *Glyptapanteles kittelae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (SAMA 32-46156). A. Lateral habitus. B. Fore wing. C. Dorsal metasoma. D. Lateral metasoma. E. Dorsal mesosoma. F. Lateral mesosoma. G. Anterior head. H. Dorsal head.

Description

Female

COLOURATION. Gena with a pale spot; labrum reddish-brown; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally or uniformly pale; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa light brown; hind coxa dark; fore femur orange to light brown; mid femur orange to light brown; hind femur orange to light brown; fore tibia orange to light brown; mid tibia orange to light brown; hind tibia darkening posteriorly; hind basitarsus dark reddish-brown; T1 dark; T2 sclerotised area dark; T2 lateral area same colour as sclerotised area, or only slightly paler; T3 uniformly brown; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.2 mm; fore wing length 2.1 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 14 length/width 1.12–1.40; antennal flagellomere 2 length/width 2.42–3.00; OOD/POD 1.71; IOD/POD 1.71.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures; scutellar disk sculpturing with only very shallow punctures; eight pits in scutellar sulcus; propodeum with median carina absent, very smooth and shiny, only very shallow punctures associated with setae.

WINGS. Pterostigma length 0.51 mm; pterostigma width 0.22 mm; r 0.13 mm; 2RS 0.14 mm; 2m 0.1 mm; (RS+M)b 0.07 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 smooth and shiny; T1 length 0.31 mm; T1 width at posterior edge 0.06 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.31 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles kittelae sp. nov. constitutes BIN: BOLD:ADL3407 and is 5.13% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL4089; *Glyptapanteles ferrugineus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 5.3% different from the most similar *COI* sequence from an Australian specimen (AUMIC042-18; *Glyptapanteles ferrugineus* sp. nov.). The holotype was able to be sequenced for the *wingless* gene and has a unique barcode, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

Glyptapanteles kittelae sp. nov. is known from central SA and eastern NSW.

Glyptapanteles kurandaensis Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:14DC77FB-9FEB-4DE4-8D02-4678EBB481E5](https://zoobank.org/act:14DC77FB-9FEB-4DE4-8D02-4678EBB481E5)
 Figs 12D, 14A, 41

Diagnosis

Glyptapanteles kurandaensis sp. nov. can be separated from the other species of *Glyptapanteles* described from Australia by having the gena without a pale spot, T1 and T2 dark, anteromesoscutum

punctures not extremely sparse and deep (i.e., not as in the *G. niveus* species group), no orange markings on the postero-lateral anteromesoscutum, propodeum without strong rugose sculpturing (sometimes with shallow sculpturing in centre, but never over most of propodeum), hind femur solidly dark in colouration, ventral side of antennal scape (at least in distal half) the same colour or darker than the flagellomeres, and fore wing veins r and 2SR narrow and smoothly joined, both significantly longer than vein 2m (compared to the sharply angled, thicker and shorter r and 2SR veins more typical of this genus in Australia).

Etymology

Named for the collection locality, Kuranda, a small town near Cairns in northern QLD.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Kuranda; -16.8135; 145.6430586; 317 m a.s.l.; 12 Feb.–6 Apr. 2020; M.S. Moulds leg.; Malaise Trap EFJ2020MT36; Extraction1568, BOLD: AUGLY110-21; QM T250958.

Paratypes

AUSTRALIA – **Queensland** • 1 ♀; same collection data as for holotype: Extraction1520, BOLD: AUGLY092-21; QM T250959 • 1 ♀; same collection data as for holotype; Extraction1542, BOLD: AUGLY098-21; QM T250960 • 1 ♀; same collection data as for holotype; Extraction1543, BOLD: AUGLY099-21; QM T250961 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1546, BOLD: AUGLY101-21; QM T250962 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1551, BOLD: AUGLY103-21; QM T250963 • 1 ♀; same collection data as for holotype; Extraction1554, BOLD: AUGLY104-21; QM T250964 • 1 ♀ (ethanol); same collection data as for holotype; Extraction1565, BOLD: AUGLY109-21; QM T250965 • 1 ♀; same collection data as for holotype; Extraction1569, BOLD: AUGLY111-21; QM T250966 • 1 ♀; same collection data as for holotype; Extraction1570, BOLD: AUGLY112-21; QM T250967 • 1 ♀; same collection data as for holotype; Extraction1508, BOLD: AUGLY129-21; QM T250968 • 1 ♀; same collection data as for holotype; Extraction296, BOLD: AUMIC161-18; QM T250969 • 1 ♀ (ethanol); same collection data as for holotype; 10 Feb.–15 Mar. 2017; Extraction500, BOLD: AUMIC306-18; QM T250970 • 1 ♀; same collection data as for holotype; 16 Mar.–12 Apr. 2017; Extraction551, BOLD: AUMIC342-18; QM T250971.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark or reddish-brown; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres darkening distally; tegula dark; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur dark reddish-brown or dark; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark or dark reddish-brown; T2 lateral area same colour as sclerotised area, or only slightly paler, or dark extends past indentation, but then pale; T3 dark, mostly dark with paler lateral areas or uniformly brown; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 1.8 mm; fore wing length 1.9 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 14 length/width 1.80–2.25; antennal flagellomere 2 length/width 2.42–3.80; OOD/POD 1.86–2.33; IOD/POD 1.29–2.00.

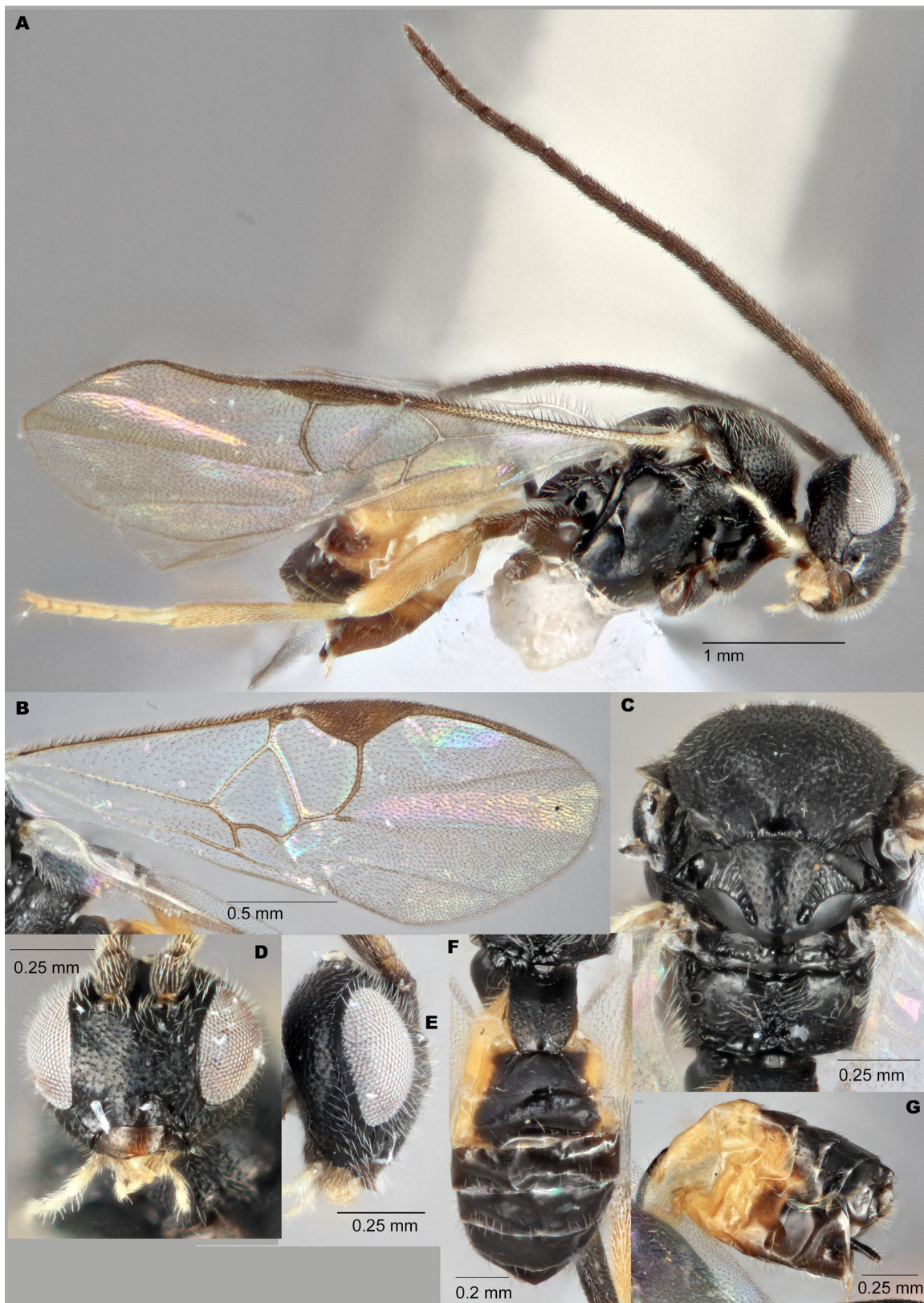


Fig. 41. *Glyptapanteles kurandaensis* Fagan-Jeffries, Bird & Austin sp. nov., paratypes, ♀. **A–B,** **D–E.** (QM T250969). **C, F–G.** QM T250971. **A.** Lateral habitus. **B.** Fore wing. **C.** Dorsal mesosoma. **D.** Anterior head. **E.** Lateral head. **F.** Dorsal metasoma. **G.** Lateral metasoma.

MESOSOMA. Anteromesoscutum sculpturing with shallow punctures, space between punctures generally smaller than diameter of punctures, slightly denser and deeper punctures anteriorly, smoother in posterior centre; scutellar disk sculpturing with only very shallow punctures; 7–9 pits in scutellar sulcus; propodeum with median carina absent, some indistinct sculpturing in posterior centre, or in centre along length of propodeum, or median carina faintly indicated for all of length by indistinct sculpturing.

WINGS. Pterostigma length 0.48 mm; pterostigma width 0.15 mm; r 0.17 mm; 2RS 0.12 mm; 2m 0.09 mm; (RS+M)b 0.1 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly, or lateral edges parallel for entirety of length, posterior corners sometimes rounded at boundary with T2; T1 mostly smooth, some punctures in posterior half or mostly smooth, some shallow rugose sculpturing in posterior half; T1 length 0.29 mm; T1 width at posterior edge 0.14 mm; T2 an isosceles trapezoid, lateral edges straight, with curved lateral and anterior edges, becoming arch- or semicircle-shaped or almost square, lateral edges almost parallel, only broadening posteriorly very slightly; T2 smooth and shiny; T2 length 0.13 mm; T2 width at posterior edge 0.24 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles kurandaensis sp. nov. constitutes BIN BOLD:ADL2798 and is 2.4% (p-dist.) divergent from the closet BIN in the database (BOLD:AAH1268, *Glyptapanteles creatonoti* (Viereck, 1912)). We have examined the images of *G. creatonoti* in Gupta *et al.* (2016) and are confident that *G. kurandaensis* sp. nov. is a different species, most easily separated by the following morphological characteristics: *G. kurandaensis* sp. nov. has the hind coxa dark (*G. creatonoti* hind coxa pale; “stamineous” in original description); *G. kurandaensis* sp. nov. has the propodeum much smoother than that of *G. creatonoti* and *G. kurandaensis* sp. nov. has T1 completely black, whilst *G. creatonoti* has the anterior area of T1 pale (“stamineous” in original description, appearing pale orange in images of Gupta *et al.* (2016)).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 7.7% different from the most similar *COI* sequence from an Australian specimen (AUMIC291-18; *Glyptapanteles austini* sp. nov.). The type specimen was sequenced for the *wingless* gene, which differs by a minimum of 14 bp from all other species with available sequence data.

Distribution

This species is currently known from one collection site; Kuranda in north-eastern QLD.

Glyptapanteles lambkinae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:AB18FD64-2820-46A6-A7CD-F3869B4CA991](https://zoobank.org/act:AB18FD64-2820-46A6-A7CD-F3869B4CA991)

Fig. 42

Diagnosis

Glyptapanteles lambkinae sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the species group, other than from *G. erucadesolator* sp. nov., by having the propodeum with very coarse and strong rugose sculpting and T1 with strong sculpturing. *Glyptapanteles arcanus* sp. nov., *G. vergrandiacus* sp. nov. and *G. goodwinnoakes* sp. nov. also have coarse sculpturing on the propodeum (although to a lesser degree), but T1 is either smooth or with clear punctures (but not rugose sculpturing). It is noted, however, that all these species are morphologically very similar and identifications should be made with DNA barcodes. There was not a morphological character found to

easily differentiate *G. lambkinae* sp. nov. from *G. erucadesolator* sp. nov., which are sister lineages in the current phylogeny (Fig. 2); however, as the species differ by 2 bp in the *wingless* sequence and the *COI* barcodes are >6% divergent, we feel confident that these are different species.

Etymology

Named for Dr Christine Lambkin, who collected the type material and EPF-J and ARD thank her for all her support and help over the years.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Lamington National Park; -28.155, 153.139; 282 m a.s.l.; 13–23 Jan. 2007; C. Lambkin and N. Starick leg.; IBISCA Plot # IQ-300-B, rainforest, Malaise trap; bulk vial 22139; Extraction601, BOLD: AUMIC387-18; QM T208402.

Paratypes

AUSTRALIA • 1 ♀; same collection data as for holotype; Extraction1467, BOLD: AUGLY085-21; QM T250972 • 1 ♀; same collection data as for holotype; Extraction1471, BOLD: AUGLY089-21; QM T250973 • 1 ♀ (ethanol); same collection data as for holotype; Extraction599, BOLD: AUMIC383-18; QM T208401.

Description

Female

COLOURATION. Gena without a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly reddish-brown; anteromesoscutum all dark or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur orange to light brown; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus dark reddish-brown; T1 dark; T2 sclerotised area dark reddish-brown; T2 lateral area dark extends past indentation, but then pale; T3 mostly pale with darker patch in centre or mostly dark with paler lateral areas; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.7 mm; fore wing length 2.5 mm; antennal length similar to body length.

HEAD. Antennal flagellomere 14 length/width 1.12–1.62; antennal flagellomere 2 length/width 3.00–3.85; OOD/POD 1.43–1.71; IOD/POD 1.33–1.71.

MESOSOMA. Anteromesoscutum sculpturing with deep punctures, space between punctures a mixture of smaller than diameter of punctures, and of similar size; scutellar disk sculpturing with deep, sparse, irregularly spaced punctures, more common on anterior lateral edges; 10–11 pits in scutellar sulcus; propodeum with median carina absent, strongly rugose, sometimes with punctures in centre.

WINGS. Pterostigma length 0.62 mm; pterostigma width 0.21 mm; r 0.19 mm; 2RS 0.17 mm; 2m 0.11 mm; (RS+M)b 0.12 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly; T1 coarsely sculptured; T1 length 0.46 mm; T1 width at posterior edge 0.09 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.16 mm; T2 width at posterior edge 0.27 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

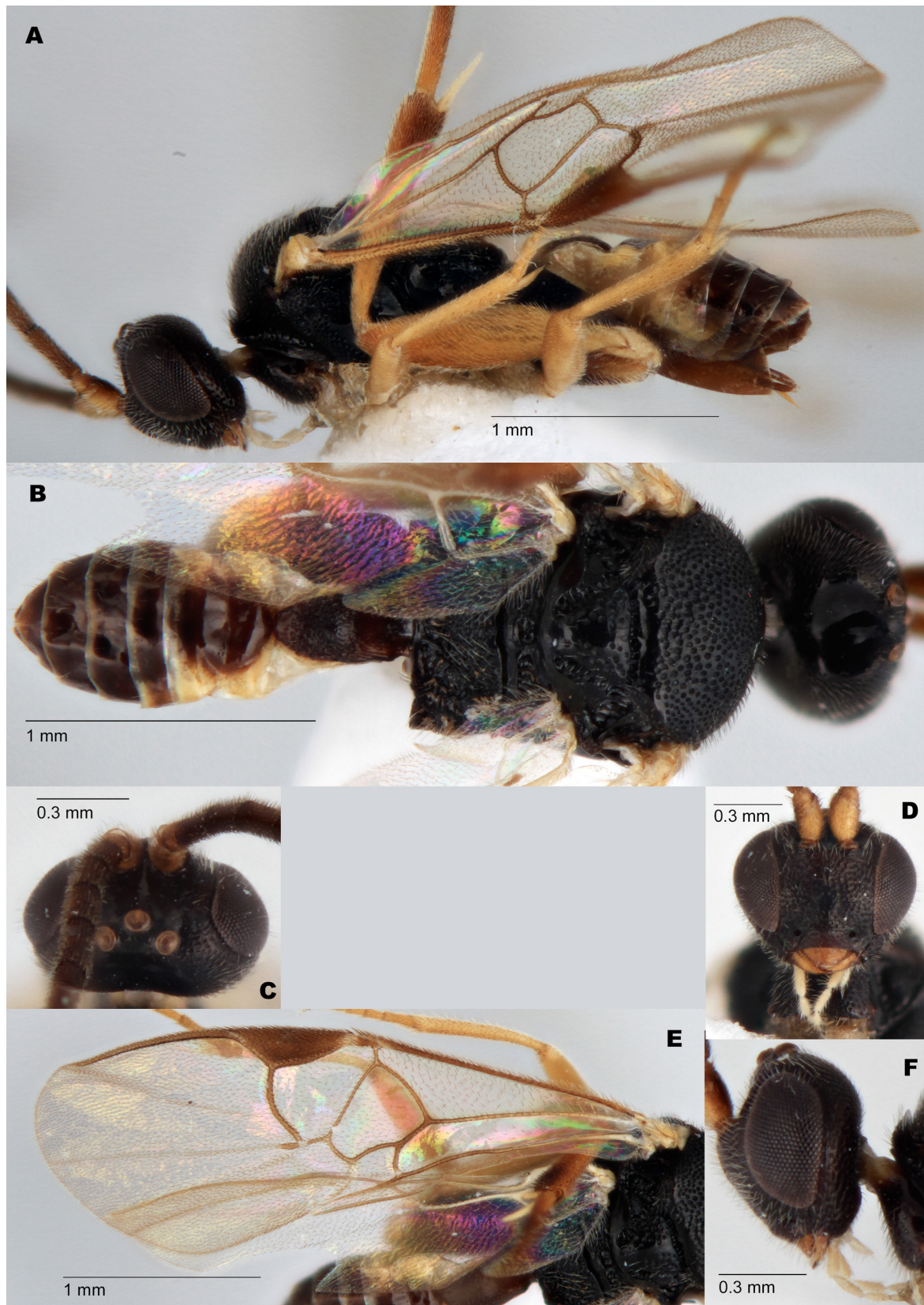


Fig. 42. *Glyptapanteles lambkinae* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250973). A. Lateral habitus. B. Dorsal habitus. C. Dorsal head. D. Anterior head. E. Fore wing. F. Lateral head.

Remarks

Glyptapanteles lambkinae sp. nov. constitutes BIN BOLD:ADL3041 and is 6.09% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL3293; *Glyptapanteles wrightae* sp. nov.).

Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 6.1% different from the most similar *COI* sequence from an Australian specimen (AUGLY141-21; an undescribed lineage, with a single specimen). All four five specimens were sequenced for the *wingless* gene, which is identical within the species and differs by a minimum of 2 bp from all other species with available sequence data.

Distribution

This species is known only from Lamington National Park in southern QLD.

Glyptapanteles lessardi Fagan-Jeffries, Bird & Austin sp. nov.

[urn:lsid:zoobank.org:act:3DDE9CAF-3AD2-49C2-BBD6-875FF512E6C9](https://zoobank.org/act:3DDE9CAF-3AD2-49C2-BBD6-875FF512E6C9)

Fig. 43

Diagnosis

Glyptapanteles lessardi sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the group as follows:

Glyptapanteles lessardi sp. nov. can be separated from *G. rodriguezae* sp. nov. and *G. ruhri* sp. nov. by T1 being smooth and shiny, not having punctures that cover at least a third of the area of the posterior half of the tergite.

Glyptapanteles lessardi sp. nov. can be separated from *G. goodwinnoakes* sp. nov., *G. erucadesolator* sp. nov., *G. lambkinae* sp. nov., *G. arcanus* sp. nov. and *G. vergrandiacus* sp. nov. by the propodeum being less coarsely (and less consistently), strongly rugose sculptured across the anterior half of the propodeum. *Glyptapanteles wrightae* sp. nov. has the propodeum with strong punctures in the anterior half, the posterior half with shallow or strong rugose sculpturing.

Glyptapanteles lessardi sp. nov. can be separated from *G. doreyi* sp. nov. by having the tegula pale in colouration (the tegula in *G. doreyi* sp. nov. is generally dark in colouration), the hind femur mostly pale or light brown (mostly dark in *G. doreyi* sp. nov.) and the indentation in the centre of the mesopleuron being smooth, not strongly canaliculated as it is in *G. doreyi* sp. nov.

There was not a morphological character found that easily separates *G. lessardi* sp. nov. from *G. wrightae* sp. nov.; we do not diagnose these species morphologically. The *wingless* barcodes of the two species differ by 3 bp and the *COI* is greater than 4% divergent.

Etymology

Named for Dr Bryan Lessard, part of the team who collected the holotype, for his constant endeavour to raise awareness about the importance of taxonomy in the community and his support of other early career researchers.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Blue Tongue Scrub, Steve Irwin Reserve, Cape York; -12.286584, 142.343274; 42.9 m a.s.l.; 23–26 Jul. 2018; C. Lambkin and B. Lessard leg.; vine thicket rainforest; Extraction888, BOLD: AUGLY015-21; QM T250974.

Paratype

AUSTRALIA • 1 ♀; same collection data as for preceding; Extraction890, BOLD: AUGLY016-21; QM T250975.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres uniformly reddish-brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with significant orange patches on posterolateral corners or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark reddish-brown; T2 sclerotised area dark reddish-brown; T2 lateral area dark extends past indentation, but then pale; T3 uniformly brown; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 1.9 mm; fore wing length 1.8 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 1.14–1.28; antennal flagellomere 2 length/width 3.00–3.16; OOD/POD 2.17–2.20; IOD/POD 1.80–1.83.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size, often smoother in posterior centre; scutellar disk sculpturing with shallow punctures scattered over most of area; seven pits in scutellar sulcus; propodeum with median carina absent and with punctured sculpturing in ‘v’ shape in centre of propodeum, or with shallow sculpturing over most of propodeum other than smoother area in anterior centre.

WINGS. Pterostigma length 0.46 mm; pterostigma width 0.15 mm; r 0.15 mm; 2RS 0.12 mm; 2m 0.09 mm; (RS+M)b 0.05 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 smooth and shiny; T1 length 0.32 mm; T1 width at posterior edge 0.09 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.12 mm; T2 width at posterior edge 0.3 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles lessardi sp. nov. constitutes BIN BOLD:AEI3823 and is 4.49% (p-dist.) divergent from the closet BIN in the database (BOLD:AEI5416, an undescribed lineage from Australia, possibly part of *G. wrightae* sp. nov. (see details under *G. wrightae* sp. nov. Remarks section).

Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 4.8% different from the most similar *COI* sequence from an Australian specimen (AUMIC084-18; *Glyptapanteles wrightae* sp. nov.).

Distribution

Currently known only from a single reserve in Cape York, the most northern region of QLD.

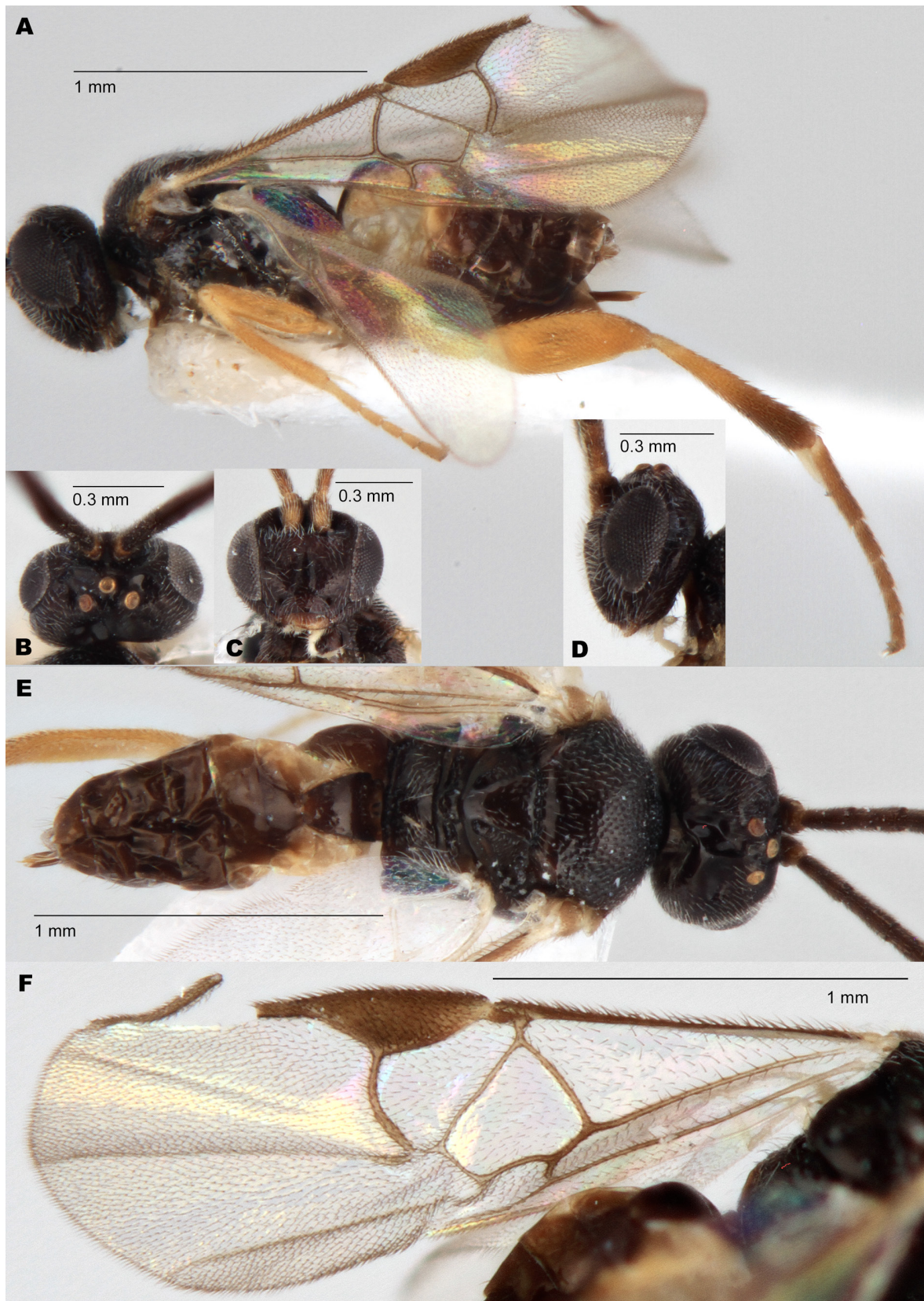


Fig. 43. *Glyptapanteles lessardi* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QM T250975). A. Lateral habitus. B. Dorsal head. C. Anterior head. D. Lateral head. E. Dorsal habitus. F. Fore wing.



Fig. 44. *Glyptapanteles mnesampela* Austin, 2000 holotype, ♀ (ANIC 32-141445). A. Lateral habitus. B. Dorsal propodeum and metasoma. C. Anterior head. Images courtesy of O. Evangelista (ANIC).

Glyptapanteles mnesampela Austin, 2000

Figs 6A, 44

Diagnosis

Glyptapanteles mnesampela is in the *G. albigena* species group and can be separated from the other members of the species group by having a pale T2, whilst all other described species with a pale spot on the gena have T2 dark in colouration.

Material examined

Holotype

AUSTRALIA • ♀; Australian Capital Territory, Lyneham Ridge; 30 Feb. 1993; ex *Mnesampela privata*; 22 Jun. 1993; ANIC 32-141445. (Only images examined).

Remarks

Reared from *Mnesampela privata* Guenée, 1858 (Geometridae).

Distribution

Known from Australia, currently only from the ACT.

Glyptapanteles mouldsi Fagan-Jeffries, Bird & Austin sp. nov.

[urn:lsid:zoobank.org:act:03F851F6-74B6-4195-8254-B4BA372F1361](https://zoobank.org/act:03F851F6-74B6-4195-8254-B4BA372F1361)

Figs 6D, 11A–B, 45

Diagnosis

Glyptapanteles mouldsi sp. nov. is in the *G. mouldsi* species group and can be separated from the other two species in the group (*G. rixi* sp. nov. and *G. dowtoni* sp. nov.) by the metasomal sternites being all pale yellow other than the hypopygium, which is dark and T4–5 all pale/yellow. *Glyptapanteles rixi* sp. nov. and *G. dowtoni* sp. nov. have S5+ dark and T4–6 with at least some dark areas, or completely dark. The *wingless* sequences of the three species differ by at least 6 bp and the *COI* differs by at least 4.3%.

Etymology

This species is named for Dr Max Moulds, who collected the type series, along with many other specimens, whilst kindly running a Malaise trap for this and other microgastrine projects at Kuranda, north QLD.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Kuranda; -16.8135, 145.6430586; 317 m a.s.l.; 12 Feb.–6 Apr. 2020; M.S. Moulds leg.; Malaise Trap EFJ2020MT36; Extraction1562, BOLD: AUGLY108-21; QM T250976.

Paratypes

AUSTRALIA • 1 ♀; same collection data as for holotype; Extraction1559, BOLD: AUGLY106-21; QM T250977 • 1 ♀; same collection data as for holotype; 16 Mar.–12 Apr. 2017; Extraction552, BOLD: AUMIC343-18; QM T250978 • 1 ♀; same collection data as for holotype; 18 Apr.–8 Jul. 2017; Extraction735, BOLD: AUMIC488-18; QM T250979.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark or reddish-brown; scape colour in ventral half either uniformly paler than flagellomeres, or the same colour or darker than flagellomeres;

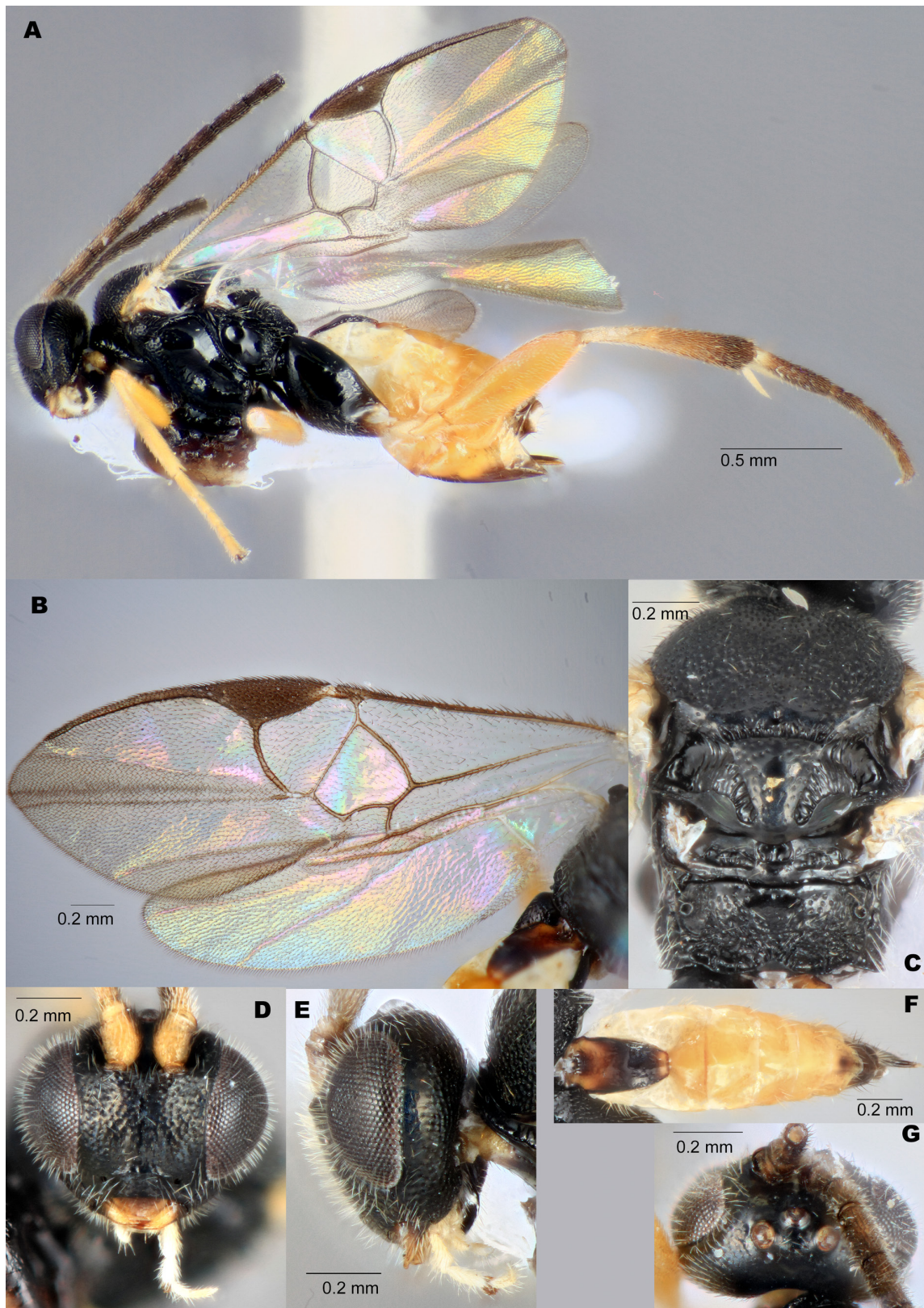


Fig. 45. *Glyptapanteles mouldsi* Fagan-Jeffries, Bird & Austin sp. nov., paratypes, ♀. **A–F.** QM T250978. **G.** QM T250979. **A.** Lateral habitus. **B.** Fore wing. **C.** Dorsal mesosoma. **D.** Anterior head. **E.** Lateral head. **F.** Dorsal metasoma. **G.** Dorsal head.

flagellomeres darkening distally; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur orange to light brown; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus dark reddish-brown; T1 dark or dark reddish-brown; T2 sclerotised area pale; T2 lateral area same colour as sclerotised area, or only slightly paler; T3–T5 pale, T6–7 darkening posteriorly.

HOLOTYPE BODY MEASUREMENTS. Body length 2.3 mm; fore wing length 2.2 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 1.83–2.16; antennal flagellomere 2 length/width 1.77–3.00; OOD/POD 1.37–1.71; IOD/POD 1.57–1.75.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size. Often smoother in posterior centre; scutellar disk sculpturing with deep or shallow, sparse, irregularly spaced punctures, more common on anterior lateral edges; 9–12 pits in scutellar sulcus; propodeum with median carina absent, strong punctures in anterior half, rugose sculpturing in posterior half.

WINGS. Pterostigma length 0.54 mm; pterostigma width 0.18 mm; r 0.28 mm; 2RS 0.12 mm; 2m 0.05 mm; (RS+M)b 0.1 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{4}$ of length, then gently narrowing posteriorly, lateral edges often with slight curve or lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly; T1 smooth and shiny, sometimes with shallow sculpturing along lateral edges or some shallow scattered punctures on lateral edges; T1 length 0.3 mm; T1 width at posterior edge 0.09 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.21 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles mouldsi sp. nov. constitutes BIN: BOLD:ADL3640 and is 4.87% (p-dist). divergent from the closet BIN in the database (BOLD:AEI8040, an undescribed lineage from Australia, with one specimen).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 5.2% different from the most similar *COI* sequence from an Australian specimen (AUGLY141-21; an undescribed lineage, with a single specimen). All four of the type specimens were able to be sequenced for the *wingless* gene and share a unique barcode, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

This species is currently known from Kuranda in QLD.

Glyptapanteles niveus Fagan-Jeffries, Bird & Austin sp. nov.

[urn:lsid:zoobank.org:act:DB934891-C1E7-4C64-97A8-0F1E5D31D6B1](https://zoobank.org/act:DB934891-C1E7-4C64-97A8-0F1E5D31D6B1)

Fig. 46

Diagnosis

Glyptapanteles niveus sp. nov. is in the *G. niveus* species group and can be differentiated from *G. bradfordae* sp. nov. by the presence of white distal flagellomeres (at least flagellomeres 10–11), a dark labrum and

sparse punctures on the hind coxa (*G. bradfordae* sp. nov. has the flagellomeres 10–11 dark, the labrum pale in colour and denser punctures on the hind coxa). *Glyptapanteles niveus* sp. nov. can be separated from *G. cooperi* sp. nov. by having a dark labrum (*G. cooperi* sp. nov. has the labrum pale in colour).

Etymology

The species epithet ‘*niveus*’ is a Latin adjective meaning ‘snowy’ and refers to the white flagellomeres and white fore coxa, mid coxa, hind- trochanter and anterior metasomal sternites.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Kuranda; -16.8135, 145.6430586; 317 m a.s.l.; 12 Feb.–6 Apr. 2020; M.S. Moulds leg.; Malaise Trap EFJ2020MT36; Extraction1527, BOLD: AUGLY096-21; QM T250980.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark; scape colour in ventral half the same colour or darker than flagellomeres; proximal nine flagellomere segments dark, at least some distal segments (flagellomeres 10–11) white (final five segments missing from type so colour unknown); tegula dark; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum dark with significant orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa white; mid coxa white; hind coxa dark; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia light brown; hind basitarsus light brown; T1 dark; T2 sclerotised area dark reddish-brown; T2 lateral area dark extends past indentation, but then pale; T3 uniformly brown; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.1 mm; fore wing length 1.8 mm; antennal length slightly shorter than body length.

HEAD. Antennal flagellomere 2 length/width 4.33; antennal flagellomere 14 missing; OOD/POD 1.57–1.83; IOD/POD 1.17.

MESOSOMA. Anteromesoscutum sculpturing with very sparse, deep punctures; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; seven pits in scutellar sulcus; propodeum with median carina present, in some parts difficult to differentiate from surrounding course rugosity.

WINGS. Pterostigma length 0.52 mm; pterostigma width 0.14 mm; r 0.1 mm; 2RS 0.11 mm; 2m 0.09 mm; (RS+M)b 0.07 mm.

METASOMA. T1 lateral edges parallel for entirety of length, posterior corners rounded at boundary with T2; T1 mostly smooth, some punctures in posterior half; T1 length 0.32 mm; T1 width at posterior edge 0.15 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth, with some very shallow punctures in posterior half; T2 length 0.11 mm; T2 width at posterior edge 0.28 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles niveus sp. nov. constitutes BIN BOLD:AEI1197 and is 3.55% (p-dist.) divergent from the closet BIN in the database (BOLD:ADD5918, a BIN containing one specimen (GMNGU216-16) from Papua New Guinea).

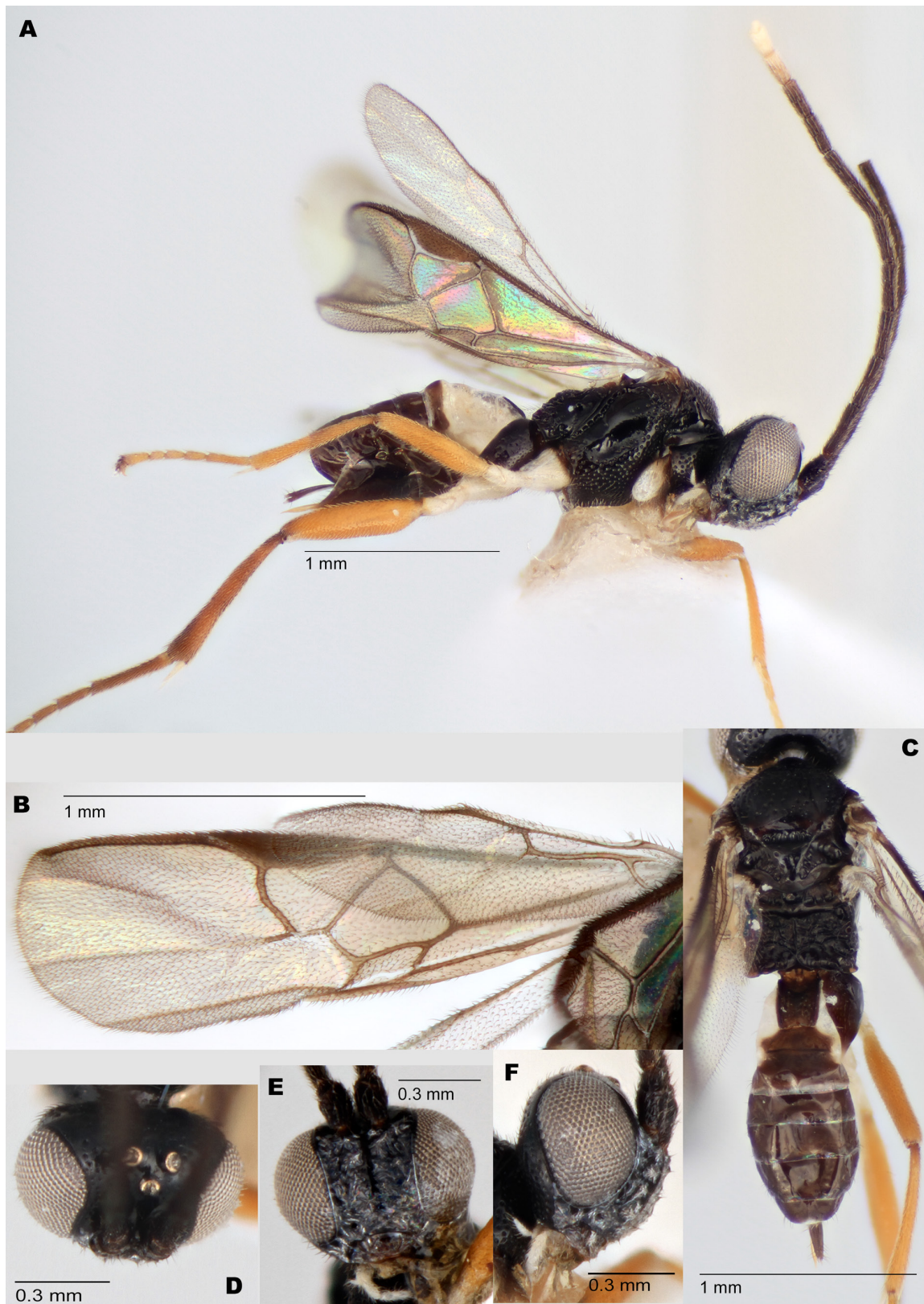


Fig. 46. *Glyptapanteles niveus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250980). A. Lateral habitus. B. Fore wing. C. Dorsal habitus. D. Dorsal head. E. Anterior head. F. Lateral head.

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 5.8% different from the most similar *COI* sequence from an Australian specimen (GMAQJ131-17; an undescribed lineage, with two specimens). The type specimen was sequenced for the *wingless* gene and the sequence differs by a minimum of 3 bp from all other species with available sequence data.

Distribution

This species is only known from Kuranda in northern QLD.

Glyptapanteles operculinae (Fullaway, 1941)

Fig. 16D

Diagnosis

This species has a distinctive scutellum, with the lateral band extremely broad so that the lateral scutellum is compressed into a narrow crenulated sulcus. This, along with the T1 being extremely sharply narrowing posteriorly from a broad anterior width, means this species is very unlikely to be conspecific with any of the newly described Australian species.

Material examined

No specimens examined; illustrations and redescription in Austin & Dangerfield (1992) used as reference (Fig. 16D).

Distribution

American Samoa, Western Samoa.

Glyptapanteles phytometrae (Wilkinson, 1928)

Fig. 47C, E

Diagnosis

Glyptapanteles phytometrae can be separated from many of the Australian species as follows: from the *G. albigena* species group by having the gena without a pale spot, from the *G. mouldsi* and *G. eburneus* species groups by having T1–2 dark and from the *niveus* species group by having a relatively smooth anteromesoscutum. The propodeum of *G. phytometrae* is mostly smooth with rugose sculpturing in the centre (Fig. 47C), which is distinct (at least on the available specimens) from any of the newly described species from Australia. Whilst propodeal sculpturing can vary slightly amongst individuals, we do not consider it likely that *G. phytometrae* is conspecific with any of the Australian species.

Material examined

Holotype

SAMOA • ♀; “Samoa Is. P.A. Buxton and G.H. Hopkins”; “Pres. by Imp. Bur. Ent. Brit. Mus. 1928-318”; NHMUK: 3.c.1016 (only images examined).

Remarks

Known hosts in Samoa include *Chrysodeixis eriosoma* (Doubleday, 1843) and an unknown Plusiinae (Noctuidae).

Distribution (in the Australasian region)

Samoa, Fiji, Sumatra (Austin & Dangerfield 1992).



Fig. 47. A–B, D. *Glyptapanteles taylori* (Wilkinson, 1928), holotype, ♀ (NHMUK 3.c.1018). C, E. *Glyptapanteles phytometrae* (Wilkinson, 1928), holotype, ♀ (NHMUK 3.c.1016). A. Lateral habitus. B. Dorsal propodeum and metasoma. C. Dorsal habitus. D. Dorsal mesosoma. E. Lateral habitus and fore wing. Images © The Trustees of the Natural History Museum, London (CC-BY 4.0).

Glyptapanteles rixi Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:4BD5D589-8647-4922-8CC3-3457114C71AC](https://zoobank.org/act:4BD5D589-8647-4922-8CC3-3457114C71AC)
Figs 6C, 10B, 11E–F, 48

Diagnosis

Glyptapanteles rixi sp. nov. can be separated from *G. mouldsi* sp. nov. by having S5+ dark and T4–6 completely dark. *Glyptapanteles mouldsi* sp. nov. has the metasomal sternites all pale yellow other than the hypopygium, which is dark and T4–5 all pale/yellow. *Glyptapanteles rixi* sp. nov. can be separated from *G. dowtoni* sp. nov. by T4 being solidly dark (mostly pale in *G. dowtoni* sp. nov.) and T1 being orange to light brown (dark in *G. dowtoni* sp. nov.). The *wingless* sequences of the three species differ by at least 6 bp and the *COI* differs by at least 4.3%.

Etymology

This species is named for Dr Mike Rix, who collected the type series, along with many other specimens, whilst kindly running a Malaise trap for this and other microgastrine projects at Toohey State Forest in Brisbane.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Toohey State Forest; -27.5375, 153.057; 11 Dec. 2016–2 Jan. 2017; M. Rix leg.; Malaise trap; Extraction283, BOLD: AUMIC150-18; QM T250981.

Paratypes

AUSTRALIA • 1 ♀; same collection data as for holotype; Extraction282, BOLD: AUMIC149-18; QM T250982 • 1 ♂; same collection data as for holotype; Extraction1284, BOLD: AUGLY075-21; QM T250983 • 1 ♂; same collection data as for holotype; 29 Nov.–11 Dec. 2016; Extraction1285, BOLD: AUGLY076-21; QM T250984 • 1 ♂; same collection data as for holotype; Extraction1286, BOLD: AUGLY077-21; QM T250985.

Description

Female

COLOURATION. Gena without a pale spot; labrum mostly dark or reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres darkening distally; tegula pale or light brown; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur orange to light brown; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 orange to light brown; T2 sclerotised area dark reddish-brown, pale or orange to light brown; T2 lateral area same colour as sclerotised area, or only slightly paler or much paler; T3 mostly pale with darker patch in centre, mostly dark with paler lateral areas or uniformly brown; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.0 mm; fore wing length 2.0 mm; antennal length similar to body length.

HEAD. Antennal flagellomere 14 length/width 1.75; antennal flagellomere 2 length/width 2.89–4.00; OOD/POD 1.63–1.86; IOD/POD 1.38–2.00.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size, often smoother in posterior centre; scutellar disk sculpturing with deep, sparse, irregularly spaced punctures, more common on anterior lateral edges or shallow punctures scattered over most of area; 8–9 pits in scutellar sulcus; propodeum with median carina absent, shallow rugose sculpturing with some punctures in anterior corners or median carina absent, smooth in centre and postero-lateral corners, rugose elsewhere.

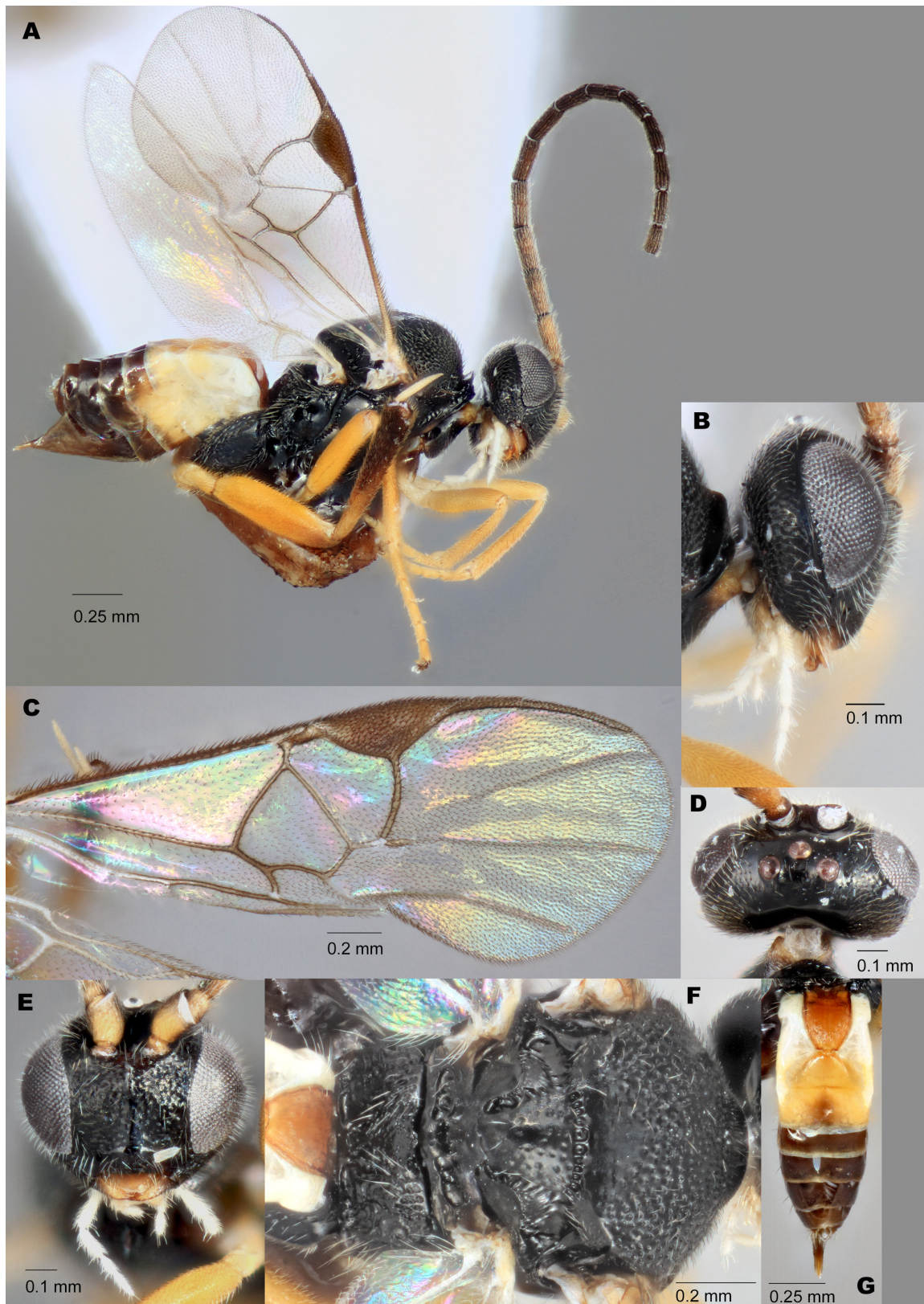


Fig. 48. *Glyptapanteles rixi* Fagan-Jeffries, Bird & Austin sp. nov. A–C, E–G. Holotype, ♀ (QM T250981). D. Paratype, ♀ (QM T250982). A. Lateral habitus. B. Lateral head. C. Fore wing. D. Dorsal head. E. Anterior head. F. Dorsal mesosoma. G. Dorsal metasoma.

WINGS. Pterostigma length 0.52 mm; pterostigma width 0.19 mm; r 0.18 mm; 2RS 0.13 mm; 2m 0.11 mm; (RS+M)b 0.12 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 mostly smooth, some punctures in posterior half; T1 length 0.32 mm; T1 width at posterior edge 0.08 mm; T2 an isosceles trapezoid, lateral edges straight or with curved lateral and anterior edges, becoming arch- or semicircle-shaped; T2 smooth and shiny; T2 length 0.15 mm; T2 width at posterior edge 0.32 mm; ovipositor slightly protruding from end of metasoma.

Male

As female.

Remarks

Glyptapanteles rixi sp. nov. constitutes BIN: BOLD:ADL2963 and is 3.69% (p-dist.) divergent from the closet BIN in the database (BOLD:AEI5416, an undescribed lineage from Australia, with two specimens).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 3.5% different from the most similar *COI* sequence from an Australian specimen (AUGLY139-21; an undescribed lineage). All five of the type specimens were able to be sequenced for the *wingless* gene and share a unique barcode, which differs by a minimum of 5 bp from all other species with available sequence data.

Distribution

This species is currently known from Toohey State Forest in Brisbane, QLD.

Glyptapanteles rodriguezae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:04BFEC74-5C1A-40F1-B10D-FAD918E18883](https://zoobank.org/act:04BFEC74-5C1A-40F1-B10D-FAD918E18883)

Fig. 49

Diagnosis

Glyptapanteles rodriguezae sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the species group, other than from *Glyptapanteles ruhri* sp. nov., by the hind femur being pale in colouration and propodeum punctate with smooth areas between punctures and with little-to-no rugosity and T1 punctured in the posterior half. There was no external morphological character found which easily separates *G. rodriguezae* sp. nov. from *G. ruhri* sp. nov. and we do not provide a morphological diagnosis between these two species. The two species are sister lineages in the current phylogeny (Fig. 2), but *Glyptapanteles ruhri* sp. nov. is 6 bp different in the *wingless* sequence from *G. rodriguezae* sp. nov. and >6% divergent in the *COI* barcode and we therefore feel confident that they are different species.

Etymology

This species is named for Dr Juanita Rodriguez, who was part of the team who collected the type specimen and who has an endlessly enthusiastic and collaborative approach to hymenopteran research.

Material examined

Holotype

AUSTRALIA • ♀; Australian Capital Territory, CSIRO Black Mountain; -35.2744, 149.1115; 6 Dec. 2017–5 Jan. 2018; J. Rodriguez, C. Waichert, K.M. Bayless and T. Pleines leg.; Malaise 2, green dry wash; Extraction979, BOLD: AUGLY031-21; ANIC 32 130332.

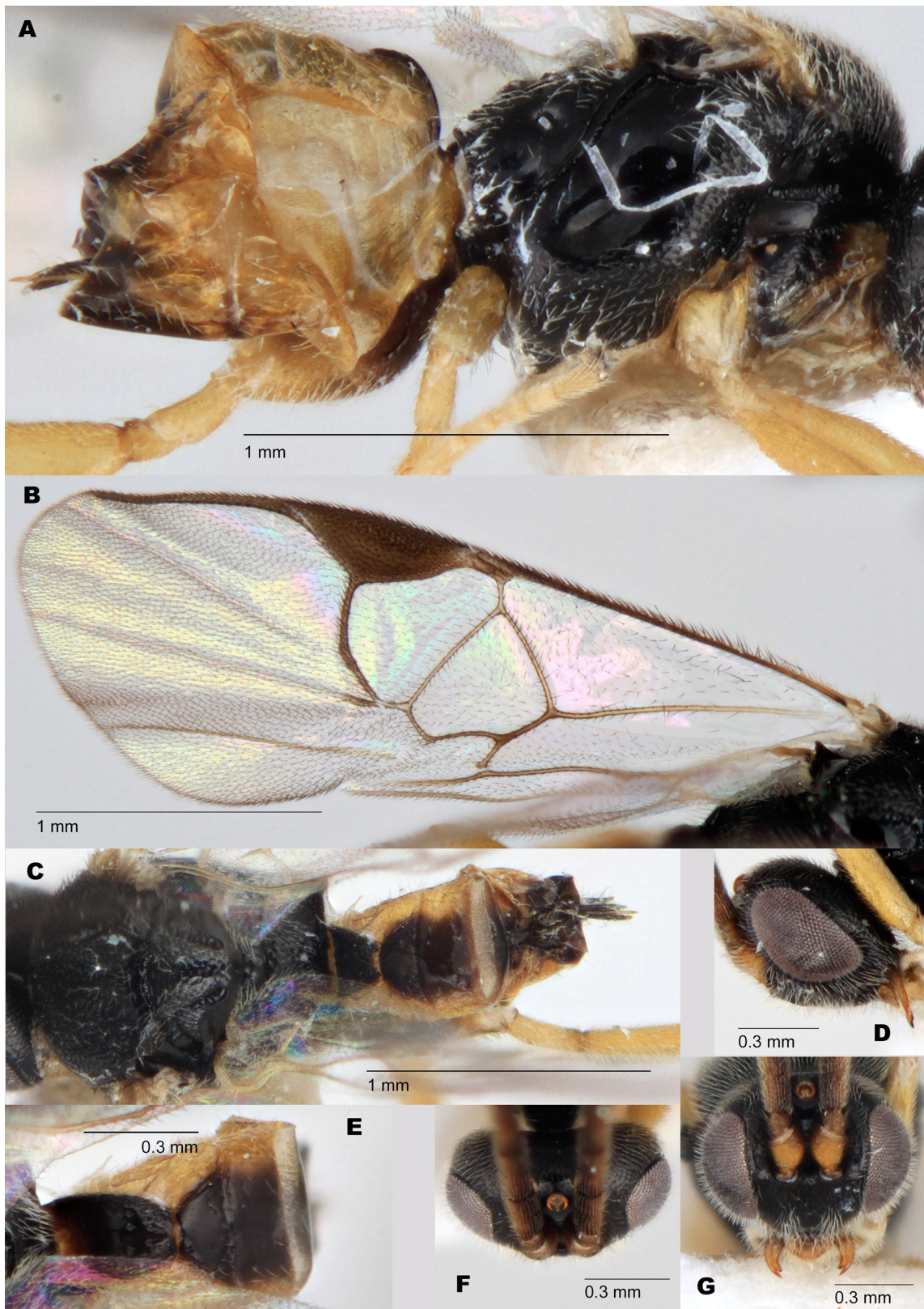


Fig. 49. *Glyptapanteles rodriguezae* Fagan-Jeffries, Bird & Austin sp. nov., **A, C, E.** Paratype, ♀ (ANIC 32 130333). **B, D, F–G.** Holotype, ♀ (ANIC 32 130332). **A.** Lateral habitus. **B.** Fore wing. **C.** Dorsal habitus. **D.** Lateral head. **E.** Dorsal metasoma. **F.** Dorsal head. **G.** Anterior head.

Paratype

AUSTRALIA • 1 ♀; Australian Capital Territory, CSIRO Black Mountain; -35.268, 149.1066; 650 m a.s.l.; 11–25 Feb. 2020; K.M. Bayless leg.; Y-shaped wash, Malaise trap collapsed by flood; Extraction1138, BOLD: AUGLY062-21; ANIC 32 130333.

Description

Female

COLOURATION. Gena without a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres all black/dark brown or darkening distally; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark; T2 lateral area much paler; T3 mostly pale with darker patch in centre or mostly dark with paler lateral areas; T4–5 pale with dark patch in centre, then darkening posteriorly on T6–7.

HOLOTYPE BODY MEASUREMENTS. Body length 2.4 mm; fore wing length 2.4 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 1.83–2.16; antennal flagellomere 2 length/width 2.87–3.00; OOD/POD 1.75–2.17; IOD/POD 1.38–2.00.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size, often smoother in posterior centre; scutellar disk sculpturing with shallow punctures scattered over most of area; 10–11 pits in scutellar sulcus; propodeum with median carina absent, propodeum punctate.

WINGS. Pterostigma length 0.63 mm; pterostigma width 0.22 mm; r 0.23 mm; 2RS 0.17 mm; 2m 0.12 mm; (RS+M)b 0.07 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 mostly smooth, sometimes shiny, sometimes with punctures in posterior half and/or on lateral edges; T1 length 0.42 mm; T1 width at posterior edge 0.12 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny or with some shallow indistinct sculpturing; T2 length 0.15 mm; T2 width at posterior edge 0.26 mm; ovipositor slightly protruding from end of metasoma.

Male

Unknown.

Remarks

Glyptapanteles rodriguezae sp. nov. constitutes BIN BOLD:AAU5026 and is 6.25% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL3583; *Glyptapanteles ruhri* sp. nov.).

Using the BOLD Batch ID engine, the DNA barcode of the holotype is 6.3% different from the most similar sequence from an Australian specimen (AUMIC418-18; *Glyptapanteles ruhri* sp. nov.). Both type specimens were sequenced for the *wingless* gene, which is identical between them and differs by a minimum of 6 bp from all other species with available sequence data. There is another specimen in the BOLD database, ASQAS187-11, which has an identical *COI* sequence, but the location data is listed only as Australia and there are no images available.

Distribution

This species is known from the ACT.

Glyptapanteles ruhri Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:983624B0-EA3C-4AA3-B31B-54955164B7CB](https://doi.org/10.21203/rs.3.rs-983624B0-EA3C-4AA3-B31B-54955164B7CB)

Figs 13B, 50

Diagnosis

Glyptapanteles ruhri sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the species group, other than from *Glyptapanteles rodriguezae* sp. nov., by having the hind femur pale in colouration, the propodeum punctate with smooth areas between punctures and with little-to-no rugosity and T1 punctured in the posterior half. There was no external morphological character found which easily separates *G. ruhri* sp. nov. from *G. rodriguezae* sp. nov. and we do not provide a morphological diagnosis between these two species. The two species are sister lineages in the current phylogeny (Fig. 2), but *Glyptapanteles ruhri* sp. nov. is 6 bp different in the *wingless* sequence from *G. rodriguezae* sp. nov. and >6% divergent in the *COI* barcode and we therefore feel confident that they are different species.

Etymology

Named for researcher Peter Rühr, who was part of the collection of the type specimen and contributed to the success of a field trip in northern QLD in 2019 that led to the collection of many important microgastrine specimens.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Hallorans Hill Lookout, Atherton; -17.268, 145.4917268; 858 m a.s.l.; 21 Nov. 2019; E. Fagan-Jeffries, J.B. Dorey and P. Rühr leg.; sweeping vegetation; Extraction1198, BOLD: AUGLY071-21; QM T250986.

Paratypes

AUSTRALIA – Queensland • 1 ♂; Woonoonooroon National Park scenic drive; -17.585, 145.7028957; 674 m a.s.l.; 21 Nov. 2019; E. Fagan-Jeffries, J.B. Dorey and P. Rühr leg.; sweeping vegetation; Extraction832, BOLD: AUGLY001-21; QM T250987 • 1 ♂; same collection data as for preceding; Extraction844, BOLD: AUGLY002-21; QM T250988 • 1 ♀ (ethanol); Lamington National Park; -28.21, 153.139; 474 m a.s.l.; 9–19 Apr. 2007; C. Lambkin and N. Starick leg.; IBISCA Plot # IQ-500-C rainforest Malaise trap; Extraction636, BOLD: AUMIC418-18; QM T208406 • 1 ♀; Lamington National Park; -28.148, 153.137; 267 m a.s.l.; 8–18 Jul. 2007; C. Lambkin and N. Starick leg.; IBISCA Plot # IQ-300-A rainforest Malaise trap; Extraction639, BOLD: AUMIC420-18; QM T208407.

Description

Female

COLOURATION. Gena without a pale spot; labrum pale or reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres all black/dark brown or uniformly reddish-brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark or dark with significant orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa white; mid coxa white; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark or dark reddish-brown; T2 lateral area same colour as sclerotised area, or only slightly paler or dark extends past indentation, but then pale; T3 dark or uniformly brown; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 1.9 mm; fore wing length 2.1 mm; antennal length slightly longer than body length.

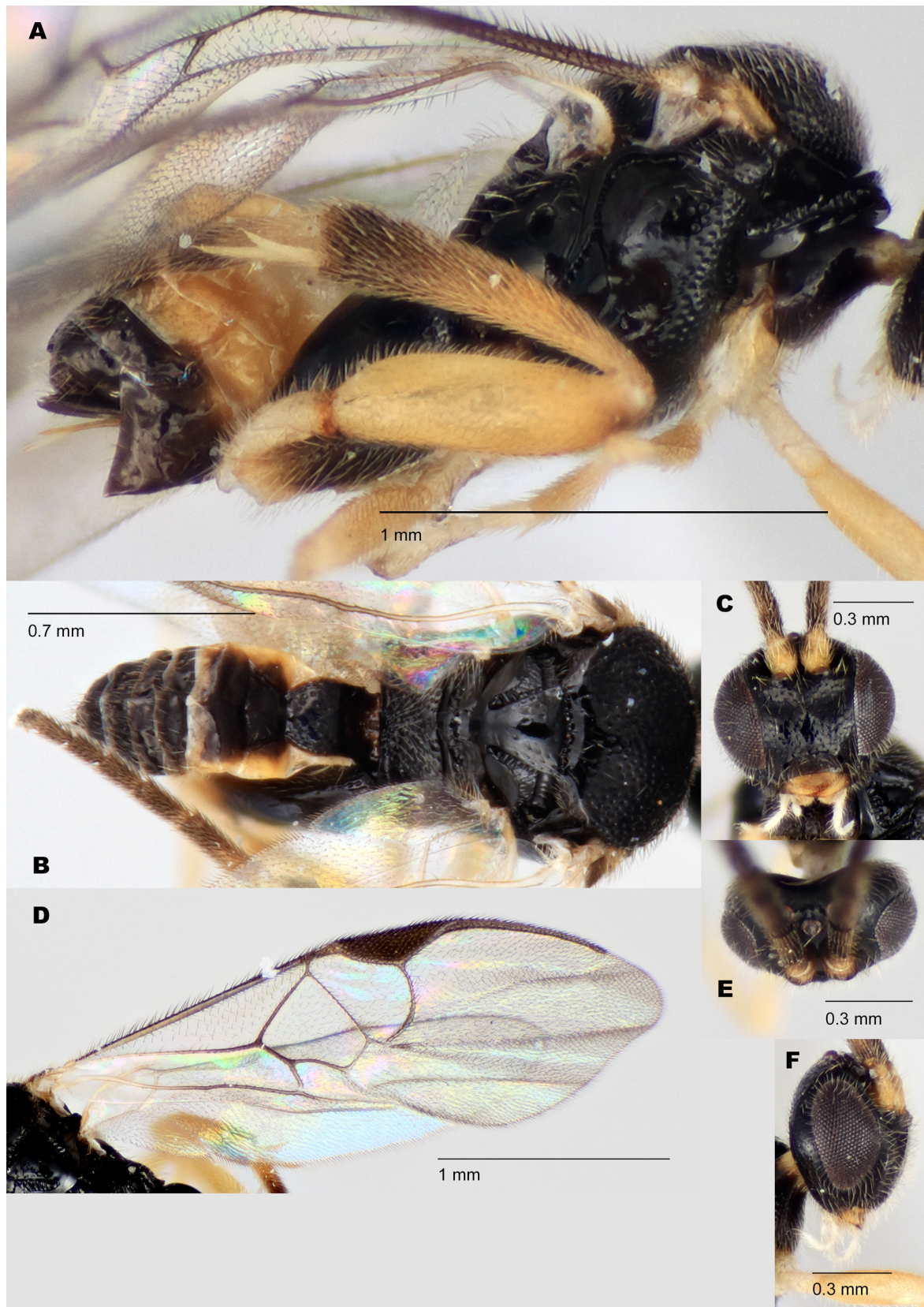


Fig. 50. *Glyptapanteles ruhri* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T250986). A. Lateral habitus. B. Dorsal habitus. C. Anterior head. D. Fore wing. E. Dorsal head. F. Lateral head.

HEAD. Antennal flagellomere 14 length/width 1.57–2.00; antennal flagellomere 2 length/width 2.62–3.00; OOD/POD 1.57–1.71; IOD/POD 1.43.

MESOSOMA. Anteromesoscutum sculpturing with very sparse, deep punctures; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; 9–10 pits in scutellar sulcus; propodeum with median carina absent, irregular rugose sculpturing along centre line with punctures in anterior half of propodeum, postero-lateral areas smooth or median carina absent, propodeum mostly punctate with some irregular shallow rugosity in centre, postero-lateral corners smooth and shiny.

WINGS. Pterostigma length 0.53 mm; pterostigma width 0.16 mm; r 0.22 mm; 2RS 0.14 mm; 2m 0.08 mm; (RS+M)b 0.06 mm.

METASOMA. T1 lateral edges parallel for anterior 3/4 of length, then narrowing posteriorly; T1 mostly smooth, some punctures in posterior half; T1 length 0.30 mm; T1 width at posterior edge 0.14 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny, or smooth with some very shallow punctures in posterior half; T2 length 0.14 mm; T2 width at posterior edge 0.2 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, 7–10 pits in scutellar sulcus.

Remarks

Glyptapanteles ruhri sp. nov. constitutes BIN BOLD:ADL3583 and is 6.25% (p-dist.) divergent from the closet BIN in the database (BOLD:AAU5026; *Glyptapanteles rodriguezae* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 6.4% different from the most similar *COI* sequence from an Australian specimen (BOLD: ASQAS187-11, which was not included in the original study and AUGLY062-21; *Glyptapanteles rodriguezae* sp. nov.). All five type specimens were sequenced for the *wingless* gene, which is identical among the type specimens and differs by a minimum of 6 bp from all other species with available sequence data.

Distribution

This species is known from a significant stretch of eastern QLD, from Atherton in northern QLD to Lamington National Park, south of Brisbane.

Glyptapanteles sanniopulus Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:B70D5575-E6F6-496F-8B47-453AD631CC2E](https://zoobank.org/urn:lsid:zoobank.org:act:B70D5575-E6F6-496F-8B47-453AD631CC2E)

Figs 5B, 5I

Diagnosis

Glyptapanteles sanniopulus sp. nov. is in the *G. albigena* species group and can be separated from most other members of the species group by having the gena with a large pale spot (Fig. 51G) rather than small (e.g., Fig. 40A) and from *G. albigena* sp. nov. by having the gena spot covering less than a third of the gena height and more rounded at the dorsal edge.

Etymology

The species epithet ‘*sanniopulus*’ is a combination of the Latin words ‘*sannio*’ (‘one who makes faces’) and ‘*polus*’ (‘pole of the earth’) and refers to the pale spot on the gena characteristic of this species and of the broader *G. albigena* species group. It is a noun in apposition.

Material examined

Holotype

AUSTRALIA • ♀; New South Wales, Monga National Park near Penance Grove Walk; -35.597372, 149.912126; 9 Nov. 2019–16 Jan. 2020; K.M. Bayless and J.G. Lumbers leg.; Malaise trap over stream, trap destroyed by bushfire 30 Dec. 2019, partially dried out; Extraction1666, BOLD: AUGLY127-21; ANIC 32 130370.

Paratypes

AUSTRALIA – **New South Wales** • 1 ♂; same collection data as for preceding; Extraction1638, BOLD: AUGLY121-21; ANIC 32 130371. – **Tasmania** • 1 ♂; Hobart, Kingston Beach; -42.986, 147.317; 14–18 Sep. 2010; B. Ward leg.; BIOUG00996-D09, BOLD: HYAT425-11; ANIC 32 130372.

Description

Female

COLOURATION. Gena with a pale spot; labrum pale; scape colour in ventral half the same colour or darker than flagellomeres; flagellomeres all black/dark brown; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark or dark with very slight orange patches on posterolateral corners; scutellar disk and metanotum dark; propodeum dark; fore coxa pale yellow; mid coxa pale yellow; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow or light brown; mid tibia light brown; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark or dark reddish-brown; T2 lateral area same colour as sclerotised area, or only slightly paler, or dark extends past indentation, but then pale; T3 dark or uniformly brown; T4+ dark or reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 2.4 mm; fore wing length 2.6 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 2.6; antennal flagellomere 2 length/width 4.33; OOD/POD 1.86; IOD/POD 1.43.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size. Often smoother in posterior centre; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; nine pits in scutellar sulcus; propodeum with median carina present and complete, rest of propodeum strongly rugose.

WINGS. Pterostigma length 0.6 mm; pterostigma width 0.23 mm; r 0.16 mm; 2RS 0.15 mm; 2m 0.13 mm; (RS+M)b 0.1 mm.

METASOMA. T1 wedge-shaped, narrowing posteriorly for entirety of length, lateral edges straight (but not parallel); T1 smooth and shiny, some shallow scattered punctures on lateral edges and posterior half or smooth in anterior half, indistinct sculpturing in posterior half; T1 length 0.62 mm; T1 width at posterior edge 0.1 mm; T2 an isosceles trapezoid, lateral edges straight; T2 with some shallow indistinct sculpturing or smooth and shiny, some shallow punctures along posterior edge; T2 length 0.16 mm; T2 width at posterior edge 0.27 mm; ovipositor slightly protruding from end of metasoma.

Male

As female, 7–9 pits in scutellar sulcus.

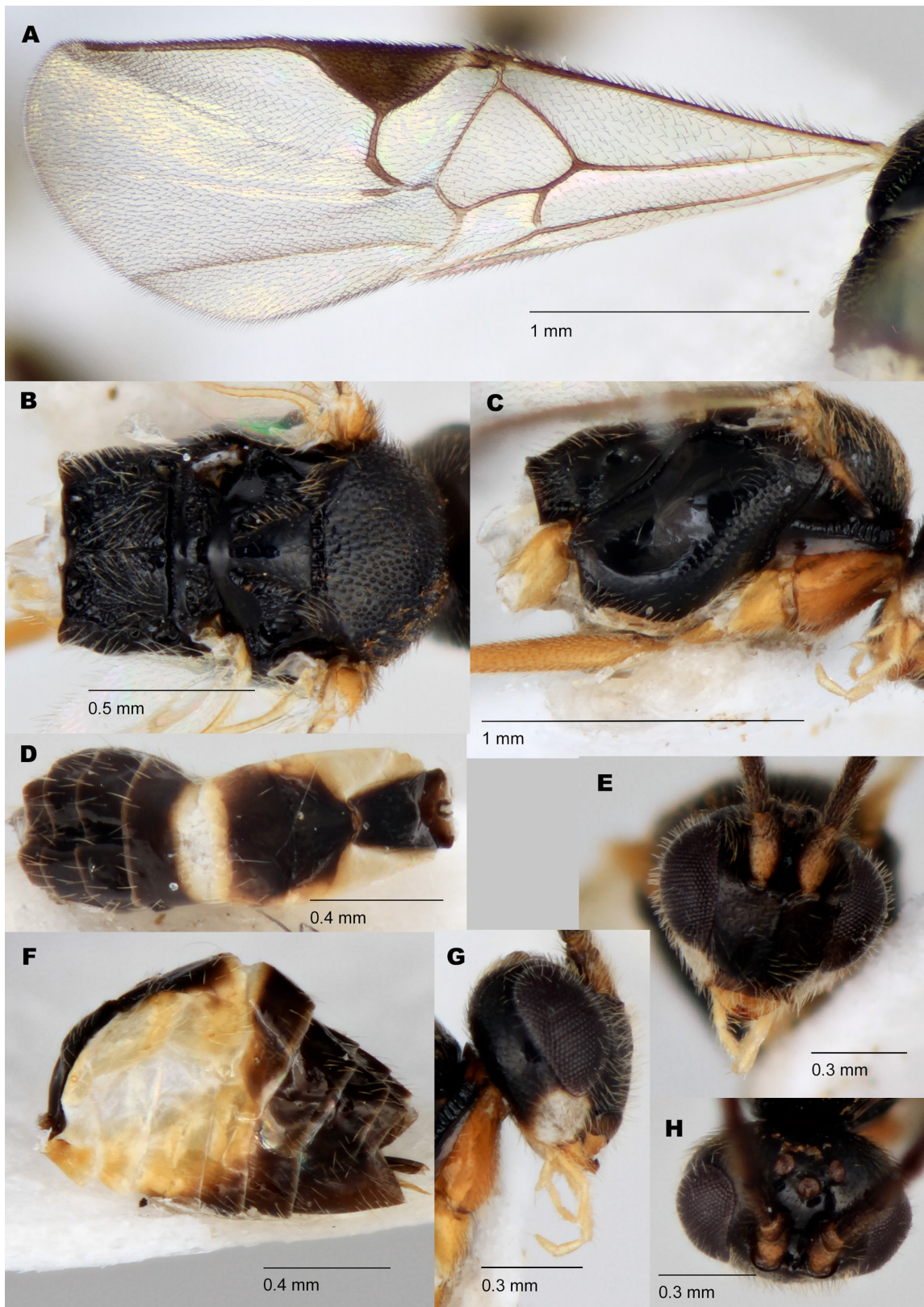


Fig. 51. *Glyptapanteles sannipolus* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (ANIC 32 130370). A. Fore wing. B. Dorsal mesosoma. C. Lateral mesosoma. D. Dorsal metasoma. E. Anterior head. F. Lateral metasoma. G. Lateral head. H. Dorsal head.

Remarks

Glyptapanteles sanniopulus sp. nov. constitutes BIN: BOLD:ABA6208 and is 5.48% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL3908; *Glyptapanteles albigena* sp. nov.).

Using the BOLD Batch ID engine, the *COI* barcode of the holotype is 5.6% different from the most similar *COI* sequence from an Australian specimen (AUMIC394-18; *Glyptapanteles albigena* sp. nov.). The two specimens able to be sequenced for the *wingless* gene share a unique barcode, which differs by a minimum of 3 bp from all other species with available sequence data.

Distribution

This species is known from southern NSW and TAS.

Glyptapanteles taylori (Wilkinson, 1928)
Fig. 47A–B, D

Diagnosis

As Austin & Dangerfield (1992) state that this species is unlikely to actually occur in the Australasian region and as we were unable to find morphological characters that definitively separate this species from all others in Australia, we do not diagnose it. There is a very low risk of this species being conspecific with any of the newly described species from Australia due to the likelihood that it does not actually occur in the region.

Material examined

Holotype

JAVA • ♀; “Java 1926 T.N.C. Taylor”; “Pres. by Imp. Bur. Ent. Brit. Mus. 1928-318”; NHMUK: 3.c.1018 (only images examined).

Distribution (in the Australasian region)

Fiji (likely erroneous record), introduced.

Glyptapanteles vergrandiacus Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:C7A23777-0E24-4E31-80E4-2A398EC889F0](https://zoobank.org/urn:lsid:zoobank.org:act:C7A23777-0E24-4E31-80E4-2A398EC889F0)
Fig. 52

Diagnosis

Glyptapanteles vergrandiacus sp. nov. is in the *G. arcanus* species group and can be separated from the other members of the species group (other than from *G. arcanus* sp. nov.) by having the propodeum with generally coarser sculpturing than most species, other than *G. lambkinae* sp. nov. and *G. erucadesolator* sp. nov. and with T1 less strongly sculptured than in *G. lambkinae* sp. nov. and *G. erucadesolator* sp. nov. It is noted, however, that all these species (along with *G. goodwinnoakes* sp. nov., which has T1 smooth) are morphologically very similar and identifications should be made with DNA barcodes. There was not a morphological character found to easily differentiate *G. vergrandiacus* sp. nov. from *G. arcanus* sp. nov., which are sister lineages in the current phylogeny (Fig. 2), but these two species differ in the *wingless* barcode by a single base pair and can also be separated by the *COI* barcodes, which have a >3% divergence.

Etymology

The species epithet ‘*vergrandiacus*’ is a combination of the Latin words ‘*vergrandis*’, meaning ‘little’ and ‘*acus*’, meaning ‘needle’ and refers to the short ovipositor present on this species. It is a noun in apposition.

Material examined**Holotype**

AUSTRALIA • ♀; Queensland, Kuranda; -16.8135, 145.6430586; 317 m a.s.l.; 12 Feb.–6 Apr. 2020; M.S. Moulds leg.; Malaise Trap EFJ2020MT36; Extraction1548, BOLD: AUGLY102-21; QM T250989.

Paratypes

AUSTRALIA – **Queensland** • 1 ♀; same collection data as for holotype; Extraction1521, BOLD: AUGLY093-21; QM T250990 • 1 ♀; same collection data as for holotype; Extraction1525, BOLD: AUGLY095-21; QM T250991 • 1 ♀; same collection data as for holotype; Extraction1541, BOLD: AUGLY097-21; QM T250992 • 1 ♀; same collection data as for holotype; Extraction1555, BOLD: AUGLY105-21; QM T250993 • 1 ♀; same collection data as for holotype; Extraction1560, BOLD: AUGLY107-21; QM T250994 • 1 ♀; same collection data as for preceding; 16 Nov.–13 Dec. 2019; Extraction924, BOLD: AUGLY019-21; QM T250995 • 1 ♀; same collection data as for holotype; 14 Dec. 2019 –6 Jan. 2020; Extraction1097, BOLD: AUGLY047-21; QM T250996 • 1 ♀ (ethanol); same collection data as for holotype; 8 Jan.–11 Feb. 2020: Extraction1511, BOLD: AUGLY090-21; QM T250997 • 1 ♀; same collection data as for holotype; Extraction1513, BOLD: AUGLY091-21; QM T250998 • 1 ♀ (ethanol); same collection data as for holotype; 19 May–8 Jul. 2017; Extraction731, BOLD: AUMIC484-18; QM T250999.

Description**Female**

COLOURATION. Gena without a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres uniformly reddish-brown or darkening distally; tegula dark; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa dark; mid coxa dark; hind coxa dark; fore femur pale yellow; mid femur pale yellow or dark proximally, lightening distally; hind femur dark; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark; T2 lateral area dark extends past indentation, but then pale; T3 mostly dark with paler lateral areas; T4+ dark.

HOLOTYPE BODY MEASUREMENTS. Body length 2.2 mm; fore wing length 2.2 mm; antennal length similar to body length.

HEAD. Antennal flagellomere 14 length/width 1.62–2.16; antennal flagellomere 2 length/width 3.00–3.42; OOD/POD 1.71–2.17; IOD/POD 1.29–1.57.

MESOSOMA. Anteromesoscutum sculpturing with shallow to deep punctures, space between punctures varying from smaller than diameter of punctures to larger than diameter of punctures, very sparse, deep punctures or very sparse, shallow to deep punctures; scutellar disk sculpturing with only very shallow punctures, smooth and shiny; 7–11 pits in scutellar sulcus; propodeum with median carina absent and propodeum strongly rugose.

WINGS. Pterostigma length 0.52 mm; pterostigma width 0.16 mm; r 0.16 mm; 2RS 0.13 mm; 2m 0.09 mm; (RS+M)b 0.06 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{3}{4}$ of length, then narrowing posteriorly or broadest at centre of length, narrowing both posteriorly and anteriorly from mid-point; T1 smooth in anterior half, indistinct sculpturing in posterior half; T1 length 0.35 mm; T1 width at posterior edge 0.14 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.15 mm; T2 width at posterior edge 0.27 mm; ovipositor slightly protruding from end of metasoma.

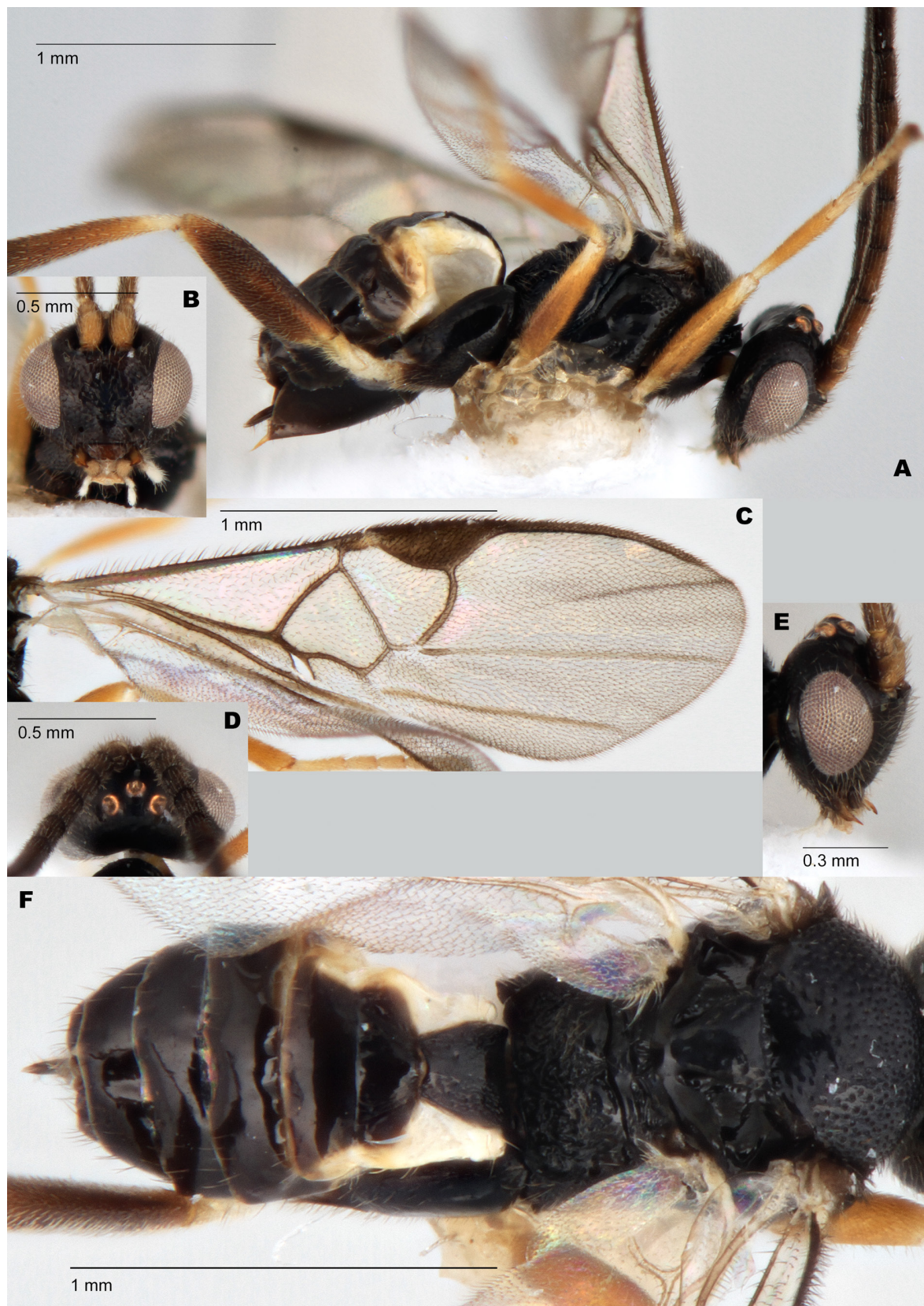


Fig. 52. *Glyptapanteles vergrandiacus* Fagan-Jeffries, Bird & Austin sp. nov., paratype, ♀ (QMT250990). A. Lateral habitus. B. Anterior head. C. Fore wing. D. Dorsal head. E. Lateral head. F. Dorsal habitus.

Male

Unknown.

Remarks

Glyptapanteles vergrandiacus sp. nov. constitutes BIN BOLD:ADL3012 and is 3.67% (p-dist.) divergent from the closet BIN in the database (BOLD:ADL5262; *Glyptapanteles arcanus* sp. nov.).

Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 3.6% different from the most similar *COI* sequence from an Australian specimen (AUMIC423-18; *Glyptapanteles arcanus* sp. nov.). All of the type specimens were sequenced for the *wingless* gene, which is identical within the species and differs by a minimum of 1 bp from all other species with available sequence data.

Distribution

This species is known only from Kuranda in northern QLD.

Glyptapanteles wrightae Fagan-Jeffries, Bird & Austin sp. nov.
[urn:lsid:zoobank.org:act:569C2146-14FF-4453-BA4E-A6A8787A044A](https://doi.org/10.3896/urn:lsid:zoobank.org:act:569C2146-14FF-4453-BA4E-A6A8787A044A)

Fig. 53

Diagnosis

Glyptapanteles wrightae sp. nov. can be separated from the other members of the *arcanus* species group as follows:

Glyptapanteles wrightae sp. nov. can be separated from *G. rodriguezae* sp. nov. and *G. ruhri* sp. nov. by T1 being smooth and shiny, not having punctures that cover at least a third of the area of the posterior half of the tergite.

Glyptapanteles wrightae sp. nov. can be separated from *G. goodwinnoakes* sp. nov., *G. erucadesolator* sp. nov., *G. lambkinae* sp. nov., *G. arcanus* sp. nov. and *G. vergrandiacus* sp. nov. by the propodeum being less coarsely and less consistently rugose sculptured across the anterior half of the propodeum. *Glyptapanteles wrightae* sp. nov. has the propodeum with strong punctures in the anterior half, the posterior half with shallow or strong rugose sculpturing.

Glyptapanteles wrightae sp. nov. can be separated from *G. doreyi* sp. nov. by the tegula being pale in colouration (the tegula in *G. doreyi* sp. nov. is generally dark in colouration), the hind femur mostly pale or light brown (mostly dark in *G. doreyi* sp. nov.) and the indentation in the centre of the mesopleuron being smooth, not strongly canaliculated as it is in *G. doreyi* sp. nov.

There was not a morphological character found that easily separates *G. wrightae* sp. nov. from *G. lessardi* sp. nov. and we do not diagnose these species morphologically. The *wingless* barcodes of the two species differ by 3 bp and the *COI* is greater than 4% divergent.

Etymology

Named for Susan Wright, who collected the holotype specimen. EPF-J would like to acknowledge Susan's support of visiting researchers at the QM and her continual generosity with her time and advice.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Samsonvale Cemetery, 8.5 km SSE of Dayboro; -27.2703, 152.856; 50 m a.s.l.; 6 Jan.–8 Feb. 2015; S. Wright leg.; *Casuarinal*/open forest Malaise trap; Extraction198, BOLD: AUMIC084-18; QM T208400.

Paratypes

AUSTRALIA – New South Wales • 1 ♂; Royal National Park, near Waterfall Couranga Track near Hacking River; -34.1486, 151.0221; 40 m a.s.l.; 20 Jan.–6 Feb. 2020; K.M. Bayless and J.G. Lumbers leg.; 6 m Malaise trap over Waterfall Creek; Extraction857, BOLD: AUGLY128-21; ANIC 32 130373 • 1 ♂; same collection data as for preceding; Extraction863, BOLD: AUGLY010-21; ANIC 32 130374.

Description

Female

COLOURATION. Gena without a pale spot; labrum reddish-brown; scape colour in ventral half uniformly paler than flagellomeres; flagellomeres darkening distally; tegula pale; wing veins uniformly black or brown, or with small lighter area proximally; anteromesoscutum all dark; scutellar disk and metanotum dark; propodeum dark; fore coxa white; mid coxa white; hind coxa dark; fore femur pale yellow; mid femur pale yellow; hind femur pale yellow; fore tibia pale yellow; mid tibia pale yellow; hind tibia darkening posteriorly; hind basitarsus light brown; T1 dark; T2 sclerotised area dark reddish-brown; T2 lateral area dark extends past indentation, but then pale; T3 mostly dark with paler lateral areas; T4+ reddish-brown.

HOLOTYPE BODY MEASUREMENTS. Body length 1.8 mm; fore wing length 1.8 mm; antennal length slightly longer than body length.

HEAD. Antennal flagellomere 14 length/width 3.5; antennal flagellomere 2 length/width 2.71; OOD/POD 1.83; IOD/POD 1.67.

MESOSOMA. Anteromesoscutum sculpturing with deep punctures, space between punctures a mixture of smaller than diameter of punctures and of similar size; scutellar disk sculpturing with deep, sparse, irregularly spaced punctures, more common on anterior lateral edges or scutellar disk with shallow punctures scattered over most of area; eight pits in scutellar sulcus; propodeum with median carina absent, strong punctures in anterior half, rugose sculpturing in posterior half.

WINGS. Pterostigma length 0.49 mm; pterostigma width 0.16 mm; r 0.13 mm; 2RS 0.14 mm; 2m 0.06 mm; (RS+M)b 0.07 mm.

METASOMA. T1 lateral edges parallel for anterior $\frac{1}{2}$ to $\frac{2}{3}$ of length, then narrowing posteriorly; T1 mostly smooth, some punctures in posterior half, smooth and shiny, sometimes with some shallow scattered punctures on lateral edges; T1 length 0.25 mm; T1 width at posterior edge 0.08 mm; T2 an isosceles trapezoid, lateral edges straight; T2 smooth and shiny; T2 length 0.14 mm; T2 width at posterior edge 0.19 mm; ovipositor slightly protruding from end of metasoma.

Male

As female.

Remarks

Glyptapanteles wrightae sp. nov. constitutes BIN BOLD:ADL3293 and is 1.76% (p-dist.) divergent from the closet BIN in the database (BOLD:AEI5416; see information below about the specimens within this BIN).

Using the BOLD Batch ID engine, the *COI* sequence of the holotype is 1.7% different from the most similar *COI* sequence from an Australian specimen (AUGLY139-21, an undescribed lineage, with three sequences). This closely related lineage, (BOLD:AEI5416; also including BOLD AUGLY135-21 and AUGLY130-21) requires further study to determine whether it is within the species limits of *G. wrightae* sp. nov. or whether it represents a distinct species.

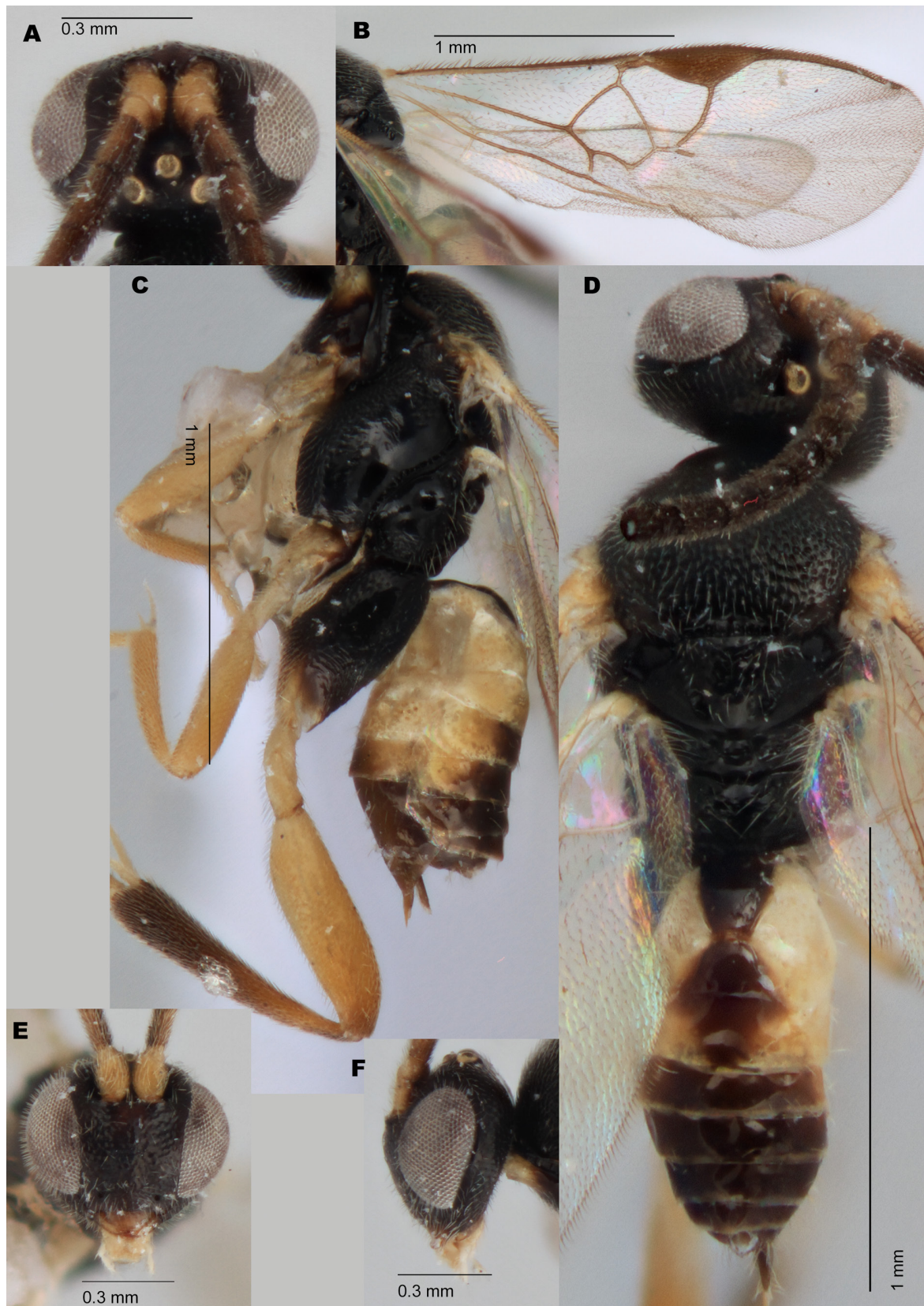


Fig. 53. *Glyptapanteles wrightae* Fagan-Jeffries, Bird & Austin sp. nov., holotype, ♀ (QM T208400). A. Dorsal head. B. Fore wing. C. Lateral habitus. D. Dorsal habitus. E. Anterior head. F. Lateral head.

Distribution

This species is currently known from southern QLD and north-eastern NSW.

Discussion

In this study we attempted to find a balance between producing robust species hypotheses which are accompanied by descriptions that are useful for end-user identifications and adhering to the Taxonomy Australia mission of documenting Australia's biodiversity at an accelerated pace. Whilst we believe the majority of our species descriptions will stand the test of time, this project makes only a tiny contribution to treating the huge number of undescribed species of Australian Microgastrinae. Whilst not worked on full-time, this study took 18 months of a postdoctoral researcher's time, with significant assistance from the co-authors. At this rate, it would take over 65 years to describe the remainder of the estimated 1300 undescribed microgastrine species in Australia. Changes to the traditional way of delimiting and describing species are needed to tackle these hyper-diverse, morphologically conserved taxa. Some authors have attempted to create this change by describing species based almost entirely on *COI* barcode BIN assignments on BOLD (Sharkey *et al.* 2021) with morphology barely used at all in descriptions or diagnoses, but this approach has received criticism (Ahrens *et al.* 2021; Meier *et al.* 2021). We have included morphology to a reasonably large extent in the diagnoses and key provided here, but have also described several species which are only diagnosable from their closest relative(s) by molecular information.

This approach is simply not tenable going forward, however, as even in this one genus and in the limited geographical scope of this study, there are nearly as many undescribed lineages as those that we were able to describe. Finding morphological characters to delimit a further 30 or so species, that are consistent and useful for non-experts, would not be an efficient or practical way to document the remaining Australian *Glyptapanteles* fauna that have DNA barcoded specimens available, let alone those that are yet to be collected and incorporated into a revision. A recent revision of *Glyptapanteles* from Costa Rica and Ecuador (Arias-Penna *et al.* 2019) described an impressive 136 species in a single study and provided a morphological key to species. Many of the characters used to separate species in Arias-Penna *et al.* (2019) are variable for the Australian species examined here and therefore not useful as diagnostic characters for all regions. We do not provide the answers to this dilemma here, but simply note that our approach, whilst limiting the application of morphological diagnoses, has not gone far enough to make the needed impact on increasing the rate at which new species of microgastrines are described.

As we relied on specimens that were preserved in ethanol and therefore suitable for DNA extraction, the sampling in this study is limited. Along with the lineages not described here, but recognised by sequenced specimens (Fig. 2), there is likely many more *Glyptapanteles* species yet to be collected, DNA barcoded and described in Australia. We make an assumption in this study that the species described here are endemic to Australia and Papua New Guinea and we do not morphologically diagnose them against *Glyptapanteles* species described from outside of the region. Whilst it is theoretically possible that there are either recent introductions or cosmopolitan species present in Australia, morphology in this group is not often reliable enough to make confident conclusions about species identification. We have ensured that none of our newly described species have *COI* barcodes <2% different to anything available on BOLD (other than those mentioned in the remarks sections from Papua New Guinea) and consider that until a global effort to sequence historical type material is conducted, we are unable to categorically ensure that none of the new species have not already been described from another region.

A significant impediment to a modern revision of Australian parasitic wasps is the lack of host data for specimens that are of DNA-quality. Most specimens able to be DNA sequenced and incorporated into the current revision were collected by Malaise traps and sweep-netting and are therefore devoid of the

critical biological information that can a) be used as a third piece of delimitation evidence alongside morphology and molecular information and b) allow the newly described species to be contextually placed in the environment. As such, we were not able to draw many conclusions about whether, for example, any of these new species are at risk of co-extinction due to parasitising endangered hosts, or whether any of these new species may be useful biological control agents for lepidopteran pests. A concerted effort to rear lepidopterans and their parasitoids is needed in Australia to allow more robust and more useful, taxonomic revisions to occur.

Acknowledgements

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Supplementary files

Supp. file 1. Specimen data for the phylogeny presented in Fig. 1, in the standard downloaded BOLD format (.xlsx). <https://doi.org/10.5852/ejt.2022.792.1647.5927>

Supp. file 2. Alignment file in PHYLIP format for the phylogeny presented in Fig. 1 (.txt). <https://doi.org/10.5852/ejt.2022.792.1647.5929>

Supp. file 3. Specimen data for the phylogeny presented in Fig. 2, in the standard downloaded BOLD format (.xlsx). <https://doi.org/10.5852/ejt.2022.792.1647.5931>

Supp. file 4. Alignment file in PHYLIP format for the phylogeny presented in Fig. 2 (.txt). <https://doi.org/10.5852/ejt.2022.792.1647.5933>

Supp. file 5. Expanded phylogeny of the circular phylogeny presented in Fig. 1, support values are SH-aLRT support (%) / ultrafast bootstrap support (%) (.pdf). Due to the characteristics of this file, it is recommended to open it in a browser (i.e., Chrome, Safari) or with an application such as Preview Mac or similar. <https://doi.org/10.5852/ejt.2022.792.1647.5935>