

Structure Elucidation of Erythrosotidienone and Erythromotidienone — Two New Isoquinoline Alkaloids from *Erythrina variegata* Flowers†

SHIV K. SHARMA and H. MOHINDRA CHAWLA*

Department of Chemistry, Indian Institute of Technology, Hauz Khas, New Delhi-110 016

Manuscript received 9 November 1998

Careful examination of the acetone extract of the *Erythrina variegata* flowers has revealed the presence of five compounds, three of which are identified as stigmasterol (1), cycloartenol (2) and erysotramidine (13). The remaining two compounds are new isoquinolines whose structures have been established on the basis of chemical correlation, derivatization and ir, ^1H nmr, ^{13}C nmr and mass spectral analysis. They are tentatively named as erythrosotidienone (4) and erythromotidienone (10). Cycloartenol (2) has been isolated for the first time from *Erythrina* species. The occurrence of the new compounds 4 and 10 is of interest from the biogenetic point of view.

Erythrina (Family-Leguiminosae, subfamily-Papilionaceae) is an important genus among the trees and shrubs consisting of about fifty species and is found in the tropic and subtropic zones of all parts of the world¹⁻³. Various parts of some species such as *arborescens*, *stigmoides*, *stricta*, *suberosa* and *variegata* are reported to possess pharmacological and medicinal properties⁴⁻⁶. These were usually attributed to their rich alkaloidal content which is normally considered as a reservoir of protein synthesis, as a protective material discouraging animal or insect attacks, as plant stimulants or regulators for growth, metabolism and reproduction, or as detoxicating agents for substances whose accumulation might otherwise cause damage to the plant^{7,8}.

Erythrina variegata is a tall ornamental tree, reaching about 10 m in height, and is widely distributed throughout the upper Gangetic plains of India, including Assam^{9,10}. Various extracts of different parts of this plant find use in the indigenous system of medicine and have been found antiasthmatic and a collyrium in ophthalmia¹⁰⁻¹². The leaves of the plant are used as a diuretic and for relieving pain in joints. A thorough literature survey indicates that leaves, seeds, stem bark and root bark of *E. variegata* have been examined^{1,12-16} for their chemical components but very little work seems to have been reported on its flowers which was taken up in the present study.

In this paper, we wish to report an account of the isolation and structural elucidation of five compounds (designated as A₁-A₅) from the acetone extract of flowers of *E. variegata* two of which are new natural products.

Identification of compounds :

The concentrated acetone extract of the shade-dried flowers of *Erythrina variegata* on maceration with methanol afforded the methanol-soluble and methanol-insoluble fractions. The methanol-insoluble fraction when subjected to column chromatography over silica gel, gave two com-

pounds, designated A₁ and A₂, while the methanol-soluble fraction, when chromatographed over neutral alumina, afforded three compounds, designated A₃, A₄ and A₅.

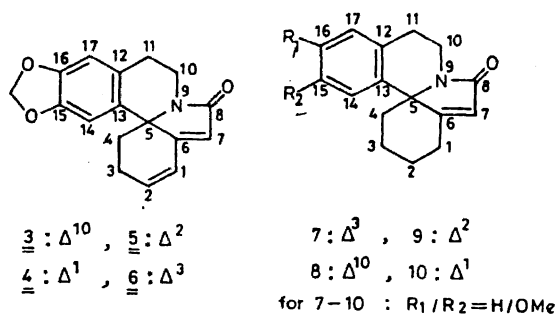
Compound A₁, crystallized as white needles, m.p. 168°, was identified as stigmasterol (1) by colour reaction, ir, ^1H nmr and mass spectral fragmentation pattern, and by preparation of the derived acetate^{17,18}.

Compound A₂ was crystallized from benzene-chloroform as white needles, m.p. 116°, and gave a positive Liebermann-Burchard test. From a detailed analysis of its ir, ^1H nmr and mass spectral fragmentation pattern, it was identified as cycloartenol (2), which was confirmed by its literature data^{19,20}. This is the first report of the occurrence of this compound in *Erythrina* species.

Compound A₃ designated as erythrosotidienone, C₁₇H₁₅NO₃ (also, M⁺ 281), neither gave any colour with FeCl₃ nor a positive reaction with Liebermann-Burchard reagent, which indicated the absence of a phenolic group and a steroidal or a triterpenoidal skeleton in the molecule respectively. Since it responded to a colour reaction with Meyers reagent, it was considered to be an alkaloid. Strong ir absorption at 1740 cm⁻¹ suggested the presence of a five-membered lactam ring which is a common structural feature among *Erythrina* alkaloids, and this was confirmed by the ^{13}C nmr signal at δ 184.0. The other absorptions at 1610 and 1160 cm⁻¹ reflected the presence of an unsaturation and an ether linkage respectively in the molecule. The ^1H nmr spectrum of compound A₃ showed the presence of a methylenedioxy group (δ 5.94, 2H, s) and three aromatic protons (δ 7.32, 7.23 and 6.62, 1H, s each, H-14, 17, 7 respectively). In the mass spectrum, the molecular ion peak at m/z 281 suffered two successive losses of 28 and 26 units to give respective intense fragments, at m/z 253 and 227. Since its ^1H nmr showed a total number of 15 protons (also confirmed by MS), there should be one more double bond

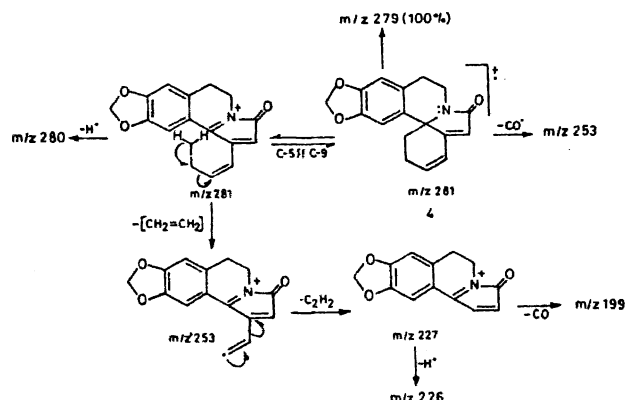
†Dedicated to Professor Sukh Dev on his 75th. birthday.

present, which could be accommodated in any one of the four structures 3-6.



The structures 3 and 5 were rejected as only one doublet and a multiplet in the low field, i.e. at δ 6.02 and 5.72 respectively, were observed in the 1H nmr spectrum of compound A_4 , whereas the requirements for the structure 3 should be two doublets and for structure 5 there should be two multiplets. On the other hand, both the structures 4 and 6 equally explained the above observation and 1H nmr results could not resolve the constitution of A_3 . The problem was solved by the mass spectral analysis. The observed successive loss of 28 and 26 m.u. from the M^+ are only possible for 4 as structure 6 should first lose an acetylene molecule to give the fragment at m/z 225, which was not observed.

The derived structure 4 of compound A_3 was compatible with the major mass spectral fragmentation pattern (Scheme 1) and the high resolution 1H nmr spectral data (vide experimental). The proposed structure 4 was fully in agreement with its ^{13}C nmr (including SFORD). The tentative assignments have been shown in the experimental section.



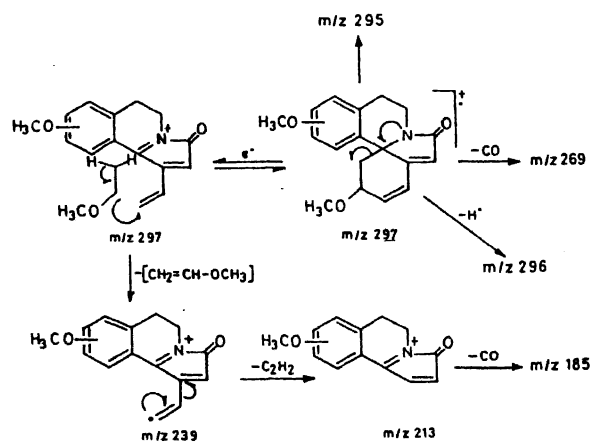
Scheme 1. Mass spectral fragmentation of erythrosotidienone (4). Except for M^+ , only peaks with r.a. of 15% and above have been shown. The peak at m/z 81 remains unexplained.

Compound A_4 designated as erythromotidienone, $C_{18}H_{19}NO_3$, gave a positive Meyers reagent test and the DNP test for a ketonic functionality but did not respond to the $FeCl_3$ and Liebermann-Burchard reactions. It showed strong ir bands at 1760, 1600 and 1170 cm^{-1} which are suggestive of a five-membered lactam ring, an unsaturation and an ether linkage respectively, in the compound. The other physical and chemical properties of A_4 indicate that it has an *Erythrina*-type skeleton, similar to that of A_3 .

In the 1H nmr spectrum, A_4 exhibited signals for two methoxyls – one aryl (δ 3.86, s) and the other aliphatic (δ 3.36, s) – and those typical of an *Erythrina* alkaloid skeleton, viz. three aromatic protons (δ 7.13, 7.21, 7.32), three olefinic protons (δ 6.02, 6.34, 6.62) and seven aliphatic protons (δ 1.92–3.76, m).

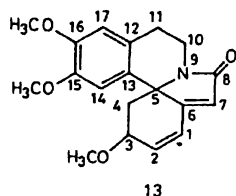
In the mass spectrum, compound A_4 showed peaks at m/z 297 (M^+), 296 ($M-1$), 295 ($M-2$), 269 ($M-CO$) and 266 ($M-OCH_3$). An intense fragment at m/z 239 could be ascribed to the loss of methyl vinyl ether molecule (a loss of 58 units) which revealed that the second methoxyl is attached to the C-3 position, as found in other *Erythrina* alkaloids¹²⁻¹⁶. To establish the position of double bond, four possible structures 7-10 were considered for compound A_3 .

Structure 7 was rejected as only one singlet (δ 6.62, 1H) was observed in its 1H nmr spectrum as against the expected two singlets for it. The structures 8 and 9 were ruled out as there would be two doublets and a triplet respectively in the low field but we observed only a doublet and a double doublet in the 1H nmr spectrum. This left structure 10 to be the likely structure for A_4 . This was confirmed by analysis of the remaining peaks in the 1H nmr spectrum, so the doublet (δ 6.34) and the double doublet (δ 6.02) could be ascribed to the C-1 and C-2 protons respectively. Structure 10 was also in agreement with its mass spectral fragmentation, as depicted in Scheme 2.



Scheme 2. Mass spectral fragmentation of erythrosotidienone (10).

The exact position of the methoxyl group, i.e. whether it is attached to C-15 (11) or C-16 (12) could not be ascertained due to insufficient amount of the sample which did not allow execution of chemical reactions that might have shed light on the true structure for the compound.



Compound A₅ was identified as erysotramidine (13) by its MS data and was confirmed by comparison of its IR and ¹H NMR data reported in literature²¹.

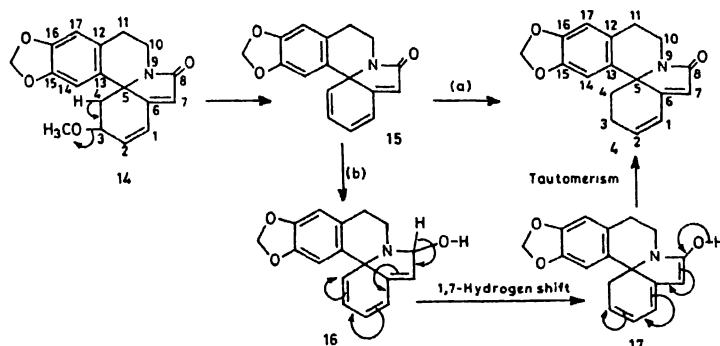
Though analogous synthetic compounds have been described in the literature. Erythrosotidienone (4) is the only naturally occurring compound among *Erythrina* alkaloids which does not have a methoxyl group in the C-3 position. At this stage, a comment on possible biogenesis of erythrosotidienone (4) is in order. Prelog *et al.*^{22,23} and Koniuszy and Folkers²⁴ reported that an *Erythrina* alkaloid with a methoxyl functionality at C-3 incurs a loss of methanol to yield a compound which has a double bond between C-3 and C-4 on treatment with a mineral acid at room temperature. In some cases, the newly generated double bond has been determined to undergo an apo-rearrangement to give other isomers^{25,26}. We believe that a similar pathway might be operative during the biogenesis of erythro-sotidienone via and intermediate 15 which can be formed from compound 14 (Scheme 3). The carbonyl group of 15 might undergo biological reduction to afford 16 which yields 17 and

While the petroleum ether extract²⁷ revealed the presence of steroids, fatty acids and only one known alkaloid, the acetone extract seems to be rich in alkaloids. Cycloartenol (2) is being reported for the first time from an *Erythrina* species.

Experimental

All m.ps. were determined on an electric melting point apparatus (Adair Dutt) in open capillaries and are uncorrected. ¹H and ¹³C NMR spectra were recorded in CDCl₃ on 99.50 MHz JOEL FT NMR spectrometer using TMS as the internal standard and chemical shifts are reported in δ scale. IR spectra were run on KBr discs on SP-1200 and 5DX-Nicolet FT IR spectrometers. Mass spectra were recorded on a JEOL JMS-D-300 spectrometer at 70 eV. The purity of the compounds were monitored by TLC performed on silica gel (B.D.H.) plates using iodine for visualizing the spots. Column chromatography was carried out by using silica gel (60–120 mesh) and neutral alumina. The organic solvent extracts were usually dried over anhydrous MgSO₄ and the solvents used were freshly distilled and purified before use. Petroleum ether, b.p. 60–80°, was used.

Extraction and isolation procedure : Shade-dried flowers (1.2 kg) of *E. variegata* (collected from the main campus of the Indian Institute of Technology, Hauz Khas, New Delhi, in the month of April 1984) were extracted with petroleum ether (3 × 2 dm³, 12 h). The defatted plant material was then extracted with acetone (3 × 2 dm³, 12 h). The combined acetone extracts was concentrated under reduced pressure to give a greyish semisolid mass (13.4 g) which on maceration with methanol gave a solid. It showed six spots on TLC plates, but when subjected to column chromatography over silica gel, it afforded only two compounds A₁ and



Scheme 3 Suggested biogenetic pathways for the formation of erythrosotidienone (4)

4 via 1,7-hydrogen shift and prototypic tautomerism respectively. On the other hand, there is a possibility that 15 might directly lead to a product by reduction of a double bond between positions C-3 and C-4.

A₂. On the other hand, methanol-soluble part on concentration under reduced pressure and chromatography over neutral alumina yielded three major compounds A₅, A₃ and A₄. Compound A₁ (78 mg) was eluted from a mixture of

petroleum ether-benzene (1 : 3) and crystallized from methanol as white silky needles, $[\alpha]_{25} -52$, m.p. 168°. It was identified as stigmasterol (lit.^{17,18}, m.p. 170°), which was confirmed by preparation of its acetate (Py/Ac₂O); R.T., 48 h), a white powder, $[\alpha]_{25} -54$, m.p. 143° (lit.^{17,18}, 143–144°).

Compound A₂ (42 mg) was eluted and crystallized from a mixture of benzene-chloroform (4 : 1) as white needles, m.p. 116°. It was established as cycloartenol (lit.^{19,20}, m.p. 115°) and was confirmed by preparation of its acetate derivative as white plates, m.p. 122° (lit.^{19,20}, 122°) and direct comparison with literature data.

Compound A₃ (4; 92 mg) was isolated as a light brown semisolid from benzene-chloroform (10 : 3) eluate and was found to be soluble in dilute HCl and H₂SO₄. It did not respond to the Liebermann-Burchard test and FeCl₃ reactions, but gave a clear pale yellow bright colour with Meyers reagent, and analyzed for C₁₇H₁₅NO₃ (Found : C, 72.39; H, 5.28; N, 4.90. Requires : C, 72.59; H, 5.34; N, 4.98%); v_{\max} 2930s, 2860s, 1740s, 1610s, 1460s, 1440s, 1380s, 1280–60bs, 1160s, 1130s, 1075s, 1040s, 975s, 960s, 860–40bs, 730s,sh, 720s, 675s, 645s, 610s, 580s cm⁻¹, ¹H nmr (CDCl₃) δ 3.63 (2H, t, *J* 1.5 and 4.5 Hz, C-10-H), 5.72 (1H, m, C-2-H), 5.94 (2H, s, O-CH₂-O), 7.32 (1H, s, C-14-H), 7.23 (1H, s, C-17-H), 6.62 (1H, s, C-7-H), 6.02 (1H, d, *J* 10.0 Hz, C-1-H), 1.87–3.27 (6H, m, methylene and methine protons); *m/z* (% rel. abundance) 281 (5.0), 280 (22.5), 279 (100), 267 (7.5), 265 (7.5), 253 (17.5), 227 (2.5), 226, 199 (15.4), 174 (15.0), 167 (10.0), 133 (5.0), 81 (31.6), 120 (8.0), 77 (5.0), 76 (80.0), 56 (27.3), 55 (42.5), ¹³C nmr (CDCl₃) δ 184.0 (C=O), 152.0 (C-16), 152.4 (C-15), 132 (C-2), 131.0 (C-1), 129.6 (C-7), 128.3 (C-12), 128.1 (C-13), 114.0 (C-17), 110.1 (C-14), 104.0 (C-6), 102.8 (-O-CH₂-O), 68.1 (C-5), 40.3 (C-10), 32.0 (C-3), 30.4 (C-4), 23.2 (C-11).

Compound A₄ (10; 56 mg) was isolated from a mixture of benzene-chloroform (1 : 1) eluate, purified by preparative TLC using the same solvent system and crystallized from methanol as white needles, m.p. 172°. It responded to the 2,4-DNP test for ketones and the Meyers test for alkaloids, but gave no reaction with FeCl₃. It analyzed for C₁₈H₁₉NO₃ (Found : C, 72.98; H, 6.50; N, 4.63. Requires : C, 72.72; H, 6.39; N, 4.71%); v_{\max} 2920s, 2880s, 1760s, 1600s, 1450s, 1380s, 1280–60bs, 1335s, 1280–60bs, 1250s, 1170s, 1110s,sh, 1075a, 1055s, 1025wb, 975s, 960s, 925s, 885w,sh, 835s, 800s, 765w,sh, 730s, 700s cm⁻¹, ¹H nmr (CDCl₃) δ 3.86 (3H, s, OCH₃), 3.36 (3H, s, OCH₃), 7.21 (1H, d, *J* 2.7 Hz, C-14-H), 7.32 (1H, dd, *J* 2.5 and 9.0 Hz, C-16 or C-15-H), 7.13 (1H, d, *J* 2.3 Hz, C-17-H), 6.62 (1H, s, C-7-H), 6.34 (1H, d, *J* 9.6 Hz, C-1-H), 6.02 (1H, dd, *J* 2.5 and 7.5 Hz, C-2-H), 1.92–3.76 (7H, m, methylene and methine protons); *m/z*(% rel. abundance) 297

(5), 296 (8), 295 (11), 269 (19), 255 (30), 239 (8), 235 (10), 213 (16), 185 (16), 83 (CHCl₂⁺), 81 (43).

Compound A₅ (13; 52 mg) eluted from petroleum ether-benzene (5 : 4) eluates as a light yellow semisolid which could not be crystallized. It responded positively to a 2,4-DNP test and the Meyers test (HgCl₂ and KI in H₂O), but gave no colour change with FeCl₃. It analyzed for C₁₉H₂₁NO₄ (Found : C, 69.56; H, 6.30; N, 4.18. Requires : C, 69.72; H, 6.42; N, 4.28%); v_{\max} 2930s, 2860s, 1760s, 1600s, 1475s, 1380s, 1310s, 1170s, 1030s, 980–70bs, 915s,sh, 885–60bs, 810s,sh, 740s, 720s, 695s, 610s, cm⁻¹; ¹H nmr (CDCl₃) δ 3.86, 3.62 and 3.36 (3H, s, each, 3 × OCH₃), 7.16 (1H, s, C-17-H), 6.97 (1H, s, C-14-H), 6.52 (1H, s, C-7-H), 6.34 (1H, d, *J* 5.5, C-1-H), 6.01 (dd, *J* 3.5 and 9.5 Hz, C-2-H), 3.18–1.72 (7H, m, methylene and methine protons); *m/z* (% rel. abundance) 327 (19), 299 (16), 269 (15), 268 (13), 265 (19), 264 (15), 243 (19), 159 (18), 82 (CCl₂), 81 (31).

Acknowledgement

One of the authors (S.K.S.) thanks C.S.I.R., New Delhi, for the award of a Research Fellowship, and Department of Chemistry, University of St. Andrews, Scotland, U.K., for computing facility.

References

1. A. MONDON in "Chemistry of the Alkaloids", ed. S. W. PELLETIER, van Nostrand-Reinhold, New York, 1970, p. 173.
2. T. A. HENRY, "The Plant Alkaloids", 4th ed., J & A Churchill Ltd., London, 1949, p. 386.
3. L. E. GRAIG in "The Alkaloids, Chemistry and Physiology", ed. R.H.F. MANSKE, Academic, New York, 1995, Vol. V, Chap.46, p. 281.
4. K. UNNA, M. KNIAZUK and J. G. GRESLIN, *J. Pharmacol. Exp. Therap.*, 1944, **80**, 39.
5. D. MEGIRIAN, D. E. LEAVY and I. H. SLATER, *J. Pharmacol. Exp. Therap.*, 1955, **113**, 212.
6. H. LULLMANN, A. MONDON and P. R. SEIDAL, *J. Pharmacol. Exp. Therap.*, 1967, **158**, 91.
7. S. W. PELLETIER in "Chemistry of the Alkaloids", van Nostrand-Reinhold, New York, 1970, p. 1.
8. K. WADA, S. MARUMO and K. MUNAKATA, *Tetrahedron Lett.*, 1996, 5179.
9. R. N. CHOPRA, S. L. NAYR and I. C. CHOPRA, "Glossary of Indian Medicinal Plants", C.S.I.R., New Delhi, 1956, p. 11.
10. J. D. HOOKER, "Flora of British India", 1879, Vol. II, p. 190 (Published under the authority of the Secretary of the State for India in Council, London, UK).
11. V. H. DESHPANDE, A. D. PENDSE and R. PENDSE, *Indian J. Chem., Sect. B*, 1977, **15**, 205.

12. S. GHOSAL, S. K. DUTTA and S. K. BHATTACHARYA, *J. Pharm. Sci.*, 1972, **61**, 1274.
13. H. K. SINGH and A. S. CHAWLA, *J. Pharm. Sci.*, 1970, **59**, 1179.
14. A. S. CHAWLA, A. H. JACKSON and P. LUDGATA, *J. Chem. Soc., Perkin Trans. 1*, 1982, 2903.
15. K. ITO, M. HARUNA, V. JINNO and H. FURUKAWA, *Chem. Pharm. Bull.*, 1976, **24**, 52.
16. M. H. SARRAGIOTTO, H. L. FILHO and A. J. MARSAIOLI, *Can. J. Chem.*, 1981, **59**, 2771.
17. J. BUCKINGHAM, "Dictionary of Organic Compounds", 5th ed., Chapman and Hall, New York, 1980, Vol. 5, p. 5061 (Compound no. S00825).
18. I. RUBINSTEIN, L. J. GOAD, A. D. H. CLAGUE and L. J. MULHEIRN, *Phytochemistry*, 1976, **15**, 195.
19. S. CHAPON and S. DAVID, *Bull. Chem. Soc. Fr.*, 1952, 456.
20. D. H. R. BARTON, *J. Chem. Soc.*, 1951, 1444.
21. K. ITO, M. HARUNA and H. FURUKAWA, *Yakugaku Zasshi*, 1973, **193**, 1611.
22. V. PRELOG, A. LANGEMANN, O. RODIG and M. TERNBAH, *Helv. Chim. Acta*, 1959, **42**, 1301.
23. M. CARMACK, B. C. KUSICH and V. PRELOG, *Helv. Chim. Acta*, 1951, **34**, 1601.
24. F. KONIUSZY and K. FOLKERS, *J. Am. Chem. Soc.*, 1950, **72**, 5579; 1951, **73**, 333.
25. G. L. SAUVAGE and V. BOEKELHEIDE, *J. Am. Chem. Soc.*, 1950, **72**, 2062.
26. G. L. SUVAGE, F. M. BERGER and V. BOEKELHEIDE, *Science*, 1949, **109**, 627.
27. H. M. CHAWLA and S. K. SHARMA, *Fitoterapia*, 1993, **64**, 88.