# 1-Tigloyl-3-acetylazadirachtol, a New Limonoid from the Marrango Tree, Azadirachta excelsa Jack (Meliaceae) $^{\Psi}$

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Manuscript received 1 September 1997

# The structure of a new limonoid from the Viarrango tree, *Azadirachta excelsa* Jack, has been established as 1-tigloyl-acetylazadirachtol (4) by 1D and 2D <sup>1</sup>H and <sup>13</sup>C nmr spectra.

The constituents of A. indica A. Juss (Meliaceae), also known as 'neem tree', have been investigated intensively by different research groups during the past decades because of their effects on feeding behaviour and development of insects and other biological activities<sup>2</sup>. Up till now, azadirachtin<sup>2-9</sup> is the most interesting component present in neem seed extracts since it has been found to be not only a very potent insect antifeedant but also an insect growth regulating agent<sup>2</sup>. Recently, another Azadirachta species occurring in the Philippines and other South Asian countries. A. excelsa Jack (Meliaceae), the 'marrango tree', was found to be a promising source for natural products with considerable insect feeding or development controlling properties<sup>10,11</sup>. We report on the isolation and structure determination by nmr of a new compound, 1-tigloyl-3acetylazadirachtol (4) from A. excelsa.

ether and aqueous methanol, and 3-tigloylazadirachtol (2)<sup>12</sup> and nimbolide (5)<sup>13</sup> by extraction of *A. excelsa* bark with ethyl acetate. MPLC over silica gel (Si60, 45–63  $\mu$ m 300 × 40 mm, gradient system PE/EtOAc/MeOH) and HPLC (LiChrospher RP-18, 5  $\mu$ m 250 × 4.6 mm, MeCN/water = 3.5/6.5) led to the pure compounds.

The structure of 4 was established by <sup>1</sup>H and <sup>13</sup>C nmr spectroscopy (Tables 1 and 2). The assignments of the <sup>1</sup>H nmr signals were achieved by means of H,H-COSY and nuclear Overhauser difference spectroscopy (n.O.e., Table 3). The <sup>13</sup>C signals were assigned on the basis of DEPT spectra and by comparison to already described azadirachtin analogs. Comparison of the <sup>13</sup>C-spectroscopic data of 1<sup>4,7</sup> and 4 showed that the hemiacetal carbon signal of C-11 is shifted from  $\delta$  104.10 in 1 to  $\delta$  79.19 in 4 (Table 2) As proved by D<sub>2</sub>O exchange, only one tertiary OH (20-OH.  $\delta$ 



# **Results and Discussion**

Azadirachtin (1). marrangin  $(3)^{11}$  and 1-tigloyl-3-acetylazadirachtol (4) were isolated by extraction of *A. excelsa* seeds with petrol ether, followed by partition between petrol 2.78) and one secondary OH group (7-OH,  $\delta$  3.01) arc present in the spectrum of 4. This led us to assume that 4 is an azadirachtin-ll-desoxo compound of the tigloy-lazadirachtol type. In the H,H-COSY and long-range H,H-

1		TABLE 1-	- <sup>1</sup> H Nmr Dat	ra (250 MHz	; CDCl <sub>3</sub> , TMS =	= 0) OF COMPOUNDS	1-4		
•н		1		2		3			4
1-H	4.75 d	d (2.9, 3.1)		3.52 ddd (2.3	3, 2.7, 5.8)	5.30 t (	2.8)	4./100(2.9,2.9) 2 20 44/2 0 2 0)	
2-H <sub>α</sub>	2.34 dd (	16.7, 2.9, 2.7)	2	2.32 ddd (16.	3, 2.3, 2.8)	2.34	m	2.29 dd	(2.9, 2.9)
2-H <sub>β</sub>	2.13 ddd	(16.7, 3.1, 2.9)	2	2.22 ddd (16.	3, 2.7, 2.8)	2.34	m 2 (1)	2.99 dd	(2.9, 2.9)
3-H	5.50 d	d (2.9, 2.7)		5.53 dd (2	.8, 2.8)	5.54 [ (	2.9) 12.5)	5.50 dd	(2.9, 2.9)
5-H	5.53			) D 25.55 C	12.7)	) 0 66.6	12.5)	) 45.5 1	1(12.3) (135.38)
0-11	4.00 00	1(12.3, 2.7)		4.55 00 (1.	(2.5)	4.46 uu (12	2.3, 2.7)	4.55 001	(12.3, 2.0) d (2.8)
/-H	4.7.	5 d (2.7)		4.72 U	(2.3)	4./10	,2.0) s	4.//	u (2.0) D8 d
9-n	-	5.54 \$		3.194	(1.3)	2.81	5	J., 4 50	s (hr)
11-11	A 6	-		4.470	(1.5)	4 58 4 4	3 7)	4.53	d (3.6)
13-H	4.0	/u(3.4) /12.0 5.1 3.4)		1 33 4 (	(3. <i>3)</i> 12 9)	1 20 ddd (13	4 5 3 3 9)	4.05 d (5.0)	
10-Π <sub>α</sub>	1.75 000 (	A (13 0)	1	65 ddd (12	0 3 0 5 3)	1 29 d (	(3.4)	1.27 d (13.0)	
17.U	2.31	a(13.0)			(5 3)	2 34	m	7 37 4 (5 3)	
17-11	2.3	01.0		2.500	5.5)	1.83	s	2.37 U (3.3) 1 98 e	
10-FI 10 U	2.63	2 4 (0 6)		3 40 4 (	(9.4)	3 75 d (	9.8)	1.70 S 3 70 4 (0 2)	
19-Πα 10 μ	J.U. A 14	5 d (9.6)		1 3 05 d (	(9.4) (9.4)	3 93 d (	9.8)	3 88	d (9.8)
<sup>19-Πβ</sup>	4.1.	5 G ( 9.0)		5.55 @ (	,,) S	5.67	s,	3.00 U (7.0) 5 67 e	
21-11	5.04	() () S () () () () () () () () () () () () () (		5 03 d (	2 9)	5 06 đ (	2.9)	5 ()6 A (2 Q)	
22-11	6.44	(2.)) (d (2.9)		6 43 d (	2.9)	6.47 d (	2.9)	6 45 d (2.9)	
23-11	4.05	3 d (9 0)		4.04 d (	9.0)	4.12 d (	8.9)	4.09 d (8.9)	
28-11α 28-11-	3 76	(0.0)		3.83 d (	9.0)	3.78 d (	8.9)	3.71 0	1 (8.9)
20-Πβ 30-H	5.70	74 s		1.45	s	1.39	S	1.42 s	
1-04		_		3.41 dd (	- (5.8)	-		-	_
7-0H	3	.02 s		3 29 s (br)		2.70 s		3.01 s	
11-01	5	.05 s		-		-			
20-01	3	.11 s		2.78 s (	br)	2.71 s		2.78 s (br)	
12-0CH	3	.68 s		3.76	S	-	-		6 s
29-0CH.	3	.76 s		3.76 s		3.80 s		3.8	0 s
3.04c	1	95 s		-		1.97 s		1.9	96
U-OAC	2.	-		-		2.00 s	;	-	-
Tiglovi-									
3'-H	6.93 ag	(7.0, 1.5)		6.95 qq (7.0	), 1.3)	6.92 qq (7.)	i, 1.4)	6.92 gg (	7.0, 1.5)
4'-H	1.78 dq	(7.0, 1.1)		1.79 dq (7.0	), 1.3)	1.78 dg (7.1	l, 1.1)	1.78 dq (7.0, 1.3)	
5'-H	1.85 dq	(1.5, 1.1)		1.84 dq (1.3	8, 1.3)	1.85 s (br)		1.85 dq (1.5, 1.3)	
TABLE 2-13	C Nmr Data (62.	89 MHz; CDCI	3, TMS = 0) o	ог Сом-				Table	-2(contd.)
	PO	DUNDS 1-4			C-18	18.49 q	18.56 q	18.93 q	16.10 q
<sup>13</sup> C	1	2	3	4	C-19	69.07 t	71.43 t	71.12 t	69.50 t
C-1	70.51 d	69.37 d	71.23 d	70.26 d	C-20	83.55 s	83.71 s	83.57 s	83.60 s
C-2	29.37 t	32.09 t	30.64 t	30.20 t	C-21	108.70 d	109.16 d	10 <b>8.76 d</b>	108.68 d
C-3	66.99 d	67.69 d	67.12 d	66.99 d	C-22	107.30 d	107.53 d	107.53 d	107.52 d
C-4	52.52 s	53.34 s	52.47 s	52.50 s	C-23	147.00 d	146.79 d	147.15 d	147.14 d
C-5	37.06 d	35.18 d	36.90 d	36.97 d	C-29	173.20 s	174.12 s	173.26 s	173.31 s
C-6	73.79 d	74.32 d	74.03 <b>d</b>	76.49 d	C-30	21.33 q	21.35 <b>q</b>	20.54 q	21.03 q
C-7	74.37 d	73.68 d	72.99 d	73.73 d	12-OCH3	53.52 q	53.01 q	-	52.72 q
C-8	45.41 s	44.04 s	43.26 s	44.39 s	29-OCH <sub>3</sub>	52.72 q	52.63 q	52.72 q	52.72 q
C-9	` 44.69 d	43.80 d	47.10 d	44.97 d	СН <sub>3</sub> СОО	169.50 s	-	169.71 s	169.72 s
C-10	50.19 s	51.24 s	47.40 s	49.26 s	CH3COO	20.88 q	-	20.96 q	20.91 q
C-11	104.10 s	79.48 d	99.64 d	79.19 d	CH <sub>3</sub> COO	-	-	170.00 s	-
C-12	171.70 s	173.52 s	-	172.58 s	CH <sub>3</sub> COO	-	-	20.54 q	-
C-13	68.53 s	66.59 s	66.51 s	66.79 s	C-1'	166.22 s	167.08 s	165.87 s	166.14 s
C-14	69.69 s	69.43 s	69.36 s	69.50 s	C-2'	128.60 s	128.49 s	128.77 s	128.65 s
C-15	76.43 d	76.16 d	76.18 d	76.17 d	C-3'	137.50 d	138.86 d	137.44 d	137.52 d
C-16	25.04 t	25.08 t	25.14 t	25.22 t	C-4′	14.29 q	14.69 q	14.27 q	14.33 q
C-17	48.67 d	48.99 d	48.28 d	48.39 d	C-5′	11.94 q	12.09 q	11.96 q	11.97 q

COSY 2D-nmr spectra of 4, a crosspeak was observed between 9-H  $\delta$  3.28 and the signal at  $\delta$  4.59. Irradiation at  $\delta$ 4.59 caused a n.O.e. on 9-H and 30-H (Table 3). These effects are in accordance with a proton attached to C-11. These n.O.e. effects, H,H-COSY couplings and a similar chemical shift of C-11 ( $\delta$  79.48) are also observed in the spectrum of 3-tigloylazadirachtol (2)<sup>12</sup>. All other n.O.e's (Table 3) are typical for azadirachtin and related compounds, particularly the n.O.e 3'-H/18-H indicating that the tiglate group is attached to C-1<sup>4,7</sup>.

TABLE 3-NUCLEAR OVERHAUSER EFFECTS IN THE <sup>1</sup> H Nmr SPECTRUM (250 MHz, CDCl <sub>3</sub> ) of 1-TigloyL-3-ACETYLAZADIRACHTOL (4)				
Irradiated	Observed			
18-H	9-H, 17-H, 3'-H			
30-Н	6-Н, 7-Н; 15-Н, 19-Н <sub>в</sub> ; 11-Н; 21-Н			
9-Н	18-H; 5-H, 11-H			
11-H	9-H, 30-H			
3′-Н	18-H, 4'-H			

Bioassay :

In the 24h dual choice antifeedant test against the Mexican bean beetle *E. varivestis*<sup>14</sup>, compounds 1–5 showed high activity (Table 4), marrangin (3) and 1-tigloyl-3acetylazadirachtol (4) being more potent than azadirachtin (1).

	TABLE 4-ANTIF	EEDANT ACTIV	VITY AGAINST	E. varivestis	
[ppm]	1	2	3	4	5
EC 50	13	30	6	6	90
EC100	120	150	50	50	>500

Compounds 1-5 were also tested for antimicrobial activity<sup>15</sup> against *Bacillus subtilis* (Gram-positive) and *Pseudomonas stutzeri* (Gram-negative) (Table 5). Nimbolide (5) showed inhibitory effects even at a concentration as low as 0.5  $\mu$ g/spot. All other compounds possessed no bactericidal activity up to a concentration of 100  $\mu$ g/spot.

T WILE 5-BACTERICIDAL ACTIVITY AGAINST Bacillus subtilis AND Pseudomonas stutzeri								
[µg/spot]	1	2	3	4	5			
B subtilis	>100	>100	>100	>100	5			
P stutzeri	>100	>100	>100	>100	5			

#### Experimental

A. excelsa, bark and seed kernels, were kindly provided by Prof. Dr. H. Schmutterer and Dr. K. Ermel, Institut für Phytopathologie, Universitat Giessen.

Nmr measurements were performed with a Bruker WM 250 instrument : <sup>1</sup>H nmr 250 MHz, <sup>13</sup>C nmr 62.89 MHz; solvent CDCl<sub>3</sub>, internal standard TMS. Mass spectra (El-MS, 70 eV) were recorded on a Varian MAT 311 A or Varian

MAT 44 S instrument. Optical rotation was recorded on a Perkin-Elmer 241 polarimeter.

Isolation of 1, 3, and 4 : Ground A. excelsa seed kernel (140 g) was stirred with petrol ether (5 × 3 dm<sup>3</sup>; b.p.30–50°) for 24 h. Partition between petrol ether and aqueous methanol of the extract gave 45 g oil and 1.5 g methanol/ water phase. Chromatography of the methanol/water phase on LiChroprep Si 60 (45–63,  $\mu$ m 300 × 40 mm gradient system PE/EtOAc/MeOH) gave 10 fractions.

Azadirachtin (1) : HPLC on LiChrospher (RP-18, 5  $\mu$ m, 250 × 4.6 mm, MeCN/water = 3.5/6.5; flow 0.8 ml min<sup>-1</sup>,  $\lambda$  216 nm and RI detection) of fraction 8 (120 mg) gave 1 mg 1 ( $t_{\rm R}$  = 16 min),  $R_{\rm t}$  = 0.18 (TLC, silica gel 60 F<sub>254</sub> 0.25 mm plates, Merck, Darmstadt, PE/EtOAc/acetone/EtOH = 72 : 13 : 12 : 3).

Marrangin (3) : HPLC on LiChrospher (RP-18, 5  $\mu$ m, 250 × 4.6 mm, MeCN/water = 3.5 : 6.5; flow 1 ml min<sup>-1</sup>,  $\lambda$ 216 nm and RI detection) of fraction 10 (90 mg) gave 1.5 mg 3 ( $t_{\rm R}$  = 11 min),  $R_{\rm f}$  = 0.10 (Si60, PE/EtOAc/acetone/ EtOH = 72 : 13 : 12 : 3).

*l-Acetyl-3-tigloylazadirachtol* (4) : HPLC on LiChrospher (RP-18, 5  $\mu$ m, 250 × 4.6 mm, MeOH/water = 4:6; flow 1 ml min<sup>-1</sup>,  $\lambda$  216 nm and RI detection) of fraction 9 (80 mg) gave 5.5 mg 4 ( $t_{\rm R}$  = 14 min),  $R_{\rm f}$  = 0.18 (Si60, PE/EtOAc/acetone/EtOH = 72 : 13 : 12 : 3); C<sub>35</sub>H<sub>44</sub>O<sub>15</sub>, calcd. 704.2667, found *m/z* 704.2675 (HRMS); <sup>1</sup>H and <sup>13</sup>C nmr spectra in Tables 1 and 2.

λ(nm)	589	578	546	436	405	365	313
$[\alpha]^{25}_{\lambda}$	-12	-16	-20	-51	-72	-103	-26
(CHCl <sub>1</sub> ,	0.1)						

Isolation of 2 and 5: Powdered A. excelsa bark (500 g) was successively stirred three times with 5 dm<sup>3</sup> petrol ether, ethyl acetate, and methanol for 24 h. Chromatography on LiChroprep Si60 (45–63  $\mu$ m, 300 × 40 mm, gradient system PE/EtOAc/MeOH) of the crude EtOAc extract (4.5 g) gave 9 fractions.

3-Tigloylazadirachtol (2) : HPLC on LiChrospher (RP-18, 5  $\mu$ m, 250 × 16 mm, MeCN/water = 4 : 6; flow 4.5 ml min<sup>-1</sup>,  $\lambda$  216 nm and RI detection) of fraction 8 (200 mg) gave 2.5 mg 2 ( $t_{\rm R}$  = 14 min),  $R_{\rm f}$  = 0.16 (Si60, PE/EtOAc/ acetone/EtOH = 72 : 13 : 12 : 3).

Nimbolide (5) : HPLC on LiChrospher (RP-18, 5  $\mu$ m, 250 × 16 mm, MeCN/water = 4 : 6; flow 4.5 ml min<sup>-1</sup>,  $\lambda$  216 nm and RI detection) of fraction 3 (340 mg) gave 33 mg 5 ( $t_{\rm R}$  = 20 min),  $R_{\rm f}$  = 0.33 (Si60, PE/EtOAc/acetone/EtOH = 72 : 13 : 12 : 3).

## Acknowledgement

Financial support by Fonds der Chemischen Industrie is gratefully acknowledged.

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