
“Off-Target” Therapeutic Mechanisms of “Target-Based” Drugs

Rationality or Luckiness?

Supplementary Data 2

for

Biological Sciences and Physics Unified: Internal Evolution and Urging the Second Scientific Revolution

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See the main article for the detailed method.

Below most target-tables, there is the STRING interaction network along with its permalink, from which the interactive network along with relevant data and files can be accessed; STRING png and svg image files, STRING interaction and annotation files and the downloaded and deduplicated citations are also available at doi.org/10.5281/zenodo.5732942.

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
DONEPEZIL	acetylcholinesterase	ACHE	P22303 41

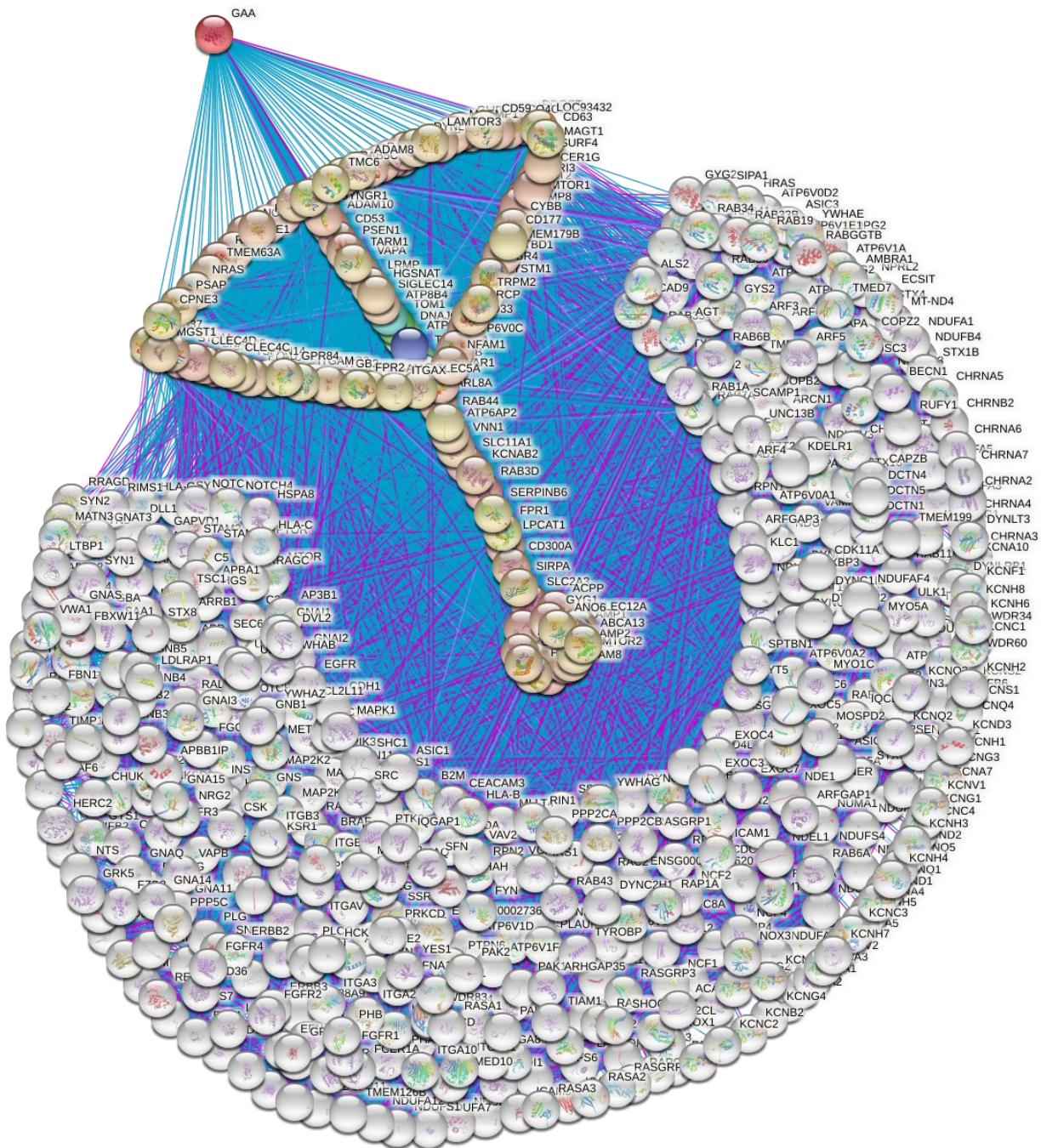
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1. prevents glutamate neurotoxicity through $\alpha 4$ - and $\alpha 7$ -nicotinic acetylcholine receptors, followed by activation of PI3-Akt pathway and downregulation of NMDA receptors(1-5)
2. protective effect against oxygen–glucose deprivation-induced injury independent of muscarinic cholinergic system and nicotinic cholinergic system(6)
3. modulates the vasoconstrictive effects of A β peptides at the level of skin microvasculature, independent of a direct action on smooth muscle cell reactivity or on endothelial cell function in the absence of A β (7)
4. neuroprotective activity against depolarization-induced toxicity via inhibition of the rapid influx of sodium and calcium ions, and via decrease of glutamate and glycine release(8)
5. anti-inflammatory properties independent of its acetylcholinesterase inhibition(9)
6. directly inhibits microglial activation induced by A β through blocking MAPK and NF- κ B signaling(10)
7. increases mitochondrial biogenesis through AMP-activated protein kinase in the hippocampus(11)

8. improves neuropathy through activation of AMPK pathway(12)
9. modulates a T helper 2 bias via α 7-nAChR leading to increased expression of naturally occurring auto-antibodies against amyloid beta (A β)(13)
10. attenuates vascular reactivity dysfunction by decreasing reactive oxygen species generation and increasing NO bioavailability; possibly via increased endothelial NO synthase activity, and inhibition of NADPH oxidase activity(14)
11. direct non-competitive inhibition of nAChR(15)
12. increases the activity of α -secretase and decreases the activity of β -secretase(16)
13. inhibitory effects on the production of PGE2, TXB2, COX-1 and COX-2 mRNA and protein expression in macrophages(17)
14. antioxidant properties(18, 19)
15. inhibits microglial activation and release of proinflammatory cytokines(20)
16. enhances A β clearance across the blood-brain barrier and liver by upregulating the expression of A β major transport proteins(21)
17. promotes phagocytic activity of microglial cells through the PI3K pathway(22)
18. agonistic activity at sigma-1 receptor(23-28)
19. inhibits a canonical inflammatory NF- κ B signaling and microglial activation independent of acetylcholine(29)
20. stimulates oligodendrocyte differentiation and myelin-related gene expression via nicotinic acetylcholine receptors in neural stem cell-derived oligodendrocyte progenitor cells(30)
21. attenuates vascular dementia through increasing BDNF by reducing HDAC6 nuclear translocation(31)
22. produces acute vasodilation induced by a selective activation of neuronal nitric oxide synthase in the cerebral parenchymal arterioles(32)
23. ameliorates A β impairments in hippocampal long-term potentiation(33, 34)
24. increases blood flow and reduces ischemia-induced cellular injury(35-37)
25. neuroprotective effects via reducing the efflux of lactate dehydrogenase which is induced by A β (1-42)(38)
26. enhances the survival of newborn neurons in the hippocampal dentate gyrus via CREB signaling(39)
27. neuroprotection by upregulation of BDNF(40)
28. inhibits neuronal apoptosis and regulates synaptic plasticity, through the upregulation of p-Akt, p-GSK-3 β Bcl-2 and the downregulation of Bax, Caspase-3(41)
29. inhibits the inflammatory response and the increase of the intracellular reactive oxygen species induced by bradykinin via nAChR and PI3K-Akt pathway in astrocytes(42)
30. promotes neurogenesis via Src signaling pathway(43)
31. inhibits apoptosis of endothelial cells(44)
32. improves cognitive function by increasing the hippocampal production of IGF-I through sensory neuron stimulation(45)
33. inhibits acetylcholine esterase activity and increases the release of sAPP α not only through a muscarinic receptor pathway, but also by

- directly enhancing trafficking and activity of both TACE and ADAM 10(46)
34. prevents A β 42-induced neurotoxicity through the activation of PI3K/Akt and inhibition of GSK-3, as well as through the activation of nAChR(47)
 35. increased neuronal viability with reduced p-tau by enhancing protein phosphatase 2A activity(48)
 36. reverses the age-related downregulation of the growth hormone/insulin-like growth factor-1 axis(49)
 37. potentiates neuronal differentiation by enhancing the activation of ERK(50)
 38. promotes neurite outgrowth(51)
 39. regulates serum adipokine level(52)
 40. modulates monocyte chemotactic protein-1 and IL-4 production, which may reflect a general shift towards type Th0/Th2 cytokines which could be protective in Alzheimer(53)
 41. inhibits microtubule affinity-regulating kinase (MARK4)(54)
 42. neuroprotective effects via inhibiting voltage-gated calcium and potassium and sodium channels(55, 56)
 43. upregulates and sensitizes α 7-nAChR(57)
 44. reduces A β by increasing the activity of phospholipase A2(58)
 45. reduces the phosphorylation of tau protein by phosphorylating and inactivating GSK-3 β (58)
 46. decreases the expression of the pro-apoptotic protein, glyceraldehyde-3-phosphate dehydrogenase(59)
 47. decreases the expression of miR-206-3p(60)
 48. attenuates A β -associated mitochondrial dysfunction and reduces mitochondrial A β accumulation(61)
 49. rescues spatial learning and memory deficits following traumatic brain injury independent of its effects on neurogenesis(62)
 50. attenuates cell apoptosis induced by oxygen-glucose deprivation via blocking Kv2.1 potassium channels(63)
 51. delays cellular senescence that is promoted under high glucose condition via activation of SIRT1 and inhibiting the generation of reactive oxygen species(64)
 52. neuroprotective effects via PINK 1 that is related to mitophagy and cellular protection from mitochondrial dysfunction(65)
 53. upregulates proteins relevant to axon guidance, cytoskeleton, and mitophagy, and modulates the proteins connected to mTOR and MAPK pathways(65)
 54. decreases the level of A β by increasing SNX33 expression and APP cleavage by α -secretase(66)
 55. neuroprotection via inhibition of sodium channels(67)
 56. modulates the insulin and IGF-1 signaling pathways(68)

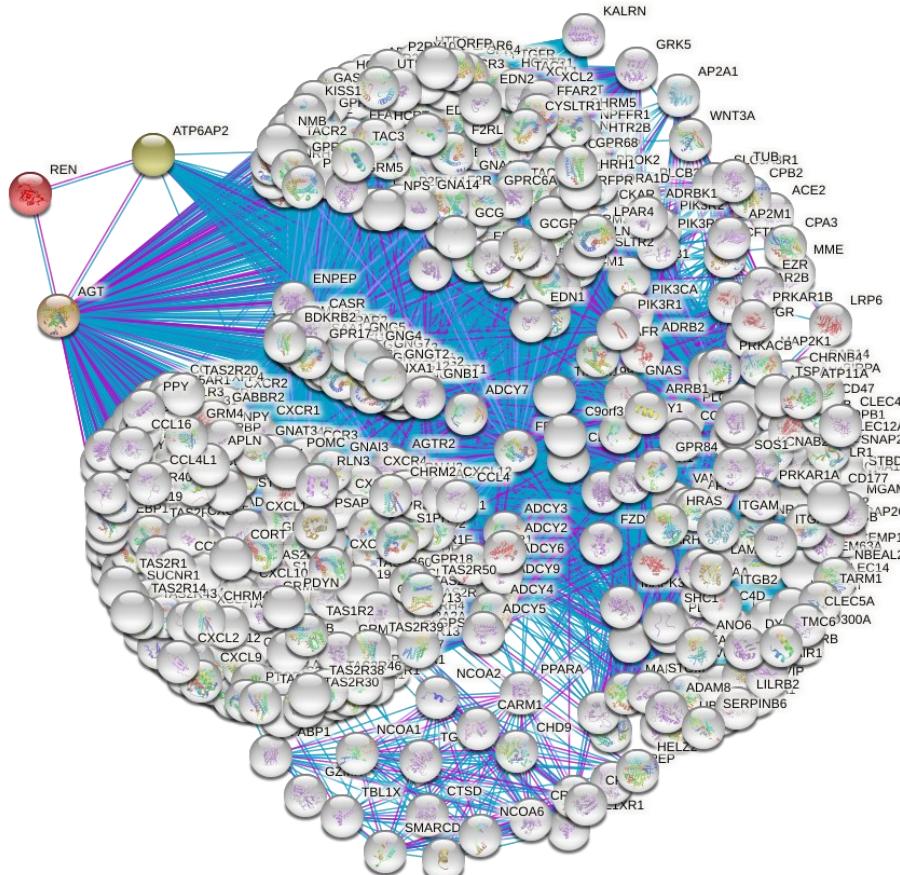
Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
ACARBOSE	alpha glucosidase	GAA	P10253 14



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1. directly inhibits the absorption of free glucose and some other hexoses(69, 70)
2. prevents pancreatic islet hypertrophy and augments islet blood flow(71)
3. regulates lipid metabolism(72)
4. downregulates sucrase-isomaltase and sGLT1 gene expressions and upregulates GLUT4 mRNA and protein expressions(73)
5. ameliorates endothelial barrier dysfunction by directly inhibiting NLRP3 inflammasome(74)
6. inhibits glucose-6-phosphatase and hepatic glucose production(75)
7. regulates thyroid hormones(76)
8. delays gastric emptying(77)
9. regulates gut microbiome and their fermentation products(78, 79)
10. activates miR-10a-5p and miR-664 and regulates the MAPK pathway(80)
11. attenuates insulitis via anti-inflammatory actions(81)

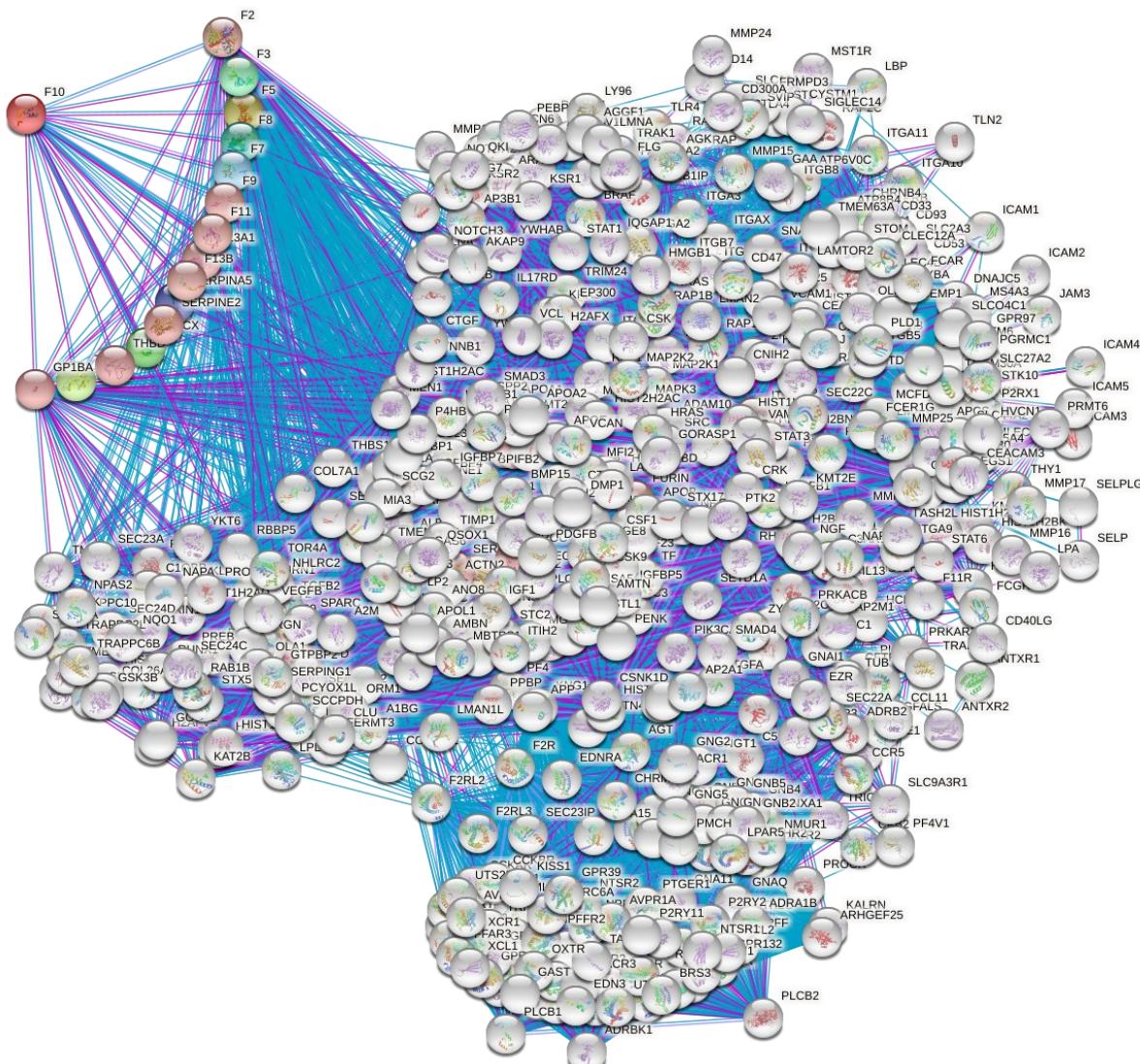
Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
ALISKIREN	renin	REN	P00797 12



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1. protects against myocardial I/R injury via eNOS dependent mechanisms(82)
2. increases cardiomyocyte survival via increasing autophagosomal formation and decreasing apoptosis and necrosis via modulating AMPK expression(83)
3. improves endothelium-dependent relaxation and oxidative stress by activating PI3K/Akt/eNOS pathway(84)
4. protects endothelial function and improves impaired nitric oxide bioavailability(85)
5. downregulates the expression of the (pro)renin and AT₁ receptors(86)
6. attenuates myocardial ischemia-reperfusion injury by a bradykinin B₂ receptor- and angiotensin AT₂ receptor-mediated mechanism(87)
7. improves vascular endothelial function and platelet-endothelium activation independent of antihypertensive effects(88)
8. cardioprotective properties and beneficial effects on oxidative stress biomarkers(89)
9. upregulates pro-angiogenic cells and reduces atherogenesis independent of blood pressure lowering(90)
10. favorable effects on endothelial function and epithelial progenitor cells, reduced arterial stiffness, and improved left ventricular twisting and untwisting, independent of blood pressure(91)
11. attenuates myocardial apoptosis and oxidative stress(92)
12. ameliorates aortic endothelial dysfunction and oxidative vascular remodeling via elevating plasma nitric oxide metabolite levels and reducing systolic hypertension, insulin resistance, dyslipidemia, aortic lipid peroxide levels and aortic wall hypertrophy(93)
13. positive effects on cardiac function via MMP-9(94)
14. inhibits intracellular angiotensin II levels without affecting (pro)renin receptor signals(95)
15. improves vascular remodeling partially via increasing vasculature NO production(96)
16. reduces pulmonary vein arrhythmogenic activity with a direct vasodilatory property(97)
17. positive inotropic effect on cardiomyocytes(97)
18. improves endothelium-dependent vasodilation and NO availability in the peripheral resistance arterioles probably because of antioxidant activities(98)
19. inhibits cardiac hypertrophy and fibrosis, independent of blood pressure reduction, by blocking Ang II-PKC β I-ERK1/2-regulated autophagy(99)
20. improves the endothelial repair capacity of endothelial progenitor cells via Tie2/PI3k/Akt/eNOS Pathway(100)
21. protects against myocardial ischemia/reperfusion injury by activating the PI3K-Akt-eNOS pathway
22. direct effects on myocardial matrix turnover and beneficial effects on diastolic function(101)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
RIVAROXABAN			7
APIXABAN			2
EDOXABAN	coagulation factor X	F10	
BETRIXABAN		P00742	1



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Rivaroxaban

1. fibrinolytic effect(102)
2. protective, repairing and fibrinolytic effects on vascular endothelium(103)
3. directly affects endothelial cells and activates the NO-mediated vasorelaxant pathway(104)
4. ameliorates angiotensin II-induced cardiac remodeling by attenuating TXNIP/Trx2 interaction(105)
5. protects against atherosclerosis by increasing the anti-inflammatory cytokine IL-10(106)
6. improves blood flow recovery and increases neovascularization and promotes the functions of endothelial progenitor cells, including migration, tube formation, and senescence via NO-related pathways(107)
7. anti-atherosclerotic effects by regulating the expression of genes in the TLR4/NF-κB pathway(108)

8. reduces TRAP-induced platelet aggregation(109)
9. inhibited tissue factor-induced platelet aggregation(110)
10. reduces DNA oxidative changes and vascular endothelial damage via inhibiting reactive oxygen species production(111)

Apixaban

1. directly affects endothelial cells and activates the NO-mediated vasorelaxant pathway(104)
2. reduces TRAP-induced platelet aggregation(109)

Edoxaban

1. improves atrial fibrillation and thromboembolism through regulation of the Wnt-β-induced PI3K/ATK-activated protein C system(112)
2. improves venous thrombosis by decreasing hydrogen sulfide and homocysteine through the PI3K/AKT pathway(113)

Drugs	Discovery Target		
ZANAMIVIR	OSELTAMIVIR	PERAMIVIR	Influenza Neuraminidase

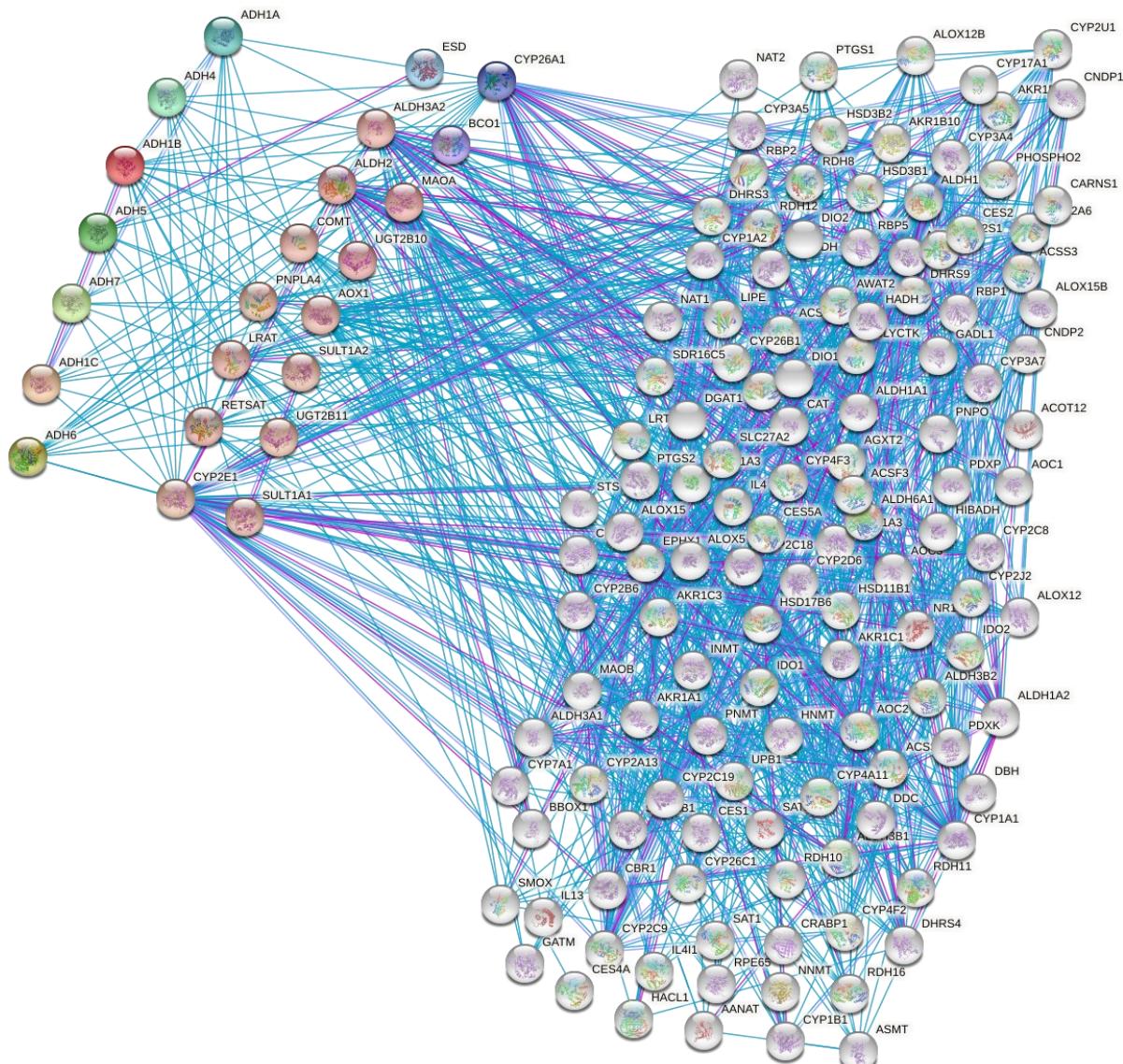
Zanamivir

1. reduces intense infiltration of pulmonary tissues by macrophages and consequent pulmonary inflammation by reducing nitric oxide production in influenza virus-infected and gamma interferon-activated RAW 264.7 macrophages(114)
2. reduces lung inflammation and damage by inhibiting nitric oxide production(115)

Oseltamivir

1. reduces intense infiltration of pulmonary tissues by macrophages and consequent pulmonary inflammation by reducing nitric oxide production in gamma interferon-activated RAW 264.7 macrophages(114)

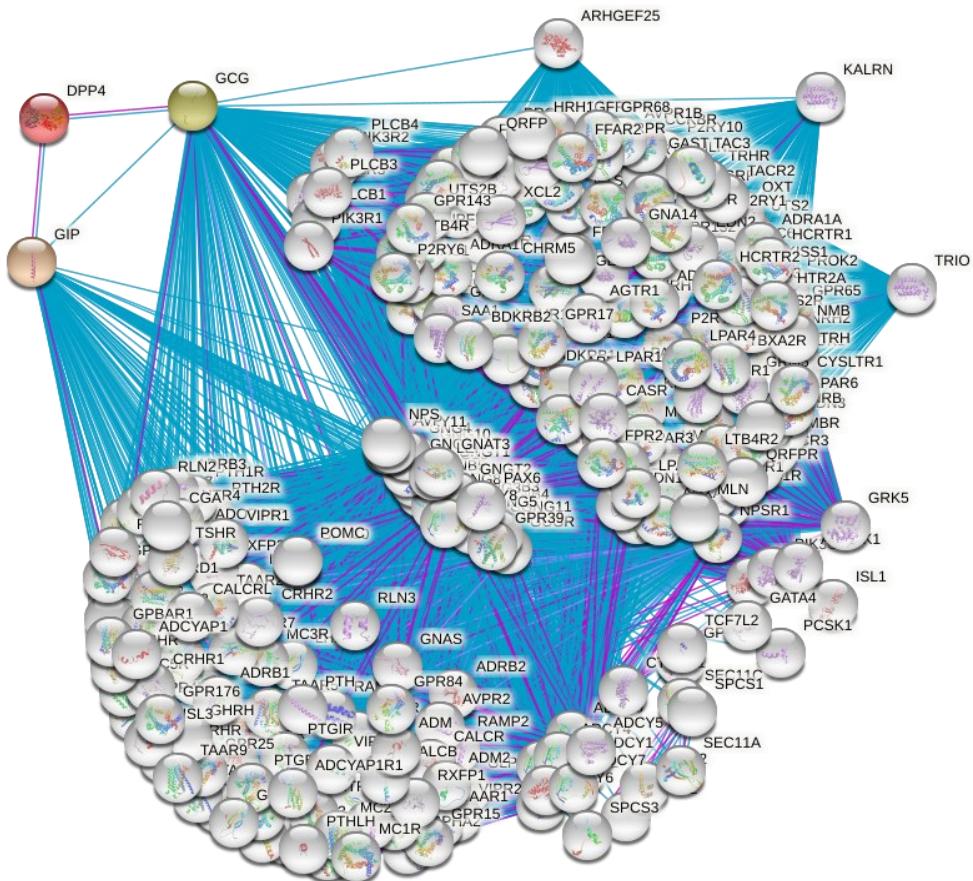
Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
FOMEPIZOLE	Alcohol dehydrogenase	ADH1A ADH4 ADH1B ADH5 ADH7 ADH1C ADH6	P07327 P08319 P00325 P11766 P40394 P00326 P28332



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1. Protection of gastric mucosa independent of alcohol dehydrogenase inhibition and via prevention of acute vascular injury through endogenous sulphydryls, prostaglandins(116, 117)
2. Gastroprotection via increasing adherent gastric mucus and reducing oxidative stress(118, 119)
3. Reduces catalase-H₂O₂ system activity by reducing H₂O₂ availability(120)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
SITAGLIPTIN			13
SAXAGLIPTIN	dipeptidyl peptidase 4	P27487	20
LINAGLIPTIN	DPP4		1
ALOGLIPTIN			1



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Sitagliptin

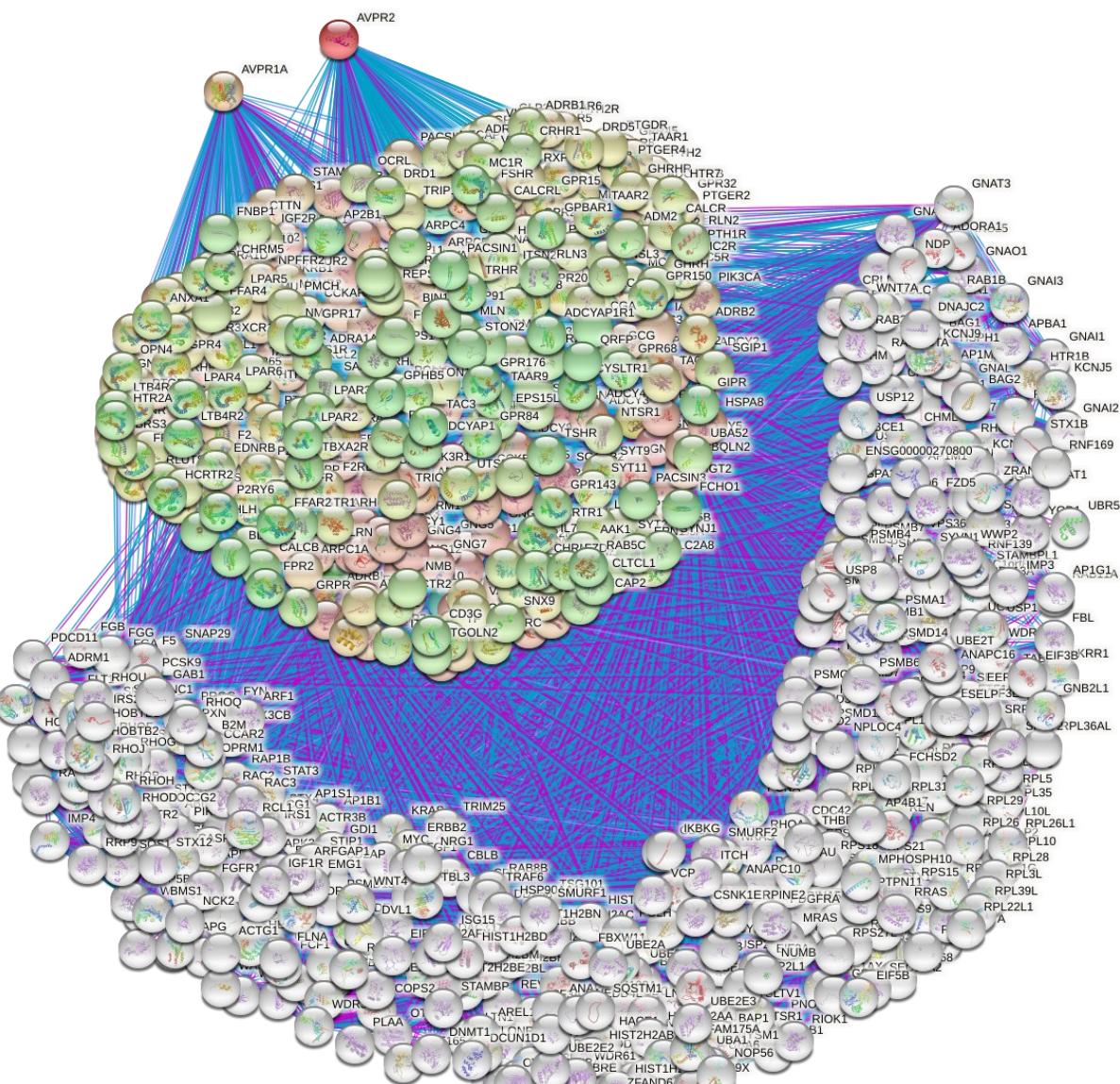
1. in addition to its well-known hypoglycemic action, may also have beneficial effects on hyperglycemia-induced vascular changes in an endothelium-dependent manner(121)
2. direct influence on the diabetes-induced dysfunctional collagen metabolism(122)
3. prevents high glucose-induced apoptosis via activation of AMP-activated protein kinase in endothelial cells(123)
4. ameliorates oxidative stress in experimental diabetic nephropathy by diminishing the miR-200a/Keap-1/Nrf2 antioxidant pathway(124)
5. anti-inflammatory effects in adipose tissue and in pancreatic islets that accompany the insulinotropic effect(125)
6. prevents nitrosative stress, inflammation and apoptosis in retinal cells induced by diabetes and exerts beneficial effects on the blood-retinal barrier integrity(126)
7. protects against vascular inflammation via the SIRT6/ROS-dependent signaling pathway(127)
8. Modulation of the RBP4-GLUT4 system may be one of the mechanisms by which sitagliptin ameliorates the symptoms of type 2 diabetes mellitus(128).
9. reverses the diabetes-induced endothelial-mesenchymal transition of human aortic endothelial cells as well as the cardiac function through the SDF-1 α /PKA pathway(129)
10. alleviates diabetes-induced cardiac injury by reducing nitrooxidative stress and promoting autophagy(130)
11. improves glucose-lipid metabolism and protects kidney and vascular endothelial cells during diabetic nephropathy pathogenesis through inhibiting iNOS expression(131)
12. attenuates endothelial cell dysfunction and atherogenesis via attenuation of TNF α -mediated induction of NF- κ B and orphan nuclear receptor NUR77 mRNA expression, inhibiting TNF α induction of PAI-1, ICAM-1 and VCAM-1 mRNA and protein expression(132)
13. ameliorates diabetes-induced renal injury via inhibiting the TGF- β /Smad pathway(133)
14. improves diabetes-induced endothelial impairment via mechanisms related to anti-peroxynitrite and promoting autophagy(134)
15. ameliorates diabetic nephropathy fibrosis via MAPK/ERK pathway(135)
16. ameliorates renal tubular injury in diabetic kidney disease via STAT3-dependent mitochondrial homeostasis through SDF-1 α /CXCR4 pathway(136)
17. ameliorates insulin resistance by inhibiting inflammatory responses and activating autophagy via AMPK/mTOR signaling pathway(137)

Saxagliptin

1. ameliorates insulin resistance by reducing endoplasmic reticulum stress(138)
2. induces β -cell proliferation through increasing stromal cell-derived factor-1 α (139)
3. additional metabolic benefits via increased serum bile acid(140)

4. vascular benefits via regulating AP-1 and NF-κB(141)
 5. prevents cell death via extrinsic and intrinsic apoptotic pathways in pancreas by insulin resistance(142)
 6. anti-inflammatory effects via upregulation of IL-10(143)
 7. induces insulin secretion(144)
 8. can dose-dependently upregulate proliferator-activated receptor γ coactivator-1α and irisin, potentially improving insulin resistance and glycolipid metabolism and inhibiting inflammation(145)
 9. ameliorates diabetes-induced renal injury via inhibiting TGF-β/Smad pathway(146)
 10. protects β-cells in the pancreas by elongating the telomere length(147)
 11. modulates immune-relevant pathways in beta cells which may protect them from inflammation, for example, downregulates HLA Class I and II expression and upregulates the immune-regulatory molecules PD-L1 and CTLA4(148)
 12. improves cardiac function and glucose homeostasis and ameliorates β-Cell dysfunction via reducing S6K1 activation and IRS-1 and IRS-2 degradation(149)
 13. attenuates high glucose-induced alterations in migration, proliferation, calcification and apoptosis of vascular smooth muscle cells via ERK1/2 pathway(150)
 14. inhibits the tubulointerstitial Wnt/β-catenin pathway in diabetic nephropathy and provides renal protection by alleviating renal tubulointerstitial transdifferentiation and fibrosis(151)
 15. upregulates nesfatin-1 secretion(152)
 16. improves function of circulating proangiogenic cells(153)
 17. delays diabetic nephropathy progression, possibly via inhibiting ERK1/2 signaling and promotion of the interaction between GLP-1 and the GLP-1 receptor(154)
 18. accelerates endothelial regeneration through SDF-1/CXCR4 in a GLP1R-independent manner(155)
 19. exerts direct, DPP-IV independent effects on intestinal L cells, activating cAMP and ERK1/2 signaling and stimulating total GLP-1 secretion(156)
 20. reduces inflammatory cytokines and improves unfavorable M1/M2-like phenotypes of peripheral blood monocytes(157)
 21. reduces insulin resistance via downregulating RBP-4
 22. exerts favorable effects on the vascular endothelium by inhibiting ET-1 via suppressing the NF-κB/IκBα system through the activation of AMPK pathway(158)
- Linagliptin**
1. direct protective effect on β-cell function and survival(159)
- Alogliptin**
1. alleviates diabetes-induced ventricular hypertrophy, interstitial fibrosis, atrial remodeling and diastolic dysfunction via decreasing the production of reactive oxygen species, mitochondrial membrane depolarization and improving mitochondrial swelling and biogenesis via PGC-1α/NRF1/Tfam pathway(160, 161)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
CONIVAPTAN	V1A receptor V2 receptor	AVPR1A	P37288
TOLVAPTAN		AVPR2	P30518 6



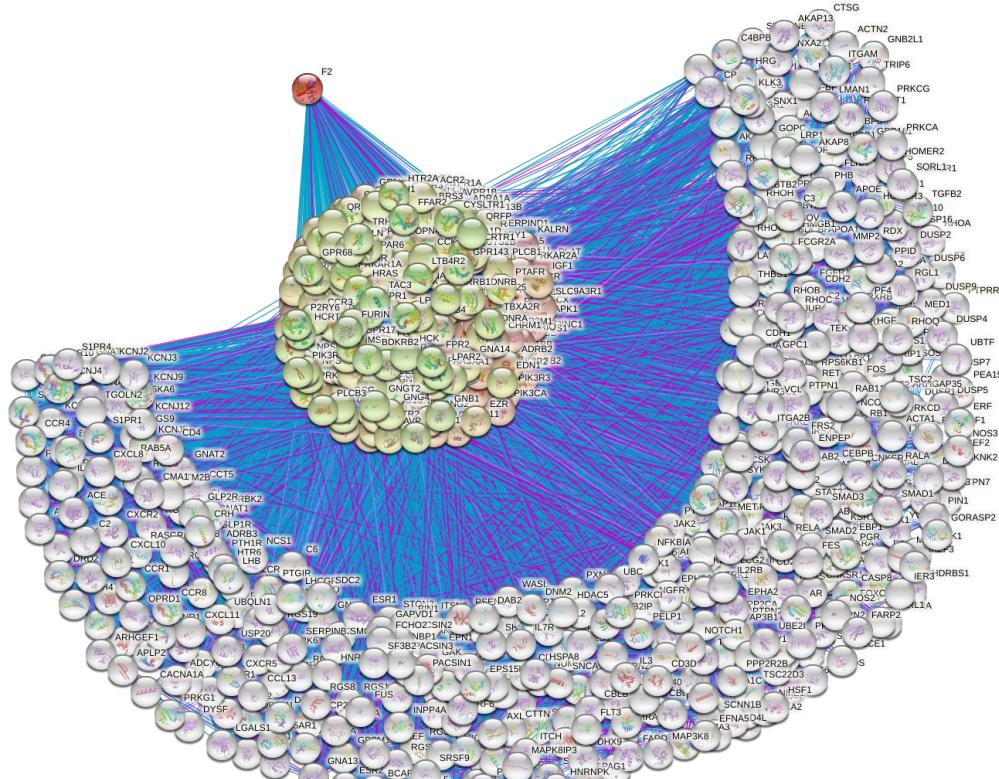
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Tolvaptan

1. inhibits adrenal aldosterone synthesis via V₂ receptor-independent pathway(162)
2. improves renal functions via activating the Nrf2/HO-1 antioxidant pathway through PERK phosphorylation(163)
3. renoprotective via inhibiting TNF α and monocyte chemoattractant protein-1 expression and NF κ B phosphorylation(164)

4. attenuates MI-induced mRNA expressions of atrial and brain natriuretic peptides, monocyte chemotactic protein-1, transforming growth factor- β 1, arginine vasopressin V(1a) receptor, and endothelin-1 in the marginal infarct region(165)
5. beneficial effects on atrial remodeling via miR-21/Spry1/ERK/MMP-9, miR-21/PTEN/PI3K/AKT, and NF- κ B pathways(166)

Drugs	Discovery Target	Uniprot ID	“Off-Target” Therapeutic Mechanisms
ARGATROBAN	coagulation factor II, thrombin	F2	1
DABIGATRAN		P00734	2



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Argatroban

1. anti-inflammatory effects via nitric oxide pathway(167)
2. improves microcirculation through increasing plasma NO(168)

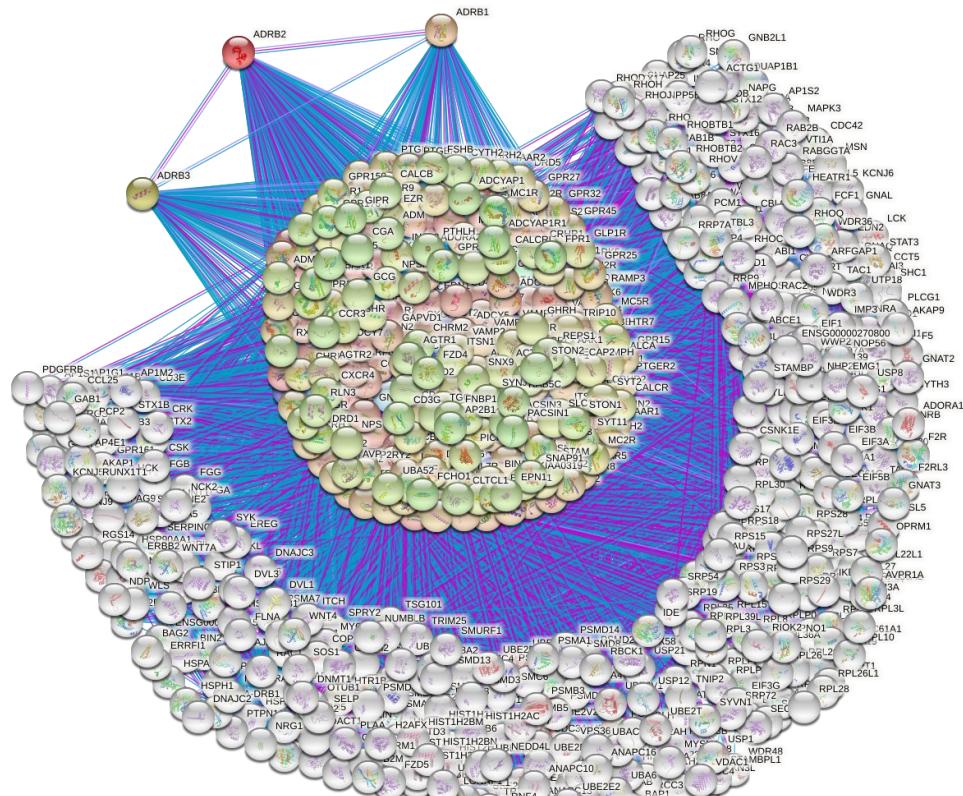
contributing to plaque stabilizing and atheroprotective effects(169)

2. reduces proinflammatory stimuli via reduced expression of cytokines and chemokines(170)
3. reduces DNA oxidative changes and vascular endothelial damage via inhibiting reactive oxygen species production(111)

Dabigatran

1. reduces proinflammatory M1 macrophages in atherosclerotic lesions, thereby

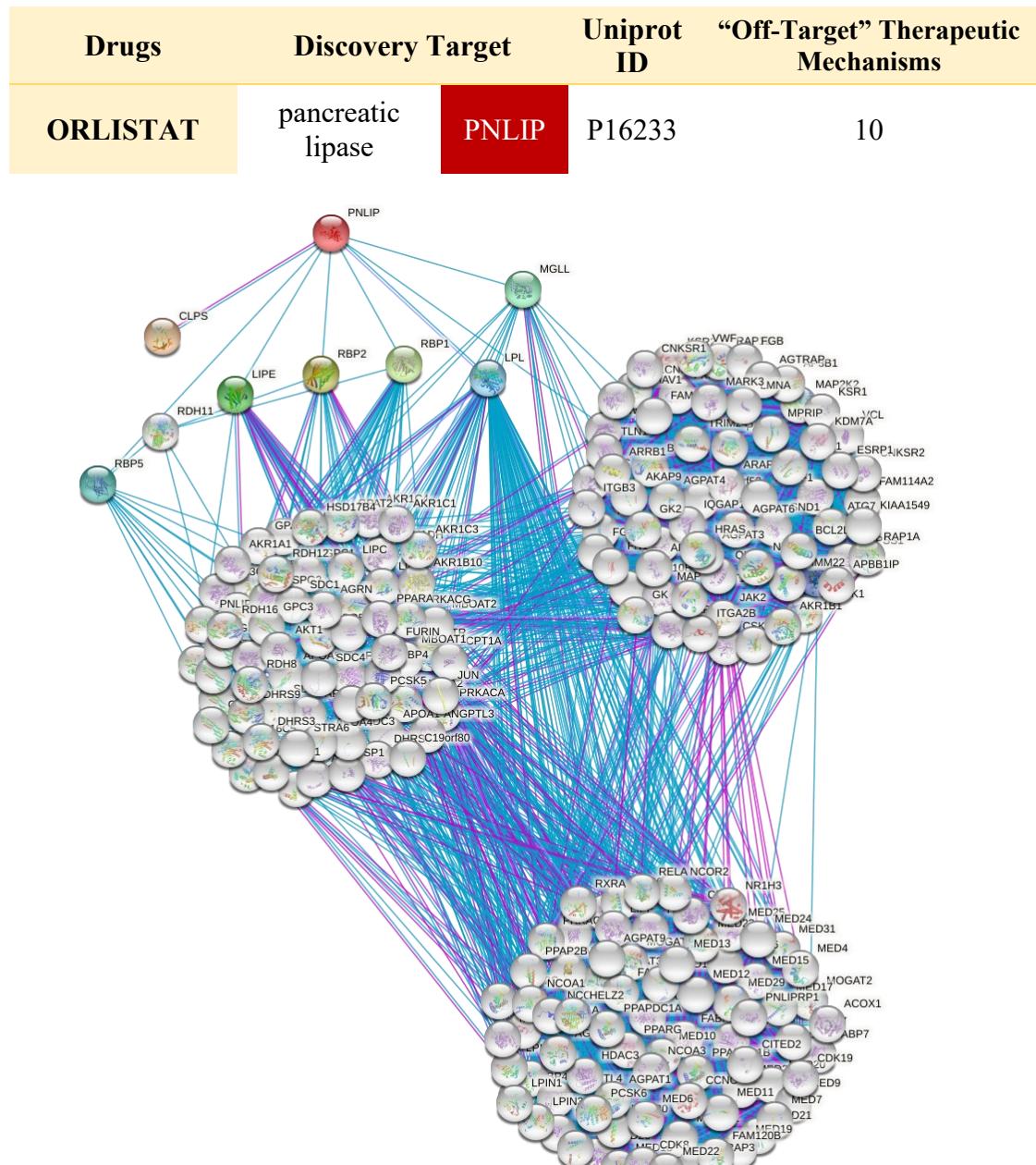
Drugs	Discovery Target	Uniprot ID	“Off-Target” Therapeutic Mechanisms
MIRABEGRON	β1-adrenoceptor β2-adrenoceptor β3-adrenoceptor	ADRB1 ADRB2 ADRB3	P08588 P07550 P13945
VIBEGRON			2



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Mirabegron

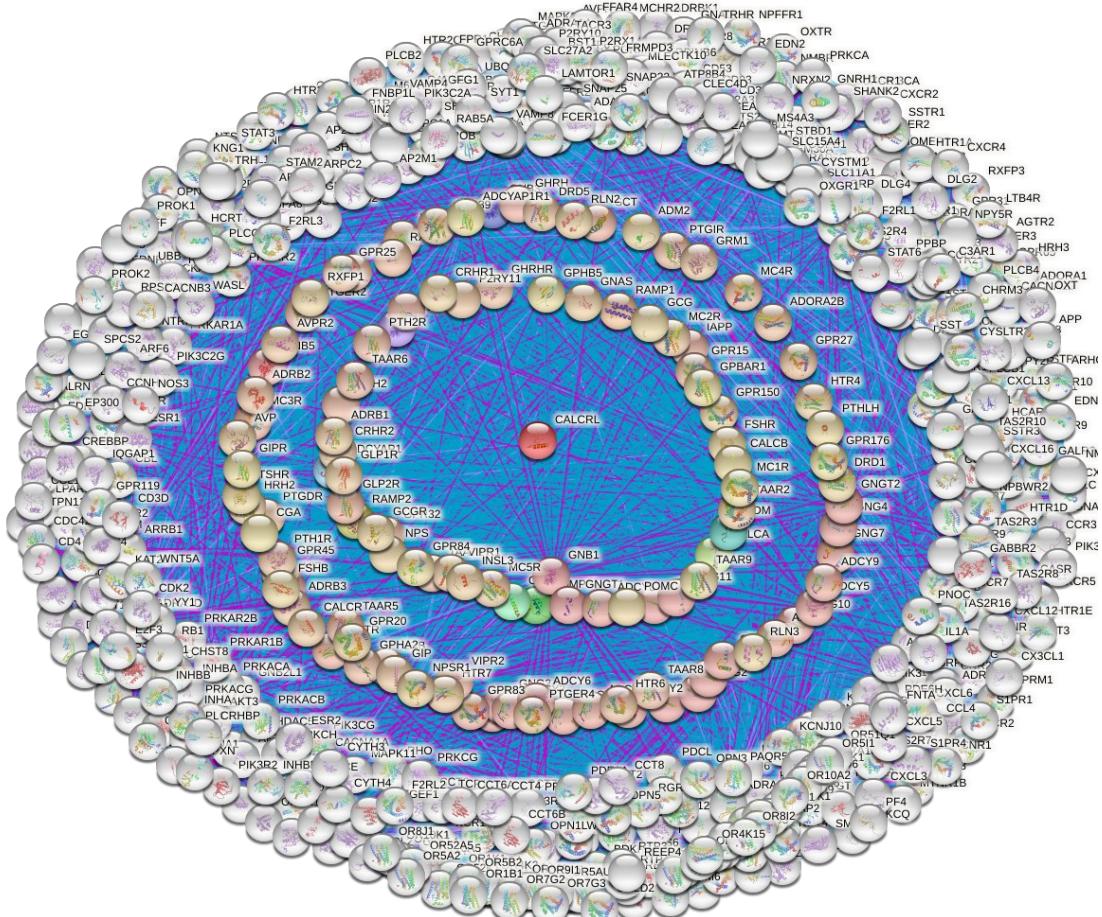
1. relaxes smooth muscle also through α_{1A} and α_{1D} -adrenoceptor antagonism(171)
2. relaxes smooth muscle via both β 3-adrenoceptor/cAMP-dependent and -independent pathways(172)
3. with the participation of endogenous adenosine may exert inhibitory effects on bladder functions increasing the storage capacity and prolonging the micturition interval, without affecting the voiding pressure or postvoid residual volume(173)



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1. increases postprandial **GLP-1** levels, thereby enhances insulin secretion and thus may lead to decreased food intake(174), also see(175, 176)
2. induces a weight-independent reduction in post-prandial **NEFA** levels along with increase in stimulated GH response, IGF-I levels, and IGF-I/IGFBP-3 ratio(177)
3. limits cholesterol absorption by the inhibition of **Niemann-Pick C1-like 1 transport protein**(178)
4. other targets identified by activity-based proteome profiling which have functions related to biogenesis, glycolysis, protein biosynthesis and GTPase activity: **Hsp90AB1, RPL14, RPS9, GAPDH, ANXA2, β-tubulin, RPL7a**(179)

Drugs	Discovery Target	Uniprot “Off-Target” Therapeutic ID	Mechanisms
UBROGEPANT			
RIMEGEPANT	Calcitonin receptor-like receptor	CALCRL Q16602	1

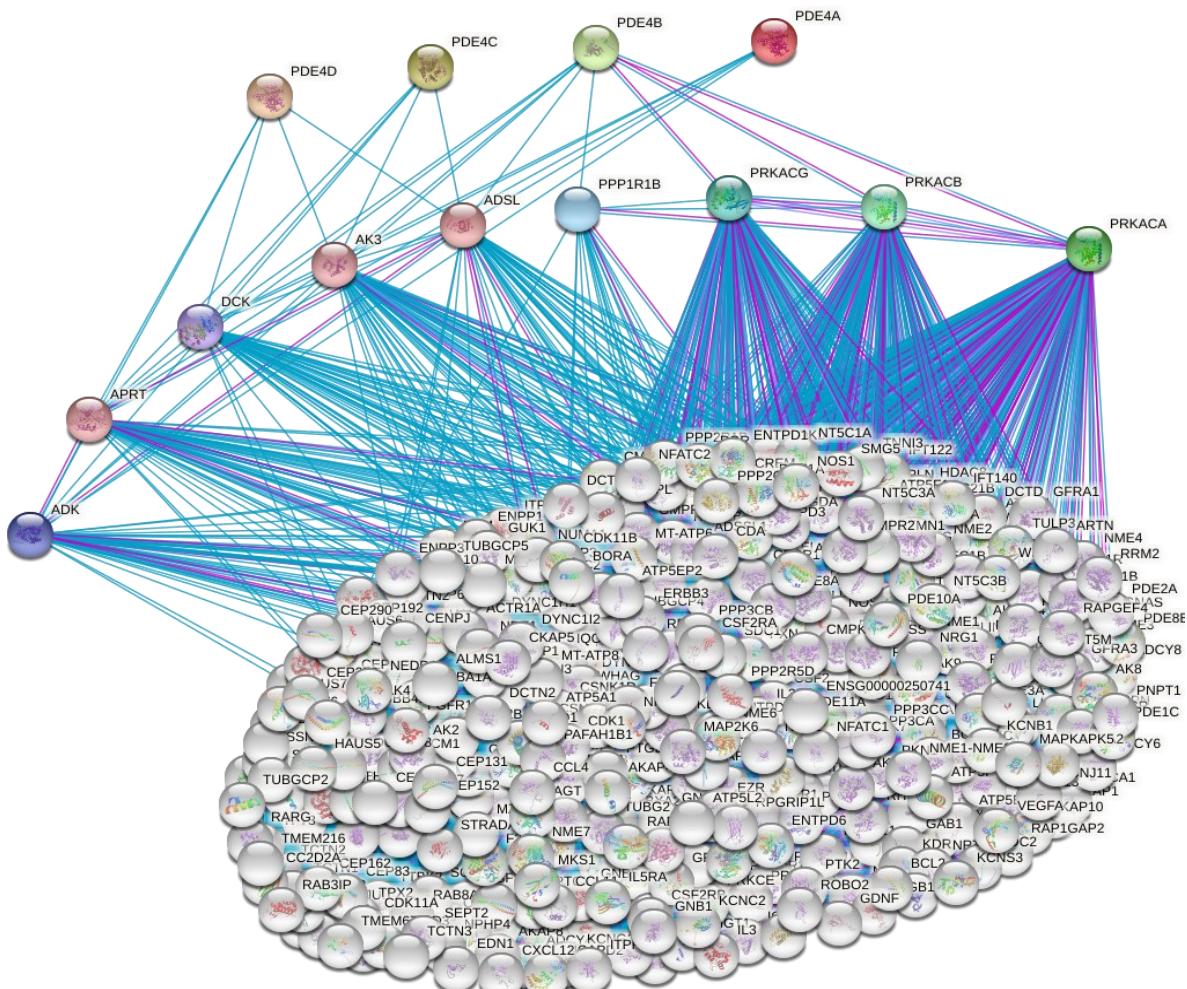


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Rimegepant

- effectively antagonizes α CGRP-mediated signaling through the **AMY₁** receptor, in addition to the CGRP receptor(180)

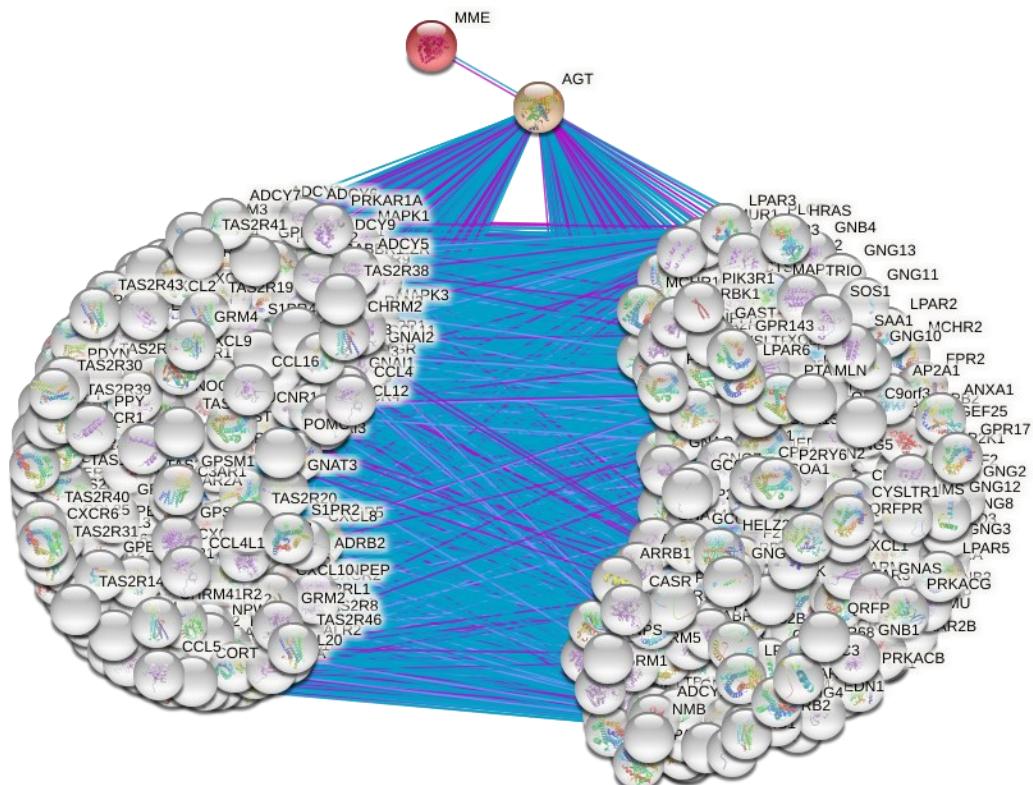
Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
ROFLUMILAST	phosphodiesterase 4A/4B/4C/4D	P27815 Q07343 Q08493 Q08499	9



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1. inhibits inflammatory mediators via suppression of NF κ B, p38 mitogen-activated protein kinase, and c-jun NH2-terminal kinase activation(181)
 2. may modulate the SCF/c-kit pathway(182)
 3. suppresses T cell proliferation by interfering with IP3-IP3R binding to inhibit calcium emission, blocking pathway activation from this phase onward, eventually decreasing the level of a growth factor for T cell proliferation, IL-2(183)
 4. modulates heme oxygenase-1 and its product carbon monoxide(184)
 5. activates cystic fibrosis transmembrane conductance regulator (CFTR)(185)
 6. upregulates glucocorticoid receptor (GR α) transcript levels(186)
 7. inhibits leukocyte-endothelial cell interactions, expression of adhesion molecules and microvascular permeability(187)

Drugs	Discovery Target	Uniprot ID	“Off-Target” Therapeutic Mechanisms
SACUBITRILAT	Neutral endopeptidase	MME	P08473 2

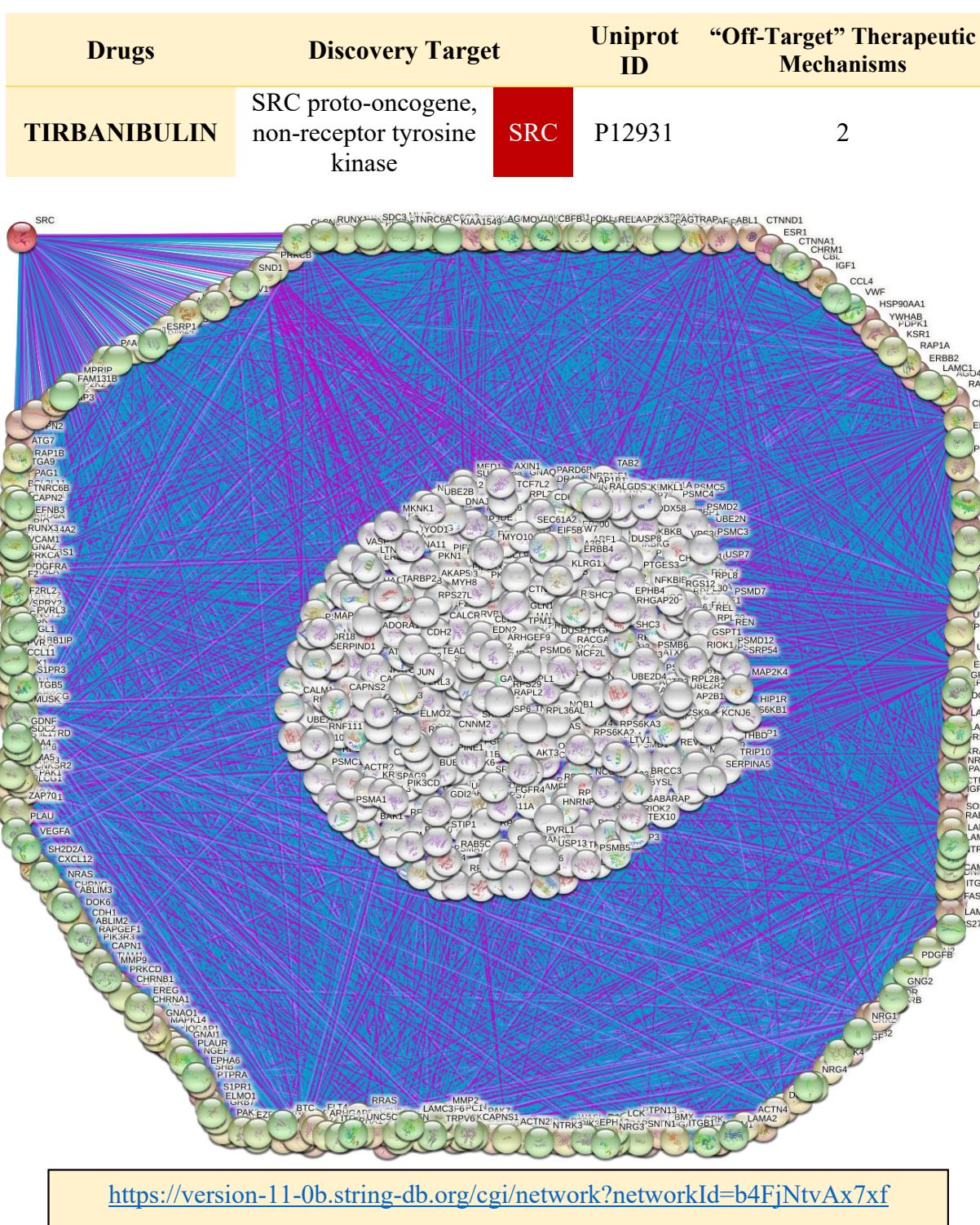


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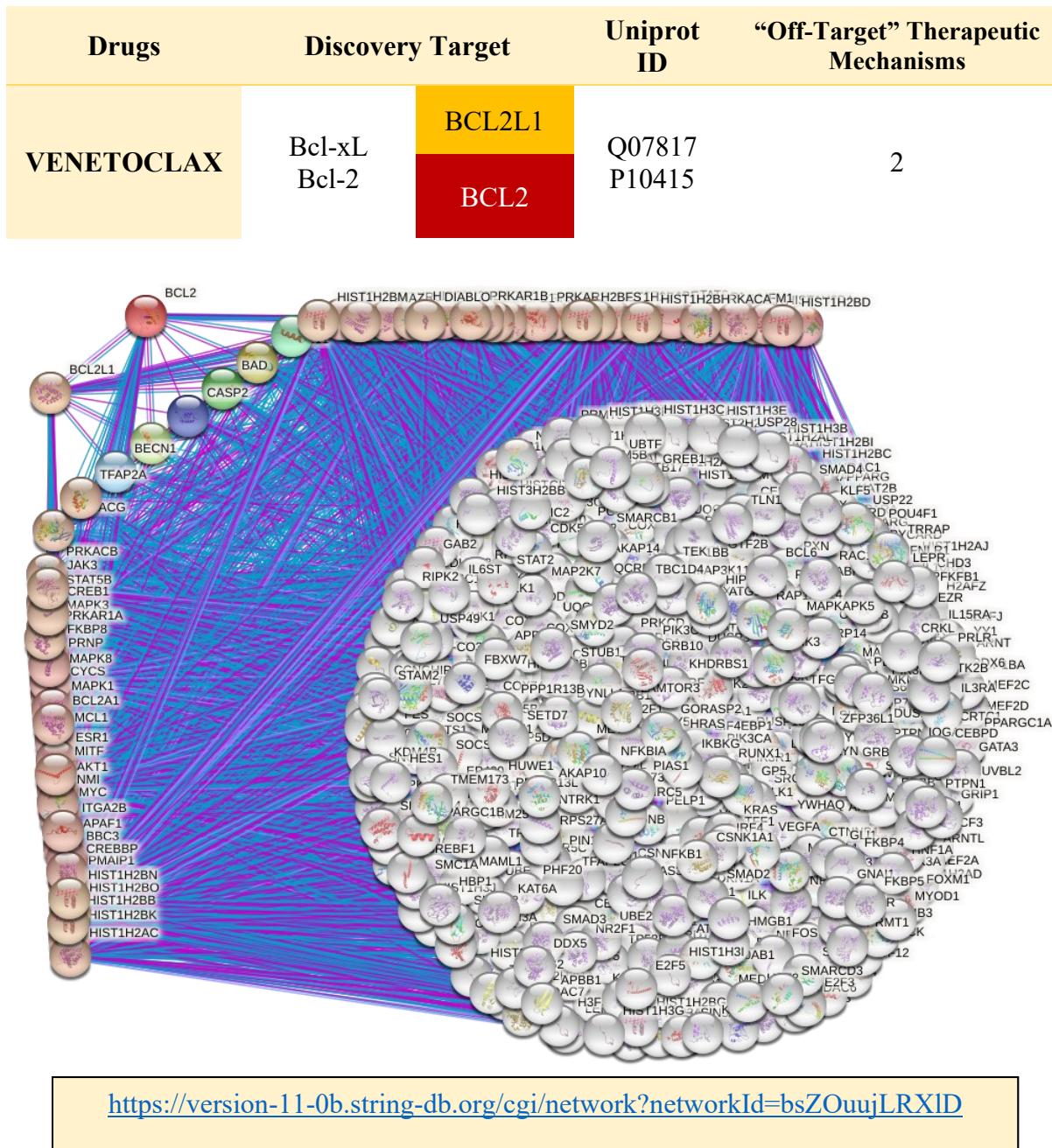
- directly improves Ca^{2+} homeostasis by reducing proarrhythmic sarcoplasmic reticulum Ca^{2+} leak without acutely affecting systolic Ca^{2+} release and

inotropy. These effects might contribute to its mortality benefits(188).

- antioxidant properties(189)



1. inhibits pre-tubulin and tubulin polymerization(190)
2. also inhibits additional key proteins involved in cell proliferation and fatty acid metabolism(190)



1. modulates the mitochondrial signaling pathway and ATP production(320, 321)

2. inhibits Nrf2 antioxidant pathway activation induced by hypomethylating agents(322)

Drugs	Discovery Target
SAQUINAVIR: 11	NELFINAVIR: 8
RITONAVIR: 8	LOPINAVIR: 2
INDINAVIR: 7	ATAZANAVIR: 1
	DARUNAVIR
	AMPRENAVIR
	HIV Protease
	TIPRANAVIR

Saquinavir

1. immunomodulation via upregulation of telomerase activity and interferon-gamma release in activated nonadherent mononuclear cells(191)
2. direct immunomodulatory effects(192)
3. enhances the humoral immune response, possibly through modulating T cell functions(193)
4. It reduces the release of interleukin 2 probably via direct interaction with one or more of the transport or regulatory drug binding sites of P-glycoprotein. Given that activated lymphocytes preferentially replicate HIV-1, this inhibitory effect may be beneficial(194). Also see (195)
5. inhibits apoptosis by mitigating oxidative stress(196)
6. increases interferon gamma production from stimulated splenocytes(195)
7. inhibits pathological inflammation by targeting the interaction of cathepsin V with TLR4/MyD88(197)
8. protective effect on ventilation-induced lung injury via inhibition of NF-κB activation and high-mobility group box 1 expression(198)

9. inhibits HIV replication not only by inhibiting the HIV protease but also by blocking microbial antigen-induced endothelial cell activation via blocking the TLR-and TNF-α-mediated NF-κB activation and proinflammatory cytokine production(199)
10. modulates antigen presentation, and T cell responses via inhibiting the chymotrypsin-like activity of the 20S proteasome(200)
11. immunomodulation via regulating the maturation of dendritic cells(201)
12. strong inhibitory effect on the T cell stimulatory capacity of mature dendritic cells(201)
13. inhibits TNF-alpha induced cell death(202)

Ritonavir

1. regulates T cell receptor excisional circles(203)
2. modulates antigen presentation, and T cell responses via inhibiting the chymotrypsin-like activity of the 20S proteasome(200, 204, 205)
3. inhibits apoptosis via inhibiting calpain(206)

4. induces cell-cycle arrest at G1-phase and apoptosis through downregulation of cell-cycle gene **cyclin D2** and antiapoptotic gene **survivin**. Also, it suppresses transcriptional activation of NF-κB in these cells. Thus, it may prevent EBV-associated lympho-proliferative diseases that occur in immunocompromised patients(207).
5. inhibits HIV replication not only by inhibiting the HIV protease but also by blocking microbial antigen-induced endothelial cell activation via blocking the TLR-and TNF-α-mediated **NF-κB** activation and proinflammatory cytokine production(199)
6. It reduces the release of interleukin 2 probably via direct interaction with one or more of the transport or regulatory drug binding sites of **P-glycoprotein**. Given that activated lymphocytes preferentially replicate HIV-1, this inhibitory effect may be beneficial(194).
7. modulates activation of peripheral blood CD4(+) T cells and decreases their susceptibility to apoptosis(208)
8. stimulates hematopoiesis and decreases apoptosis and ICE expression in CD34(+) cells(209)
9. is an immune modulator that may affect leukocyte activation and **apoptosis** as an important part of its therapeutic benefit(210)
10. immunomodulation via regulating the **maturation of dendritic cells**(201)
11. inhibits TNF-alpha induced cell death(202)
12. direct immunomodulatory effects(192)

Indinavir

1. inhibits HIV replication not only by inhibiting the HIV protease but also by blocking microbial antigen-induced endothelial cell activation via blocking the TLR-and TNF-α-mediated **NF-κB** activation and proinflammatory cytokine production(199)
2. inhibits apoptosis via inhibiting **calpain**(206, 211)
3. It reduces the release of IL-2 probably via direct interaction with one or more of the transport or regulatory drug binding sites of **P-glycoprotein**. Given that activated lymphocytes preferentially replicate HIV-1, this inhibitory effect may be beneficial(194).
4. independent of its antiviral activity, may modulate immunologic responses by prolonging cell survival via inhibiting their **entry** into cell cycle(212)
5. immunomodulation via regulating the maturation of dendritic cells(201)
6. slightly affects the dendritic cell induced proliferation of T cells(201)
7. induces restoration of both memory and naive CD8 T cells and improves T cell function, as assessed by enhanced lymphoproliferative capacity and diminished propensity to undergo **apoptosis**(213)
8. immunoregulation by regulating **dendritic cells'** functions and antifungal activity(214)
9. inhibits the trypsin-like and mostly chymotrypsin-like activities of purified **26S proteasome**(215)
10. inhibits TNF-alpha induced cell death(202)

Nelfinavir

1. inhibits HIV replication not only by inhibiting the HIV protease but also by blocking microbial antigen-induced endothelial cell activation via blocking the TLR-and TNF- α -mediated NF- κ B activation and proinflammatory cytokine production(199)
2. modulates activation of peripheral blood CD4(+) T cells and decreases their susceptibility to apoptosis(208)
3. protects against apoptosis via inhibiting the adenine nucleotide translocator pore function(216)
4. immunomodulation via regulating the maturation of dendritic cells(201)
5. inhibits host proteases and reduces immature dendritic cell transendothelial migration(217)
6. activates PP2 and inhibits MAPK signaling in macrophages and may offer beneficial effects independent of antiviral activity by reducing severity of chronic innate immune activation in HIV-1 infection(218)
7. It triggers inflammasome formation and elicits an IL-1R-dependent inflammation. Inflammasomes are critical sensors that convey cellular stress and pathogen presence to the immune system(219).
8. inhibits HIV gp41-induced mitochondrial depolarization, unlike other HIV protease inhibitors or inhibitors of calpain and cathepsin(220)
9. augments thymic output independent of HIV(221)
10. increases CD4 T cells through a non-viral effect(222)
11. may exert some beneficial effects counteracting the increased mitochondrially driven apoptosis present in HIV-infected people(223)

Lopinavir

1. increases IL-8 mRNA expression(224)
2. upregulates expression of the antiviral protein ribonuclease L(225)

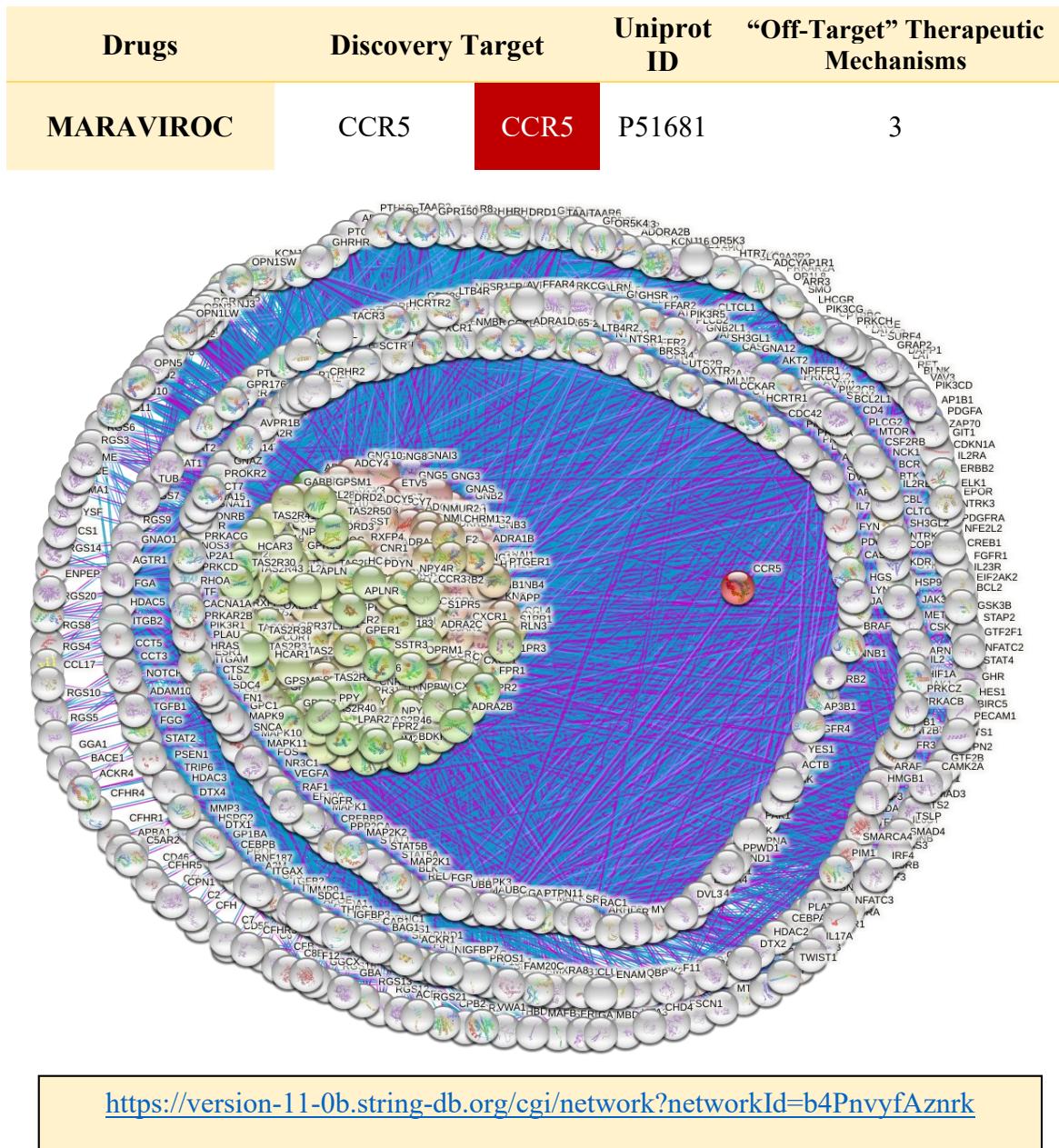
Atazanavir

1. increases IL-8 mRNA expression(224)

Drugs	Discovery Target
Raltegravir	
Dolutegravir	HIV Protease

Raltegravir

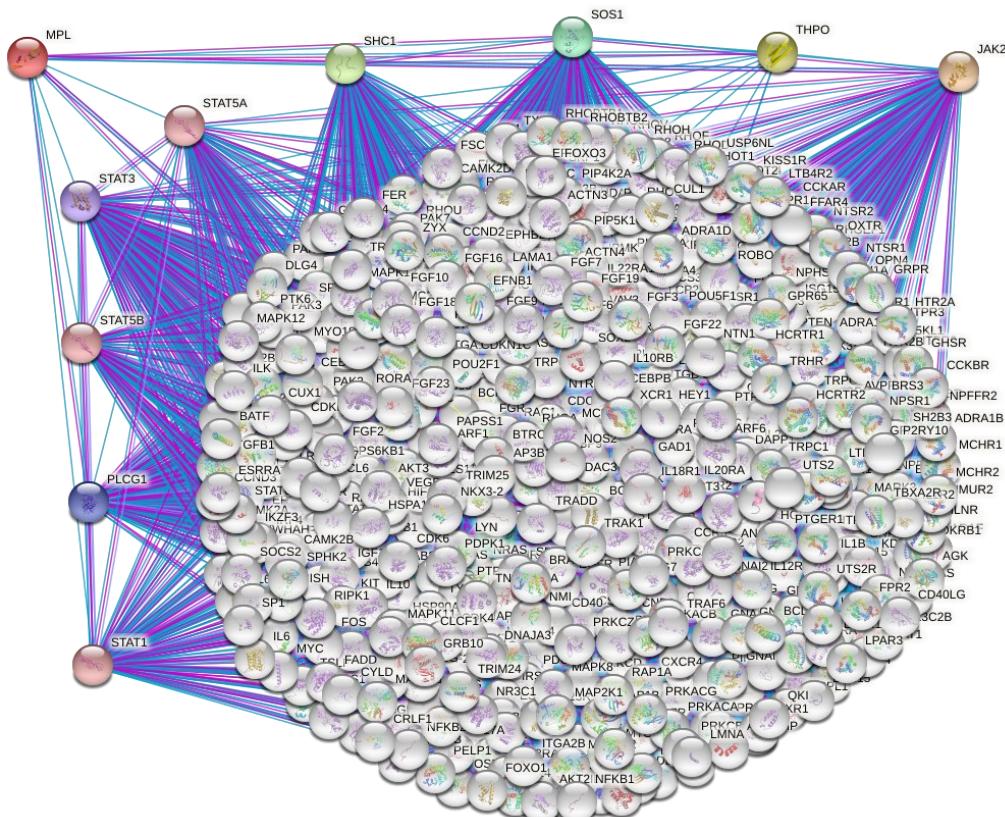
has profound and specific effects on the host transcription profile that may contribute to the overall antiviral activity of the drug(226)



1. It has additional immunological effects via decreasing some activation expression markers on T lymphocytes and also migration towards chemoattractants that might have potential capacity to inhibit HIV-associated chronic inflammation and activation, both by directly affecting T cell **activation** and by reducing **entrapment** of lymphocytes in lymph nodes(227).
2. may prevent neurologic disorders in HIV-infected individuals by also reducing inflammation in the brain(228)
3. contributes to the restoration of the homeostasis of regulatory T cell subsets(229)
4. downregulates HIV-associated chronic **inflammation** by blocking the recirculation and trafficking of macrophages and monocyte-derived dendritic cells(230)

5. reduces chronic inflammation via reducing activation and cytokine secretion of CD8+ T cells via a CCR5-independent pathway(231)
6. increases T cell activation(232)

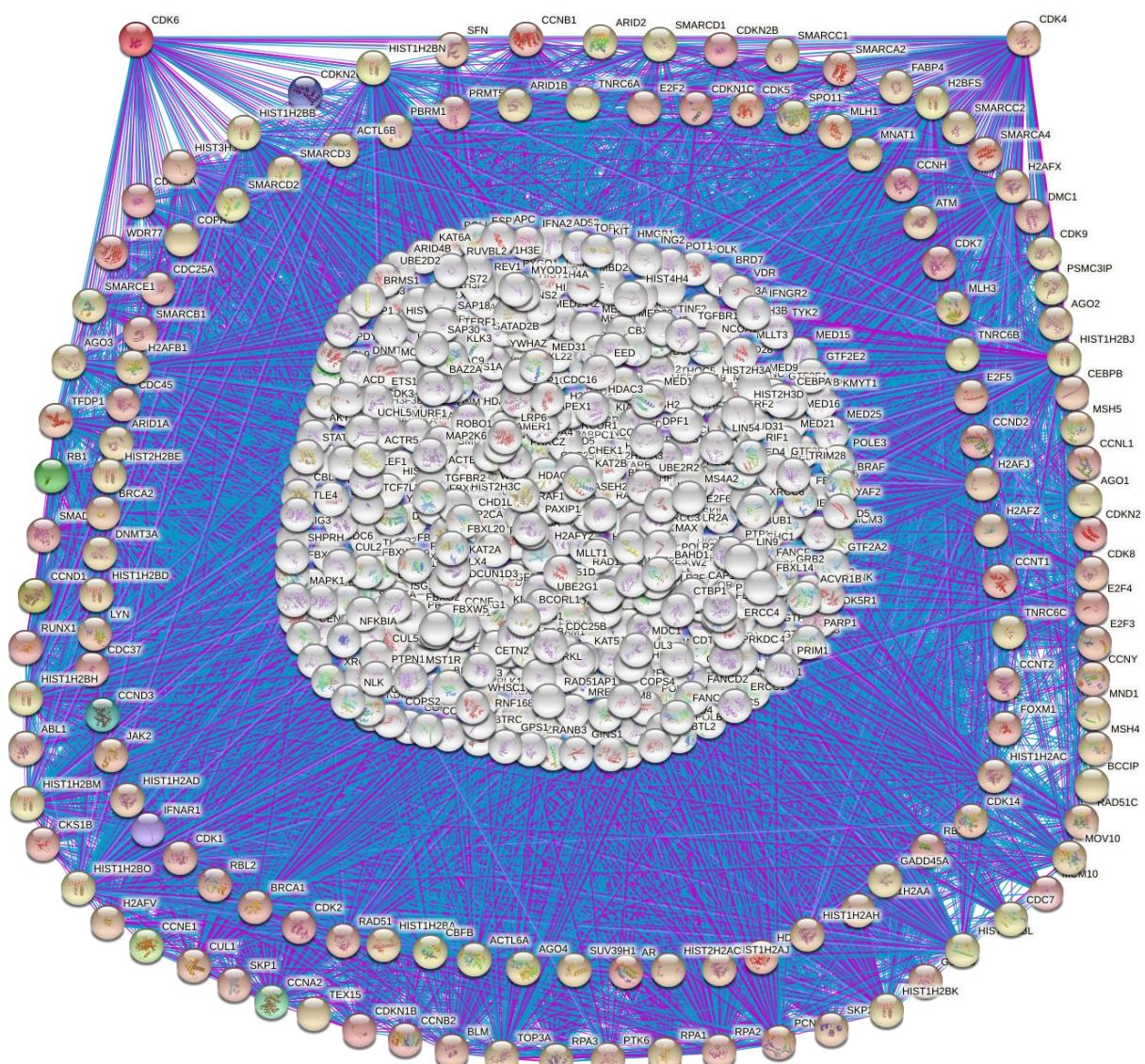
Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
ELTROMBOPAG			1
LUSUTROMBOPAG	Thrombopoietin receptor	MPL	P40238
AVATROMBOPAG			



<https://version-11-0b.string-db.org/cgi/network?networkId=bv094Im7lUmV>

Eltrombopag: stimulates hematopoiesis at the stem cell level through iron chelation-mediated molecular reprogramming, independent of thrombopoietin receptor(233)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
PALBOCICLIB	CDK4		38
RIBOCICLIB	CDK4 CDK6	P11802 Q00534	14
ABEMACICLIB	CDK6		17



<https://version-11-0b.string-db.org/cgi/network?networkId=bHxtGfTtlgsN>

Palbociclib

1. induces senescence via activating **proteasome**(234)
2. activates p53 via inhibition of the **PRMT5-MDM4** axis(235)
3. inhibits **other kinases**(236):
 - 1) AAK1
 - 2) AMPK α 1
 - 3) BMP2K
 - 4) CAMK2D
 - 5) CAMK2G
 - 6) CDK9
 - 7) CSNK2 α 1
 - 8) CSNK2 α 2
 - 9) DAPK3
 - 10) ERK2
 - 11) FAK
 - 12) FER
 - 13) GSK3 β
 - 14) JNK1
 - 15) JNK2
 - 16) PIP4K2A
 - 17) PIP4K2B
 - 18) PIP4K2C
 - 19) PRKD2
 - 20) PRKD3
 - 21) RSK1
 - 22) TNK1
 - 23) AXL
 - 24) CDK11B
 - 25) CDK17
 - 26) IRAK1
 - 27) PIK3C3
 - 28) PIK3CD
 - 29) PIK3R4
 - 30) MSK2
 - 31) ATM
 - 32) TBK1
 - 33) TTK
 - 34) PYK2
 - 35) TAOK3
 4. induces apoptosis via activating **CDK2**(237)
 5. inhibits metastasis via downregulating the **c-Jun/COX-2** pathway(238)
 6. induces apoptosis via the **PP5/AMPK** axis(239)
 7. **inhibits other kinases**(240):
 - 1) CAMK2D (CaMKII δ)
 - 2) TTK
 - 3) CLK1
 - 4) CLK2

Ribociclib

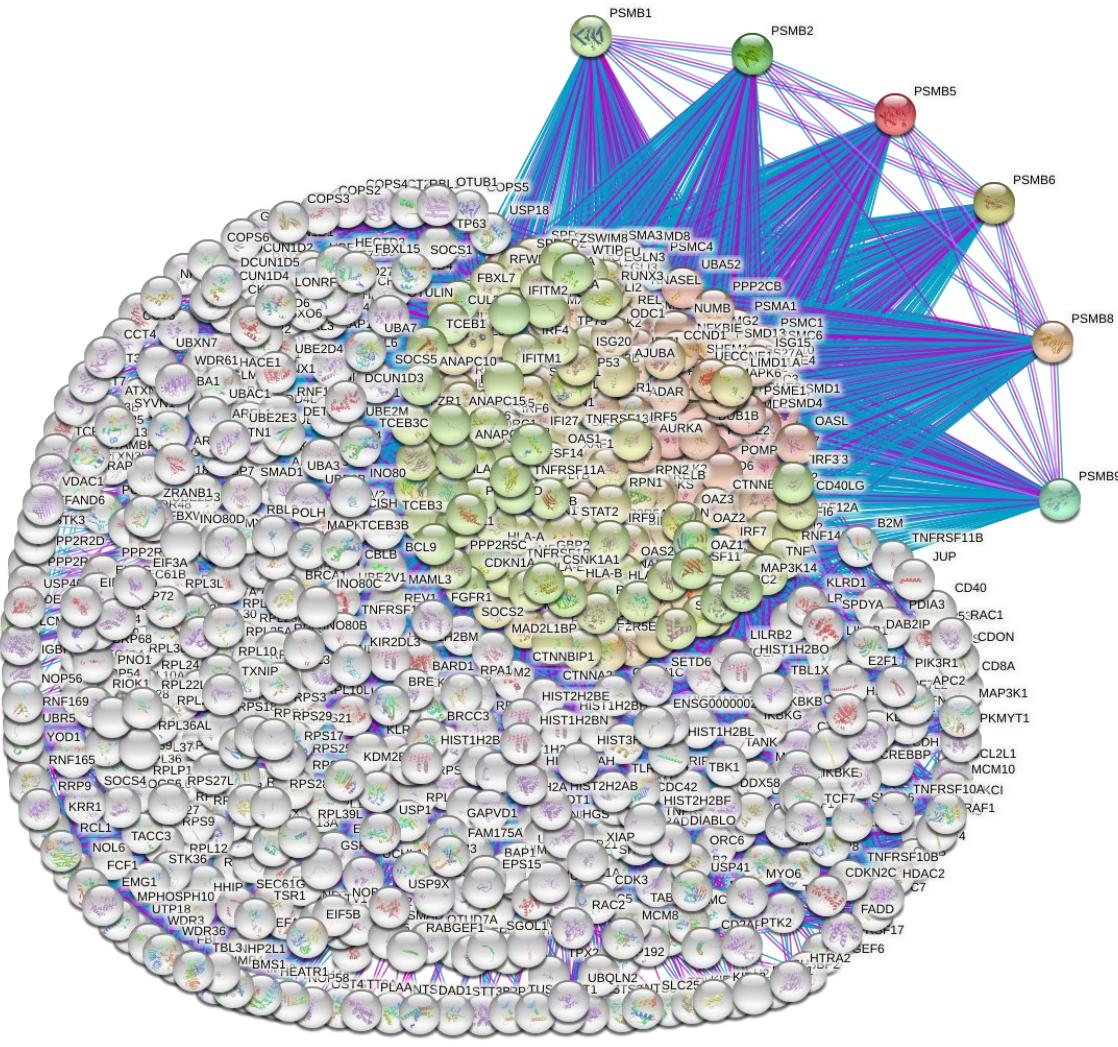
1. induces senescence through downregulation of the transcriptional expression of **MYBL2**(241)
2. **inhibits other kinases**(240):
 - 1) CDK9/cyclin T1

- 2) CAMK2D (CaMKII δ)
 3) TTK
 4) CAMK2A (CaMKII α)
3. inhibits other kinases(236):
- 1) CAMK2D
 - 2) CAMK2G
 - 3) CDK9
 - 4) GAK
 - 5) AURKA
 - 6) PRKD2
 - 7) FER
 - 8) TNK1
 - 9) FAK
- Abemaciclib**
1. upregulates antigen processing and presentation on tumor cells(242, 243)
 2. elevates secreted proinflammatory cytokines(243)
3. markedly upregulates the canonical interferon signaling pathway(243)
 4. activates T cells(242)
 5. inhibits other kinases(240):
- 1) CDK9/cyclin T1
 - 2) CDK9/cyclin K
 - 3) CDK1/cyclin B
 - 4) CDK2/cyclin A1
 - 5) CDK2/cyclin E1
 - 6) GSK3B (GSK3 β)
 - 7) CAMK2D (CaMKII δ)
 - 8) TTK
 - 9) PRKCG (PKC γ)
 - 10) GSK3A (GSK3 α)
 - 11) PRKCA (PKC α)
 - 12) PRKCB1 (PKC β 1)
 - 13) DYRK1A AAK1
 - 14) CAMK2G (CaMKII γ)

Drugs	Discovery Target	Uniprot ID	“Off-Target” Therapeutic Mechanisms
BORTEZOMIB		PSMB1 PSMB2 PSMB5 PSMB6 PSMB8 PSMB9	P20618 P49721 P28074 P28072 P28062 P28065
CARFILZOMIB	Proteasome		69 2
IXAZOMIB			

Note: Here, because of the central role of proteasome in various downstream processes, to identify the downstream effects of inhibiting proteasome, I also took benefit from the results of the study “Genome-wide siRNA

screen for modulators of cell death induced by proteasome inhibitor bortezomib(244),” in addition to using the interactors reported by STRING.



<https://version-11-0b.string-db.org/cgi/network?networkId=bGugYavdmmDI>

Bortezomib

1. primes hepatoma cells for natural killer cell antitumor reactivity(245)
2. increases cellular ceramide production to promote cell apoptosis(246)
3. downregulates Stat3 activity(247)
4. in addition to the abolishment of the pro-survival NF- κ B, directly induces apoptosis by activating proapoptotic endoplasmic reticulum stress-reactive oxygen species signaling cascades(248)
5. induces apoptosis and a decrease of both IL-6/IL-10 secretion and STAT3 phosphorylation(249)
6. disrupts tumor-dendritic cell interactions(250, 251)
7. stabilizes mitotic cyclins and prevents cell cycle progression via inhibition of UBE2C(252)
8. Microarray chips revealed multiple signaling pathways targeted by bortezomib, including endoplasmic reticulum, Wnt- β ,

- and calcium-mediated pathway, with possible targets including UBD, CUL3, HDAC6, and GADD45A(253)
9. sensitizes tumor cells to natural killer cells via endoplasmic reticulum-stress induced reduction of cell surface HLA-E which is exclusively controlled by the inhibitory receptor NKG2A(254, 255)
 10. can render tumor cells immunogenic by upregulating the cell surface expression of heat shock protein 60 and heat shock protein 90(256)
 11. improves the functions of dendritic cells and consequently exerts potent immune-mediated antitumor effects(256)
 12. exerts numerous effects on the immune system(257, 258)
 13. induces miRNAs that direct epigenetic silencing of locus genes and trigger apoptosis in leukemia, like miRNA/Ago2/YY1/PcG(259)
 14. downregulates telomerase and disrupts telomere homeostasis and function(260, 261)
 15. Dysregulation of unfolded protein response partially underlies its proapoptotic activity(262)
 16. induces a quick increase in reactive oxygen species production and subsequently decreases the glutathione levels(263)
 17. induces upregulation of the cell cycle inhibitor p21(WAF1) and the proapoptotic protein Noxa as well as cleavage of the anti-apoptotic protein Mcl-1(264-270)
 18. It induces proteasome-independent degradation of the TRAF6 protein, but not mRNA. The reduction in TRAF6 protein coincides with bortezomib-induced autophagy, and subsequently with apoptosis(271).
 19. inhibits the expression of SALL4 gene as well as C-mye and CCND1 downstream in the Wnt/β-catenin signaling pathway(272)
 20. inhibits steroid receptor coactivator-3 degradation which leads to activated Akt(273)
 21. Its apoptosis induction is dependent on caspase-2 activation, which is associated with ER stress and required for release of cytochrome c, breakdown of mitochondrial transmembrane potential, and its downstream caspase-9 activation(274).
 22. abrogates IL-6 triggered signaling cascades via caspase-dependent downregulation of gp130(275)
 23. induces G2-M arrest through reactive oxygen species-inducible phosphorylation of ATM-CHK1(276)
 24. might inhibit cells migration and invasion function by downregulation of Tyr397 expression(277)
 25. decreases ErbB family expression through lysosomal degradation pathway in a heat shock protein 90 (HSP90)-dependent manner(278)
 26. induces growth arrest and apoptosis by blockade of the androgen receptor signaling pathway and downregulating the prostate-specific antigen (PSA)(279, 280)
 27. sensitizes tumor cells to natural killer cell recognition by induction of NKG2D ligands(281)
 28. induce FLT3-ITD degradation through autophagy(282)

29. inhibits angiogenesis(283-286)
30. relieves tumor immune tolerance via STAT1 suppression and indoleamine 2,3-dioxygenase downregulation(287)
31. induces the expression of gene products associated with the endoplasmic reticulum secretory pathway like(288):
- 1) BCL2-associated athanogene 3 (BAG3)
 - 2) calreticulin, heat shock 70 kDa protein 1A (HSPA1A)
 - 3) VAMP-associated protein of 33 kDa mRNA
 - 4) heat shock 70 kDa protein 1B (HSPA1B)
 - 5) ATP-binding cassette
 - 6) subfamily F (GCN20)
 - 7) member 2 (ABCF2)
 - 8) EST pseudogene similar to UBL1 [ubiquitin-like 1 (sentrin)]
 - 9) Sec23-interacting protein p125
32. induces apoptosis via Bim and Bik upregulation(289, 290)
33. promotes the apoptosis potentially by downregulating the expression of HSP27(291)
34. It both inhibits VEGF-triggered caveolin-1 phosphorylation and markedly decreased caveolin-1 expression. Consequently, it inhibits VEGF-induced tumor cell migration. It also decreases VEGF secretion in the bone marrow microenvironment and inhibits VEGF-triggered tyrosine phosphorylation of caveolin-1, migration, and survival in human umbilical vascular endothelial cells(292).
35. It significantly affects the expression of these targets independent of p-53(293):
- 1) Heat shock cognate 71 kDa protein (HSPA8)
 - 2) Heat shock protein HSP 90 alpha (HSP90AA1)
 - 3) Heat shock 70 kDa protein 1B (HSPA1B)
 - 4) Importin subunit alpha-2 (Kpna2)
 - 5) Hist1h3b
 - 6) Psmc3
 - 7) Psmd14
 - 8) Sqstm1
 - 9) Gnas
 - 10) Rps10
 - 11) Psma5
 - 12) PPlase B
 - 13) Psmb1
 - 14) ATP5F1
 - 15) RPS15a
 - 16) Psmb2
 - 17) Hadha
 - 18) Myo1c
 - 19) Hsd17b10
36. induces apoptosis through microRNA-17-5p by targeting p21(294)
37. reduces the RANKL expression, inhibits cell proliferation and activates caspase-3 activity to induce cell apoptosis(295)
38. induces apoptosis via inhibition of cancerous inhibitor of protein phosphatase

- 2A (CIP2A)-mediated PP2A dependent Akt activation, independent of proteasome inhibition(296-299)
39. triggers apoptosis by enhancing the caspase 3 activation(300)
40. reduces tumorigenicity via downregulation of upregulated targets in side population cells(301)
41. induces apoptosis via activation of the p38 mitogen-activated protein kinase pathway(302)
42. stabilizes NOXA and triggers reactive oxygen species-associated apoptosis(303, 304)
43. inhibits prosurvival autophagy, in addition to its known function in blocking the proteasome(305)
44. induces apoptosis via activation of sterile20-like kinase 1 (MST1)(306)
45. induces an inhibitory chromatin environment at a distal enhancer of the estrogen receptor- α gene(307)
46. inhibits cellular growth of vascular endothelial cells through suppression of G2/M transition(308)
47. inhibited the progression of tumor cells via upregulation of the expression of miR-198(309)
48. sensitizes tumor cells to induction of apoptosis by type I interferons through NOXA expression and Mcl-1 cleavage(310)
49. augments lymphocyte stimulatory cytokine signaling in the tumor microenvironment to sustain CD8+T cell antitumor function(311)
50. induces lysosomal cathepsin B release and a caspase-2-dependent mitochondrial permeabilization and apoptosis(312)

Carfilzomib

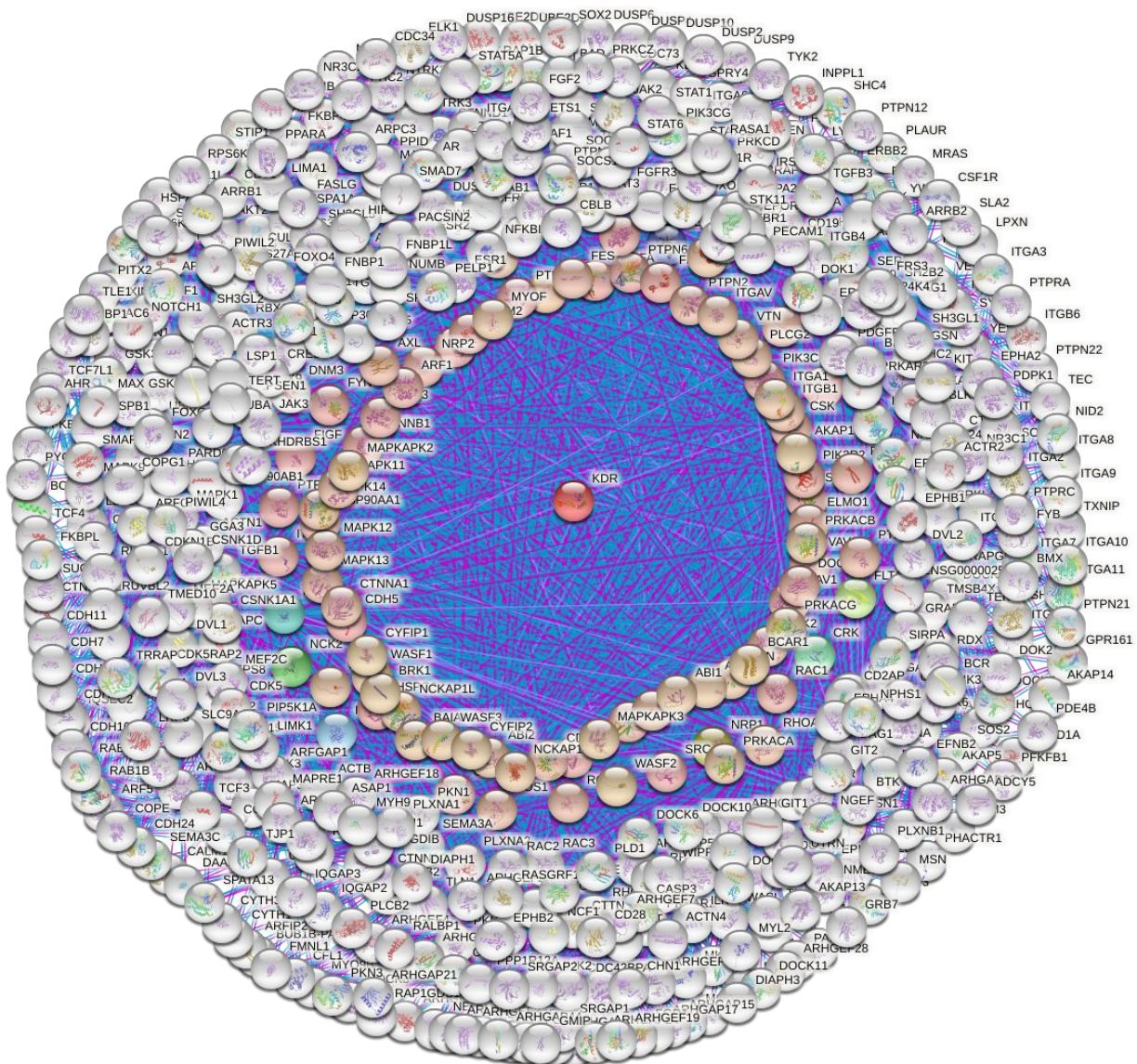
1. induces apoptosis via inhibiting ELK1/KIAA1524 (Elk-1/CIP2A) and activating PP2A not related to proteasome inhibition(313)
2. anti-tumor activity via the parathyroid hormone receptor pathway(314)

Drugs	Discovery Target		
NEVIRAPINE	EFAVIRENZ: 2		
ETRAVIRINE	RILPIVIRINE	DORAVIRINE	
			HIV Reverse Transcriptase

Efavirenz

1. inhibits the late stage of the HIV-1 replication and enhances the intracellular processing of Gag and Gag-Pol polyproteins. This is associated with a decrease in viral particle production from HIV-1-transfected cells(315, 316).
2. enhances Gag-Pol precursor dimerization after plasma membrane targeting but before complete particle assembly, resulting in an aberrant distribution of Gag/Gag-Pol processing products(317).

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
NINTEDANIB	VEGFR-2	KDR	P35968 229



<https://version-11-0b.string-db.org/cgi/network?networkId=b9X6KFnZSJWp>

*As nintedanib was approved in approximately the same time for both idiopathic pulmonary fibrosis (US – 2014) and non-small-

cell lung cancer (European Union – 2014), its “off-target” mechanisms in both these disorders were sought.

1. exhibits an anti-fibrosis effect via directly blocking Src and inhibiting the expression of genes downstream of Wnt signaling such as Cyclin D1, Wisp1, and S100a4(318)
2. inhibits several kinases in nanomolar range that are related to antitumor effects (IC_{50} (nmol/l))(319):
 - 1) ABL1(12 ± 5)
 - 2) BLK(42 ± 9)
 - 3) BTK(34 ± 14)
 - 4) CSF1R(5 ± 2)
 - 5) DDR1(17 ± 7)
 - 6) DDR2(16 ± 4)
 - 7) FYN(74 ± 24)
 - 8) JAK3(67 ± 27)
 - 9) KIT(6 ± 3)
 - 10) MAP3K3 (MEKK3)(58 ± 25)
 - 11) MAP3K7(46 ± 31)
 - 12) MELK(3 ± 2)
 - 13) MST4(84 ± 9)
 - 14) NTRK1 (TRKA)(30 ± 8)
 - 15) NTRK3 (TRKC)(48 ± 25)
 - 16) NUAK1(50 ± 8)
 - 17) RET(2 ± 1)
 - 18) SIK2(11 ± 4)
 - 19) STK24(61 ± 12)
 - 20) TGFBR1(77 ± 20)
 - 21) YES1(14 ± 4)
3. induces TNBC apoptosis by acting as a SHP-1 agonist(320)
4. directly inhibits tumor cell growth and induces tumor shrinkage in addition to its antiangiogenic effect on the tumor stroma(319)
5. inhibits mast cell survival and activation and thus provides a novel additional mechanism by which this drug may exert anti-fibrotic effects(321)
6. independent of TGF-β signaling, down-regulates protein and mRNA expression of extracellular matrix proteins, fibronectin, and collagen 1a1 and induces non-canonical autophagy(322)
7. reduces the expression of collagen I and V and inhibits collagen I fibril formation and causes a reduction in and an altered appearance of collagen fibril bundles(323)
8. Its derivate, which was used for binding-profiling of nintedanib itself, directly binds these kinases (K_d , nM) (NP: non-phosphorylated, P: phosphorylated)(324):
 - 1) ZAP70: 3500
 - 2) YSK4: 5.2
 - 3) YES: 80
 - 4) WEE1: 2900
 - 5) TSSK1B: 6100
 - 6) TTK: 22
 - 7) TXK: 860
 - 8) TYK2(JH1domain-catalytic): 37
 - ✓ TYK2(JH2domain-pseudokinase): 500
 - 9) TYRO3: 4900
 - 10) ULK1: 340

- 11) ULK2: 380
- 12) ULK3: 60
- 13) VEGFR2: 2.9
- 14) TRKA: 4.5
- 15) TRKB: 19
- 16) TRKC: 32
- 17) TNIK: 53
- 18) TNK1: 210
- 19) TNK2: 1600
- 20) TIE1: 2200
- 21) TIE2: 1400
- 22) TGFBR1: 4200
- 23) TBK1: 150
- 24) TAK1: 4.1
- 25) STK39: 520
- 26) SRPK1: 37
- 27) SRPK2: 36
- 28) SRPK3: 12
- 29) STK16: 18
- 30) STK33: 1100
- 31) STK35: 810
- 32) SRC: 580
- 33) SNARK: 21
- 34) S6K1: 190
- 35) SBK1: 110
- 36) SgK110: 2000
- 37) SGK3: 1000
- 38) SIK: 670
- 39) SIK2: 280
- 40) SLK: 51
- 41) RSK4(Kin.Dom.1-N-terminal): 230
- 42) RSK2(Kin.Dom.1-N-terminal): 65
- 43) RSK3(Kin.Dom.1-N-terminal): 58
- 44) RSK1(Kin.Dom.1-N-terminal): 180
- 45) RPS6KA5(Kin.Dom.1-N-terminal): 200
- 46) RPS6KA4(Kin.Dom.1-N-terminal): 620
- 47) ROS1: 3700
- 48) RIPK4: 440
- 49) RIOK3: 36
- 50) RIPK1: 240
- 51) RET: 31
 - ✓ RET(M918T): 30
 - ✓ RET(V804L): 9.8
 - ✓ RET(V804M): 9.6
- 52) RIOK1: 23
- 53) PRP4: 8.4
- 54) PYK2: 81
- 55) PRKCQ: 5500
- 56) PRKD1: 990
- 57) PRKD2: 720
- 58) PRKD3: 640
- 59) PRKG1: 14
- 60) PLK4: 53
- 61) PKN1: 740
- 62) PKN2: 1400
- 63) PKNB(*M. tuberculosis*): 3.6

- 64) PIP5K2B: 17
 65) PIP5K1A: 49
 66) PFTK1: 470
 67) PHKG1: 140
 68) PHKG2: 260
 69) PAK7: 1300
 70) PCTK1: 21
 71) PCTK2: 93
 72) PCTK3: 510
 73) PDGFRA: 16
 74) PDGFRB: 15
 75) PDPK1: 2800
 76) PFCDPK1(*P. falciparum*): 1200
 77) PAK2: 3300
 78) PAK3: 210
 79) PAK4: 3000
 80) OSR1: 1300
 81) MYO3B: 2700
 82) MUSK: 590
 83) MYLK: 290
 84) MYLK2: 4000
 85) MYLK4: 440
 86) MST2: 38
 87) MST3: 110
 88) MST4: 49
 89) MST1: 9
 90) MLCK: 110
 91) MLK1: 220
 92) MLK2: 2300
 93) MLK3: 700
 94) MELK: 4.9
 95) MERTK: 8.5
 96) MET: 200
 ✓ MET(M1250T): 340
 ✓ MET(Y1235D): 460
 97) MINK: 82
 98) MARK1: 1400
 99) MARK2: 1400
 100) MARK3: 770
 101) MARK4: 2000
 102) MAST1: 4600
 103) MEK1: 10
 104) MEK2: 42
 105) MEK3: 2100
 106) MEK4: 3700
 107) MEK5: 1.8
 108) MAP4K2: 290
 109) MAP4K3: 290
 110) MAP4K4: 150
 111) MAP4K5: 390
 112) MAP3K15: 1400
 113) MAP3K2: 9.4
 114) MAP3K3: 34
 115) LKB1: 18
 116) LOK: 87
 117) LRRK2: 46

- ✓ LRRK2(G2019S): 37
- 118) LTK: 150
- 119) LYN: 940
- 120) LZK: 140
- 121) JNK3: 270
- 122) KIT: 5.7
 - ✓ KIT(A829P): 87
 - ✓ KIT(D816H): 310
 - ✓ KIT(D816V): 47
 - ✓ KIT(L576P): 2.7
 - ✓ KIT(V559D): 6.3
 - ✓ KIT(V559D,T670I): 2.9
 - ✓ KIT(V559D,V654A): 29
- 123) LATS1: 420
- 124) LATS2: 380
- 125) LCK: 6.2
- 126) IKK-epsilon: 170
- 127) INSR: 24
- 128) INSRR: 21
- 129) IRAK1: 120
- 130) IRAK3: 5300
- 131) IRAK4: 810
- 132) ITK: 210
- 133) JNK1: 630
 - ✓ JAK1(JH1domain-catalytic): 2500
 - ✓ JAK1(JH2domain-pseudokinase): 4.8
- 134) JAK2(JH1domain-catalytic): 14
- 135) JAK3(JH1domain-catalytic): 8.2
- 136) ICK: 1700
- 137) IGF1R: 62
- 138) IKK-alpha: 4500
- 139) GCN2(Kin.Dom.2,S808G): 2000
- 140) GRK1: 100
- 141) GRK4: 17
- 142) GRK7: 1200
- 143) GSK3A: 430
- 144) GSK3B: 83
- 145) HCK: 5300
- 146) HIPK1: 1300
- 147) HIPK2: 790
- 148) HIPK3: 850
- 149) HIPK4: 1200
- 150) HPK1: 35
- 151) ERN1: 2000
- 152) FAK: 210
- 153) FER: 73
- 154) FES: 1200
- 155) FGFR1: 92
- 156) FGFR2: 350
- 157) FGFR3: 93
 - ✓ FGFR3(G697C): 140
- 158) FGFR4: 1800
- 159) FGR: 300
- 160) FLT1: 63
- 161) FLT3: 3.8
 - ✓ FLT3(D835H): 0.71

✓	FLT3(D835Y): 0.42	185)	CSF1R: 48
✓	FLT3(ITD): 0.7	186)	CLK1: 8.8
✓	FLT3(K663Q): 4.5	187)	CLK2: 760
✓	FLT3(N841I): 2.6	188)	CHEK1: 290
✓	FLT3(R834Q): 17	189)	CDKL2: 1300
162)	FLT4: 95	190)	CDK7: 300
163)	FRK: 2700	191)	CDK4-cyclinD1: 160
164)	FYN: 630	192)	CDK4-cyclinD3: 720
165)	ERK5: 2500	193)	CDK2: 1400
166)	EPHB4: 1100	194)	CAMK4: 3700
167)	EPHB6: 140	195)	CAMKK1: 630
168)	EPHB1: 550	196)	CAMKK2: 920
169)	EPHA6: 940	197)	BTK: 310
170)	DYRK1B: 1600	198)	CAMK1: 5900
171)	DRAK1: 110	199)	CAMK1D: 2200
172)	DRAK2: 670	200)	CAMK1G: 260
173)	DCAMKL3: 770	201)	CAMK2A: 3600
174)	DDR1: 12	202)	BMPR2: 56
175)	DDR2: 42	203)	AURKB: 420
176)	DLK: 140	204)	AURKC: 190
177)	DAPK2: 3200	205)	AXL: 12
178)	DAPK3: 2100	206)	BIKE: 2.2
179)	DCAMKL1: 540	207)	BLK: 380
180)	CSNK2A1: 7500	208)	ARK5: 160
181)	CSNK2A2: 890	209)	ALK: 31
182)	CSNK1D: 1300	210)	AMPK-alpha1: 87
183)	CSNK1E: 230	211)	AMPK-alpha2: 84
184)	CLK4: 18	212)	ACVRL1: 4200

213) AAK1: 63

214) ABL1(E255K)-P: 63

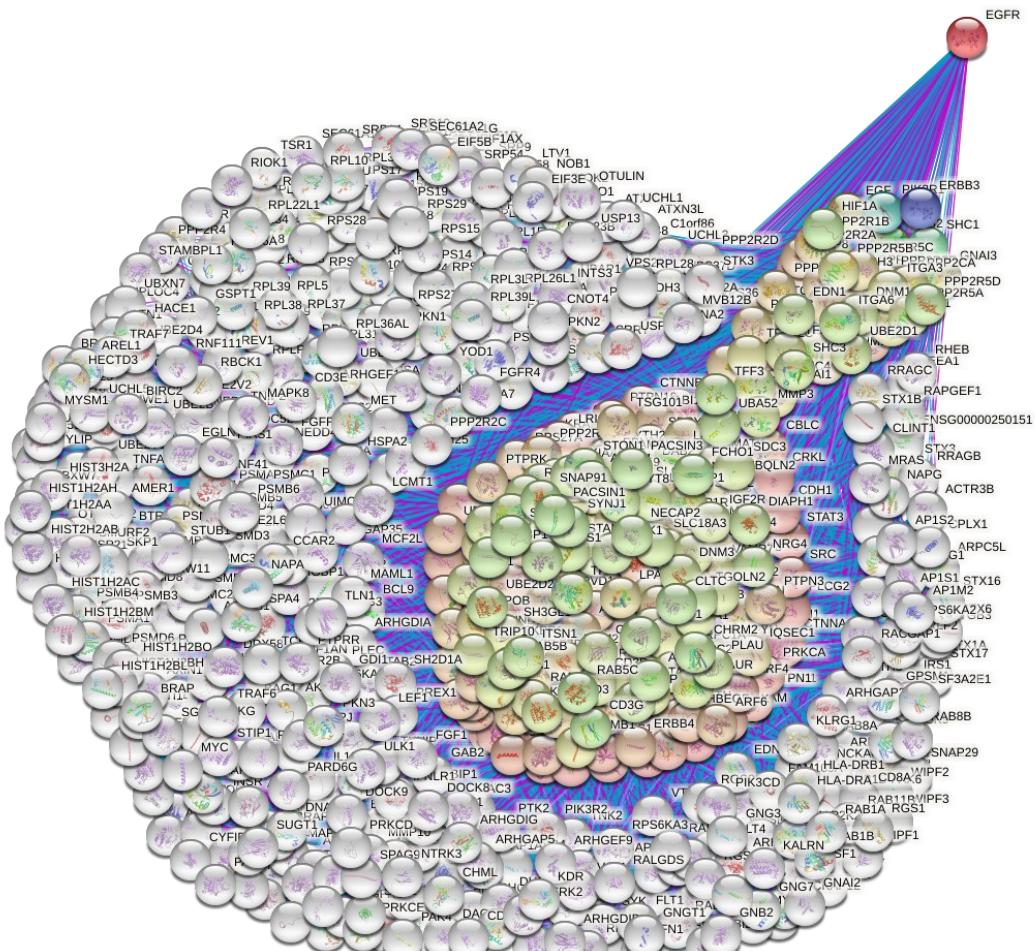
- ✓ ABL1(F317I)-NP: 6900
- ✓ ABL1(F317I)-P: 2600
- ✓ ABL1(F317L)-NP: 2300
- ✓ ABL1(F317L)-P: 640
- ✓ ABL1(H396P)-NP: 45
- ✓ ABL1(H396P)-P: 49
- ✓ ABL1(M351T)-P: 52
- ✓ ABL1(Q252H)-NP: 73
- ✓ ABL1(Q252H)-P: 28
- ✓ ABL1(T315I)-NP: 66
- ✓ ABL1(T315I)-P: 10
- ✓ ABL1(Y253F)-P: 37
- ✓ ABL1-NP: 230
- ✓ ABL1-P: 64

215) ABL2: 2000

216) ACVR1: 600

217) ACVR1B: 4900

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
GEFITINIB ERLOTINIB			111 109
LAPATINIB VANDETANIB			16 114
AFATINIB DACOMITINIB	epidermal growth factor receptor	EGFR	P00533 41
OSIMERTINIB NERATINIB			8 4
AVAPRITINIB TUCATINIB			



<https://version-11-0b.string-db.org/cgi/network?networkId=bpVYSKoHpYIY>

Gefitinib

1. induces apoptosis and antitumor effects via epidermal growth factor receptor (EGFR)-independent mechanisms as it also has activity in tumors lacking EGFR(325, 326)
2. induces apoptosis by inducing cytoplasmic translocation of the CDK inhibitor p27 and its binding to a cleaved intermediate of caspase 8(327)
3. induces apoptosis through activation of Bax(328)
4. directly inhibits other protein kinases(IC_{50} (μ mol/L))(329-331):
 - 1) Aurora A(60 ± 7)
 - 2) Aurora B(11.1 ± 2.9)
 - 3) BLK(3.1 ± 0.4)
 - 4) BRK(0.82 ± 0.06)
 - 5) CaMKII(13.5 ± 1.4)
 - 6) CK1 δ (61 ± 7)
 - 7) CSK(41 ± 17)
 - 8) EphB4(1.22 ± 0.49)
 - 9) HCK(2.09 ± 0.35)
 - 10) GAK(0.090 ± 0.018)
 - 11) LYN(0.95 ± 0.46)
 - 12) MET(3.2 ± 1.1)
 - 13) p38 α (1.19 ± 0.03)
 - 14) RICK(0.049 ± 0.001)
 - 15) YES(1.75 ± 0.13)
 - 16) BTK(6.27)
5. induces apoptosis in human glioma cells by targeting Bad phosphorylation(332)
6. reduces the proliferation of glioma cells, presumably by autophagic mechanisms involving AMPK activation(333)
7. induces apoptosis probably via upregulation of Fas(334)
8. induces apoptosis through a p53-dependent upregulation of proapoptotic molecules and downregulation of antiapoptotic molecules(335)
9. increases the expression of both p27KIP1 and p21CIP1/WAF1 cyclin-dependent kinase (CDK) inhibitors(336)
10. Inhibition of angiogenesis by gefitinib seems to be independent of the EGFR genetic status of the tumors(337).
11. enhances natural killer cell cytotoxicity to lung cancer cells via upregulating the expression of NKG2D ligands ULBP1, ULBP2 or MICA on tumor cells and NKG2D on natural killer cells(338), also see (339)
12. inhibits retina angiogenesis by affecting VEGF signaling pathway(340)
13. Upregulation of YY1 and E-cadherin may account for the efficacy of gefitinib in bladder cancer(341).
14. apoptosis via translocating p53 from cytosol to nucleus and upregulating Fas(342)
15. inhibits EGF-independent angiogenesis, by acting through Fes as an inhibitor of fibroblast growth factor-2 (FGF-2)-driven angiogenesis, but not toward vascular endothelial growth factor-A (VEGF-A)(343)
16. cytotoxicity via phosphorylation of the eukaryotic initiation factor 2 alpha independent of EGFR(344)

17. may have an additional beneficiary effect on tumor cell proliferation and migration via **decreasing the synthesis of hyaluronic acid** by mesothelioma cells(345)
18. targets the transcription factor **FOXO3a** to mediate cell cycle arrest and cell death(346, 347)
19. directly binds these kinases (K_d (μM))(348):
- 1) CSNK1E: 2
 - 2) EGFR: 0.0018
 - 3) EPHA6: 1.4
 - 4) EPHB1: 7.3
 - 5) ERBB2: 1.1
 - 6) GAK: 0.007
 - 7) JNK2: 1.4
 - 8) JNK3: 2.3
 - 9) LCK: 1.1
 - 10) MKNK2: 0.36
 - 11) PHKG2: 6.4
 - 12) RIPK2: 0.8
 - 13) SLK: 1.1
 - 14) SRC: 5
 - 15) STK10: 0.87
 - 16) STK17A: 2.9
 - 17) STK17B: 6.5
 - 18) ULK3_m: 1.4
20. exerts **antimetastatic** effects and reduces cell adhesion via downregulation of integrin $\alpha 3$, αv , $\beta 1$, $\beta 4$, $\beta 5$, $\beta 6$ and FAK phosphorylation independent of EGFR blockade(349)
21. modulates stress fibers and tubular-like structure formation and attenuates angiogenesis(350)
22. represses **FOXM1** expression via **FOXO3a** activation and is mediated at the transcriptional level and gene promoter level(347)
23. induces apoptosis via downregulation of Bcl-2 and caspase-3 activation(351)
24. demonstrates amphiregulin-dependant activity, even in the absence of sensitizing EGFR mutations(352)
25. antitumor activity via downregulating the expression of cyclin-dependent kinase 2 (**CDK2**), **CDK4**, **CDK6** and the antiapoptotic protein Bcl-2, upregulation of p27Kip1 and upregulation and activation of apoptosis related protein Bax(353)
26. induces cytostasis and primes the extrinsic (Fas) and intrinsic (mitochondrial and endoplasmic reticulum) apoptotic pathways(354)
27. accelerates Fas-mediated apoptosis by enhancing caspase-8 activation in cancer cells independent of EGFR(355)
28. reduces the expression of metastasis-related proteins(356):
- 1) basic fibroblast growth factor (**bFGF**)
 - 2) matrix metalloproteinases-2 (**MMP-2**)
 - 3) matrix metalloproteinases-9 (**MMP-9**)
29. causes cell arrest via inhibition of cyclin G-associated kinase and induction of **miR-630**(357)
30. antimetastatic activity via decreasing the synthesis of matrix metalloproteinase and adhesion of cancer cells to extracellular matrix proteins(358)

31. upregulates p27KIP1 and induces G1 arrest(359)
32. The antiproliferative effects of gefitinib may be, at least in part, due to the inhibition of E2F-1 expression and telomerase activity(360).
33. downregulates the expression of histone deacetylase(361)
34. downregulates the expression of VEGF(361, 362)
35. modulates cell growth and differentiation of acute myeloid leukemia cells via histamine receptors(363)
36. upregulates death receptor 5 expression to mediate apoptosis(364)
37. induces apoptosis through downregulating tumor necrosis factor-related apoptosis-inducing ligand expression levels(365)
38. inhibits cell growth via elevation of p21 levels (through inducing protein stability) and suppression of cdk2/4 and cyclinE/D1 activities(366)
39. directly binds these kinases (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):
- 1) ABL1-NP: 2200
 - ✓ ABL1-P: 480
 - ✓ ABL1(E255K)-P: 400
 - ✓ ABL1(F317I)-P: 4700
 - ✓ ABL1(F317L)-NP: 2700
 - ✓ ABL1(F317L)-P: 780
 - ✓ ABL1(H396P)-NP: 680
 - ✓ ABL1(H396P)-P: 460
 - ✓ ABL1(M351T)-P: 520
 - ✓ ABL1(Q252H)-NP: 1100
 - ✓ ABL1(Q252H)-P: 230
 - ✓ ABL1(Y253F)-P: 360
 - 2) ABL2: 1600
 - 3) AXL: 1800
 - 4) BLK: 1200
 - 5) CDK7: 610
 - 6) CHEK2: 800
 - 7) CIT: 1300
 - 8) CSNK1D: 3200
 - 9) CSNK1E: 430
 - 10) DAPK3: 5700
 - 11) DCAMKL3: 2900
 - 12) DMPK: 6900
 - 13) DRAK1: 2000
 - 14) DRAK2: 3800
 - 15) EGFR: 1
 - ✓ EGFR(E746-A750del): 0.54
 - ✓ EGFR(G719C): 2
 - ✓ EGFR(G719S): 1.1
 - ✓ EGFR(L747-E749del, A750P): 0.57
 - ✓ EGFR(L747-S752del, P753S): 0.57
 - ✓ EGFR(L747-T751del,Sins): 0.52
 - ✓ EGFR(L858R): 0.94
 - ✓ EGFR(L858R,T790M): 140
 - ✓ EGFR(L861Q): 1.4
 - ✓ EGFR(S752-I759del): 0.98

- ✓ EGFR(T790M): 40
- 16) EPHA1: 4000
- 17) EPHA3: 5500
- 18) EPHA5: 1500
- 19) EPHA6: 590
- 20) EPHA8: 1800
- 21) EPHB1: 1300
- 22) EPHB4: 2500
- 23) EPHB6: 3200
- 24) ERBB2: 3500
- 25) ERBB3: 790
- 26) ERBB4: 410
- 27) ERK3: 1600
- 28) ERK4: 3100
- 29) FGR: 2600
- 30) FLT3: 3100
 - ✓ FLT3(D835H): 1100
 - ✓ FLT3(D835Y): 1000
 - ✓ FLT3(ITD): 2900
 - ✓ FLT3(K663Q): 5100
 - ✓ FLT3(N841I): 3000
 - ✓ FLT3(R834Q): 3500
- 31) FRK: 2000
- 32) GAK: 13
- 33) HCK: 4400
- 34) HIPK4: 310
- 35) IRAK1: 69
- 36) IRAK3: 1500
- 37) IRAK4: 540
- 38) JNK2: 1700
- 39) JNK3: 3200
- 40) KIT(A829P): 1800
 - ✓ KIT(D816H): 5500
 - ✓ KIT(D816V): 4300
- 41) LCK: 630
- 42) LOK: 470
- 43) LTK: 5500
- 44) LYN: 990
- 45) MAP3K2: 3300
- 46) MAP3K3: 2100
- 47) MEK5: 600
- 48) MET(Y1235D): 3500
- 49) MINK: 1800
- 50) MKNK1: 290
- 51) MKNK2: 1200
- 52) MYLK2: 1900
- 53) NLK: 4200
- 54) PHKG1: 3700
- 55) PHKG2: 2700
- 56) PIM3: 5800
- 57) PIP5K2C: 7500
- 58) PRKD1: 3500
- 59) RIPK2: 530
- 60) RPS6KA4(Kin.Dom.2-C-terminal): 1200
- 61) SBK1: 560
- 62) SIK2: 2100

- 63) SLK: 920
- 64) SNARK: 4600
- 65) SRC: 3800
- 66) STK36: 5700
- 67) TNIK: 6900
- 68) TXK: 6000
- 69) YSK4: 240

Erlotinib

1. induces apoptosis and antitumor effects via EGFR-independent mechanisms as it also has activity in tumors lacking EGFR(367-369)
2. prevents epithelial-to-mesenchymal transition by acting on E-cadherin expression and displacing the ternary complex formed by integrin-linked kinase (ILK), α -parvin, and PINCH (IPP)(370)
3. directly binds these kinases (K_d (μM))(348):
 - 1) AAK1: 4.4
 - 2) ABL1: 0.77
 - ✓ ABL1(E255K): 2.4
 - ✓ ABL1(H396P): 0.69
 - ✓ ABL1(M351T): 0.73
 - ✓ ABL1(Q252H): 0.28
 - ✓ ABL1(T315I): 0.6
 - ✓ ABL1(Y253F): 0.54
 - 3) ABL2: 0.3
 - 4) Aurora2: 3.8
 - 5) BIKE: 1.8
 - 6) EGFR: 0.0014
- 7) EPHA6: 0.93
- 8) ERBB2: 5.1
- 9) GAK: 0.04
- 10) JNK2: 4
- 11) LCK: 0.53
- 12) MKNK2: 1.6
- 13) RIPK2: 0.41
- 14) SLK: 0.11
- 15) SRC: 1.9
- 16) STK10: 0.083
- 17) ULK3_m: 0.63
4. sensitizes cancer cells to natural killer cell mediated cytotoxicity by increasing the levels of mRNA transcripts and surface protein of UL16-binding protein-1 (ULBP1) via inhibition of the PKC pathway(371)
5. activates mitochondrial death pathways related to the production of reactive oxygen species through inducing loss of mitochondrial membrane potential, the release of cytochrome c and apoptosis-inducing factor (AIF) and activation of JNK(372, 373)
6. exhibits antineoplastic off-target effects via JAK2 and nucleocytoplasmic translocation of nucleophosmin-1 (NPM-1) and p14ARF(368)
7. targets FLT3(374)
8. immunomodulation via increasing T cell mediated cytotoxicity on lung cancer through inhibiting the expression of PD-L1 and genes related to antigen presentation and inflammation(375)

9. decreases Rad51 protein levels by enhancing **Rad51** mRNA and protein instability(376)
10. directly binds to six protein kinases with even higher affinity (K_d : ranging from 0.09 to 0.358 μM) compared to EGFR (K_d : 0.434 μM)(330):
- 1) STK10
 - 2) **MAP3K1**
 - 3) **ILK**
 - 4) SLK
 - 5) Ripk2
 - 6) ABL2
11. prevents bone **metastases** by affecting host microenvironments irrespective of its direct effect on tumor cells and in cancers with no EGFR expression(377)
12. inhibits JAK2V617F activity, a mutant of tyrosine kinase JAK2(378)
13. directly binds(331):
- 1) Lyn ($\text{IC}_{50} = 1.46\mu\text{m}$)
 - 2) Yes ($\text{IC}_{50} = 3.99\mu\text{m}$)
 - 3) **Btk** ($\text{IC}_{50} = 5.62\mu\text{m}$)
14. induces **p27KIP1** upregulation and nuclear translocation in association with cell growth inhibition and G1/S phase arrest(379)
15. exerts antitumor effects via **CIP2A**(380)
16. increases the susceptibility of cancer cells to immune-cell-mediated cytotoxicity through **PI3** and independent of EGFR(381, 382)
17. decrease **VEGF** expression also by hypoxia-inducible factor (HIF)-1-independent mechanisms(383)
18. directly inhibits HER2 kinase activation and downstream signaling(384)
19. induces apoptosis through inducing via p73 the transcription of **PUMA**(385)
20. triggers excessive mitochondrial fragmentation which promotes apoptosis via activating the mROS-HtrA2/Omi pathways(386)
21. suppresses the transcription of **miR-9-1** and the consequent downregulation of FoxO1(387)
22. directly binds these kinases (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):
- 1) AAK1: 1200
 - 2) ABL1-NP: 330
 - ✓ ABL1-P: 76
 - ✓ ABL1(E255K)-P: 63
 - ✓ ABL1(F317I)-NP: 8200
 - ✓ ABL1(F317I)-P: 1100
 - ✓ ABL1(F317L)-NP: 640
 - ✓ ABL1(F317L)-P: 150
 - ✓ ABL1(H396P)-NP: 89
 - ✓ ABL1(H396P)-P: 58
 - ✓ ABL1(M351T)-P: 66
 - ✓ ABL1(Q252H)-NP: 230
 - ✓ ABL1(Q252H)-P: 61
 - ✓ ABL1(T315I)-NP: 620
 - ✓ ABL1(T315I)-P: 59

- ✓ ABL1(Y253F)-P: 76
- 3) ABL2: 200
- 4) ADCK3: 1900
- 5) ADCK4: 2500
- 6) ALK: 1200
- 7) AURKA: 2200
- 8) AURKB: 1400
- 9) AURKC: 600
- 10) AXL: 4000
- 11) BIKE: 1200
- 12) BLK: 190
- 13) CAMK2D: 6400
- 14) CIT: 680
- 15) CSK: 9600
- 16) CSNK1D: 3500
- 17) CSNK1E: 500
- 18) DAPK3: 6000
- 19) DDR1: 790
- 20) DMPK: 2900
- 21) DMPK2: 3400
- 22) DRAK1: 6300
- 23) DRAK2: 9400
- 24) DYRK2: 1300
- 25) EGFR: 0.67
 - ✓ EGFR(E746-A750del): 0.48
 - ✓ EGFR(G719C): 0.85
 - ✓ EGFR(G719S): 0.52
 - ✓ EGFR(L747-E749del, A750P): 0.52
- ✓ EGFR(L747-S752del, P753S): 0.47
- ✓ EGFR(L747-T751del,Sins): 0.35
- ✓ EGFR(L858R): 0.97
- ✓ EGFR(L858R,T790M): 190
- ✓ EGFR(L861Q): 1.2
- ✓ EGFR(S752-I759del): 1.6
- ✓ EGFR(T790M): 140
- 26) EPHA3: 2400
- 27) EPHA5: 710
- 28) EPHA6: 440
- 29) EPHA7: 1400
- 30) EPHA8: 940
- 31) EPHB1: 1100
- 32) EPHB4: 2200
- 33) EPHB6: 1700
- 34) ERBB2: 2900
- 35) ERBB3: 1100
- 36) ERBB4: 230
- 37) ERK4: 2500
- 38) FGR: 1100
- 39) FLT1: 4400
- 40) FLT3: 1200
 - ✓ FLT3(D835H): 350
 - ✓ FLT3(D835Y): 130
 - ✓ FLT3(ITD): 820
 - ✓ FLT3(K663Q): 1300
 - ✓ FLT3(N841I): 500
- 41) FLT4: 2100

- 42) FRK: 2000
 43) GAK: 3.1
 44) GCN2(Kin.Dom.2,S808G): 4400
 45) GRK4: 7300
 46) HCK: 1800
 47) HIPK4: 960
 48) JAK2(JH1domain-catalytic): 3700
 49) JAK3(JH1domain-catalytic): 700
 50) JNK2: 2000
 51) JNK3: 4300
 52) KIT: 1700
- ✓ KIT(D816V): 1600
 - ✓ KIT(L576P): 3500
 - ✓ KIT(V559D): 3100
 - ✓ KIT(V559D,T670I): 1300
- 53) LCK: 250
 54) LOK: 19
 55) LRRK2: 2900
- ✓ LRRK2(G2019S): 4800
- 56) LTK: 890
 57) LYN: 530
 58) MAP3K2: 2500
 59) MAP3K3: 2300
 60) MEK5: 96
 61) MERTK: 980
 62) MET: 3800
- ✓ MET(M1250T): 2800
 - ✓ MET(Y1235D): 1100
- 63) MINK: 1300
 64) MKNK1: 690
 65) MKNK2: 1000
 66) MYLK2: 970
 67) PDGFRA: 1800
 68) PDGFRB: 1400
 69) PIP5K2C: 1000
 70) PLK4: 1900
 71) PRKR: 1300
 72) RET: 1300
- ✓ RET(M918T): 330
- 73) RIPK2: 680
 74) RIPK4: 4900
 75) RPS6KA4(Kin.Dom.2-C-terminal):
 7000
 76) SBK1: 1200
 77) SIK2: 2600
 78) SLK: 26
 79) SRC: 700
 80) SRPK3: 8300
 81) STK36: 4400
 82) TBK1: 3100
 83) TIE1: 850
 84) TNIK: 4500
 85) TNK1: 630
 86) TNNI3K: 570
 87) TTK: 3700
 88) TXK: 3800
 89) TYK2(JH2domain-pseudokinase): 2400

90) TYRO3: 3900

91) ULK3: 920

92) VEGFR2: 5700

93) YES: 2200

94) YSK4: 25

3) ERBB3: 5500

4) ERBB4: 54

5) LOK: 4400

6) MEK5: 1100

7) MKK7: 4400

8) PIK3C2B: 670

9) PIK3C2G: 7500

10) PIK4CB: 940

11) RIPK2: 3600

12) SLK: 3300

1. negatively regulates general translation and induces stress granules formation through the kinase PERK(388)

2. induces G1 arrest via upregulation of p27 by targeting c-jun via miR-1470(389)

3. promotes apoptosis through mROS-HtrA2/Omi pathways(390)

4. Activity of lapatinib is independent of EGFR expression level in HER2-overexpressing(391)

5. directly binds these kinases (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):

1) EGFR: 2.4

✓ EGFR(E746-A750del): 8.6

✓ EGFR(G719C): 0.92

✓ EGFR(G719S): 2.1

✓ EGFR(L747-E749del, A750P): 2.2

✓ EGFR(L747-S752del, P753S): 3.9

✓ EGFR(L747-T751del,Sins): 3.5

✓ EGFR(L858R): 2.8

✓ EGFR(L861Q): 1.2

✓ EGFR(S752-I759del): 4.2

✓ EGFR(T790M): 860

2) ERBB2: 7

6. induces p27 expression via both transcriptional and post-translational upregulations, leading to cell cycle arrest and cell proliferation inhibition(392)

7. directly binds these kinases (K_d (μM))(348):

1) EGFR: 0.0055

2) ERBB2: 0.011

3) SLK: 9.3

4) STK10: 2.6

Vandetanib

1. **Vandetanib, designed to inhibit VEGFR2 and EGFR signaling, had no clinical activity as monotherapy for recurrent ovarian cancer and no detectable modulation of VEGFR2. Proteomic analysis of paired biopsies detected both phosphorylated-EGFR and phosphorylated-VEGF receptor-2 in ovarian tumor tissue, but only phosphorylated-EGFR was measurably inhibited by vandetanib(463).

2. directly binds these kinases (K_d (μM))(348):
- 1) ABL1: 0.27
 - ✓ ABL1(E255K): 0.68
 - ✓ ABL1(H396P): 0.12
 - ✓ ABL1(M351T): 0.17
 - ✓ ABL1(Q252H): 0.18
 - ✓ ABL1(T315I): 0.12
 - ✓ ABL1(Y253F): 0.16
 - 2) ABL2: 0.13
 - 3) CSK: 3.7
 - 4) CSNK1E: 1.5
 - 5) EGFR: 0.017
 - 6) EPHA2: 1.9
 - 7) EPHA3: 1.8
 - 8) EPHA4: 4.3
 - 9) EPHA5: 0.31
 - 10) EPHA6: 0.065
 - 11) EPHA7: 3.4
 - 12) EPHA8: 0.16
 - 13) EPHB1: 0.54
 - 14) EPHB4: 2.6
 - 15) FGFR1: 5.3
 - 16) FGFR2: 5.5
 - 17) FGFR3: 0.24
 - 18) FGR: 2.2
 - 19) FLT3: 1.2
 - 20) FLT4: 0.3
 - 21) FRK: 0.48
 - 22) FYN: 5.7
 - 23) GAK: 0.33
 - 24) HCK: 4.2
 - 25) KIT: 1.1
 - 26) LCK: 0.061
 - 27) LYN: 0.71
 - 28) MAP3K4: 4.8
 - 29) MAP4K5: 0.51
 - 30) MKNK2: 3.4
 - 31) PDGFRB: 0.25
 - 32) PHKG1: 7.1
 - 33) PHKG2: 9.5
 - 34) PRKAA1: 10
 - 35) PTK6: 0.033
 - 36) RIPK2: 0.031
 - 37) SLK: 0.096
 - 38) SRC: 0.17
 - 39) STK10: 2.4
 - 40) STK18: 1.5
 - 41) TEK: 3.6
 - 42) TNIK: 3.9
 - 43) VEGFR2: 0.47
 - 44) YES: 0.28
3. inhibits cancer cell migration and invasion via downregulating the SDF-1/CXCR4 axis and matrix metalloproteinase 14(393), also see (394)
4. directly binds these kinases (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):

- 1) ABL1-NP: 48
 ✓ ABL1-P: 16
 ✓ ABL1(E255K)-P: 13
 ✓ ABL1(F317I)-NP: 770
 ✓ ABL1(F317I)-P: 170
 ✓ ABL1(F317L)-NP: 75
 ✓ ABL1(F317L)-P: 33
 ✓ ABL1(H396P)-NP: 9.8
 ✓ ABL1(H396P)-P: 14
 ✓ ABL1(M351T)-P: 15
 ✓ ABL1(Q252H)-NP: 18
 ✓ ABL1(Q252H)-P: 16
 ✓ ABL1(T315I)-NP: 150
 ✓ ABL1(T315I)-P: 20
 ✓ ABL1(Y253F)-P: 9
- 2) ABL2: 69
- 3) ACVR1: 150
- 4) ACVRL1: 470
- 5) ADCK3: 4500
- 6) ADCK4: 1700
- 7) ALK: 2100
- 8) AMPK-alpha1: 3000
- 9) AURKC: 1500
- 10) AXL: 250
- 11) BLK: 66
- 12) BMPR1B: 240
- 13) BRK: 160
- 14) BTK: 1700
- 15) CDK7: 4100
 16) CIT: 1800
 17) CSF1R: 1200
 18) CSK: 2500
 19) CSNK1E: 3000
 20) DDR1: 11
 21) DDR2: 320
 22) DMPK2: 2200
 23) EGFR: 9.5
 ✓ EGFR(E746-A750del): 4.8
 ✓ EGFR(G719C): 9.6
 ✓ EGFR(G719S): 5.9
 ✓ EGFR(L747-E749del, A750P): 12
 ✓ EGFR(L747-S752del, P753S): 7.9
 ✓ EGFR(L747-T751del,Sins): 8.9
 ✓ EGFR(L858R): 8.7
 ✓ EGFR(L858R,T790M): 230
 ✓ EGFR(L861Q): 11
 ✓ EGFR(S752-I759del): 12
 ✓ EGFR(T790M): 100
- 24) EPHA1: 230
 25) EPHA2: 1100
 26) EPHA3: 2000
 27) EPHA4: 1600
 28) EPHA5: 240
 29) EPHA6: 50
 30) EPHA7: 2400
 31) EPHA8: 91

- | | |
|----------------------|--------------------------|
| 32) EPHB1: 290 | 53) HUNK: 4100 |
| 33) EPHB2: 440 | 54) IRAK1: 1200 |
| 34) EPHB4: 520 | 55) IRAK4: 75 |
| 35) EPHB6: 76 | 56) KIT: 260 |
| 36) ERBB2: 2600 | ✓ KIT(A829P): 34 |
| 37) ERBB3: 160 | ✓ KIT(D816H): 420 |
| 38) ERBB4: 480 | ✓ KIT(D816V): 290 |
| 39) ERK3: 1500 | ✓ KIT(L576P): 140 |
| 40) FGFR1: 560 | ✓ KIT(V559D): 180 |
| 41) FGFR2: 1100 | ✓ KIT(V559D,T670I): 2000 |
| 42) FGFR3: 1600 | ✓ KIT(V559D,V654A): 560 |
| ✓ FGFR3(G697C): 6900 | |
| 43) FGFR4: 2300 | 57) LCK: 17 |
| 44) FGR: 270 | 58) LOK: 81 |
| 45) FLT1: 260 | 59) LTK: 550 |
| 46) FLT3: 850 | 60) LYN: 110 |
| ✓ FLT3(D835H): 560 | 61) MAP3K4: 3000 |
| ✓ FLT3(D835Y): 830 | 62) MAP4K2: 1600 |
| ✓ FLT3(ITD): 1800 | 63) MAP4K3: 1500 |
| ✓ FLT3(K663Q): 190 | 64) MAP4K4: 1400 |
| ✓ FLT3(N841I): 1200 | 65) MAP4K5: 450 |
| ✓ FLT3(R834Q): 1300 | 66) MEK1: 1800 |
| 47) FLT4: 1100 | 67) MEK2: 1100 |
| 48) FRK: 170 | 68) MEK5: 49 |
| 49) FYN: 360 | 69) MERTK: 1400 |
| 50) GAK: 86 | 70) MET: 5700 |
| 51) HCK: 360 | ✓ MET(Y1235D): 4100 |
| 52) HPK1: 5500 | 71) MINK: 3400 |
| | 72) MKNK1: 360 |

- 73) MKNK2: 1700
 74) MRCKA: 2600
 75) MRCKB: 2500
 76) PDGFRA: 230
 77) PDGFRB: 88
 78) PHKG1: 1100
 79) PHKG2: 7900
 80) PLK4: 620
 81) QSK: 3900
 82) RET: 34
 ✓ RET(M918T): 14
 83) RIPK2: 4.6
 84) RIPK4: 620
 85) ROCK2: 3600
 86) RSK1(Kin.Dom.2-C-terminal): 400
 87) RSK4(Kin.Dom.2-C-terminal): 240
 88) S6K1: 1600
 89) SIK: 1900
 90) SIK2: 430
 91) SLK: 95
 92) SRC: 70
 93) SRMS: 1900
 94) STK33: 1400
 95) STK35: 56
 96) TESK1: 3700
 97) TIE1: 1500
 98) TIE2: 1000
 99) TNIK: 2300
 100) TNNI3K: 2800
 101) TXK: 3700
 102) TYRO3: 93
 103) ULK3: 6400
 104) VEGFR2: 820
 105) YANK1: 5000
 106) YES: 120
 107) YSK4: 980
 108) ZAK: 5100

Afatinib

1. induces apoptosis in cancer cells without EGFR mutation via suppressing CIP2A at the gene transcription level by reducing the promoter binding activity of Elk-1(395)
2. decreases epithelial-mesenchymal transition and tumorigenesis by regulating the ERK-VEGF/MMP9 signaling pathway(396)
3. hampers AKT-mTOR activation by stimulating PERK-eIF2 α -ATF4 signaling pathway, giving rise to MCL-1 downregulation mediated apoptosis(397)
4. directly inhibits ErBB-4 with an EC₅₀ of 1nM(398)
5. downregulates ribonucleotide reductase(399)
6. directly binds these kinases (K_d, nM) (NP: nonphosphorylated, P: phosphorylated)(324):
 - 1) TXK: 3100
 - 2) SRC: 2800
 - 3) SLK: 3700

- 4) SBK1: 4800
- 5) RSK4(Kin.Dom.2-C-terminal): 3100
- 6) RIPK2: 2700
- 7) PHKG2: 470
- 8) PHKG1: 1300
- 9) p38-alpha: 1300
- 10) MKNK1: 1800
- 11) MKNK2: 1600
- 12) MET: 2200
- ✓ MET(M1250T): 6000
 - ✓ MET(Y1235D): 2600
- 13) MEK5: 1300
- 14) LOK: 4300
- 15) LCK: 570
- 16) JNK2: 2100
- 17) JNK3: 2000
- 18) IRAK1: 240
- 19) HIPK4: 360
- 20) HCK: 2200
- 21) GAK: 79
- 22) FRK: 1400
- 23) FLT3(N841I): 6200
- ✓ FLT3(D835H): 1800
 - ✓ FLT3(D835Y): 1400
- 24) EPHB6: 3100
- 25) ERBB2: 5
- 26) ERBB3: 4500
- 27) ERBB4: 6.3
- 28) EPHA6: 340
- 29) DYRK1A: 970
- 30) DYRK1B: 2800
- 31) DYRK2: 1800
- 32) DRAK1: 2300
- 33) CSNK1E: 1300
- 34) CIT: 2900
- 35) BLK: 220
- 36) AXL: 5300
- 37) ABL1-NP: 1300
- ✓ ABL1-P: 570
 - ✓ ABL1(E255K)-P: 420
 - ✓ ABL1(F317I)-NP: 2900
 - ✓ ABL1(F317I)-P: 750
 - ✓ ABL1(F317L)-NP: 580
 - ✓ ABL1(F317L)-P: 230
 - ✓ ABL1(H396P)-NP: 500
 - ✓ ABL1(H396P)-P: 500
 - ✓ ABL1(M351T)-P: 1200
 - ✓ ABL1(Q252H)-NP: 790
 - ✓ ABL1(Q252H)-P: 380
 - ✓ ABL1(T315I)-NP: 2600
 - ✓ ABL1(T315I)-P: 870
 - ✓ ABL1(Y253F)-P: 830

Osimertinib

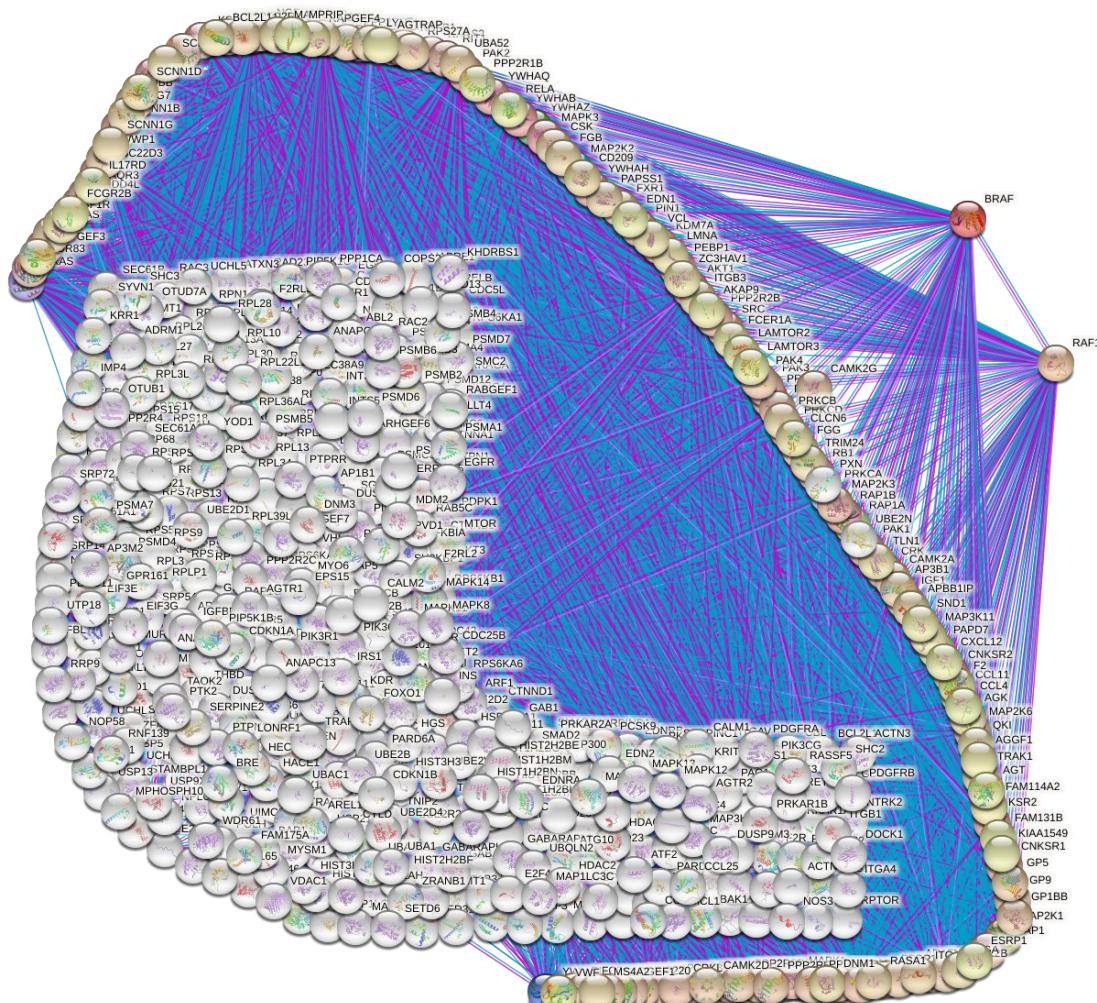
1. may reactivate immune activity of T cells in tumor microenvironment via reducing **PD-L1** mRNA expression and inducing its protein degradation(400)

2. reduces EZH2 mRNA expression via up-regulating the expression of a tumor suppressor, miR-34a(401)
3. induces apoptosis via reactive oxygen species generation(402)
4. disrupts the interaction of EZH2–EED, leading to impairment of PRC2 activity and downregulation of EZH2 protein(401)
5. exerts antitumor effects via directly inhibiting MNK1 and MNK2 kinases with IC₅₀ values of 324 nM and 48.6 nM, respectively, and subsequently suppressing the phosphorylation of eIF4E(403)
6. can trigger autophagy-mediated cell death by increasing the expression of phosphatidylethanolamine-modified microtubule-associated protein light-chain 3 (LC3) and decreasing the expression of p62(404)
7. directly inhibits LSD1 with an IC₅₀ of 3.98 ± 0.3 μM(405)

Neratinib

1. It enhances LATS1/2 phosphorylation independently of RAP2A/MAP4K4 and that MST4 degradation and Ezrin dephosphorylation may represent a universal trigger for the biological actions of neratinib(406).
2. inhibits Hippo/YAP signaling, reduces mutant K-RAS expression, and kills pancreatic and blood cancer cells(407)
3. induces ubiquitylation-mediated endocytic degradation of ErbB2(408)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
SORAFENIB		RAF1	140
PAZOPANIB	c-Raf	P04049	107
AXITINIB	B-Raf	P15056	102
REGORAFENIB		BRAF	20
LENVATINIB			10



<https://version-11-0b.string-db.org/cgi/network?networkId=bkp1TxEsAQKB>

Sorafenib

1. downregulates the respiratory chain complex I (**NADH dehydrogenase**) which is accompanied with the loss of mitochondrial transmembrane potential ($\Delta\Psi_m$) and complete impairment of complex I enzyme activity(409)
2. promotes immune responsiveness via selectively increasing effector T cell activation while blocking regulatory T cell function(410), also see (411-417)
3. enhances antitumor immunity via modulating immunosuppressive cell populations(418)
4. augments antitumor immunity via relieving cell-intrinsic and cell-extrinsic inhibitions of effector T cells in tumor microenvironment(419)
5. inhibits TGF- β signaling mainly by inducing caveolae/lipid raft-mediated internalization and degradation of cell-surface **T β R-II** in target cells(420)
6. dose-dependently induces the generation of ROS in tumor cells(421)
7. inhibits cell viability, migration, invasion, and STAT3 activation via **ARHGAP24**(422)
8. could potentially reverse the immunosuppressive **cytokine** profile of tumor-associated macrophages, rendering the tumor microenvironment more conducive to an anti-tumor immune response(423)
9. modulate the expression of a wide range of ncRNAs and specifically, **GAS5** and **miR-126-3p**(424)
10. hinders oxidative phosphorylation, whereas at the same time stimulates aerobic glycolysis in glucose-grown cells, hence attenuating the cellular **ATP depletion**(425)
11. inhibits cell proliferation and induces apoptosis through the mitochondrial pathway, independent of MAPK and AKT and via AMPK-dependent inhibition of the mTORC1 pathway(426)
12. induces apoptosis and has antiproliferative properties via exerting free radical scavenging properties associated with the downregulation of nuclear factor E2-related factor 2 (Nrf2)-regulated thioredoxin 1 (**Trx1**) expression(427)
13. inhibits migration and invasion of cancer cells through suppression of matrix metalloproteinase expression(428)
14. induces apoptosis and exerts its effect – at least in part – by CSF1R inhibition(429)
15. directly binds these kinases (K_d (μM))(348):
 - 1) ABL1: 0.13
 - ✓ ABL1(E255K): 4.4
 - ✓ ABL1(H396P): 1.2
 - ✓ ABL1(M351T): 0.23
 - ✓ ABL1(Q252H): 0.45
 - ✓ ABL1(T315I): 0.17
 - ✓ ABL1(Y253F): 0.58
 - 2) ABL2: 1.3
 - 3) CDK5: 6.2
 - 4) EPHA2: 4.8
 - 5) EPHA3: 6.4
 - 6) EPHA4: 1.3
 - 7) EPHA5: 0.36

- 8) EPHA6: 0.24
- 9) EPHA7: 0.67
- 10) EPHA8: 0.96
- 11) EPHB1: 1.7
- 12) EPHB4: 3.9
- 13) FGFR1: 2.5
- 14) FLT3: 0.02
- 15) FLT4: 0.016
- 16) FRK: 0.44
- 17) JNK2: 3.6
- 18) KIT: 0.74
- 19) LCK: 6.9
- 20) LIMK1: 2.2
- 21) MAP4K5: 1.7
- 22) MKNK2: 0.25
- 23) MYLK2: 1.3
- 24) NTRK1: 3.6
- 25) p38-alpha: 0.26
- 26) p38-beta: 0.2
- 27) p38-gamma: 9.9
- 28) PDGFRB: 0.041
- 29) RIPK2: 2.9
- 30) SLK: 1.6
- 31) STK10: 0.14
- 32) STK18: 3.4
- 33) STK36: 5.4
- 34) TEK: 3
- 35) VEGFR2: 0.093
16. induces ferroptosis, a novel form of programmed cell death(430) probably via the **retinoblastoma protein**(431)
17. sensitization of cancer cells to TRAIL- and Fas-induced apoptosis, dependent on **FLIP** and independent of Raf/MEK/ERK inhibition(432, 433)
18. promotes oxidative stress and mitochondrial dysfunction by inducing mitochondrial dysfunction via increasing reactive oxygen species formation and activating JNKs, leading to translocation of activated JNK to mitochondria(434)
19. Besides functioning as a multiple tyrosine kinase, sorafenib also acts via a kinase-independent mechanism to target **STAT3** signaling in cancer cells. Sorafenib inactivates STAT3 and STAT3-related signaling by inducing a conformational change in and releasing the autoinhibition of Src homology region 2 domain-containing phosphatase-1(435).
20. induces the formation of Atg5-deficient **autophagosomes** which promotes the interaction of p62 with RIPK, leading to caspase-independent cell death by necroptosis(436)
21. Sorafenib downregulates AIB1 protein expression by inhibiting AIB1 mRNA translation through simultaneously blocking **eIF4E** and mTOR/p70S6K/RP-S6 signaling. Downregulation of AIB1 contributes to sorafenib-induced cell death at least in part through upregulating the levels of reactive oxygen species(437).
22. can inhibit the proliferation of non-small cell lung cancer H460 and A549 cells, and block the cells in the G1 phase, which may be related to the regulation of Cyclin E1 and **E2F1** protein(438)

23. Sorafenib suppresses ATP production, resulting in AMPK activation via phosphorylation. Further secondary effects include reduction of the levels of SREBP1, phosphorylation of mTOR, disruption of lipogenesis and liver cancer cell death(439).
24. Inhibits soluble epoxide hydrolase(440)
25. decreases proliferation and induces apoptosis of prostate cancer cells by inhibition of the androgen receptor and prostate-specific antigen expression(441)
26. induces cell death through a process involving induction of endoplasmic reticulum stress independent of MEK1/2-ERK1/2(442)
27. induces alteration of cell death receptor S-nitrosylation status which may have a relevant repercussion on cell death signaling(443)
28. may suppress mTOR signaling by direct inhibition of FLT3(444)
29. induces autophagic cell death through Akt inhibition in a RAF-MEK-ERK1/2 independent fashion(445)
30. inhibits macrophage-induced growth of cancer cells by interference with insulin-like growth factor-1 secretion(417)
31. exerts antitumor effects via a kinase-independent mechanism: SHP-1 dependent STAT3 inactivation(446, 447)
32. induces cell death via increasing the expression of extracellular vesicle long noncoding RNA (linc-VLDLR) in tumor cells(448)
33. inhibits invasion and metastasis by regulating the expression of MMP and TIMP gene families(449)
34. directly inhibits CSF-1R(450)
35. osteosarcoma cells respond to sorafenib chemotherapy by downregulation of the tumor progression factors S100A4, CXCR4 and the oncogene FOS(451)
36. induces apoptosis via upregulating ATF4
37. reduces infiltrated regulatory T cells by suppressing TGF-beta signaling(452)
38. suppresses growth and survival of cancer cells by accelerating degradation of EZH2(453)
39. Besides inhibition of the Raf-MAPK pathway, Sorafenib might also regulate hepatoma cell growth via alteration of receptor-mediated cytoskeletal rearrangement(454).
40. inhibits cancer cell proliferation and invasion by inhibiting MMP-2 and Ki-67 expression due to up-regulation of P53 and suppressing FoxM1(455)
41. inhibits tumor invasion and induces apoptosis via blocking the HIF-1 α /VEGFA pathway(456)
42. These data indicate that the inhibition of STAT3 activity by sorafenib involves both the inhibition of upstream kinases (JAK1 and JAK2) of STAT3 and increased phosphatase activity. Phosphorylation of AKT was also reduced by sorafenib. In contrast, mitogen-activated protein kinases were not consistently inhibited by sorafenib in these cells(457).
43. inhibits STAT3 signaling associated with growth arrest and apoptosis(458)
44. inhibits AAA+ ATPase p97/VCP which leads to disruption of the secretory pathway, endoplasmic reticulum stress, and cancer cell death(459)

45. It inhibits ribonucleotide reductase regulatory subunit M2 (**RRM2**). This partially contributes to its anticancer activity(460).
46. directly binds **MAPK14**(461)
47. suppresses extrahepatic metastasis through inhibition of mesenchymal cancer stem cells characterized by the expression of CD90 and inhibiting the production of extracellular vesicles which contain TGF- β mRNA and regulate distant metastasis(462)
48. targets the mitochondrial electron transport chain complexes and ATP synthase to activate the PINK1-Parkin pathway and modulate cellular drug response(463)
49. In addition to its anticancer effects, it has an immunoregulatory property via inducing survival and differentiation of **bone marrow cells** that is apparent at low doses(464).
50. directly binds these kinases (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):
- 1) ABL1-NP: 130
 - ✓ ABL1-P: 1400
 - ✓ ABL1(E255K)-P: 3400
 - ✓ ABL1(F317I)-NP: 1700
 - ✓ ABL1(F317I)-P: 4900
 - ✓ ABL1(F317L)-NP: 770
 - ✓ ABL1(F317L)-P: 7100
 - ✓ ABL1(H396P)-NP: 520
 - ✓ ABL1(H396P)-P: 2400
 - ✓ ABL1(M351T)-P: 1200
 - ✓ ABL1(Q252H)-NP: 130
 - ✓ ABL1(Q252H)-P: 5500
 - ✓ ABL1(T315I)-NP: 590
 - ✓ ABL1(T315I)-P: 1500
 - ✓ ABL1(Y253F)-P: 660
 - 2) ABL2: 2900
 - 3) AURKB: 440
 - 4) AURKC: 210
 - 5) AXL: 4500
 - 6) BRAF: 540
 - ✓ BRAF(V600E): 260
 - 7) CDK11: 250
 - 8) CDK2: 8700
 - 9) CDK3: 3800
 - 10) CDK5: 8300
 - 11) CDK7: 140
 - 12) CDK8: 310
 - 13) CDKL2: 130
 - 14) CDKL3: 490
 - 15) CIT: 6200
 - 16) CLK1: 7500
 - 17) CSF1R: 28
 - 18) DDR1: 1.5
 - 19) DDR2: 6.6
 - 20) EPHA1: 3100
 - 21) EPHA2: 2000
 - 22) EPHA3: 1900
 - 23) EPHA4: 3000

- 24) EPHA5: 3500
 25) EPHA6: 370
 26) EPHA7: 5300
 27) EPHA8: 2400
 28) EPHB1: 3000
 29) EPHB2: 1900
 30) EPHB4: 1800
 31) EPHB6: 240
 32) ERK8: 46
 33) FGFR1: 2800
 34) FGFR2: 2700
 35) FGFR3: 4200
 ✓ FGFR3(G697C): 5900
 36) FGR: 7800
 37) FLT1: 31
 38) FLT3: 13
 ✓ FLT3(D835H): 30
 ✓ FLT3(D835Y): 82
 ✓ FLT3(ITD): 79
 ✓ FLT3(K663Q): 4.5
 ✓ FLT3(N841I): 11
 ✓ FLT3(R834Q): 17
 39) FLT4: 95
 40) FRK: 510
 41) FYN: 8400
 42) HCK: 8500
 43) HIPK1: 3300
 44) HIPK2: 1700
 45) HIPK3: 440
 46) HIPK4: 3.3
 47) HPK1: 4800
 48) JAK3(JH1domain-catalytic): 7300
 49) JNK2: 7400
 50) KIT: 28
 ✓ KIT(A829P): 20
 ✓ KIT(D816H): 430
 ✓ KIT(D816V): 310
 ✓ KIT(L576P): 25
 ✓ KIT(V559D): 16
 ✓ KIT(V559D,T670I): 18
 ✓ KIT(V559D,V654A): 240
 51) LCK: 2700
 52) LIMK1: 1600
 53) LIMK2: 9600
 54) LOK: 150
 55) LYN: 3000
 56) MAP4K4: 4800
 57) MAP4K5: 1600
 58) MEK5: 190
 59) MERTK: 3600
 60) MKNK1: 230
 61) MKNK2: 130
 62) MLCK: 9600
 63) MUSK: 130
 64) MYLK2: 5400
 65) MYO3B: 7100

- 66) NLK: 640
 67) p38-alpha: 370
 68) p38-beta: 230
 69) p38-delta: 6600
 70) p38-gamma: 7600
 71) PCTK2: 1600
 72) PDGFRA: 62
 73) PDGFRB: 37
 74) PFCDPK1(*P. falciparum*): 220
 75) PFTAIRE2: 2900
 76) PFTK1: 2900
 77) PLK4: 4500
 78) RAF1: 230
 79) RET: 13
- ✓ RET(M918T): 7.4
 - ✓ RET(V804L): 39
 - ✓ RET(V804M): 22
- 80) RIPK2: 1300
 81) RSK4(Kin.Dom.2-C-terminal): 7500
 82) SLK: 1000
 83) SRMS: 9800
 84) STK33: 2400
 85) STK36: 3800
 86) TAK1: 690
 87) TAOK1: 3100
 88) TAOK2: 540
 89) TAOK3: 2700
 90) TGFBR2: 6900
- 91) TIE1: 68
 92) TIE2: 2100
 93) TNIK: 7900
 94) TNK1: 2300
 95) TNNI3K: 280
 96) TRKA: 6300
 97) TRKB: 2100
 98) TRKC: 600
 99) TTK: 3500
 100) VEGFR2: 59
 101) YSK4: 99
 102) ZAK: 6.3
51. triggers cell growth inhibition and apoptosis by directly targeting the mitochondria and inducing rapid mitochondrial fragmentation, which is associated with the deregulation of mitochondria fusion-related protein optic atrophy 1 (OPA1)(465)
52. ID1 might be a potential target for the antitumor activity of sorafenib by contributing to its antitumor activity by suppressing epithelial to mesenchymal transition(466).
53. upregulates MiR-1274a which can repress the expression of ADAM9(467)
- Pazopanib**
1. triggers autophagic cell death via inducing cathepsin B activation, upregulating the glucosidase and downregulating the TP73 mRNA expression(468)
 2. potently inhibits several mammalian carbonic anhydrases and in addition to its tyrosine kinase inhibitory action, it may

- exert antitumor/antimetastatic effects also due to the potent inhibition of the tumor-associated, hypoxia-inducible enzymes carbonic anhydrase IX and XII(469)
3. immunomodulation through priming dendritic cells by downregulation of the β -catenin pathway(470)
 4. directly binds and inhibits MEKK2 with an IC₅₀ of 698 ± 163 nM(471)
 5. directly binds these kinases (K_d, nM) (NP: nonphosphorylated, P: phosphorylated)(324):
 - 1) AAK1: 2900
 - 2) ABL1-NP: 620
 - ✓ ABL1-P: 650
 - ✓ ABL1(E255K)-P: 800
 - ✓ ABL1(F317L)-NP: 4400
 - ✓ ABL1(F317L)-P: 2700
 - ✓ ABL1(H396P)-NP: 380
 - ✓ ABL1(H396P)-P: 700
 - ✓ ABL1(M351T)-P: 560
 - ✓ ABL1(Q252H)-NP: 530
 - ✓ ABL1(Q252H)-P: 530
 - ✓ ABL1(T315I)-NP: 3800
 - ✓ ABL1(T315I)-P: 2100
 - ✓ ABL1(Y253F)-P: 480
 - 3) ABL2: 3000
 - 4) ACVR2B: 2400
 - 5) ALK: 2200
 - 6) AURKA: 7100
 - 7) AURKC: 750
 - 8) BIKE: 8800
 - 9) BLK: 2600
 - 10) BMPR1B: 1000
 - 11) BRAF: 730
 - 12) BRAF(V600E): 430
 - 13) BRK: 2300
 - 14) CAMK1: 2100
 - 15) CAMK1G: 3700
 - 16) CDC2L1: 2100
 - 17) CDC2L2: 1300
 - 18) CSF1R: 7.9
 - 19) DDR1: 57
 - 20) DDR2: 98
 - 21) EPHB6: 81
 - 22) FER: 2700
 - 23) FES: 1400
 - 24) FGFR1: 990
 - 25) FGFR2: 210
 - 26) FGFR3: 740
 - ✓ FGFR3(G697C): 620
 - 27) FGFR4: 2800
 - 28) FGR: 1600
 - 29) FLT1: 14
 - 30) FLT3: 1100
 - ✓ FLT3(D835H): 1100
 - ✓ FLT3(D835Y): 810
 - ✓ FLT3(ITD): 2500

- ✓ FLT3(K663Q): 740
- ✓ FLT3(N841I): 1600
- ✓ FLT3(R834Q): 9600
- 31) FLT4: 27
- 32) FRK: 750
- 33) FYN: 2700
- 34) GAK: 200
- 35) HCK: 5700
- 36) HPK1: 750
- 37) IRAK1: 5900
- 38) IRAK3: 800
- 39) JAK2(JH1domain-catalytic): 1700
- 40) JAK3(JH1domain-catalytic): 6900
- 41) JNK1: 2000
- 42) JNK3: 1900
- 43) KIT: 2.8
 - ✓ KIT(A829P): 33
 - ✓ KIT(D816H): 1000
 - ✓ KIT(D816V): 500
 - ✓ KIT(L576P): 1.8
 - ✓ KIT(V559D): 2.3
 - ✓ KIT(V559D,T670I): 6.5
 - ✓ KIT(V559D,V654A): 30
- 44) LCK: 1200
- 45) LIMK1: 720
- 46) LIMK2: 390
- 47) LOK: 84
- 48) LTK: 6800
- 49) LYN: 1100
- 50) MAP3K1: 2900
- 51) MAP3K2: 290
- 52) MAP4K2: 2700
- 53) MAP4K3: 1600
- 54) MAP4K4: 2200
- 55) MAP4K5: 3000
- 56) MARK3: 4000
- 57) MEK4: 590
- 58) MEK5: 480
- 59) MEK6: 4100
- 60) MERTK: 3300
- 61) MET: 2000
 - ✓ MET(M1250T): 3400
 - ✓ MET(Y1235D): 2100
- 62) MLK1: 290
- 63) MLK2: 2100
- 64) MLK3: 740
- 65) MYLK2: 2000
- 66) NEK2: 980
- 67) NEK5: 7300
- 68) NLK: 4400
- 69) PCTK1: 1200
- 70) PDGFRA: 4.9
- 71) PDGFRB: 2
- 72) PFCDPK1(*P. falciparum*): 370
- 73) PIK4CB: 960
- 74) PIP5K1C: 1900

- 75) PIP5K2C: 280
- 76) PLK4: 290
- 77) PRKR: 1900
- 78) RAF1: 900
- 79) RET: 310
- ✓ RET(M918T): 270
 - ✓ RET(V804L): 5900
 - ✓ RET(V804M): 2300
- 80) RIOK2: 610
- 81) RIPK1: 260
- 82) RIPK2: 580
- 83) ROS1: 920
- 84) SIK: 2200
- 85) SIK2: 7300
- 86) SLK: 240
- 87) SRC: 2800
- 88) SRMS: 2500
- 89) STK16: 360
- 90) STK36: 470
- 91) SYK: 4700
- 92) TAOK1: 240
- 93) TAOK2: 1800
- 94) TAOK3: 45
- 95) TGFBR2: 3000
- 96) TIE1: 700
- 97) TIE2: 3300
- 98) TNIK: 310
- 99) TTK: 150
- 100) TXK: 2600
- 101) TYK2(JH1domain-catalytic): 3400
- 102) VEGFR2: 14
- 103) YES: 5000
- 104) YSK4: 940
- Axitinib**
1. induces DNA damage response characterized by γ -H2AX phosphorylation and Chk1 kinase activation leading to G2/M cell cycle arrest and mitotic catastrophe(472)
 2. triggers senescence through ROS accumulation and ATM activation(473)
 3. induces DNA damage response leading to senescence, mitotic catastrophe, and increased NK cell recognition through inducing a DNA damage response initially characterized by γ -H2AX phosphorylation and Chk1 kinase activation and at later time points by p21 overexpression(474)
 4. blocks Wnt/ β -catenin signaling and directs asymmetric cell division in cancer via directly stabilizing SHPRH and thereby increasing the ubiquitination and degradation of β -catenin(475)
 5. induces the apoptosis of cancer cells possibly through decreasing the mitochondrial membrane potential(476)
 6. has the potential to modulate antitumor immunity by downregulating STAT3 expression and reversing MDSC-mediated tumor-induced immunosuppression(477)
 7. exerts an anticancer effect in melanoma through promoting antitumor immunity(478)

- 8) directly inhibits CSF-1R(450)
- 9) directly binds these kinases (K_d , nM)
(NP: nonphosphorylated, P: phosphorylated)(324):
- 1) ZAK: 2600
 - 2) YSK4: 270
 - 3) YES: 2700
 - 4) VEGFR2: 5.9
 - 5) ULK3: 670
 - 6) ULK2: 9900
 - 7) TYK2(JH1domain-catalytic): 3600
 - 8) TXK: 2400
 - 9) TTK: 3900
 - 10) TRKA: 1800
 - 11) TNK2: 6000
 - 12) TNK1: 160
 - 13) TNIK: 180
 - 14) TIE2: 310
 - 15) TIE1: 97
 - 16) TESK1: 4300
 - 17) STK16: 370
 - 18) SRPK3: 2300
 - 19) SRPK1: 1800
 - 20) SNARK: 1300
 - 21) SLK: 1000
 - 22) SIK: 2100
 - 23) ROS1: 3200
 - 24) RIPK4: 1400
 - 25) RIPK2: 9900
 - 26) RIPK1: 2500
 - 27) RIOK3: 1000
 - 28) RIOK1: 1200
 - 29) RET: 120
 - ✓ RET(V804M): 1600
 - ✓ RET(V804L): 1300
 - ✓ RET(M918T): 100
 - 30) PRKR: 7400
 - 31) PLK4: 16
 - 32) PKN2: 2700
 - 33) PDGFRB: 0.57
 - 34) PDGFRA: 0.51
 - 35) PCTK1: 5300
 - 36) PAK4: 4500
 - 37) MYO3A: 1900
 - 38) MYLK2: 1300
 - 39) MST2: 2200
 - 40) MST1: 1400
 - 41) MRCKB: 4800
 - 42) MLK1: 3000
 - 43) MLCK: 1700
 - 44) MINK: 560
 - 45) MET: 820
 - ✓ MET(Y1235D): 990
 - ✓ MET(M1250T): 1100
 - 46) MERTK: 3200
 - 47) MEK5: 140
 - 48) MEK2: 2600

49) MEK1: 2700	68) GAK: 2700
50) MAP4K5: 550	69) FLT4: 170
51) MAP4K4: 350	70) FLT1: 5.8
52) MAP4K3: 1200	71) FLT3: 42
53) MAP4K2: 1300	✓ FLT3(D835H): 330
54) MAP3K4: 5600	✓ FLT3(D835Y): 270
55) MAP3K3: 2800	✓ FLT3(ITD): 250
56) LRRK2: 920	✓ FLT3(K663Q): 31
✓ LRRK2(G2019S): 990	✓ FLT3(N841I): 200
57) LOK: 1200	✓ FLT3(R834Q): 2400
58) LCK: 2700	72) FGR: 1800
59) KIT: 3.2	73) FGFR1: 380
✓ KIT(A829P): 1800	74) FGFR2: 110
✓ KIT(D816H): 4200	75) FGFR3: 210
✓ KIT(D816V): 1300	✓ FGFR3(G697C): 210
✓ KIT(L576P): 1.7	76) EPHB6: 360
✓ KIT(V559D): 0.49	77) EGFR(G719C): 2300
✓ KIT(V559D,T670I): 1.4	✓ EGFR(G719S): 6200
✓ KIT(V559D,V654A): 3.5	78) DDR1: 340
60) JNK1: 3900	79) DDR2: 5300
61) JNK3: 1100	80) DLK: 1700
✓ JAK3(JH1domain-catalytic): 3100	81) CSNK2A2: 2800
62) JAK2(JH1domain-catalytic): 3300	82) CSK: 1100
63) ITK: 1200	83) CSF1R: 21
64) IRAK1: 3100	84) CDKL2: 530
65) HUNK: 1500	85) CAMKK1: 1400
66) HPK1: 330	86) CAMKK2: 1500
67) GRK4: 2300	87) BMPR1B: 2900

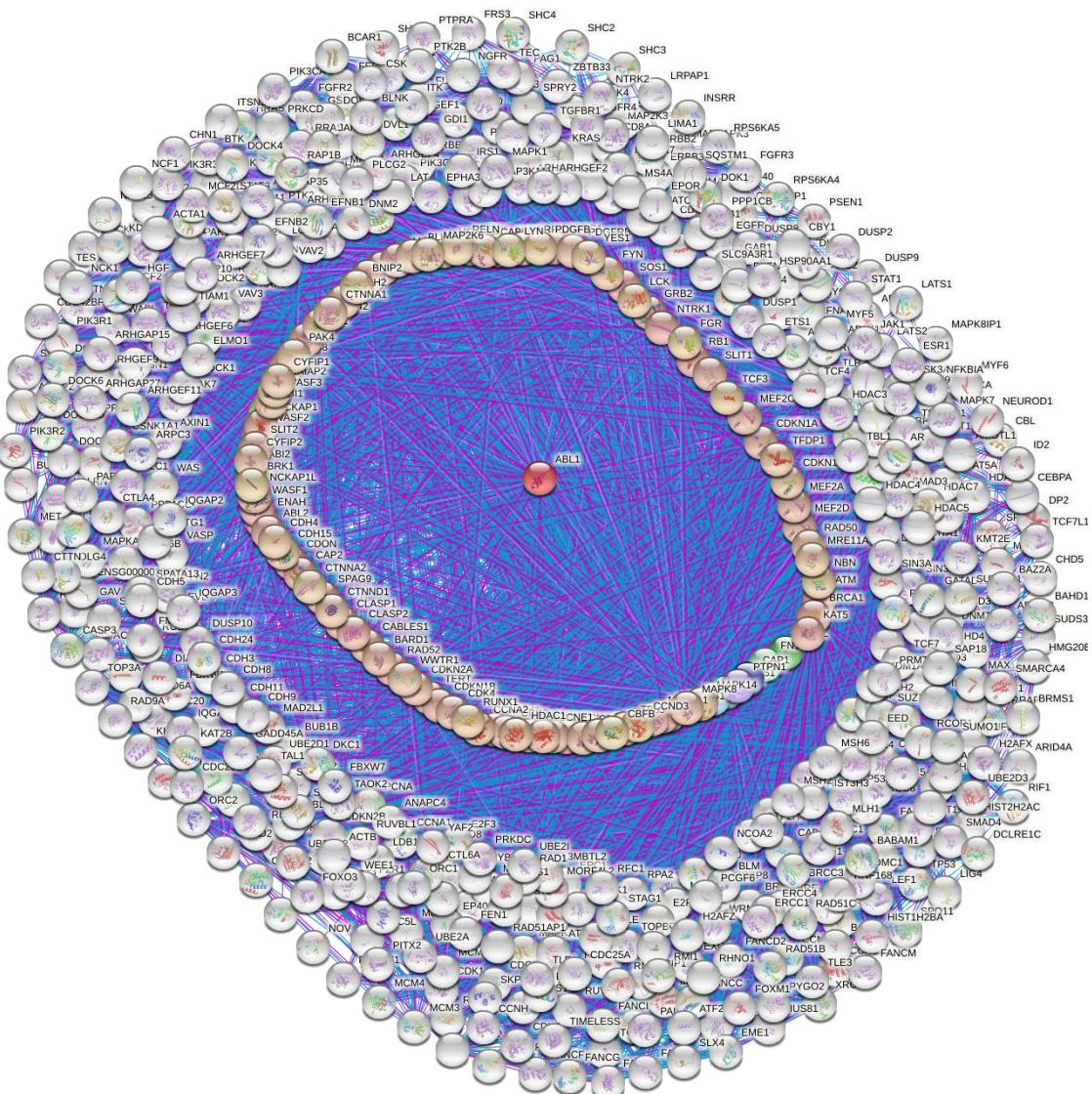
- 88) AURKA: 72
- 89) AURKB: 11
- 90) AURKC: 1.3
- 91) AXL: 420
- 92) BIKE: 1000
- 93) BLK: 3100
- 94) ANKK1: 3600
- 95) ACVR2B: 5200
- 96) AAK1: 1200
- 97) ABL1-NP: 84
- ✓ ABL1-P: 36
 - ✓ ABL1(E255K)-P: 63
 - ✓ ABL1(F317I)-NP: 2600
 - ✓ ABL1(F317I)-P: 800
 - ✓ ABL1(F317L)-NP: 830
 - ✓ ABL1(F317L)-P: 330
 - ✓ ABL1(H396P)-NP: 20
 - ✓ ABL1(H396P)-P: 23
 - ✓ ABL1(M351T)-P: 36
 - ✓ ABL1(Q252H)-NP: 290
 - ✓ ABL1(Q252H)-P: 200
 - ✓ ABL1(T315I)-NP: 3.6
 - ✓ ABL1(T315I)-P: 1.5
 - ✓ ABL1(Y253F)-P: 230
- 98) ABL2: 70
- Regorafenib**
1. can exert antitumor effects via directly binding microRNA-21 pre-element(479)
 2. exerts superior antitumor effects by enhancing SHP-1 activity that directly targets p-STAT3Tyr705(480)
 3. Hippo signaling pathway is a mediator of regorafenib efficacy(481).
 4. directly stabilizes PSAT1 to trigger PRKAA-dependent autophagy initiation and inhibit RAB11A-mediated autophagosome-lysosome fusion, resulting in lethal autophagy arrest(482)
 5. abolishes epithelial-to-mesenchymal transition-related invasion/metastasis via enhancing SHP-1 activity which impedes TGF-β1-induced epithelial-to-mesenchymal transition/invasion through low p-STAT3Tyr705 level(483)
 6. induces significant tumor inhibition by relieving the autoinhibited N-SH2 domain of SHP-1 directly and inhibiting p-STAT3 signals(484)
 7. can efficiently block noncanonical Ang-2-driven angiogenesis by inhibiting the Ang-2/Tie2 axis(485)
 8. inhibits invasion and metastasis by regulating the expression of MMP and TIMP gene families(449)
 9. promotes antitumor immunity via inhibiting PD-L1 and IDO1 expression(486, 487)
 10. inhibits Fos/Jun pathway(488)
 11. inhibits cell proliferation and invasion of cancer cells via decreasing the expression of CXCR4 and further reducing the transcriptional activity of Wnt/β-Catenin pathway(489)
 12. directly inhibits RIPK2(490)
 13. inhibits these protein kinases(491):

- 1) VEGFR-1 ($IC_{50}=13\pm0.4\text{nm}$)
- 2) MURINE VEGFR-2($IC_{50}=4.2\pm1.6\text{nm}$)
- 3) MURINE VEGFR-3 ($IC_{50}=46\pm10\text{nm}$)
- 4) TIE2 ($IC_{50}=311\pm46\text{nm}$)
- 5) FGFR1 ($IC_{50}=202\pm18\text{nm}$)
- 6) PDGFR- β ($IC_{50}=22\pm3\text{nm}$)
- 7) KIT ($IC_{50}=7\pm2\text{nm}$)
- 8) RET ($IC_{50}=1.5\pm0.7\text{nm}$)
- 9) RAF-1 ($IC_{50}=2.5\pm0.6\text{nm}$)

Lenvatinib

1. inhibits invasion and metastasis by regulating the expression of **MMP and TIMP** gene families(449)
2. has **immunomodulatory** activity that contributes to its antitumor activity(492)
3. directly inhibits these kinases(493):
 - 1) VEGFR1($K_i = 1.3\text{nm}$)
 - 2) VEGFR2($K_i = 0.74\text{nm}$)
 - 3) VEGFR3($K_i = 0.71\text{nm}$)
 - 4) FGFR1($K_i = 22\text{nm}$)
 - 5) FGFR2($K_i = 8.2\text{nm}$)
 - 6) FGFR3($K_i = 15\text{nm}$)
 - 7) RET($K_i = 1.5\text{nm}$)
 - 8) KIT($K_i = 11\text{nm}$)

Drugs	Discovery Target	Uniprot "Off-Target" Therapeutic ID	Therapeutic Mechanisms
IMATINIB			78
DASATINIB	NILOTINIB	ABL1	ABL1 P00519 158 63
PONATINIB	BOSUTINIB		13 74
RIPRETTINIB	SELPERCATINIB		



<https://version-11-0b.string-db.org/cgi/network?networkId=bwgtq8WeUcGW>

Imatinib

1. induces a significant increase in Type I (IFN- γ) cytokine-producing T cells(494)
2. potentiates antitumor T cell responses through the inhibition of IDO(495), see (496)
3. inhibition of topoisomerases may be a significant factor in imatinib-induced apoptosis(497)
4. induces apoptosis through Bim accumulation independently of cell cycle arrest(498)
5. increases apoptosis index through modulation of survivin subcellular localization in the blast phase of CML cells(499)
6. exerts natural killer cell-dependent anti-tumor effects via increasing the IFN- γ production by natural killer cells(500), see (496)
7. inhibits DDR1 and DDR2(501, 502)
8. induces apoptosis through production of reactive oxygen species(503)
9. reduces myeloid suppressor cells and releases effector lymphocyte responses(504)
10. inhibits the RET tyrosine kinase activity in a dose-dependent manner(505, 506)
11. Its anti-leukemic mechanism may involve not only the inhibition of BCR/ABL, but also DNA damage in the cells expressing this fusion protein(507).
12. induces downregulation of c-Kit by targeting the ATP pocket(508)
13. promotes apoptosis by targeting microRNA-30a-mediated runt-related transcription factor 2(509)
14. targets the macrophage colony-stimulating factor (M-CSF) receptor, c-FMS, and can inhibit cell proliferation and metastasis(510, 510-513)
15. reduces cancer stem cell ability and induces cell differentiation via continuous inhibition of PDGFR and c-Kit signaling through activation of MAPK signalling(514)
16. might act as an effective inhibitor of vacuolar H⁺-ATPase function(515)
17. has an antifibrotic effect on human breast stromal fibroblasts that may inhibit desmoplastic reaction and thus tumor progression via downregulation of mRNA synthesis of collagen I and collagen III(516)
18. directly inhibits these kinases(517):
 - 1) PDGFR α
 - 2) PDFGFR β
 - 3) Axl
 - 4) RYK
 - 5) EGFR
 - 6) EphA2
 - 7) EphA10
 - 8) IGF1R
 - 9) CSF-1R
19. Antiapoptotic proteins Bcl2 and BclX do not protect chronic myeloid leukemia cells from imatinib-mediated growth arrest. We conclude that, besides its apoptotic effect, imatinib acts through an apoptosis-independent mechanism to arrest cell growth(518).
20. directly binds MAPK14(461)

21. induces up-regulation of **NM23**, a metastasis suppressor gene(519)
22. It sensitizes T1 cells by directly downregulating **c-FLIPL**, with the use of an alternative pathway for antitumor activity, because PDGFR α is not activated in T1 cells and these cells do not express c-kit, c-ABL or PDGFR β (520). See (496).
23. directly targets these proteins(521, 522):
- 1) **ARG**(523, 524)
 - 2) **KIT**(525, 526)
 - 3) **PDGFR**(526, 527)
 - 4) **DDR1**
 - 5) **NQO2**
 - 6) **CDC2**
24. has antitumor effects on the GIST cells with respect to not only the inhibition of cell growth but also the suppression of VEGF at both the transcriptional and translational levels(528)
25. modulates the expression of various **microRNAs**(529):
26. immunomodulation via impairing the immunosuppressive function of **regulatory T cells** and enhancing antitumor immune responses to dendritic cell-based immunization(530, 531), see (496)
27. inhibits tumor angiogenesis by inhibiting c-Kit and consequently downregulating HIF-1 α -mediated enhancement of VEGF expression(532)
28. causes **a transcriptional shift** toward alternative splicing in a large number of apoptotic genes(533)
29. exerts cytotoxic effects and modulates erythroid differentiation of CML cells by inducing endogenous **Spred2** expression(534, 535)
30. **binds DNA** and causes conformational changes in DNA double helix(536)
31. triggers apoptosis in part through the up-regulation of soluble histone **H2AX**(537)
32. induces a caspase-independent, necrosis-like programmed cell death via the serine protease activity of **Omi/HtrA2**(538), also see (539)
33. independent of its target, induces DNA damage which itself increases the activity of p53(540)
34. inhibits the expression of **SCO2** and **FRATAXIN** genes that encode mitochondrial proteins(541)
35. can help to direct natural immunity toward an anti-leukemic reactivity by acting on the bone marrow microenvironment(542), see (496)
36. directly inhibits CSF-1R(450)
37. The mechanism for the apoptosis-inducing effects of imatinib may be associated with the upregulation of **SHIP** and caspase-9 genes(543).
38. not only has a direct cytotoxic effect on bcr-abl gene rearranged cells but also an indirect effect associated with increased anti-leukemic immunological function due to an **intensified antigen presentation**(544), see (496)
39. directly binds these kinases (K_d (μM))(348):
- 1) ABL1: 0.0022
 - 2) ABL2: 0.013
 - 3) CLK1: 4.5

- 4) CLK4: 4.2
- 5) EPHA8: 2.1
- 6) FRK: 3.5
- 7) FYN: 5.5
- 8) GAK: 3.6
- 9) JNK1: 3.2
- 10) **JNK2**: 5.2
- 11) JNK3: 3.3
- 12) KIT: 0.83
- 13) LCK: 0.062
- 14) PDGFRB: 0.028
- 15) **STK17A**: 2.8
- 16) **STK18**: 9
40. prevents lung cancer metastasis by inhibiting M2-like polarization of macrophages via inhibiting **STAT6** phosphorylation and nuclear translocation(545)
41. may modulate metastasis and exert anti-cancer activity via upregulating **KAI1/CD82** gene expression(546)
42. enhances antigen-presenting cell function and overcomes tumor-induced CD4+ T-cell tolerance(547), see (496)
43. induces growth inhibition via production of spliced **osteocalcin-mRNA**(548)
44. modulates extracellular **ATP** catabolism and expression of **NTPDases** in a chronic myeloid leukemia cell line(549)
45. has the potential to exert its antileukemia effects by down-regulating **hERG1** K⁺ channels in chronic myelogenous leukemia, independent of targeting tyrosine kinase(550)
46. inhibits **glucose-6-phosphate dehydrogenase** and **glutathione reductase** which are important for redox homeostasis and play key roles in many cellular processes and represent an attractive approach to the development of anticancer agents(551)
47. It downregulates telomerase activity and inhibits proliferation in telomerase-expressing cell lines. This study demonstrates an additional cellular target of imatinib, not necessarily mediated via known tyrosine kinases(552).
48. has antiangiogenic activities like down-regulating **VE cadherin**, reducing the population of **endothelial cell** and reducing cell-to-cell cohesiveness, which possibly contribute to its therapeutic potential(553)
49. directly binds these kinases (K_d, nM) (NP: nonphosphorylated, P: phosphorylated)(324):
- 1) ABL1-NP: 1.1
 - ✓ ABL1-P: 21
 - ✓ ABL1(E255K)-P: 170
 - ✓ ABL1(F317I)-NP: 8.3
 - ✓ ABL1(F317I)-P: 580
 - ✓ ABL1(F317L)-NP: 2.5
 - ✓ ABL1(F317L)-P: 110
 - ✓ ABL1(H396P)-NP: 5.9
 - ✓ ABL1(H396P)-P: 65
 - ✓ ABL1(M351T)-P: 44
 - ✓ ABL1(Q252H)-NP: 1.8
 - ✓ ABL1(Q252H)-P: 92

- ✓ ABL1(Y253F)-P: 130
 - 2) ABL2: 10
 - 3) BLK: 520
 - 4) BRAF(V600E): 3300
 - 5) CDK11: 5500
 - 6) CLK1: 4500
 - 7) CLK4: 2100
 - 8) CSF1R: 11
 - 9) DDR1: 0.7
 - 10) DDR2: 15
 - 11) DRAK1: 5300
 - 12) EGFR(L747-E749del,A750P): 7600
 - 13) EPHA8: 1400
 - 14) FGR: 2400
 - 15) FLT3(ITD): 6300
 - 16) FRK: 1500
 - 17) FYN: 3100
 - 18) GAK: 1000
 - 19) HIPK4: 960
 - 20) IRAK1: 1200
 - 21) JNK1: 5000
 - 22) JNK3: 3100
 - 23) KIT: 13
 - ✓ KIT(A829P): 15
 - ✓ KIT(D816H): 560
 - ✓ KIT(D816V): 980
 - ✓ KIT(L576P): 14
 - ✓ KIT(V559D): 15
 - ✓ KIT(V559D,T670I): 2500
 - ✓ KIT(V559D,V654A): 91
 - 24) LCK: 40
 - 25) LYN: 890
 - 26) MELK: 1900
 - 27) PDGFRA: 31
 - 28) PDGFRB: 14
 - 29) PIP5K2C: 380
 - 30) PLK4: 7800
 - 31) RAF1: 1700
 - 32) TNNI3K: 4300
 - 33) TYK2(JH2domain-pseudokinase): 8700
 - 34) ZAK: 2600
- Dasatinib**
1. directly targets these proteins(521, 522, 554):
 - 1) ACK
 - 2) ARG
 - 3) BTK
 - 4) KIT
 - 5) PDGFR
 - 6) SRC(555, 556)
 - 7) YES
 - 8) FYN
 - 9) LYN(557, 558)
 - 10) HCK
 - 11) LCK
 - 12) FGR

13) BLK	41) HH498/TNNI3K
14) FRK	42) ILK
15) KHS2	43) LIMK1
16) CSK	44) LIMK2
17) RTK	45) MAP2K5
18) TEC	46) MAP3K1
19) BMX	47) MAP3K2
20) TXK	48) MAP3K3
21) DDR1(502)	49) MAP3K4
22) DDR2(502, 559)	50) MAP4K1
23) ACTR2B	51) MAP4K5/KHS1
24) ACVR2	52) MAPK11/p38 beta
25) BRAF	53) MAPK14/p38 alpha
26) EGFR/ERBB1(560)	54) MYT1(564)
27) EPHA2(561-563)	55) NLK
28) EPHA3	56) PTK6/Brk
29) EPHA4	57) QIK
30) EPHA5	58) QSK
31) EPHA8	59) RAF1
32) EPHB1	60) RET
33) EPHB2	61) RIPK2
34) EPHB4	62) SLK
35) EPHB6	63) STK36/ULK
36) ERBB2	64) SYK
37) ERBB4	65) TAO3
38) FAK	66) TESK2
39) GAK	67) TYK2/ZAK
40) GCK	68) ZAK

2. inhibits DDR1 and DDR2(502, 559)
3. reduces myeloid suppressor cells and releases effector lymphocyte responses(504)
4. transcriptional and post-translational inhibition of **telomerase**, independent of BRC/ABL(565)
5. **Angiogenesis/blood vessel-related pathways** and human vascular endothelial cell function (tube formation/viability) were adversely affected by dasatinib, ponatinib, and nilotinib but not by imatinib or bosutinib. These results correspond to the differences in vascular adverse event profiles of these tyrosine kinase inhibitors, imply a direct effect on vascular cells(566).
6. Btk tyrosine kinase is a major direct target of the Bcr-Abl inhibitor dasatinib(554)
7. reduces the expression of **CDK8** probably independent of BCR/ABL(567)
8. induces apoptosis through downregulating expression levels of antiapoptotic **SK-1** but not GCS, and upregulating expression levels of ceramide synthase (**CerS**) genes, especially CerS1(568)
9. inhibits FMS phosphorylation and cell proliferation(513)
10. **directly binds these kinases** (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):
 - 1) ABL1-NP: 0.029
 - ✓ ABL1-P: 0.046
 - ✓ ABL1(E255K)-P: 0.047
 - ✓ ABL1(F317I)-NP: 0.1
 - ✓ ABL1(F317I)-P: 0.041
 - ✓ ABL1(F317L)-NP: 0.032
 - ✓ ABL1(F317L)-P: 0.019
 - ✓ ABL1(H396P)-NP: 0.025
 - ✓ ABL1(H396P)-P: 0.046
 - ✓ ABL1(M351T)-P: 0.016
 - ✓ ABL1(Q252H)-NP: 0.037
 - ✓ ABL1(Q252H)-P: 0.064
 - ✓ ABL1(T315I)-NP: 890
 - ✓ ABL1(T315I)-P: 120
 - ✓ ABL1(Y253F)-P: 0.058
- 2) ABL2: 0.17
- 3) ACVR1: 620
- 4) ACVR1B: 330
- 5) ACVR2A: 210
- 6) ACVR2B: 570
- 7) ACVRL1: 460
- 8) ADCK3: 190
- 9) AURKA: 9300
- 10) BLK: 0.21
- 11) BMPR1A: 7000
- 12) BMPR1B: 53
- 13) BMX: 1.4
- 14) BRAF: 500
 - ✓ BRAF(V600E): 570
- 15) BRK: 7.8
- 16) BTK: 1.4
- 17) CSF1R: 0.58

- 18) CSK: 1
 19) CSNK1E: 1500
 20) DDR1: 0.69
 21) DDR2: 3.2
 22) DMPK: 1300
 23) DMPK2: 1200
 24) EGFR: 120
- ✓ EGFR(E746-A750del): 130
 - ✓ EGFR(G719C): 170
 - ✓ EGFR(G719S): 79
 - ✓ EGFR(L747-E749del, A750P): 110
 - ✓ EGFR(L747-S752del, P753S): 320
 - ✓ EGFR(L747-T751del,Sins): 160
 - ✓ EGFR(L858R): 120
 - ✓ EGFR(L858R,T790M): 2200
 - ✓ EGFR(L861Q): 110
 - ✓ EGFR(S752-I759del): 330
 - ✓ EGFR(T790M): 2300
- 25) EPHA1: 4.1
 26) EPHA2: 0.85
 27) EPHA3: 0.093
 28) EPHA4: 1.2
 29) EPHA5: 0.24
 30) EPHA6: 2100
 31) EPHA8: 0.24
 32) EPHB1: 0.45
 33) EPHB2: 0.39
 34) EPHB3: 6.9
- 35) EPHB4: 0.34
 36) EPHB6: 0.039
 37) ERBB2: 1400
 38) ERBB3: 18
 39) ERBB4: 55
 40) FGFR1: 3700
 41) FGFR2: 1400
 42) FGFR3: 3900
 43) FGR: 0.5
 44) FLT1: 5000
 45) FLT3: 4800
- ✓ FLT3(D835H): 8100
 - ✓ FLT3(D835Y): 4800
 - ✓ FLT3(ITD): 9900
 - ✓ FLT3(K663Q): 3200
 - ✓ FLT3(N841I): 7000
- 46) FRK: 0.31
 47) FYN: 0.79
 48) GAK: 2.6
 49) GCN2(Kin.Dom.2,S808G): 1600
 50) HCK: 0.35
 51) HPK1: 980
 52) JAK2(JH1domain-catalytic): 1000
 53) JAK3(JH1domain-catalytic): 640
 54) KIT: 0.81
- ✓ KIT(A829P): 0.66
 - ✓ KIT(D816H): 1.6
 - ✓ KIT(D816V): 2.6

- ✓ KIT(L576P): 0.57
- ✓ KIT(V559D): 0.68
- ✓ KIT(V559D,V654A): 2.7
- 55) LCK: 0.2
- 56) LIMK1: 570
- 57) LIMK2: 86
- 58) LOK: 1200
- 59) LYN: 0.57
- 60) LZK: 5300
- 61) MAP3K2: 140
- 62) MAP3K3: 280
- 63) MAP3K4: 310
- 64) MAP4K2: 1300
- 65) MAP4K3: 640
- 66) MAP4K4: 3100
- 67) MAP4K5: 45
- 68) MEK1: 1000
- 69) MEK2: 1400
- 70) MEK5: 3.3
- 71) MINK: 430
- 72) MRCKA: 2000
- 73) MRCKB: 2100
- 74) MST1: 3800
- 75) MST4: 1900
- 76) MYLK2: 3500
- 77) NEK11: 470
- 78) NEK2: 6500
- 79) NLK: 260
- 80) p38-alpha: 27
- 81) p38-beta: 410
- 82) PDGFRA: 0.47
- 83) PDGFRB: 0.63
- 84) PFCDPK1(*P. falciparum*): 640
- 85) PKMYT1: 130
- 86) QSK: 28
- 87) RAF1: 570
- 88) RET: 730
- ✓ RET(M918T): 390
- ✓ RET(V804L): 3200
- 89) RIPK2: 31
- 90) SBK1: 1200
- 91) SIK: 3.9
- 92) SIK2: 6.4
- 93) SLK: 720
- 94) SRC: 0.21
- 95) SRMS: 13
- 96) STK35: 770
- 97) STK36: 210
- 98) SYK: 2900
- 99) TAK1: 3700
- 100) TAOK2: 5400
- 101) TAOK3: 2300
- 102) TEC: 13
- 103) TESK1: 33
- 104) TGFBR1: 230
- 105) TGFBR2: 2900

- 106) TNIK: 2000
- 107) TNK2: 5.6
- 108) TNNI3K: 11
- 109) TXK: 2.1
- 110) TYK2(JH1domain-catalytic): 1800
 ✓ TYK2(JH2domain-pseudokinase):
 110
- 111) ULK3: 4600
- 112) VEGFR2: 2900
- 113) VRK2: 3200
- 114) WEE1: 7000
- 115) WEE2: 200
- 116) YES: 0.3
- 117) YSK1: 3400
- 118) YSK4: 79
- 119) ZAK: 45
11. an immunological component of its anti-tumoral “off-target” effect is that it also influences **non-conventional T-αβ cell** subsets known for their potential anti-tumoral properties, namely iNKT cells and the distinct new innate CD8 T-cell subset(569)
12. targets several off-target proteins and peptide in tumor environment(570)
13. directly inhibits EphA2 in a dose-dependent manner and may exert anticancer effects due to EphA2 inhibition, besides its effects on Src(561-563)
14. increases the activation of allogenic T cells via modulating dendritic cell activity via confining c-Kit signaling and IDO-mediated **tryptophan metabolism**(571)
15. directly binds 40 different kinases (identified reproducibly in three human cell lines using affinity chromatography approach)(572):
- 1) ABL1
 - 2) ABL2 isoform IB
 ✓ ABL2 isoform 4
 - 3) ACVR1
 - 4) ACVR1B
 - 5) CSK
 - 6) DDR1
 - 7) EGFR(560)
 - 8) EPHA1
 - 9) EPHA2(561-563)
 - 10) EPHA4
 - 11) EPHA5
 - 12) EPHA8
 - 13) EPHB1
 - 14) EPHB2
 - 15) EPHB3
 - 16) EPHB4
 - 17) FRK
 - 18) FYN
 - 19) GAK
 - 20) ILK**
 - 21) QSK
 - 22) LCK
 - 23) LIMK2
 - 24) LYK5**

- 25) LYN isoform A
- ✓ LYN isoform B(557, 558)
- 26) MAP2K5
- 27) MAP3K1
- 28) MAP3K2
- 29) MAP4K5
- 30) MAPK14 isoform1
- ✓ MAPK14 isoform2
- 31) PRO3078**
- 32) BRK
- 33) RICK**
- 34) SIK
- 35) QIK**
- 36) SRC
- 37) TEC
- 38) TESK1
- 39) TGFBR1
- 40) TNK2(573)
- 41) TYK2
- 42) YES1
- 43) ZAK isoform 1
- ✓ ZAK isoform 2
16. has immunostimulatory effects in the form of persistent monoclonal/oligoclonal LGL lymphocytosis(574-577), at least partly through directly stimulating the **proliferation of natural killer cells**(578)
17. promotes cell death by targeting c-KIT(579)
18. is associated with a rapid mobilization of cytotoxic lymphocytes through direct effect on signal transduction pathways(580, 581)
19. exerts antitumor effects via directly inhibiting TNK2(572, 573)
20. induces apoptosis through degradation of EGFR(582)
21. can inhibit **AR Tyr267 and AR Tyr 534** phosphorylation, and it may play a significant role in anti-prostate cancer cells by inhibiting Ack1-mediated AR Tyr-267 phosphorylation and the expression of PSA mRNA and hk2 mRNA induced by heregulin(583)
22. enhances the activity of immune system, particularly via natural killer cell differentiation through inhibition of regulatory T cells(584)
23. enhances T-cell responses and interleukin 12 (IL-12) production via enhancing the stimulatory activity of dendritic cells(585)
24. exerts antitumor effects via decreasing levels of T regulatory cells while specifically enhancing tumor antigen-specific T cell responses(586)
25. can induce **pyroptosis** in tumor cells and increase the protein levels of GSDMD and GSDME (gasdermin E) in a p53-independent manner(587)
26. directly binds and inhibits MEKK2(324, 471)
27. directly binds and targets these kinases(588):
- 1) **AXL**
 - 2) c-KIT

✓ c-KIT(D816 V)

✓ c-KIT(D816 H)

3) c-RAF

4) EGFR

✓ EGFR(T790M)

5) FES

6) IRAK4

7) LIMK1

8) PDGFR α

9) RIPK2

10) TIE2

11) TRKA

12) TRKB

13) TXK

Nilotinib

1. induces apoptosis through **Bim** accumulation independently of cell cycle arrest(498)

2. inhibits DDR1(589) and DDR2(502)

3. **Angiogenesis/blood vessel**-related pathways and human vascular endothelial cell function (tube formation/viability) were adversely affected by dasatinib, ponatinib, and nilotinib but not by imatinib or bosutinib. These results correspond to the differences in vascular adverse event profiles of these tyrosine kinase inhibitors, imply a direct effect on vascular cells(566).

4. Directly targets these proteins(521, 522):

1) ARG

2) KIT

3) PDGFR(590)

4) DDR1

5) NQO2

5. transcriptional and post-translational inhibition of **telomerase**, independent of BRC/ABL(565)

6. induces apoptosis through upregulating **ceramide synthase** genes and downregulating **SK-1** in CML cells in addition to inhibition of BCR/ABL(591)

7. in addition to its known activity against several tyrosine-kinase-mediated proliferative pathways, exerts antitumor effects via directly binding receptor Smoothened (**SMO**) and direct inhibition of the Hedgehog pathway(592)

8. induces autophagic cell death through deactivating phosphatase **PP2A** and subsequently increasing AMPK phosphorylation(593)

9. can exert antitumor effects in not only Philadelphia-positive but also Philadelphia-negative acute lymphoblastic leukemia via inhibiting **MDM2** and inducing a p53-independent apoptosis pathway by downregulating XIAP(594)

10. **directly binds these kinases** (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):

1) ABL1-NP: 10

✓ ABL1-P: 13

✓ ABL1(E255K)-P: 36

✓ ABL1(F317I)-NP: 18

✓ ABL1(F317I)-P: 56

✓ ABL1(F317L)-NP: 12

- ✓ ABL1(F317L)-P: 14
- ✓ ABL1(H396P)-NP: 4.9
- ✓ ABL1(H396P)-P: 21
- ✓ ABL1(M351T)-P: 15
- ✓ ABL1(Q252H)-NP: 10
- ✓ ABL1(Q252H)-P: 21
- ✓ ABL1(T315I)-NP: 660
- ✓ ABL1(Y253F)-P: 13
- 2) ABL2: 26
- 3) BLK: 500
- 4) BRAF: 1700
 - ✓ BRAF(V600E): 570
- 5) CLK1: 2100
- 6) CLK4: 7100
- 7) CSF1R: 45
- 8) CSK: 2400
- 9) DDR1: 1.1
- 10) DDR2: 33
- 11) EPHA1: 590
- 12) EPHA2: 230
- 13) EPHA3: 110
- 14) EPHA4: 330
- 15) EPHA5: 1900
- 16) EPHA6: 640
- 17) EPHA8: 37
- 18) EPHB1: 1300
- 19) EPHB2: 640
- 20) EPHB3: 1000
- 21) EPHB4: 730
- 22) EPHB6: 500
- 23) FGR: 320
- 24) FRK: 86
- 25) FYN: 1600
- 26) HCK: 390
- 27) HPK1: 890
- 28) JNK1: 450
- 29) JNK2: 5800
- 30) JNK3: 2000
- 31) KIT: 29
 - ✓ KIT(A829P): 46
 - ✓ KIT(D816H): 540
 - ✓ KIT(D816V): 770
 - ✓ KIT(L576P): 22
 - ✓ KIT(V559D): 46
 - ✓ KIT(V559D,T670I): 150
 - ✓ KIT(V559D,V654A): 260
- 32) LCK: 47
- 33) LOK: 6800
- 34) LYN: 100
- 35) MEK5: 190
- 36) MRCKB: 910
- 37) p38-alpha: 460
- 38) p38-beta: 36
- 39) PDGFRA: 180
- 40) PDGFRB: 73
- 41) PFCDPK1(*P. falciparum*): 790

- 42) PIP5K2C: 8800
- 43) RAF1: 3900
- 44) RET: 870
- ✓ RET(M918T): 1100
- 45) SRC: 1900
- 46) TAOK1: 4700
- 47) TAOK3: 1700
- 48) TIE1: 1000
- 49) TIE2: 1000
- 50) TNNI3K: 360
- 51) TRKB: 490
- 52) TRKC: 600
- 53) YES: 1100
- 54) ZAK: 11

Ponatinib

1. inhibits **DDR1** and **DDR2**(501, 502)
2. Angiogenesis/blood vessel-related pathways and human vascular endothelial cell function (tube formation/viability) were adversely affected by dasatinib, ponatinib, and nilotinib but not by imatinib or bosutinib. These differences in vascular adverse event profiles of these tyrosine kinase inhibitors, imply a direct effect on vascular cells(566).
3. modulates the expression of various **microRNAs**(529):
4. directly binds and inhibits **MEKK2** with an IC₅₀ of 16 ± 3(595)
5. exerts antiangiogenic effects via blocking **VEGFR** signaling at its receptor level and downstream pathways(596)

6. directly inhibits **RIPK2**(490)
7. exerts antitumor effects via inhibiting various **FGFRs**(597)
8. inhibits the phosphorylation of(598):
 - 1) **RYK**
 - 2) **CSF1R**
 - 3) **ALK**
 - 4) **TEK**
9. Reduces endothelial survival and angiogenesis and induces their senescence and apoptosis via **Notch-1** pathway(599)
10. has **microtubule stabilizing** activity(600)

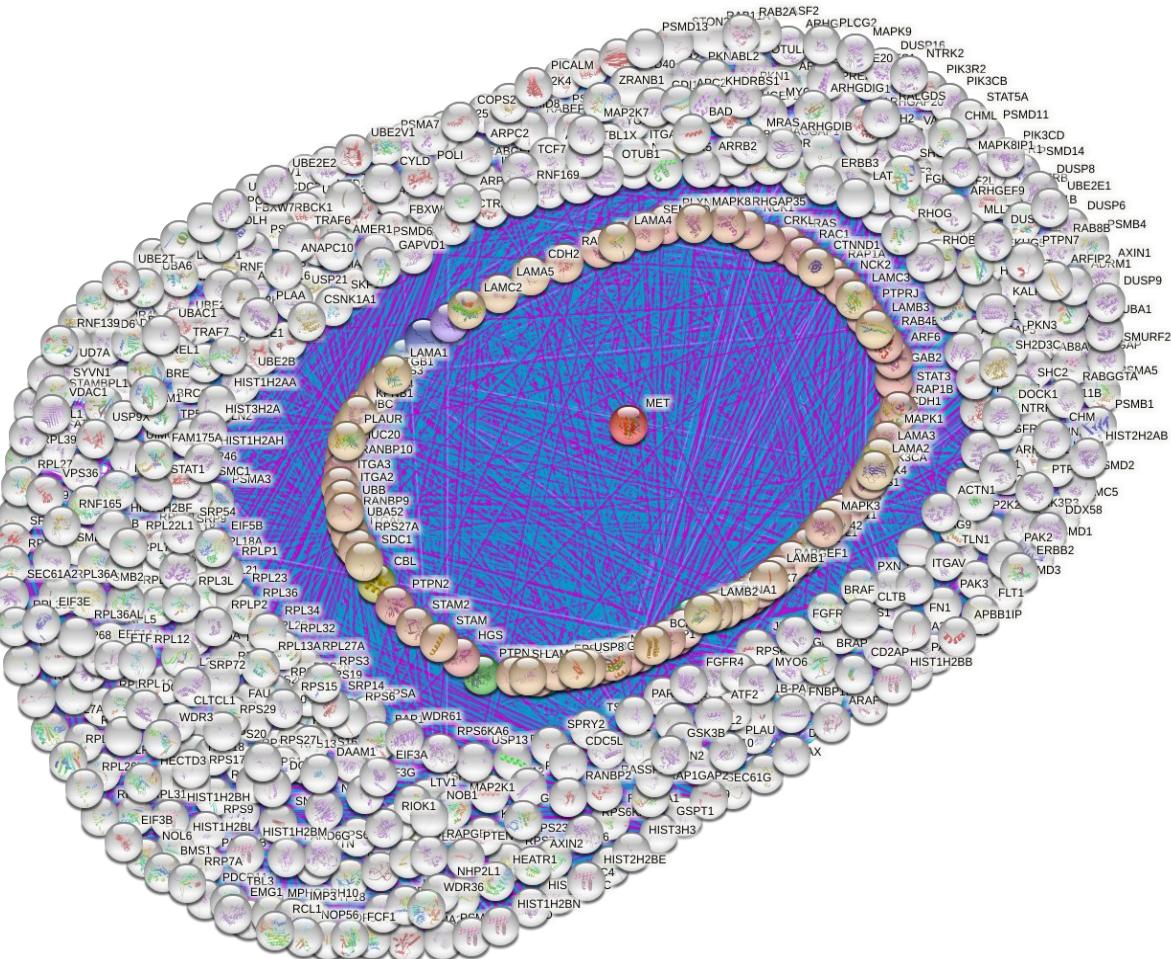
Bosutinib

1. targets about 30 proteins in tumor environments(570)
2. directly binds and inhibits MEKK2 with an IC₅₀ of 59 nM(471, 595)
3. directly inhibits these proteins(521):
 - 1) **ACK**
 - 2) **ALK4**
 - 3) **ARG**
 - 4) **BCR-ABL**
 - 5) **BTK**
 - 6) **CaMKK2**
 - 7) **CK1d**
 - 8) **CK1e**
 - 9) **CLK2**
 - 10) **CSK**
 - 11) **DDR1**
 - 12) **EphB1**

- | | |
|----------------|--|
| 13) EphB4 | 41) Wee1 |
| 14) FAK | 42) YES |
| 15) FER | 43) ZAK |
| 16) FRK | 44) ZC2_TNIK |
| 17) FYN | 4. directly binds and prevents auto-phosphorylation of ACK1(588) and attenuates migration and invasion of tumors via inhibition of ACK1(601) |
| 18) GAK | |
| 19) GCK | 5. directly binds these kinases(588): |
| 20) HCK | 1) MAP4K5 (KHS) |
| 21) HRI | 2) YES1 |
| 22) KHS2 | 3) SRC |
| 23) LYN | 4) FGR |
| 24) MAP3K1 | 5) LCK |
| 25) MAP3K2 | 6) FYN |
| 26) MAP3K3 | 7) FRK (PTK5) |
| 27) MAP3K4 | 8) BTK |
| 28) MER | 9) TNK2 (ACK1) |
| 29) MYT1 | 10) HCK |
| 30) NLK | 11) STK24 (MST3) |
| 31) PKC τ | 12) EPHB4 |
| 32) PYK2 | 13) BMX |
| 33) QIK | 14) LYN (A and B) |
| 34) QSK | 15) MAP4K2 (GCK) |
| 35) RSK2 | 16) MINK1 |
| 36) SLK | 17) STK10 (LOK) |
| 37) SRC | 18) SYK |
| 38) SYK | 19) CSK |
| 39) TBK1 | 20) CAMK1D |
| 40) TEC | |

- | | |
|------------------------|--------------------------|
| 21) PTK2B (FAK2, PYK2) | 36) ZAK (Isoforms 1 & 2) |
| 22) CAMK2G | 37) STK35 (CLIK1) |
| 23) STK4 (MST1) | 38) MAPK14 (p38a) |
| 24) TEC | 39) BMP2K (BIKE) |
| 25) TBK1 | 40) ILK |
| 26) FER | 41) STK25 (YSK1) |
| 27) PTK2 (FAK) | 42) TNIK |
| 28) TAOK3 (JIK) | 43) SLK |
| 29) PRKAA1 (AMPK) | 44) MAP4K4 (HGK, NIK) |
| 30) MAP2K1 (MEK1) | 45) MAP3K1 |
| 31) MST4 (MASK) | 46) GAK |
| 32) MAP2K2 (MEK2) | 47) MAP4K1 (HPK1) |
| 33) AAK1 | 48) PKMYT1 |
| 34) MAP3K2 | 49) MAP2K5 |
| 35) MAP3K3 | 50) MAP3K |

Drugs	Discovery Target	Uniprot "Off-Target" Therapeutic ID	Mechanisms
CRIZOTINIB			148
CABOZANTINIB			5
LAROTRECTINIB LORLATINIB	MET	P08581	
CERITINIB BRIGATINIB			9
ENTRECTINIB CAPMATINIB			



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Crizotinib

1. “Shortly after in 2007, anaplastic lymphoma kinase (ALK) was fortuitously identified as a drug target in NSCLC. Initially, the industry researchers who developed crizotinib, the first-in-class oral ALK TKI, were searching for a mesenchymal–epithelial transition factor (MET) inhibitor.(602)”
2. directly binds and inhibits MEKK2 with an IC₅₀ of 75 ± 35 nM(471)
3. reduces gastric cancer growth through oxidative DNA damage and triggers pro-survival Akt signal(603)
4. induces apoptosis via lethal endoplasmic reticulum stress response in cancer cells by increasing intracellular levels of reactive oxygen species, independent of MTH1 inhibition(604, 605)
5. induces apoptosis via downregulation of BCL-2 family proteins including MCL-1(606)
6. stimulates antitumor immune responses via inducing Immunogenic cell death(607)
7. Crizotinib-induced antitumor activity is not solely dependent on ALK and MET inhibition(605).
8. Exerts antitumor effects via directly inhibiting FAK1 (PTK2)(608)
9. suppresses the growth of aggressive thyroid cancer cells, and this potential therapeutic benefit results from their non-MET-targeting effects(609)
10. reduces tumor burden and metastasis via reducing the phosphorylation of Akt and the activity of matrix metalloproteinases and adhesion to various extracellular matrices(610)
11. directly binds these kinases (K_d, nM) (NP: nonphosphorylated, P: phosphorylated)(324):
 - 1) AAK1: 2300
 - 2) ABL1-NP: 78
 - ✓ ABL1-P: 110
 - ✓ ABL1(E255K)-P: 170
 - ✓ ABL1(F317I)-NP: 2700
 - ✓ ABL1(F317I)-P: 3200
 - ✓ ABL1(F317L)-NP: 610
 - ✓ ABL1(F317L)-P: 590
 - ✓ ABL1(H396P)-NP: 33
 - ✓ ABL1(H396P)-P: 87
 - ✓ ABL1(M351T)-P: 97
 - ✓ ABL1(Q252H)-NP: 97
 - ✓ ABL1(Q252H)-P: 100
 - ✓ ABL1(T315I)-NP: 12
 - ✓ ABL1(T315I)-P: 10
 - ✓ ABL1(Y253F)-P: 74
 - 3) ABL2: 460
 - 4) ACVR1: 440
 - 5) ACVR1B: 860
 - 6) ADCK4: 1100
 - 7) ALK: 3.3
 - 8) AMPK-alpha1: 2400
 - 9) ANKK1: 780
 - 10) AURKA: 260

- | | |
|-----------------------|----------------------|
| 11) AURKB: 76 | 38) EPHA6: 65 |
| 12) AURKC: 4300 | 39) EPHA7: 470 |
| 13) AXL: 7.8 | 40) EPHA8: 280 |
| 14) BIKE: 740 | 41) EPHB1: 120 |
| 15) BLK: 110 | 42) EPHB4: 570 |
| 16) BMPR1B: 230 | 43) EPHB6: 6 |
| 17) BMX: 3600 | 44) ERK5: 510 |
| 18) BTK: 7800 | 45) FAK: 310 |
| 19) CAMKK2: 1500 | 46) FER: 270 |
| 20) CASK: 140 | 47) FES: 450 |
| 21) CDC2L1: 760 | 48) FGFR3: 2700 |
| 22) CDC2L2: 420 | ✓ FGFR3(G697C): 3500 |
| 23) CDK7: 330 | 49) FGR: 670 |
| 24) CDKL2: 2100 | 50) FLT1: 2300 |
| 25) CSF1R: 210 | 51) FLT3: 850 |
| 26) DCAMKL1: 330 | ✓ FLT3(D835H): 500 |
| 27) DCAMKL2: 370 | ✓ FLT3(D835Y): 210 |
| 28) DDR1: 510 | ✓ FLT3(ITD): 730 |
| 29) DLK: 170 | ✓ FLT3(K663Q): 900 |
| 30) DMPK: 1400 | ✓ FLT3(N841I): 860 |
| 31) DYRK2: 4000 | ✓ FLT3(R834Q): 3000 |
| 32) EGFR(G719C): 2700 | 52) FRK: 2900 |
| ✓ EGFR(L861Q): 2700 | 53) FYN: 1300 |
| 33) EPHA1: 140 | 54) GRK4: 4200 |
| 34) EPHA2: 100 | 55) HIPK4: 3600 |
| 35) EPHA3: 700 | 56) HPK1: 39 |
| 36) EPHA4: 360 | 57) IGF1R: 780 |
| 37) EPHA5: 1000 | 58) IKK-beta: 5500 |

- 59) IKK-epsilon: 710
 60) INSR: 340
 61) INSRR: 600
 62) IRAK1: 49
 63) IRAK3: 31
 64) IRAK4: 3600
 65) ITK: 2000
 66) JAK1(JH1domain-catalytic): 330
 67) JAK2(JH1domain-catalytic): 290
 68) JAK3(JH1domain-catalytic): 200
 69) KIT(D816V): 3700
 ✓ KIT(V559D,T670I): 800
 70) LCK: 30
 71) LIMK1: 830
 72) LIMK2: 690
 73) LOK: 44
 74) LTK: 12
 75) LYN: 940
 76) LZK: 230
 77) MAP3K1: 1100
 78) MAP3K15: 2800
 79) MAP3K2: 72
 80) MAP3K3: 110
 81) MAP4K2: 80
 82) MAP4K3: 75
 83) MAP4K5: 79
 84) MEK3: 3300
 85) MERTK: 3.6
 86) MET: 2.1
 ✓ MET(M1250T): 0.55
 ✓ MET(Y1235D): 1.5
 87) MINK: 4400
 88) MST1: 580
 89) MST1R: 25
 90) MST2: 990
 91) MST4: 2800
 92) MUSK: 230
 93) NEK7: 5700
 94) NEK9: 790
 95) NIM1: 4600
 96) PIP5K1A: 3200
 97) PLK4: 60
 98) PRKD1: 990
 99) PRKD3: 510
 100) PRKR: 1100
 101) PYK2: 190
 102) RET(M918T): 1300
 ✓ RET(V804L): 2100
 103) RIOK1: 6100
 104) RIOK3: 4000
 105) RIPK1: 1600
 106) RIPK2: 900
 107) RIPK4: 2100
 108) RIPK5: 450
 109) ROCK1: 3700
 110) ROCK2: 3300

- 111) ROS1: 4.1
 112) S6K1: 640
 113) SBK1: 1300
 114) SgK110: 240
 115) SIK2: 200
 116) SLK: 18
 117) SNARK: 120
 118) SRC: 560
 119) SRPK1: 1800
 120) SRPK3: 8200
 121) STK35: 610
 122) SYK: 2500
 123) TAK1: 1800
 124) TAOK1: 1200
 125) TAOK2: 900
 126) TAOK3: 490
 127) TBK1: 690
 128) TESK1: 380
 129) TGFBR1: 1000
 130) TIE1: 110
 131) TIE2: 270
 132) TNK1: 320
 133) TNK2: 760
 134) TNNI3K: 110
 135) TRKA: 95
 136) TRKB: 37
 137) TRKC: 82
 138) TXK: 850
- 139) TYK2(JH1domain-catalytic): 210
 ✓ TYK2(JH2domain-pseudokinase): 2000
 140) TYRO3: 800
 141) ULK1: 1000
 142) ULK2: 1600
 143) ULK3: 6700
 144) YES: 770
 145) YSK4: 980
 146) ZAP70: 4200

Cabozantinib

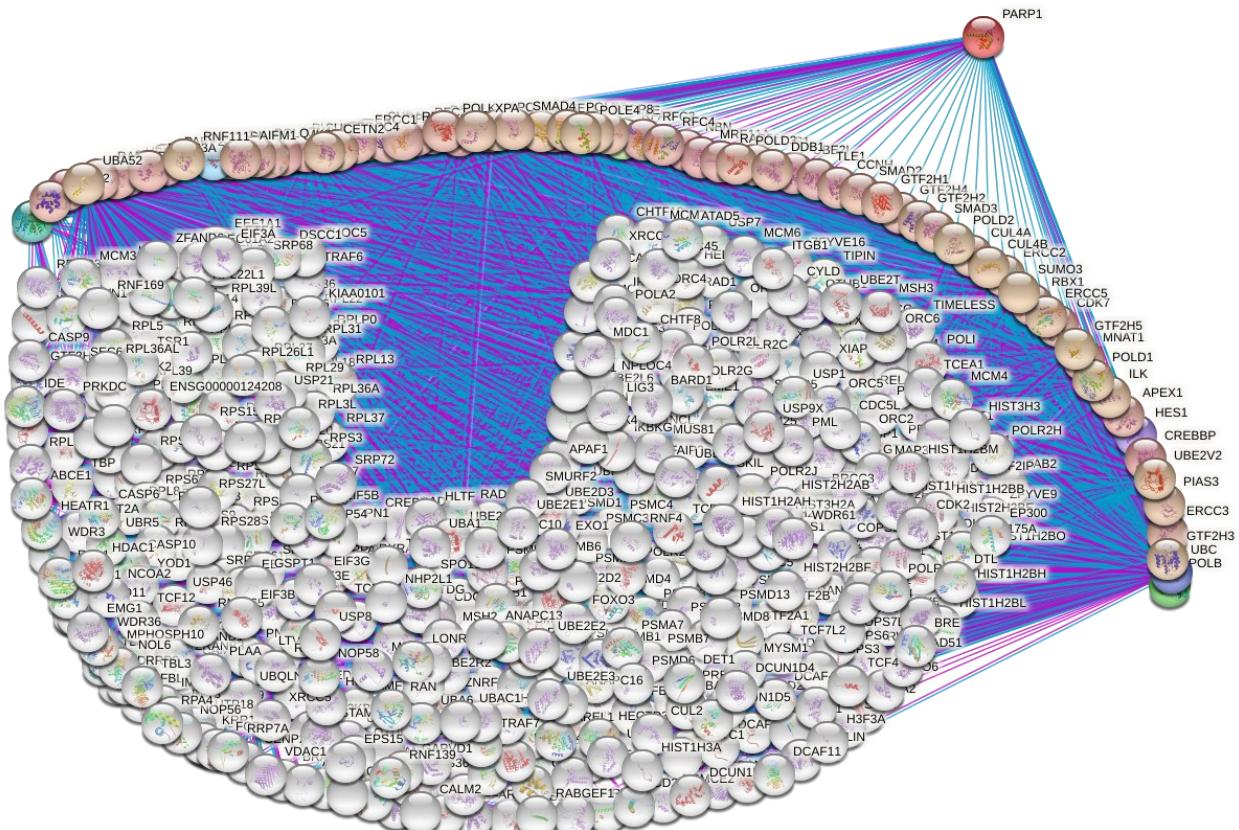
1. targets a broad range of targets, including(611):
 - 1) MET
 - 2) RET
 - 3) AXL
 - 4) VEGFR2
 - 5) FLT3
 - 6) c-KIT
2. inhibits cancer cell migration and invasion via inhibition of GAS6-AXL pathway independent of HGF-MET pathway(612)
3. exerts antitumoral and antimigratory effects which are most probably off-target effects, not mediated by c-Met(613)

Ceritinib

1. suppresses mTORC1 signaling in the presence of trametinib through direct inhibition of IGF1R and/or ACK1(614)
2. has activity even in ALK-negative cancer cell lines via off-targets like(615):

- 1) IGF1R
- 2) FAK1 (encoded by *PTK2*)
- 3) RSK1/2 (encoded by *RPS6KA1/3*)
- 4) ERK1/2
- 5) CAMKK2
- 6) FER
- 7) CAMKK2
- 8) AMPK α 1

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
OLAPARIB			4
RUCAPARIB		P09874	17
NIRAPARIB	poly(ADP-ribose) polymerase 1	PARP1	2
TALAZOPARIB			3



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Olaparib

1. induces upregulation of death receptors in primary acute myeloid leukemia blasts by NF- κ B activation(616)
2. inhibits the PI3K/AKT/mTOR signaling pathway(617)
3. induces cellular senescence through P16-RB/P53-RB signaling pathway(618)
4. traps PARP1 and PARP2 to the sites of DNA damage and prevents DNA repair, replication, and transcription, leading to cell death(619)

Rucaparib

1. micromolar affinities (IC₅₀ values) for nine protein kinases(620):
 - 1) PIM1 (1.2 μ M)
 - 2) PIM2 (7.7 μ M)
 - 3) PRKD2 (9.7 μ M)
 - 4) DYRK1A (1.4 μ M)
 - 5) CDK1 (1.4 μ M)
 - 6) CDK9 (2.7 μ M)
 - 7) HIPK2 (4.4 μ M)
 - 8) CK2 (7.8 μ M)
 - 9) ALK (18 μ M)
2. inhibits hexose-6-phosphate dehydrogenase(621)
3. induces vasodilation and potentiates response antitumor effects of other drugs through a complex process that involves myosin light chain kinase, P2 receptors, and PARP itself(622)
4. suppresses the lactate dehydrogenase pathway(623)

5. activates necrotic apoptosis via inducing the production of excessive reactive oxygen species and upregulating the expression of RIP1 and RIP3(624)
6. traps PARP1 and PARP2 to the sites of DNA damage and prevents DNA repair, replication, and transcription, leading to cell death(619)

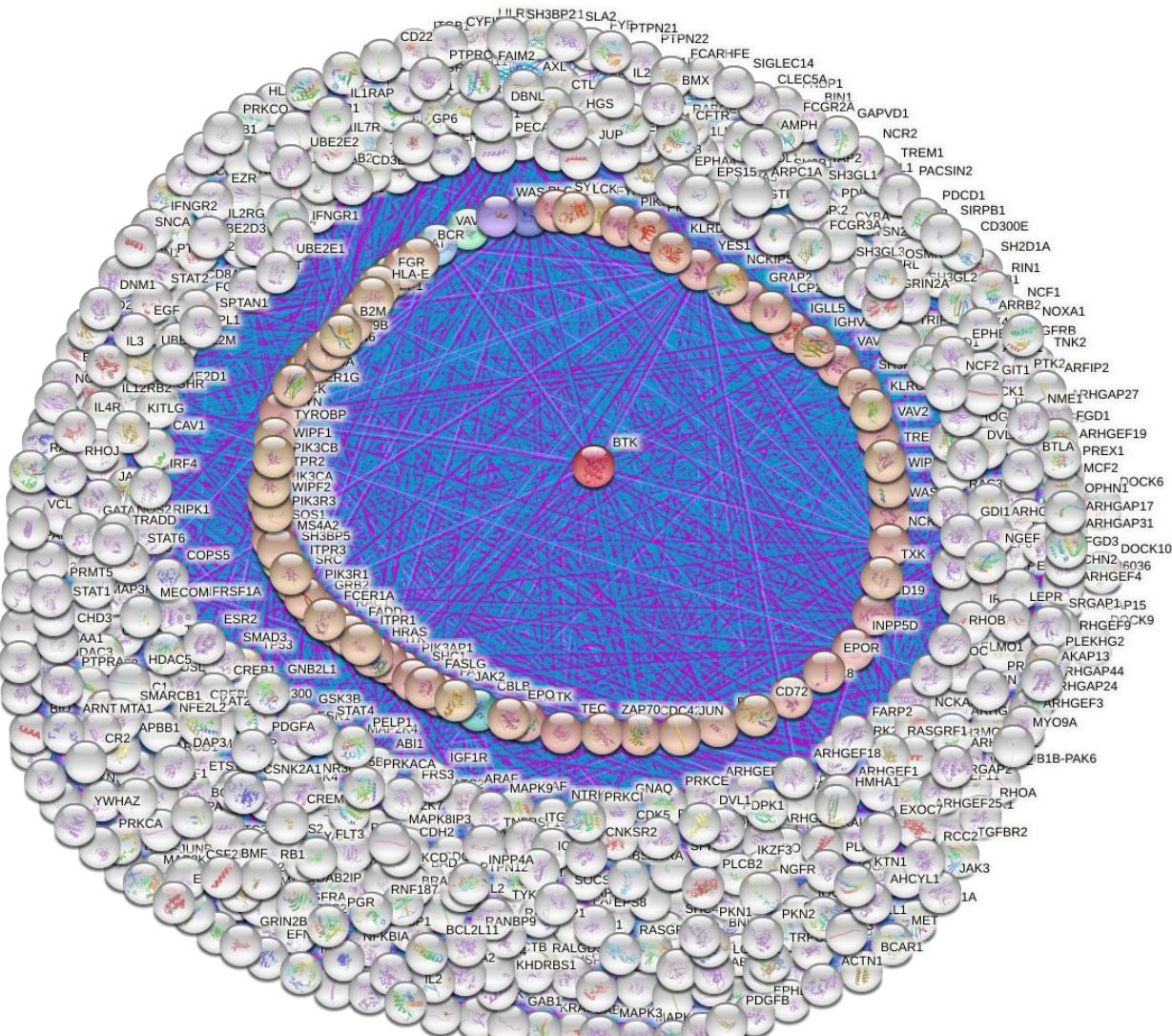
Niraparib

1. inhibits deoxycytidine kinase and reduces cytarabine activity(621)
2. traps PARP1 and PARP2 to the sites of DNA damage and prevents DNA repair, replication, and transcription, leading to cell death(619)

Talazoparib

1. induces not only cancer cell-intrinsic apoptosis but also cancer cell-extrinsic antitumor immune effects via increasing the number of peritoneal CD8⁺ T cells and NK cells as well as their production of IFN- γ and TNF- α (625)
2. G2/M cell cycle arrest via the upregulation of p53(626)
3. traps PARP1 and PARP2 to the sites of DNA damage and prevents DNA repair, replication, and transcription, leading to cell death(619)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
IBRUTINIB			40
ACALABRUTINIB	Bruton tyrosine kinase	BTK	Q06187
ZANUBRUTINIB			3



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Ibrutinib

1. Inhibits VEGFR2(627)
2. Inhibits Janus Kinase 2 (JAK2)(628)
3. also binds other kinases that are relevant to antitumor effects (IC₅₀ in nM)(629):
 - 1) TEC (78)
 - 2) ITK (10.7)
 - 3) BMX (0.8)
 - 4) BLK (0.5)
 - 5) EGFR (5.6)
 - 6) ErbB2/HER2 (9.4)
 - 7) JAK3 (16.1)
4. inhibits ERBB receptor tyrosine kinases and tumor growth(630)
5. inhibits IL-2-inducible kinase (ITK)(630-632)
6. immunomodulation via modifying the function of monocyte/macrophage populations(633)
7. causes cellular reactive oxygen species elevation and induces cancer cell apoptosis through irreversible inhibition of mammalian thioredoxin reductase enzymes, aside from its therapeutic mechanism through BTK inhibition(634)
8. both BTK and B lymphocyte kinase (BLK) are relevant targets of ibrutinib(635)
9. inhibits Wnt signaling pathway(636)
10. activates endoplasmic reticulum stress-induced cell death(637)
11. inhibits LCK and SRC phosphorylation in normal T cells(638)
12. increases the in vivo persistence of activated T cells, decreases the Treg/CD4⁺ T cell ratio, and diminishes the immune-suppressive properties of CLL cells through BTK-dependent and -independent mechanisms(639)
13. targets microRNA-21 in multiple myeloma cells by inhibiting NF-κB and STAT3(640)
14. downregulates activation-induced cytidine deaminase (AID) enzyme and proliferative fractions in chronic lymphocytic leukemia(641)
15. induces rapid down-regulation of inflammatory markers and altered transcription of chronic lymphocytic leukemia-related genes in blood and lymph nodes(642)
16. exerts immunomodulatory effects through regulation of tumor infiltrating macrophages(643)
17. induces changes in nuclear morphology and causes apoptosis via caspase-dependent extrinsic apoptosis pathway with the activation of caspases-8, caspase-3, and cleavage of PARP1(644)
18. inhibits ERBB4 activity in the same nM range as its canonical target, BTK, and consequently reduces cell growth in a WNT5A-dependent manner(645)
19. inhibits free fatty acid metabolism in chronic lymphocytic leukemia(646)
20. inhibits wild and mutant-EGFR kinase(647, 648)
21. increases apoptosis and G1-S cell-cycle delay via inhibiting EGFR, HER2, ErbB3, and ErbB4 and consequently inhibiting AKT and ERK signaling pathways(648)

22. downregulates a subset of miRNAs leading to upregulation of tumor suppressors and inhibition of cell proliferation in chronic lymphocytic leukemia(649)

- 1) miR-22
- 2) miR-34a
- 3) miR-146b
- 4) miR-181b

23. blocks immunophenotypic changes associated with the adhesion and activation of CLL cells in the tumor microenvironment through changing the expression of these 11 antigens(650):

- 1) CD23
- 2) CD27
- 3) CD53
- 4) CD58
- 5) CD71
- 6) CD80
- 7) CD84
- 8) CD97
- 9) CD126
- 10) CD150
- 11) FMC7

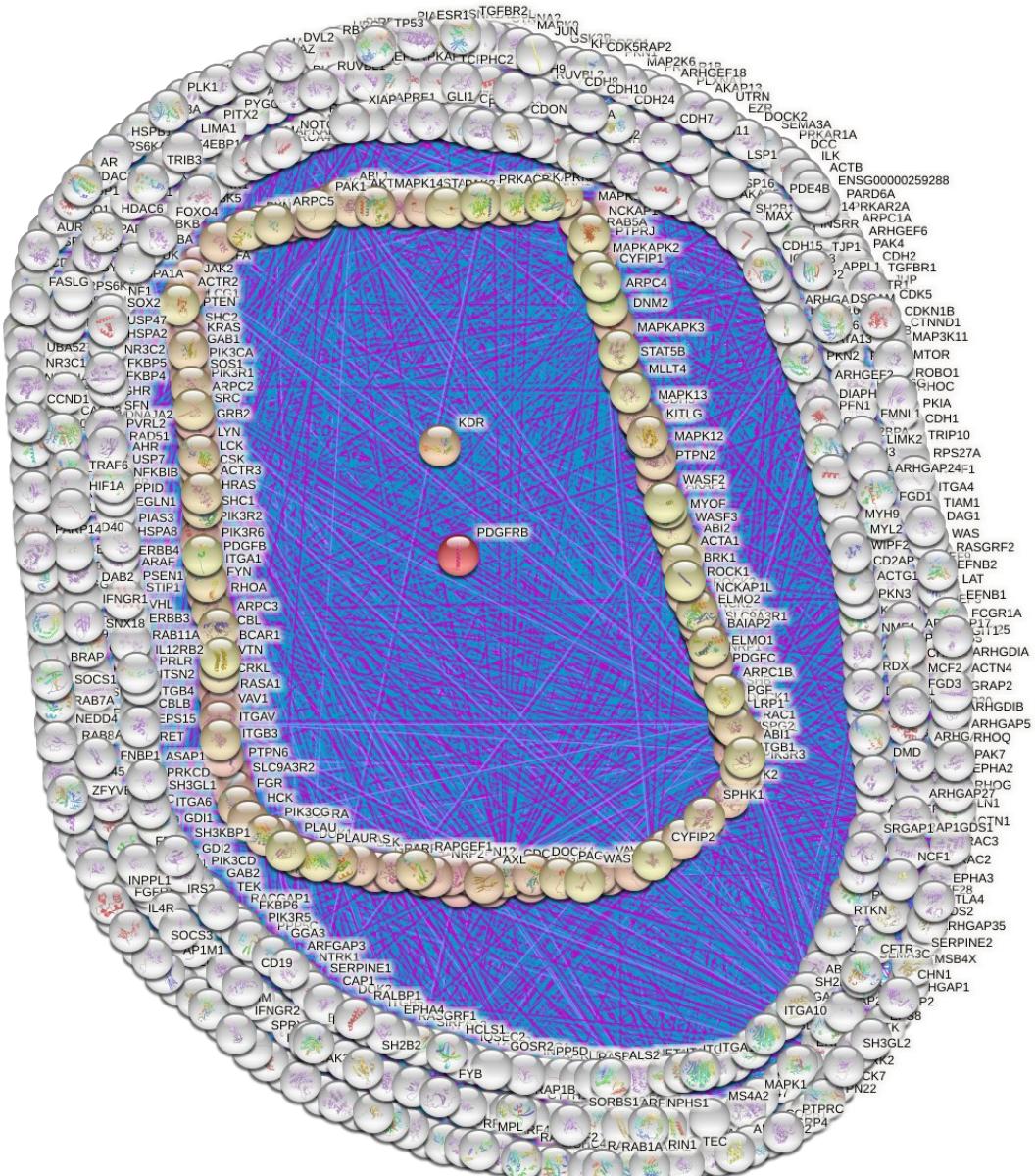
24. hematopoietic cell kinase (HCK) is a survival determinant transactivated by mutated MYD88, and a direct target of ibrutinib(651)

25. changes the quantity of about 1000 unique proteins, with nearly 400 significant changes (p-value < 0.05)(652)

Acalabrutinib

1. reduces expression of the immunosuppressive molecules CD200 and BTLA as well as IL-10 production by CLL cells(639)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
SUNITINIB	kinase insert domain receptor	KDR	P35968
	platelet derived growth factor receptor beta	PDGFR β	P09619 270



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1. inhibits the autophosphorylation of the receptor tyrosine kinase fms-like tyrosine kinase 3 (FLT3) in a dose-dependent manner and reduces survival and migration of human meningioma cells(653)
2. Sunitinib but not VEGF blockade inhibits cancer stem cell endothelial differentiation, suggesting a VEGF-independent mechanism(654).
3. inhibits other tyrosine kinases including, KIT, FLT3, colony-stimulating factor 1 (CSF-1), and RET which are associated with neoplasms(450, 655)
4. inhibits autophagy independent of AMPK signaling and through **SQSTM1/p62**(656)
5. inhibits the migration and invasion of MCF-7 through downregulation of the interaction between **furin** and its substrates (pro-MT1-MMP, pro-VEGF-C)(657)
6. concentration-dependent induction of oxidative stress genes (heme oxygenase 1 and glutathione transferase A1) through the **nuclear factor erythroid 2-related factor 2** pathway(658)
7. directly inhibits the activity of mammalian 5'-AMP-activated protein kinase (AMPK)(659)
8. directly induces cell death in renal cancer cells and simultaneously affect the expression levels of their apoptosis-related microRNAs, **miR-145**, **miR-15a** and **miR-16**, upon this process(660)
9. directly targets **vascular endothelial-cadherin** and inhibits phosphorylation which is induced by VEGF(661)
10. directly binds and inhibits MEKK2(324, 471)
11. induces apoptosis in colon cancer cells via **PUMA**(662)
12. inhibits hemangioma cell growth and migration by suppressing focal adhesion kinase signaling(663)
13. directly binds these kinases (K_d (μM))(348):
 - 1) AAK1: 0.13
 - 2) ABL1: 1
 - ✓ ABL1(H396P): 0.87
 - ✓ ABL1(M351T): 0.53
 - ✓ ABL1(Q252H): 2.3
 - ✓ ABL1(T315I): 0.21
 - ✓ ABL1(Y253F): 0.72
 - 3) ABL2: 1.2
 - 4) ACK1: 2.2
 - 5) Aurora2: 6.6
 - 6) Aurora3: 0.31
 - 7) BIKE: 0.038
 - 8) CAMK1: 2
 - 9) CAMK1D: 1.1
 - 10) CAMK1G: 0.99
 - 11) CAMK2A: 0.37
 - 12) CAMK2B: 3.2
 - 13) CAMK2D: 1.1
 - 14) CAMK2G: 0.76
 - 15) CAMKK1: 0.9
 - 16) CAMKK2: 5.8
 - 17) CLK1: 0.1
 - 18) CLK2: 0.19

- 19) CLK4: 0.08
 20) DAPK2: 0.47
 21) DAPK3: 0.3
 22) EPHA5: 5.9
 23) EPHA6: 2
 24) EPHA7: 0.71
 25) EPHB1: 0.96
 26) EPHB4: 2.2
 27) FER: 1.4
 28) FGFR1: 1.8
 29) FGFR2: 0.53
 30) FGFR3: 0.3
 31) FGR: 0.29
 32) FLT3: 0.0008
 33) FLT4: 0.035
 34) FYN: 3.9
 35) GAK: 0.12
 36) HCK: 3.3
 37) INSR: 0.18
 38) JAK1 (Kin.Dom.1): 0.0092
 39) JAK2 (Kin.Dom.2): 0.94
 40) KIT: 0.00071
 41) LCK: 1.2
 42) LYN: 0.54
 43) MAP3K4: 3.3
 44) MAP4K5: 0.077
 45) MARK2: 0.32
 46) MYLK2: 0.057
 47) NEK2: 0.5
 48) NTRK1: 0.22
 49) PCTK1: 0.13
 50) PDGFRB: 0.00021
 51) PHKG1: 0.07
 52) PHKG2: 0.039
 53) **PIM2**: 5
 54) PRKAA1: 0.052
 55) **PRKACA**: 6
 56) **PTK2**: 0.61
 57) RPS6KA2 (Kin.Dom.1): 0.058
 58) RPS6KA3 (Kin.Dom.1): 0.055
 59) RPS6KA5 (Kin.Dom.1): 0.25
 60) SLK: 0.081
 61) SRC: 3.1
 62) STK10: 0.64
 63) STK16: 0.36
 64) STK17A: 0.021
 65) STK17B: 0.22
 66) STK18: 0.38
 67) STK3_m: 0.16
 68) STK38L: 1.5
 69) STK4: 0.1
 70) TNIK: 0.025
 71) TTK: 0.23
 72) ULK3_m: 0.11
 73) VEGFR2: 0.00023
 74) YES: 0.24

14. inhibits renal cancer cell migration and invasion via attenuating the expression of miR-452-5p(664)
15. indirectly enhances antitumor cytotoxicity of cytokine-induced killer cells and CD3⁺CD56⁺ subset through the coculturing dendritic cells(665)
16. immunomodulation via reversing MDSC-mediated tumor-induced immunosuppression(666-668)
17. triggers incomplete autophagy, impairs cathepsin B activation and stimulates a lysosomal-dependent necrosis(468)
18. directly inhibits FMS phosphorylation and cell proliferation(513)
19. may exert antitumor effects via direct inhibition of tumor growth as opposed to an antiangiogenic mechanism(669)
20. may contribute to immunologic recovery via inducing myeloid lineage redistribution(670)
21. directly binds these kinases (Kd, nM) (NP: nonphosphorylated, P: phosphorylated)(324):
- 1) AAK1: 11
 - 2) ABL1-NP: 270
 - ✓ ABL1-P: 150
 - ✓ ABL1(E255K)-P: 180
 - ✓ ABL1(F317I)-NP: 3600
 - ✓ ABL1(F317I)-P: 890
 - ✓ ABL1(F317L)-NP: 730
 - ✓ ABL1(F317L)-P: 370
 - ✓ ABL1(H396P)-NP: 74
 - ✓ ABL1(H396P)-P: 140
 - ✓ ABL1(M351T)-P: 120
 - ✓ ABL1(Q252H)-NP: 240
 - ✓ ABL1(Q252H)-P: 76
 - ✓ ABL1(T315I)-NP: 150
 - ✓ ABL1(T315I)-P: 55
 - ✓ ABL1(Y253F)-P: 140
 - 3) ABL2: 1000
 - 4) AKT2: 2700
 - 5) ALK: 170
 - 6) AMPK-alpha1: 19
 - 7) AMPK-alpha2: 89
 - 8) ANKK1: 310
 - 9) ARK5: 48
 - 10) AURKA: 1700
 - 11) AURKB: 380
 - 12) AURKC: 220
 - 13) AXL: 9
 - 14) BIKE: 5.5
 - 15) BLK: 65
 - 16) BMPR1B: 2400
 - 17) BMPR2: 570
 - 18) BRK: 4600
 - 19) BRSK1: 3500
 - 20) BRSK2: 1100
 - 21) BTK: 2100
 - 22) CAMK1: 970
 - 23) CAMK1D: 510
 - 24) CAMK1G: 440

- 25) CAMK2A: 80
 26) CAMK2B: 1400
 27) CAMK2D: 420
 28) CAMK2G: 690
 29) CAMK4: 890
 30) CAMKK1: 420
 31) CAMKK2: 1500
 32) CDK4-cyclinD1: 6200
 33) CDK4-cyclinD3: 2000
 34) CDK5: 6900
 35) CDK7: 330
 36) CDKL2: 1100
 37) CHEK1: 300
 38) CHEK2: 10
 39) CIT: 3900
 40) CLK1: 22
 41) CLK2: 20
 42) CLK4: 29
 43) CSF1R: 2.5
 44) CSNK1A1: 99
 45) CSNK1A1L: 550
 46) CSNK1D: 15
 47) CSNK1E: 13
 48) CSNK1G1: 930
 49) CSNK1G2: 110
 50) CSNK1G3: 240
 51) CSNK2A1: 900
 52) CSNK2A2: 160
 53) CTK: 7700
 54) DAPK1: 120
 55) DAPK2: 150
 56) DAPK3: 22
 57) DCAMKL1: 370
 58) DCAMKL2: 2700
 59) DCAMKL3: 110
 60) DDR1: 2000
 61) DDR2: 2900
 62) DLK: 100
 63) DRAK1: 1
 64) DRAK2: 110
 65) DYRK1A: 200
 66) DYRK1B: 2300
 67) DYRK2: 680
 68) EGFR(G719C): 6800
 ✓ EGFR(L858R,T790M): 860
 ✓ EGFR(T790M): 2400
 69) EPHA3: 2100
 70) EPHA5: 1200
 71) EPHA6: 960
 72) EPHA7: 2400
 73) EPHB1: 480
 74) EPHB4: 3100
 75) EPHB6: 1000
 76) ERK5: 9900
 77) ERN1: 600
 78) FAK: 440

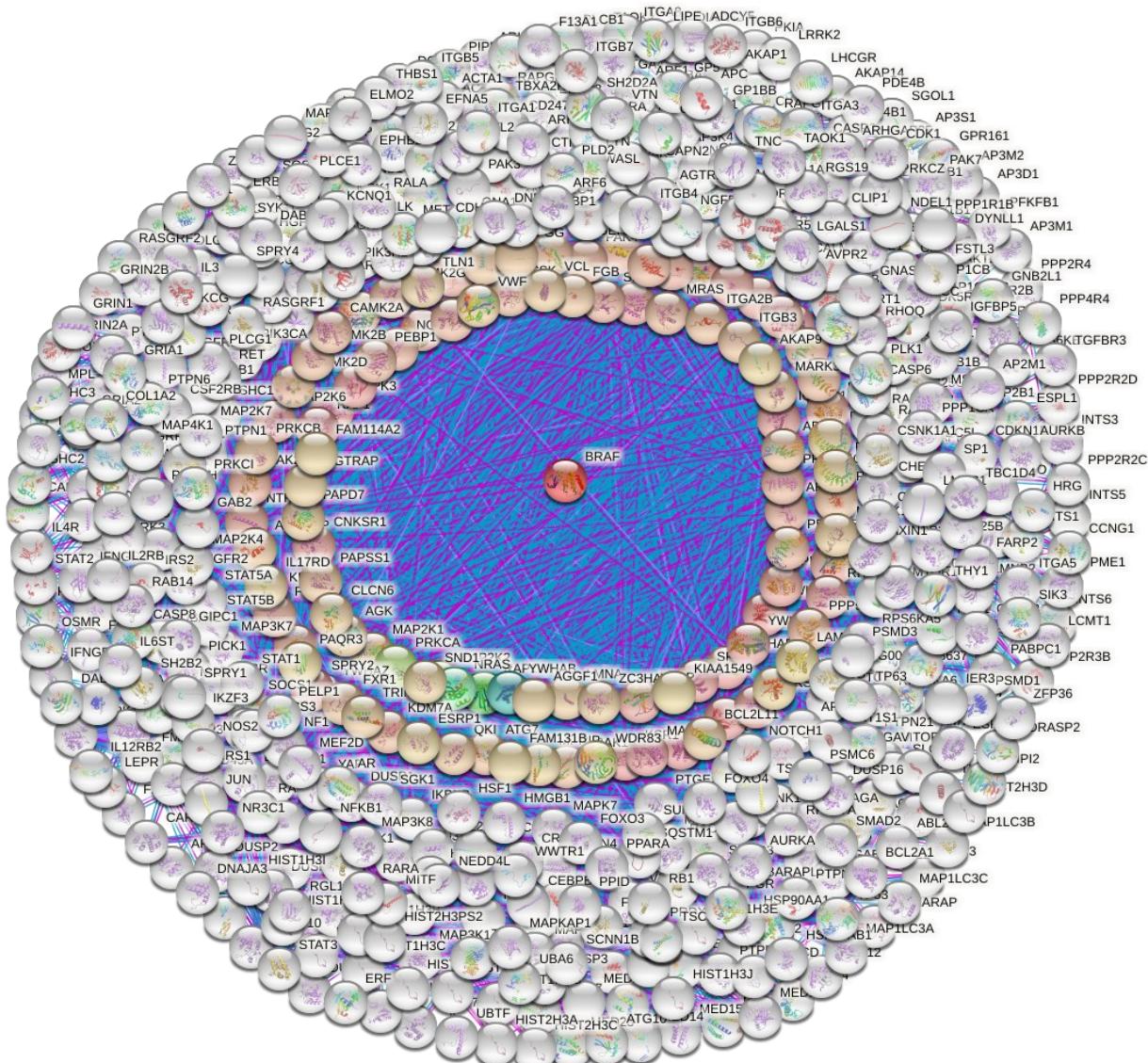
- 79) FER: 1100
 80) FES: 960
 81) FGFR1: 520
 82) FGFR2: 570
 83) FGFR3: 290
 - ✓ FGFR3(G697C): 1400
 84) FGFR4: 2100
 85) FGR: 270
 86) FLT1: 1.8
 87) FLT3: 0.41
 - ✓ FLT3(D835H): 4.3
 - ✓ FLT3(D835Y): 2.3
 - ✓ FLT3(ITD): 0.99
 - ✓ FLT3(K663Q): 0.22
 - ✓ FLT3(N841I): 2.4
 - ✓ FLT3(R834Q): 11
 88) FLT4: 50
 89) FRK: 530
 90) FYN: 520
 91) GAK: 20
 92) GCN2(Kin.Dom.2,S808G): 180
 93) GRK1: 290
 94) GRK4: 140
 95) GRK7: 180
 96) HCK: 880
 97) HIPK1: 55
 98) HIPK2: 31
 99) HIPK3: 41
 - 100) HIPK4: 160
 - 101) HPK1: 16
 - 102) HUNK: 500
 - 103) ICK: 470
 - 104) IGF1R: 2600
 - 105) IKK-alpha: 520
 - 106) IKK-epsilon: 620
 - 107) INSR: 500
 - 108) INSRR: 430
 - 109) IRAK1: 14
 - 110) IRAK3: 940
 - 111) IRAK4: 66
 - 112) ITK: 13
 - 113) JAK1(JH1domain-catalytic): 6000
 - ✓ JAK1(JH2domain-pseudokinase): 49
 - 114) JAK2(JH1domain-catalytic): 410
 - 115) JAK3(JH1domain-catalytic): 1200
 - 116) JNK2: 2400
 - 117) JNK3: 4300
 - 118) KIT: 0.37
 - ✓ KIT(A829P): 43
 - ✓ KIT(D816H): 110
 - ✓ KIT(D816V): 380
 - ✓ KIT(L576P): 1.3
 - ✓ KIT(V559D): 0.41
 - ✓ KIT(V559D,T670I): 0.28
 - ✓ KIT(V559D,V654A): 0.21
 - 119) LATS1: 630

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|------|---------------------|------|---------------------|
| 120) | LATS2: 460 | 147) | MELK: 350 |
| 121) | LCK: 230 | 148) | MERTK: 26 |
| 122) | LKB1: 38 | 149) | MET: 7200 |
| 123) | LOK: 19 | | ✓ MET(M1250T): 1200 |
| 124) | LRRK2: 110 | | ✓ MET(Y1235D): 4700 |
| | ✓ LRRK2(G2019S): 76 | 150) | MINK: 29 |
| 125) | LTK: 1800 | 151) | MKNK1: 3900 |
| 126) | LYN: 270 | 152) | MKNK2: 5700 |
| 127) | LZK: 95 | 153) | MLCK: 23 |
| 128) | MAP3K15: 1300 | 154) | MLK1: 3400 |
| 129) | MAP3K2: 57 | 155) | MLK3: 1300 |
| 130) | MAP3K3: 220 | 156) | MST1: 19 |
| 131) | MAP3K4: 4800 | 157) | MST2: 56 |
| 132) | MAP4K2: 33 | 158) | MST3: 63 |
| 133) | MAP4K3: 180 | 159) | MST4: 340 |
| 134) | MAP4K4: 140 | 160) | MUSK: 490 |
| 135) | MAP4K5: 41 | 161) | MYLK: 280 |
| 136) | MARK1: 1200 | 162) | MYLK2: 49 |
| 137) | MARK2: 310 | 163) | MYLK4: 15 |
| 138) | MARK3: 410 | 164) | MYO3A: 3100 |
| 139) | MARK4: 3600 | 165) | MYO3B: 4500 |
| 140) | MAST1: 200 | 166) | NDR1: 410 |
| 141) | MEK1: 130 | 167) | NDR2: 970 |
| 142) | MEK2: 110 | 168) | NEK2: 1400 |
| 143) | MEK3: 1700 | 169) | NEK7: 4100 |
| 144) | MEK4: 700 | 170) | NIM1: 850 |
| 145) | MEK5: 46 | 171) | OSR1: 530 |
| 146) | MEK6: 5200 | 172) | PAK3: 16 |

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|------|---------------------------------------|------|--|
| 173) | PAK4: 2300 | 201) | RET: 13 |
| 174) | PAK6: 2400 | | ✓ RET(M918T): 19 |
| 175) | PAK7: 640 | | ✓ RET(V804L): 8.7 |
| 176) | PCTK1: 150 | | ✓ RET(V804M): 5.8 |
| 177) | PCTK2: 1200 | 202) | RIOK1: 35 |
| 178) | PCTK3: 1700 | 203) | RIOK2: 49 |
| 179) | PDGFRA: 0.79 | 204) | RIOK3: 3800 |
| 180) | PDGFRB: 0.075 | 205) | RIPK1: 370 |
| 181) | PDPK1: 3500 | 206) | RIPK4: 2100 |
| 182) | PFCDPK1(<i>P. falciparum</i>): 1300 | 207) | RIPK5: 1300 |
| 183) | PFTK1: 270 | 208) | ROCK1: 460 |
| 184) | PHKG1: 5.5 | 209) | ROCK2: 140 |
| 185) | PHKG2: 5.9 | 210) | RPS6KA4(Kin.Dom.1-N-terminal):
96 |
| 186) | PIM3: 2400 | | ✓ RPS6KA4(Kin.Dom.2-C-terminal):
2000 |
| 187) | PIP5K1A: 5400 | 211) | RPS6KA5(Kin.Dom.1-N-terminal):
28 |
| 188) | PIP5K2B: 39 | | ✓ RPS6KA5(Kin.Dom.2-C-terminal):
1700 |
| 189) | PKN1: 710 | 212) | RSK1(Kin.Dom.1-N-terminal): 140 |
| 190) | PKN2: 1300 | | ✓ RSK1(Kin.Dom.2-C-terminal):
5000 |
| 191) | PKNB(<i>M. tuberculosis</i>): 87 | 213) | RSK2(Kin.Dom.1-N-terminal): 580 |
| 192) | PLK2: 4700 | 214) | RSK3(Kin.Dom.1-N-terminal): 17 |
| 193) | PLK4: 190 | 215) | RSK3(Kin.Dom.2-C-terminal): 8400 |
| 194) | PRKCQ: 4300 | 216) | RSK4(Kin.Dom.1-N-terminal): 2400 |
| 195) | PRKD1: 310 | 217) | S6K1: 48 |
| 196) | PRKD2: 380 | 218) | SBK1: 200 |
| 197) | PRKD3: 280 | 219) | SgK110: 1900 |
| 198) | PRKR: 670 | | |
| 199) | PRP4: 390 | | |
| 200) | PYK2: 82 | | |

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|------|--------------|------|-------------------------------------|
| 220) | SGK3: 220 | 248) | TTK: 63 |
| 221) | SIK: 3200 | 249) | TYK2(JH1domain-catalytic): 1600 |
| 222) | SIK2: 580 | | ✓ TYK2(JH2domain-pseudokinase): 360 |
| 223) | SLK: 56 | 250) | TYRO3: 49 |
| 224) | SNARK: 150 | 251) | ULK1: 23 |
| 225) | SNRK: 640 | 252) | ULK2: 13 |
| 226) | SRC: 2100 | 253) | ULK3: 42 |
| 227) | SRPK1: 250 | 254) | VEGFR2: 1.5 |
| 228) | SRPK2: 190 | 255) | WEE1: 1100 |
| 229) | SRPK3: 59 | 256) | YES: 120 |
| 230) | STK16: 250 | 257) | YSK1: 290 |
| 231) | STK33: 17 | 258) | YSK4: 17 |
| 232) | STK35: 1300 | | |
| 233) | STK39: 140 | | |
| 234) | TAK1: 93 | | |
| 235) | TAOK1: 890 | | |
| 236) | TAOK3: 210 | | |
| 237) | TBK1: 120 | | |
| 238) | TIE1: 3900 | | |
| 239) | TLK1: 740 | | |
| 240) | TLK2: 330 | | |
| 241) | TNIK: 25 | | |
| 242) | TNK1: 680 | | |
| 243) | TNK2: 8900 | | |
| 244) | TRKA: 100 | | |
| 245) | TRKB: 590 | | |
| 246) | TRKC: 5100 | | |
| 247) | TSSK1B: 6300 | | |

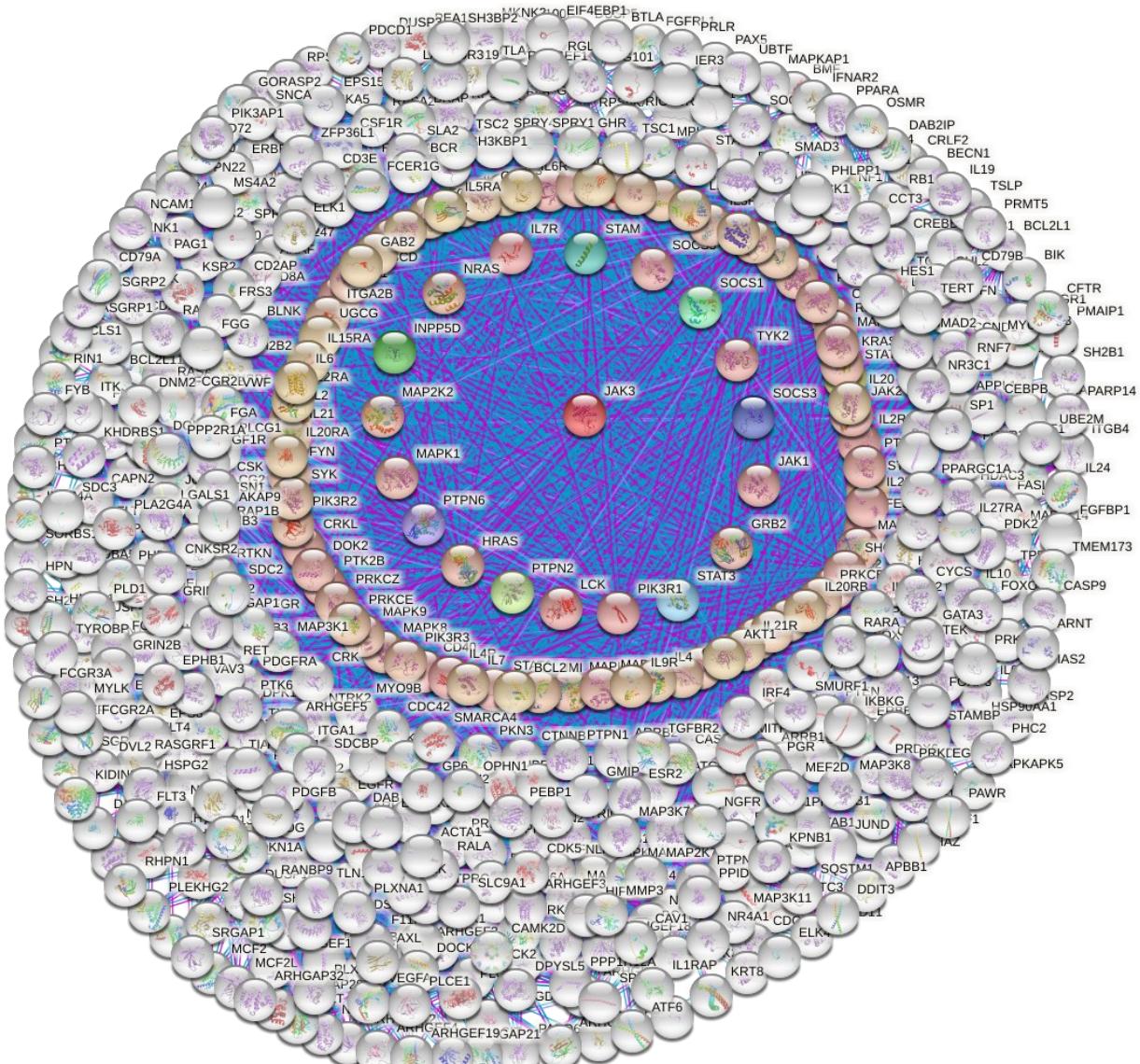
Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
VEMURAFENIB	B-Raf proto-oncogene, serine/threonine kinase	BRAF	P15056 18



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1. induces endoplasmic reticulum stress-mediated apoptosis(671)
2. downmodulates aggressiveness mediators of colorectal cancer: Low Molecular Weight Protein Tyrosine Phosphatase (**LMWPTP**)(672)
3. stimulate inflammasome activation and **IL-1 β** production in dendritic cells which may affect the dendritic cell-mediated course of anti-tumor immune responses(673)
4. compromise the ATP binding capacity of MAP2K5 and inhibit its kinase activity(674)
5. upregulates 58 kinases and downregulates 5 kinases(674)
6. reduces the ATP binding affinities of these kinases(674):
 - 1) ARAF
 - 2) ZAK
 - 3) SRC
 - 4) IP6K1
 - 5) MAP2K5
 - 6) FER
 - 7) PRPS1
 - 8) AK6
 - 9) ABR
 - 10) FGFR3
 - 11) CHUK
 - 12) PEAK1
7. increases **TRAIL**-induced apoptosis(675)
8. reverses **immunosuppression** by myeloid derived suppressor cells(676)
9. binds the active site of Protein tyrosine kinase 6 (**PTK6**, also called BRK) and inhibits its activation(677)
10. triggers production of **superoxide and nitric oxide** and the consequent depolarization of mitochondrial membranes, and potentially apoptosis and growth inhibition(678)

Drugs	Discovery Target	Uniprot ID	"Off-Target" Therapeutic Mechanisms
TOFACITINIB	Janus kinase 3	JAK3	P52333 34



<https://version-11-0b.string-db.org/cgi/network?networkId=bD9gJDmjupjZ>

1. restores the balance of $\gamma\delta$ Treg/ $\gamma\delta$ T17 cells in rheumatoid arthritis by inhibiting the **NLRP3 inflammasome**(679)
2. modulates inflammation and exerts protective effects in endothelial cells via suppressing ox-LDL-mediated activation of NF- κ B inhibitor α (I κ B- α), accumulation of nuclear p65, activation of nuclear factor κ B (NF- κ B) promoter and **adhesion of THP-1 monocytes to endothelial cells**(680)
3. **directly binds these kinases** (K_d , nM) (NP: nonphosphorylated, P: phosphorylated)(324):
 - 1) ABL1(H396P)-NP: 4500
 - ✓ ABL1(H396P)-P: 6700
 - ✓ ABL1(T315I)-P: 2200
 - 2) BIKE: 7100
 - 3) CAMK1: 1200
 - 4) CAMK1D: 1600
 - 5) CAMK2A: 3500
 - 6) CAMK2D: 2700
 - 7) DCAMKL1: 6700
 - 8) DCAMKL3: 12
 - 9) DMPK: 1200
 - 10) FYN: 1100
 - 11) GRK7: 4300
 - 12) JAK1(JH1domain-catalytic): 1.6
 - 13) JAK2(JH1domain-catalytic): 0.58
 - 14) JAK3(JH1domain-catalytic): 0.16
 - 15) LCK: 460
- 16) LRRK2: 1300
- 17) LRRK2(G2019S): 550
- 18) MAP4K2: 4800
- 19) MKNK2: 1600
- 20) PKN1: 170
- 21) PKN2: 1200
- 22) PKNB(*M. tuberculosis*): 2000
- 23) PRKCD: 2000
- 24) RET(M918T): 3300
- 25) ROCK1: 470
- 26) ROCK2: 420
- 27) RSK1(Kin.Dom.2-C-terminal): 1400
- 28) RSK3(Kin.Dom.2-C-terminal): 600
- 29) RSK4(Kin.Dom.2-C-terminal): 540
- 30) SNARK: 240
- 31) TNK1: 120
- 32) TYK2(JH1domain-catalytic): 4.8
- 33) ULK3: 6400

References

1. H. Shen, T. Kihara, H. Hongo, X. Wu, W. R. Kem, S. Shimohama, A. Akaike, T. Niidome, H. Sugimoto, Neuroprotection by donepezil against glutamate excitotoxicity involves stimulation of $\alpha 7$ nicotinic receptors and internalization of NMDA receptors. *British Journal of Pharmacology* **161**, 127-39 (2010). doi.org/10.1111/j.1476-5381.2010.00894.x ncbi.nlm.nih.gov/pmc/PMC2962822
2. A. Akaike, Y. Takada-Takatori, T. Kume, Y. Izumi, Mechanisms of neuroprotective effects of nicotine and acetylcholinesterase inhibitors: Role of $\alpha 4$ and $\alpha 7$ receptors in neuroprotection. *Journal of Molecular Neuroscience* **40**, 211-6 (2010). doi.org/10.1007/s12031-009-9236-1
3. E. Arias, S. Gallego-Sandín, M. Villarroya, A. G. García, M. G. López, Unequal neuroprotection afforded by the acetylcholinesterase inhibitors galantamine, donepezil, and rivastigmine in SHSY5Y neuroblastoma cells: Role of nicotinic receptors. *Journal of Pharmacology and Experimental Therapeutics* **315**, 1346-53 (2005). doi.org/10.1124/jpet.105.090365
4. Y. Takada-Takatori, T. Kume, Y. Ohgi, Y. Izumi, T. Niidome, T. Fujii, H. Sugimoto, A. Akaike, Mechanism of neuroprotection by donepezil pretreatment in rat cortical neurons chronically treated with donepezil. *Journal of Neuroscience Research* **86**, 3575-83 (2008). doi.org/10.1002/jnr.21798
5. A. Akaike, H. Katsuki, T. Kume, Y. Takada-Takatori, Neuroprotective action of donepezil mediated by neuronal nicotinic receptors. *Psychogeriatrics* **6**, S47-S56 (2006). doi.org/10.1111/j.1479-8301.2006.00171.x
6. S. Akasofu, T. Kosasa, M. Kimura, A. Kubota, Protective effect of donepezil in a primary culture of rat cortical neurons exposed to oxygen-glucose deprivation. *European Journal of Pharmacology* **472**, 57-63 (2003). [doi.org/10.1016/s0014-2999\(03\)01865-x](https://doi.org/10.1016/s0014-2999(03)01865-x)
7. G. Doganay, B. Khodr, G. Georgiou, Z. Khalil, Pharmacological manipulation of the vasoconstrictive effects of amyloid-beta peptides by donepezil and rivastigmine. *Current Alzheimer Research* **3**, 137-45 (2006). doi.org/10.2174/156720506776383086
8. S. Akasofu, K. Sawada, T. Kosasa, H. Hihara, H. Ogura, A. Akaike, Donepezil attenuates excitotoxic damage induced by membrane depolarization of cortical neurons exposed to veratridine. *European Journal of Pharmacology* **588**, 189-97 (2008). doi.org/10.1016/j.ejphar.2008.03.064
9. M. Arikawa, Y. Kakinuma, T. Noguchi, H. Todaka, T. Sato, Donepezil, an acetylcholinesterase inhibitor, attenuates LPS-induced inflammatory response in murine macrophage cell line RAW 264.7 through inhibition of nuclear factor kappa B translocation. *European Journal of Pharmacology* **789**, 17-26 (2016). doi.org/10.1016/j.ejphar.2016.06.053
10. H. G. Kim, M. Moon, J. G. Choi, G. Park, A. J. Kim, J. Hur, K. T. Lee, M. S. Oh, Donepezil inhibits the amyloid-beta oligomer-induced microglial activation in vitro and in vivo. *Neurotoxicology* **40**, 23-32 (2014). doi.org/10.1016/j.neuro.2013.10.004
11. E. Kim, M. Park, J. Jeong, H. Kim, S. K. Lee, E. Lee, B. H. Oh, K. Namkoong, Cholinesterase inhibitor donepezil increases mitochondrial biogenesis through AMP-activated protein kinase in the hippocampus. *Neuropsychobiology* **73**, 81-91 (2016). doi.org/10.1159/000441522
12. M. M. Atef, N. M. El-Sayed, A. A. M. Ahmed, Y. M. Mostafa, Donepezil improves neuropathy through activation of AMPK signalling pathway in streptozotocin-induced diabetic mice. *Biochemical Pharmacology* **159**, 1-10 (2019). doi.org/10.1016/j.bcp.2018.11.006
13. E. Conti, L. Tremolizzo, M. E. Santarone, M. Tironi, I. Radice, C. P. Zoia, A. Aliprandi, A. Salmaggi, R. Dominici, M. Casati, I. Appollonio, C. Ferrarese, Donepezil modulates the endogenous immune response: Implications for Alzheimer's disease. *Human Psychopharmacology* **31**, 296-303 (2016). doi.org/10.1002/hup.2538
14. R. M. Lataro, M. A. B. Silva, F. L. Mestriner, S. B. A. Cau, R. C. A. Tostes, H. C. Salgado, Chronic treatment with acetylcholinesterase inhibitors attenuates vascular dysfunction in spontaneously hypertensive rats. *American Journal of Hypertension* **32**, 579-87 (2019). doi.org/10.1093/ajh/hpz036
15. S. Di Angelantonio, G. Bernardi, N. B. Mercuri, Donepezil modulates nicotinic receptors of substantia nigra dopaminergic neurones. *British Journal of Pharmacology* **141**, 644-52 (2004). doi.org/10.1038/sj.bjp.0705660 ncbi.nlm.nih.gov/pmc/PMC1574242
16. G. S. Dong, X. Li, Q. H. Jiang, H. Q. Yang, [Effects of donepezil treatment on platelets alpha and beta secretase activities in Alzheimer's disease patients]. *Zhonghua Yi Xue Za Zhi* **91**, 3341-5 (2011). doi.org/10.3760/cma.j.issn.0376-2491.2011.47.008
17. M. Goschorska, I. Baranowska-Bosiacka, I. Gutowska, M. Tarnowski, K. Piotrowska, E. Metryka, K. Safranow, D. Chlubek, Effect of acetylcholinesterase inhibitors donepezil and rivastigmine on the activity and expression of cyclooxygenases in a model of the inflammatory action of fluoride on macrophages obtained from THP-1 monocytes. *Toxicology* **406-407**, 9-20 (2018). doi.org/10.1016/j.tox.2018.05.007
18. M. Goschorska, I. Gutowska, I. Baranowska-Bosiacka, K. Piotrowska, E. Metryka, K. Safranow, D. Chlubek, Influence of acetylcholinesterase inhibitors used in Alzheimer's disease treatment on the activity of antioxidant enzymes and the concentration of glutathione in THP-1 macrophages under fluoride-induced oxidative stress. *International Journal of Environmental Research and Public Health* **16**, (2019). doi.org/10.3390/ijerph16010010 ncbi.nlm.nih.gov/pmc/PMC6339019
19. S. Umukoro, F. A. Adewole, A. T. Eduviere, A. O. Aderibigbe, C. Onwuchekwa, Free radical scavenging effect of donepezil as the possible contribution to its memory enhancing activity in mice. *Drug Research* **64**, 236-9 (2014). doi.org/10.1055/s-0033-1357126
20. H. B. Guo, Y. F. Cheng, J. G. Wu, C. M. Wang, H. T. Wang, C. Zhang, Z. K. Qiu, J. P. Xu, Donepezil improves learning and memory deficits in APP/PS1 mice by inhibition of microglial activation. *Neuroscience* **290**, 530-42 (2015). doi.org/10.1016/j.neuroscience.2015.01.058
21. L. A. Mohamed, H. Qosa, A. Kaddoumi, Age-related decline in brain and hepatic clearance of amyloid-beta is rectified by the cholinesterase inhibitors donepezil and rivastigmine in rats. *ACS Chemical Neuroscience* **6**, 725-36 (2015). doi.org/10.1021/acscchemneuro.5b00040 ncbi.nlm.nih.gov/pmc/PMC5248655

22. Y. Haraguchi, Y. Mizoguchi, M. Ohgidani, Y. Imamura, T. Murakawa-Hirachi, H. Nabeta, H. Tateishi, T. A. Kato, A. Monji, Donepezil suppresses intracellular Ca²⁺ mobilization through the PI3K pathway in rodent microglia. *Journal of Neuroinflammation* **14**, (2017). doi.org/10.1186/s12974-017-1033-0 ncbi.nlm.nih.gov/pmc/PMC5741946
23. K. Hashimoto, Potential role of the sigma-1 receptor chaperone in the beneficial effects of donepezil in dementia with lewy bodies. *Clinical Psychopharmacology and Neuroscience* **11**, 43-4 (2013). doi.org/10.9758/cpn.2013.11.1.43 ncbi.nlm.nih.gov/pmc/PMC3650298
24. M. Ishikawa, M. Sakata, K. Ishii, Y. Kimura, K. Oda, J. Toyohara, J. Wu, K. Iishiwa, M. Iyo, K. Hashimoto, High occupancy of σ1 receptors in the human brain after single oral administration of donepezil: A positron emission tomography study using [11C]SA4503. *International Journal of Neuropsychopharmacology* **12**, 1127-31 (2009). doi.org/10.1017/S1461145709990204
25. T. Ishima, T. Nishimura, M. Iyo, K. Hashimoto, Potentiation of nerve growth factor-induced neurite outgrowth in PC12 cells by donepezil: Role of sigma-1 receptors and IP3 receptors. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* **32**, 1656-9 (2008). doi.org/10.1016/j.pnpbp.2008.06.011
26. J. Meunier, J. Ieni, T. Maurice, The anti-amnesic and neuroprotective effects of donepezil against amyloid B25-35 peptide-induced toxicity in mice involve an interaction with the σ1 receptor. *British Journal of Pharmacology* **149**, 998-1012 (2006). doi.org/10.1038/sj.bjp.0706927 ncbi.nlm.nih.gov/pmc/PMC2014636
27. S. Kunitachi, Y. Fujita, T. Ishima, M. Kohno, M. Horio, Y. Tanibuchi, Y. Shirayama, M. Iyo, K. Hashimoto, Phencyclidine-induced cognitive deficits in mice are ameliorated by subsequent subchronic administration of donepezil: Role of sigma-1 receptors. *Brain Research* **1279**, 189-96 (2009). doi.org/10.1016/j.brainres.2009.05.004
28. T. Maurice, J. Meunier, B. Feng, J. Ieni, D. T. Monaghan, Interaction with σ1 protein, but not N-Methyl-D-aspartate receptor, is involved in the pharmacological activity of donepezil. *Journal of Pharmacology and Experimental Therapeutics* **317**, 606-14 (2006). doi.org/10.1124/jpet.105.097394
29. J. Hwang, H. Hwang, H. W. Lee, K. Suk, Microglia signaling as a target of donepezil. *Neuropharmacology* **58**, 1122-9 (2010). doi.org/10.1016/j.neuropharm.2010.02.003
30. O. Imamura, M. Arai, M. Dateki, T. Ogata, R. Uchida, H. Tomoda, K. Takishima, Nicotinic acetylcholine receptors mediate donepezil-induced oligodendrocyte differentiation. *Journal of Neurochemistry* **135**, 1086-98 (2015). doi.org/10.1111/jnc.13294
31. W. X. Jian, Z. Zhang, J. H. Zhan, S. F. Chu, Y. Peng, M. Zhao, Q. Wang, N. H. Chen, Donepezil attenuates vascular dementia in rats through increasing BDNF induced by reducing HDAC6 nuclear translocation. *Acta Pharmacologica Sinica* **41**, 588-98 (2020). doi.org/10.1038/s41401-019-0334-5 ncbi.nlm.nih.gov/pmc/PMC7470853
32. H. Kinoshita, K. Nakahata, K. Hama-Tomioka, Y. Ishida, N. Matsuda, N. Hatakeyama, M. Haba, T. Kondo, Y. Hatano, Cholinesterase inhibitor donepezil dilates cerebral parenchymal arterioles via the activation of neuronal nitric oxide synthase. *Anesthesiology* **109**, 124-9 (2008). doi.org/10.1097/ALN.0b013e31817c0316
33. N. A. Kapai, J. V. Bukanova, E. I. Solntseva, V. G. Skrebitsky, Donepezil in a narrow concentration range augments control and impaired by beta-amyloid peptide hippocampal LTP in NMDAR-independent manner. *Cellular and Molecular Neurobiology* **32**, 219-26 (2012). doi.org/10.1007/s10571-011-9751-9
34. N. A. Kapai, E. I. Solntseva, V. G. Skrebitskii, Donepezil eliminates suppressive effects of beta-amyloid peptide (1-42) on long-term potentiation in the hippocampus. *Bulletin of Experimental Biology and Medicine* **149**, 33-6 (2010). doi.org/10.1007/s10517-010-0868-5
35. M. Fujiki, H. Kobayashi, S. Uchida, R. Inoue, K. Ishii, Neuroprotective effect of donepezil, a nicotinic acetylcholine-receptor activator, on cerebral infarction in rats. *Brain Research* **1043**, 236-41 (2005). doi.org/10.1016/j.brainres.2005.02.063
36. S. Nakano, T. Asada, H. Matsuda, M. Uno, M. Takasaki, Donepezil hydrochloride preserves regional cerebral blood flow in patients with Alzheimer's disease. *Journal of Nuclear Medicine* **42**, 1441-5 (2001).
37. D. Min, X. Mao, K. Wu, Y. Cao, F. Guo, S. Zhu, N. Xie, L. Wang, T. Chen, C. Shaw, J. Cai, Donepezil attenuates hippocampal neuronal damage and cognitive deficits after global cerebral ischemia in gerbils. *Neuroscience Letters* **510**, 29-33 (2012). doi.org/10.1016/j.neulet.2011.12.064
38. M. Kimura, H. Komatsu, H. Ogura, K. Sawada, Comparison of donepezil and memantine for protective effect against amyloid-beta(1-42) toxicity in rat septal neurons. *Neuroscience Letters* **391**, 17-21 (2005). doi.org/10.1016/j.neulet.2005.08.036
39. S. Kotani, T. Yamauchi, T. Teramoto, H. Ogura, Donepezil, an acetylcholinesterase inhibitor, enhances adult hippocampal neurogenesis. *Chemico-Biological Interactions* **175**, 227-30 (2008). doi.org/10.1016/j.cbi.2008.04.004
40. T. Leyhe, E. Stransky, G. W. Eschweiler, G. Buchkremer, C. Laske, Increase of BDNF serum concentration during donepezil treatment of patients with early Alzheimer's disease. *European Archives of Psychiatry and Clinical Neuroscience* **258**, 124-8 (2008). doi.org/10.1007/s00406-007-0764-9
41. Y. X. Li, Z. H. Ye, T. Chen, X. F. Jia, L. He, The effects of donepezil on phencyclidine-induced cognitive deficits in a mouse model of schizophrenia. *Pharmacology Biochemistry and Behavior* **175**, 69-76 (2018). doi.org/10.1016/j.pbb.2018.09.006
42. K. Makitani, S. Nakagawa, Y. Izumi, A. Akaike, T. Kume, Inhibitory effect of donepezil on bradykinin-induced increase in the intracellular calcium concentration in cultured cortical astrocytes. *Journal of Pharmacological Sciences* **134**, 37-44 (2017). doi.org/10.1016/j.jphs.2017.03.008
43. J. Man, K. Cui, X. Fu, D. Zhang, Z. Lu, Y. Gao, L. Yu, N. Li, J. Wang, Donepezil promotes neurogenesis via Src signaling pathway in a rat model of chronic cerebral hypoperfusion. *Brain Research* **1736**, 146782 (2020). doi.org/10.1016/j.brainres.2020.146782
44. S. M. Mortazavian, H. Parsaei, S. H. Mousavi, Z. Tayarani-Najaran, A. Ghorbani, H. R. Sadeghnia, Acetylcholinesterase inhibitors promote angiogenesis in chick chorioallantoic membrane and inhibit apoptosis of endothelial cells. *International Journal of*

- Alzheimer's Disease* **2013**, 121068 (2013).
doi.org/10.1155/2013/121068 ncbi.nlm.nih.gov/pmc/PMC3789489
45. N. Narimatsu, N. Harada, H. Kurihara, N. Nakagata, K. Sobue, K. Okajima, Donepezil improves cognitive function in mice by increasing the production of insulin-like growth factor-I in the hippocampus. *Journal of Pharmacology and Experimental Therapeutics* **330**, 2-12 (2009). doi.org/10.1124/jpet.108.147280
46. M. Zimmermann, F. Gardoni, E. Marcello, F. Colciaghi, B. Borroni, A. Padovani, F. Cattabeni, M. Di Luca, Acetylcholinesterase inhibitors increase ADAM10 activity by promoting its trafficking in neuroblastoma cell lines. *Journal of Neurochemistry* **90**, 1489-99 (2004). doi.org/10.1111/j.1471-4159.2004.02680.x
47. M. Y. Noh, S. H. Koh, Y. Kim, H. Y. Kim, G. W. Cho, S. H. Kim, Neuroprotective effects of donepezil through inhibition of GSK-3 activity in amyloid- β -induced neuronal cell death. *Journal of Neurochemistry* **108**, 1116-25 (2009). doi.org/10.1111/j.1471-4159.2008.05837.x
48. M. Y. Noh, S. H. Koh, S. M. Kim, T. Maurice, S. K. Ku, S. H. Kim, Neuroprotective effects of donepezil against A β 42-induced neuronal toxicity are mediated through not only enhancing PP2A activity but also regulating GSK-3 β and nAChRs activity. *Journal of Neurochemistry* **127**, 562-74 (2013). doi.org/10.1111/jnc.12319
49. R. P. Obermayr, L. Mayerhofer, M. Knechtelsdorfer, N. Mersich, E. R. Huber, G. Geyer, K. H. Tragl, The age-related down-regulation of the growth hormone/insulin-like growth factor-1 axis in the elderly male is reversed considerably by donepezil, a drug for Alzheimer's disease. *Experimental Gerontology* **40**, 157-63 (2005). doi.org/10.1016/j.exger.2004.11.001
50. T. Oda, T. Kume, H. Katsuki, T. Niidome, H. Sugimoto, A. Akaike, Donepezil potentiates nerve growth factor-induced neurite outgrowth in PC12 cells. *Journal of Pharmacological Sciences* **104**, 349-54 (2007). doi.org/10.1254/jphs.fp0070563
51. M. Page, N. Pacico, S. Ourtionalous, T. Deprez, K. Koshibu, Procognitive compounds promote neurite outgrowth. *Pharmacology* **96**, 131-6 (2015). doi.org/10.1159/000436974
52. M. Páksáki, A. Fehér, A. Juhász, G. Drótós, O. C. Fazekas, J. Kovács, Z. Janka, J. Kálman, Serum adipokine levels modified by donepezil treatment in Alzheimer's disease. *Journal of Alzheimer's Disease* **38**, 371-7 (2014). doi.org/10.3233/JAD-131139
53. M. Reale, C. Iarlori, F. Gambi, C. Feliciani, L. Isabella, D. Gambi, The acetylcholinesterase inhibitor, Donepezil, regulates a Th2 bias in Alzheimer's disease patients. *Neuropharmacology* **50**, 606-13 (2006). doi.org/10.1016/j.neuropharm.2005.11.006
54. A. Shamsi, S. Anwar, T. Mohammad, M. F. Alajmi, A. Hussain, M. T. Rehman, G. M. Hasan, A. Islam, M. I. Hassan, MARK4 inhibited by AChE inhibitors, donepezil and rivastigmine tartrate: Insights into alzheimer's disease therapy. *Biomolecules* **10**, (2020). doi.org/10.3390/biom10050789 ncbi.nlm.nih.gov/pmc/PMC7277793
55. E. I. Solntseva, J. V. Bukanova, E. Marchenko, V. G. Skrebitsky, Donepezil is a strong antagonist of voltage-gated calcium and potassium channels in molluscan neurons. *Comparative Biochemistry and Physiology - C Toxicology and Pharmacology* **144**, 319-26 (2007). doi.org/10.1016/j.cbpc.2006.10.001
56. B. Yu, G. Y. Hu, Donepezil blocks voltage-gated ion channels in rat dissociated hippocampal neurons. *European Journal of Pharmacology* **508**, 15-21 (2005). doi.org/10.1016/j.ejphar.2004.12.004

57. Y. Takada-Takatori, T. Kume, Y. Ohgi, T. Fujii, T. Niidome, H. Sugimoto, A. Akaike, Mechanisms of α 7-nicotinic receptor up-regulation and sensitization to donepezil induced by chronic donepezil treatment. *European Journal of Pharmacology* **590**, 150-6 (2008). doi.org/10.1016/j.ejphar.2008.06.027
58. L. L. Talib, S. R. Hototian, H. P. G. Joaquim, O. V. Forlenza, W. F. Gattaz, Increased iPLA2 activity and levels of phosphorylated GSK3B in platelets are associated with donepezil treatment in Alzheimer's disease patients. *European Archives of Psychiatry and Clinical Neuroscience* **265**, 701-6 (2015). doi.org/10.1007/s00406-015-0600-6
59. K. Tsuchiya, H. Tajima, M. Yamada, H. Takahashi, T. Kuwae, K. Sunaga, N. Katsube, R. Ishitani, Disclosure of a pro-apoptotic glyceraldehyde-3-phosphate dehydrogenase promoter: Anti-dementia drugs depress its activation in apoptosis. *Life Sciences* **74**, 3245-58 (2004). doi.org/10.1016/j.lfs.2003.11.029
60. C. N. Wang, Y. J. Wang, H. Wang, L. Song, Y. Chen, J. L. Wang, Y. Ye, B. Jiang, The anti-dementia effects of donepezil involve miR-206-3p in the hippocampus and cortex. *Biological and Pharmaceutical Bulletin* **40**, 465-72 (2017). doi.org/10.1248/bpb.b16-00898
61. C. Y. Ye, Y. Lei, X. C. Tang, H. Y. Zhang, Donepezil attenuates A β -associated mitochondrial dysfunction and reduces mitochondrial A β accumulation in vivo and in vitro. *Neuropharmacology* **95**, 29-36 (2015). doi.org/10.1016/j.neuropharm.2015.02.020
62. T. S. Yu, A. Kim, S. G. Kernie, Donepezil rescues spatial learning and memory deficits following traumatic brain injury independent of its effects on neurogenesis. *PloS One* **10**, e0118793 (2015). doi.org/10.1371/journal.pone.0118793 ncbi.nlm.nih.gov/pmc/PMC4340948
63. H. Yuan, W. P. Wang, N. Feng, L. Wang, X. L. Wang, Donepezil attenuated oxygen-glucose deprivation insult by blocking Kv2.1 potassium channels. *European Journal of Pharmacology* **657**, 76-83 (2011). doi.org/10.1016/j.ejphar.2011.01.054
64. T. Zhang, F. Tian, J. Wang, S. Zhou, X. Dong, K. Guo, J. Jing, Y. Zhou, Y. Chen, Donepezil attenuates high glucose-accelerated senescence in human umbilical vein endothelial cells through SIRT1 activation. *Cell Stress and Chaperones* **20**, 787-92 (2015). doi.org/10.1007/s12192-015-0601-4 ncbi.nlm.nih.gov/pmc/PMC4529865
65. X. Zhou, W. Xiao, Z. Su, J. Cheng, C. Zheng, Z. Zhang, Y. Wang, L. Wang, B. Xu, S. Li, X. Yang, M. Pui Man Hoi, Hippocampal proteomic alteration in triple transgenic mouse model of Alzheimer's disease and implication of PINK 1 regulation in donepezil treatment. *Journal of Proteome Research* **18**, 1542-52 (2019). doi.org/10.1021/acs.jproteome.8b00818
66. Y. Takada-Takatori, S. Nakagawa, R. Kimata, Y. Nao, Y. Mizukawa, T. Urushidani, Y. Izumi, A. Akaike, K. Tsuchida, T. Kume, Donepezil modulates amyloid precursor protein endocytosis and reduction by up-regulation of SNX33 expression in primary cortical neurons. *Scientific Reports* **9**, 11922 (2019). doi.org/10.1038/s41598-019-47462-4 ncbi.nlm.nih.gov/pmc/PMC6695423

67. T. T. Guo, W. P. Wang, X. L. Wang, Mechanism of donepezil protecting neurons by inhibiting sodium channel and NMDA receptor channel. *Chinese Pharmaceutical Journal* **52**, 2166-71 (2017).
68. Z. Zhu, L. Zhang, Y. Cui, M. Li, R. Ren, G. Li, X. Sun, Q. Li, Functional compensation and mechanism of choline acetyltransferase in the treatment of cognitive deficits in aged dementia mice. *Neuroscience* **442**, 41-53 (2020). doi.org/10.1016/j.neuroscience.2020.05.016
69. A. J. Hirsh, S. Y. Yao, J. D. Young, C. I. Cheeseman, Inhibition of glucose absorption in the rat jejunum: A novel action of alpha-D-glucosidase inhibitors. *Gastroenterology* **113**, 205-11 (1997). [doi.org/10.1016/s0016-5085\(97\)70096-9](https://doi.org/10.1016/s0016-5085(97)70096-9)
70. A. Merat, M. Sahmani, Effect of acarbose on in vitro intestinal absorption of monosaccharides in diabetic rats. *Archives of Iranian Medicine* **6**, 40-3 (2003).
71. M. Iwase, H. Imoto, M. Oku, S. Shibata, K. Sonoki, M. Iida, Acarbose feeding increases pancreatic islet blood flow in obese glucose-intolerant Otsuka Long-Evans Tokushima fatty rats. *Pancreas* **37**, 228-30 (2008). doi.org/10.1097/MPA.0b013e318164c41e
72. T. Kawamura, G. Egusa, R. Fujikawa, T. Watanabe, K. Oda, S. Kataoka, S. Takayama, K. Kubo, S. Yamamoto, K. Noma, R. Orita, M. Yamakido, Effect of acarbose on glycemic control and lipid metabolism in patients with non-insulin-dependent diabetes mellitus. *Current Therapeutic Research* **59**, 97-106 (1998). [doi.org/10.1016/S0011-393X\(98\)85004-2](https://doi.org/10.1016/S0011-393X(98)85004-2)
73. Y. L. Kim, S. H. Chung, Antidiabetic activity and mechanisms of acarbose in KKAY mice. *Korean Journal of Physiology and Pharmacology* **5**, 183-8 (2001).
74. X. X. Li, S. K. Ling, M. Y. Hu, Y. Ma, Y. Li, P. L. Huang, Protective effects of acarbose against vascular endothelial dysfunction through inhibiting Nox4/NLRP3 inflammasome pathway in diabetic rats. *Free Radical Biology and Medicine* **145**, 175-86 (2019). doi.org/10.1016/j.freeradbiomed.2019.09.015
75. R. Petlevski, D. Juretić, L. Mayer, M. Hadžija, M. Slijepčević, J. Lukač-Bajalo, Effect of acarbose on glucose-6-phosphatase in liver of CBA diabetic mice. *Periodicum biologorum* **104**, 73-5 (2002).
76. J. Rameshwar, K. Anand, Antihyperglycaemic and antiperoxidative roles of acarbose in type 2 diabetes mellitus are possibly mediated through changes in thyroid function. *Clinical and Experimental Pharmacology and Physiology* **33**, 1104-6 (2006). doi.org/10.1111/j.1440-1681.2006.04499.x
77. L. Ranganath, F. Morris, L. Morgan, J. Wright, V. Marks, Delayed gastric emptying occurs following acarbose administration and is a further mechanism for its anti-hyperglycaemic effect. *Diabetic Medicine* **15**, 120-4 (1998). [doi.org/10.1002/\(SICI\)1096-9136\(199802\)15:2<120::AID-DIA529>3.0.CO;2-I](https://doi.org/10.1002/(SICI)1096-9136(199802)15:2<120::AID-DIA529>3.0.CO;2-I)
78. B. J. Smith, R. A. Miller, A. C. Ericsson, D. C. Harrison, R. Strong, T. M. Schmidt, Changes in the gut microbiome and fermentation products concurrent with enhanced longevity in acarbose-treated mice. *BMC Microbiology* **19**, 130 (2019). doi.org/10.1186/s12866-019-1494-7 ncbi.nlm.nih.gov/pmc/PMC6567620
79. B. Su, H. Liu, J. Li, Y. Sunli, B. Liu, D. Liu, P. Zhang, X. Meng, Acarbose treatment affects the serum levels of inflammatory cytokines and the gut content of bifidobacteria in Chinese patients with type 2 diabetes mellitus. *Journal of Diabetes* **7**, 729-39 (2015). doi.org/10.1111/1753-0407.12232
80. Q. Zhang, X. Xiao, M. Li, W. Li, M. Yu, H. Zhang, Z. Wang, H. Xiang, Acarbose reduces blood glucose by activating miR-10a-5p and miR-664 in diabetic rats. *PLoS One* **8**, e79697 (2013). doi.org/10.1371/journal.pone.0079697 ncbi.nlm.nih.gov/pmc/PMC3832586
81. B. Zhao, F. Wu, X. Han, W. Zhou, Q. Shi, H. Wang, Protective effects of acarbose against insulitis in multiple low-dose streptozotocin-induced diabetic mice. *Life Sciences* **263**, 118490 (2020). doi.org/10.1016/j.lfs.2020.118490
82. Y. Chen, G. Meng, W. Bai, Y. Ma, L. Xie, N. Altaf, Y. Qian, Y. Han, Y. Ji, Aliskiren protects against myocardial ischaemia-reperfusion injury via an endothelial nitric oxide synthase dependent manner. *Clinical and Experimental Pharmacology and Physiology* **44**, 266-74 (2017). doi.org/10.1111/1440-1681.12692
83. M. H. Chiang, C. J. Liang, C. W. Liu, B. J. Pan, W. P. Chen, Y. F. Yang, I. T. Lee, J. S. Tsai, C. W. Lee, Y. L. Chen, Aliskiren improves ischemia- and oxygen glucose deprivation-induced cardiac injury through activation of autophagy and AMP-activated protein kinase. *Frontiers in Pharmacology* **8**, 819 (2017). doi.org/10.3389/fphar.2017.00819 ncbi.nlm.nih.gov/pmc/PMC5694452
84. Y. Gu, X. Tang, L. Xie, G. Meng, Y. Ji, Aliskiren improves endothelium-dependent relaxation of thoracic aorta by activating PI3K/Akt/eNOS signal pathway in SHR. *Clinical and Experimental Pharmacology and Physiology* **43**, 450-8 (2016). doi.org/10.1111/1440-1681.12550
85. T. Imanishi, H. Tsujioka, H. Ikejima, A. Kuroi, S. Takarada, H. Kitabata, T. Tanimoto, Y. Muragaki, S. Mochizuki, M. Goto, K. Yoshida, T. Akasaka, Renin inhibitor aliskiren improves impaired nitric oxide bioavailability and protects against atherosclerotic changes. *Hypertension* **52**, 563-72 (2008). doi.org/10.1161/HYPERTENSIONAHA.108.111120
86. G. Jagadeesh, P. Balakumar, N. Stockbridge, How well do aliskiren's purported mechanisms track its effects on cardiovascular and renal disorders? *Cellular Signalling* **24**, 1583-91 (2012). doi.org/10.1016/j.cellsig.2012.04.003
87. S. S. Koid, J. Ziogas, D. J. Campbell, Aliskiren reduces myocardial ischemia-reperfusion injury by a bradykinin B2 receptor- and angiotensin AT2 receptor-mediated mechanism. *Hypertension* **63**, 768-73 (2014). doi.org/10.1161/HYPERTENSIONAHA.113.02902
88. H. Moriya, S. Kobayashi, T. Ohtake, D. Tutumi, Y. Mochida, K. Ishioka, M. Oka, K. Maesato, S. Hidaka, S. Nomura, Aliskiren, a direct renin inhibitor, improves vascular endothelial function in patients on hemodialysis independent of antihypertensive effect approximately a pilot study approximately. *Kidney and Blood Pressure Research* **37**, 190-8 (2013). doi.org/10.1159/000350144
89. S. Plecevic, O. Pechanova, A. Barta, A. Vranic, J. Jeremic, L. Arsenijevic, N. Jeremic, V. Jakovljevic, M. Jevdjevic, D. Stanojevic, Effects of the direct renin inhibitor aliskiren on oxidative stress in isolated rat heart. *Serbian Journal of Experimental and Clinical Research* **16**, 193-9 (2015). doi.org/10.1515/sjercr-2015-0025

90. J. Poss, C. Werner, D. Lorenz, C. Gensch, M. Bohm, U. Laufs, The renin inhibitor aliskiren upregulates pro-angiogenic cells and reduces atherogenesis in mice. *Basic Research in Cardiology* **105**, 725-35 (2010). doi.org/10.1007/s00395-010-0120-5
91. A. E. Raptis, K. P. Markakis, M. C. Mazioti, I. Ikonomidis, E. P. Maratou, D. V. Vlahakos, E. E. Kotsifaki, A. N. Voumvourakis, A. G. Tsirogianni, V. A. Lambadiari, J. P. Lekakis, S. A. Raptis, G. D. Dimitriadis, Effect of aliskiren on circulating endothelial progenitor cells and vascular function in patients with type 2 diabetes and essential hypertension. *American Journal of Hypertension* **28**, 22-9 (2015). doi.org/10.1093/ajh/hpu119
92. A. Rashikh, S. J. Ahmad, K. K. Pillai, K. Kohli, A. K. Najmi, Aliskiren attenuates myocardial apoptosis and oxidative stress in chronic murine model of cardiomyopathy. *Biomedicine and Pharmacotherapy* **66**, 138-43 (2012). doi.org/10.1016/j.biopha.2011.11.020
93. C. L. Chou, C. Y. Pang, T. J. Lee, T. C. Fang, Direct renin inhibitor prevents and ameliorates insulin resistance, aortic endothelial dysfunction and vascular remodeling in fructose-fed hypertensive rats. *Hypertension Research* **36**, 123-8 (2013). doi.org/10.1038/hr.2012.124
94. T. A. Ramirez, R. P. Iyer, O. Ghasemi, E. F. Lopez, D. B. Levin, J. Zhang, R. Zamilpa, Y. M. Chou, Y. F. Jin, M. L. Lindsey, Aliskiren and valsartan mediate left ventricular remodeling post-myocardial infarction in mice through MMP-9 effects. *Journal of Molecular and Cellular Cardiology* **72**, 326-35 (2014). doi.org/10.1016/j.yjmcc.2014.04.007 ncbi.nlm.nih.gov/pmc/PMC4095995
95. M. Sakoda, A. Ichihara, A. Kurauchi-Mito, T. Narita, K. Kinouchi, K. Murohashi-Bokuda, M. A. Saleem, A. Nishiyama, F. Suzuki, H. Itoh, Aliskiren inhibits intracellular angiotensin II levels without affecting (pro)renin receptor signals in human podocytes. *American Journal of Hypertension* **23**, 575-80 (2010). doi.org/10.1038/ajh.2009.273
96. C. Savoia, E. Arrabito, R. Parente, L. Sada, L. Madaro, C. Nicoletti, L. Zezza, A. Alonso, S. Rubattu, S. Michelini, D. N. Muller, M. Volpe, The direct renin inhibitor aliskiren improves vascular remodelling in transgenic rats harbouring human renin and angiotensinogen genes. *Clin Sci (Lond)* **125**, 183-9 (2013). doi.org/10.1042/CS20120395
97. C. F. Tsai, Y. C. Chen, Y. K. Lin, S. A. Chen, Y. J. Chen, Electromechanical effects of the direct renin inhibitor (aliskiren) on the pulmonary vein and atrium. *Basic Research in Cardiology* **106**, 979-93 (2011). doi.org/10.1007/s00395-011-0206-8
98. A. Virdis, L. Ghiadoni, A. A. Qasem, G. Lorenzini, E. Duranti, G. Cartoni, R. M. Bruno, G. Bernini, S. Taddei, Effect of aliskiren treatment on endothelium-dependent vasodilation and aortic stiffness in essential hypertensive patients. *European Heart Journal* **33**, 1530-8 (2012). doi.org/10.1093/eurheartj/ehs057
99. L. Q. Weng, W. B. Zhang, Y. Ye, P. P. Yin, J. Yuan, X. X. Wang, L. Kang, S. S. Jiang, J. Y. You, J. Wu, H. Gong, J. B. Ge, Y. Z. Zou, Aliskiren ameliorates pressure overload-induced heart hypertrophy and fibrosis in mice. *Acta Pharmacologica Sinica* **35**, 1005-14 (2014). doi.org/10.1038/aps.2014.45 ncbi.nlm.nih.gov/pmc/PMC4125714
100. S. Yao, C. Su, S. H. Wu, D. J. Hu, X. Liu, Aliskiren improved the endothelial repair capacity of endothelial progenitor cells from patients with hypertension via the Tie2/PI3k/Akt/eNOS signalling pathway. *Cardiology Research and Practice* **2020**, 6534512 (2020). doi.org/10.1155/2020/6534512 ncbi.nlm.nih.gov/pmc/PMC7275222
101. H. Zhi, I. Luptak, G. Alreja, J. Shi, J. Guan, N. Metes-Kosik, J. Joseph, Effects of direct Renin inhibition on myocardial fibrosis and cardiac fibroblast function. *PloS One* **8**, e81612 (2013). doi.org/10.1371/journal.pone.0081612 ncbi.nlm.nih.gov/pmc/PMC3859492
102. C. Allegra, P. L. Antignani, Does rivaroxaban have a fibrinolytic effect? *Acta Phlebologica* **16**, 107-9 (2015).
103. E. Alvarez, B. Paradela-Dobarro, S. Raposeiras-Roubin, J. R. Gonzalez-Juanatey, Protective, repairing and fibrinolytic effects of rivaroxaban on vascular endothelium. *British Journal of Clinical Pharmacology* **84**, 280-91 (2018). doi.org/10.1111/bcp.13440 ncbi.nlm.nih.gov/pmc/PMC5777430
104. J. Mabley, J. P. Patel, A. Sayed, R. Arya, G. Scutt, Direct oral anticoagulant (DOAC)-mediated vasodilation: Role of nitric oxide. *Thrombosis Research* **176**, 36-8 (2019). doi.org/10.1016/j.thromres.2019.02.014
105. Y. Rao, J. Chen, Y. Guo, T. Ji, P. Xie, Rivaroxaban ameliorates angiotensin II-induced cardiac remodeling by attenuating TXNIP/Trx2 interaction in KKAY mice. *Thrombosis Research* **193**, 45-52 (2020). doi.org/10.1016/j.thromres.2020.05.030
106. M. Laurent, U. Joimel, R. Varin, L. Cazin, C. Gest, V. Le-Cam-Duchez, J. Jin, J. Liu, J. P. Vannier, H. Lu, J. Soria, H. Li, C. Soria, Comparative study of the effect of rivaroxaban and fondaparinux on monocyte's coagulant activity and cytokine release. *Experimental Hematology and Oncology* **3**, 30 (2014). doi.org/10.1186/2162-3619-3-30 ncbi.nlm.nih.gov/pmc/PMC4298120
107. T. C. Wu, J. S. Chan, C. Y. Lee, H. B. Leu, P. H. Huang, J. S. Chen, S. J. Lin, J. W. Chen, Rivaroxaban, a factor Xa inhibitor, improves neovascularization in the ischemic hindlimb of streptozotocin-induced diabetic mice. *Cardiovascular Diabetology* **14**, 81 (2015). doi.org/10.1186/s12933-015-0243-y ncbi.nlm.nih.gov/pmc/PMC4473833
108. X. Lou, Z. Yu, X. Yang, J. Chen, Protective effect of rivaroxaban on arteriosclerosis obliterans in rats through modulation of the toll-like receptor 4/NF- κ B signaling pathway. *Experimental and Therapeutic Medicine* **18**, 1619-26 (2019). doi.org/10.3892/etm.2019.7726 ncbi.nlm.nih.gov/pmc/PMC6676094
109. F. Nehaj, J. Sokol, J. Ivankova, M. Mokan, F. Kovar, J. Stasko, M. Mokan, First evidence: TRAP-induced platelet aggregation is reduced in patients receiving Xabans. *Clinical and Applied Thrombosis/Hemostasis* **24**, 914-9 (2018). doi.org/10.1177/1076029617734310 ncbi.nlm.nih.gov/pmc/PMC6714725
110. E. Perzborn, S. Heitmeier, V. Laux, Effects of rivaroxaban on platelet activation and platelet-coagulation pathway interaction: In vitro and in vivo studies. *Journal of Cardiovascular Pharmacology and Therapeutics* **20**, 554-62 (2015). doi.org/10.1177/1074248415578172 ncbi.nlm.nih.gov/pmc/PMC4598653
111. E. Woźniak, M. Broncel, B. Bukowska, P. Gorzelak-Pabiś, The protective effect of dabigatran and rivaroxaban on DNA oxidative

- changes in a model of vascular endothelial damage with oxidized cholesterol. *International Journal of Molecular Sciences* **21**, (2020). doi.org/10.3390/ijms21061953 ncbi.nlm.nih.gov/pmc/PMC7139915
112. X. Shan, Z. Liu, M. Wulasihan, S. Ma, Edoxaban improves atrial fibrillation and thromboembolism through regulation of the Wnt-beta-induced PI3K/ATK-activated protein C system. *Experimental and Therapeutic Medicine* **17**, 3509-17 (2019). doi.org/10.3892/etm.2019.7379 ncbi.nlm.nih.gov/pmc/PMC6447810
113. W. Song, H. Ci, G. Tian, Y. Zhang, X. Ge, Edoxaban improves venous thrombosis via increasing hydrogen sulfide and homocysteine in rat model. *Molecular Medicine Reports* **16**, 7706-14 (2017). doi.org/10.3892/mmr.2017.7574
114. T. Kacergius, A. Ambrozaitis, Y. Deng, S. Gravenstein, Neuraminidase inhibitors reduce nitric oxide production in influenza virus-infected and gamma interferon-activated RAW 264.7 macrophages. *Pharmacological Reports* **58**, 924-30 (2006).
115. B. Zablockiene, T. Kacergius, A. Ambrozaitis, E. Zurauskas, M. Bratchikov, L. Jurgauskiene, R. Zablockis, S. Gravenstein, Zanamivir diminishes lung damage in influenza A virus-infected mice by inhibiting nitric oxide production. *In Vivo* **32**, 473-8 (2018). doi.org/10.21873/invivo.11263 ncbi.nlm.nih.gov/pmc/PMC6000778
116. J. Hauser, S. Szabo, Extremely long protection by pyrazole derivatives against chemically induced gastric mucosal injury. *Journal of Pharmacology and Experimental Therapeutics* **256**, 592-8 (1991).
117. G. Iaquinto, The several activities of 4-methyl pyrazole in animals and humans. *Current Pharmaceutical Design* **24**, 1957-60 (2018). doi.org/10.2174/1381612824666180529081328
118. G. Iaquinto, M. Del Tacca, L. Cuccurullo, M. C. Parodi, N. Giardullo, V. D'Onofrio, G. Natale, D. Carignani, F. Ferraraccio, S. Szabo, Gastroprotection by 4-methylpyrazole against ethanol in humans. *Digestive Diseases and Sciences* **43**, 816-25 (1998). doi.org/10.1023/a:1018878400935
119. K. Sommerfeld, B. Zielinska-Psuja, J. Przystanowicz, J. Kowalowka-Zawieja, J. Orlowski, Effect of 4-methylpyrazole on antioxidant enzyme status and lipid peroxidation in the liver of rats after exposure to ethylene glycol and ethyl alcohol. *Pharmacological Reports* **64**, 1547-53 (2012). [doi.org/10.1016/s1734-1140\(12\)70952-0](https://doi.org/10.1016/s1734-1140(12)70952-0)
120. A. T. Peana, F. A. Pintus, F. Bennardini, G. Rocchitta, G. Bazzu, P. A. Serra, S. Porru, M. Rosas, E. Acquas, Is catalase involved in the effects of systemic and pVTA administration of 4-methylpyrazole on ethanol self-administration? *Alcohol* **63**, 61-73 (2017). doi.org/10.1016/j.alcohol.2017.04.001
121. C. F. Amber, T. K. Zeynep, O. Evren, B. Yusuf, A. K. Can, T. Belma, Di-peptidyl peptidase-4 inhibitor sitagliptin protects vascular function in metabolic syndrome: Possible role of epigenetic regulation. *Molecular Biology Reports* **41**, 4853-63 (2014). doi.org/10.1007/s11033-014-3392-2
122. L. Baerts, L. Glorie, W. Maho, A. Eelen, A. Verhulst, P. D'Haese, A. Covaci, I. De Meester, Potential impact of sitagliptin on collagen-derived dipeptides in diabetic osteoporosis. *Pharmacological Research* **100**, 336-40 (2015). doi.org/10.1016/j.phrs.2015.08.023
123. C. Wu, S. Hu, N. Wang, J. Tian, Dipeptidyl peptidase-4 inhibitor sitagliptin prevents high glucose-induced apoptosis via activation of AMP-activated protein kinase in endothelial cells. *Molecular Medicine Reports* **15**, 4346-51 (2017). doi.org/10.3892/mmr.2017.6501
124. E. Civantos, E. Bosch, E. Ramirez, O. Zhenyukh, J. Egido, O. Lorenzo, S. Mas, Sitagliptin ameliorates oxidative stress in experimental diabetic nephropathy by diminishing the miR-200a/Keap-1/Nrf2 antioxidant pathway. *"Diabetes, Metabolic Syndrome and Obesity"* **10**, 207-22 (2017). doi.org/10.2147/DMSO.S132537 ncbi.nlm.nih.gov/pmc/PMC5473486
125. A. D. Dobrian, Q. Ma, J. W. Lindsay, K. A. Leone, K. Ma, J. Coben, E. V. Galkina, J. L. Nadler, Dipeptidyl peptidase IV inhibitor sitagliptin reduces local inflammation in adipose tissue and in pancreatic islets of obese mice. *American Journal of Physiology: Endocrinology and Metabolism* **300**, E410-21 (2011). doi.org/10.1152/ajpendo.00463.2010 ncbi.nlm.nih.gov/pmc/PMC3043624
126. A. Gonçalves, E. Leal, A. Paiva, E. Teixeira Lemos, F. Teixeira, C. F. Ribeiro, F. Reis, A. F. Ambrósio, R. Fernandes, Protective effects of the dipeptidyl peptidase IV inhibitor sitagliptin in the blood-retinal barrier in a type 2 diabetes animal model. *Diabetes, Obesity and Metabolism* **14**, 454-63 (2012). doi.org/10.1111/j.1463-1326.2011.01548.x
127. Y. He, G. Yang, F. Yao, Y. Xian, G. Wang, L. Chen, X. Lv, H. Gao, Z. Zheng, L. Sun, W. Wang, R. Lin, Sitagliptin inhibits vascular inflammation via the SIRT6-dependent signaling pathway. *International Immunopharmacology* **75**, 105805 (2019). doi.org/10.1016/j.intimp.2019.105805
128. H. Hu, M. Xu, R. Qi, Y. Wang, C. Wang, J. Liu, L. Luo, L. Xia, Z. Fang, Sitagliptin downregulates retinol-binding protein 4 and upregulates glucose transporter type 4 expression in a type 2 diabetes mellitus rat model. *International Journal of Clinical and Experimental Medicine* **8**, 17902-11 (2015). doi.org/10.4292/ijcem.v8.17902 ncbi.nlm.nih.gov/pmc/PMC4694284
129. Y. Wu, M. Xu, H. Bao, J. H. Zhang, Sitagliptin inhibits EndMT In Vitro ana improves cardiac function of diabetic rats through the SDF-1a/PKA pathway. *European Review for Medical and Pharmacological Sciences* **23**, 841-8 (2019). doi.org/10.26355/eurrev_201901_16899
130. Y. Zhou, H. Wang, F. Man, Z. Guo, J. Xu, W. Yan, J. Li, Q. Pan, W. Wang, Sitagliptin protects cardiac function by reducing nitroxidative stress and promoting autophagy in Zucker diabetic fatty (ZDF) rats. *Cardiovascular Drugs and Therapy* **32**, 541-52 (2018). doi.org/10.1007/s10557-018-6831-9
131. H. Y. Zhang, Y. Li, Y. H. Zhong, L. B. Ruan, T. R. Yang, H. P. Yin, Effect of sildigiline on glucose lipid metabolism and the expression of iNOS and GLP-1 receptors in diabetic rats. *European Review for Medical and Pharmacological Sciences* **22**, 8984-9 (2018). doi.org/10.26355/eurrev_201812_16669
132. M. A. Ibrahim, A. Geddawy, S. Abdel-Wahab, Sitagliptin prevents isoproterenol-induced myocardial infarction in rats by modulating nitric oxide synthase enzymes. *European Journal of Pharmacology* **829**, 63-9 (2018). doi.org/10.1016/j.ejphar.2018.04.005 ncbi.nlm.nih.gov/pmc/Merck
133. D. Wang, G. Zhang, X. Chen, T. Wei, C. Liu, C. Chen, Y. Gong, Q. Wei, Sitagliptin ameliorates diabetic nephropathy by blocking

- TGF- β 1/Smad signaling pathway. *International Journal of Molecular Medicine* **41**, 2784-92 (2018). doi.org/10.3892/ijmm.2018.3504 ncbi.nlm.nih.gov/pmc/PMC5846674
134. H. Wang, Y. Zhou, Z. Guo, Y. Dong, J. Xu, H. Huang, H. Liu, W. Wang, Sitagliptin attenuates endothelial dysfunction of Zucker diabetic fatty rats: Implication of the antiperoxynitrite and autophagy. *Journal of Cardiovascular Pharmacology and Therapeutics* **23**, 66-78 (2018). doi.org/10.1177/1074248417715001
135. S. Cheng, L. Li, C. Song, H. Jin, S. Ma, P. Kang, Sitagliptin relieves diabetic nephropathy fibrosis via the MAPK/ERK signaling pathway. *Minerva Endocrinologica* **45**, 273-5 (2020). doi.org/10.23736/S0391-1977.20.03122-3
136. Q. Zhang, L. He, Y. Dong, Y. Fei, J. Wen, X. Li, J. Guan, F. Liu, T. Zhou, Z. Li, Y. Fan, N. Wang, Sitagliptin ameliorates renal tubular injury in diabetic kidney disease via STAT3-dependent mitochondrial homeostasis through SDF-1 α /CXCR4 pathway. *FASEB Journal* **34**, 7500-19 (2020). doi.org/10.1096/fj.201903038R ncbi.nlm.nih.gov/pmc/PMC6079465
137. W. Zheng, J. Zhou, S. Song, W. Kong, W. Xia, L. Chen, T. Zeng, Dipeptidyl-peptidase 4 inhibitor sitagliptin ameliorates hepatic insulin resistance by modulating inflammation and autophagy in ob/ob mice. *International Journal of Endocrinology* **2018**, 8309723 (2018). doi.org/10.1155/2018/8309723 ncbi.nlm.nih.gov/pmc/PMC6079465
138. Z. M. Coskun, ER stress amelioration by saxagliptin protects the liver against fructose-induced insulin resistance. *Archives of Medical Research* **51**, 303-9 (2020). doi.org/10.1016/j.arcmed.2020.03.007
139. C. J. Li, B. Sun, Q. H. Fang, M. Ding, Y. Z. Xing, L. M. Chen, D. M. Yu, Saxagliptin induces beta-cell proliferation through increasing stromal cell-derived factor-1alpha in vivo and in vitro. *Front Endocrinol (Lausanne)* **8**, 326 (2017). doi.org/10.3389/fendo.2017.00326 ncbi.nlm.nih.gov/pmc/PMC5711777
140. W. Li, R. Liu, X. Li, B. Tao, N. Zhai, X. Wang, Q. Li, Y. Zhang, W. Gu, W. Wang, G. Ning, Saxagliptin alters bile acid profiles and yields metabolic benefits in drug-naïve overweight or obese type 2 diabetes patient. *Journal of Diabetes* **11**, 982-92 (2019). doi.org/10.1111/1753-0407.12956
141. S. Ma, Z. Bai, H. Wu, W. Wang, The DPP-4 inhibitor saxagliptin ameliorates ox-LDL-induced endothelial dysfunction by regulating AP-1 and NF-kappaB. *European Journal of Pharmacology* **851**, 186-93 (2019). doi.org/10.1016/j.ejphar.2019.01.008
142. G. Nephan, Z. M. Coskun, S. Bolkent, Dipeptidyl peptidase-4 inhibition prevents cell death via extrinsic and intrinsic apoptotic pathways in rat pancreas with insulin resistance. *Cell Biochemistry & Function* **36**, 212-20 (2018). doi.org/10.1002/cbf.3333
143. I. U. Njerve, S. Akra, T. W. Weiss, S. Solheim, R. Ovstebo, H. C. D. Aass, R. Byrkjeland, H. Arnesen, I. Seljeflot, A double-blinded randomized study investigating a possible anti-inflammatory effect of saxagliptin versus placebo as add-on therapy in patients with both type 2 diabetes and stable coronary artery disease. *Mediators of Inflammation* **2017**, 5380638 (2017). doi.org/10.1155/2017/5380638 ncbi.nlm.nih.gov/pmc/PMC5449736
144. S. S. Shankar, R. R. Shankar, L. A. Mixson, D. L. Miller, H. O. Steinberg, C. R. Beals, D. E. Kelley, Insulin secretory effect of sitagliptin: Assessment with a hyperglycemic clamp combined with a meal challenge. *American Journal of Physiology: Endocrinology and Metabolism* **314**, E406-E12 (2018). doi.org/10.1152/ajpendo.00238.2017 ncbi.nlm.nih.gov/pmc/PMC5846674
145. Y. Liu, F. Xu, P. Jiang, Effect of sitagliptin on expression of skeletal muscle peroxisome proliferator-activated receptor γ coactivator-1 α and irisin in a rat model of type 2 diabetes mellitus. *Journal of International Medical Research* **48**, (2020). doi.org/10.1177/0300060519885569 ncbi.nlm.nih.gov/pmc/PMC7218978
146. L. I. Luxin, X. Lian, Z. Wang, J. Zheng, L. I. U. Jieting, C. H. U. Yanhui, Y. Teng, Z. Zhang, The dipeptidyl peptidase-4 inhibitor sitagliptin ameliorates renal injury in type 1 diabetic mice via inhibiting the TGF- β /Smad signal pathway. *Pharmazie* **74**, 239-42 (2019). doi.org/10.1681/ph.2019.8918
147. D. Ma, Y. Yu, X. Yu, M. Zhang, Y. Yang, The changes of leukocyte telomere length and telomerase activity after sitagliptin intervention in newly diagnosed type 2 diabetes. *Diabetes/Metabolism Research and Reviews* **31**, 256-61 (2015). doi.org/10.1002/dmrr.2578
148. A. M. Malvandi, C. Loretelli, M. Ben Nasr, G. V. Zuccotti, P. Fiorina, Sitagliptin favorably modulates immune-relevant pathways in human beta cells. *Pharmacological Research* **148**, 104405 (2019). doi.org/10.1016/j.phrs.2019.104405
149. S. Qiao, G. Mao, H. Li, Z. Ma, L. Hong, H. Zhang, C. Wang, J. An, DPP-4 inhibitor sitagliptin improves cardiac function and glucose homeostasis and ameliorates beta-cell dysfunction together with reducing S6K1 activation and IRS-1 and IRS-2 degradation in obesity female mice. *Journal of Diabetes Research* **2018**, 3641516 (2018). doi.org/10.1155/2018/3641516 ncbi.nlm.nih.gov/pmc/PMC6079488
150. L. Shi, Y. Ji, D. Liu, Y. Liu, Y. Xu, Y. Cao, X. Jiang, C. Xu, Sitagliptin attenuates high glucose-induced alterations in migration, proliferation, calcification and apoptosis of vascular smooth muscle cells through ERK1/2 signal pathway. *Oncotarget* **8**, 77168-80 (2017). doi.org/10.18632/oncotarget.20417 ncbi.nlm.nih.gov/pmc/PMC5652771
151. X. Ren, R. Zhu, G. Liu, F. Xue, Y. Wang, J. Xu, W. Zhang, W. Yu, R. Li, Effect of sitagliptin on tubulointerstitial Wnt/ β -catenin signalling in diabetic nephropathy. *Nephrology* **24**, 1189-97 (2019). doi.org/10.1111/nep.13641
152. K. Chen, T. Zhuo, J. Wang, Q. Mei, Saxagliptin upregulates nesfatin-1 secretion and ameliorates insulin resistance and metabolic profiles in type 2 diabetes mellitus. *Metabolic Syndrome and Related Disorders* **16**, 336-41 (2018). doi.org/10.1089/met.2018.0010
153. N. Poncina, M. Albiero, L. Menegazzo, R. Cappellari, A. Avogaro, G. P. Fadini, The dipeptidyl peptidase-4 inhibitor saxagliptin improves function of circulating pro-angiogenic cells from type 2 diabetic patients. *Cardiovascular Diabetology* **13**, 92 (2014). doi.org/10.1186/1475-2840-13-92 ncbi.nlm.nih.gov/pmc/PMC4033689
154. X. Ren, G. Liu, Y. Wang, W. Zhang, F. Xue, R. Li, W. Yu, Influence of dipeptidyl peptidase-IV inhibitor sitagliptin on extracellular signal-regulated kinases 1/2 signaling in rats with diabetic nephropathy. *Pharmacology* **100**, 1-13 (2017). doi.org/10.1159/000455874
155. F. Remm, N. Krinkel, D. Lener, D. J. Drucker, S. Sopper, C. Brenner, Sitagliptin Accelerates Endothelial Regeneration after Vascular Injury Independent from GLP1 Receptor Signaling. *Stem*

Cells International **2018**, 5284963 (2018).

doi.org/10.1155/2018/5284963 ncbi.nlm.nih.gov/pmc/PMC5822806

156. G. V. Sangle, L. M. Lauffer, A. Grieco, S. Trivedi, R. Iakoubov, P. L. Brubaker, Novel biological action of the dipeptidylpeptidase-IV inhibitor, sitagliptin, as a glucagon-like peptide-1 secretagogue. *Endocrinology* **153**, 564-73 (2012). doi.org/10.1210/en.2011-1732

157. N. Satoh-Asahara, Y. Sasaki, H. Wada, M. Tochiya, A. Iguchi, R. Nakagawachi, S. Odori, S. Kono, K. Hasegawa, A. Shimatsu, A dipeptidyl peptidase-4 inhibitor, sitagliptin, exerts anti-inflammatory effects in type 2 diabetic patients. *Metabolism* **62**, 347-51 (2013). doi.org/10.1016/j.metabol.2012.09.004

158. S. T. Tang, H. Su, Q. Zhang, H. Q. Tang, C. J. Wang, Q. Zhou, W. Wei, H. Q. Zhu, Y. Wang, Sitagliptin inhibits endothelin-1 expression in the aortic endothelium of rats with streptozotocin-induced diabetes by suppressing the nuclear factor-B/IB system through the activation of AMP-activated protein kinase. *International Journal of Molecular Medicine* **37**, 1558-66 (2016). doi.org/10.3892/ijmm.2016.2578

159. P. Shah, A. Ardestani, G. Dharmadhikari, S. Laue, D. M. Schumann, J. Kerr-Conte, F. Pattou, T. Klein, K. Maedler, The DPP-4 inhibitor linagliptin restores β -cell function and survival in human isolated islets through GLP-1 stabilization. *Journal of Clinical Endocrinology and Metabolism* **98**, E1163-E72 (2013). doi.org/10.1210/jc.2013-1029

160. X. Zhang, Z. Zhang, Y. Yang, Y. Suo, R. Liu, J. Qiu, Y. Zhao, N. Jiang, C. Liu, G. Tse, G. Li, T. Liu, Alogliptin prevents diastolic dysfunction and preserves left ventricular mitochondrial function in diabetic rabbits. *Cardiovascular Diabetology* **17**, 160 (2018). doi.org/10.1186/s12933-018-0803-z

ncbi.nlm.nih.gov/pmc/PMC6307280

161. X. Zhang, Z. Zhang, Y. Zhao, N. Jiang, J. Qiu, Y. Yang, J. Li, X. Liang, X. Wang, G. Tse, G. Li, T. Liu, Alogliptin, a dipeptidyl peptidase-4 inhibitor, alleviates atrial remodeling and improves mitochondrial function and biogenesis in diabetic rabbits. *Journal of the American Heart Association* **6**, (2017). doi.org/10.1161/JAHA.117.005945

ncbi.nlm.nih.gov/pmc/PMC5524117

162. Y. Ali, K. Dohi, R. Okamoto, K. Katayama, M. Ito, Novel molecular mechanisms in the inhibition of adrenal aldosterone synthesis: Action of tolvaptan via vasopressin V2 receptor-independent pathway. *British Journal of Pharmacology* **176**, 1315-27 (2019). doi.org/10.1111/bph.14630

ncbi.nlm.nih.gov/pmc/PMC6468255

163. T. Fujiki, F. Ando, K. Murakami, K. Isobe, T. Mori, K. Susa, N. Nomura, E. Sohara, T. Rai, S. Uchida, Tolvaptan activates the Nrf2/HO-1 antioxidant pathway through PERK phosphorylation. *Scientific Reports* **9**, 9245 (2019). doi.org/10.1038/s41598-019-45539-8 ncbi.nlm.nih.gov/pmc/PMC6592894

164. M. Ishikawa, N. Kobayashi, F. Sugiyama, S. Onoda, T. Ishimitsu, Renoprotective effect of vasopressin v2 receptor antagonist tolvaptan in Dahl rats with end-stage heart failure. *International Heart Journal* **54**, 98-106 (2013). doi.org/10.1536/ihj.54.98

165. T. Yamazaki, Y. Izumi, Y. Nakamura, N. Yamashita, H. Fujiki, M. Osada-Oka, M. Shiota, A. Hanatani, K. Shimada, H. Iwao, M. Yoshiyama, Tolvaptan improves left ventricular dysfunction after

myocardial infarction in rats. *Circulation: Heart Failure* **5**, 794-802 (2012). doi.org/10.1161/circheartfailure.112.968750

166. K. Zhang, Z. Ma, W. Wang, R. Liu, Y. Zhang, M. Yuan, G. Li, Beneficial effects of tolvaptan on atrial remodeling induced by chronic intermittent hypoxia in rats. *Cardiovascular Therapeutics* **36**, e12466 (2018). doi.org/10.1111/1755-5922.12466

167. D. Hoppensteadt, W. Jeske, J. Walenga, R. Bick, J. Fareed, The anti-inflammatory effects of argatroban can be differentiated from other direct thrombin inhibitors: Experimental and clinical observations. *Seminars in Thrombosis and Hemostasis* **34**, 097-102 (2008). doi.org/10.1055/s-0028-1086089

168. Y. Ueki, K. Matsumoto, Y. Kizaki, K. Yoshida, Y. Matsunaga, M. Yano, S. Miyake, Y. Tominaga, K. Eguchi, Argatroban increases nitric oxide levels in patients with peripheral arterial obstructive disease: Placebo-controlled study. *Journal of Thrombosis & Thrombolysis* **8**, 131-7 (1999). doi.org/10.1023/a:1008963118789

169. K. Feldmann, M. Grandoch, C. Kohlmorgen, B. Valentin, S. Gerfer, N. Nagy, S. Hartwig, S. Lehr, A. C. Fender, J. W. Fischer, Decreased M1 macrophage polarization in dabigatran-treated Ldlr-deficient mice: Implications for atherosclerosis and adipose tissue inflammation. *Atherosclerosis* **287**, 81-8 (2019). doi.org/10.1016/j.atherosclerosis.2019.06.897

170. K. Song, Y. Wang, J. Sheng, C. Ma, H. Li, Effects of dabigatran regulates no-reflow phenomenon in acute myocardial infarction mice through anti-inflammatory and anti-oxidative activities and connective tissue growth factor expression. *Molecular Medicine Reports* **17**, 580-5 (2018). doi.org/10.3892/mmr.2017.7861

171. E. C. Alexandre, L. R. Kiguti, F. B. Calmasini, F. H. Silva, K. P. da Silva, R. Ferreira, C. A. Ribeiro, F. Z. Monica, A. S. Pupo, E. Antunes, Mirabegron relaxes urethral smooth muscle by a dual mechanism involving beta3 -adrenoceptor activation and alpha1 -adrenoceptor blockade. *British Journal of Pharmacology* **173**, 415-28 (2016). doi.org/10.1111/bph.13367

ncbi.nlm.nih.gov/pmc/PMC4728418

172. T. Maki, S. Kajioka, M. Itsumi, E. Kareman, K. Lee, M. Shiota, M. Eto, Mirabegron induces relaxant effects via cAMP signaling-dependent and -independent pathways in detrusor smooth muscle. *Lower Urinary Tract Symptoms* **11**, O209-O17 (2019). doi.org/10.1111/luts.12247

173. I. Silva, A. F. Costa, S. Moreira, F. Ferreira, M. T. Magalhães-Cardoso, I. Calejo, M. Silva-Ramos, P. Correia-de-Sá, Inhibition of cholinergic neurotransmission by beta3-adrenoceptors depends on adenosine release and A1-receptor activation in human and rat urinary bladders. *American Journal of Physiology-Renal Physiology* **313**, F388-F403 (2017). doi.org/10.1152/ajpregn.00392.2016

174. T. Damci, S. Yalin, H. Balci, Z. Osar, U. Korugan, M. Ozyazar, H. Ilkova, Orlistat augments postprandial increases in glucagon-like peptide 1 in obese type 2 diabetic patients. *Diabetes Care* **27**, 1077-80 (2004). doi.org/10.2337/diacare.27.5.1077

175. M. Ellrichmann, M. Kapelle, P. R. Ritter, J. J. Holst, K. H. Herzig, W. E. Schmidt, F. Schmitz, J. J. Meier, Orlistat inhibition of intestinal lipase acutely increases appetite and attenuates postprandial glucagon-like peptide-1-(7-36)-amide-1, cholecystokinin, and peptide YY concentrations. *Journal of Clinical Endocrinology & Metabolism* **93**, 3995-8 (2008). doi.org/10.1210/jc.2008-0924

176. M. Sahin, N. Tanaci, M. Yucel, N. B. Tutuncu, N. Guvener, The effect of single-dose orlistat on postprandial serum glucose, insulin and glucagon-like peptide-1 levels in nondiabetic obese patients. *Clinical Endocrinology* **67**, 346-50 (2007). doi.org/10.1111/j.1365-2265.2007.02888.x
177. C. Di Somma, A. Rivellese, G. Pizza, L. Patti, A. De Rosa, P. Cipriano, V. Nedi, A. Rossi, G. Lombardi, A. Colao, S. Savastano, Effects of short-term treatment with orlistat on growth hormone/insulin-like growth factor-I axis in obese postmenopausal women. *Journal of Endocrinological Investigation* **34**, 90-6 (2011). doi.org/10.1007/BF03347036
178. S. Alqahtani, H. Qosa, B. Primeaux, A. Kaddoumi, Orlistat limits cholesterol intestinal absorption by Niemann-pick C1-like 1 (NPC1L1) inhibition. *European Journal of Pharmacology* **762**, 263-9 (2015). doi.org/10.1016/j.ejphar.2015.05.060
179. P. Y. Yang, K. Liu, M. H. Ngai, M. J. Lear, M. R. Wenk, S. Q. Yao, Activity-based proteome profiling of potential cellular targets of Orlistat--an FDA-approved drug with anti-tumor activities. *Journal of the American Chemical Society* **132**, 656-66 (2010). doi.org/10.1021/ja907716f
180. K. S. Pan, A. Siow, D. L. Hay, C. S. Walker, Antagonism of CGRP signaling by rimegepant at two receptors. *Frontiers in Pharmacology* **11**, 1240 (2020). doi.org/10.3389/fphar.2020.01240 ncbi.nlm.nih.gov/pmc/PMC7468408
181. H. J. Kwak, J. S. Song, J. Y. Heo, S. D. Yang, J. Y. Nam, H. G. Cheon, Roflumilast inhibits lipopolysaccharide-induced inflammatory mediators via suppression of nuclear factor-kappaB, p38 mitogen-activated protein kinase, and c-Jun NH₂-terminal kinase activation. *Journal of Pharmacology and Experimental Therapeutics* **315**, 1188-95 (2005). doi.org/10.1124/jpet.105.092056
182. S. W. Kim, J. H. Kim, C. K. Park, T. J. Kim, S. Y. Lee, Y. K. Kim, S. S. Kwon, C. K. Rhee, H. K. Yoon, Effect of roflumilast on airway remodelling in a murine model of chronic asthma. *Clinical and Experimental Allergy* **46**, 754-63 (2016). doi.org/10.1111/cea.12670
183. S. Y. Kim, T. J. An, C. K. Rhee, C. K. Park, J. H. Kim, H. Yoon, The effect and associated mechanism of action of phosphodiesterase 4 (PDE4) inhibitor on CD4+ lymphocyte proliferation. *Clinical and Experimental Pharmacology and Physiology* **48**, 221-6 (2021). doi.org/10.1111/1440-1681.13417
184. H. J. Kwak, J. S. Song, Z. S. No, J. H. Song, S. D. Yang, H. G. Cheon, The inhibitory effects of roflumilast on lipopolysaccharide-induced nitric oxide production in RAW264.7 cells are mediated by heme oxygenase-1 and its product carbon monoxide. *Inflammation Research* **54**, 508-13 (2005). doi.org/10.1007/s00011-005-1386-1
185. J. A. Lambert, S. V. Raju, L. P. Tang, C. M. McNicholas, Y. Li, C. A. Courville, R. F. Farris, G. E. Coricor, L. H. Smoot, M. M. Mazur, M. T. Dransfield, G. B. Bolger, S. M. Rowe, Cystic fibrosis transmembrane conductance regulator activation by roflumilast contributes to therapeutic benefit in chronic bronchitis. *American Journal of Respiratory Cell and Molecular Biology* **50**, 549-58 (2014). doi.org/10.1165/rccm.2013-0228OC ncbi.nlm.nih.gov/pmc/PMC4068936
186. J. A. Meyers, J. Taverna, J. Chaves, A. Makkinje, A. Lerner, Phosphodiesterase 4 inhibitors augment levels of glucocorticoid receptor in B cell chronic lymphocytic leukemia but not in normal circulating hematopoietic cells. *Clinical Cancer Research* **13**, 4920-7 (2007). doi.org/10.1158/1078-0432.CCR-07-0276 ncbi.nlm.nih.gov/pmc/PMC2656255
187. M. J. Sanz, J. Cortijo, M. A. Taha, M. Cerdá-Nicolás, E. Schatton, B. Burgbacher, J. Klar, H. Tenor, C. Schudt, A. C. Issekutz, A. Hatzelmann, E. J. Morcillo, Roflumilast inhibits leukocyte-endothelial cell interactions, expression of adhesion molecules and microvascular permeability. *British Journal of Pharmacology* **152**, 481-92 (2007). doi.org/10.1038/sj.bjp.0707428 ncbi.nlm.nih.gov/pmc/PMC2050829
188. J. Eiringhaus, C. M. Wunsche, P. Tirilomis, J. Herting, N. Bork, V. O. Nikolaev, G. Hasenfuss, S. Sossalla, T. H. Fischer, Sacubitrilat reduces pro-arrhythmogenic sarcoplasmic reticulum Ca(2+) leak in human ventricular cardiomyocytes of patients with end-stage heart failure. *ESC Heart Fail* **7**, 2992-3002 (2020). doi.org/10.1002/ehf2.12918 ncbi.nlm.nih.gov/pmc/PMC7586991
189. M. Imran, M. Q. Hassan, M. S. Akhtar, O. Rahman, M. Akhtar, A. K. Najmi, Sacubitril and valsartan protect from experimental myocardial infarction by ameliorating oxidative damage in Wistar rats. *Clinical and Experimental Hypertension* **41**, 62-9 (2019). doi.org/10.1080/10641963.2018.1441862
190. C. Tu, J. Li, Y. Bu, D. Hangauer, J. Qu, An ion-current-based, comprehensive and reproducible proteomic strategy for comparative characterization of the cellular responses to novel anti-cancer agents in a prostate cell model. *Journal of Proteomics* **77**, 187-201 (2012). doi.org/10.1016/j.jprot.2012.08.020 ncbi.nlm.nih.gov/pmc/PMC4073256
191. F. A. Comandini, A. Lombardi, A. Saponiero, E. Bonmassar, Saquinavir up-regulates telomerase activity in lymphocytes activated with monoclonal antibodies against CD3/CD28. *Journal of Chemotherapy* **13**, (2001). doi.org/10.1179/joc.2001.13.4.384
192. O. M. Delmonte, G. Bertolotto, E. Ricotti, P. A. Tovo, Immunomodulatory effects of two HIV protease inhibitors, saquinavir and ritonavir, on lymphocytes from healthy seronegative individuals. *Immunology Letters* **111**, (2007). doi.org/10.1016/j.imlet.2007.06.003
193. T. L. Guo, D. R. Germolec, D. M. Roesh, K. L. White, Jr., Immunomodulation in female B₆C₅₇F₁ mice following treatment with the HIV protease inhibitor saquinavir for 28 days by gavage. *Journal of Immunotoxicology* **7**, (2010). doi.org/10.3109/1547691X.2010.495097
194. M. B. Lucia, S. Rutella, G. Leone, R. Cauda, HIV protease inhibitors reduce IL-2 release from normal human phytohaemagglutinin-activated T cells. *AIDS* **15**, 2339-41 (2001). doi.org/10.1097/00002030-200111230-00023
195. R. Pacifici, S. Di Carlo, A. Bacosi, S. Pichini, P. Zuccaro, Cytokine production in saquinavir treated mice. *International Journal of Immunopharmacology* **19**, (1997). [doi.org/10.1016/s0192-0561\(97\)00031-3](https://doi.org/10.1016/s0192-0561(97)00031-3)
196. A. Mongia, M. Bhaskaran, K. Reddy, N. Manjappa, N. Baqi, P. C. Singhal, Protease inhibitors modulate apoptosis in mesangial cells derived from a mouse model of HIVAN. *Kidney International* **65**, 860-70 (2004). doi.org/10.1111/j.1523-1755.2004.00464.x
197. J. P. Pribis, Y. Al-Abed, H. Yang, D. Gero, H. Xu, M. F. Montenegro, E. M. Bauer, S. Kim, S. S. Chavan, C. Cai, T. Li, P. Szoleczky, C. Szabo, K. J. Tracey, T. R. Billiar, The HIV protease

inhibitor saquinavir inhibits HMGB1-driven inflammation by targeting the interaction of cathepsin V with TLR4/MyD88. *Molecular Medicine* **21**, 749–57 (2015).

doi.org/10.2119/molmed.2015.00197
ncbi.nlm.nih.gov/pmc/PMC4749497

198. X. Wang, R. Zhang, Y. Tong, X. Ding, S. Jin, X. Zhao, J. Zong, Z. Chen, T. R. Billiar, Q. Li, High-mobility group box 1 protein is involved in the protective effect of Saquinavir on ventilation-induced lung injury in mice. *Acta Biochim Biophys Sin (Shanghai)* **49**, (2017). doi.org/10.1093/abbs/gmx085

199. O. Equils, A. Shapiro, Z. Madak, C. Liu, D. Lu, Human immunodeficiency virus type 1 protease inhibitors block toll-like receptor 2 (TLR2)- and TLR4-induced NF- κ B activation. *Antimicrobial Agents and Chemotherapy* **48**, 3905-11 (2004). doi.org/10.1128/AAC.48.10.3905-3911.2004
ncbi.nlm.nih.gov/pmc/PMC521905

200. P. André, M. Groettrup, P. Klennerman, R. de Giuli, B. L. Booth, Jr., V. Cerundolo, M. Bonneville, F. Jotereau, R. M. Zinkernagel, V. Lotteau, An inhibitor of HIV-1 protease modulates proteasome activity, antigen presentation, and T cell responses. *Proceedings of the National Academy of Sciences of the United States of America* **95**, 13120-4 (1998). doi.org/10.1073/pnas.95.22.13120
ncbi.nlm.nih.gov/pmc/PMC23730

201. A. Gruber, J. C. Wheat, K. L. Kuhen, D. J. Looney, F. Wong-Staal, Differential effects of HIV-1 protease inhibitors on dendritic cell immunophenotype and function. *Journal of Biological Chemistry* **276**, 47840-3 (2001). doi.org/10.1074/jbc.M105582200

202. T. Wolf, S. Findhammer, B. Nolte, E. B. Helm, H. R. Brodt, Inhibition of TNF-alpha mediated cell death by HIV-1 specific protease inhibitors. *European Journal of Medical Research* **8**, 17-24 (2003).

203. H. Aladdin, T. Katzenstein, A. M. Dreves, L. Ryder, J. Gerstoft, P. Skinhøj, B. K. Pedersen, H. Ullum, T-cell receptor excisional circles, telomere length, proliferation and apoptosis in peripheral blood mononuclear cells of human immunodeficiency virus-infected individuals after 18 months of treatment induced viral suppression. *Scandinavian Journal of Immunology* **57**, 485-92 (2003). doi.org/10.1046/j.1365-3083.2003.01258.x

204. A. D. Kelleher, B. L. Booth, Jr., A. K. Sewell, A. Oxenius, V. Cerundolo, A. J. McMichael, R. E. Phillips, D. A. Price, Effects of retroviral protease inhibitors on proteasome function and processing of HIV-derived MHC class I-restricted cytotoxic T lymphocyte epitopes. *AIDS Research and Human Retroviruses* **17**, 1063-6 (2001). doi.org/10.1089/08892201300343744

205. G. Schmidtke, H. G. Holzhütter, M. Bogyo, N. Kairies, M. Groll, R. de Giuli, S. Emch, M. Groettrup, How an inhibitor of the HIV-I protease modulates proteasome activity. *Journal of Biological Chemistry* **274**, 35734-40 (1999). doi.org/10.1074/jbc.274.50.35734

206. L. Ghibelli, F. Mengoni, M. Lichtner, S. Coppola, M. De Nicola, A. Bergamaschi, C. Mastroianni, V. Vullo, Anti-apoptotic effect of HIV protease inhibitors via direct inhibition of calpain. *Biochemical Pharmacology* **66**, 1505-12 (2003). [doi.org/10.1016/s0006-2952\(03\)00505-7](https://doi.org/10.1016/s0006-2952(03)00505-7)

207. M. Z. Dewan, M. Tomita, H. Katano, N. Yamamoto, S. Ahmed, M. Yamamoto, T. Sata, N. Mori, N. Yamamoto, An HIV protease inhibitor, ritonavir targets the nuclear factor-kappaB and inhibits the

tumor growth and infiltration of EBV-positive lymphoblastoid B cells. *International Journal of Cancer* **124**, 622-9 (2009). doi.org/10.1002/ijc.23993

208. E. M. Sloand, P. N. Kumar, S. Kim, A. Chaudhuri, F. F. Weichold, N. S. Young, Human immunodeficiency virus type 1 protease inhibitor modulates activation of peripheral blood CD4(+) T cells and decreases their susceptibility to apoptosis in vitro and in vivo. *Blood* **94**, 1021-7 (1999). doi.org/10.1182/blood.V94.3.1021.415k29_1021_1027

209. E. M. Sloand, J. Maciejewski, P. Kumar, S. Kim, A. Chaudhuri, N. Young, Protease inhibitors stimulate hematopoiesis and decrease apoptosis and ICE expression in CD34(+) cells. *Blood* **96**, 2735-9 (2000). doi.org/10.1182/blood.V96.8.2735

210. F. F. Weichold, J. L. Bryant, S. Pati, O. Barabitskaya, R. C. Gallo, M. S. Reitz, Jr., HIV-1 protease inhibitor ritonavir modulates susceptibility to apoptosis of uninfected T cells. *Journal of Human Virology* **2**, 261-9 (1999).

211. M. Lichtner, F. Mengoni, C. M. Mastroianni, I. Sauzullo, R. Rossi, M. De Nicola, V. Vullo, L. Ghibelli, HIV protease inhibitor therapy reverses neutrophil apoptosis in AIDS patients by direct calpain inhibition. *Apoptosis* **11**, 781-7 (2006). doi.org/10.1007/s10495-006-5699-5

212. S. Chavan, S. Kodoth, R. Pahwa, S. Pahwa, The HIV protease inhibitor Indinavir inhibits cell-cycle progression in vitro in lymphocytes of HIV-infected and uninfected individuals. *Blood* **98**, 383-9 (2001). doi.org/10.1182/blood.v98.2.383

213. M. Opravil, Improvement in immune function due to treatment with indinavir despite severe immune deficiency. *Antiviral Therapy* **3**, 159-67 (1998).

214. E. Pericolini, E. Cenci, E. Gabrielli, S. Perito, P. Mosci, F. Bistoni, A. Vecchiarelli, Indinavir influences biological function of dendritic cells and stimulates antifungal immunity. *Journal of Leukocyte Biology* **83**, 1286-94 (2008). doi.org/10.1189/jlb.0707454

215. M. Piccinini, M. T. Rinaudo, N. Chiapello, E. Ricotti, S. Baldovino, M. Mostert, P. A. Tovo, The human 26S proteasome is a target of antiretroviral agents. *AIDS* **16**, 693-700 (2002). doi.org/10.1097/00002030-200203290-00004

216. J. G. Weaver, A. Tarze, T. C. Moffat, M. Lebras, A. Deniaud et al., Inhibition of adenine nucleotide translocator pore function and protection against apoptosis in vivo by an HIV protease inhibitor. *Journal of Clinical Investigation* **115**, 1828-38 (2005). doi.org/10.1172/JCI22954 ncbi.nlm.nih.gov/pmc/PMC1142110

217. K. T. Whelan, C. L. Lin, M. Celli, A. J. McMichael, J. M. Austyn, S. L. Rowland-Jones, The HIV protease inhibitor indinavir reduces immature dendritic cell transendothelial migration. *European Journal of Immunology* **33**, 2520-30 (2003). doi.org/10.1002/eji.200323646

218. M. A. Wallet, C. M. Reist, J. C. Williams, S. Appelberg, G. L. Guiulfo, B. Gardner, J. W. Sleasman, M. M. Goodenow, The HIV-1 protease inhibitor nelfinavir activates PP2 and inhibits MAPK signaling in macrophages: A pathway to reduce inflammation. *Journal of Leukocyte Biology* **92**, 795-805 (2012). doi.org/10.1189/jlb.0911447 ncbi.nlm.nih.gov/pmc/PMC3441314

219. A. Di Micco, G. Frera, J. Lugrin, Y. Jamilloux, E. T. Hsu, A. Tardivel, A. De Gassart, L. Zaffalon, B. Bujisic, S. Siegert, M.

Quadroni, P. Broz, T. Henry, C. A. Hrycyna, F. Martinon, AIM2 inflamasome is activated by pharmacological disruption of nuclear envelope integrity. *Proceedings of the National Academy of Sciences of the United States of America* **113**, E4671-80 (2016).

doi.org/10.1073/pnas.1602419113
ncbi.nlm.nih.gov/pmc/PMC4987819

220. H. Garg, R. Blumenthal, HIV gp41-induced apoptosis is mediated by caspase-3-dependent mitochondrial depolarization, which is inhibited by HIV protease inhibitor nelfinavir. *Journal of Leukocyte Biology* **79**, 351-62 (2006). doi.org/10.1189/jlb.0805430

221. D. B. Graham, M. P. Bell, C. J. Huntoon, J. G. Weaver, N. Hawley, A. D. Badley, D. J. McKean, Increased thymic output in HIV-negative patients after antiretroviral therapy. *AIDS* **19**, 1467-72 (2005). doi.org/10.1097/01.aids.0000182520.69159.8a

222. S. Kravcik, A. Magill, B. Sanghvi, R. Ogden, D. W. Cameron, R. Lewis, G. Yu, A. D. Badley, Comparative CD4 T-cell responses of reverse transcriptase inhibitor therapy with or without nelfinavir matched for viral exposure. *HIV Clinical Trials* **2**, 160-70 (2001). doi.org/10.1310/F45L-FDKK-Y48N-N2BT

223. O. Miró, J. Villarroya, G. Garrabou, S. López, M. Rodríguez de la Concepción, E. Pedrol, E. Martínez, M. Giralt, J. M. Gatell, F. Cardellach, J. Casademont, F. Villarroya, In vivo effects of highly active antiretroviral therapies containing the protease inhibitor nelfinavir on mitochondrially driven apoptosis. *Antiviral Therapy* **10**, 945-51 (2005).

224. R. J. Danaher, C. S. Kaetzel, R. N. Greenberg, C. Wang, M. E. Bruno, C. S. Miller, HIV protease inhibitors alter innate immune response signaling to double-stranded RNA in oral epithelial cells: Implications for immune reconstitution inflammatory syndrome? *AIDS* **24**, 2587-90 (2010). doi.org/10.1097/QAD.0b013e32833f4022
ncbi.nlm.nih.gov/pmc/PMC3166643

225. G. Batman, A. W. Oliver, I. Zehbe, C. Richard, L. Hampson, I. N. Hampson, Lopinavir up-regulates expression of the antiviral protein ribonuclease L in human papillomavirus-positive cervical carcinoma cells. *Antiviral Therapy* **16**, 515-25 (2011). doi.org/10.3851/IMP1786

226. M. R. Pennington, J. K. Grenier, G. R. Van de Walle, Transcriptome profiling of alphaherpesvirus-infected cells treated with the HIV-integrase inhibitor raltegravir reveals profound and specific alterations in host transcription. *Journal of General Virology* **99**, 1115-28 (2018). doi.org/10.1099/jgv.0.001090

227. H. Arberas, A. C. Guardo, M. E. Bargallo, M. J. Maleno, M. Calvo, J. L. Blanco, F. Garcia, J. M. Gatell, M. Plana, In vitro effects of the CCR5 inhibitor maraviroc on human T cell function. *Journal of Antimicrobial Chemotherapy* **68**, 577-86 (2013). doi.org/10.1093/jac/dks432

228. K. M. Kelly, S. E. Beck, K. A. Metcalf Pate, S. E. Queen, J. L. Dorsey, R. J. Adams, L. B. Avery, W. Hubbard, P. M. Tarwater, J. L. Mankowski, Neuroprotective maraviroc monotherapy in simian immunodeficiency virus-infected macaques: Reduced replicating and latent SIV in the brain. *AIDS* **27**, F21-8 (2013). doi.org/10.1097/QAD.0000000000000074
ncbi.nlm.nih.gov/pmc/PMC4235167

229. M. M. Pozo-Balado, I. Rosado-Sanchez, G. Mendez-Lagares, M. M. Rodriguez-Mendez, E. Ruiz-Mateos, M. R. Benhnia, M. A. Munoz-Fernandez, M. Leal, Y. M. Pacheco, Maraviroc contributes to

the restoration of the homeostasis of regulatory T-cell subsets in antiretroviral-naïve HIV-infected subjects. *Clinical Microbiology and Infection* **22**, 461 e1-5 (2016). doi.org/10.1016/j.cmi.2015.12.025

230. R. Rossi, M. Lichtner, A. De Rosa, I. Sauzullo, F. Mengoni, A. P. Massetti, C. M. Mastroianni, V. Vullo, In vitro effect of anti-human immunodeficiency virus CCR5 antagonist maraviroc on chemotactic activity of monocytes, macrophages and dendritic cells. *Clinical and Experimental Immunology* **166**, 184-90 (2011). doi.org/10.1111/j.1365-2249.2011.04409.x
ncbi.nlm.nih.gov/pmc/PMC3219893

231. X. Wang, K. E. Russell-Lodrigue, M. S. Ratterree, R. S. Veazey, H. Xu, Chemokine receptor CCR5 correlates with functional CD8(+) T cells in SIV-infected macaques and the potential effects of maraviroc on T-cell activation. *FASEB Journal* **33**, 8905-12 (2019). doi.org/10.1096/fj.201802703R
ncbi.nlm.nih.gov/pmc/PMC6662974

232. P. W. Hunt, N. S. Shulman, T. L. Hayes, V. Dahl, M. Somsouk, N. T. Funderburg, B. McLaughlin, A. L. Landay, O. Adeyemi, L. E. Gilman, B. Clagett, B. Rodriguez, J. N. Martin, T. W. Schacker, B. L. Shacklett, S. Palmer, M. M. Lederman, S. G. Deeks, The immunologic effects of maraviroc intensification in treated HIV-infected individuals with incomplete CD41 T-cell recovery: A randomized trial. *Blood* **121**, (2013). doi.org/10.1182/blood-2012-06-436345
ncbi.nlm.nih.gov/pmc/PMC3685899

233. Y. R. Kao, J. Chen, S. R. Narayananari, T. I. Todorova, M. M. Aivalioti, M. Ferreira, P. M. Ramos, C. Pallaud, I. Mantzaris, A. Shastry, J. B. Bussel, A. Verma, U. Steidl, B. Will, Thrombopoietin receptor-independent stimulation of hematopoietic stem cells by eltrombopag. *Science Translational Medicine* **10**, (2018). doi.org/10.1126/scitranslmed.aas9563

234. T. P. Miettinen, J. Peltier, A. Hartlova, M. Gierlinski, V. M. Jansen, M. Trost, M. Bjorklund, Thermal proteome profiling of breast cancer cells reveals proteasomal activation by CDK4/6 inhibitor palbociclib. *EMBO Journal* **37**, (2018). doi.org/10.15252/embj.201798359
ncbi.nlm.nih.gov/pmc/PMC5978322

235. S. AbuHammad, C. Cullinane, C. Martin, Z. Bacolas, T. Ward et al., Regulation of PRMT5-MDM4 axis is critical in the response to CDK4/6 inhibitors in melanoma. *Proceedings of the National Academy of Sciences of the United States of America* **116**, 17990-8000 (2019). doi.org/10.1073/pnas.1901323116
ncbi.nlm.nih.gov/pmc/PMC6731642

236. N. J. Sumi, B. M. Kuenzi, C. E. Knezevic, L. L. Remsing Rix, U. Rix, Chemoproteomics reveals novel protein and lipid kinase targets of clinical CDK4/6 inhibitors in lung cancer. *ACS Chemical Biology* **10**, 2680-6 (2015). doi.org/10.1021/acscchembio.5b00368
ncbi.nlm.nih.gov/pmc/PMC4684772

237. G. Zhang, F. Ma, L. Li, J. Li, P. Li, S. Zeng, H. Sun, E. Li, Palbociclib triggers apoptosis in bladder cancer cells by Cdk2-induced Rad9-mediated reorganization of the Bak-Bcl-xL complex. *Biochemical Pharmacology* **163**, 133-41 (2019). doi.org/10.1016/j.bcp.2019.02.017

238. G. Qin, F. Xu, T. Qin, Q. Zheng, D. Shi, W. Xia, Y. Tian, Y. Tang, J. Wang, X. Xiao, W. Deng, S. Wang, Palbociclib inhibits epithelial-mesenchymal transition and metastasis in breast cancer via c-Jun/COX-2 signaling pathway. *Oncotarget* **6**, 41794-808 (2015). doi.org/10.18632/oncotarget.5993
ncbi.nlm.nih.gov/pmc/PMC4747189

239. F. S. Hsieh, Y. L. Chen, M. H. Hung, P. Y. Chu, M. H. Tsai, L. J. Chen, Y. J. Hsiao, C. T. Shih, M. J. Chang, T. I. Chao, C. W. Shiau, K. F. Chen, Palbociclib induces activation of AMPK and inhibits hepatocellular carcinoma in a CDK4/6-independent manner. *Molecular Oncology* **11**, 1035-49 (2017). doi.org/10.1002/1878-0261.12072 ncbi.nlm.nih.gov/pmc/PMC5537702
240. M. Hafner, C. E. Mills, K. Subramanian, C. Chen, M. Chung, S. A. Boswell, R. A. Everley, C. Liu, C. S. Walmsley, D. Juric, P. K. Sorger, Multiomics profiling establishes the polypharmacology of FDA-approved CDK4/6 inhibitors and the potential for differential clinical activity. *Cell Chemical Biology* **26**, 1067-80 e8 (2019). doi.org/10.1016/j.chembiol.2019.05.005 ncbi.nlm.nih.gov/pmc/PMC6936329
241. Y. F. Tao, N. N. Wang, L. X. Xu, Z. H. Li, X. L. Li et al., Molecular mechanism of G1 arrest and cellular senescence induced by LEE011, a novel CDK4/CDK6 inhibitor, in leukemia cells. *Cancer Cell International* **17**, 35 (2017). doi.org/10.1186/s12935-017-0405-y ncbi.nlm.nih.gov/pmc/PMC5340031
242. D. A. Schaer, R. P. Beckmann, J. A. Dempsey, L. Huber, A. Forest et al., The CDK4/6 inhibitor abemaciclib induces a T cell inflamed tumor microenvironment and enhances the efficacy of PD-L1 checkpoint blockade. *Cell Reports* **22**, 2978-94 (2018). doi.org/10.1016/j.celrep.2018.02.053
243. M. Dowless, C. D. Lowery, T. Shackleford, M. Renschler, J. Stephens, R. Flack, W. Blosser, S. Gupta, J. Stewart, Y. Webster, J. Dempsey, A. B. VanWye, P. Ebert, P. Iversen, J. B. Olsen, X. Gong, S. Buchanan, P. Houghton, L. Stancato, Abemaciclib is active in preclinical models of Ewing sarcoma via multipronged regulation of cell cycle, DNA methylation, and interferon pathway signaling. *Clinical Cancer Research* **24**, 6028-39 (2018). doi.org/10.1158/1078-0432.CCR-18-1256 ncbi.nlm.nih.gov/pmc/PMC6279561
244. S. Chen, J. L. Blank, T. Peters, X. J. Liu, D. M. Rappoli et al., Genome-wide siRNA screen for modulators of cell death induced by proteasome inhibitor bortezomib. *Cancer Research* **70**, 4318-26 (2010). doi.org/10.1158/0008-5472.CAN-09-4428
245. S. Armeanu, M. Krusch, K. M. Baltz, T. S. Weiss, I. Smirnow, A. Steinle, U. M. Lauer, M. Bitzer, H. R. Salih, Direct and natural killer cell-mediated antitumor effects of low-dose bortezomib in hepatocellular carcinoma. *Clinical Cancer Research* **14**, 3520-8 (2008). doi.org/10.1158/1078-0432.CCR-07-4744
246. L. Gong, B. Yang, M. Xu, B. Cheng, X. Tang, P. Zheng, Y. Jing, G. J. Wu, Bortezomib-induced apoptosis in cultured pancreatic cancer cells is associated with ceramide production. *Cancer Chemotherapy and Pharmacology* **73**, 69-77 (2014). doi.org/10.1007/s00280-013-2318-3
247. X. Bao, T. Ren, Y. Huang, C. Ren, K. Yang, H. Zhang, W. Guo, Bortezomib induces apoptosis and suppresses cell growth and metastasis by inactivation of Stat3 signaling in chondrosarcoma. *International Journal of Oncology* **50**, 477-86 (2017). doi.org/10.3892/ijo.2016.3806
248. A. Fribley, Q. Zeng, C. Y. Wang, Proteasome inhibitor PS-341 induces apoptosis through induction of endoplasmic reticulum stress-reactive oxygen species in head and neck squamous cell carcinoma cells. *Molecular and Cellular Biology* **24**, 9695-704 (2004).
249. F. Baran-Marszak, M. Boukhiar, S. Harel, C. Laguillier, C. Roger, R. Gressin, A. Martin, R. Fagard, N. Varin-Blank, F. Ajchenbaum-Cymbalista, D. Ledoux, Constitutive and B-cell receptor-induced activation of STAT3 are important signaling pathways targeted by bortezomib in leukemic mantle cell lymphoma. *Haematologica* **95**, 1865-72 (2010). doi.org/10.3324/haematol.2009.019745 ncbi.nlm.nih.gov/pmc/PMC2966908
250. A. Kukreja, A. Hutchinson, A. Mazumder, D. Vesole, R. Angitapalli, S. Jagannath, A. O'Connor O, M. V. Dhodapkar, Bortezomib disrupts tumour-dendritic cell interactions in myeloma and lymphoma: Therapeutic implications. *British Journal of Haematology* **136**, 106-10 (2007). doi.org/10.1111/j.1365-2141.2006.06369.x
251. X. Zhao, W. Qiu, J. Kung, X. Zhao, X. Peng, M. Yegappan, B. Yen-Lieberman, E. D. Hsi, Bortezomib induces caspase-dependent apoptosis in Hodgkin lymphoma cell lines and is associated with reduced c-FLIP expression: A gene expression profiling study with implications for potential combination therapies. *Leukemia Research* **32**, 275-85 (2008). doi.org/10.1016/j.leukres.2007.05.024
252. P. Bavi, S. Uddin, M. Ahmed, Z. Jehan, R. Bu, J. Abubaker, M. Sultana, N. Al-Sanea, A. Abduljabbar, L. H. Ashari, S. Alhomoud, F. Al-Dayel, S. Prabhakaran, A. R. Hussain, K. S. Al-Kuraya, Bortezomib stabilizes mitotic cyclins and prevents cell cycle progression via inhibition of UBE2C in colorectal carcinoma. *American Journal of Pathology* **178**, 2109-20 (2011). doi.org/10.1016/j.ajpath.2011.01.034 ncbi.nlm.nih.gov/pmc/PMC3081207
253. N. Ao, Y. Dai, Q. Chen, Y. Feng, J. Yu, C. Wang, F. Liu, M. Li, G. Liu, Genome-wide profiling of the toxic effect of bortezomib on human esophageal carcinoma epithelial cells. *Technology in Cancer Research & Treatment* **18**, 153033819842546 (2019). doi.org/10.1177/153033819842546 ncbi.nlm.nih.gov/pmc/PMC6457034
254. M. Carlsten, A. Namazi, R. Reger, E. Levy, M. Berg, C. St Hilaire, R. W. Childs, Bortezomib sensitizes multiple myeloma to NK cells via ER-stress-induced suppression of HLA-E and upregulation of DR5. *Oncoimmunology* **8**, e1534664 (2019). doi.org/10.1080/2162402X.2018.1534664 ncbi.nlm.nih.gov/pmc/PMC6343814
255. J. Shi, G. J. Tricot, T. K. Garg, P. A. Malaviarachchi, S. M. Szmania, R. E. Kellum, B. Storrie, A. Mulder, J. D. Shaughnessy, Jr., B. Barlogie, F. van Rhee, Bortezomib down-regulates the cell-surface expression of HLA class I and enhances natural killer cell-mediated lysis of myeloma. *Blood* **111**, 1309-17 (2008). doi.org/10.1182/blood-2007-03-078535 ncbi.nlm.nih.gov/pmc/PMC2214736
256. C. L. Chang, Y. T. Hsu, C. C. Wu, Y. C. Yang, C. Wang, T. C. Wu, C. F. Hung, Immune mechanism of the antitumor effects generated by bortezomib. *Journal of Immunology* **189**, 3209-20 (2012). doi.org/10.4049/jimmunol.1103826
257. L. H. Mattingly, R. A. Gault, W. J. Murphy, Use of systemic proteasome inhibition as an immune-modulating agent in disease. *Endocrine, Metabolic and Immune Disorders - Drug Targets* **7**, 29-34 (2007). doi.org/10.2174/187153007780059397
258. M. Mohty, E. Brissot, B. N. Savani, B. Gaugler, Effects of bortezomib on the immune system: A focus on immune regulation. *Biology of Blood and Marrow Transplantation* **19**, 1416-20 (2013). doi.org/10.1016/j.bbmt.2013.05.011

259. Y. Y. Chu, C. Y. Ko, S. M. Wang, P. I. Lin, H. Y. Wang, W. C. Lin, D. Y. Wu, L. H. Wang, J. M. Wang, Bortezomib-induced miRNAs direct epigenetic silencing of locus genes and trigger apoptosis in leukemia. *Cell Death & Disease* **8**, e3167 (2017). doi.org/10.1038/cddis.2017.520 ncbi.nlm.nih.gov/pmc/PMC5775404
260. X. Ci, B. Li, X. Ma, F. Kong, C. Zheng, M. Bjorkholm, J. Jia, D. Xu, Bortezomib-mediated down-regulation of telomerase and disruption of telomere homeostasis contributes to apoptosis of malignant cells. *Oncotarget* **6**, 38079-92 (2015). doi.org/10.1863/oncotarget.5752 ncbi.nlm.nih.gov/pmc/PMC4741985
261. C. Weiss, O. Uziel, O. Wolach, J. Nordenberg, E. Beery, S. Bulvick, G. Kanfer, O. Cohen, R. Ram, M. Bakhanashvili, H. Magen-Nativ, N. Shilo, M. Lahav, Differential downregulation of telomerase activity by bortezomib in multiple myeloma cells-multiple regulatory pathways in vitro and ex vivo. *British Journal of Cancer* **107**, 1844-52 (2012). doi.org/10.1038/bjc.2012.460 ncbi.nlm.nih.gov/pmc/PMC3504947
262. H. Dong, L. Chen, X. Chen, H. Gu, G. Gao, Y. Gao, B. Dong, Dysregulation of unfolded protein response partially underlies proapoptotic activity of bortezomib in multiple myeloma cells. *Leukemia & Lymphoma* **50**, 974-84 (2009). doi.org/10.1080/10428190902895780
263. Z. X. Du, H. Y. Zhang, X. Meng, Y. Guan, H. Q. Wang, Role of oxidative stress and intracellular glutathione in the sensitivity to apoptosis induced by proteasome inhibitor in thyroid cancer cells. *BMC Cancer* **9**, 56 (2009). doi.org/10.1186/1471-2407-9-56 ncbi.nlm.nih.gov/pmc/PMC2666756
264. S. K. Edwards, Y. Han, Y. Liu, B. Z. Kreider, Y. Liu, S. Grewal, A. Desai, J. Baron, C. R. Moore, C. Luo, P. Xie, Signaling mechanisms of bortezomib in TRAF3-deficient mouse B lymphoma and human multiple myeloma cells. *Leukemia Research* **41**, 85-95 (2016). doi.org/10.1016/j.leukres.2015.12.005 ncbi.nlm.nih.gov/pmc/PMC4740239
265. P. Gomez-Bougie, S. Wuilleme-Toumi, E. Menoret, V. Trichet, N. Robillard, M. Philippe, R. Bataille, M. Amiot, Noxa up-regulation and Mcl-1 cleavage are associated to apoptosis induction by bortezomib in multiple myeloma. *Cancer Research* **67**, 5418-24 (2007). doi.org/10.1158/0008-5472.CAN-06-4322
266. A. M. Fribley, B. Evenchik, Q. Zeng, B. K. Park, J. Y. Guan, H. Zhang, T. J. Hale, M. S. Soengas, R. J. Kaufman, C. Y. Wang, Proteasome inhibitor PS-341 induces apoptosis in cisplatin-resistant squamous cell carcinoma cells by induction of Noxa. *Journal of Biological Chemistry* **281**, 31440-7 (2006). doi.org/10.1074/jbc.M604356200
267. M. Maynadier, J. Shi, O. Vaillant, M. Gary-Bobo, I. Basile, M. Gleizes, A. M. Cathiard, J. L. T. Wah, M. S. Sheikh, M. Garcia, Roles of estrogen receptor and p21Waf1 in bortezomib-induced growth inhibition in human breast cancer cells. *Molecular Cancer Research* **10**, 1473-81 (2012). doi.org/10.1158/1541-7786.MCR-12-0133
268. K. Podar, S. L. Gouill, J. Zhang, J. T. Opferman, E. Zorn, Y. T. Tai, T. Hideshima, M. Amiot, D. Chauhan, J. L. Harousseau, K. C. Anderson, A pivotal role for Mcl-1 in Bortezomib-induced apoptosis. *Oncogene* **27**, 721-31 (2008). doi.org/10.1038/sj.onc.1210679
269. M. Ri, S. Iida, T. Ishida, A. Ito, H. Yano, A. Inagaki, J. Ding, S. Kusumoto, H. Komatsu, A. Utsunomiya, R. Ueda, Bortezomib-induced apoptosis in mature T-cell lymphoma cells partially depends on upregulation of Noxa and functional repression of Mcl-1. *Cancer Science* **100**, 341-8 (2009). doi.org/10.1111/j.1349-7006.2008.01038.x
270. B. Z. Yuan, J. Chapman, S. H. Reynolds, Proteasome inhibitors induce apoptosis in human lung cancer cells through a positive feedback mechanism and the subsequent Mcl-1 protein cleavage. *Oncogene* **28**, 3775-86 (2009). doi.org/10.1038/onc.2009.240
271. J. Fang, G. Rhyasen, L. Bolanos, C. Rasch, M. Varney, M. Wunderlich, S. Goyama, G. Jansen, J. Cloos, C. Rigolino, A. Cortelezzi, J. C. Mulloy, E. N. Oliva, M. Cuzzola, D. T. Starczynowski, Cytotoxic effects of bortezomib in myelodysplastic syndrome/acute myeloid leukemia depend on autophagy-mediated lysosomal degradation of TRAF6 and repression of PSMA 1. *Blood* **120**, 858-67 (2012). doi.org/10.1182/blood-2012-02-407999 ncbi.nlm.nih.gov/pmc/PMC3412348
272. L. H. Fu, L. Wang, Y. C. Yu, C. L. Hu, L. Chen, Influence of Bortezomib on apoptosis of acute myelogenous leukemia cells as well as on SALL4 gene and Wnt/β-catenin signaling pathway. *Chinese Journal of Biologicals* **25**, 458-61+68 (2012).
273. G. Ayala, J. Yan, R. Li, Y. Ding, T. C. Thompson, M. P. Mims, T. G. Hayes, V. MacDonnell, R. G. Lynch, A. Frolov, B. J. Miles, T. M. Wheeler, J. W. Harper, M. J. Tsai, M. M. Ittmann, D. Kadmon, Bortezomib-mediated inhibition of steroid receptor coactivator-3 degradation leads to activated Akt. *Clinical Cancer Research* **14**, 7511-8 (2008). doi.org/10.1158/1078-0432.CCR-08-0839 ncbi.nlm.nih.gov/pmc/PMC2820291
274. H. Gu, X. Chen, G. Gao, H. Dong, Caspase-2 functions upstream of mitochondria in endoplasmic reticulum stress-induced apoptosis by bortezomib in human myeloma cells. *Molecular Cancer Therapeutics* **7**, 2298-307 (2008). doi.org/10.1158/1535-7163.MCT-08-0186
275. T. Hideshima, D. Chauhan, T. Hayashi, M. Akiyama, N. Mitsiades, C. Mitsiades, K. Podar, N. C. Munshi, P. G. Richardson, K. C. Anderson, Proteasome inhibitor PS-341 abrogates IL-6 triggered signaling cascades via caspase-dependent downregulation of gp130 in multiple myeloma. *Oncogene* **22**, 8386-93 (2003). doi.org/10.1038/sj.onc.1207170
276. Y. S. Hong, S. W. Hong, S. M. Kim, D. H. Jin, J. S. Shin, D. H. Yoon, K. P. Kim, J. L. Lee, D. S. Heo, J. S. Lee, T. W. Kim, Bortezomib induces G2-M arrest in human colon cancer cells through ROS-inducible phosphorylation of ATM-CHK1. *International Journal of Oncology* **41**, 76-82 (2012). doi.org/10.3892/ijo.2012.1448
277. W. Hu, W. Wang, Z. Gao, The influence of bortezomib on migration and invasion in cultured human prostatic cancer cells and its mechanism. *Cancer Research and Clinic* **26**, 98-101 (2014).
278. T. K. Huynh, C. Y. Ho, C. H. Tsai, C. K. Wang, Y. J. Chen, D. T. Bau, C. Y. Tu, T. S. Li, W. C. Huang, Proteasome inhibitors suppress ErbB family expression through HSP90-mediated lysosomal degradation. *International Journal of Molecular Sciences* **20**, (2019). doi.org/10.3390/ijms20194812 ncbi.nlm.nih.gov/pmc/PMC6801459
279. T. Ikezoe, Y. Yang, T. Saito, H. P. Koeffler, H. Taguchi, Proteasome inhibitor PS-341 down-regulates prostate-specific antigen (PSA) and induces growth arrest and apoptosis of androgen-

- dependent human prostate cancer LNCaP cells. *Cancer Science* **95**, 271-5 (2004). doi.org/10.1111/j.1349-7006.2004.tb02215.x
280. K. Kuroda, H. Liu, The proteasome inhibitor, bortezomib, induces prostate cancer cell death by suppressing the expression of prostate-specific membrane antigen, as well as androgen receptor. *International Journal of Oncology* **54**, 1357-66 (2019). doi.org/10.3892/ijo.2019.4706
281. L. Jardine, S. Hambleton, V. Bigley, S. Pagan, X. N. Wang, M. Collin, Sensitizing primary acute lymphoblastic leukemia to natural killer cell recognition by induction of NKG2D ligands. *Leukemia & Lymphoma* **54**, 167-73 (2013). doi.org/10.3109/10428194.2012.708026 ncbi.nlm.nih.gov/pmc/PMC6588533
282. C. Larrue, E. Saland, H. Boutzen, F. Vergez, M. David, C. Joffre, M. A. Hospital, J. Tamburini, E. Delabesse, S. Manenti, J. E. Sarry, C. Récher, Proteasome inhibitors induce FLT3-ITD degradation through autophagy in AML cells. *Blood* **127**, 882-92 (2016). doi.org/10.1182/blood-2015-05-646497
283. D. Belloni, L. Veschini, C. Foglieni, G. Dell'Antonio, F. Caligaris-Cappio, M. Ferrarini, E. Ferrero, Bortezomib induces autophagic death in proliferating human endothelial cells. *Experimental Cell Research* **316**, 1010-8 (2010). doi.org/10.1016/j.yexcr.2009.11.005
284. M. Politou, K. Naresh, E. Terpos, D. Crawley, I. Lampert, J. F. Apperley, A. Rahemtulla, Anti-angiogenic effect of bortezomib in patients with multiple myeloma. *Acta Haematologica* **114**, 170-3 (2005). doi.org/10.1159/000087894
285. J. B. Hamner, P. V. Dickson, T. L. Sims, J. Zhou, Y. Spence, C. Y. Ng, A. M. Davidoff, Bortezomib inhibits angiogenesis and reduces tumor burden in a murine model of neuroblastoma. *Surgery* **142**, 185-91 (2007). doi.org/10.1016/j.surg.2007.04.012
286. J. B. Sunwoo, Z. Chen, G. Dong, N. Yeh, C. C. Bancroft, E. Sausville, J. Adams, P. Elliott, C. Van Waes, Novel proteasome inhibitor PS-341 inhibits activation of nuclear factor-κB, cell survival, tumor growth, and angiogenesis in squamous cell carcinoma. *Clinical Cancer Research* **7**, 1419-28 (2001).
287. G. M. Jiang, H. S. Wang, J. Du, W. F. Ma, H. Wang, Y. Qiu, Q. G. Zhang, W. Xu, H. F. Liu, J. P. Liang, Bortezomib relieves immune tolerance in nasopharyngeal carcinoma via STAT1 suppression and indoleamine 2,3-dioxygenase downregulation. *Cancer Immunology Research* **5**, 42-51 (2017). doi.org/10.1158/2326-6066.CIR-16-0102
288. T. H. Landowski, C. J. Megli, K. D. Nullmeyer, R. M. Lynch, R. T. Dorr, Mitochondrial-mediated disregulation of Ca²⁺ is a critical determinant of velcade (PS-341/Bortezomib) cytotoxicity in myeloma cell lines. *Cancer Research* **65**, 3828-36 (2005). doi.org/10.1158/0008-5472.CAN-04-3684
289. C. Li, R. Li, J. R. Grandis, D. E. Johnson, Bortezomib induces apoptosis via Bim and Bik up-regulation and synergizes with cisplatin in the killing of head and neck squamous cell carcinoma cells. *Molecular Cancer Therapeutics* **7**, 1647-55 (2008). doi.org/10.1158/1535-7163.MCT-07-2444 ncbi.nlm.nih.gov/pmc/PMC2534142
290. H. Zhu, W. Guo, L. Zhang, S. Wu, F. Teraishi, J. J. Davis, F. Dong, B. Fang, Proteasome inhibitors-mediated TRAIL resensitization and Bik accumulation. *Cancer Biology & Therapy* **4**, 781-6 (2005). doi.org/10.4161/cbt.4.7.1897 ncbi.nlm.nih.gov/pmc/PMC1592469
291. J. Li, X. Zhang, J. Shen, J. Guo, X. Wang, J. Liu, Bortezomib promotes apoptosis of multiple myeloma cells by regulating HSP27. *Molecular Medicine Reports* **20**, 2410-8 (2019). doi.org/10.3892/mmr.2019.10467
292. K. Podar, R. Shringarpure, Y. T. Tai, M. Simoncini, M. Sattler, K. Ishitsuka, P. G. Richardson, T. Hideshima, D. Chauhan, K. C. Anderson, Caveolin-1 is required for vascular endothelial growth factor-triggered multiple myeloma cell migration and is targeted by bortezomib. *Cancer Research* **64**, 7500-6 (2004). doi.org/10.1158/0008-5472.CAN-04-0124
293. A. Yerlikaya, E. Okur, A. T. Baykal, C. Acilan, I. Boyaci, E. Ulukaya, A proteomic analysis of p53-independent induction of apoptosis by bortezomib in 4T1 breast cancer cell line. *Journal of Proteomics* **113**, 315-25 (2015). doi.org/10.1016/j.jprot.2014.09.010
294. Y. Shen, L. Lu, J. Xu, W. Meng, Y. Qing, Y. Liu, B. Zhang, H. Hu, Bortezomib induces apoptosis of endometrial cancer cells through microRNA-17-5p by targeting p21. *Cell Biology International* **37**, 1114-21 (2013). doi.org/10.1002/cbin.10139
295. L. Lin, D. Chen, Z. F. Xiang, R. Z. Pei, P. S. Zhang, X. H. Liu, X. H. Du, Y. Lu, Bortezomib could down-regulate the expression of RANKL, inhibit cell proliferation and induce cell apoptosis in the human myeloma cell line RPMI 8226 by activating caspase-3. *Cancer Biomarkers* **20**, 217-24 (2017). doi.org/10.3233/CBM-170584
296. Y. C. Lin, K. C. Chen, C. C. Chen, A. L. Cheng, K. F. Chen, CIP2A-mediated Akt activation plays a role in bortezomib-induced apoptosis in head and neck squamous cell carcinoma cells. *Oral Oncology* **48**, 585-93 (2012). doi.org/10.1016/j.oraloncology.2012.01.012
297. Y. Ding, Y. Wang, S. Ju, X. Wu, W. Zhu, F. Shi, L. Mao, Role of CIP2A in the antitumor effect of bortezomib in colon cancer. *Molecular Medicine Reports* **10**, 387-92 (2014). doi.org/10.3892/mmr.2014.2173
298. L. M. Tseng, C. Y. Liu, K. C. Chang, P. Y. Chu, C. W. Shiao, K. F. Chen, CIP2A is a target of bortezomib in human triple negative breast cancer cells. *Breast Cancer Research* **14**, R68 (2012). doi.org/10.1186/bcr3175 ncbi.nlm.nih.gov/pmc/PMC3446403
299. H. C. Yu, D. R. Hou, C. Y. Liu, C. S. Lin, C. W. Shiao, A. L. Cheng, K. F. Chen, Cancerous inhibitor of protein phosphatase 2A mediates bortezomib-induced autophagy in hepatocellular carcinoma independent of proteasome. *PloS One* **8**, e55705 (2013). doi.org/10.1371/journal.pone.0055705 ncbi.nlm.nih.gov/pmc/PMC3562236
300. P. Luo, M. Lin, M. Lin, D. Zhu, Z. Wang, J. Shen, B. Yang, Q. He, Bortezomib induces apoptosis in human neuroblastoma CHP126 cells. *Pharmazie* **65**, 213-8 (2010).
301. M. Nara, K. Teshima, A. Watanabe, M. Ito, K. Iwamoto, A. Kitabayashi, M. Kume, Y. Hatano, N. Takahashi, S. Iida, K. Sawada, H. Tagawa, Bortezomib reduces the tumorigenicity of multiple myeloma via downregulation of upregulated targets in clonogenic side population cells. *PloS One* **8**, e56954 (2013). doi.org/10.1371/journal.pone.0056954 ncbi.nlm.nih.gov/pmc/PMC3587640

302. M. Lioni, K. Noma, A. Snyder, A. Klein-Szanto, J. A. Diehl, A. K. Rustgi, M. Herlyn, K. S. M. Smalley, Bortezomib induces apoptosis in esophageal squamous cell carcinoma cells through activation of the p38 mitogen-activated protein kinase pathway. *Molecular Cancer Therapeutics* **7**, 2866-75 (2008). doi.org/10.1158/1535-7163.MCT-08-0391 ncbi.nlm.nih.gov/pmc/PMC2903039
303. S. Ohshima-Hosoyama, M. A. Davare, T. Hosoyama, L. D. Nelon, C. Keller, Bortezomib stabilizes NOXA and triggers ROS-associated apoptosis in medulloblastoma. *Journal of Neuro-Oncology* **105**, 475-83 (2011). doi.org/10.1007/s11060-011-0619-0
304. P. Pérez-Galán, G. Roue, N. Villamor, E. Montserrat, E. Campo, D. Colomer, The proteasome inhibitor bortezomib induces apoptosis in mantle-cell lymphoma through generation of ROS and Noxa activation independent of p53 status. *Blood* **107**, 257-64 (2006). doi.org/10.1182/blood-2005-05-2091
305. S. Periyasamy-Thandavan, W. H. Jackson, J. S. Samaddar, B. Erickson, J. R. Barrett, L. Raney, E. Gopal, V. Ganapathy, W. D. Hill, K. N. Bhalla, P. V. Schoenlein, Bortezomib blocks the catabolic process of autophagy via a cathepsin-dependent mechanism, affects endoplasmic reticulum stress and induces caspase-dependent cell death in antiestrogen-sensitive and resistant ER+ breast cancer cells. *Autophagy* **6**, 19-35 (2010). doi.org/10.4161/auto.6.1.10323
306. F. Teraishi, W. Guo, L. Zhang, F. Dong, J. J. Davis, T. Sasazuki, S. Shirasawa, J. Liu, B. Fang, Activation of sterile20-like kinase 1 in proteasome inhibitor bortezomib-induced apoptosis in oncogenic K-ras-transformed cells. *Cancer Research* **66**, 6072-9 (2006). doi.org/10.1158/0008-5472.CAN-06-0125 ncbi.nlm.nih.gov/pmc/PMC1482805
307. G. L. Powers, P. Rajbhandari, N. M. Solodin, B. Bickford, E. T. Alarid, The proteasome inhibitor bortezomib induces an inhibitory chromatin environment at a distal enhancer of the estrogen receptor- α gene. *PloS One* **8**, (2013). doi.org/10.1371/journal.pone.0081110 ncbi.nlm.nih.gov/pmc/PMC3855213
308. D. Tamura, T. Arao, K. Tanaka, H. Kaneda, K. Matsumoto, K. Kudo, K. Aomatsu, Y. Fujita, T. Watanabe, N. Saijo, Y. Kotani, Y. Nishimura, K. Nishio, Bortezomib potentially inhibits cellular growth of vascular endothelial cells through suppression of G2/M transition. *Cancer Science* **101**, 1403-8 (2010). doi.org/10.1111/j.1349-7006.2010.01544.x
309. R. Wang, J. Shen, Q. Wang, M. Zhang, Bortezomib inhibited the progression of diffuse large B-cell lymphoma via targeting miR-198. *Biomedicine and Pharmacotherapy* **108**, 43-9 (2018). doi.org/10.1016/j.biopha.2018.08.151
310. R. Wang, A. M. Davidoff, L. M. Pfeffer, Bortezomib sensitizes human glioblastoma cells to induction of apoptosis by type I interferons through NOXA expression and Mcl-1 cleavage. *Biochemical and Biophysical Research Communications* **478**, 128-34 (2016). ncbi.nlm.nih.gov/pmc/PMC4991636
311. S. T. Pellom, Jr., D. F. Dudimah, M. C. Thounaojam, R. V. Uzhachenko, A. Singhal, A. Richmond, A. Shanker, Bortezomib augments lymphocyte stimulatory cytokine signaling in the tumor microenvironment to sustain CD8+T cell antitumor function. *Oncotarget* **8**, 8604-21 (2017). doi.org/10.18632/oncotarget.14365 ncbi.nlm.nih.gov/pmc/PMC5352426
312. B. H. Yeung, D. C. Huang, F. A. Sinicrope, PS-341 (bortezomib) induces lysosomal cathepsin B release and a caspase-2-dependent mitochondrial permeabilization and apoptosis in human pancreatic cancer cells. *Journal of Biological Chemistry* **281**, 11923-32 (2006). doi.org/10.1074/jbc.M508533200
313. C. Y. Liu, F. S. Hsieh, P. Y. Chu, W. C. Tsai, C. T. Huang, Y. B. Yu, T. T. Huang, P. S. Ko, M. H. Hung, W. L. Wang, C. W. Shiao, K. F. Chen, Carfilzomib induces leukaemia cell apoptosis via inhibiting ELK1/KIAA1524 (Elk-1/CIP2A) and activating PP2A not related to proteasome inhibition. *British Journal of Haematology* **177**, 726-40 (2017). doi.org/10.1111/bjh.14620
314. M. Zangari, T. Berno, Y. Yang, M. Zeng, H. Xu, L. Pappas, G. Tricot, A. Kamalakar, D. Yoon, L. J. Suva, Parathyroid hormone receptor mediates the anti-myeloma effect of proteasome inhibitors. *Bone* **61**, 39-43 (2014). doi.org/10.1016/j.bone.2013.12.025 ncbi.nlm.nih.gov/pmc/PMC3967551
315. A. Figueiredo, K. L. Moore, J. Mak, N. Sluis-Cremer, M. P. de Bethune, G. Tachedjian, Potent nonnucleoside reverse transcriptase inhibitors target HIV-1 Gag-Pol. *PLoS Pathog* **2**, e119 (2006). doi.org/10.1371/journal.ppat.0020119 ncbi.nlm.nih.gov/pmc/PMC1635531
316. G. Tachedjian, K. L. Moore, S. P. Goff, N. Sluis-Cremer, Efavirenz enhances the proteolytic processing of an HIV-1 pol polyprotein precursor and reverse transcriptase homodimer formation. *FEBS Letters* **579**, 379-84 (2005). doi.org/10.1016/j.febslet.2004.11.099
317. S. Sudo, H. Haraguchi, Y. Hirai, H. Gatanaga, J. Sakuragi, F. Momose, Y. Morikawa, Efavirenz enhances HIV-1 gag processing at the plasma membrane through Gag-Pol dimerization. *Journal of Virology* **87**, 3348-60 (2013). doi.org/10.1128/JVI.02306-12 ncbi.nlm.nih.gov/pmc/PMC3592135
318. X. Li, X. Liu, R. Deng, S. Gao, H. Yu, K. Huang, Q. Jiang, R. Liu, X. Li, L. Zhang, H. Zhou, C. Yang, Nintedanib inhibits Wnt3a-induced myofibroblast activation by suppressing the Src/beta-catenin pathway. *Frontiers in Pharmacology* **11**, 310 (2020). doi.org/10.3389/fphar.2020.00310 ncbi.nlm.nih.gov/pmc/PMC7087487
319. F. Hilberg, U. Tontsch-Grunt, A. Baum, A. T. Le, R. C. Doebele, S. Lieb, D. Gianni, T. Voss, P. Garin-Chesa, C. Haslinger, N. Kraut, Triple angiokinase inhibitor nintedanib directly inhibits tumor cell growth and induces tumor shrinkage via blocking oncogenic receptor tyrosine kinases. *Journal of Pharmacology and Experimental Therapeutics* **364**, 494-503 (2018). doi.org/10.1124/jpet.117.244129 ncbi.nlm.nih.gov/pmc/PMC6040086
320. C. Y. Liu, T. T. Huang, P. Y. Chu, C. T. Huang, C. H. Lee, W. L. Wang, K. Y. Lau, W. C. Tsai, T. I. Chao, J. C. Su, M. H. Chen, C. W. Shiao, L. M. Tseng, K. F. Chen, The tyrosine kinase inhibitor nintedanib activates SHP-1 and induces apoptosis in triple-negative breast cancer cells. *Experimental and Molecular Medicine* **49**, e366 (2017). doi.org/10.1038/emm.2017.114 ncbi.nlm.nih.gov/pmc/PMC5579508
321. C. Overed-Sayer, E. Miranda, R. Dunmore, E. Liarte Marin, L. Beloki, D. Rassl, H. Parfrey, A. Carruthers, A. Chahboub, S. Koch, G. Guler-Gane, M. Kuziora, A. Lewis, L. Murray, R. May, D. Clarke, Inhibition of mast cells: A novel mechanism by which nintedanib may elicit anti-fibrotic effects. *Thorax* **75**, 754-63 (2020).

doi.org/10.1136/thoraxjnl-2019-214000

ncbi.nlm.nih.gov/pmc/PMC7476277

322. S. Rangarajan, A. Kurundkar, D. Kurundkar, K. Bernard, Y. Y. Sanders, Q. Ding, V. B. Antony, J. Zhang, J. Zmijewski, V. J. Thannickal, Novel mechanisms for the antifibrotic action of nintedanib. *American Journal of Respiratory Cell and Molecular Biology* **54**, 51-9 (2016). doi.org/10.1165/rcmb.2014-0445OC
ncbi.nlm.nih.gov/pmc/PMC4742925

323. L. Knuppel, Y. Ishikawa, M. Aichler, K. Heinzelmann, R. Hatz, J. Behr, A. Walch, H. P. Bachinger, O. Eickelberg, C. A. Staab-Weijnitz, A novel antifibrotic mechanism of nintedanib and pirfenidone. inhibition of collagen fibril assembly. *American Journal of Respiratory Cell and Molecular Biology* **57**, 77-90 (2017).
doi.org/10.1165/rcmb.2016-0217OC

324. M. I. Davis, J. P. Hunt, S. Herrgard, P. Ciceri, L. M. Wodicka, G. Pallares, M. Hocker, D. K. Treiber, P. P. Zarrinkar, Comprehensive analysis of kinase inhibitor selectivity. *Nature Biotechnology* **29**, 1046-51 (2011). doi.org/10.1038/nbt.1990

325. K. Stegmaier, S. M. Corsello, K. N. Ross, J. S. Wong, D. J. Deangelo, T. R. Golub, Gefitinib induces myeloid differentiation of acute myeloid leukemia. *Blood* **106**, 2841-8 (2005).
doi.org/10.1182/blood-2005-02-0488
ncbi.nlm.nih.gov/pmc/PMC1895296

326. E. Lindhagen, A. Eriksson, M. Wickstrom, K. Danielsson, B. Grundmark, R. Henriksson, P. Nygren, A. Aleskog, R. Larsson, M. Hoglund, Significant cytotoxic activity in vitro of the EGFR tyrosine kinase inhibitor gefitinib in acute myeloblastic leukaemia. *European Journal of Haematology* **81**, 344-53 (2008). doi.org/10.1111/j.1600-0609.2008.01120.x

327. S. H. Ahn, E. H. Jeong, T. G. Lee, S. Y. Kim, H. R. Kim, C. H. Kim, Gefitinib induces cytoplasmic translocation of the CDK inhibitor p27 and its binding to a cleaved intermediate of caspase 8 in non-small cell lung cancer cells. *Cell Oncol (Dordr)* **37**, 377-86 (2014). doi.org/10.1007/s13402-014-0198-0

328. H. Ariyama, B. Qin, E. Baba, R. Tanaka, K. Mitsugi, M. Harada, S. Nakano, Gefitinib, a selective EGFR tyrosine kinase inhibitor, induces apoptosis through activation of Bax in human gallbladder adenocarcinoma cells. *Journal of Cellular Biochemistry* **97**, 724-34 (2006). doi.org/10.1002/jcb.20678

329. D. Brehmer, Z. Greff, K. Godl, S. Blencke, A. Kurtenbach, M. Weber, S. Muller, B. Klebl, M. Cotten, G. Keri, J. Wissing, H. Daub, Cellular targets of gefitinib. *Cancer Research* **65**, 379-82 (2005).

330. L. Conradt, K. Godl, C. Schaab, A. Tebbe, S. Eser, S. Diersch, C. W. Michalski, J. Kleeff, A. Schnieke, R. M. Schmid, D. Saur, G. Schneider, Disclosure of erlotinib as a multikinase inhibitor in pancreatic ductal adenocarcinoma. *Neoplasia* **13**, 1026-34 (2011).
doi.org/10.1593/neo.111016 ncbi.nlm.nih.gov/pmc/PMC3223607

331. C. Weber, T. B. Schreiber, H. Daub, Dual phosphoproteomics and chemical proteomics analysis of erlotinib and gefitinib interference in acute myeloid leukemia cells. *Journal of Proteomics* **75**, 1343-56 (2012). doi.org/10.1016/j.jprot.2011.11.004

332. C. Y. Chang, C. C. Shen, H. L. Su, C. J. Chen, Gefitinib induces apoptosis in human glioma cells by targeting Bad phosphorylation. *Journal of Neuro-Oncology* **105**, 507-22 (2011).
doi.org/10.1007/s11060-011-0632-3

333. C. Y. Chang, Y. H. Kuan, Y. C. Ou, J. R. Li, C. C. Wu, P. H. Pan, W. Y. Chen, H. Y. Huang, C. J. Chen, Autophagy contributes to gefitinib-induced glioma cell growth inhibition. *Experimental Cell Research* **327**, 102-12 (2014). doi.org/10.1016/j.yexcr.2014.05.011

334. G. C. Chang, S. L. Hsu, J. R. Tsai, F. P. Liang, S. Y. Lin, G. T. Sheu, C. Y. Chen, Molecular mechanisms of ZD1839-induced G1-cell cycle arrest and apoptosis in human lung adenocarcinoma A549 cells. *Biochemical Pharmacology* **68**, 1453-64 (2004).
doi.org/10.1016/j.bcp.2004.06.006

335. G. C. Chang, C. T. R. Yu, C. H. Tsai, J. R. Tsai, J. C. Chen, C. C. Wu, W. J. Wu, S. L. Hsu, An epidermal growth factor inhibitor, Gefitinib, induces apoptosis through a p53-dependent upregulation of pro-apoptotic molecules and downregulation of anti-apoptotic molecules in human lung adenocarcinoma A549 cells. *European Journal of Pharmacology* **600**, 37-44 (2008).
doi.org/10.1016/j.ejphar.2008.10.024

336. E. Di Gennaro, M. Barbarino, F. Buzzese, S. De Lorenzo, M. Caraglia, A. Abbruzzese, A. Avallone, P. Comella, F. Caponigro, S. Pepe, A. Budillon, Critical role of both p27KIP1 and p21CIP1/WAF1 in the antiproliferative effect of ZD1839 ('Iressa'), an epidermal growth factor receptor tyrosine kinase inhibitor, in head and neck squamous carcinoma cells. *Journal of Cellular Physiology* **195**, 139-50 (2003). doi.org/10.1002/jcp.10239

337. J. S. Guillamo, S. de Bouard, S. Valable, L. Marteau, P. Leuraud, Y. Marie, M. F. Poupon, J. J. Parienti, E. Raymond, M. Peschanski, Molecular mechanisms underlying effects of epidermal growth factor receptor inhibition on invasion, proliferation, and angiogenesis in experimental glioma. *Clinical Cancer Research* **15**, 3697-704 (2009). doi.org/10.1158/1078-0432.CCR-08-2042

338. S. He, T. Yin, D. Li, X. Gao, Y. Wan, X. Ma, T. Ye, F. Guo, J. Sun, Z. Lin, Y. Wang, Enhanced interaction between natural killer cells and lung cancer cells: Involvement in gefitinib-mediated immunoregulation. *Journal of Translational Medicine* **11**, (2013).
doi.org/10.1186/1479-5876-11-186
ncbi.nlm.nih.gov/pmc/PMC3766712

339. R. Okita, D. Wolf, K. Yasuda, A. Maeda, T. Yukawa, S. Saisho, K. Shimizu, Y. Yamaguchi, M. Oka, E. Nakayama, A. Lundqvist, R. Kiessling, B. Seliger, M. Nakata, Contrasting effects of the cytotoxic anticancer drug gemcitabine and the EGFR tyrosine kinase inhibitor gefitinib on NK cell-mediated cytotoxicity via regulation of NKG2D ligand in non-small-cell lung cancer cells. *PloS One* **10**, e0139809 (2015). doi.org/10.1371/journal.pone.0139809
ncbi.nlm.nih.gov/pmc/PMC4595469

340. H. Hu, L. Hao, B. Yan, X. Li, Y. Zhu, J. Yao, G. Wang, Q. Jiang, Gefitinib inhibits retina angiogenesis by affecting VEGF signaling pathway. *Biomedicine and Pharmacotherapy* **102**, 115-9 (2018). doi.org/10.1016/j.biopha.2018.02.110

341. R. Inoue, H. Matsuyama, S. Yano, Y. Yamamoto, N. Iizuka, K. Naito, Gefitinib-related gene signature in bladder cancer cells identified by a cDNA microarray. *Anticancer Research* **26**, 4195-202 (2006).

342. K. R. Jin, J. C. Yun, B. Y. Ryoo, I. N. Im, H. Y. Sung, H. K. Cheol, C. L. Jae, p53 enhances gefitinib-induced growth inhibition and apoptosis by regulation of Fas in non-small cell lung cancer. *Cancer Research* **67**, 1163-9 (2007). doi.org/10.1158/0008-5472.CAN-06-2037

343. S. Kanda, A. Naba, Y. Miyata, Inhibition of endothelial cell chemotaxis toward FGF-2 by gefitinib associates with downregulation of Fes activity. *International Journal of Oncology* **35**, 1305-12 (2009). doi.org/10.3892/ijo_00000448
344. S. Koyama, T. Omura, A. Yonezawa, S. Imai, S. Nakagawa, T. Nakagawa, I. Yano, K. Matsubara, Gefitinib and erlotinib lead to phosphorylation of eukaryotic initiation factor 2 alpha independent of epidermal growth factor receptor in A549 cells. *PloS One* **10**, e0136176 (2015). doi.org/10.1371/journal.pone.0136176 ncbi.nlm.nih.gov/pmc/PMC4545731
345. J. Klangas, D. Zardavas, J. Zhong, M. Roth, G. Karakiulakis, E. Papakonstantinou, The effect of the highly selective epidermal growth factor receptor tyrosine kinase inhibitor gefitinib on glycosaminoglycan synthesis by human malignant mesothelioma cells. *Epitheorese Klinikes Farmakologias kai Farmakokinetikas* **25**, 62-3 (2007).
346. J. Krol, R. E. Francis, A. Albergaria, A. Sunters, A. Polychronis, R. C. Coombes, E. W. F. Lam, The transcription factor FOXO3a is a crucial cellular target of gefitinib (Iressa) in breast cancer cells. *Molecular Cancer Therapeutics* **6**, 3169-79 (2007). doi.org/10.1158/1535-7163.MCT-07-0507
347. U. B. McGovern, R. E. Francis, B. Peck, S. K. Guest, J. Wang, S. S. Myatt, J. Krol, J. M. Kwok, A. Polychronis, R. C. Coombes, E. W. Lam, Gefitinib (Iressa) represses FOXM1 expression via FOXO3a in breast cancer. *Molecular Cancer Therapeutics* **8**, 582-91 (2009). doi.org/10.1158/1535-7163.MCT-08-0805
348. M. A. Fabian, W. H. Biggs, 3rd, D. K. Treiber, C. E. Atteridge, M. D. Azimioara et al., A small molecule-kinase interaction map for clinical kinase inhibitors. *Nature Biotechnology* **23**, 329-36 (2005). doi.org/10.1038/nbt1068
349. S. Shintani, C. Li, M. Mihara, K. I. Nakashiro, H. Hamakawa, Gefitinib ('Iressa'), an epidermal growth factor receptor tyrosine kinase inhibitor, mediates the inhibition of lymph node metastasis in oral cancer cells. *Cancer Letters* **201**, 149-55 (2003). [doi.org/10.1016/s0304-3835\(03\)00464-6](https://doi.org/10.1016/s0304-3835(03)00464-6)
350. T. C. Lin, Gefitinib modulates stress fibers and tubular-like structure formation and attenuates angiogenesis in an in vivo chicken model of chorioallantoic membrane angiogenesis. *Biochemical and Biophysical Research Communications* **526**, 568-73 (2020). doi.org/10.1016/j.bbrc.2020.03.102
351. D. O. Moon, M. O. Kim, J. D. Lee, Y. H. Choi, M. K. Lee, G. Y. Kim, Molecular mechanisms of ZD1839 (Iressa)-induced apoptosis in human leukemic U937 cells. *Acta Pharmacologica Sinica* **28**, 1205-14 (2007). doi.org/10.1111/j.1745-7254.2007.00615.x
352. I. D. O'Neill, Gefitinib as targeted therapy for mucoepidermoid carcinoma of the lung: Possible significance of CRTC1-MAML2 oncogene. *Lung Cancer* **64**, 129-30 (2009). doi.org/10.1016/j.lungcan.2009.01.003
353. J. L. Ouyang, N. J. Chen, Y. C. Liu, J. Q. Liao, H. Wei, H. Li, Antitumor activity and molecular mechanism of Gefitinib in human glioma cell line U251. *Chinese Journal of Cancer Prevention and Treatment* **20**, 21-6 (2013).
354. M. P. Piechocki, G. H. Yoo, S. K. Dibbley, E. H. Amjad, F. Lonardo, Iressa induces cytostasis and augments Fas-mediated apoptosis in acinic cell adenocarcinoma overexpressing HER2/neu. *International Journal of Cancer* **119**, 441-54 (2006). doi.org/10.1002/ijc.21837
355. Y. Sekiguchi, M. Yamada, T. Noguchi, C. Noomote, M. Tsuchida, Y. Kudoh, Y. Hirata, A. Matsuzawa, The anti-cancer drug gefitinib accelerates Fas-mediated apoptosis by enhancing caspase-8 activation in cancer cells. *Journal of Toxicological Sciences* **44**, 435-40 (2019). doi.org/10.2131/jts.44.435
356. J. Song, J. Zhu, Q. Zhao, B. Tian, Gefitinib causes growth arrest and inhibition of metastasis in human chondrosarcoma cells. *Journal of B.U.ON.* **20**, 894-901 (2015).
357. M. A. Sakurai, Y. Ozaki, D. Okuzaki, Y. Naito, T. Sasakura, A. Okamoto, H. Tabara, T. Inoue, M. Hagiya, A. Ito, N. Yabuta, H. Nojima, Gefitinib and luteolin cause growth arrest of human prostate cancer PC-3 cells via inhibition of cyclin G-associated kinase and induction of miR-630. *PloS One* **9**, e100124 (2014). doi.org/10.1371/journal.pone.0100124 ncbi.nlm.nih.gov/pmc/PMC4074034
358. D. Toda, T. Ota, K. Tsukuda, K. Watanabe, T. Fujiyama, M. Murakami, M. Naito, N. Shimizu, Gefitinib decreases the synthesis of matrix metalloproteinase and the adhesion to extracellular matrix proteins of colon cancer cells. *Anticancer Research* **26**, 129-34 (2006).
359. S. Shintani, C. Li, M. Mihara, J. Yano, N. Terakado, K. I. Nakashiro, H. Hamakawa, Gefitinib ('Iressa', ZD1839), an epidermal growth factor receptor tyrosine kinase inhibitor, up-regulates p27KIP1 and induces G1 arrest in oral squamous cell carcinoma cell lines. *Oral Oncology* **40**, 43-51 (2004). [doi.org/10.1016/s1368-8375\(03\)00131-3](https://doi.org/10.1016/s1368-8375(03)00131-3)
360. M. Suenaga, A. Yamaguchi, H. Soda, K. Orihara, Y. Tokito, Y. Sakaki, M. Umehara, K. Terashi, N. Kawamata, M. Oka, S. Kohno, C. Tei, Antiproliferative effects of gefitinib are associated with suppression of E2F-1 expression and telomerase activity. *Anticancer Research* **26**, 3387-91 (2006).
361. Y. K. Tak, P. K. Naoghare, E. Han, J. M. Song, VEGF inhibitor (Iressa) arrests histone deacetylase expression: Single-cell cotransfection imaging cytometry for multi-target-multi-drug analysis. *Journal of Cellular Physiology* **226**, 2115-22 (2011). doi.org/10.1002/jcp.22540
362. X. Wan, Y. Zhu, L. Zhang, W. Hou, Gefitinib inhibits malignant melanoma cells through the VEGF/AKT signaling pathway. *Molecular Medicine Reports* **17**, 7351-5 (2018). doi.org/10.3892/mmr.2018.8728
363. M. Yadav, A. K. Singh, H. Kumar, G. Rao, B. Chakravarti, A. Gurjar, S. Dogra, S. Kushwaha, A. L. Vishwakarma, P. N. Yadav, D. Datta, A. K. Tripathi, N. Chattopadhyay, A. K. Trivedi, S. Sanyal, Epidermal growth factor receptor inhibitor cancer drug gefitinib modulates cell growth and differentiation of acute myeloid leukemia cells via histamine receptors. *Biochimica et Biophysica Acta* **1860**, 2178-90 (2016). doi.org/10.1016/j.bbagen.2016.05.011
364. D. Yan, Y. Ge, H. Deng, W. Chen, G. An, Gefitinib upregulates death receptor 5 expression to mediate rmh TRAIL-induced apoptosis in Gefitinib-sensitive NSCLC cell line. *Oncotargets and Therapy* **8**, 1603-10 (2015). doi.org/10.2147/OTT.S73731 ncbi.nlm.nih.gov/pmc/PMC4498723
365. H. Yi, S. Li, H. Li, P. Wang, H. Zheng, X. Cheng, Gefitinib induces non-small cell lung cancer H1650 cell apoptosis through

- downregulating tumor necrosis factor-related apoptosis-inducing ligand expression levels. *Oncology Letters* **16**, 4768-72 (2018). doi.org/10.3892/ol.2018.9162 ncbi.nlm.nih.gov/pmc/PMC6126167
366. Y. F. Zhao, C. R. Wang, Y. M. Wu, S. L. Ma, Y. Ji, Y. J. Lu, P21 (wafl/cip1) is required for non-small cell lung cancer sensitive to Gefitinib treatment. *Biomedicine and Pharmacotherapy* **65**, 151-6 (2011). doi.org/10.1016/j.biopha.2011.02.009
367. G. Chan, M. Pilichowska, Complete remission in a patient with acute myelogenous leukemia treated with erlotinib for non small-cell lung cancer. *Blood* **110**, 1079-80 (2007). doi.org/10.1182/blood-2007-01-069856
368. S. Boehler, L. Ades, T. Braun, L. Galluzzi, J. Grosjean, C. Fabre, G. Le Roux, C. Gardin, A. Martin, S. de Botton, P. Fenoux, G. Kroemer, Erlotinib exhibits antineoplastic off-target effects in AML and MDS: A preclinical study. *Blood* **111**, 2170-80 (2008). doi.org/10.1182/blood-2007-07-100362
369. V. Pitini, C. Arrigo, G. Altavilla, Erlotinib in a patient with acute myelogenous leukemia and concomitant non-small-cell lung cancer. *Journal of Clinical Oncology* **26**, 3645-6 (2008). doi.org/10.1200/JCO.2008.17.0357
370. A. Augustin, J. Lamerz, H. Meistermann, S. Golling, S. Scheiblich, J. C. Hermann, G. Duchateau-Nguyen, M. Tzouros, D. W. Avila, H. Langen, L. ESSIOUTX, B. Klughammer, Quantitative chemical proteomics profiling differentiates erlotinib from gefitinib in EGFR wild-type non-small cell lung carcinoma cell lines. *Molecular Cancer Therapeutics* **12**, 520-9 (2013). doi.org/10.1158/1535-7163.MCT-12-0880
371. J. H. Bae, S. J. Kim, M. J. Kim, S. O. Oh, J. S. Chung, S. H. Kim, C. D. Kang, Susceptibility to natural killer cell-mediated lysis of colon cancer cells is enhanced by treatment with epidermal growth factor receptor inhibitors through UL16-binding protein-1 induction. *Cancer Science* **103**, 7-16 (2012). doi.org/10.1111/j.1349-7006.2011.02109.x
372. X. Qian, J. Li, J. Ding, Z. Wang, W. Zhang, G. Hu, Erlotinib activates mitochondrial death pathways related to the production of reactive oxygen species in the human non-small cell lung cancer cell line A549. *Clinical and Experimental Pharmacology and Physiology* **36**, 487-94 (2009). doi.org/10.1111/j.1440-1681.2008.05091.x
373. F. Shan, Z. Shao, S. Jiang, Z. Cheng, Erlotinib induces the human non-small-cell lung cancer cells apoptosis via activating ROS-dependent JNK pathways. *Cancer Medicine* **5**, 3166-75 (2016). doi.org/10.1002/cam4.881 ncbi.nlm.nih.gov/pmc/PMC5119972
374. Z. X. Cao, C. J. Guo, X. Song, J. L. He, L. Tan, S. Yu, R. Q. Zhang, F. Peng, C. Peng, Y. Z. Li, Erlotinib is effective against FLT3-ITD mutant AML and helps to overcome intratumoral heterogeneity via targeting FLT3 and Lyn. *FASEB Journal* **34**, 10182-90 (2020). doi.org/10.1096/fj.201902922RR
375. J. S. Im, A. C. Herrmann, C. Bernatchez, C. Haymaker, J. J. Molldrem, W. K. Hong, R. Perez-Soler, Immune-modulation by epidermal growth factor receptor inhibitors: Implication on anti-tumor immunity in lung cancer. *PloS One* **11**, e0160004 (2016). doi.org/10.1371/journal.pone.0160004 ncbi.nlm.nih.gov/pmc/PMC4965069
376. J. C. Ko, S. C. Ciou, J. Y. Jhan, C. M. Cheng, Y. J. Su, S. M. Chuang, S. T. Lin, C. C. Chang, Y. W. Lin, Roles of MKK1/2-ERK1/2 and phosphoinositide 3-kinase-AKT signaling pathways in erlotinib-induced Rad51 suppression and cytotoxicity in human non-small cell lung cancer cells. *Molecular Cancer Research* **7**, 1378-89 (2009). doi.org/10.1158/1541-7786.MCR-09-0051
377. A. G. M. Gabr, H. Goto, M. Hanibuchi, H. Ogawa, T. Kuramoto, M. Suzuki, A. Sajio, S. Kakiuchi, V. T. Trung, S. Sakaguchi, Y. Moriya, S. Sone, Y. Nishioka, Erlotinib prevents experimental metastases of human small cell lung cancer cells with no epidermal growth factor receptor expression. *Clinical and Experimental Metastasis* **29**, 207-16 (2012). doi.org/10.1007/s10585-011-9443-3
378. Z. Li, M. Xu, S. Xing, W. T. Ho, T. Ishii, Q. Li, X. Fu, Z. J. Zhao, Erlotinib effectively inhibits JAK2V617F activity and polycythemia vera cell growth. *Journal of Biological Chemistry* **282**, 3428-32 (2007). doi.org/10.1074/jbc.C600277200 ncbi.nlm.nih.gov/pmc/PMC2096634
379. Y. H. Ling, T. Li, Z. Yuan, M. Haigentz, Jr., T. K. Weber, R. Perez-Soler, Erlotinib, an effective epidermal growth factor receptor tyrosine kinase inhibitor, induces p27KIP1 up-regulation and nuclear translocation in association with cell growth inhibition and G1/S phase arrest in human non-small-cell lung cancer cell lines. *Molecular Pharmacology* **72**, 248-58 (2007). doi.org/10.1124/mol.107.034827
380. C. Y. Wang, T. T. Chao, F. Y. Chang, Y. L. Chen, Y. T. Tsai, H. I. Lin, Y. C. T. Huang, C. W. Shiao, C. J. Yu, K. F. Chen, CIP2A mediates erlotinib-induced apoptosis in non-small cell lung cancer cells without EGFR mutation. *Lung Cancer* **85**, 152-60 (2014). doi.org/10.1016/j.lungcan.2014.05.024
381. G. Liu, J. Mei, X. Zhang, J. Zhao, R. Feng, Erlotinib enhanced the susceptibility of human lung cancer A549 cells to CIK cell-mediated lysis. *Chinese Journal of Clinical Oncology* **40**, 617-20 (2013).
382. J. Z. Mei, G. J. Liu, X. J. Zhang, J. Z. Zhao, R. T. Feng, Erlotinib enhances the CIK cell-killing sensitivity of lung adenocarcinoma A549 cells. *Genetics and Molecular Research* **14**, 3082-9 (2015). doi.org/10.4238/2015.April.10.18
383. N. Pore, Z. Jiang, A. Gupta, G. Cerniglia, G. D. Kao, A. Maity, EGFR tyrosine kinase inhibitors decrease VEGF expression by both hypoxia-inducible factor (HIF)-1-independent and HIF-1-dependent mechanisms. *Cancer Research* **66**, 3197-204 (2006). doi.org/10.1158/0008-5472.CAN-05-3090
384. G. Schaefer, L. Shao, K. Totpal, R. W. Akita, Erlotinib directly inhibits HER2 kinase activation and downstream signaling events in intact cells lacking epidermal growth factor receptor expression. *Cancer Research* **67**, 1228-38 (2007). doi.org/10.1158/0008-5472.CAN-06-3493
385. Q. Sun, L. Ming, S. M. Thomas, Y. Wang, Z. G. Chen, R. L. Ferris, J. R. Grandis, L. Zhang, J. Yu, PUMA mediates EGFR tyrosine kinase inhibitor-induced apoptosis in head and neck cancer cells. *Oncogene* **28**, 2348-57 (2009). doi.org/10.1038/onc.2009.108 ncbi.nlm.nih.gov/pmc/PMC2872091
386. J. Wan, J. Cui, L. Wang, K. Wu, X. Hong, Y. Zou, S. Zhao, H. Ke, Excessive mitochondrial fragmentation triggered by erlotinib promotes pancreatic cancer PANC-1 cell apoptosis via activating the mROS-HtrA2/Omi pathways. *Cancer Cell International* **18**, 165 (2018). doi.org/10.1186/s12935-018-0665-1 ncbi.nlm.nih.gov/pmc/PMC6196464

387. X. Chen, L. Zhu, Z. Ma, G. Sun, X. Luo, M. Li, S. Zhai, P. Li, X. Wang, Oncogenic miR-9 is a target of erlotinib in NSCLCs. *Scientific Reports* **5**, 17031 (2015). doi.org/10.1038/srep17031 ncbi.nlm.nih.gov/pmc/PMC4655475
388. P. Adjibade, B. Simoneau, N. Ledoux, W. N. Gauthier, M. Nkurunziza, E. W. Khandjian, R. Mazroui, Treatment of cancer cells with Lapatinib negatively regulates general translation and induces stress granules formation. *PloS One* **15**, e0231894 (2020). doi.org/10.1371/journal.pone.0231894 ncbi.nlm.nih.gov/pmc/PMC7197775
389. W. Nie, W. Song, W. Zhang, Y. Wang, A. Zhu, J. Shao, X. Guan, miR-1470 mediates lapatinib induced p27 upregulation by targeting c-jun. *Journal of Cellular Physiology* **230**, 1630-9 (2015). doi.org/10.1002/jcp.24910
390. Y. Yang, H. Zhang, W. Chen, X. Chen, X. Yu, Lapatinib promotes ovarian cancer cell apoptosis through mROS-HtrA2/Omi pathways. *European Journal of Gynaecological Oncology* **41**, 598-603 (2020). doi.org/10.31083/ejgo.2020.04.5247
391. D. Zhang, A. Pal, W. G. Bornmann, F. Yamasaki, F. J. Esteva, G. N. Hortobagyi, C. Bartholomeusz, N. T. Ueno, Activity of lapatinib is independent of EGFR expression level in HER2-overexpressing breast cancer cells. *Molecular Cancer Therapeutics* **7**, 1846-50 (2008). doi.org/10.1158/1535-7163.MCT-08-0168 ncbi.nlm.nih.gov/pmc/PMC2525738
392. L. Tang, Y. Wang, A. Strom, J. A. Gustafsson, X. Guan, Lapatinib induces p27kip1-dependent G1 arrest through both transcriptional and post-translational mechanisms. *Cell Cycle* **12**, 2665-74 (2013). doi.org/10.4161/cc.25728 ncbi.nlm.nih.gov/pmc/PMC3865056
393. X. Ding, L. Xiang, N. Wang, Z. Zhao, X. Jin, Y. Sun, W. Duan, S. Wang, X. Jin, Vandetanib-induced inhibition of neuroblastoma cell migration and invasion is associated with downregulation of the SDF-1/CXCR4 axis and matrix metalloproteinase 14. *Oncology Reports* **31**, 1165-74 (2014). doi.org/10.3892/or.2013.2963 ncbi.nlm.nih.gov/pmc/LC/United States
394. K. Pham, D. Luo, D. W. Siemann, B. K. Law, B. A. Reynolds, P. Hothi, G. Foltz, J. K. Harrison, VEGFR inhibitors upregulate CXCR4 in VEGF receptor-expressing glioblastoma in a TGF β R signaling-dependent manner. *Cancer Letters* **360**, 60-7 (2015). doi.org/10.1016/j.canlet.2015.02.005 ncbi.nlm.nih.gov/pmc/AstraZeneca
- Tocris(United Kingdom)
395. T. T. Chao, C. Y. Wang, Y. L. Chen, C. C. Lai, F. Y. Chang, Y. T. Tsai, C. H. H. Chao, C. W. Shiao, Y. C. T. Huang, C. J. Yu, K. F. Chen, Afatinib induces apoptosis in NSCLC without EGFR mutation through Elk-1-mediated suppression of CIP2A. *Oncotarget* **6**, 2164-79 (2015). doi.org/10.18632/oncotarget.2941 ncbi.nlm.nih.gov/pmc/PMC4385843
396. Y. Chen, X. Chen, X. Ding, Y. Wang, Afatinib, an EGFR inhibitor, decreases EMT and tumorigenesis of Huh-7 cells by regulating the ERK-VEGF/MMP9 signaling pathway. *Molecular Medicine Reports* **20**, 3317-25 (2019). doi.org/10.3892/mmr.2019.10562 ncbi.nlm.nih.gov/pmc/PMC6755195
397. X. Liu, Z. Lv, J. Zou, X. Liu, J. Ma, J. Wang, N. Sa, P. Jing, W. Xu, Afatinib down-regulates MCL-1 expression through the PERK-eIF2a-ATF4 axis and leads to apoptosis in head and neck squamous cell carcinoma. *American Journal of Cancer Research* **6**, 1708-19 (2016). doi.org/10.1038/s41388-019-0849-8 ncbi.nlm.nih.gov/pmc/PMC5004074
398. F. Solca, G. Dahl, A. Zoephel, G. Bader, M. Sanderson, C. Klein, O. Kraemer, F. Himmelsbach, E. Haaksma, G. R. Adolf, Target binding properties and cellular activity of afatinib (BIBW 2992), an irreversible ErbB family blocker. *Journal of Pharmacology and Experimental Therapeutics* **343**, 342-50 (2012). doi.org/10.1124/jpet.112.197756
399. C. H. Yu, C. C. Chou, H. F. Tu, W. C. Huang, Y. Y. Ho, K. H. Khoo, M. S. Lee, G. D. Chang, Antibody-assisted target identification reveals afatinib, an EGFR covalent inhibitor, down-regulating ribonucleotide reductase. *Oncotarget* **9**, 21512-29 (2018). doi.org/10.18632/oncotarget.25177 ncbi.nlm.nih.gov/pmc/PMC5940374
400. X. M. Jiang, Y. L. Xu, M. Y. Huang, L. L. Zhang, M. X. Su, X. Chen, J. J. Lu, Osimertinib (AZD9291) decreases programmed death ligand-1 in EGFR-mutated non-small cell lung cancer cells. *Acta Pharmacologica Sinica* **38**, 1512-20 (2017). doi.org/10.1038/aps.2017.123 ncbi.nlm.nih.gov/pmc/PMC5672073
401. K. L. Zhang, Q. Q. Shen, Y. F. Fang, Y. M. Sun, J. Ding, Y. Chen, AZD9291 inactivates the PRC2 complex to mediate tumor growth inhibition. *Acta Pharmacologica Sinica* **40**, 1587-95 (2019). doi.org/10.1038/s41401-019-0248-2 ncbi.nlm.nih.gov/pmc/PMC7468275
402. Z. H. Tang, W. X. Cao, M. X. Su, X. Chen, J. J. Lu, Osimertinib induces autophagy and apoptosis via reactive oxygen species generation in non-small cell lung cancer cells. *Toxicology and Applied Pharmacology* **321**, 18-26 (2017). doi.org/10.1016/j.taap.2017.02.017
403. C. Chen, C. D. Cheng, H. Wu, Z. W. Wang, L. Wang, Z. R. Jiang, A. L. Wang, C. Hu, Y. F. Dong, W. X. Niu, S. Qi, Z. P. Qi, J. Liu, W. C. Wang, C. S. Niu, Q. S. Liu, Osimertinib successfully combats EGFR-negative glioblastoma cells by inhibiting the MAPK pathway. *Acta Pharmacologica Sinica* **42**, 108-14 (2021). doi.org/10.1038/s41401-020-0418-2 ncbi.nlm.nih.gov/pmc/PMC7921414
404. Z. Zhang, M. Zhang, H. Liu, W. Yin, AZD9291 promotes autophagy and inhibits PI3K/Akt pathway in NSCLC cancer cells. *Journal of Cellular Biochemistry* **120**, 756-67 (2019). doi.org/10.1002/jcb.27434
405. Z. R. Li, F. Z. Suo, B. Hu, Y. J. Guo, D. J. Fu, B. Yu, Y. C. Zheng, H. M. Liu, Identification of osimertinib (AZD9291) as a lysine specific demethylase 1 inhibitor. *Bioorganic Chemistry* **84**, 164-9 (2019). doi.org/10.1016/j.bioorg.2018.11.018
406. P. Dent, L. Booth, A. Poklepovic, J. Martinez, D. V. Hoff, J. F. Hancock, Neratinib degrades MST4 via autophagy that reduces membrane stiffness and is essential for the inactivation of PI3K, ERK1/2, and YAP/TAZ signaling. *Journal of Cellular Physiology* **235**, 7889-99 (2020). doi.org/10.1002/jcp.29443
407. P. Dent, L. Booth, J. L. Roberts, J. Liu, A. Poklepovic, A. S. Lalani, D. Tuveson, J. Martinez, J. F. Hancock, Neratinib inhibits Hippo/YAP signaling, reduces mutant K-RAS expression, and kills pancreatic and blood cancer cells. *Oncogene* **38**, 5890-904 (2019). doi.org/10.1038/s41388-019-0849-8 ncbi.nlm.nih.gov/pmc/PMC7133220

408. S. Wang, J. Zhang, T. Wang, F. Ren, X. Liu, Y. Lu, L. Xu, Y. Zhang, D. Wang, L. Xu, Y. Wu, F. Liu, Q. Li, M. Y. Zaky, S. Liu, W. Dong, F. Liu, K. Zou, Y. Zhang, Endocytic degradation of ErbB2 mediates the effectiveness of neratinib in the suppression of ErbB2-positive ovarian cancer. *International Journal of Biochemistry & Cell Biology* **117**, 105640 (2019). doi.org/10.1016/j.biocel.2019.105640
409. V. H. Bull, K. Rajalingam, B. Thiede, Sorafenib-induced mitochondrial complex I inactivation and cell death in human neuroblastoma cells. *Journal of Proteome Research* **11**, 1609-20 (2012). doi.org/10.1021/pr200790e
410. R. Cabrera, M. Ararat, Y. Xu, T. Brusko, C. Wasserfall, M. A. Atkinson, L. J. Chang, C. Liu, D. R. Nelson, Immune modulation of effector CD4+ and regulatory T cell function by sorafenib in patients with hepatocellular carcinoma. "Cancer Immunology, Immunotherapy" **62**, 737-46 (2013). doi.org/10.1007/s00262-012-1380-8 ncbi.nlm.nih.gov/pmc/PMC3863727
411. R. Houben, H. Voigt, C. Noelke, V. Hofmeister, J. C. Becker, D. Schrama, MAPK-independent impairment of T-cell responses by the multikinase inhibitor sorafenib. *Molecular Cancer Therapeutics* **8**, 433-40 (2009). doi.org/10.1158/1535-7163.MCT-08-1051
412. M. M. Hipp, N. Hilf, S. Walter, D. Werth, K. M. Brauer, M. P. Radsak, T. Weinschenk, H. Singh-Jasuja, P. Brossart, Sorafenib, but not sunitinib, affects function of dendritic cells and induction of primary immune responses. *Blood* **111**, 5610-20 (2008). doi.org/10.1182/blood-2007-02-075945
413. R. V. Iyer, O. Maguire, M. Kim, L. I. Curtin, S. Sexton, D. T. Fisher, S. A. Schihl, G. Fetterly, S. Menne, H. Minderman, Dose-dependent sorafenib-induced immunosuppression is associated with aberrant NFAT activation and expression of PD-1 in T cells. *Cancers* **11**, (2019). doi.org/10.3390/cancers11050681 ncbi.nlm.nih.gov/pmc/PMC6562672
414. M. Krusch, J. Salih, M. Schlicke, T. Baessler, K. M. Kampa, F. Mayer, H. R. Salih, The kinase inhibitors sunitinib and sorafenib differentially affect NK cell antitumor reactivity in vitro. *Journal of Immunology* **183**, 8286-94 (2009). doi.org/10.4049/jimmunol.0902404
415. C. Li, S. Wei, X. Xu, Y. Jiang, L. Xue, P. Jiang, J. Wang, Sorafenib attenuated the function of natural killer cells infiltrated in HCC through inhibiting ERK1/2. *International Immunopharmacology* **76**, (2019). doi.org/10.1016/j.intimp.2019.105855
416. J. C. Lin, C. L. Liu, J. J. Lee, T. P. Liu, W. C. Ko, Y. C. Huang, C. H. Wu, Y. J. Chen, Sorafenib induces autophagy and suppresses activation of human macrophage. *International Immunopharmacology* **15**, 333-9 (2013). doi.org/10.1016/j.intimp.2013.01.006 ncbi.nlm.nih.gov/pmc/PMC7106104
417. M. F. Sprinzl, A. Puschnik, A. M. Schlitter, A. Schad, K. Ackermann, I. Esposito, H. Lang, P. R. Galle, A. Weinmann, M. Heikenwalder, U. Protzer, Sorafenib inhibits macrophage-induced growth of hepatoma cells by interference with insulin-like growth factor-1 secretion. *Journal of Hepatology* **62**, 863-70 (2015). doi.org/10.1016/j.jhep.2014.11.011
418. M. Cao, Y. Xu, J. I. Youn, R. Cabrera, X. Zhang, D. Gabrilovich, D. R. Nelson, C. Liu, Kinase inhibitor Sorafenib modulates immunosuppressive cell populations in a murine liver cancer model. *Laboratory Investigation* **91**, 598-608 (2011). doi.org/10.1038/labinvest.2010.205 ncbi.nlm.nih.gov/pmc/PMC3711234
419. M. L. Chen, B. S. Yan, W. C. Lu, M. H. Chen, S. L. Yu, P. C. Yang, A. L. Cheng, Sorafenib relieves cell-intrinsic and cell-extrinsic inhibitions of effector T cells in tumor microenvironment to augment antitumor immunity. *International Journal of Cancer* **134**, 319-31 (2014). doi.org/10.1002/ijc.28362
420. C. L. Chung, S. W. Wang, W. C. Sun, C. W. Shu, Y. C. Kao, M. S. Shiao, C. L. Chen, Sorafenib suppresses TGF-beta responses by inducing caveolae/lipid raft-mediated internalization/degradation of cell-surface type II TGF-beta receptors: Implications in development of effective adjunctive therapy for hepatocellular carcinoma. *Biochemical Pharmacology* **154**, 39-53 (2018). doi.org/10.1016/j.bcp.2018.04.014
421. R. Coriat, C. Nicco, C. Chereau, O. Mir, J. Alexandre, S. Ropert, B. Weill, S. Chaussade, F. Goldwasser, F. Batteux, Sorafenib-induced hepatocellular carcinoma cell death depends on reactive oxygen species production in vitro and in vivo. *Molecular Cancer Therapeutics* **11**, 2284-93 (2012). doi.org/10.1158/1535-7163.MCT-12-0093
422. X. Dai, F. Geng, J. Dai, M. Li, M. Liu, Rho GTPase activating protein 24 (ARHGAP24) regulates the anti-cancer activity of sorafenib against breast cancer MDA-MB-231 cells via the signal transducer and activator of transcription 3 (STAT3) signaling pathway. *Medical Science Monitor* **24**, 8669-77 (2018). doi.org/10.12659/MSM.911394 ncbi.nlm.nih.gov/pmc/PMC6284358
423. J. P. Edwards, L. A. Emens, The multikinase inhibitor Sorafenib reverses the suppression of IL-12 and enhancement of IL-10 by PGE2 in murine macrophages. *International Immunopharmacology* **10**, 1220-8 (2010). doi.org/10.1016/j.intimp.2010.07.002 ncbi.nlm.nih.gov/pmc/PMC2949513
424. T. Faranda, I. Grossi, M. Manganelli, E. Marchina, G. Baiocchi, N. Portolani, M. Crosatti, G. De Petro, A. Salvi, Differential expression profiling of long non-coding RNA GAS5 and miR-126-3p in human cancer cells in response to sorafenib. *Scientific Reports* **9**, 9118 (2019). doi.org/10.1038/s41598-019-45604-2 ncbi.nlm.nih.gov/pmc/PMC6591391
425. L. Fiume, M. Manerba, M. Vettraino, G. Di Stefano, Effect of sorafenib on the energy metabolism of hepatocellular carcinoma cells. *European Journal of Pharmacology* **670**, 39-43 (2011). doi.org/10.1016/j.ejphar.2011.08.038
426. C. Fumarola, C. Caffarra, S. La Monica, M. Galetti, R. R. Alfieri, A. Cavazzoni, E. Galvani, D. Generali, P. G. Petronini, M. A. Bonelli, Effects of sorafenib on energy metabolism in breast cancer cells: Role of AMPK-mTORC1 signaling. *Breast Cancer Research and Treatment* **141**, 67-78 (2013). doi.org/10.1007/s10549-013-2668-x
427. R. González, M. A. Rodríguez-Hernández, M. Negrete, K. Ranguelova, A. Rossin, C. Choya-Foces, P. D. L. Cruz-Ojeda, A. Miranda-Vizuete, A. Martínez-Ruiz, S. Rius-Pérez, J. Sastre, J. A. Bárcena, A. O. Hueber, C. A. Padilla, J. Muntané, Downregulation of thioredoxin-1-dependent CD95 S-nitrosation by Sorafenib reduces liver cancer. *Redox Biology* **34**, (2020). doi.org/10.1016/j.redox.2020.101528 ncbi.nlm.nih.gov/pmc/PMC7210585

428. T. Y. Ha, S. Hwang, K. I. M. Moon, Y. J. Won, G. I. W. Song, N. Kim, E. Tak, B. Y. Ryoo, H. N. Hong, Sorafenib inhibits migration and invasion of hepatocellular carcinoma cells through suppression of matrix metalloproteinase expression. *Anticancer Research* **35**, 1967-76 (2015).
doi.org/10.18632/oncotarget.8812
[ncbi.nlm.nih.gov/pmc/PMC5045420](https://www.ncbi.nlm.nih.gov/pmc/PMC5045420)
429. K. Ullrich, K. D. Wurster, B. Lamprecht, K. Köchert, A. Engert, B. Dörken, M. Janz, S. Mathas, BAY 43-9006/Sorafenib blocks CSF1R activity and induces apoptosis in various classical Hodgkin lymphoma cell lines. *British Journal of Haematology* **155**, 398-402 (2011). doi.org/10.1111/j.1365-2141.2011.08685.x
430. C. Louandre, Z. Ezzoukhry, C. Godin, J. C. Barbare, J. C. Maziere, B. Chauffert, A. Galmiche, Iron-dependent cell death of hepatocellular carcinoma cells exposed to sorafenib. *International Journal of Cancer* **133**, 1732-42 (2013). doi.org/10.1002/ijc.28159
431. C. Louandre, I. Marcq, H. Bouhla, E. Lachaiher, C. Godin, Z. Saidak, C. Francois, D. Chatelain, V. Debuysscher, J. C. Barbare, B. Chauffert, A. Galmiche, The retinoblastoma (Rb) protein regulates ferroptosis induced by sorafenib in human hepatocellular carcinoma cells. *Cancer Letters* **356**, 971-7 (2015).
doi.org/10.1016/j.canlet.2014.11.014
432. D. Llobet, N. Eritja, A. Yeramian, J. Pallares, A. Sorolla, M. Domingo, M. Santacana, F. J. Gonzalez-Tallada, X. Matias-Guiu, X. Dolcet, The multikinase inhibitor Sorafenib induces apoptosis and sensitises endometrial cancer cells to TRAIL by different mechanisms. *European Journal of Cancer* **46**, 836-50 (2010).
doi.org/10.1016/j.ejca.2009.12.025
433. R. R. Rosato, J. A. Almenara, S. Coe, S. Grant, The multikinase inhibitor sorafenib potentiates TRAIL lethality in human leukemia cells in association with Mcl-1 and cFLIPL down-regulation. *Cancer Research* **67**, 9490-500 (2007). doi.org/10.1158/0008-5472.CAN-07-0598
434. K. A. Heslop, A. Rovini, E. G. Hunt, D. Fang, M. E. Morris, C. F. Christie, M. B. Gooz, D. N. DeHart, Y. Dang, J. J. Lemasters, E. N. Maldonado, JNK activation and translocation to mitochondria mediates mitochondrial dysfunction and cell death induced by VDAC opening and sorafenib in hepatocarcinoma cells. *Biochemical Pharmacology* **171**, 113728 (2020).
doi.org/10.1016/j.bcp.2019.113728
[ncbi.nlm.nih.gov/pmc/PMC7309270](https://www.ncbi.nlm.nih.gov/pmc/PMC7309270)
435. M. H. Hung, W. T. Tai, C. W. Shiao, K. F. Chen, Downregulation of signal transducer and activator of transcription 3 by sorafenib: A novel mechanism for hepatocellular carcinoma therapy. *World Journal of Gastroenterology* **20**, 15269-74 (2014).
doi.org/10.3748/wjg.v20.i41.15269
[ncbi.nlm.nih.gov/pmc/PMC4223260](https://www.ncbi.nlm.nih.gov/pmc/PMC4223260)
436. P. Kharaziha, D. Chioureas, G. Baltatzis, P. Fonseca, P. Rodriguez, V. Gogvadze, L. Lennartsson, A. C. Bjorklund, B. Zhivotovsky, D. Grander, L. Egevad, S. Nilsson, T. Panaretakis, Sorafenib-induced defective autophagy promotes cell death by necrosis. *Oncotarget* **6**, 37066-82 (2015).
doi.org/10.18632/oncotarget.5797
[ncbi.nlm.nih.gov/pmc/PMC4741916](https://www.ncbi.nlm.nih.gov/pmc/PMC4741916)
437. M. Li, W. Wang, Y. Dan, Z. Tong, W. Chen, L. Qin, K. Liu, W. Li, P. Mo, C. Yu, Downregulation of amplified in breast cancer 1 contributes to the anti-tumor effects of sorafenib on human hepatocellular carcinoma. *Oncotarget* **7**, 29605-19 (2016).
438. J. Xu, N. Cao, Y. Dai, J. Feng, L. Liu, Inhibitory effect of sorafenib on non-small cell lung cancer H460 and A549 cell lines. *Anti-Tumor Pharmacy* **8**, 26-30 (2018).
doi.org/10.1097/JTO.0000000000000107
439. G. Liu, S. Kuang, R. Cao, J. Wang, Q. Peng, C. Sun, Sorafenib kills liver cancer cells by disrupting SCD1-mediated synthesis of monounsaturated fatty acids via the ATP-AMPK-mTOR-SREBP1 signaling pathway. *FASEB Journal* **33**, 10089-103 (2019).
doi.org/10.1096/fj.201802619RR
440. J. Y. Liu, S. H. Park, C. Morisseau, H. H. Sung, B. D. Hammock, R. H. Weiss, Sorafenib has soluble epoxide hydrolase inhibitory activity, which contributes to its effect profile in vivo. *Molecular Cancer Therapeutics* **8**, 2193-203 (2009).
doi.org/10.1158/1535-7163.MCT-09-0119
[ncbi.nlm.nih.gov/pmc/PMC2728155](https://www.ncbi.nlm.nih.gov/pmc/PMC2728155)
441. S. J. Oh, H. H. Erb, A. Hobisch, F. R. Santer, Z. Culig, Sorafenib decreases proliferation and induces apoptosis of prostate cancer cells by inhibition of the androgen receptor and Akt signaling pathways. *Endocrine-Related Cancer* **19**, 305-19 (2012).
doi.org/10.1530/ERC-11-0298 [ncbi.nlm.nih.gov/pmc/PMC3353237](https://www.ncbi.nlm.nih.gov/pmc/PMC3353237)
442. M. Rahmani, E. M. Davis, T. R. Crabtree, J. R. Habibi, T. K. Nguyen, P. Dent, S. Grant, The kinase inhibitor sorafenib induces cell death through a process involving induction of endoplasmic reticulum stress. *Molecular and Cellular Biology* **27**, 5499-513 (2007). doi.org/10.1128/MCB.01080-06
[ncbi.nlm.nih.gov/pmc/PMC1952105](https://www.ncbi.nlm.nih.gov/pmc/PMC1952105)
443. A. Rodríguez-Hernández, E. Navarro-Villarán, R. González, S. Pereira, L. B. Soriano-De Castro et al., Regulation of cell death receptor S-nitrosylation and apoptotic signaling by sorafenib in hepatoblastoma cells. *Redox Biology* **6**, 174-82 (2015).
doi.org/10.1016/j.redox.2015.07.010
[ncbi.nlm.nih.gov/pmc/PMC4534573](https://www.ncbi.nlm.nih.gov/pmc/PMC4534573)
444. C. Roolf, N. Dybowski, A. Sekora, S. Mueller, G. Knuebel, A. Tebbe, H. Murua Escobar, K. Godl, C. Junghanss, C. Schaab, Phosphoproteome analysis reveals differential mode of action of sorafenib in wildtype and mutated FLT3 acute myeloid leukemia (AML) cells. *Molecular and Cellular Proteomics* **16**, 1365-76 (2017).
doi.org/10.1074/mcp.M117.067462
[ncbi.nlm.nih.gov/pmc/PMC5500767](https://www.ncbi.nlm.nih.gov/pmc/PMC5500767)
445. L. Serrano-Oviedo, M. Ortega-Muelas, J. García-Cano, M. L. Valero, F. J. Cimas, R. Pascual-Serra, D. M. Fernandez-Aroca, O. Roche, M. J. Ruiz-Hidalgo, B. Belandia, J. M. Giménez-Bachs, A. S. Salinas, R. Sanchez-Prieto, Autophagic cell death associated to sorafenib in renal cell carcinoma is mediated through Akt inhibition in an ERK1/2 independent fashion. *PLoS One* **13**, e0200878 (2018).
doi.org/10.1371/journal.pone.0200878
[ncbi.nlm.nih.gov/pmc/PMC6062059](https://www.ncbi.nlm.nih.gov/pmc/PMC6062059)
446. W. T. Tai, A. L. Cheng, C. W. Shiao, H. P. Huang, J. W. Huang, P. J. Chen, K. F. Chen, Signal transducer and activator of transcription 3 is a major kinase-independent target of sorafenib in hepatocellular carcinoma. *Journal of Hepatology* **55**, 1041-8 (2011).
doi.org/10.1016/j.jhep.2011.01.047
447. C. Y. Wang, T. T. Chao, W. T. Tai, F. Y. Chang, W. P. Su, Y. L. Chen, P. T. Chen, C. Y. Weng, A. Yuan, C. W. Shiao, C. J. Yu, K. F.

- Chen, Signal transducer and activator of transcription 3 as molecular therapy for non-small-cell lung cancer. *Journal of Thoracic Oncology* **9**, 488-96 (2014). doi.org/10.1097/JTO.0000000000000107
448. K. Takahashi, I. K. Yan, J. Wood, H. Haga, T. Patel, Involvement of extracellular vesicle long noncoding RNA (linc-VLDR) in tumor cell responses to chemotherapy. *Molecular Cancer Research* **12**, 1377-87 (2014). doi.org/10.1158/1541-7786.MCR-13-0636 ncbi.nlm.nih.gov/pmc/PMC4201956
449. X. X. He, L. L. Shi, M. J. Qiu, Q. T. Li, M. M. Wang, Z. F. Xiong, S. L. Yang, Molecularly targeted anti-cancer drugs inhibit the invasion and metastasis of hepatocellular carcinoma by regulating the expression of MMP and TIMP gene families. *Biochemical and Biophysical Research Communications* **504**, 878-84 (2018). doi.org/10.1016/j.bbrc.2018.08.203
450. J. Guo, P. A. Marcotte, J. O. McCall, Y. Dai, L. J. Pease, M. R. Michaelides, S. K. Davidsen, K. B. Glaser, Inhibition of phosphorylation of the colony-stimulating factor-1 receptor (c-Fms) tyrosine kinase in transfected cells by ABT-869 and other tyrosine kinase inhibitors. *Molecular Cancer Therapeutics* **5**, 1007-13 (2006). doi.org/10.1158/1535-7163.MCT-05-0359
451. I. Walter, B. Wolfesberger, I. Miller, G. Mair, S. Burger, B. Galle, R. Steinborn, Human osteosarcoma cells respond to sorafenib chemotherapy by downregulation of the tumor progression factors S100A4, CXCR4 and the oncogene FOS. *Oncology Reports* **31**, 1147-56 (2014). doi.org/10.3892/or.2013.2954
452. Q. Wang, T. Yu, Y. Yuan, H. Zhuang, Z. Wang, X. Liu, M. Feng, Sorafenib reduces hepatic infiltrated regulatory T cells in hepatocellular carcinoma patients by suppressing TGF-beta signal. *Journal of Surgical Oncology* **107**, 422-7 (2013). doi.org/10.1002/jso.23227
453. S. Wang, Y. Zhu, H. He, J. Liu, L. Xu, H. Zhang, H. Liu, W. Liu, Y. Liu, D. Pan, L. Chen, Q. Wu, J. Xu, J. Gu, Sorafenib suppresses growth and survival of hepatoma cells by accelerating degradation of enhancer of zeste homolog 2. *Cancer Science* **104**, 750-9 (2013). doi.org/10.1111/cas.12132 ncbi.nlm.nih.gov/pmc/PMC7657235
454. Z. Wang, M. Wang, B. I. Carr, Involvement of receptor tyrosine phosphatase DEP-1 mediated PI3K-cofilin signaling pathway in sorafenib-induced cytoskeletal rearrangement in hepatoma cells. *Journal of Cellular Physiology* **224**, 559-65 (2010). doi.org/10.1002/jcp.22160
455. J. C. Wei, F. D. Meng, K. Qu, Z. X. Wang, Q. F. Wu, L. Q. Zhang, Q. Pang, C. Liu, Sorafenib inhibits proliferation and invasion of human hepatocellular carcinoma cells via up-regulation of p53 and suppressing FoxM1. *Acta Pharmacologica Sinica* **36**, 241-51 (2015). doi.org/10.1038/aps.2014.122 ncbi.nlm.nih.gov/pmc/PMC4326788
456. M. Xu, Y. L. Zheng, X. Y. Xie, J. Y. Liang, F. S. Pan, S. G. Zheng, M. D. Lü, Sorafenib blocks the HIF-1 α /VEGFA pathway, inhibits tumor invasion, and induces apoptosis in hepatoma cells. *DNA and Cell Biology* **33**, 275-81 (2014). doi.org/10.1089/dna.2013.2184
457. F. Yang, C. Brown, R. Buettner, M. Hedvat, R. Starr, A. Scuto, A. Schroeder, M. Jensen, R. Jove, Sorafenib induces growth arrest and apoptosis of human glioblastoma cells through the dephosphorylation of signal transducers and activators of transcription 3. *Molecular Cancer Therapeutics* **9**, 953-62 (2010).
- doi.org/10.1158/1535-7163.MCT-09-0947 ncbi.nlm.nih.gov/pmc/PMC2852467
458. F. Yang, T. E. Van Meter, R. Buettner, M. Hedvat, W. Liang, C. M. Kowolik, N. Mepani, J. Mirosevich, S. Nam, M. Y. Chen, G. Tye, M. Kirschbaum, R. Jove, Sorafenib inhibits signal transducer and activator of transcription 3 signaling associated with growth arrest and apoptosis of medulloblastomas. *Molecular Cancer Therapeutics* **7**, 3519-26 (2008). doi.org/10.1158/1535-7163.MCT-08-0138 ncbi.nlm.nih.gov/pmc/PMC2592687
459. P. Yi, A. Higa, S. Taouji, M. G. Bexiga, E. Marza, D. Arma, C. Castain, B. Le Bail, J. C. Simpson, J. Rosenbaum, C. Balabaud, P. Bioulac-Sage, J. F. Blanc, E. Chevet, Sorafenib-mediated targeting of the AAA(+) ATPase p97/VCP leads to disruption of the secretory pathway, endoplasmic reticulum stress, and hepatocellular cancer cell death. *Molecular Cancer Therapeutics* **11**, 2610-20 (2012). doi.org/10.1158/1535-7163.MCT-12-0516
460. P. M. Yang, L. S. Lin, T. P. Liu, Sorafenib inhibits ribonucleotide reductase regulatory subunit M2 (RRM2) in hepatocellular carcinoma cells. *Biomolecules* **10**, (2020). doi.org/10.3390/biom10010117 ncbi.nlm.nih.gov/pmc/PMC7022495
461. H. V. Namboodiri, M. Bukhtiyarova, J. Ramcharan, M. Karpusas, Y. Lee, E. B. Springman, Analysis of imatinib and sorafenib binding to p38 α Compared with c-Abl and b-Raf provides structural insights for understanding the selectivity of inhibitors targeting the DFG-out form of protein kinases. *Biochemistry* **49**, 3611-8 (2010). doi.org/10.1021/bi100070r
462. M. Yoshida, T. Yamashita, H. Okada, N. Oishi, K. Nio, T. Hayashi, Y. Nomura, T. Hayashi, Y. Asahina, M. Ohwada, H. Sunagozaka, H. Takatori, F. Colombo, L. Porretti, M. Honda, S. Kaneko, Sorafenib suppresses extrahepatic metastasis de novo in hepatocellular carcinoma through inhibition of mesenchymal cancer stem cells characterized by the expression of CD90. *Scientific Reports* **7**, 11292 (2017). doi.org/10.1038/s41598-017-11848-z ncbi.nlm.nih.gov/pmc/PMC5596021
463. C. Zhang, Z. Liu, E. Bunker, A. Ramirez, S. Lee, Y. Peng, A. C. Tan, S. G. Eckhardt, D. A. Chapnick, X. Liu, Sorafenib targets the mitochondrial electron transport chain complexes and ATP synthase to activate the PINK1-Parkin pathway and modulate cellular drug response. *Journal of Biological Chemistry* **292**, 15105-20 (2017). doi.org/10.1074/jbc.M117.783175 ncbi.nlm.nih.gov/pmc/PMC5592685
464. X. Zhao, M. Cao, Z. Lu, T. Wang, Y. Ren, C. Liu, D. Nelson, Small-molecule inhibitor sorafenib regulates immunoreactions by inducing survival and differentiation of bone marrow cells. *Innate Immunity* **22**, 493-502 (2016). doi.org/10.1177/1753425916659702
465. X. Zhao, C. Tian, W. M. Puszyk, O. O. Ogunwobi, M. Cao, T. Wang, R. Cabrera, D. R. Nelson, C. Liu, OPA1 downregulation is involved in sorafenib-induced apoptosis in hepatocellular carcinoma. *Laboratory Investigation* **93**, 8-19 (2013). doi.org/10.1038/labinvest.2012.144 ncbi.nlm.nih.gov/pmc/PMC3860369
466. M. Zheng, H. Xu, X. H. Liao, C. P. Chen, A. L. Zhang, W. Lu, L. Wang, D. Yang, J. Wang, H. Liu, X. Z. Zhou, K. P. Lu, Inhibition of the prolyl isomerase Pin1 enhances the ability of sorafenib to induce cell death and inhibit tumor growth in hepatocellular carcinoma. *Oncotarget* **8**, 29771-84 (2017).

doi.org/10.18632/oncotarget.15967
ncbi.nlm.nih.gov/pmc/PMC5444702

467. C. Zhou, J. Liu, Y. Li, L. Liu, X. Zhang, C. Y. Ma, S. C. Hua, M. Yang, Q. Yuan, microRNA-1274a, a modulator of sorafenib induced a disintegrin and metalloproteinase 9 (ADAM9) down-regulation in hepatocellular carcinoma. *FEBS Letters* **585**, 1828-34 (2011). doi.org/10.1016/j.febslet.2011.04.040
468. M. Santoni, C. Amantini, M. B. Morelli, S. Liberati, V. Farfariello, M. Nabissi, L. Bonfili, A. M. Eleuteri, M. Mozzicafreddo, L. Burattini, R. Berardi, S. Cascinu, G. Santoni, Pazopanib and sunitinib trigger autophagic and non-autophagic death of bladder tumour cells. *British Journal of Cancer* **109**, 1040-50 (2013). doi.org/10.1038/bjc.2013.420 ncbi.nlm.nih.gov/pmc/PMC3749583
469. J. Y. Winum, A. Maresca, F. Carta, A. Scozzafava, C. T. Supuran, Polypharmacology of sulfonamides: Pazopanib, a multitargeted receptor tyrosine kinase inhibitor in clinical use, potently inhibits several mammalian carbonic anhydrases. *Chem Commun (Camb)* **48**, 8177-9 (2012). doi.org/10.1039/c2cc33415a
470. I. G. Zizzari, C. Napoletano, A. Botticelli, S. Caponnetto, F. Calabro, A. Celibter, A. Rughetti, I. Ruscito, H. Rahimi, E. Rossi, G. Schinzari, P. Marchetti, M. Nuti, TK inhibitor pazopanib primes DCs by downregulation of the beta-catenin pathway. *Cancer Immunology Research* **6**, 711-22 (2018). doi.org/10.1158/2326-6066.CIR-17-0594
471. S. Ahmad, M. A. Hughes, G. L. Johnson, J. E. Scott, Development and validation of a high-throughput intrinsic ATPase activity assay for the discovery of MEKK2 inhibitors. *Journal of Biomolecular Screening* **18**, 388-99 (2013). doi.org/10.1177/1087057112466430 ncbi.nlm.nih.gov/pmc/PMC3723327
472. M. B. Morelli, C. Amantini, M. Nabissi, C. Cardinali, M. Santoni, G. Bernardini, A. Santoni, G. Santoni, Axitinib induces senescence-associated cell death and necrosis in glioma cell lines: The proteasome inhibitor, bortezomib, potentiates axitinib-induced cytotoxicity in a p21(Waf/Cip1) dependent manner. *Oncotarget* **8**, 3380-95 (2017). doi.org/10.18632/oncotarget.13769 ncbi.nlm.nih.gov/pmc/PMC5356889
473. M. P. Mongiardi, G. Radice, M. Piras, V. Stagni, S. Pacioni, A. Re, S. Putti, F. Ferrè, A. Farsetti, R. Pallini, D. Barilà, A. Levi, M. L. Falchetti, Axitinib exposure triggers endothelial cells senescence through ROS accumulation and ATM activation. *Oncogene* **38**, 5413-24 (2019). doi.org/10.1038/s41388-019-0798-2
474. M. B. Morelli, C. Amantini, M. Santoni, A. Soriani, M. Nabissi, C. Cardinali, A. Santoni, G. Santoni, Axitinib induces DNA damage response leading to senescence, mitotic catastrophe, and increased NK cell recognition in human renal carcinoma cells. *Oncotarget* **6**, 36245-59 (2015). doi.org/10.18632/oncotarget.5768 ncbi.nlm.nih.gov/pmc/PMC4742174
475. Y. Qu, N. Gharbi, X. Yuan, J. R. Olsen, P. Blicher, B. Dalhus, K. A. Brokstad, B. Lin, A. M. Øyan, W. Zhang, K. H. Kalland, X. Ke, Axitinib blocks Wnt/β-catenin signaling and directs asymmetric cell division in cancer. *Proceedings of the National Academy of Sciences of the United States of America* **113**, 9339-44 (2016). doi.org/10.1073/pnas.1604520113 ncbi.nlm.nih.gov/pmc/PMC4995957
476. Y. Song, X. Zhang, X. Cheng, X. Zhang, Effect of axitinib on the proliferation and apoptosis of human lung adenocarcinoma PC9

cells and its mechanism. *Cancer Research and Clinic* **31**, 232-6 (2019).

477. H. Yuan, P. Cai, Q. Li, W. Wang, Y. Sun, Q. Xu, Y. Gu, Axitinib augments antitumor activity in renal cell carcinoma via STAT3-dependent reversal of myeloid-derived suppressor cell accumulation. *Biomedicine and Pharmacotherapy* **68**, 751-6 (2014). doi.org/10.1016/j.biopha.2014.07.002
478. X. Zhang, X. Fang, Z. Gao, W. Chen, F. Tao, P. Cai, H. Yuan, Y. Shu, Q. Xu, Y. Sun, Y. Gu, Axitinib, a selective inhibitor of vascular endothelial growth factor receptor, exerts an anticancer effect in melanoma through promoting antitumor immunity. *Anti-Cancer Drugs* **25**, 204-11 (2014). doi.org/10.1097/CAD.0000000000000033
479. X. Chen, B. Xie, L. Cao, F. Zhu, B. Chen, H. Lv, X. Fan, L. Han, L. Bie, X. Cao, X. Shen, F. Cao, Direct binding of microRNA-21 pre-element with Regorafenib: An alternative mechanism for anti-colorectal cancer chemotherapy? *Journal of Molecular Graphics and Modelling* **73**, 48-53 (2017). doi.org/10.1016/j.jmgm.2017.02.004
480. L. C. Fan, H. W. Teng, C. W. Shiao, H. Lin, M. H. Hung, Y. L. Chen, J. W. Huang, W. T. Tai, H. C. Yu, K. F. Chen, SHP-1 is a target of regorafenib in colorectal cancer. *Oncotarget* **5**, 6243-51 (2014). doi.org/10.18632/oncotarget.2191 ncbi.nlm.nih.gov/pmc/PMC4171626
481. S. Suemura, T. Kodama, Y. Myojin, R. Yamada, M. Shigekawa, H. Hikita, R. Sakamori, T. Tatsumi, T. Takehara, CRISPR loss-of-function screen identifies the hippo signaling pathway as the mediator of regorafenib efficacy in hepatocellular carcinoma. *Cancers* **11**, (2019). doi.org/10.3390/cancers11091362 ncbi.nlm.nih.gov/pmc/PMC6770429
482. J. Jiang, L. Zhang, H. Chen, Y. Lei, T. Zhang et al., Regorafenib induces lethal autophagy arrest by stabilizing PSAT1 in glioblastoma. *Autophagy* **16**, 106-22 (2020). doi.org/10.1080/15548627.2019.1598752 ncbi.nlm.nih.gov/pmc/PMC6984601
483. L. C. Fan, H. W. Teng, C. W. Shiao, W. T. Tai, M. H. Hung, S. H. Yang, J. K. Jiang, K. F. Chen, Regorafenib (Stivarga) pharmacologically targets epithelial-mesenchymal transition in colorectal cancer. *Oncotarget* **7**, 64136-47 (2016). doi.org/10.18632/oncotarget.11636 ncbi.nlm.nih.gov/pmc/PMC5325431
484. W. T. Tai, P. Y. Chu, C. W. Shiao, Y. L. Chen, Y. S. Li, M. H. Hung, L. J. Chen, P. L. Chen, J. C. Su, P. Y. Lin, H. C. Yu, K. F. Chen, STAT3 mediates regorafenib-induced apoptosis in hepatocellular carcinoma. *Clinical Cancer Research* **20**, 5768-76 (2014). doi.org/10.1158/1078-0432.CCR-14-0725
485. P. Todesca, L. Marzi, R. M. Critelli, B. Cuffari, C. Caporali, L. Turco, G. Pinelli, F. Schepis, L. Carulli, N. de Maria, F. Casari, R. Scaglioni, E. Villa, Angiopoietin-2/Tie2 inhibition by regorafenib associates with striking response in a patient with aggressive hepatocellular carcinoma. *Hepatology* **70**, 745-7 (2019). doi.org/10.1002/hep.30588
486. R. Y. Wu, P. F. Kong, L. P. Xia, Y. Huang, Z. L. Li et al., Regorafenib Promotes Antitumor Immunity via Inhibiting PD-L1 and IDO1 Expression in Melanoma. *Clinical Cancer Research* **25**, 4530-41 (2019). doi.org/10.1158/1078-0432.CCR-18-2840

487. M. J. Qiu, X. X. He, N. R. Bi, M. M. Wang, Z. F. Xiong, S. L. Yang, Effects of liver-targeted drugs on expression of immune-related proteins in hepatocellular carcinoma cells. *Clinica Chimica Acta* **485**, 103-5 (2018). doi.org/10.1016/j.cca.2018.06.032
488. D. Subramonian, N. Phanhthilath, H. Rinehardt, S. Flynn, Y. Huo, J. Zhang, K. Messer, Q. Mo, S. Huang, J. Lesperance, P. E. Zage, Regorafenib is effective against neuroblastoma in vitro and in vivo and inhibits the RAS/MAPK, PI3K/Akt/mTOR and Fos/Jun pathways. *British Journal of Cancer* **123**, 568-79 (2020). doi.org/10.1038/s41416-020-0905-8 ncbi.nlm.nih.gov/pmc/PMC7434894
489. X. L. Lin, Q. Xu, L. Tang, L. Sun, T. Han, L. W. Wang, X. Y. Xiao, Regorafenib inhibited gastric cancer cells growth and invasion via CXCR4 activated Wnt pathway. *PloS One* **12**, e0177335 (2017). doi.org/10.1371/journal.pone.0177335 ncbi.nlm.nih.gov/pmc/PMC542513
490. P. Canning, Q. Ruan, T. Schwerd, M. Hrdinka, J. L. Maki, D. Saleh, C. Suebsuwong, S. Ray, P. E. Brennan, G. D. Cuny, H. H. Uhlig, M. Gyrd-Hansen, A. Degterev, A. N. Bullock, Inflammatory signaling by NOD-RIPK2 Is inhibited by clinically relevant type II kinase inhibitors. *Chemistry and Biology* **22**, 1174-84 (2015). doi.org/10.1016/j.chembiol.2015.07.017 ncbi.nlm.nih.gov/pmc/PMC4579271
491. S. M. Wilhelm, J. Dumas, L. Adhane, M. Lynch, C. A. Carter, G. Schutz, K. H. Thierauch, D. Zopf, Regorafenib (BAY 73-4506): A new oral multikinase inhibitor of angiogenic, stromal and oncogenic receptor tyrosine kinases with potent preclinical antitumor activity. *International Journal of Cancer* **129**, 245-55 (2011). doi.org/10.1002/ijc.25864
492. T. Kimura, Y. Kato, Y. Ozawa, K. Kodama, J. Ito, K. Ichikawa, K. Yamada, Y. Hori, K. Tabata, K. Takase, J. Matsui, Y. Funahashi, K. Nomoto, Immunomodulatory activity of lenvatinib contributes to antitumor activity in the Hepa1-6 hepatocellular carcinoma model. *Cancer Science* **109**, 3993-4002 (2018). doi.org/10.1111/cas.13806 ncbi.nlm.nih.gov/pmc/PMC6272102
493. Y. Yamamoto, J. Matsui, T. Matsushima, H. Obaishi, K. Miyazaki et al., Lenvatinib, an angiogenesis inhibitor targeting VEGFR/FGFR, shows broad antitumor activity in human tumor xenograft models associated with microvessel density and pericyte coverage. *Vascular Cell* **6**, 18 (2014). doi.org/10.1186/2045-824X-6-18 ncbi.nlm.nih.gov/pmc/PMC4156793
494. J. M. Aswald, J. H. Lipton, S. Aswald, H. A. Messner, Increased IFN-gamma synthesis by T cells from patients on imatinib therapy for chronic myeloid leukemia. "Cytokines, Cellular & Molecular Therapy" **7**, 143-9 (2002). doi.org/10.1080/13684730210002319
495. V. P. Balachandran, M. J. Cavnar, S. Zeng, Z. M. Bamboat, L. M. Ocuin, H. Obaid, E. C. Sorenson, R. Popow, C. Ariyan, F. Rossi, P. Besmer, T. Guo, C. R. Antonescu, T. Taguchi, J. Yuan, J. D. Wolchok, J. P. Allison, R. P. Dematteo, Imatinib potentiates antitumor T cell responses in gastrointestinal stromal tumor through the inhibition of Ido. *Nature Medicine* **17**, 1094-100 (2011). doi.org/10.1038/nm.2438 ncbi.nlm.nih.gov/pmc/PMC3278279
496. L. Zitvogel, S. Rusakiewicz, B. Routy, M. Ayyoub, G. Kroemer, Immunological off-target effects of imatinib. *Nature Reviews Clinical Oncology* **13**, 431-46 (2016). doi.org/10.1038/nrclinonc.2016.41
497. Y. Baran, S. Zencir, Z. Cakir, E. Ozturk, Z. Topcu, Imatinib-induced apoptosis: A possible link to topoisomerase enzyme inhibition. *Journal of Clinical Pharmacy and Therapeutics* **36**, 673-9 (2011). doi.org/10.1111/j.1365-2710.2010.01224.x
498. F. Belloc, F. Moreau-Gaudry, M. Uhalde, L. Cazalis, M. Jeanneteau, F. Lacombe, V. Praloran, F. X. Mahon, Imatinib and nilotinib induce apoptosis of chronic myeloid leukemia cells through a bim-dependant pathway modulated by cytokines. *Cancer Biology & Therapy* **6**, 912-9 (2007). doi.org/10.4161/cbt.6.6.4101
499. P. S. Bernardo, F. R. S. Reis, R. C. Maia, Imatinib increases apoptosis index through modulation of survivin subcellular localization in the blast phase of CML cells. *Leukemia Research* **36**, 1510-6 (2012). doi.org/10.1016/j.leukres.2012.08.014
500. C. Borg, M. Terme, J. Taieb, C. Menard, C. Flament et al., Novel mode of action of c-kit tyrosine kinase inhibitors leading to NK cell-dependent antitumor effects. *Journal of Clinical Investigation* **114**, 379-88 (2004). doi.org/10.1172/JCI21102 ncbi.nlm.nih.gov/pmc/PMC489961
501. P. Canning, L. Tan, K. Chu, S. W. Lee, N. S. Gray, A. N. Bullock, Structural mechanisms determining inhibition of the collagen receptor DDR1 by selective and multi-targeted type II kinase inhibitors. *Journal of Molecular Biology* **426**, 2457-70 (2014). doi.org/10.1016/j.jmb.2014.04.014 ncbi.nlm.nih.gov/pmc/PMC4058747
502. E. Day, B. Waters, K. Spiegel, T. Alnafid, P. W. Manley, E. Buchdunger, C. Walker, G. Jarai, Inhibition of collagen-induced discoidin domain receptor 1 and 2 activation by imatinib, nilotinib and dasatinib. *European Journal of Pharmacology* **599**, 44-53 (2008). doi.org/10.1016/j.ejphar.2008.10.014
503. S. P. Chang, S. C. Shen, W. R. Lee, L. L. Yang, Y. C. Chen, Imatinib mesylate induction of ROS-dependent apoptosis in melanoma B16F0 cells. *Journal of Dermatological Science* **62**, 183-91 (2011). doi.org/10.1016/j.jdermsci.2011.03.001
504. L. Christiansson, S. Soderlund, S. Mansbo, H. Hjorth-Hansen, M. Hoglund, B. Markevarn, J. Richter, L. Stenke, S. Mustjoki, A. Loskog, U. Olsson-Stromberg, The tyrosine kinase inhibitors imatinib and dasatinib reduce myeloid suppressor cells and release effector lymphocyte responses. *Molecular Cancer Therapeutics* **14**, 1181-91 (2015). doi.org/10.1158/1535-7163.MCT-14-0849
505. M. S. Cohen, H. B. Hussain, J. F. Moley, Inhibition of medullary thyroid carcinoma cell proliferation and RET phosphorylation by tyrosine kinase inhibitors. *Surgery* **132**, 960-7 (2002). doi.org/10.1067/msy.2002.128562
506. S. Ezzat, P. Huang, A. Dackiw, S. L. Asa, Dual inhibition of RET and FGFR4 restrains medullary thyroid cancer cell growth. *Clinical Cancer Research* **11**, 1336-41 (2005).
507. A. Czechowska, T. Poplawski, J. Drzewoski, J. Blasiak, Imatinib (ST1571) induces DNA damage in BCR/ABL-expressing leukemic cells but not in normal lymphocytes. *Chemico-Biological Interactions* **152**, 139-50 (2005). doi.org/10.1016/j.cbi.2005.03.002
508. D. D'Allard, J. Gay, C. Descarpentries, E. Frisan, K. Adam, F. Verdier, C. Floquet, P. Dubreuil, C. Lacombe, M. Fontenay, P. Mayeux, O. Kosmider, Tyrosine kinase inhibitors induce down-regulation of c-Kit by targeting the ATP pocket. *PloS One* **8**, e60961 (2013). doi.org/10.1371/journal.pone.0060961 ncbi.nlm.nih.gov/pmc/PMC3634048

509. Y. Liao, G. Lv, B. Wang, L. Kuang, X. Wang, Imatinib promotes apoptosis of giant cell tumor cells by targeting microRNA-30a-mediated runt-related transcription factor 2. *Molecular Medicine Reports* **13**, 1739-45 (2016). doi.org/10.3892/mmr.2015.4722
510. A. L. Dewar, A. C. Cambareri, A. C. W. Zannettino, B. L. Miller, K. V. Doherty, T. P. Hughes, A. Bruce Lyons, Macrophage colony-stimulating factor receptor c-fms is a novel target of imatinib. *Blood* **105**, 3127-32 (2005). doi.org/10.1182/blood-2004-10-3967
511. I. El Hajj Dib, M. Gallet, R. Mentaverri, N. Sévenet, M. Brazier, S. Kamel, Imatinib mesylate (Gleevec®) enhances mature osteoclast apoptosis and suppresses osteoclast bone resorbing activity. *European Journal of Pharmacology* **551**, 27-33 (2006). doi.org/10.1016/j.ejphar.2006.09.007
512. T. Hiraga, H. Nakamura, Imatinib mesylate suppresses bone metastases of breast cancer by inhibiting osteoclasts through the blockade of c-Fms signals. *International Journal of Cancer* **124**, 215-22 (2009). doi.org/10.1002/ijc.23903
513. B. Mashkani, R. Griffith, L. K. Ashman, Colony stimulating factor-1 receptor as a target for small molecule inhibitors. *Bioorganic and Medicinal Chemistry* **18**, 1789-97 (2010). doi.org/10.1016/j.bmc.2010.01.056
514. Y. Dong, Q. Han, Y. Zou, Z. Deng, X. Lu, X. Wang, W. Zhang, H. Jin, J. Su, T. Jiang, H. Ren, Long-term exposure to imatinib reduced cancer stem cell ability through induction of cell differentiation via activation of MAPK signaling in glioblastoma cells. *Molecular and Cellular Biochemistry* **370**, 89-102 (2012). doi.org/10.1007/s11010-012-1401-0
515. S. C. Dos Santos, I. Sá-Correia, Genome-Wide identification of genes required for yeast growth under imatinib stress: Vacuolar h⁺-atpase function is an important target of this anticancer drug. *OMICS: A Journal of Integrative Biology* **13**, 185-98 (2009). doi.org/10.1089/omi.2008.0086
516. V. Gioni, T. Karampinas, G. Voutsinas, A. E. Roussidis, S. Papadopoulos, N. K. Karamanos, D. Kletsas, Imatinib mesylate inhibits proliferation and exerts an antifibrotic effect in human breast stroma fibroblasts. *Molecular Cancer Research* **6**, 706-14 (2008). doi.org/10.1158/1541-7786.MCR-07-0355
517. B. Gobin, G. Moriceau, B. Ory, C. Charrier, R. Brion, F. Blanchard, F. Redini, D. Heymann, Imatinib mesylate exerts anti-proliferative effects on osteosarcoma cells and inhibits the tumour growth in immunocompetent murine models. *PloS One* **9**, e90795 (2014). doi.org/10.1371/journal.pone.0090795 ncbi.nlm.nih.gov/pmc/PMC3944320
518. M. T. Gómez-Casares, J. P. Vaqué, A. Lemes, T. Molero, N. Ferrández, J. León, Antiapoptotic proteins Bcl2 and BclX do not protect chronic myeloid leukemia cells from imatinib-mediated growth arrest. *Anales de la Real Academia Nacional de Farmacia* **72**, 27-36 (2006).
519. B. Keshavarz-Pakseresht, S. A. Shandiz, F. Baghbani-Arani, Imatinib induces up-regulation of NM23, a metastasis suppressor gene, in human hepatocarcinoma (HepG2) cell line. *Gastroenterology and Hepatology from Bed to Bench* **10**, 29-33 (2017). doi.org/10.22037/ghfb.vo10.940 ncbi.nlm.nih.gov/pmc/PMC5346821
520. A. Hamai, C. Richon, F. Meslin, F. Faure, A. Kauffmann, Y. Lecluse, A. Jalil, L. Larue, M. F. Avril, S. Chouaib, M. Mehrpour, Imatinib enhances human melanoma cell susceptibility to TRAIL-induced cell death: Relationship to Bcl-2 family and caspase activation. *Oncogene* **25**, 7618-34 (2006). doi.org/10.1038/sj.onc.1209738
521. M. Bantscheff, D. Eberhard, Y. Abraham, S. Bastuck, M. Boesche et al., Quantitative chemical proteomics reveals mechanisms of action of clinical ABL kinase inhibitors. *Nature Biotechnology* **25**, 1035-44 (2007). doi.org/10.1038/nbt1328
522. U. Rix, O. Hantschel, G. Durnberger, L. L. Remsing Rix, M. Planyavsky, N. V. Fernbach, I. Kaupe, K. L. Bennett, P. Valent, J. Colinge, T. Kocher, G. Superti-Furga, Chemical proteomic profiles of the BCR-ABL inhibitors imatinib, nilotinib, and dasatinib reveal novel kinase and nonkinase targets. *Blood* **110**, 4055-63 (2007). doi.org/10.1182/blood-2007-07-102061
523. N. Nishimura, Y. Furukawa, K. Sutheesophon, M. Nakamura, K. Kishi, K. Okuda, Y. Sato, Y. Kano, Suppression of ARG kinase activity by ST1571 induces cell cycle arrest through up-regulation of CDK inhibitor p18/INK4c. *Oncogene* **22**, 4074-82 (2003). doi.org/10.1038/sj.onc.1206498
524. K. Okuda, E. Weisberg, D. G. Gilliland, J. D. Griffin, ARG tyrosine kinase activity is inhibited by ST1571. *Blood* **97**, 2440-8 (2001). doi.org/10.1182/blood.v97.8.2440
525. C. D. Mol, D. R. Dougan, T. R. Schneider, R. J. Skene, M. L. Kraus, D. N. Scheibe, G. P. Snell, H. Zou, B. C. Sang, K. P. Wilson, Structural basis for the autoinhibition and STI-571 inhibition of c-Kit tyrosine kinase. *Journal of Biological Chemistry* **279**, 31655-63 (2004). doi.org/10.1074/jbc.M403319200
526. A. Kadivar, M. I. Noordin, A. Aditya, B. Kamalidehghan, E. T. Davoudi, R. Sedghi, H. A. Javar, Antiproliferative effects of imatinib mesylate on ZR-75-1 and MDA-MB-231 cell lines via PDGFR-β, PDGF-BB, c-Kit and SCF expression. *International Journal of Molecular Medicine* **42**, 414-24 (2018). doi.org/10.3892/ijmm.2018.3590
527. D. Matei, D. D. Chang, M. H. Jeng, Imatinib mesylate (Gleevec) inhibits ovarian cancer cell growth through a mechanism dependent on platelet-derived growth factor receptor α and Akt inactivation. *Clinical Cancer Research* **10**, 681-90 (2004). doi.org/10.1158/1078-0432.ccr-0754-03
528. T. Jin, H. Nakatani, T. Taguchi, T. Nakano, T. Okabayashi, T. Sugimoto, M. Kobayashi, K. Araki, STI571 (Glivec) suppresses the expression of vascular endothelial growth factor in the gastrointestinal stromal tumor cell line, GIST-T1. *World Journal of Gastroenterology* **12**, 703-8 (2006). doi.org/10.3748/wjg.v12.i5.703 ncbi.nlm.nih.gov/pmc/PMC4066119
529. C. Kayabasi, T. B. Okcanoglu, B. O. Yelken, A. Asik, S. Y. Susluer, C. B. Avci, G. Saydam, C. Gunduz, Comparative effect of imatinib and ponatinib on autophagy and miRNome in chronic myeloid leukemia. *Gene* **637**, 173-80 (2017). doi.org/10.1016/j.gene.2017.09.036
530. N. Larmonier, N. Janikashvili, C. J. LaCasse, C. B. Larmonier, J. Cantrell, E. Situ, T. Lundeen, B. Bonnotte, E. Katsanis, Imatinib mesylate inhibits CD4+ CD25+ regulatory T cell activity and enhances active immunotherapy against BCR-ABL- tumors. *Journal of Immunology* **181**, 6955-63 (2008). doi.org/10.4049/jimmunol.181.10.6955 ncbi.nlm.nih.gov/pmc/PMC2579962

531. A. Tanaka, H. Nishikawa, S. Noguchi, D. Sugiyama, H. Morikawa et al., Tyrosine kinase inhibitor imatinib augments tumor immunity by depleting effector regulatory T cells. *The Journal of experimental medicine* **217**, (2020). doi.org/10.1084/jem.20191009 ncbi.nlm.nih.gov/pmc/PMC7041710
532. J. Litz, G. W. Krystal, Imatinib inhibits c-Kit-induced hypoxia-inducible factor-1 α activity and vascular endothelial growth factor expression in small cell lung cancer cells. *Molecular Cancer Therapeutics* **5**, 1415-22 (2006). doi.org/10.1158/1535-7163.MCT-05-0503
533. J. Liu, Y. Xiao, H. M. Xiong, J. Li, B. Huang, H. B. Zhang, D. Q. Feng, X. M. Chen, X. Z. Wang, Alternative splicing of apoptosis-related genes in imatinib-treated K562 cells identified by exon array analysis. *International Journal of Molecular Medicine* **29**, 690-8 (2012). doi.org/10.3892/ijmm.2011.872 ncbi.nlm.nih.gov/pmc/PMC3577368
534. X. Y. Liu, Y. F. Yang, C. T. Wu, F. J. Xiao, Q. W. Zhang, X. N. Ma, Q. F. Li, J. Yan, H. Wang, L. S. Wang, Spred2 is involved in imatinib-induced cytotoxicity in chronic myeloid leukemia cells. *Biochemical and Biophysical Research Communications* **393**, 637-42 (2010). doi.org/10.1016/j.bbrc.2010.02.044
535. Y. Yang, X. Liu, F. Xiao, S. Xue, Q. Xu, Y. Yin, H. Sun, J. Xu, H. Wang, Q. Zhang, H. Wang, L. Wang, Spred2 modulates the erythroid differentiation induced by imatinib in chronic myeloid leukemia cells. *PLoS One* **10**, e0117573 (2015). doi.org/10.1371/journal.pone.0117573 ncbi.nlm.nih.gov/pmc/PMC4331423
536. A. H. Hegde, J. Seetharamappa, Fluorescence and circular dichroism studies on binding and conformational aspects of an anti-leukemic drug with DNA. *Molecular Biology Reports* **41**, 67-71 (2014). doi.org/10.1007/s11033-013-2838-2
537. Y. Liu, M. Tseng, S. A. Perdreau, F. Rossi, C. Antonescu, P. Besmer, J. A. Fletcher, S. Duensing, A. Duensing, Histone H2AX is a mediator of gastrointestinal stromal tumor cell apoptosis following treatment with imatinib mesylate. *Cancer Research* **67**, 2685-92 (2007). doi.org/10.1158/0008-5472.CAN-06-3497
538. M. Okada, S. Adachi, T. Imai, K. Watanabe, S. Y. Toyokuni, M. Ueno, A. S. Zervos, G. Kroemer, T. Nakahata, A novel mechanism for imatinib mesylate-induced cell death of BCR-ABL-positive human leukemic cells: Caspase-independent, necrosis-like programmed cell death mediated by serine protease activity. *Blood* **103**, 2299-307 (2004). doi.org/10.1182/blood-2003-05-1605
539. P. Mlejnek, Can application of serine protease inhibitors TPCK and TLCK provide evidence for possible involvement of serine protease Omi/HtrA2 in imatinib mesylate-induced cell death of BCR-ABL-positive human leukemia cell? *Leukemia* **19**, 1085-7 (2005). doi.org/10.1038/sj.leu.2403701
540. M. F. Ochs, L. Rink, C. Tarn, S. Mburu, T. Taguchi, B. Eisenberg, A. K. Godwin, Detection of treatment-induced changes in signaling pathways in gastrointestinal stromal tumors using transcriptomic data. *Cancer Research* **69**, 9125-32 (2009). doi.org/10.1158/0008-5472.CAN-09-1709 ncbi.nlm.nih.gov/pmc/PMC2789202
541. L. C. Papadopoulou, A. V. Kyriazou, I. D. Bonovolias, A. S. Tsiftsoglou, Imatinib inhibits the expression of SCO2 and FRATAxin genes that encode mitochondrial proteins in human Bcr-Abl+ leukemia cells. *Blood Cells, Molecules, and Diseases* **53**, 84-90 (2014). doi.org/10.1016/j.bcmd.2014.03.001
542. A. Poggi, M. R. Zocchi, Imatinib mesylate can help to direct natural immunity toward an anti-leukemic reactivity by acting on the bone marrow microenvironment. *Oncobiology* **1**, 214-6 (2012). doi.org/10.4161/onci.1.2.18112 ncbi.nlm.nih.gov/pmc/PMC3377005
543. J. H. Ren, J. M. Luo, X. Y. Du, J. C. Yang, L. Yao, Y. T. Shang, X. J. Liu, Effects of imatinib on the expression of SHIP gene and apoptosis of K562 cells. *Tumor* **27**, 853-6 (2007).
544. N. Sato, M. Narita, M. Takahashi, K. Yagisawa, A. Liu, T. Abe, K. Nikkuni, T. Furukawa, K. Toba, Y. Aizawa, The effects of STI571 on antigen presentation of dendritic cells generated from patients with chronic myelogenous leukemia. *Hematological Oncology* **21**, 67-75 (2003). doi.org/10.1002/hon.705
545. Z. Yao, J. Zhang, B. Zhang, G. Liang, X. Chen, F. Yao, X. Xu, H. Wu, Q. He, L. Ding, B. Yang, Imatinib prevents lung cancer metastasis by inhibiting M2-like polarization of macrophages. *Pharmacological Research* **133**, 121-31 (2018). doi.org/10.1016/j.phrs.2018.05.002
546. S. A. S. Shandiz, M. Khosravani, S. Mohammadi, H. Noorbazargan, A. Mirzaie, D. N. Inanlou, M. D. Jalali, H. Jouzaghkar, F. Baghbani-Arani, B. Keshavarz-Pakseresht, Evaluation of imatinib mesylate (Gleevec) on KAI1/CD82 gene expression in breast cancer MCF-7 cells using quantitative real-time PCR. *Asian Pacific Journal of Tropical Biomedicine* **6**, 159-63 (2016). doi.org/10.1016/j.apjtb.2015.10.006
547. H. Wang, F. Cheng, A. Cuenda, P. Horna, Z. Zheng, K. Bhalla, E. M. Sotomayor, Imatinib mesylate (STI-571) enhances antigen-presenting cell function and overcomes tumor-induced CD4+ T-cell tolerance. *Blood* **105**, 1135-43 (2005). doi.org/10.1182/blood-2004-01-0027
548. P. Wihlidal, H. Karlic, M. Pfeilstöcker, K. Klaushofer, F. Varga, Imatinib mesylate (IM)-induced growth inhibition is associated with production of spliced osteocalcin-mRNA in cell lines. *Leukemia Research* **32**, 437-43 (2008). doi.org/10.1016/j.leukres.2007.07.020
549. J. B. Willig, D. R. B. Vianna, A. Beckenkamp, L. R. Beckenkamp, J. Sevigny, M. R. Wink, A. Buffon, D. A. Pilger, Imatinib mesylate affects extracellular ATP catabolism and expression of NTPDases in a chronic myeloid leukemia cell line. *Purinergic Signal* **16**, 29-40 (2020). doi.org/10.1007/s11302-019-09686-x ncbi.nlm.nih.gov/pmc/PMC7166234
550. F. Zheng, H. Li, K. Liang, Y. Du, D. Guo, S. Huang, Imatinib has the potential to exert its antileukemia effects by down-regulating hERG1 K⁺ channels in chronic myelogenous leukemia. *Medical Oncology* **29**, 2127-35 (2012). doi.org/10.1007/s12032-011-0102-y
551. B. Tandogan, C. Sengezer, N. N. Ulusu, In vitro effects of imatinib on glucose-6-phosphate dehydrogenase and glutathione reductase. *Folia Biologica* **57**, 57-64 (2011).
552. O. Uziel, E. Fenig, J. Nordenberg, E. Beery, H. Reshef, J. Sandbank, M. Birenbaum, M. Bakhanashvili, R. Yerushalmi, D. Luria, M. Lahav, Imatinib mesylate (Gleevec) downregulates telomerase activity and inhibits proliferation in telomerase-expressing cell lines. *British Journal of Cancer* **92**, 1881-91 (2005). doi.org/10.1038/sj.bjc.6602592 ncbi.nlm.nih.gov/pmc/PMC2361771

553. T. Vrekoussis, E. N. Stathopoulos, U. De Giorgi, M. Kafousi, K. Pavlaki, A. Kalogeraki, E. Chrysos, G. Fiorentini, O. Zoras, Modulation of vascular endothelium by imatinib: A study on the EA.hy 926 endothelial cell line. *Journal of Chemotherapy* **18**, 56-65 (2006). doi.org/10.1179/joc.2006.18.1.56
554. O. Hantschel, U. Rix, U. Schmidt, T. Bürkstümmer, M. Kneidinger, G. Schütze, J. Colinge, K. L. Bennett, W. Ellmeier, P. Valent, G. Superti-Furga, The Btk tyrosine kinase is a major target of the Bcr-Abl inhibitor dasatinib. *Proceedings of the National Academy of Sciences of the United States of America* **104**, 13283-8 (2007). doi.org/10.1073/pnas.0702654104
ncbi.nlm.nih.gov/pmc/PMC1940229
555. C. M. Chan, X. Jing, L. A. Pike, Q. Zhou, D. J. Lim, S. B. Sams, G. S. Lund, V. Sharma, B. R. Haugen, R. E. Schweppe, Targeted inhibition of Src kinase with dasatinib blocks thyroid cancer growth and metastasis. *Clinical Cancer Research* **18**, 3580-91 (2012). doi.org/10.1158/1078-0432.CCR-11-3359
ncbi.nlm.nih.gov/pmc/PMC3931551
556. A. Y. Chang, M. Wang, Molecular mechanisms of action and potential biomarkers of growth inhibition of dasatinib (BMS-354825) on hepatocellular carcinoma cells. *BMC Cancer* **13**, 267 (2013). doi.org/10.1186/1471-2407-13-267
ncbi.nlm.nih.gov/pmc/PMC3680296
557. Y. L. Choi, M. Bocanegra, M. J. Kwon, Y. K. Shin, S. J. Nam, J. H. Yang, J. Kao, A. K. Godwin, J. R. Pollack, LYN is a mediator of epithelial-mesenchymal transition and a target of dasatinib in breast cancer. *Cancer Research* **70**, 2296-306 (2010). doi.org/10.1158/0008-5472.CAN-09-3141 ncbi.nlm.nih.gov/pmc/PMC2869247
558. Y. J. Kim, S. Hong, M. Sung, M. J. Park, K. Jung, K. W. Noh, D. Y. Oh, M. S. Lee, E. Oh, Y. K. Shin, Y. L. Choi, LYN expression predicts the response to dasatinib in a subpopulation of lung adenocarcinoma patients. *Oncotarget* **7**, 82876-88 (2016). doi.org/10.18632/oncotarget.12657
ncbi.nlm.nih.gov/pmc/PMC5347739
559. A. von Mässenhausen, C. Sanders, J. Brägelmann, M. Konantz, A. Queisser, W. Vogel, G. Kristiansen, S. Duensing, A. Schröck, F. Bootz, P. Brossart, J. Kirfel, C. Lengerke, S. Perner, Targeting DDR2 in head and neck squamous cell carcinoma with dasatinib. *International Journal of Cancer* **139**, 2359-69 (2016). doi.org/10.1002/ijc.30279
560. J. Nautiyal, P. Majumder, B. B. Patel, F. Y. Lee, A. P. Majumdar, Src inhibitor dasatinib inhibits growth of breast cancer cells by modulating EGFR signaling. *Cancer Letters* **283**, 143-51 (2009). doi.org/10.1016/j.canlet.2009.03.035
561. Q. Chang, C. Jorgensen, T. Pawson, D. W. Hedley, Effects of dasatinib on EphA2 receptor tyrosine kinase activity and downstream signalling in pancreatic cancer. *British Journal of Cancer* **99**, 1074-82 (2008). doi.org/10.1038/sj.bjc.6604676
ncbi.nlm.nih.gov/pmc/PMC2567084
562. H. Ishigaki, T. Minami, O. Morimura, H. Kitai, D. Horio, Y. Koda, E. Fujimoto, Y. Negi, Y. Nakajima, M. Niki, S. Kanemura, E. Shibata, K. Mikami, R. Takahashi, T. Yokoi, K. Kurabayashi, T. Kijima, EphA2 inhibition suppresses proliferation of small-cell lung cancer cells through inducing cell cycle arrest. *Biochemical and Biophysical Research Communications* **519**, 846-53 (2019). doi.org/10.1016/j.bbrc.2019.09.076
563. R. Buettner, T. Mesa, A. Vultur, F. Lee, R. Jove, Inhibition of Src family kinases with dasatinib blocks migration and invasion of human melanoma cells. *Molecular Cancer Research* **6**, 1766-74 (2008). doi.org/10.1158/1541-7786.MCR-08-0169
ncbi.nlm.nih.gov/pmc/PMC2768340
564. A. Rohe, C. Henze, F. Erdmann, W. Sippl, M. Schmidt, A fluorescence anisotropy-based Myt1 kinase binding assay. *Assay and Drug Development Technologies* **12**, 136-44 (2014). doi.org/10.1089/adt.2013.534
565. S. Shapira, G. Granot, R. Mor-Tzuntz, P. Raanani, O. Uziel, M. Lahav, O. Shpilberg, Second-generation tyrosine kinase inhibitors reduce telomerase activity in K562 cells. *Cancer Letters* **323**, 223-31 (2012). doi.org/10.1016/j.canlet.2012.04.022
566. A. Gover-Proaktor, G. Granot, M. Pasmanik-Chor, O. Pasvolsky, S. Shapira, O. Raz, P. Raanani, A. Leader, Bosutinib, dasatinib, imatinib, nilotinib, and ponatinib differentially affect the vascular molecular pathways and functionality of human endothelial cells. *Leukemia & Lymphoma* **60**, 189-99 (2019). doi.org/10.1080/10428194.2018.1466294
567. K. Nunoda, T. Tauchi, T. Takaku, S. Okabe, D. Akahane, G. Sashida, J. H. Ohyashiki, K. Ohyashiki, Identification and functional signature of genes regulated by structurally different ABL kinase inhibitors. *Oncogene* **26**, 4179-88 (2007). doi.org/10.1038/sj.onc.1210179
568. E. B. Gencer, A. U. Ural, F. Avcu, Y. Baran, A novel mechanism of dasatinib-induced apoptosis in chronic myeloid leukemia; ceramide synthase and ceramide clearance genes. *Annals of Hematology* **90**, 1265-75 (2011). doi.org/10.1007/s00277-011-1212-5
569. A. Barbarin, M. Abdallah, L. Lefèvre, N. Piccirilli, E. Cayssials, L. Roy, J. M. Gombert, A. Herbelin, Innate T- $\alpha\beta$ lymphocytes as new immunological components of anti-tumoral "off-target" effects of the tyrosine kinase inhibitor dasatinib. *Scientific Reports* **10**, 3245 (2020). doi.org/10.1038/s41598-020-60195-z
ncbi.nlm.nih.gov/pmc/PMC7039999
570. I. Chamrád, U. Rix, A. Stukalov, M. Gridling, K. Parapatics, A. C. Müller, S. Altioik, J. Colinge, G. Superti-Furga, E. B. Haura, K. L. Bennett, A miniaturized chemical proteomic approach for target profiling of clinical kinase inhibitors in tumor biopsies. *Journal of Proteome Research* **12**, 4005-17 (2013). doi.org/10.1021/pr400309p
ncbi.nlm.nih.gov/pmc/PMC4127982
571. C. L. Chu, Y. P. Lee, C. Y. Pang, H. R. Lin, C. S. Chen, R. I. You, Tyrosine kinase inhibitors modulate dendritic cell activity via confining c-Kit signaling and tryptophan metabolism. *International Immunopharmacology* **82**, 106357 (2020). doi.org/10.1016/j.intimp.2020.106357
572. J. Li, U. Rix, B. Fang, Y. Bai, A. Edwards, J. Colinge, K. L. Bennett, J. Gao, L. Song, S. Eschrich, G. Superti-Furga, J. Koomen, E. B. Haura, A chemical and phosphoproteomic characterization of dasatinib action in lung cancer. *Nature Chemical Biology* **6**, 291-9 (2010). doi.org/10.1038/nchembio.332
ncbi.nlm.nih.gov/pmc/PMC2842457
573. C. Jenkins, S. B. Luty, J. E. Maxson, C. A. Eide, M. L. Abel, C. Togiai, E. R. Nemecek, D. Bottomly, S. K. McWeeney, B. Wilmut, M. Loriaux, B. H. Chang, J. W. Tyner, Synthetic lethality of TNK2 inhibition in PTPN11-mutant leukemia. *Science Signaling* **11**,

(2018). doi.org/10.1126/scisignal.aaq5617
ncbi.nlm.nih.gov/pmc/PMC6168748

574. D. H. Kim, S. Kamel-Reid, H. Chang, R. Sutherland, C. W. Jung, H. J. Kim, J. J. Lee, J. H. Lipton, Natural killer or natural killer/T cell lineage large granular lymphocytosis associated with dasatinib therapy for Philadelphia chromosome positive leukemia. *Haematologica* **94**, 135-9 (2009). doi.org/10.3324/haematol.13151
ncbi.nlm.nih.gov/pmc/PMC2625403

575. S. Mustjoki, M. Ekbom, T. P. Arstila, I. Dybedal, P. K. Epling-Burnette et al., Clonal expansion of T/NK-cells during tyrosine kinase inhibitor dasatinib therapy. *Leukemia* **23**, 1398-405 (2009). doi.org/10.1038/leu.2009.46

576. A. Kreutzman, V. Juvonen, V. Kairisto, M. Ekbom, L. Stenke, R. Seggewiss, K. Porkka, S. Mustjoki, Mono/oligoclonal T and NK cells are common in chronic myeloid leukemia patients at diagnosis and expand during dasatinib therapy. *Blood* **116**, 772-82 (2010). doi.org/10.1182/blood-2009-12-256800

577. C. A. Schiffer, J. E. Cortes, A. Hochhaus, G. Saglio, P. le Coutre, K. Porkka, S. Mustjoki, H. Mohamed, N. P. Shah, Lymphocytosis after treatment with dasatinib in chronic myeloid leukemia: Effects on response and toxicity. *Cancer* **122**, 1398-407 (2016). doi.org/10.1002/cncr.29933
ncbi.nlm.nih.gov/pmc/PMC5071708

578. T. Uchiyama, N. Sato, M. Narita, A. Yamahira, M. Iwabuchi, T. Furukawa, H. Sone, M. Takahashi, Direct effect of dasatinib on proliferation and cytotoxicity of natural killer cells in vitro study. *Hematological Oncology* **31**, 156-63 (2013). doi.org/10.1002/hon.2034

579. S. K. Heo, E. K. Noh, J. Y. Kim, Y. K. Jeong, J. C. Jo, Y. Choi, S. Koh, J. H. Baek, Y. J. Min, H. Kim, Targeting c-KIT (CD117) by dasatinib and radotinib promotes acute myeloid leukemia cell death. *Scientific Reports* **7**, 15278 (2017). doi.org/10.1038/s41598-017-15492-5
ncbi.nlm.nih.gov/pmc/PMC5681687

580. N. Iriyama, Y. Hatta, M. Takei, Direct effect of dasatinib on signal transduction pathways associated with a rapid mobilization of cytotoxic lymphocytes. *Cancer Medicine* **5**, 3223-34 (2016). doi.org/10.1002/cam4.925
ncbi.nlm.nih.gov/pmc/PMC5119978

581. N. Iriyama, H. Takahashi, K. Miura, Y. Uchino, M. Nakagawa, Y. Hatta, M. Takei, Enhanced perforin expression associated with dasatinib therapy in natural killer cells. *Leukemia Research* **68**, 1-8 (2018). doi.org/10.1016/j.leukres.2018.02.014

582. Y. C. Lin, M. H. Wu, T. T. Wei, S. H. Chuang, K. F. Chen, A. L. Cheng, C. C. Chen, Degradation of epidermal growth factor receptor mediates dasatinib-induced apoptosis in head and neck squamous cell carcinoma cells. *Neoplasia* **14**, 463-75 (2012). doi.org/10.1593/neo.12300
ncbi.nlm.nih.gov/pmc/PMC3394189

583. J. Liu, Y. Chen, J. Zhang, Y. Wang, Y. Zhang, Y. Liu, Mechanism study of dasatinib inhibiting phosphorylation of androgen receptor in prostatic cancer cells. *Cancer Research and Clinic* **28**, 361-5 (2016). doi.org/10.3760/cma.j.issn.1006-9801.2016.06.001

584. Y. Najima, C. Yoshida, N. Iriyama, S. Fujisawa, H. Wakita, S. Chiba, S. Okamoto, K. Kawakami, N. Takezako, T. Kumagai, K. Ohyashiki, J. Taguchi, S. Yano, T. Igarashi, Y. Kouzai, S. Morita, J. Sakamoto, H. Sakamaki, K. Inokuchi, Regulatory T cell inhibition by dasatinib is associated with natural killer cell differentiation and a favorable molecular response—The final results of the D-first study.

Leukemia Research **66**, 66-72 (2018). doi.org/10.1016/j.leukres.2018.01.010

585. M. Wölfel, S. Schwinn, Y. E. Yoo, M. L. Reß, M. Braun, M. Chopra, S. C. Schreiber, V. I. Ayala, C. Ohlen, M. Eyrich, A. Beilhack, P. G. Schlegel, Src-kinase inhibitors sensitize human cells of myeloid origin to Toll-like-receptor-induced interleukin 12 synthesis. *Blood* **122**, 1203-13 (2013). doi.org/10.1182/blood-2013-03-488072
ncbi.nlm.nih.gov/pmc/PMC3744989

586. Y. Yang, C. Liu, W. Peng, G. Lizée, W. W. Overwijk, Y. Liu, S. E. Woodman, P. Hwu, Antitumor T-cell responses contribute to the effects of dasatinib on c-KIT mutant murine mastocytoma and are potentiated by anti-OX40. *Blood* **120**, 4533-43 (2012). doi.org/10.1182/blood-2012-02-407163
ncbi.nlm.nih.gov/pmc/PMC3512233

587. J. Zhang, Y. Chen, Q. He, Distinct characteristics of dasatinib-induced pyroptosis in gasdermin E-expressing human lung cancer A549 cells and neuroblastoma SH-SY5Y cells. *Oncology Letters* **20**, 145-54 (2020). doi.org/10.3892/ol.2020.11556
ncbi.nlm.nih.gov/pmc/PMC7285962

588. L. L. Remsing Rix, U. Rix, J. Colinge, O. Hantschel, K. L. Bennett, T. Stranzl, A. Muller, C. Baumgartner, P. Valent, M. Augustin, J. H. Till, G. Superti-Furga, Global target profile of the kinase inhibitor bosutinib in primary chronic myeloid leukemia cells. *Leukemia* **23**, 477-85 (2009). doi.org/10.1038/leu.2008.334

589. M. Jeitany, C. Leroy, P. Tosti, M. Lafitte, J. Le Guet, V. Simon, D. Bonenfant, B. Robert, F. Grillet, C. Mollevi, S. El Messaoudi, A. Otandault, L. Canterel-Thouennon, M. Busson, A. R. Thierry, P. Martineau, J. Pannequin, S. Roche, A. Sirvent, Inhibition of DDR1-BCR signalling by nilotinib as a new therapeutic strategy for metastatic colorectal cancer. *EMBO Molecular Medicine* **10**, (2018). doi.org/10.15252/emmm.201707918
ncbi.nlm.nih.gov/pmc/PMC5887546

590. E. H. Stover, J. Chen, B. H. Lee, J. Cools, E. McDowell, J. Adelsperger, D. Cullen, A. Coburn, S. A. Moore, R. Okabe, D. Fabbro, P. W. Manley, J. D. Griffin, D. G. Gilliland, The small molecule tyrosine kinase inhibitor AMN107 inhibits TEL-PDGFRbeta and FIP1L1-PDGFRalpha in vitro and in vivo. *Blood* **106**, 3206-13 (2005). doi.org/10.1182/blood-2005-05-1932
ncbi.nlm.nih.gov/pmc/PMC1895333

591. A. Camgoz, E. B. Gencer, A. U. Ural, F. Avcu, Y. Baran, Roles of ceramide synthase and ceramide clearance genes in nilotinib-induced cell death in chronic myeloid leukemia cells. *Leukemia & Lymphoma* **52**, 1574-84 (2011). doi.org/10.3109/10428194.2011.568653

592. K. K. Chahal, J. Li, I. Kufareva, M. Parle, D. L. Durden, R. J. Wechsler-Reya, C. C. Chen, R. Abagyan, Nilotinib, an approved leukemia drug, inhibits smoothed signaling in Hedgehog-dependent medulloblastoma. *PloS One* **14**, e0214901 (2019). doi.org/10.1371/journal.pone.0214901

593. H. C. Yu, C. S. Lin, W. T. Tai, C. Y. Liu, C. W. Shiau, K. F. Chen, Nilotinib induces autophagy in hepatocellular carcinoma through AMPK activation. *Journal of Biological Chemistry* **288**, 18249-59 (2013). doi.org/10.1074/jbc.M112.446385
ncbi.nlm.nih.gov/pmc/PMC3689967

594. H. Zhang, L. Gu, T. Liu, K. Y. Chiang, M. Zhou, Inhibition of MDM2 by nilotinib contributes to cytotoxicity in both Philadelphia-

- positive and negative acute lymphoblastic leukemia. *PLoS One* **9**, e100960 (2014). doi.org/10.1371/journal.pone.0100960 ncbi.nlm.nih.gov/pmc/PMC4072773
595. S. Ahmad, G. L. Johnson, J. E. Scott, Identification of ponatinib and other known kinase inhibitors with potent MEKK2 inhibitory activity. *Biochemical and Biophysical Research Communications* **463**, 888-93 (2015). doi.org/10.1016/j.bbrc.2015.06.029 ncbi.nlm.nih.gov/pmc/PMC4500090
596. N. Ai, C. M. Chong, W. Chen, Z. Hu, H. Su, G. Chen, Q. W. Lei Wong, W. Ge, Ponatinib exerts anti-angiogenic effects in the zebrafish and human umbilical vein endothelial cells via blocking VEGFR signaling pathway. *Oncotarget* **9**, 31958-70 (2018). doi.org/10.18632/oncotarget.24110 ncbi.nlm.nih.gov/pmc/PMC6112840
597. J. M. Gozgit, M. J. Wong, L. Moran, S. Wardwell, Q. K. Mohammad, N. I. Narasimhan, W. C. Shakespeare, F. Wang, T. Clackson, V. M. Rivera, Ponatinib (AP24534), a multitargeted pan-FGFR inhibitor with activity in multiple FGFR-amplified or mutated cancer models. *Molecular Cancer Therapeutics* **11**, 690-9 (2012). doi.org/10.1158/1535-7163.MCT-11-0450
598. J. D. Lang, W. P. D. Hendricks, K. A. Orlando, H. Yin, J. Kiefer et al., Ponatinib shows potent antitumor activity in small cell carcinoma of the ovary hypercalcemic type (SCCOHT) through multikinase inhibition. *Clinical Cancer Research* **24**, 1932-43 (2018). doi.org/10.1158/1078-0432.CCR-17-1928 ncbi.nlm.nih.gov/pmc/PMC6526947
599. R. Madonna, D. Pieragostino, M. C. Cufaro, V. Doria, P. Del Boccio, M. Deidda, S. D. Pierdomenico, C. C. Dessalvi, R. De Caterina, G. Mercuro, Ponatinib Induces Vascular Toxicity through the Notch-1 Signaling Pathway. *Journal of Clinical Medicine* **9**, (2020). doi.org/10.3390/jcm9030820 ncbi.nlm.nih.gov/pmc/PMC7141219
600. S. Ramirez-Rios, S. Michallet, L. Peris, C. Barette, C. Rabat, Y. Feng, M. O. Fauvarque, A. Andrieux, K. Sadoul, L. Lafanechere, A new quantitative cell-based assay reveals unexpected microtubule stabilizing activity of certain kinase inhibitors, clinically approved or in the process of approval. *Frontiers in Pharmacology* **11**, 543 (2020). doi.org/10.3389/fphar.2020.00543 ncbi.nlm.nih.gov/pmc/PMC7204994
601. D. S. Tan, B. Haaland, J. M. Gan, S. C. Tham, I. Sinha, E. H. Tan, K. H. Lim, A. Takano, S. S. Krisna, M. M. Thu, H. P. Liew, A. Ullrich, W. T. Lim, B. T. Chua, Bosutinib inhibits migration and invasion via ACK1 in KRAS mutant non-small cell lung cancer. *Molecular Cancer* **13**, 13 (2014). doi.org/10.1186/1476-4598-13-13 ncbi.nlm.nih.gov/pmc/PMC3930897
602. C. C. Poon, J. J. Kelly, Development of crizotinib, a rationally designed tyrosine kinase inhibitor for non-small cell lung cancer. *International Journal of Cancer* **140**, 1945-54 (2017). doi.org/10.1002/ijc.30533
603. X. Chen, J. Ji, W. Chen, W. Lian, R. Chen, J. Yang, Q. Zhang, Q. Weng, Z. Khan, J. Hu, X. Chan, P. Zou, G. Liang, (S)-crizotinib reduces gastric cancer growth through oxidative DNA damage and triggers pro-survival akt signal. *Cell Death & Disease* **9**, (2018). doi.org/10.1038/s41419-018-0667-x ncbi.nlm.nih.gov/pmc/PMC5981313
604. X. Dai, G. Guo, P. Zou, R. Cui, W. Chen, X. Chen, C. Yin, W. He, R. Vinothkumar, F. Yang, X. Zhang, G. Liang, (S)-crizotinib induces apoptosis in human non-small cell lung cancer cells by activating ROS independent of MTH1. *Journal of Experimental and Clinical Cancer Research* **36**, 120 (2017). doi.org/10.1186/s13046-017-0584-3 ncbi.nlm.nih.gov/pmc/PMC5590185
605. F. Megiorni, H. P. McDowell, S. Camero, O. Mannarino, S. Ceccarelli, M. Paiano, P. D. Losty, B. Pizer, R. Shukla, A. Pizzuti, A. Clerico, C. Dominici, Crizotinib-induced antitumour activity in human alveolar rhabdomyosarcoma cells is not solely dependent on ALK and MET inhibition. *Journal of Experimental and Clinical Cancer Research* **34**, (2015). doi.org/10.1186/s13046-015-0228-4 ncbi.nlm.nih.gov/pmc/PMC4596370
606. F. S. Hamedani, M. Cinar, Z. Mo, M. A. Cervania, H. M. Amin, S. Alkan, Crizotinib (PF-2341066) induces apoptosis due to downregulation of pSTAT3 and BCL-2 family proteins in NPM-ALK+ anaplastic large cell lymphoma. *Leukemia Research* **38**, 503-8 (2014). doi.org/10.1016/j.leukres.2013.12.027 ncbi.nlm.nih.gov/pmc/PMC4386887
607. P. Liu, L. Zhao, J. Pol, S. Levesque, A. Petrazzuolo et al., Crizotinib-induced immunogenic cell death in non-small cell lung cancer. *Nature Communications* **10**, 1486 (2019). doi.org/10.1038/s41467-019-09415-3 ncbi.nlm.nih.gov/pmc/PMC6445096
608. S. Troutman, S. Moleirinho, S. Kota, K. Nettles, M. Fallahi, G. L. Johnson, J. L. Kissil, Crizotinib inhibits NF2-associated schwannoma through inhibition of focal adhesion kinase 1. *Oncotarget* **7**, 54515-25 (2016). doi.org/10.18632/oncotarget.10248 ncbi.nlm.nih.gov/pmc/PMC5342359
609. Y. Zhou, C. Zhao, S. Gery, G. D. Braunstein, R. Okamoto, R. Alvarez, S. A. Miles, N. B. Doan, J. W. Said, J. Gu, H. Phillip Koeffler, Off-target effects of c-MET inhibitors on thyroid cancer cells. *Molecular Cancer Therapeutics* **13**, 134-43 (2014). doi.org/10.1158/1535-7163.MCT-13-0187 ncbi.nlm.nih.gov/pmc/PMC3947168
610. M. Zillhardt, J. G. Christensen, E. Lengyel, An orally available small-molecule inhibitor of c-Met, PF-2341066, reduces tumor burden and metastasis in a preclinical model of ovarian cancer metastasis. *Neoplasia* **12**, 1-10 (2010). doi.org/10.1593/neo.09948 ncbi.nlm.nih.gov/pmc/PMC2805878
611. C. Grüllich, "Cabozantinib: Multi-kinase inhibitor of MET, AXL, RET, and VEGFR2" in *Small Molecules in Oncology* U. M. Martens, Ed, in series: Recent Results in Cancer Research. vol. 211 (Springer, 2018), pp. 67-75. http://doi.org/10.1007/978-3-319-91442-8_5
612. T. Hara, A. Kimura, T. Miyazaki, H. Tanaka, M. Morimoto, K. Nakai, J. Soeda, Cabozantinib inhibits AXL- and MET-dependent cancer cell migration induced by growth-arrest-specific 6 and hepatocyte growth factor. *Biochemistry Biophysics Reports* **21**, 100726 (2020). doi.org/10.1016/j.bbrep.2020.100726 ncbi.nlm.nih.gov/pmc/PMC7005370
613. C. Reuther, V. Heinze, M. Spampatti, G. Vlotides, E. de Toni, G. Spottl, J. Maurer, S. Nolting, B. Goke, C. J. Auernhammer, Cabozantinib and tivantinib, but not INC280, induce antiproliferative and antimigratory effects in human neuroendocrine tumor cells in vitro: Evidence for 'off-target' effects not mediated by c-Met

- inhibition. *Neuroendocrinology* **103**, 383-401 (2016). doi.org/10.1159/000439431
614. D. Verduzco, B. M. Kuenzi, F. Kinose, V. K. Sondak, Z. Eroglu, U. Rix, K. S. M. Smalley, Ceritinib enhances the efficacy of trametinib in BRAF/NRAS-wild-type melanoma cell lines. *Molecular Cancer Therapeutics* **17**, 73-83 (2018). doi.org/10.1158/1535-7163.MCT-17-0196 ncbi.nlm.nih.gov/pmc/PMC5752595
615. B. M. Kuenzi, L. L. Remsing Rix, P. A. Stewart, B. Fang, F. Kinose, A. T. Bryant, T. A. Boyle, J. M. Koomen, E. B. Haura, U. Rix, Polypharmacology-based ceritinib repurposing using integrated functional proteomics. *Nature Chemical Biology* **13**, 1222-31 (2017). doi.org/10.1038/nchembio.2489 ncbi.nlm.nih.gov/pmc/PMC5909815
616. I. Faraoni, F. Aloisio, A. De Gabrieli, M. I. Consalvo, S. Lavorgna, M. T. Voso, F. Lo-Coco, G. Graziani, The poly(ADP-ribose) polymerase inhibitor olaparib induces up-regulation of death receptors in primary acute myeloid leukemia blasts by NF-κB activation. *Cancer Letters* **423**, 127-38 (2018). doi.org/10.1016/j.canlet.2018.03.008
617. Y. Y. Ma, B. Yuan, W. B. Zhao, D. M. Zhao, Effect of olaparib on ovarian cancer SKOV3 cell line proliferation by inhibiting the PI3K/AKT/mTOR signaling pathway. *Chinese Pharmaceutical Journal* **54**, 36-41 (2019). doi.org/10.11669/cpj.2019.01.008
618. Z. Wang, J. Gao, J. Zhou, H. Liu, C. Xu, Olaparib induced senescence under P16 or P53 dependent manner in ovarian cancer. *Journal of Gynecologic Oncology* **30**, e26 (2019). doi.org/10.3802/jgo.2019.30.e26 ncbi.nlm.nih.gov/pmc/Selleck
619. Y. Shen, M. Aoyagi-Scharber, B. Wang, Trapping poly(ADP-ribose) polymerase. *Journal of Pharmacology and Experimental Therapeutics* **353**, 446-57 (2015). doi.org/10.1124/jpet.114.222448
620. A. A. Antolín, J. Mestres, Linking off-target kinase pharmacology to the differential cellular effects observed among PARP inhibitors. *Oncotarget* **5**, 3023-8 (2014). doi.org/10.18632/oncotarget.1814 ncbi.nlm.nih.gov/pmc/PMC4102788
621. C. E. Knezevic, G. Wright, L. L. Remsing Rix, W. Kim, B. M. Kuenzi, Y. Luo, J. M. Watters, J. M. Koomen, E. B. Haura, A. N. Monteiro, C. Radu, H. R. Lawrence, U. Rix, Proteome-wide profiling of clinical PARP inhibitors reveals compound-specific secondary targets. *Cell Chemical Biology* **23**, 1490-503 (2016). doi.org/10.1016/j.chembiol.2016.10.011 ncbi.nlm.nih.gov/pmc/PMC5182133
622. C. M. McCrudden, M. G. O'Rourke, K. E. Cherry, H. F. Yuen, D. O'Rourke, M. Babur, B. A. Telfer, H. D. Thomas, P. Keane, T. Nambirajan, C. Hagan, J. M. O'Sullivan, C. Shaw, K. J. Williams, N. J. Curtin, D. G. Hirst, T. Robson, Vasoactivity of rucaparib, a PARP-1 inhibitor, is a complex process that involves myosin light chain kinase, P2 receptors, and PARP itself. *PloS One* **10**, e0118187 (2015). doi.org/10.1371/journal.pone.0118187 ncbi.nlm.nih.gov/pmc/PMC4331495
623. Y. Nonomiya, K. Noguchi, K. Katayama, Y. Sugimoto, Novel pharmacological effects of poly (ADP-ribose) polymerase inhibitor rucaparib on the lactate dehydrogenase pathway. *Biochemical and Biophysical Research Communications* **510**, 501-7 (2019). doi.org/10.1016/j.bbrc.2019.01.133
624. Y. Wang, Y. Zheng, Y. Hao, Rucaparib (Rubraca®) induces necrosis via upregulating the expression of RIP1 and RIP3 in ovarian cancer cells. *Pharmazie* **75**, 242-5 (2020). doi.org/10.1691/ph.2020.9827
625. J. Huang, L. Wang, Z. Cong, Z. Amoozgar, E. Kiner, D. Xing, S. Orsulic, U. Matulonis, M. S. Goldberg, The PARP1 inhibitor BMN 673 exhibits immunoregulatory effects in a Brcal -/- murine model of ovarian cancer. *Biochemical and Biophysical Research Communications* **463**, 551-6 (2015). doi.org/10.1016/j.bbrc.2015.05.083
626. O. Kruglov, X. Wu, S. T. Hwang, O. E. Akilov, The synergistic proapoptotic effect of PARP-1 and HDAC inhibition in cutaneous T-cell lymphoma is mediated via Blimp-1. *Blood Advances* **4**, 4788-97 (2020). doi.org/10.1182/bloodadvances.2020002049 ncbi.nlm.nih.gov/pmc/PMC7556155
627. M. F. Adasme, D. Parisi, K. Van Belle, S. Salentin, V. J. Haupt, G. S. Jennings, J. C. Heinrich, J. Herman, B. Sprangers, T. Louat, Y. Moreau, M. Schroeder, Structure-based drug repositioning explains ibrutinib as VEGFR2 inhibitor. *PloS One* **15**, e0233089 (2020). doi.org/10.1371/journal.pone.0233089 ncbi.nlm.nih.gov/pmc/PMC7252619
628. V. Barbarino, S. Henschke, S. J. Blakemore, E. Izquierdo, M. Michalik, N. Nickel, I. Möllenkotte, D. Vorholt, L. Müller, R. Brinker, O. Fedorchenko, N. Mikhael, T. Seeger-Nukpezah, M. Hallek, C. P. Pallasch, Macrophage-mediated antibody dependent effector function in aggressive B-cell lymphoma treatment is enhanced by ibrutinib via inhibition of JAK2. *Cancers* **12**, 1-25 (2020). doi.org/10.3390/cancers12082303 ncbi.nlm.nih.gov/pmc/PMC7465917
629. L. A. Honigberg, A. M. Smith, M. Sirisawad, E. Verner, D. Loury, B. Chang, S. Li, Z. Pan, D. H. Thamm, R. A. Miller, J. J. Buggy, The bruton tyrosine kinase inhibitor PCI-32765 blocks B-cell activation and is efficacious in models of autoimmune disease and B-cell malignancy. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 13075-80 (2010). doi.org/10.1073/pnas.1004594107 ncbi.nlm.nih.gov/pmc/PMC2919935
630. J. Chen, T. Kinoshita, J. Sukbuntherng, B. Y. Chang, L. Elias, Ibrutinib inhibits ERBB receptor tyrosine kinases and HER2-amplified breast cancer cell growth. *Molecular Cancer Therapeutics* **15**, 2835-44 (2016). doi.org/10.1158/1535-7163.MCT-15-0923
631. I. De Weerdt, T. Hofland, R. Lameris, S. Endstra, A. Jongejan, P. D. Moerland, R. C. G. De Bruin, E. B. M. Remmerswaal, I. J. M. Ten Berge, N. Liu, M. Van Der Stelt, L. M. Faber, M. D. Levin, E. Eldering, S. H. Tonino, T. D. De Gruijl, H. J. Van Der Vliet, A. P. Kater, Improving CLL Vγ9Vδ2-T-cell fitness for cellular therapy by ex vivo activation and ibrutinib. *Blood* **132**, 2260-72 (2018). doi.org/10.1182/blood-2017-12-822569
632. J. A. Dubovsky, K. A. Beckwith, G. Natarajan, J. A. Woyach, S. Jaglowski et al., Ibrutinib is an irreversible molecular inhibitor of ITK driving a Th1-selective pressure in T lymphocytes. *Blood* **122**, 2539-49 (2013). doi.org/10.1182/blood-2013-06-507947 ncbi.nlm.nih.gov/pmc/PMC3795457
633. S. Fioreari, R. Maffei, V. Audrito, S. Martinelli, E. Hacken, P. Zucchini, G. Grisendi, L. Potenza, M. Luppi, J. A. Burger, S. Deaglio, R. Marasca, Ibrutinib modifies the function of monocyte/macrophage population in chronic lymphocytic leukemia.

Oncotarget 7, 65968-81 (2016). doi.org/10.18632/oncotarget.11782
ncbi.nlm.nih.gov/pmc/PMC5323207

634. X. Han, J. Zhang, D. Shi, Y. Wu, R. Liu, T. Liu, J. Xu, X. Yao, J. Fang, Targeting thioredoxin reductase by ibrutinib promotes apoptosis of SMMC-7721 cells. *Journal of Pharmacology and Experimental Therapeutics* 369, 212-22 (2019).
doi.org/10.1124/jpet.118.254862

635. E. Kim, C. Hurtz, S. Koehler, Z. Wang, S. Balasubramanian, B. Y. Chang, M. Müschen, R. E. Davis, J. A. Burger, Ibrutinib inhibits pre-BCR+ B-cell acute lymphoblastic leukemia progression by targeting BTK and BLK. *Blood* 129, 1155-65 (2017).
doi.org/10.1182/blood-2016-06-722900
ncbi.nlm.nih.gov/pmc/PMC5374732

636. P. P. Li, K. Lu, L. Y. Geng, X. X. Zhou, X. Y. Li, X. Wang, Bruton's tyrosine kinase inhibitor restrains Wnt signaling in chronic lymphocytic leukemia. *Molecular Medicine Reports* 13, 4934-8 (2016). doi.org/10.3892/mmr.2016.5111

637. Z. Li, J. Wu, L. Sheng, Ibrutinib improves the development of acute lymphoblastic leukemia by activating endoplasmic reticulum stress-induced cell death. *Pharmazie* 73, 294-9 (2018).
doi.org/10.1691/ph.2018.8306

638. V. Patel, K. Balakrishnan, E. Bibikova, M. Ayres, M. J. Keating, W. G. Wierda, V. Gandhi, Comparison of acalabrutinib, a selective Bruton tyrosine kinase inhibitor, with ibrutinib in chronic lymphocytic leukemia cells. *Clinical Cancer Research* 23, 3734-43 (2017). doi.org/10.1158/1078-0432.CCR-16-1446
ncbi.nlm.nih.gov/pmc/PMC5491371

639. M. Long, K. Beckwith, P. Do, B. L. Mundy, A. Gordon et al., Ibrutinib treatment improves T cell number and function in CLL patients. *Journal of Clinical Investigation* 127, 3052-64 (2017).
doi.org/10.1172/JCI89756 ncbi.nlm.nih.gov/pmc/PMC5531425

640. J. Ma, W. Gong, S. Liu, Q. Li, M. Guo, J. Wang, S. Wang, N. Chen, Y. Wang, Q. Liu, H. Zhao, Ibrutinib targets microRNA-21 in multiple myeloma cells by inhibiting NF-κB and STAT3. *Tumor Biology* 40, 1010428317731369 (2018).
doi.org/10.1177/1010428317731369

641. P. E. Morande, M. Sivina, A. Uriepero, N. Seija, C. Berca, P. Fresia, A. I. Landoni, J. M. Di Noia, J. A. Burger, P. Oppezzo, Ibrutinib therapy downregulates AID enzyme and proliferative fractions in chronic lymphocytic leukemia. *Blood* 133, 2056-68 (2019). doi.org/10.1182/blood-2018-09-876292
ncbi.nlm.nih.gov/pmc/PMC7022232

642. M. Palma, A. Krstic, L. Peña Perez, A. Berglöf, S. Meinke, Q. Wang, K. E. M. Blomberg, M. Kamali-Moghaddam, Q. Shen, G. Jaremko, J. Lundin, A. De Paepe, P. Höglund, E. Kimby, A. Österborg, R. Måansson, C. I. E. Smith, Ibrutinib induces rapid downregulation of inflammatory markers and altered transcription of chronic lymphocytic leukaemia-related genes in blood and lymph nodes. *British Journal of Haematology* 183, 212-24 (2018).
doi.org/10.1111/bjh.15516

643. L. Ping, N. Ding, Y. Shi, L. Feng, J. Li, Y. Liu, Y. Lin, C. Shi, X. Wang, Z. Pan, Y. Song, J. Zhu, The Bruton's tyrosine kinase inhibitor ibrutinib exerts immunomodulatory effects through regulation of tumor-infiltrating macrophages. *Oncotarget* 8, 39218-29 (2017). doi.org/10.18632/oncotarget.16836
ncbi.nlm.nih.gov/pmc/PMC5503608

644. C. B. Prabaharan, A. B. Yang, D. Chidambaram, K. Rajamanickam, S. Napper, M. K. Sakharkar, Ibrutinib as a potential therapeutic option for HER2 overexpressing breast cancer – the role of STAT3 and p21. *Investigational New Drugs* 38, 909-21 (2020).
doi.org/10.1007/s10637-019-00837-w

645. F. Rauf, F. Festa, J. G. Park, M. Magee, S. Eaton, C. Rinaldi, C. M. Betanzos, L. Gonzalez-Malerva, J. Labaer, Ibrutinib inhibition of ERBB4 reduces cell growth in a WNT5A-dependent manner. *Oncogene* 37, 2237-50 (2018). doi.org/10.1038/s41388-017-0079-x
ncbi.nlm.nih.gov/pmc/PMC5916919

646. U. Rozovski, D. M. Harris, P. Li, Z. Liu, P. Jain, A. Ferrajoli, J. Burger, P. Thompson, N. Jain, W. Wierda, M. J. Keating, Z. Estrov, Ibrutinib inhibits free fatty acid metabolism in chronic lymphocytic leukemia. *Leukemia and Lymphoma* 59, 2686-91 (2018).
doi.org/10.1080/10428194.2018.1439167
ncbi.nlm.nih.gov/pmc/PMC6135679

647. A. Wang, X. E. Yan, H. Wu, W. Wang, C. Hu et al., Ibrutinib targets mutant-EGFR kinase with a distinct binding conformation. *Oncotarget* 7, 69760-9 (2016). doi.org/10.18632/oncotarget.11951
ncbi.nlm.nih.gov/pmc/PMC5342513

648. X. Wang, J. Wong, C. J. Sevinsky, L. Kokabee, F. Khan, Y. Sun, D. S. Conklin, Bruton's tyrosine kinase inhibitors prevent therapeutic escape in breast cancer cells. *Molecular Cancer Therapeutics* 15, 2198-208 (2016). doi.org/10.1158/1535-7163.MCT-15-0813 ncbi.nlm.nih.gov/pmc/PMC5145257

649. L. M. Saleh, W. Wang, S. E. M. Herman, N. S. Saba, V. Anastas, E. Barber, M. Corrigan-Cummins, M. Farooqui, C. Sun, S. M. Sarasua, Z. Zhao, N. K. Abousamra, O. Elbaz, H. A. Abdelghaffar, A. Wiestner, K. R. Calvo, Ibrutinib downregulates a subset of miRNA leading to upregulation of tumor suppressors and inhibition of cell proliferation in chronic lymphocytic leukemia. *Leukemia* 31, 340-9 (2017). doi.org/10.1038/leu.2016.181

650. Y. Shen, O. G. Best, S. P. Mulligan, R. I. Christopherson, Ibrutinib and idelalisib block immunophenotypic changes associated with the adhesion and activation of CLL cells in the tumor microenvironment. *Leukemia and Lymphoma* 59, 1927-37 (2018).
doi.org/10.1080/10428194.2017.1403598

651. G. Yang, S. J. Buhrlage, L. Tan, X. Liu, J. Chen, L. Xu, N. Tsakmaklis, J. G. Chen, C. J. Patterson, J. R. Brown, J. J. Castillo, W. Zhang, X. Zhang, S. Liu, P. Cohen, Z. R. Hunter, N. Gray, S. P. Treon, HCK is a survival determinant transactivated by mutated MYD88, and a direct target of ibrutinib. *Blood* 127, 3237-52 (2016).
doi.org/10.1182/blood-2016-01-695098

652. P. Dwivedi, S. Chutipongtanate, D. E. Muench, M. Azam, H. L. Grimes, K. D. Greis, SWATH-proteomics of ibrutinib's action in myeloid leukemia initiating mutated G-CSFR signaling. *Proteomics - Clinical Applications* 14, e1900144 (2020).
doi.org/10.1002/prca.201900144
ncbi.nlm.nih.gov/pmc/PMC7492401

653. N. Andrae, E. Kirches, R. Hartig, D. Haase, G. Keilhoff, T. Kalinski, C. Mawrin, Sunitinib targets PDGF-receptor and Flt3 and reduces survival and migration of human meningioma cells. *European Journal of Cancer* 48, 1831-41 (2012).
doi.org/10.1016/j.ejca.2012.01.032

654. A. Brossa, C. Grange, L. Mancuso, L. Annaratone, M. A. Satolli, M. Mazzone, G. Camussi, B. Bussolati, Sunitinib but not

- VEGF blockade inhibits cancer stem cell endothelial differentiation. *Oncotarget* **6**, 11295-309 (2015). doi.org/10.18632/oncotarget.3123 [ncbi.nlm.nih.gov/pmc/PMC4484457](https://www.ncbi.nlm.nih.gov/pmc/PMC4484457)
655. L. Q. M. Chow, S. G. Eckhardt, Sunitinib: From rational design to clinical efficacy. *Journal of Clinical Oncology* **25**, 884-96 (2007). doi.org/10.1200/JCO.2006.06.3602
656. B. Hou, G. Wang, Q. Gao, Y. Wei, C. Zhang, Y. Wang, Y. Huo, H. Yang, X. Jiang, Z. Xi, SQSTM1/p62 loss reverses the inhibitory effect of sunitinib on autophagy independent of AMPK signaling. *Scientific Reports* **9**, 11087 (2019). doi.org/10.1038/s41598-019-47597-4 [ncbi.nlm.nih.gov/pmc/PMC6668422](https://www.ncbi.nlm.nih.gov/pmc/PMC6668422)
657. S. J. Jin, L. Ma, Q. Xu, L. P. Ren, Y. C. Ma, Sunitinib treatment inhibited human breast cancer cell migration through regulation furin interaction with substrates. *International Journal of Clinical and Experimental Medicine* **9**, 2535-41 (2016).
658. H. M. Korashy, Z. H. Maayah, F. E. Al Anazi, A. M. Alsaad, I. O. Alanazi, O. M. Belali, F. O. Al-Atawi, A. Alshamsan, Sunitinib inhibits breast cancer cell proliferation by inducing apoptosis, cell-cycle arrest and DNA repair while inhibiting NF- κ B signaling pathways. *Anticancer Research* **37**, 4899-909 (2017). doi.org/10.21873/anticancres.11899
659. K. R. Laderoute, J. M. Calaoagan, P. B. Madrid, A. E. Klon, P. J. Ehrlich, SU11248 (sunitinib) directly inhibits the activity of mammalian 5'-AMP-activated protein kinase (AMPK). *Cancer Biology & Therapy* **10**, 68-76 (2010). doi.org/10.4161/cbt.10.1.12162 [ncbi.nlm.nih.gov/pmc/PMC3087946](https://www.ncbi.nlm.nih.gov/pmc/PMC3087946)
660. E. I. Papadopoulos, G. M. Yousef, A. Scorilas, Cytotoxic activity of sunitinib and everolimus in Caki-1 renal cancer cells is accompanied by modulations in the expression of apoptosis-related microRNA clusters and BCL2 family genes. *Biomedicine and Pharmacotherapy* **70**, 33-40 (2015). doi.org/10.1016/j.biopha.2014.12.043
661. H. Polena, J. Creuzet, M. Dufies, A. Sidibé, A. Khalil-Mgharbel, A. Salomon, A. Deroux, J. L. Quesada, C. Roelants, O. Filhol, C. Cochet, E. Blanc, C. Ferlay-Segura, D. Borchelli, J. M. Ferrero, B. Escudier, S. Négrier, G. Pages, I. Vilgrain, The tyrosine-kinase inhibitor sunitinib targets vascular endothelial (VE)-cadherin: A marker of response to antitumoural treatment in metastatic renal cell carcinoma. *British Journal of Cancer* **118**, 1179-88 (2018). doi.org/10.1038/s41416-018-0054-5 [ncbi.nlm.nih.gov/pmc/PMC5943344](https://www.ncbi.nlm.nih.gov/pmc/PMC5943344)
662. J. Sun, Q. Sun, M. F. Brown, C. Dudgeon, J. Chandler, X. Xu, Y. Shu, L. Zhang, J. Yu, The multi-targeted kinase inhibitor sunitinib induces apoptosis in colon cancer cells via PUMA. *PLoS One* **7**, e43158 (2012). doi.org/10.1371/journal.pone.0043158 [ncbi.nlm.nih.gov/pmc/PMC3422222](https://www.ncbi.nlm.nih.gov/pmc/PMC3422222)
663. W. Scholtz, P. Mabeta, Sunitinib malate inhibits hemangioma cell growth and migration by suppressing focal adhesion kinase signaling. *Journal of Applied Biomedicine* **18**, 143-51 (2020). doi.org/10.32725/jab.2020.019
664. W. Zhai, S. Li, J. Zhang, Y. Chen, J. Ma, W. Kong, D. Gong, J. Zheng, W. Xue, Y. Xu, Sunitinib-suppressed miR-452-5p facilitates renal cancer cell invasion and metastasis through modulating SMAD4/SMAD7 signals. *Molecular Cancer* **17**, 157 (2018). doi.org/10.1186/s12943-018-0906-x [ncbi.nlm.nih.gov/pmc/PMC6231268](https://www.ncbi.nlm.nih.gov/pmc/PMC6231268)
665. A. Wongkajornsilp, V. Wamanuttajinda, K. Kasetsinsombat, S. Duangsa-ard, K. Sa-ngiamsuntorn, K. M. Suradej Hongeng, Sunitinib indirectly enhanced anti-tumor cytotoxicity of cytokine-induced killer cells and CD3+CD56+ subset through the co-culturing dendritic cells. *PLoS One* **8**, (2013). doi.org/10.1371/journal.pone.0078980 [ncbi.nlm.nih.gov/pmc/PMC3827292](https://www.ncbi.nlm.nih.gov/pmc/PMC3827292)
666. J. S. Ko, A. H. Zea, B. I. Rini, J. L. Ireland, P. Elson, P. Cohen, A. Golshayan, P. A. Rayman, L. Wood, J. Garcia, R. Dreicer, R. Bukowski, J. H. Finke, Sunitinib mediates reversal of myeloid-derived suppressor cell accumulation in renal cell carcinoma patients. *Clinical Cancer Research* **15**, 2148-57 (2009). doi.org/10.1158/1078-0432.Ccr-08-1332
667. J. Ozao-Choy, G. Ma, J. Kao, G. X. Wang, M. Meseck, M. Sung, M. Schwartz, C. M. Divino, P. Y. Pan, S. H. Chen, The novel role of tyrosine kinase inhibitor in the reversal of immune suppression and modulation of tumor microenvironment for immune-based cancer therapies. *Cancer Research* **69**, 2514-22 (2009). doi.org/10.1158/0008-5472.Can-08-4709 [ncbi.nlm.nih.gov/pmc/PMC4370269](https://www.ncbi.nlm.nih.gov/pmc/PMC4370269)
668. A. Guislain, J. Gadiot, A. Kaiser, E. S. Jordanova, A. Broeks, J. Sanders, H. van Boven, T. D. de Gruijl, J. B. Haanen, A. Bex, C. U. Blank, Sunitinib pretreatment improves tumor-infiltrating lymphocyte expansion by reduction in intratumoral content of myeloid-derived suppressor cells in human renal cell carcinoma. "Cancer Immunology, Immunotherapy" **64**, 1241-50 (2015). doi.org/10.1007/s00262-015-1735-z
669. M. Seandel, J. Shia, I. Linkov, R. G. Maki, C. R. Antonescu, J. Dupont, The activity of sunitinib against gastrointestinal stromal tumor seems to be distinct from its antiangiogenic effects. *Clinical Cancer Research* **12**, 6203-4 (2006). doi.org/10.1158/1078-0432.CCR-06-1292
670. H. Van Cruyjsen, A. A. M. Van Der Veldt, L. Vroeling, D. Oosterhoff, H. J. Broxterman, R. J. Scheper, G. Giaccone, J. B. A. G. Haanen, A. J. M. Van Den Eertwegh, E. Boven, K. Hockman, T. D. De Gruijl, Sunitinib-induced myeloid lineage redistribution in renal cell cancer patients: CD1c+ dendritic cell frequency predicts progression-free survival. *Clinical Cancer Research* **14**, 5884-92 (2008). doi.org/10.1158/1078-0432.CCR-08-0656
671. D. Beck, H. Niessner, K. S. M. Smalley, K. Flaherty, K. H. T. Paraiso et al., Vemurafenib potently induces endoplasmic reticulum stress-mediated apoptosis in BRAFV600E melanoma cells. *Science Signaling* **6**, ra7 (2013). doi.org/10.1126/scisignal.2003057 [ncbi.nlm.nih.gov/pmc/PMC3698985](https://www.ncbi.nlm.nih.gov/pmc/PMC3698985)
672. H. G. Cordeiro, A. V. De Sousa Faria, C. V. Ferreira-Halder, Vemurafenib downmodulates aggressiveness mediators of colorectal cancer (CRC): Low Molecular Weight Protein Tyrosine Phosphatase (LMWPTP), Protein Tyrosine Phosphatase 1B (PTP1B) and Transforming Growth Factor β (TGF β). *Biological Chemistry* **401**, 1063-9 (2020). doi.org/10.1515/hzs-2020-0124
673. E. Hajek, F. Krebs, R. Bent, K. Haas, A. Bast, I. Steinmetz, A. Tuettnerberg, S. Grabbe, M. Bros, BRAF inhibitors stimulate inflammasome activation and interleukin 1 beta production in dendritic cells. *Oncotarget* **9**, 28294-308 (2018). doi.org/10.18632/oncotarget.25511 [ncbi.nlm.nih.gov/pmc/PMC6033361](https://www.ncbi.nlm.nih.gov/pmc/PMC6033361)
674. W. Miao, Y. Wang, Quantitative Interrogation of the Human Kinome Perturbed by Two BRAF Inhibitors. *Journal of Proteome*

Research **18**, 2624-31 (2019).

doi.org/10.1021/acs.jproteome.9b00134

ncbi.nlm.nih.gov/pmc/PMC6939617

675. T. Pilli, S. Cantara, C. Marzocchi, F. Pacini, B. S. Prabhakar, M. G. Castagna, Vemurafenib may overcome TNF-related apoptosis-inducing ligand (TRAIL) resistance in anaplastic thyroid cancer cells. *Endocrine* **67**, 117-23 (2020). doi.org/10.1007/s12020-019-02028-2

676. B. Schilling, A. Sucker, K. Griewank, F. Zhao, B. Weide, A. Görgens, B. Giebel, D. Schadendorf, A. Paschen, Vemurafenib reverses immunosuppression by myeloid derived suppressor cells. *International Journal of Cancer* **133**, 1653-63 (2013). doi.org/10.1002/ijc.28168

677. D. J. Wozniak, B. Hutchinson, M. B. Gilic, W. Bie, V. Gaponenko, A. L. Tyner, Vemurafenib Inhibits Active PTK6 in PTEN-null Prostate Tumor Cells. *Molecular Cancer Therapeutics* **18**, 937-46 (2019). doi.org/10.1158/1535-7163.MCT-18-0862
ncbi.nlm.nih.gov/pmc/PMC6693941

678. L. Yu, L. X. Gao, X. Q. Ma, F. X. Hu, C. M. Li, Z. Lu, Involvement of superoxide and nitric oxide in BRAFV600E inhibitor PLX4032-induced growth inhibition of melanoma cells. *Integrative Biology* **6**, 1211-7 (2014). doi.org/10.1039/c4ib00170b

679. X. Yang, N. Zhan, Y. Jin, H. Ling, C. Xiao, Z. Xie, H. Zhong, X. Yu, R. Tang, J. Ma, J. Guan, G. Yin, G. Wu, L. Lu, J. Wang, Tofacitinib restores the balance of $\gamma\delta$ Treg/ $\gamma\delta$ T17 cells in rheumatoid arthritis by inhibiting the NLRP3 inflammasome. *Theranostics* **11**, 1446-57 (2021). doi.org/10.7150/thno.47860
ncbi.nlm.nih.gov/pmc/PMC7738886

680. X. Yang, M. Wan, Z. Cheng, Z. Wang, Q. Wu, Tofacitinib inhibits ox-LDL-induced adhesion of THP-1 monocytes to endothelial cells. "Artificial Cells, Nanomedicine, and Biotechnology" **47**, 2775-82 (2019). doi.org/10.1080/21691401.2019.1573740