

Supporting information

Electrogeneration of a free-standing cytochrome *c* – silica matrix at a soft electrified interface

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Experimental set-ups for electrochemistry and spectroelectrochemistry measurements

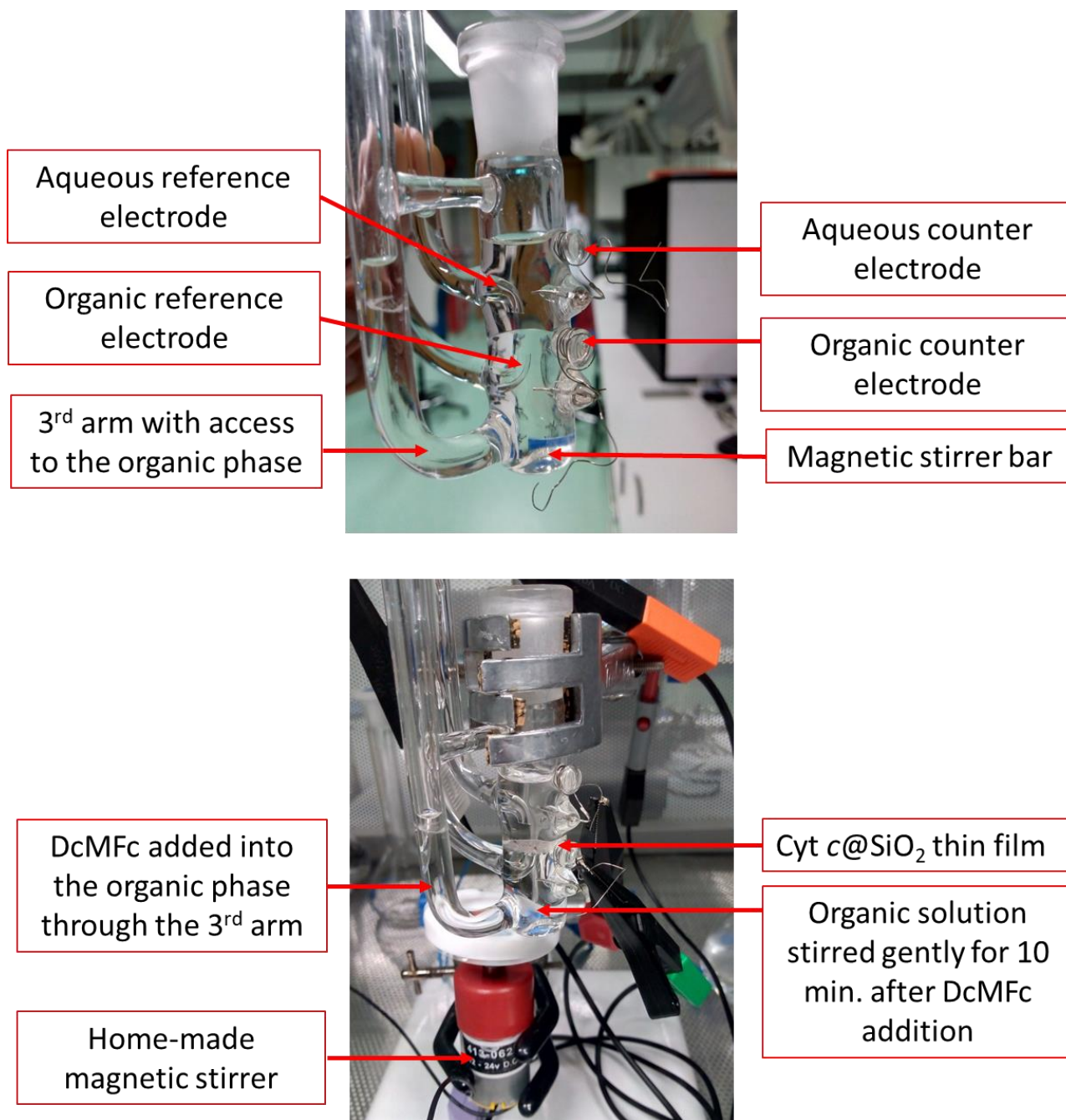


Figure S1. Homemade 4-electrode electrochemical cell with a third arm facilitating the addition of hydrophobic electron donors into the organic phase.

UV-vis spectroelectrochemistry setup

The spectrometer used was a USB 2000 Fiber Optic Spectrometer (Ocean Optics). The light source was a DH-2000-BAL deuterium–halogen (Ocean Optics) was guided through the optical fibre of 600 μm of diameter (Ocean Optics, USA). The light beam was collimated using optical lenses (Thor-lab, focal length: 2 cm) before and after the transmission of the beam through the electrochemical cell. The light beam passed through the electrochemical cell slightly above the water|TFT interface, i.e., through the aqueous phase. The interfacial Galvani potential difference ($\Delta_0^W \phi$) was controlled using an Autolab PGSTAT204 potentiostat (Metrohm, Switzerland). An image of this setup is shown in Figure S2.

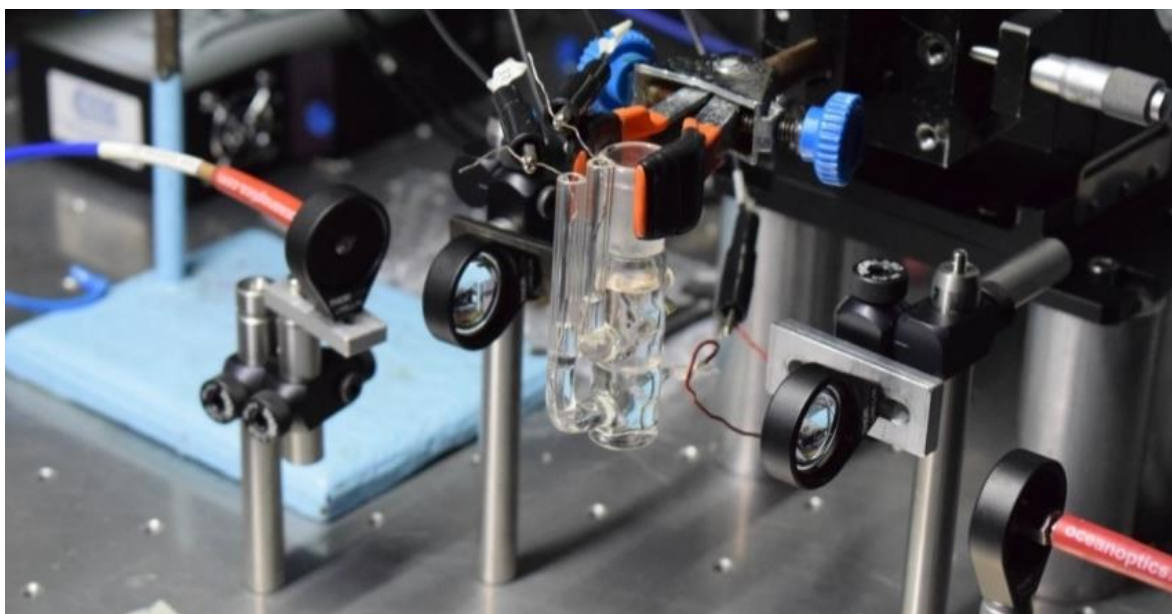


Figure S2. Optical setup for in situ parallel beam UV-vis absorbance measurements using a four-electrode electrochemical cell.

Control cyclic voltammetry of Cyt *c* in the absence of silica hydrogel

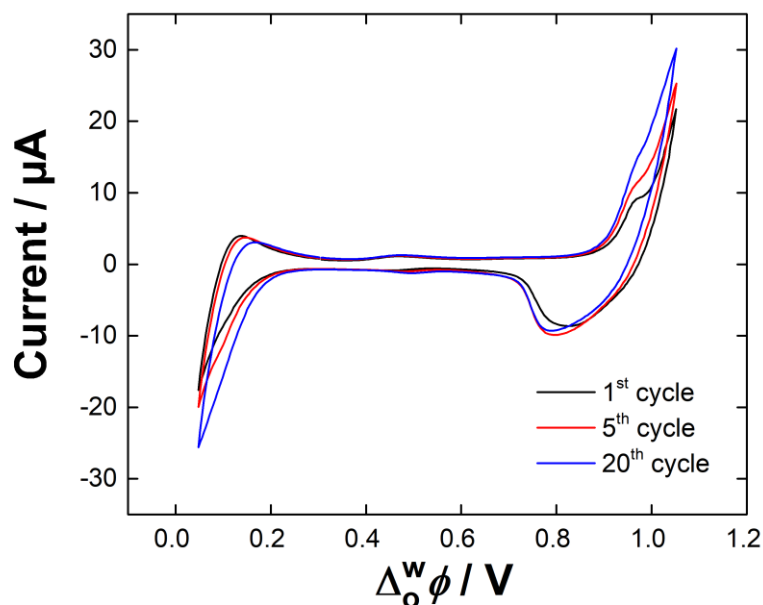


Figure S3. CV of 10 μM of Cyt *c* (pH 9) at a liquid-liquid interface in the absence of TEOS in the aqueous phase or CTA⁺ in the organic phase. $\nu = 20 \text{ mV s}^{-1}$.

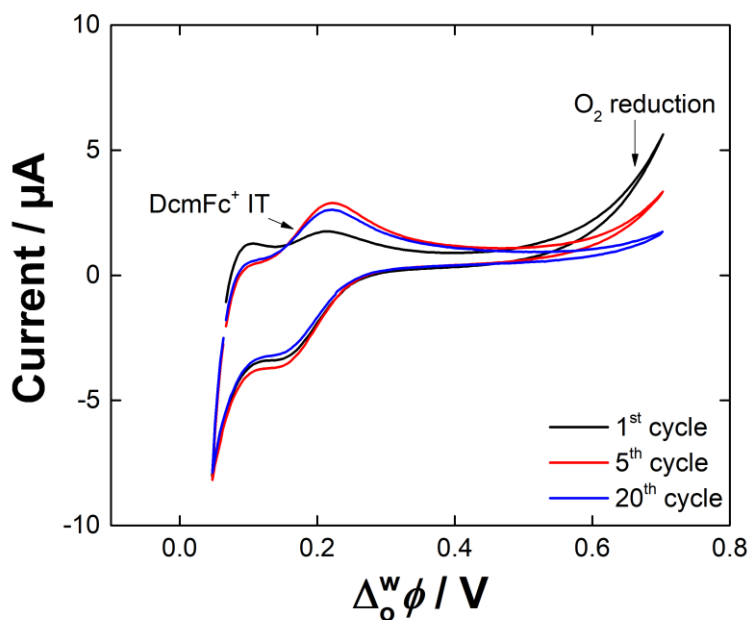


Figure S4. CV of 10 μM of Cyt *c* (aqueous phase) || 500 μM DcmFc (organic phase). $\nu = 20 \text{ mV s}^{-1}$.

Electrochemical stabilisation of Cyt *c*@SiO₂ hydrogel after aqueous phase exchange

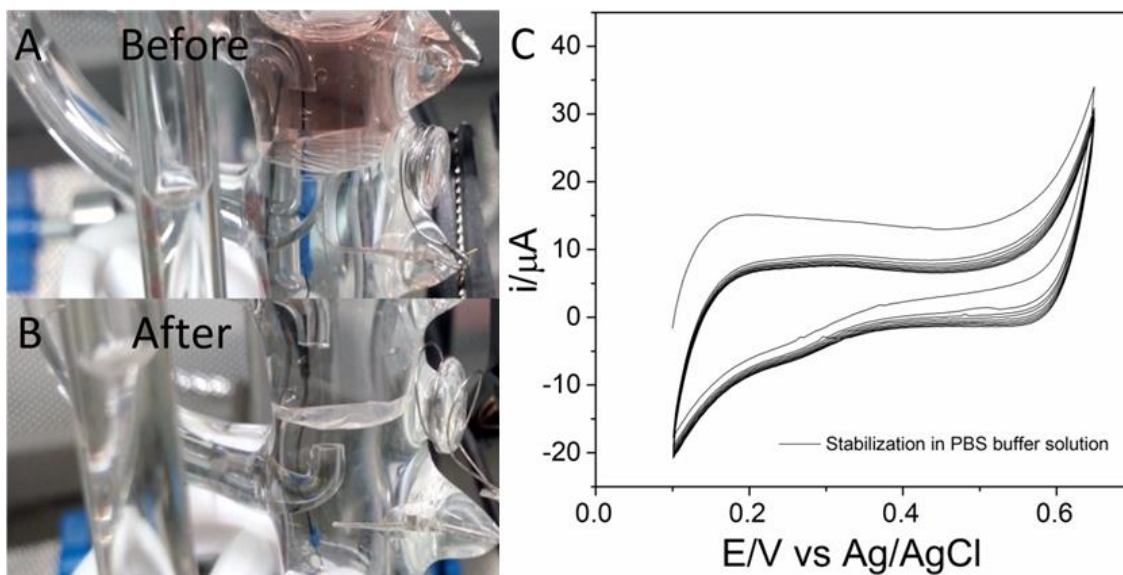


Figure S5. Image of the 4-electrode cell after (A) Cyt *c*@ SiO₂ electrogeneration and (B) after replacing the aqueous phase with 10 mM PBS solution. (C) Electrochemical stabilisation of Cyt *c*@SiO₂ by repetitive voltammetric scans at liquid-liquid interface (Electrochemical cell 3).