

**A study on the tribe Poecilonotini, with a revision
of the subtribe Nesotrinchina subtrib. nov.
and the description of a new genus and species
from Papua New Guinea
(Coleoptera: Buprestidae: Chrysochroinae)**

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Abstract. The preceding definitions of the tribe Poecilonotini Jakobson, 1913 are revised and commented and a new concept of the tribe Poecilonotini *sensu novo* is suggested. The subtribe Nesotrinchina Bílý, Kubáň & Volkovitsh, subtrib. nov. is described and revised including a study of selected taxa of the subtribe Poecilonotina. *Ulaikoilia jelineki* Bílý & Kubáň, gen. nov., sp. nov. (subtribe Nesotrinchina subtrib. nov.) is described from Papua New Guinea. The genus is diagnosed and compared with related genera and important characters at the genus and species level are illustrated. The following new synonymies are proposed: *Lamprodila (Erialata) auripilis* (Obenberger, 1922) = *L. (E.) pubescens* (Fisher, 1930), syn. nov., *Melobasina apicalis* Kerremans, 1900 = *M. ignita* (Théry, 1937), syn. nov., *Nesotrinchus australicus* (Kerremans, 1903) = *N. simonisi* Obenberger, 1924, syn. nov. and *N. caeruleipennis* (Fairmaire, 1877) = *N. orientalis* Bellamy, 1987, syn. nov. *Haplotrinchus marginifossus* Thomson, 1878, stat. restit. (Chrysochroinae Laporte de Castelnau, 1835, Dicerchini Gistel, 1848, Haplotrinchina Holyňski, 1993) is removed from synonymy with *H. pyrochlorus* (Fairmaire, 1877) and restituted as a valid species. *Nesotrinchus* Obenberger, 1924, stat. restit. is removed from synonymy with *Melobasina* Kerremans, 1900. A new substitute name is suggested: *Nesotrinchus thomsoni* Bílý & Kubáň, nom. nov. for *N. caeruleipennis* Thomson, 1878 (primary and secondary homonym). Lectotypes are designated for *Dicercomorpha caeruleipennis* Fairmaire, 1877, *D. caeruleipennis* Thomson,

1878, *Haplotrinchus australicus* Kerremans, 1903, *Melobasina apicalis* Kerremans, 1900, *Nesotrinchus simondsi* Obenberger, 1924 and *Poecilonota suturalis* Deyrolle, 1864. The genus *Pseudhyperantha* Saunders, 1869 is transferred from Stigmoderini Lacordaire, 1857 back to the tribe Buprestini Leach, 1815. Finally, the following species are transferred to other genera: *Melobasina suturalis* (Deyrolle, 1864), comb. nov. from *Lamprodila* Motschulsky, 1860; *Nesotrinchus wallisii* (Montrouzier, 1855), comb. nov. from *Haplotrinchus* Kerremans, 1903; and *N. australicus*, *N. caeruleipennis* and *N. thomsoni* Bílý & Kubáň, nom. nov., all comb. nov. from *Melobasina*.

Key words. Coleoptera, Buprestidae, Buprestini, Dicerini, Poecilonotini, Nesotrinchina subtrib. nov., taxonomy, replacement name, synonymy, new combination, lectotype designation, Oriental Region, Australian Region, Pacific Islands

Introduction

We have recently received a splendid specimen of an undescribed species of the family Buprestidae Leach, 1815. This new species belongs to an undescribed genus related to genera of the tribe Poecilonotini Jakobson, 1913 (Chrysochroinae Laporte de Castelnau, 1835). It is described in the present paper as *Ulaikoilia* Bílý & Kubáň, gen. nov. To describe and diagnose the new genus, the whole taxonomic structure of Poecilonotini had to be revised, since the tribe has been treated by various authors in rather different ways. As a result of this revision, a new concept of the tribe Poecilonotini Jakobson, 1913 *sensu novo* is suggested.

Material and methods

Wings, male and female genitalia of studied specimens were extracted and mounted on a separate label. A SZP 11 Zoom stereoscopic microscope was used for illustrations and Olympus SZX 12 microscope with fixed camera for colour images (photos by V. Kubáň unless otherwise mentioned). The nomenclature of wing venation follows FEDORENKO (2009). To study the characters of wing venation and abdominal tergite arrangement, additional species from different genera of the Poecilonotini, Dicerini Gistel, 1848, Stigmoderini Lacordaire, 1857, and Buprestini were investigated (see the list below).

Photos of the holotype of '*Nesotrinchus solomonensis* Théry, 1937' from the Database of insect primary types in the collection of the Museum of Comparative Zoology at Harvard University [<http://insects.oeb.harvard.edu/MCZ/FMPro?-db=Species.fm&-lay=web&-format=search.htm&-view>] were also used in the analyses.

We took the data on a distribution in the catalogue of each species directly from the original papers (in 'quotation marks' if cited verbatim), since many data in earlier catalogues are erroneous.

Data from locality labels of the types in the chapter 'Taxonomy' are cited 'verbatim' with our comments in square brackets, slash brackets '/' separate data from different labels, [h]

means that the preceding data within quotation marks are handwritten and [p] means that the preceding data within quotation marks are printed. All lectotype designations made in this paper are found in the chapter 'Revision of the subtribe Nesotrinchina subtrib. nov.'

The following codens are used in the text:

BMNH	Natural History Museum, London, United Kingdom;
BPBM	Bernice P. Bishop Museum, Honolulu, Hawaii, U.S.A.;
CLBC	Charles L. Bellamy research collection, Sacramento, California, U.S.A.;
DEIC	Deutsches Entomologisches Institut, Müncheberg, Germany;
FFDC	Forestry Department Collection, Suva, Fiji;
IRSN	Institut Royal des Sciences Naturelles de Belgique, Bruxelles, Belgium;
MCZC	Museum of Comparative Zoology, Harvard University, Cambridge, U.S.A.;
MNCA	Manfred Niehuis collection, Albersweiler, Germany;
MNHN	Muséum national d'Histoire Naturelle, Paris, France;
NHMB	Naturhistorisches Museum, Basel, Switzerland;
NMPC	National Museum, Prague, Czech Republic;
NZAC	New Zealand Arthropod Collection, Landcare Research, Auckland, New Zealand;
USNM	National Museum of Natural History, Washington, D.C., U.S.A.;
USPC	Insect Collection, University of the South Pacific, Suva, Fiji;
VKCP	Vítězslav Kubáň collection, deposited in National Museum, Prague, Czech Republic;
ZIN	Zoological Institute, Russian Academy of Sciences, St. Petersburg, Russia.

The following taxa were studied for the differential diagnoses:

Chrysochroinae: Poecilonotini: Poecilonotina

- Lamprodila (Ovalisia) auricollis* (Deyrolle, 1864): INDONESIA: MALUKU: Halmahera Island (1 spec., NMPC) (Figs. 98–102, 124).
- L. (O.) plasoni* (Théry, 1934): Northeastern PAPUA NEW GUINEA: MOROBE PROV.: Finschhafen dist.: Wareo, Rev. R. Wagner leg. (1 ♂, NMPC, ex coll. S. Bílý); Finschhafen, January 1930 (1 ♀, VKCP).
- L. (O.) purpuricollis* (Hoscheck, 1931): Southeastern PAPUA NEW GUINEA: GULF OF PAPUA: Ivimka Res. Station, Lakekamu Basin, 120 m, 7°44'S 146°30'E, 21.iii.2000, TASears MT (1 spec., NMPC).
- L. (O.) maculipennis* (Bílý, 1997): INDONESIA: PAPUA [formerly Irian Jaya]: Paniai dist., Nabire, Pemukiman, ca. 200 m, 17.viii.1991, A. Riedel leg. (holotype ♀, NMPC).
- Poecilonota variolosa variolosa* (Paykull, 1799): CZECH REPUBLIC: MORAVIA: Znojmo, 'Kráví hora' Hill, 16.vi.1994, S. Bílý leg. (1 ♂, NMPC).

Chrysochroinae: Dicercini: Dicercina

- Dicerca aenea aenea* (Linnaeus, 1761): BULGARIA: Melnik env., 26.v.–5.vi.1985, J. Macek leg. (1 ♀, NMPC).

Chrysochroinae: Dicercini: Haplotrinchina

- Cardiaspis mouhoutii* Saunders, 1866: Northern INDIA: 'Inde' (1 ♀, VKCP); 'British Bootang L. Durel 1898' (1 ♀, NMPC); 'Himalaya' (1 ♀, NMPC). WEST BENGAL: Darjeeling (1 ♀, NMPC); Pedong (1 ♂, NMPC). LAOS: VIANG CHAN: Ban Houay Say, 1965, A. Baudon leg. (1 ♀, NMPC). MALAYSIA: PERAK: ii.1975 (1 ♂, NMPC); January 1994 (1 ♂ 3 ♀♀, VKCP). Northeastern THAILAND: LOEI: Phu Kradung Nat. Park, 16°52'N 101°49'E, 1000 m, 16.–18.v.1999, M. Říha leg. (1 ♀, VKCP).
- C. pisciformis* Théry, 1904: Southern INDIA: TAMIL NADU: Nilgiris, Kotagiri (15 km SE), Kunchappanai, 900 m, 11°22'N 76°56'E, 7.–22.v.2000, J. Rolčík & D. Hauck leg. (1 ♂ 10 ♀♀, NMPC) (Figs. 7–13, 125); same label data but: P. Pacholátko leg. (1 ♂ 10 ♀♀, VKCP); Sri Lanka, Kandy (2 ♀♀, VKCP).
- Haplotrinchus aurocupreus* (Kerremans, 1900): PAPUA NEW GUINEA: LOUISIADIE ISLANDS: Rossel Island, i.–iii.1898, A. S. Meek leg. (1 syntype ♀, BMNH).
- H. edai* Ohmoto, 2002: PHILIPPINES: ROMBLON: 1989 (1 ♂, NMPC); 16.i.1991 (1 ♂, NMPC).
- H. embrikiellus* Obenberger, 1936: Northeastern PAPUA NEW GUINEA: MOROBE PROV.: 'Varreo' [Finschhafen dist., Wareo] (1 syntype ♀, NMPC). INDONESIA: MALUKU: Ceram Island, Wahi (12 km SE), Solea, 17.i.–6.ii.1997, from *Barringtonia* sp., S. Bílý leg. (8 larvae, NMPC); same label data but: ex larvae January 1998 (1 ♂, NMPC); Ambon Island, 1998, native collector leg. (1 ♂, VKCP).

- H. inaequalis* (Deyrolle, 1864): INDONESIA: MALUKU: Ceram Island: Wahai (6 km E), Ain Besar, 5.xi.1998, S. Bílý leg. (1 ♀, VKCP); Wahai (12 km SE), Solea, 17.i.–6.ii.1997, S. Bílý leg. (1 ♀, NMPC); same label data but: 16.x.–4.xi.1998, S. Bílý leg. (6 ♀♀ 4 larvae from *Barringtonia* sp., NMPC); same label data but: S. Bílý & J. Horák leg., ex larvae 1999–2000, reared from *Barringtonia* sp. (5 ♂♂ 3 ♀♀, NMPC, VKCP); Ambon Island: iv.2003, native collector leg. (1 ♂, NMPC); ‘Amboina’ (1 ♀, NMPC).
- H. manni* Théry, 1937: FIJI: Kandavu Island: Vanua Ava, W. M. Mann (1 paratype ♀, BMNH) (Figs. 29–33).
- H. philippinensis* Obenberger, 1928: PHILIPPINES: LUZON: Mt. Bulusan, 30.ix.1917, G. Böttcher leg. (1 syntype, NMPC) (Figs. 14–18, 130). MINDANAO: Surigao, viii.–ix.1916, G. Böttcher leg. (4 syntypes, NMPC); Northern Mindanao (1 ♀, NMPC). MINDORO: 17.ix.1992 (2 ♀♀, NMPC).
- H. pyrochlorus* (Fairmaire, 1877): FIJI: Kandavu Island (1 syntype ♀, BMNH); ‘Fidji’ (1 ♀, BMNH) (Figs. 24–28); [no locality, probably Fiji] (1 ♀, NMPC) (Fig. 126).
- H. marginifossus* (Thomson, 1878), **stat. restit.**: VANUATU: ‘Nouvelles-Hébrides’ (2 syntypes, MNHN) (Figs. 19–23). ANATOM ISLAND: ‘Aneiteum’ (4 spec., BMNH; 1 spec., MNHN, 1 spec., NMPC).
- Note.** Having studied the types and other specimens of both species, we decided to remove *Haplotrinchus marginifossus* from the synonymy with *H. pyrochlorus*. *Haplotrinchus marginifossus* (Vanuatu Islands) is somewhat more slender, distinctly bicolorous (head and pronotum bright coppery, elytra blue-violet) and with finer sculpture of the pronotal disc and the anterior third of the elytra (Fig. 19); *H. pyrochlorus* (Fiji Islands) is somewhat stouter, completely black with blue-green lustre and with rougher pronotal and elytral sculpture (Fig. 24). Both species also differ by a slightly different shape of the anal ventrite (Figs. 23, 28).
- H. splendens* Waterhouse, 1913: FIJI: September (1 syntype, BMNH).

Buprestinae: Buprestini

- Pseudhyperantha bloetei* Théry, 1935: INDONESIA: SUMATRA: Mt. Dempo, vi.2007, local collector (1 ♂ 1 ♀, NMPC) (Fig. 2).
- P. jucunda* Saunders, 1869: BORNEO: MALAYSIA: SABAH: Mt. Trus Mardi, March 2005, native collector leg. (1 spec., NMPC) (Figs. 1–6); Ranau, 05°58'N 116°41'E, 18.v.2004, native collector (1 ♂, ZIN, N. Woodley det.) (Fig. 120).
- P. trifasciata* Tôyama, 1989: PHILIPPINES: LEYTE (1 spec., NMPC).
- Buprestis (Buprestis) octoguttata octoguttata* Linnaeus, 1758: CZECH REPUBLIC: BOHEMIA: Plzeň – Sulkov, viii.1956, J. Brožik leg. (1 ♀, NMPC).

Buprestinae: Stigmoderini

- Stigmodera macularia* (Donovan, 1805): AUSTRALIA: VICTORIA: Melbourne env., B. M. Vosáhlo leg. (1 spec., NMPC).

Tribe level taxonomy

Tribe Poecilonotini Jakobson, 1913 *sensu novo*

- Poecilonotina Jakobson, 1913: 786 (as ‘fam. Buprestini: tribus Poecilonotina’; list; characters in key). Type genus: *Poecilonota* Eschscholtz, 1829.
- Poecilonotina: OBENBERGER (1930): 310 (catalogue; in synonymy of Buprestini: Dicercites).
- Poecilonotini (*sensu novo*): ALEXEEV & BEBKA (1970a): 6; ALEXEEV & BEBKA (1970b): 158 (both: comment; characters; in ‘Ancylocheirinae’ (now Buprestinae)).
- Poecilonotini (*sensu novo*): VOLKOVITSH (2001): 55 (catalogue of higher taxa; in Chalcophorinae (now Chrysochroinae)), 84, 96 (comment; characters), 108 (summary); BÍLÝ & VOLKOVITSH (2003): 108 (catalogue of higher taxa; in Chalcophorinae (in present Chrysochroinae)); BELLAMY (2003): 40 (catalogue of higher taxa; in Chrysochroinae); NELSON et al. (2008): 61 (catalogue; in Chrysochroinae); BELLAMY (2008): 594 (catalogue; in Chrysochroinae).

History of classification. The name ‘Poecilonotina’ was first used by JAKOBSON (1913: 773, 786), who used the name ‘Poecilonotina’ both for the category of tribe (*sensu lato*) and for the

category of subtribe (*sensu stricto*). He replaced by 'Poecilonotina' *sensu lato* and *sensu stricto* the formerly used names 'Buprestini' and 'Dicercites', respectively, of the previously widely used system by KERREMANS (1893); the reason for this change is not clear. JAKOBSON (1913: 786) included three subtribes into 'Poecilonotina' (*sensu lato*), 'Poecilonotina = Dicercina' (*sensu stricto*), 'Ancylochirina [nov.] = Buprestina' and 'Anthaxiina', and the following genera into the subtribe Poecilonotina (*sensu stricto*): *Cardiaspis* Saunders, 1866, *Dicerca* Eschscholtz, 1829, *Poecilonota* Eschscholtz, 1829 and *Lampra* Dejean, 1833 (now *Lamprodila* Motschulsky, 1860). JAKOBSON (1913) initially used only the form of antennal pores to define the Poecilonotini (*sensu stricto*); RICHTER (1952) mentioned difference between wing venation (position of R-M vein) in *Poecilonota* and *Lampra* (now *Lamprodila*) versus other Dicercini and also differences between the larvae of *Lampra* (now *Lamprodila*) and *Poecilonota* versus *Dicerca*.

The tribe Poecilonotini with the genera *Poecilonota* and *Lampra* (now *Lamprodila*) was independently created also by ALEXEEV & BEBKA (1970a,b), who separated the Poecilonotini in the subfamily Ancylocheirinae Jakobson, 1913 (now Buprestinae) on the basis of seven visible abdominal tergites, contrary to *Dicerca* and other genera in the subfamily Buprestinae *sensu* ALEXEEV & BEBKA (1970a,b) (now Chrysochroinae) with eight visible tergites.

COBOS (1986) newly defined two subfamilies: Chalcophorinae Lacordaire, 1857 (now Chrysochroinae) and Buprestinae and included the genera *Dicerca*, *Poecilonota* and *Ovalisia* Kerremans, 1900 (= *Lampra*, now subgenus of *Lamprodila*) into the tribe Dicercini of the subfamily Buprestinae (he did not mention the name Poecilonotini).

HOLYŃSKI (1993) defined the family Buprestidae in quite a new and revolutionary way. In his concept, the tribe Buprestini includes 18 subtribes (seven of them new and also, e.g., the subtribes Chrysochroina, Chalcophorina, Buprestina and Psilopterina). *Dicerca*, *Poecilonota* and *Ovalisia* (*sensu lato*, now *Lamprodila*) and further 11 genera were placed in the subtribe Psilopterina Lacordaire, 1857. One of his new subtribes was the subtribe Haplotrinchina Holyński, 1993, in which the genera *Cardiaspis* (erroneously as '*Bellamyola* nom. nov.', see VOLKOVITSH (1997)), *Haplotrinchus* Kerremans, 1903 and *Melobasina* Kerremans, 1900 were included. In a short comment he wrote: 'The Haplotrinchina str. n. make a very well-defined group, though superficially *Bellamyola* n.n. [= *Cardiaspis*...] looks quite dissimilar from *Melobasina* KERR. [including *Nesotrinchus* OBB., which, in my view, cannot be separated even as a subgenus, "*N.*" *coeruleipennis* (THS.) being much more closely related to *M. fossicollis* KERR. than to "*N.*" *australicus* (KERR.)]'. BELLAMY (1997: 76, resp. 2002: 62) interpreted this comment as a synonymisation of the genera *Melobasina* and *Nesotrinchus* Obenberger, 1924 but this interpretation is wrong.

From the comments by HOLYŃSKI (1993), we can judge that he based his study of the genus *Melobasina* on *M. fossicollis* (Kerremans, 1906) but without any specification of studied specimens. He probably saw specimens from BMNH, identified by B. Levey as '*fossicollis*' but in fact belonging to *M. apicalis* Kerremans, 1900 (see below under *M. apicalis*). Characters used for the subtribe Haplotrinchina in his key (couplets u(b), x(cc) and cc(z)) do not explicitly define either *Melobasina* or *Nesotrinchus*. The definition of the subtribe Haplotrinchina by HOLYŃSKI (1993) is quite insufficient and the reasons for the treatment of genera *Melobasina* and *Nesotrinchus* in Haplotrinchina is so vague that it is impossible to validate them.

BÍLÝ (1999) mentioned differences between the larvae of *Dicerca* and *Capnodis* Eschscholtz, 1829 and the larvae of *Lampra* (now *Lamprodila*) and *Poecilonota*. BÍLÝ & VOLKOVITSH (2002: Table 2) published a set of larval characters differentiating *Lampra* (now *Lamprodila*) and *Poecilonota* from the larvae of *Dicerca*, *Dicercomorpha* Deyrolle, 1864, *Haplotrinchus* and *Polybothris* Dupont, 1833 [sic!].

JENDEK (2001) found differences in the general structure of the abdomen between the Dicerini on one hand and *Lampra* (now *Lamprodila*) and *Poecilonota* and *Kisanthobia* Marseul, 1865 on the other.

VOLKOVITSH (2001) principally disagreed with the concept of HOLYŃSKI (1993) and presented results of a detailed study of the antennal structures in the Buprestidae (without the Agrilinae) based on taxa of generic rank. The arrangement and status of suprageneric taxa reflect similarities and presumed evolutionary trends in antennal structures (VOLKOVITSH 2001: 52–59); the list used several informal categories: ‘complex’, ‘lineage’, ‘branch’ and ‘generic group’. The term ‘generic group’ was used at three different levels: tribal, subtribal and lower than the subtribal level.

Unlike HOLYŃSKI (1993), VOLKOVITSH (2001) re-introduced again the subfamily Chalcophorinae (now Chrysochroinae) but in a rather different concept than did COBOS (1986). Contrary to COBOS (1986) (see above), he placed the genera *Dicerca*, *Poecilonota* and *Palmar* (now a subgenus of *Lamprodila*) into the Chalcophorinae (now Chrysochroinae) and divided this subfamily into two ‘lineages’, Chalcophorioid (now Chrysochroioid) and Psilopteroid (now Dicercioid), and placed the tribe Poecilonotini in the former lineage. The tribe Psilopterini (now Dicerini) was placed together with the tribe Sphenopterini Lacordaire, 1857 in the latter lineage and divided into four subtribes, including the subtribe Haplotrinchina with the genera *Cardiaspis* and *Haplotrinchus*. The definition of the tribes Poecilonotini and Dicerini (as Psilopterini) was based mainly on the antennal structures (U2 sensillae absent in Poecilonotini but present in Dicerini) but also other characters such as morphological differences between the larvae of both tribes (see also BÍLÝ & VOLKOVITSH 2002) or a serrate elytral margin in the Poecilonotini (VOLKOVITSH 2001: 69, 84, 96). Both VOLKOVITSH (2001) and BÍLÝ & VOLKOVITSH (2003) erroneously attributed the authorship of the tribe Poecilonotini to ALEXEEV & BEBKA (1970a,b).

VOLKOVITSH (2001) studied the antennal structures of the following taxa of the Poecilonotini and Haplotrinchina: *Palmar* subgen. *Palmar* Schaefer, 1949 (now in *Lamprodila*) (one species), *Palmar* subgen. *Scintillatrix* Obenberger, 1956 (now *Lamprodila*) (two species), *Poecilonota* (one species), *Nesotrinchus* (one species), *Cardiaspis* (one species) and *Haplotrinchus* (one species); *Pseudhyperantha* Saunders, 1869 and *Melobasina* were not studied. As a result of this comparison, VOLKOVITSH (2001: 55, 68, 96) confirmed *Nesotrinchus* as a valid genus of the tribe Poecilonotini and created for it an independent, informal ‘*Nesotrinchus* generic group’.

BELLAMY (2003, 2008) accepted the results of VOLKOVITSH (2001) but treated *Nesotrinchus* as a junior subjective synonym of the genus *Melobasina* (misinterpretation, see above). He also included the genus *Pseudhyperantha* (Figs. 1–6, 120) into the subtribe Haplotrinchina

(in Chrysochroinae: Dicerini), a traditional concept introduced by KERREMANS (1893). On the contrary he did not accept the opinion of TOYAMA (1989), who transferred the genus *Pseudhyperantha* into the tribe Buprestini (in Buprestinae) near to the genus *Buprestis* Linnaeus, 1758. Finally, HOLYŃSKI (2008) transferred *Pseudhyperantha* back to the Buprestinae but into the tribe Stigmoderini.

Placement of the genus *Pseudhyperantha*. Antennal structures of *Pseudhyperantha* correspond with those of the Buprestini (Fig. 120): antennomeres triangular, bilaterally flattened, apical pits large, irregular, situated somewhat ventrally. Lateral sensory fields large, formed by B4c sensillae, U2 sensillae missing (see VOLKOVITSH 2001: 71, Figs. 128–130). The antennal structure of the Stigmoderini were treated in detail by GARDNER (1989: 303, Figs. 36–41) and VOLKOVITSH (2001: 72, Figs. 144–149). The prosternal process is of buprestine type in *Pseudhyperantha* (Fig. 5), with preapical lobes missing (see also KOLIBÁČ 2000, Fig. 11); the prosternal processes of Stigmoderini was discussed in GARDNER (1989: 305, Figs. 61–62). Wing venation of *Pseudhyperantha* (see TOYAMA 1989: 194, Fig. 1) corresponds to *Buprestis* (see KOLIBÁČ 2000: 152, Fig. 138; FEDORENKO 2009: 260, Fig. 57) rather than a stigmoderine type (see GARDNER 1989: 311, Fig. 66): radial cell (rc) long, vein Rr distinct, Mr stretching far beyond intersection with 'r-m', 'wedge' cell (2a) closed; for the nomenclature of veins (Fig. 121) see VOLKOVITSH (2008, Fig. 54) and FEDORENKO (2009: 260, Figs. A103–A106).

Moreover, the ovipositor of *Pseudhyperantha* (Fig. 2) possesses all characters of Buprestini: it is simple and tubular with simple styli and without a dorsal valve (more complex structures and a dorsal valve are the synapomorphies of the Stigmoderini (see GARDNER 1989)). The placement of *Pseudhyperantha* into the tribe Stigmoderini by HOLYŃSKI (2008) thus cannot be maintained. We accept the opinion of TOYAMA (1989) who attributed this genus to the tribe Buprestini. The subtribal position and closest affinities of *Pseudhyperantha* within the Buprestini remain unclear. We suppose it should be separated as distinct monogeneric subtribe but this assumption requires further investigations.

The genera *Melobasina* and *Nesotrinchus*. We have studied seven specimens of *Melobasina* (four species) and 29 specimens of *Nesotrinchus* (three species). Species of *Nesotrinchus* are very similar to each other; they are undoubtedly congeneric and distributed only on the Pacific islands of Fiji, Tonga, Niue and Wallis. Species of *Melobasina* are also very similar to each other. They are distributed on the Solomon and Maluku Islands and differ from *Nesotrinchus* by the colouration and body shape (see Table 1 and Figs. 59–97). The frons of *Melobasina* species is regularly convex and much wider than that of *Nesotrinchus*, and the latter genus also possesses well-developed supraantennal carinae and deep frontal depression.

New concept of the tribe Poecilonotini. Having studied representatives of the genera mentioned above (see also the list in Material and methods) and further representatives of the genus *Lamprodila* (subgenera *Cinyrisia* Holyński, 2000, *Erialata* Zykov, 1998 and *Mabomisia* Holyński, 2000), we define newly the tribe Poecilonotini and the subtribe Poecilonotina and suggest a new concept of the tribe Poecilonotini (*sensu novo*) containing subtribes Poecilonotina and Nesotrinchina subtrib. nov.

Differential diagnosis of *Poecilonotini sensu novo*

Adults. Antennae with antennomeres triangular, apically rounded, apical pits large and regularly rounded, lateral fields and U2 sensillae missing (VOLKOVITSH 2001). Elytra with apical portion of elytral margin finely and densely serrate (VOLKOVITSH 2001). Wing venation with 'r-m' vein joining Rs and Mr near their proximal ends, 'anal cell' (1a) open (Figs. 121–124); in the Dicercini including the Haplotrinchina, 'r-m' vein located much distally starting from rc and 1a closed by transverse vein (Figs. 125–126). Abdomen with seven visible tergites in all studied representatives of the Poecilonotini (Figs. 127–129); eight in the Dicercini (ALEXEEV & BEBKA 1970a,b; JENDEK 2001), including the Haplotrinchina (Fig. 130).

Larva. Prementum with a simple field of microspinulae, fountain-like field of rough asperities around anterior portion of pronotal grooves absent (BÍLÝ 1999, BÍLÝ & VOLKOVITSH 2002).

Key to subtribes of *Poecilonotini sensu novo*

- 1(2) Posterior half of lateral pronotal margins forming sharp keel; hypomera flat, punctate, with fine and narrow groove along lateral pronotal keel (Figs. 35, 45, 50, 99); anal ventrite apically emarginate, more or less sharply bispinose (Figs. 48, 102), rarely rounded (except for *Lamprodila* subgen. *Mabomisia* as in Figs. 53), evenly convex (Figs. 38–39, 43, 102) (except for *Lamprodila* subgen. *Cinyrisia* as in Fig. 48); apical third of elytral margin very finely and densely serrate, elytral apices indistinctly bi- or trispinose (except for *Lamprodila* subgen. *Erialata* and *Mabomisia* as in Figs. 43, 53), sometimes only obliquely truncate or rounded (Figs. 44, 98).
 **Poecilonotina** Jakobson, 1913 sensu Volkovitsh (2001)
- 2(1) Posterior half of lateral pronotal margins with obtuse or callus-like keel, sometimes formed only by nearly isolated swellings; hypomera uneven with irregular elevations and with wide and deep groove along lateral pronotal keel (Figs. 55, 61, 67, 72–73, 83–84, 91–92, 104); anal ventrite very sharply bispinose in male (Figs. 81, 89, 97, 108, 112) or trispinose in female (Figs. 58, 64, 76, 87, 95) (rarely with reduced median spine as in Fig. 70), spines long, nearly needle-shaped; each lateral spine bearing elevated carina, sometimes reaching posterior third of anal ventrite (Figs. 58, 64, 70, 76, 81, 87, 89, 95, 97, 108, 112); apical third of elytral margins sparsely but sharply and roughly serrate, elytral apices always distinctly trispinose (Figs. 54, 59, 66, 71, 82, 90, 103).
 **Nesotrinchina subtrib. nov.**

Notes on selected taxa of the subtribe *Poecilonotina*

Subtribe *Poecilonotina* Jakobson, 1913 sensu VOLKOVITSH (2001)

(Figs. 34–53, 98–102, 124)

Poecilonotina Jakobson, 1913: 786 (in key; as 'subtribus *Poecilonotina* = *Dicercina*?; list; characters in key). Type genus: *Poecilonota* Eschscholtz, 1829.

'*Poecilonotina sensu* Volkovitsh': VOLKOVITSH (2001): 55 (catalogue of higher taxa; in *Poecilonotini*); BÍLÝ & VOLKOVITSH (2003): 108 (catalogue of higher taxa; in *Poecilonotini*); BELLAMY (2003): 40 (catalogue; in *Poecilonotini*); BELLAMY (2008): 594 (catalogue; in *Poecilonotini*).

Diagnosis. See the key above.

Genera included. *Poecilonota* Eschscholtz, 1829, *Lamprodila* Motschulsky, 1860 (with subgenera *Cynirisia* Holyński, 2000, *Erialata* Zykov, 1998, *Lamprodila* Motschulsky, 1860, *Mabomisia* Holyński, 2000, *Ovalisia* Kerremans, 1900, *Palmar* Schaefer, 1949, *Poecilisia* Holyński, 2000 and *Zykovisia* Holyński, 2000).

Remarks. Representatives of some subgenera of *Lamprodila* share some characters with *Nesotrinchina* subtrib. nov., e.g. the anal ventrite of female is bispinose with an obtuse medial lobe in the subgenus *Mabomisia* and bispinose or (in some specimens) bispinose with very poorly marked medial lobe in the subgenus *Erialata*, the lateral keels on the anal ventrite are elevated in the subgenus *Cynirisia* and the elytral apex is distinctly trispinose in the subgenera *Erialata* and *Mabomisia* (see Table 1).

***Lamprodila* Motschulsky, 1860** **Subgenus *Erialata* Zykov, 1998**

Erialata Zykov, 1998: 2 (characters), 4 (key). Type species: *Lampra auripilis* Obenberger, 1922, by original designation.

Erialata: HOLYŃSKI (2000): 4 (as subgenus of *Ovalisia*; new status; characters), 6 (key), Fig. 1 (distribution), Fig. 12 (cladogram); BELLAMY (2003): 40 (as subgenus of *Ovalisia*; catalogue); BELLAMY (2008): 601 (as subgenus of *Lamprodila*; catalogue; new combination).

Remarks. Females of this subgenus possess trispinose elytral apices (Figs. 34, 39–40, 43) and (in some specimens) poorly developed median spine on anal ventrite (Fig. 39, 43) similar to those in *Nesotrinchina* subtrib. nov.

***Lamprodila (Erialata) auripilis* (Obenberger, 1922)** (Figs. 34–43)

Lampra auripilis Obenberger, 1922: 81. Type locality: Borneo (without exact data).

Lampra auripilis: OBENBERGER (1930): 347 (catalogue).

Lampra pubescens Fisher, 1930: 32, **syn. nov.** Type locality: Malaysia, Perak, Batang Padang, Jor Camp, 1800 ft.

Erialata auripilis: ZYKOV (1998): 1 (comment; new combination), 2 (characters), Figs. 2–6.

Ovalisia (Erialata) auripilis: HOLYŃSKI (2000): 2, 4 (notes; new combination); BELLAMY (2003): 40 (catalogue), Fig. 178.

Ovalisia (Erialata) pubescens: HOLYŃSKI (2000): 4 (characters; new combination), Figs. 7, 10.

Lamprodila (Erialata) auripilis: BELLAMY (2008): 603 (catalogue; new combination).

Lamprodila (Erialata) pubescens: BELLAMY (2008): 618 (catalogue; new combination).

Type material examined. *Lamprodila (Erialata) auripilis*: SYNTYPE: 1 ♀ (NMPC), **BORNEO**: ‘Borneo [h] / Lampra auripilis m. Type [h] Det. D^f Obenberger [p] / Typus [p] [red label] / Mus. Nat. Pragae Inv. [p] 20137 [h] [orange label]’. Number of syntypes unknown. *Lamprodila (E.) pubescens*: HOLOTYPE (by monotypy): sex not examined (USNM), **PENINSULAR MALAYSIA**: **PERAK**: ‘Perak, F.M.S. [= Federated Malay States] Batang Padang Jor Camp [p]1800 [h] ft. [p] June 4th [h] 1923 H.M. Pendlebury. [p] [underside:] Ex coll. F. M. S. Museum. [p] / Type No [p] 57404 [h] USNM [p] [red label] / Holotype [red p] Lampra pubescens Fisher [h] [red framed label]’.

Additional material examined. **PENINSULAR MALAYSIA**: **PAHANG**: Benom Mts., Kampong Dong (15 km E), 03°53'N 102°01'E, 300–900 m, 24.iii.–15.iv.1998, D. Hauck leg. (1 ♀, VKCP). **PERAK**: Ipoh (25 km NE), Banjaran Titi Wangsa Mts., Korbu mt., 1200 m, 6.–12.v.2001, P. Čechovský leg. (1 ♀, VKCP).

Remarks. Having studied the types and other specimens of *Lamprodila (Erialata) pubescens* and *L. (E.) auripilis*, we failed to find any substantial differences and consider them conspecific, *L. (E.) pubescens* (Figs. 40–43) being a junior subjective synonym of *L. (E.) auripilis* (Figs. 34–39).

Distribution. Peninsular Malaysia (Pahang, Perak); Borneo (no further details available).

Subgenus *Cinyrisia* Holyński, 2000

Cinyrisia Holyński, 2000: 3 (as subgenus of *Ovalisia*; characters), 5 (key), Fig. 1 (distribution), Fig. 12 (in cladogram). Type species: *Poecilonota psilopterooides* Deyrolle, 1864, by original designation.

Cinyrisia: BELLAMY (2003): 40 (as subgenus of *Ovalisia*; catalogue); BELLAMY (2008): 601 (as subgenus of *Lamprodila*; catalogue).

Remarks. Representatives of the subgenus *Cinyrisia* possess an elevated carina on each lateral spine of the anal ventrite (Fig. 48) like in *Nesotrinchina* subtrib. nov.

Lamprodila (*Cinyrisia*) *psilopterooides* (Deyrolle, 1864) species-group

Psilopterooides-circle: HOLYŃSKI (2000): 3 (in *Ovalisia* subgen. *Cinyrisia*; characters).

psilopterooides-circle: BELLAMY (2008): 601 (in *Lamprodila* subgen. *Cinyrisia*; catalogue).

Lamprodila (*Cinyrisia*) *psilopterooides* (Deyrolle, 1864)

(Figs. 44–48)

Poecilonota psilopterooides Deyrolle, 1864: 58 (key), 59 (characters). Type locality: Borneo (without exact data).

Poecilonota psilopterooides: SAUNDERS (1867): 511 (list; new distribution); GEMMINGER & HAROLD (1869): 1376 (catalogue); SAUNDERS (1871): 38 (catalogue); KERREMANS (1892): 91 (catalogue).

Lampra psilopterooides: KERREMANS (1903): 135 (list of species; new combination); MOULTON (1911): 168 (list); MOULTON (1912): 84 (list; new locality in Sarawak); OBENBERGER (1930): 352 (catalogue).

Ovalisia (*Cinyrisia*) *psilopterooides*: HOLYŃSKI (2000): 3 (characters; new combination), Fig. 5; BELLAMY (2003): 40 (catalogue), Fig. 177.

Lamprodila (*Cinyrisia*) *psilopterooides*: BELLAMY (2008): 618 (catalogue; new combination).

Type material examined. SYNTYPE: sex not examined (MNHN), BORNEO: ‘Bornéo [h] / Psilopterooides HDeyr. [h] / Type [p] / Ex-Musaeo Mniszech [p] / Muséum Paris 1952 Coll. R. Oberthür [p]’. Number of syntypes unknown.

Additional material examined. PENINSULAR MALAYSIA: ‘Malacca [h] / Type [p] / Ex-Musaeo Mniszech [p] / Muséum Paris 1952 Coll. R. Oberthür [p]’ (1 spec., MNHN). PAHANG: Benom Mts., Kampong Dong (15 km E), 03°53’N 102°01’E, 300–900 m, 24.iii.–15.iv.1998, D. Hauck leg. (2 ♀♀, VKCP); same label data but 700 m, 1.iv.1998, L. Dembický & P. Pacholátko leg. (2 ♀♀, NMPC). PINANG: C. Lamb leg. (2 ♀♀, NMPC). BORNEO: INDONESIA: WEST KALIMANTAN: Pontianak, 1898 (2 ♀♀, NMPC).

Remarks. The carina on the lateral spines of the anal ventrite is sharp and very well developed (Fig. 48) in this species. The specimen from Malacca (MNHN) is not a syntype.

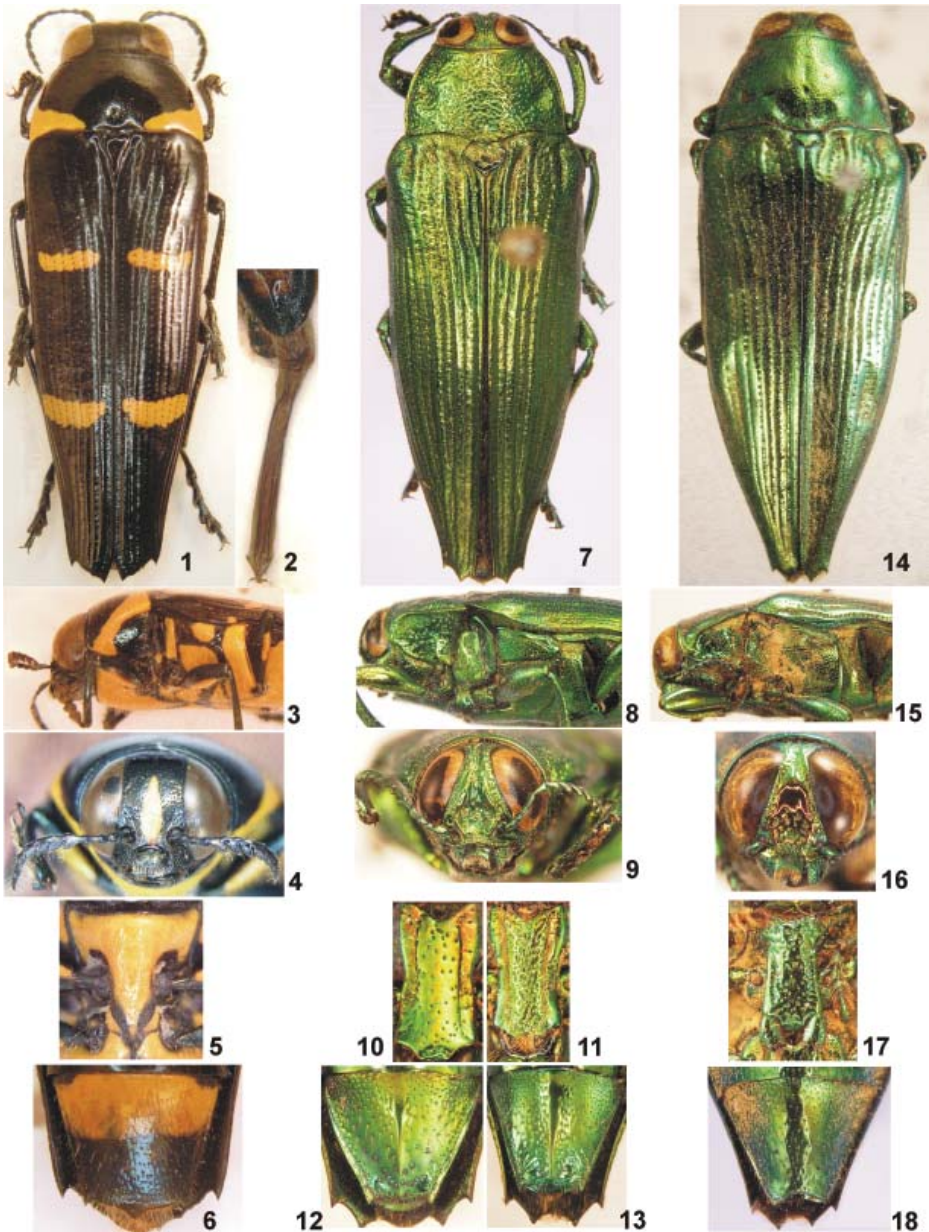
Distribution. Peninsular Malaysia (Pahang, Pinang). Borneo (no further details available); Malaysia (Sarawak); Indonesia (West Kalimantan).

Subgenus *Mabomisia* Holyński, 2000

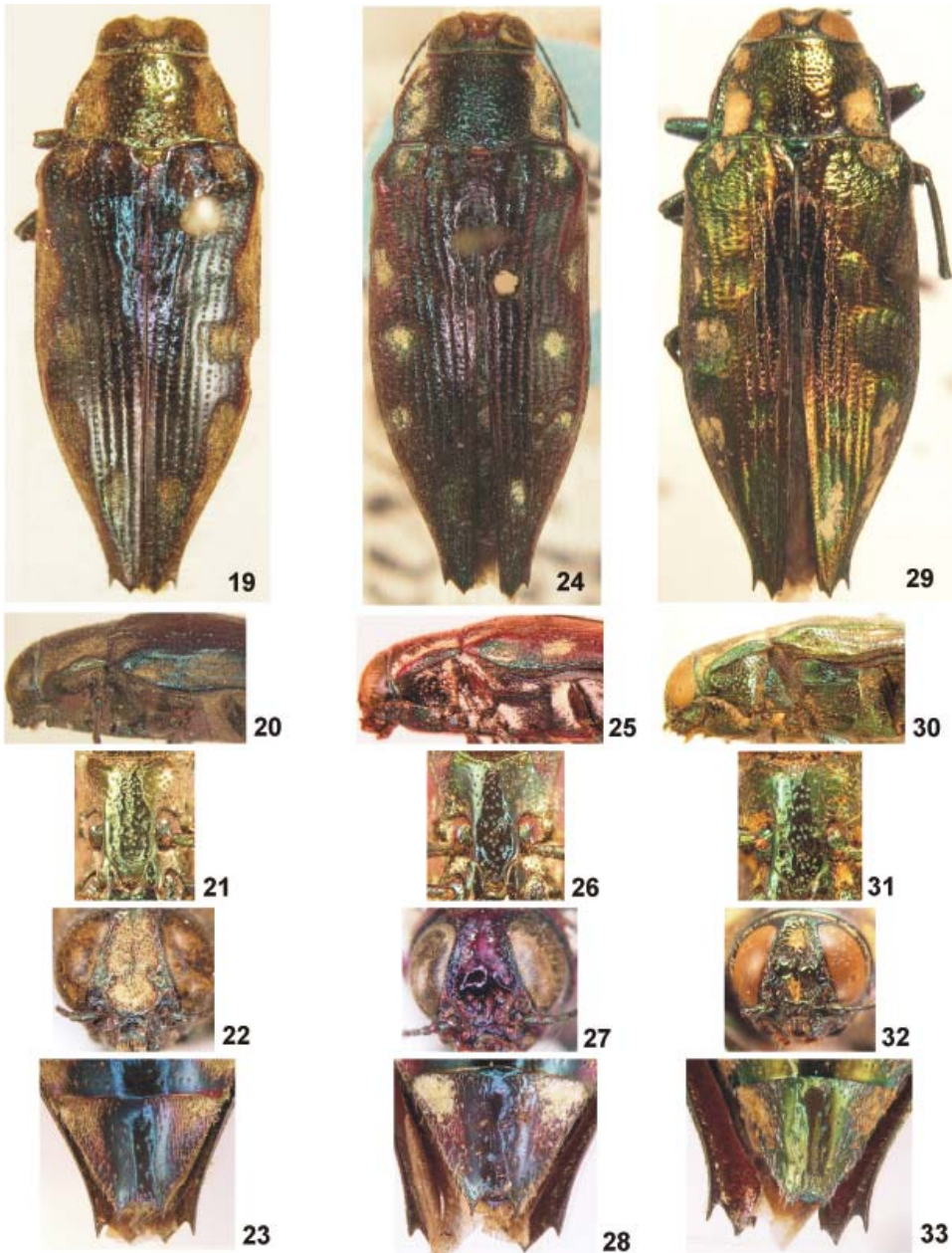
Mabomisia Holyński, 2000: 3 (as subgenus of *Ovalisia*; characters), 6 (key), Fig. 1 (distribution), Fig. 12 (in cladogram). Type species: *Chalcophora* (*Evides*) *sexspinosa* Thomson, 1857, by original designation.

Mabomisia: BELLAMY (2003): 40 (as subgenus of *Ovalisia*; catalogue); BELLAMY (2008): 601 (as subgenus of *Lamprodila*; catalogue).

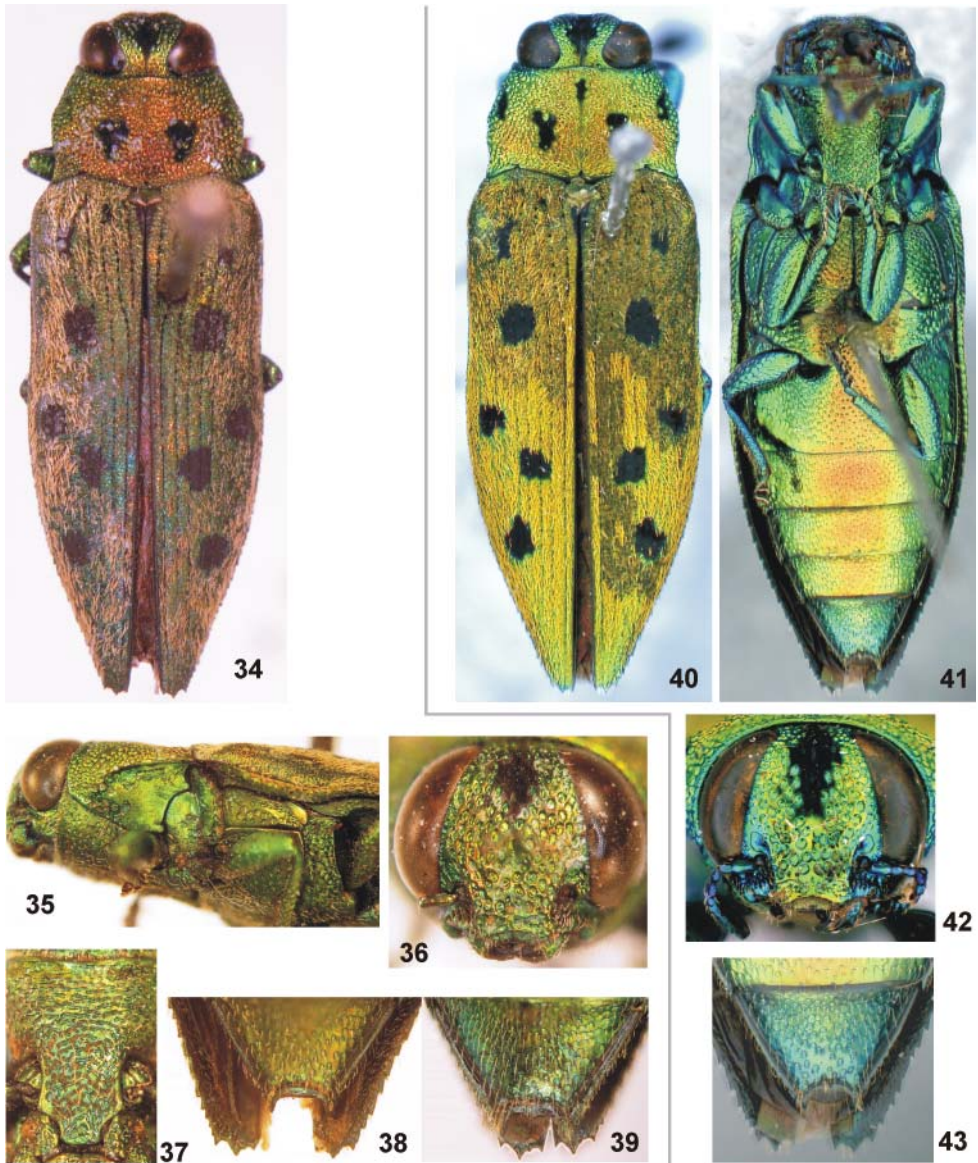
Remarks. Like in *Nesotrinchina* subtrib. nov., the representatives of this subgenus also possess distinctly trispinose elytral apices (Figs. 49, 53) and a well-developed median lobe of the anal ventrite in female (Figs. 53).



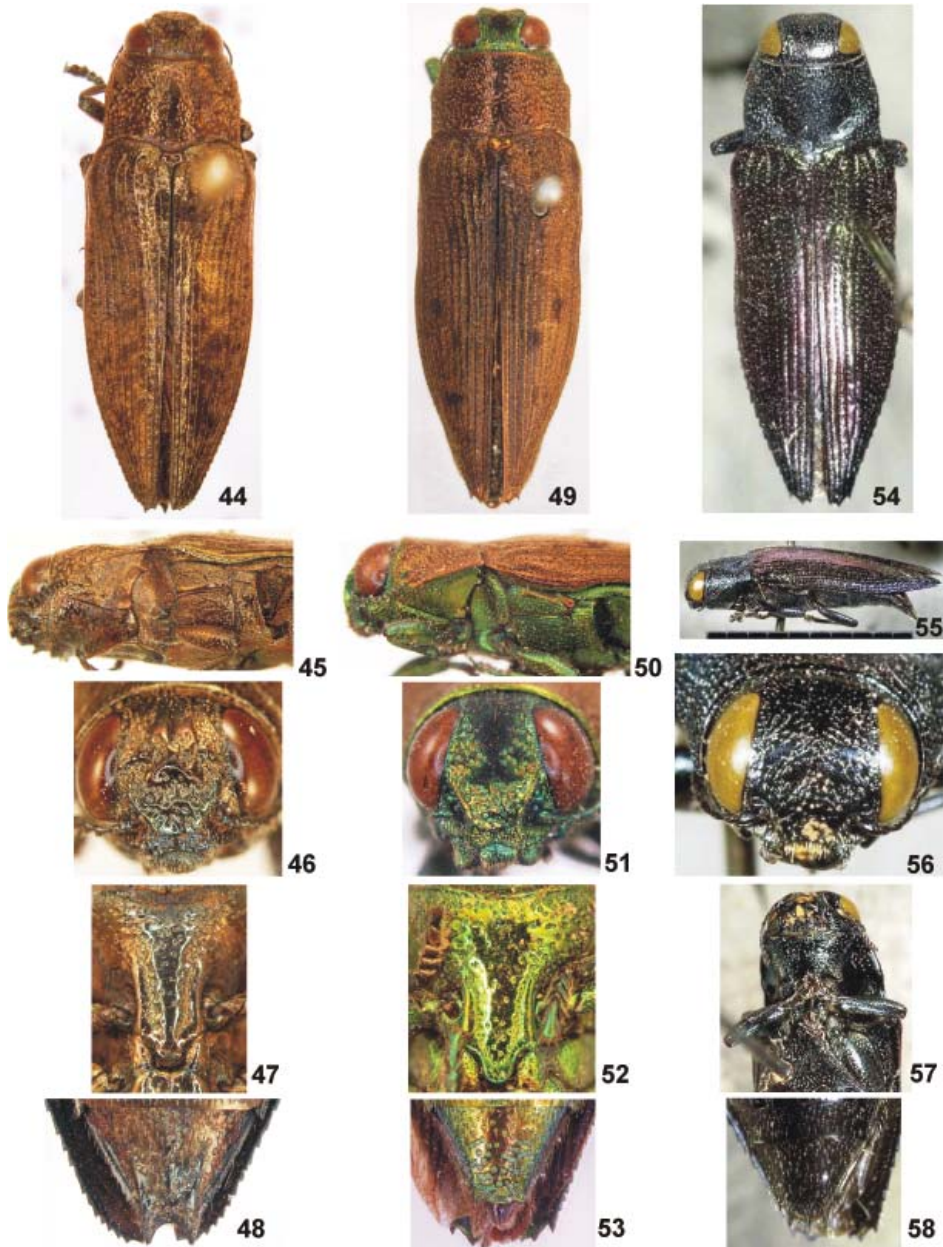
Figs. 1–18. 1, 3–6 – *Pseudhyperantha jucunda* Saunders, 1869, male, 17.0 mm, Borneo. 2 – *P. bloetei* Théry, 1935, female, Sumatra. 7–13 – *Cardiaspis pisciformis* Théry, 1904, India (7–9, 11, 13 – female, NMPC, 17.0 mm; 10, 12 – male, NMPC). 14–18 – *Haplotrinchus philippinensis* Obenberger, 1928, syntype, female, 15.5 mm, Philippines, Luzon. 1, 7, 14 – habitus, dorsal view; 2 – ovipositor, dorsal view; 3, 8, 15 – habitus, lateral view; 4, 9, 16 – head, frontal view; 5, 10–11, 17 – prosternal process; 6, 12–13, 18 – anal ventrite.



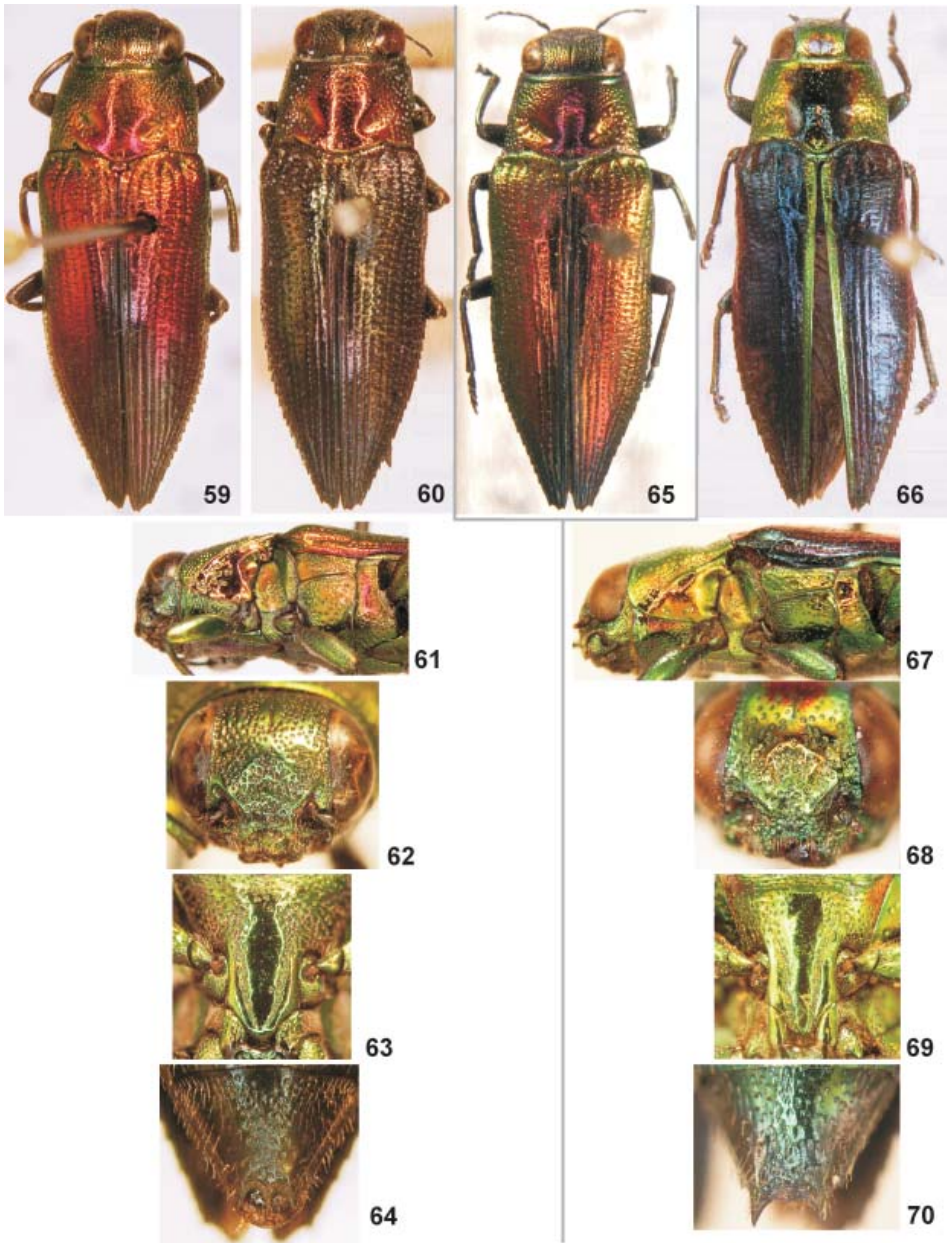
Figs. 19–33. 19–23 – *Haplotrinchus marginefossus* (Thomson, 1878), syntype, female, 14.5 mm, Vanuatu. 24–28 – *H. pyrochlorus* (Fairmaire, 1877), syntype, female, 15 mm, Fiji. 29–33 – *H. manni* Théry, 1937, paratype, female, 13.0 mm, Fiji. 19, 24, 29 – habitus, dorsal view; 20, 25, 30 – habitus, lateral view; 21, 26, 31 – prosternal process; 22, 27, 32 – head, frontal view; 23, 28, 33 – anal ventrite.



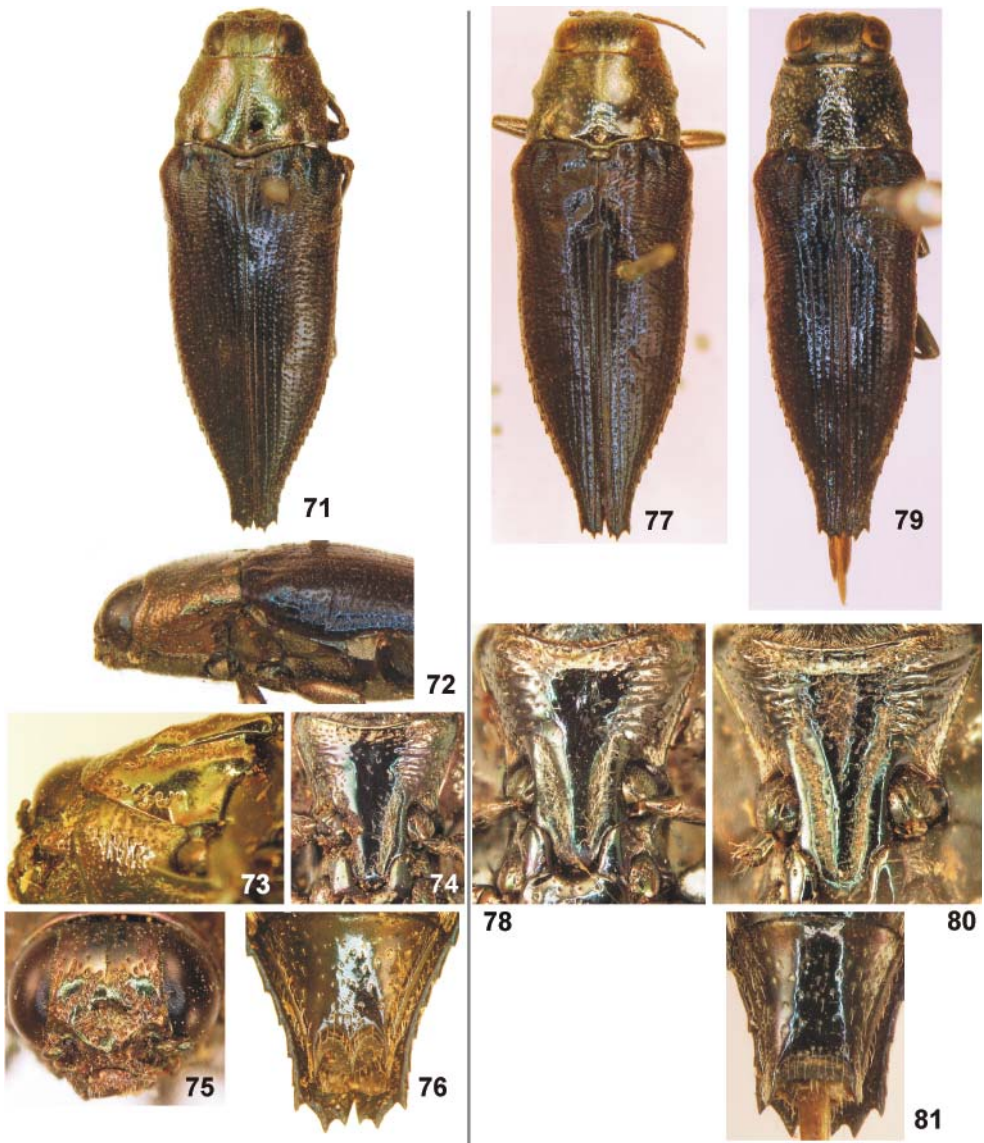
Figs. 34–43. 34–39 – *Lamprodila (Erialata) auripilis* (Obenberger, 1922), syntype, female, 10.5 mm, Borneo; 39 – the same, female from Pahang. 40–43 – *L. (E.) pubescens* (Fisher, 1930), holotype, 10.5 mm, Perak (photos by M. G. Volkovitsh). 34, 40 – habitus, dorsal view; 41 – habitus, ventral view; 35 – habitus, lateral view; 36, 42 – head, frontal view; 37 – prosternal process; 38–39, 43 – anal ventrite.



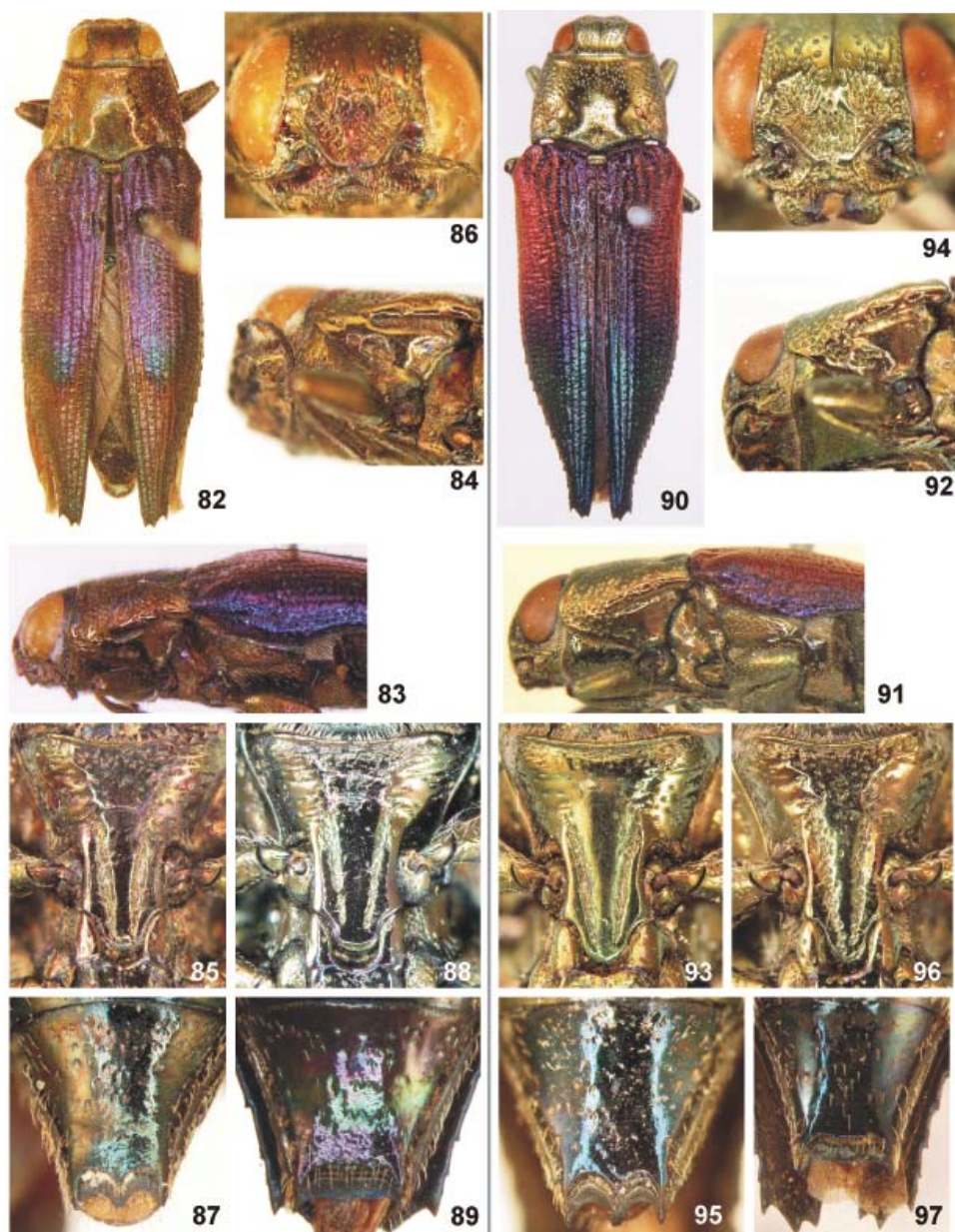
Figs. 44–58. 44–48 – *Lamprodila (Cinyrisia) psilopteroides* (Deyrolle, 1864), female (NMPC), 19.0 mm, Borneo. 49–53 – *L. (Mabomisia) sexspinosa* (Thomson, 1857), syntype, 15.5 mm, Borneo. 54–58 – *Melobasina solomonensis* (Théry, 1937), holotype, female, 12.0 mm, Solomons. 44, 49, 54 – habitus, dorsal view; 45, 50, 55 – habitus, lateral view; 46, 51, 56 – head, frontal view; 47, 52 – prosternal process; 57 – habitus, ventral view; 48, 53, 58 – anal ventrite.



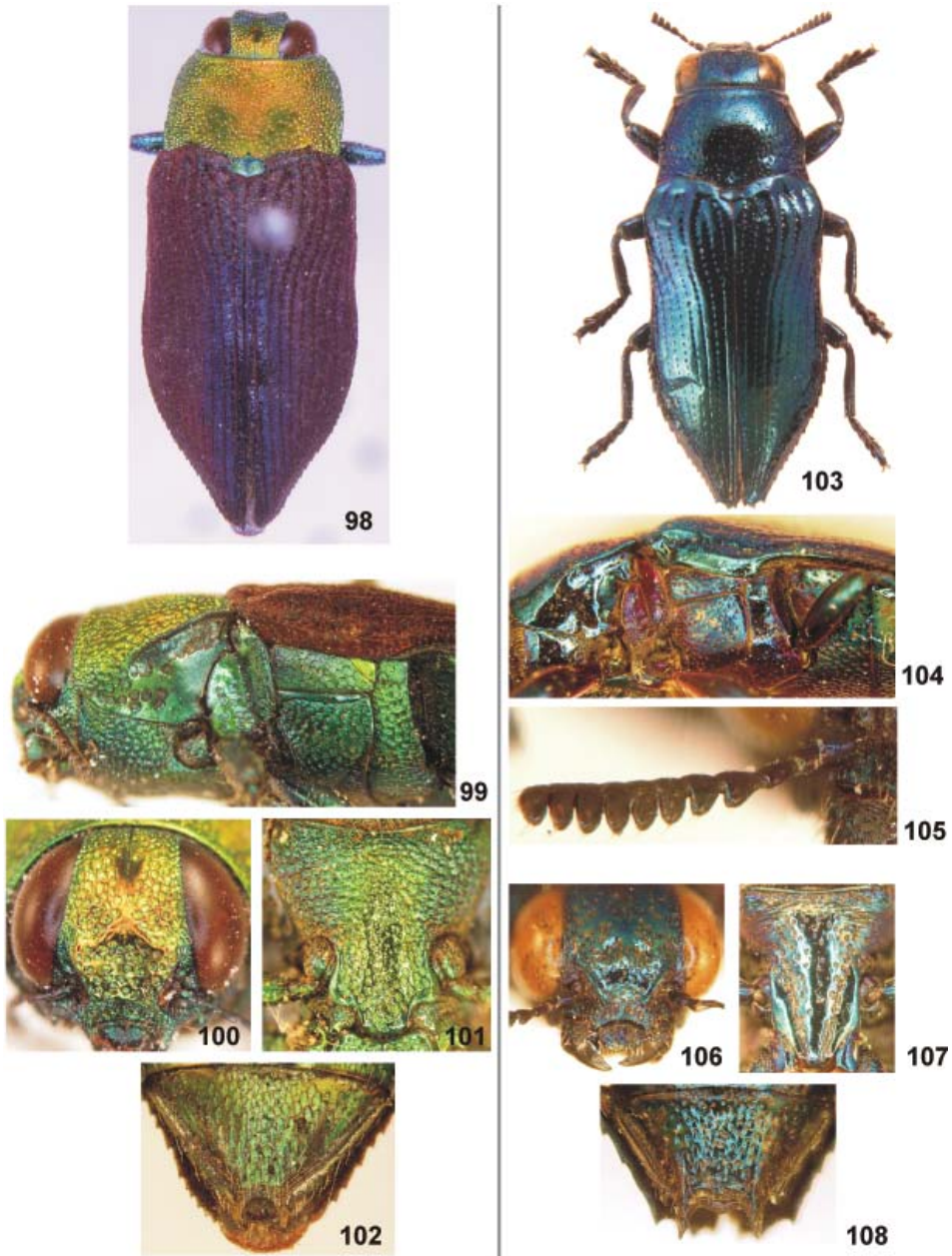
Figs. 59–70. 59–64 – *Melobasina apicalis* Kerremans, 1900, lectotype, female, 13.5 mm, Tagula Island (except 60 – female, 11.5 mm, Guadalcanal Island). 65 – *M. ignita* (Théry, 1937) (synonym of *M. apicalis*), holotype, female, 12.5 mm, Bougainville Island. 66–70 – *M. suturalis* (Deyrolle, 1864), female (NMPC), 13.0 mm, Halmahera Island. 59–60, 65–66 – habitus, dorsal view; 61, 67 – habitus, lateral view; 62, 68 – head, frontal view; 63, 69 – prosternal process; 64, 70 – anal ventrite.



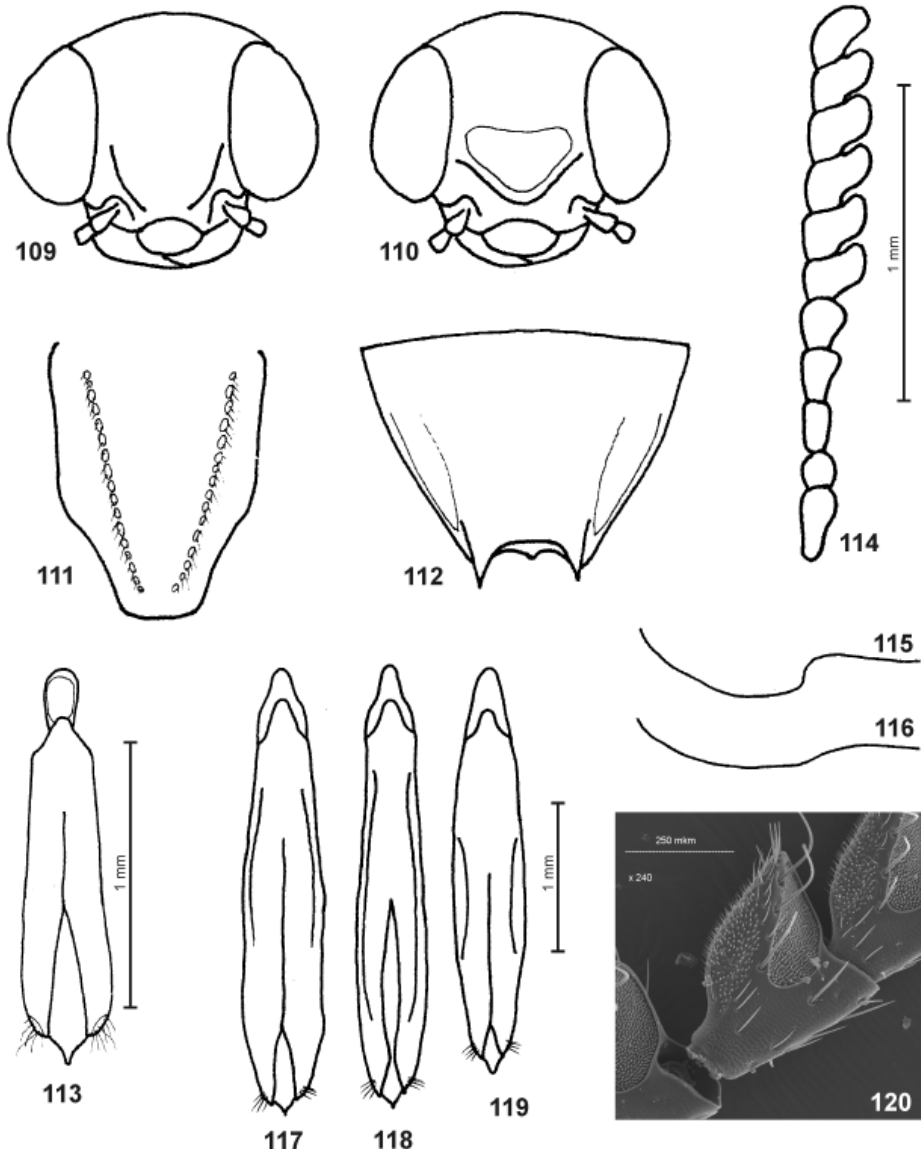
Figs. 71–81. 71–76 – *Nesotrinchus australicus* (Kerremans, 1903), lectotype, female, 14.0 mm, ‘Australie’. 77–78 – *N. simondsi* Obenberger, 1924 (synonym of *N. australicus*), paralectotype, female (BMNH), 15.0 mm, Fiji. 79–81 – *N. simondsi*, lectotype, male, 12.0 mm, Fiji. 71, 77, 79 – habitus, dorsal view; 72 – habitus, lateral view; 73 – thorax, lateral view; 74, 78, 80 – prosternal process; 75 – head, frontal view. 76, 81 – anal ventrite.



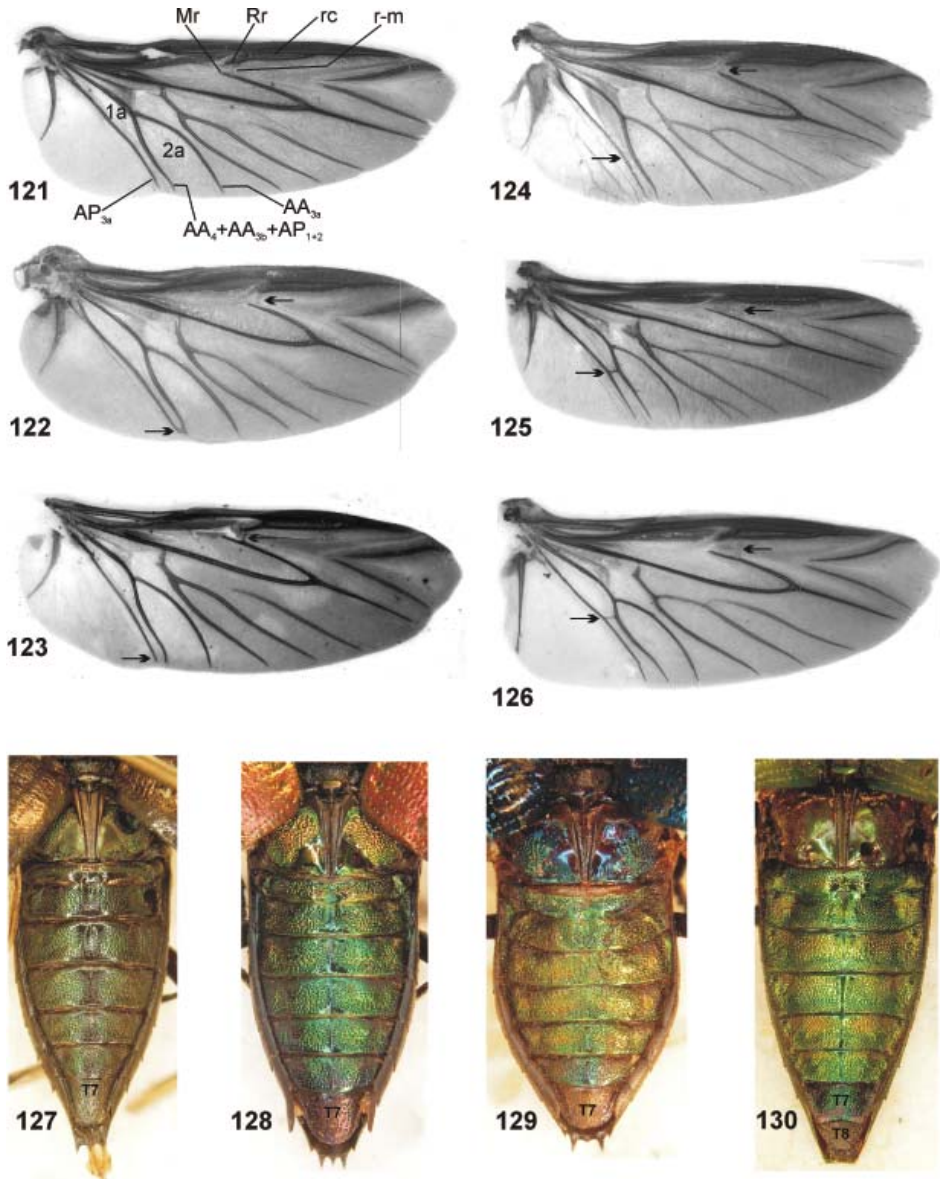
Figs. 82–97. 82–87 – *Nesotrinchus caeruleipennis* (Fairmaire, 1877), lectotype, female, 11.0 mm, Tonga. 88–89 – *N. orientalis* Bellamy, 1987 (synonym of *N. caeruleipennis* Fairmaire), paratype, male (BMNH), 11.0 mm, Tonga. 90–95 – *N. caeruleipennis* (Thomson, 1878) (= *thomsoni* Bílý & Kubáň, nom. nov.), lectotype, female, 13.5 mm, Fiji. 96–97 – *N. thomsoni* Bílý & Kubáň, nom. nov., male, 11.5 mm, Fiji. 82, 90 – habitus, dorsal view; 83, 91 – habitus, lateral view; 84, 92 – thorax, lateral view; 85, 88, 93, 96 – prosternal process; 86, 94 – head, frontal view; 87, 89, 95, 97 – anal ventrite.



Figs. 98–108. 98–102 – *Lamprodila (Ovalisia) auricollis* (Deyrolle, 1864), female, 11.0 mm, Halmahera Island. 103–108 – *Ulaikoilia jelineki* Bílý & Kubáň, gen. nov., sp. nov., holotype, male, 9.8 mm, Papua New Guinea. 98, 103 – habitus, dorsal view; 99, 104 – habitus, lateral view; 105 – right antenna; 100, 106 – head, frontal view; 101, 107 – prosternal process; 102, 108 – anal ventrite.



Figs. 109–120. 109, 111–115 – *Ulaikoilia jelineki* Bílý & Kubáň, gen. nov., sp. nov., holotype, male (109 – head, frontal view; 111 – prosternal process; 112 – anal ventrite; 113 – aedeagus; 114 – right antenna; 115 – subhumeral portion of elytral margin). 110, 116–117 – *Nesotrinchus coeruleipennis* (Fairmaire, 1877) (110 – head, frontal view; 116 – subhumeral portion of elytral margin; 117 – aedeagus, dorsal view). 118 – *N. thomsoni* Bílý & Kubáň, nom. nov., aedeagus, dorsal view. 119 – *N. australicus* (Kerremans, 1903), aedeagus, dorsal view. 120 – *Pseudhyperantha jucunda* Saunders, 1869, male, Malaysia, right antennomere 7, ventral view, apical fossa and lateral field (Photo: M. G. Volkovitsh).



Figs. 121–130. 121, 127 – *Nesotrinchus australicus* (Kerremans, 1903): paralectotype of a synonym *N. simondsi* Obenberger, 1924, female, Fiji, NMPC. 122, 129 – *Ulaikoilia jelineki* Bílý & Kubáň, gen. nov., sp. nov., holotype, Papua New Guinea. 123 – *Melobasina apicalis* Kerremans, 1900, female, Guadalcanal Island. 124 – *Lamprodila (Ovalisia) auricollis* (Deyrolle, 1864), female, Halmahera Island. 125 – *Cardiaspis pisciformis* Théry, 1904, female (NMPC), India. 126 – *Haplotrinchus pyrochlorus* (Fairmaire, 1877), syntype, female (NMPC), Fiji. 128 – *Melobasina ignita* (Théry, 1937) (synonym of *M. apicalis*), holotype, female, Bougainville Island. 130 – *Haplotrinchus philippinensis* Obenberger, 1928, syntype, female, Philippines, Luzon. 121–126 – right wing, dorsal view; 127–130 – abdomen, dorsal view; T7 = visible tergite 7; T8 = visible tergite 8.

***Lamprodila (Mabomis) sexspinosa* (Thomson, 1857)**

(Figs. 49–53)

Chalcophora (Evides) sexspinosa Thomson, 1857: 110. Type locality: Borneo (without exact data).
Dicercomorpha sexspinosa: DEYROLLE (1864): 54 (key; new combination), 58 (list); GEMMINGER & HAROLD (1869): 1372 (catalogue); SAUNDERS (1871): 35 (catalogue); KERREMANS (1892): 84 (catalogue); WATERHOUSE (1913): 182 (list); OBENBERGER (1930): 358 (catalogue); BELLAMY (1997): 76 (note).
Haplotrinchus sexspinosa: KERREMANS (1903): 127 (list of species; new combination); MOULTON (1911): 168 (list).
Ovalisia (Mabomis) sexspinosa: HOLYŃSKI (2000): 3 (characters; new combination), Fig. 9; BELLAMY (2003): 40 (catalogue), Fig. 179.
Lamprodila (Mabomis) sexspinosa: BELLAMY (2008): 621 (catalogue; new combination).

Type material examined. SYNTYPE: ♀ (MNHN), BORNEO: ‘Borneo [h] / Sexspinosa Type Thoms. [h] / Th. Type [p] / Ex Musaeo James Thomson [p] / Type [p] [red label] / Muséum Paris 1952 Coll. R. Oberthür [p]’. Number of syntypes unknown.

Additional material examined. BORNEO: ‘Borneo [h] / Ex-Musaeo Mniszecz [p] / Muséum Paris 1952 Coll. R. Oberthür [p]’ (1 ♀, MNHN); ‘Ex-Musaeo Mniszecz [p] / Muséum Paris 1952 Coll. R. Oberthür [p]’ (1 ♀, MNHN). MALAYSIA: SABAH: km 53 Kota Kinabalu – Tambunan, Gn. Emas, 1650 m, March 2000, ex Coll. S. Chew (1 ♀, VKCP).

Distribution. Borneo (no further details available); Malaysia (Sabah).

Subgenus *Ovalisia* Kerremans, 1900

Ovalisia Kerremans, 1900b: 67. Type species: *Ovalisia insularis* Kerremans, 1900, by monotypy.
Ovalisia: KERREMANS (1900a): 294 (comment); KERREMANS (1903): 134 (list; synonym of *Lampra*); OBENBERGER (1930): 347 (catalogue; synonym of *Lampra*); OBENBERGER (1956): 41 (comment; synonym or subgen. of *Lampra*); DESCARPENTRIES & VILLIERS (1963): 266 (comment; as valid genus of Dicercini); LERAUT (1983): 6 (as subgenus of *Poecilonota*; comment; new status); BELLAMY (1985): 418 (catalogue; as valid genus of Buprestinae: Dicercini); COBOS (1986): 113 (key), 118 (characters; comment; as valid genus), 303 (list; in Buprestinae: Dicercini); HOLYŃSKI (1993): 13 (list; in Buprestinae: Buprestini: Psilopterina), 25 (comment; as valid genus); BÍLÝ (1997): 10 (characters; key; comment; synonymy; as valid genus of Buprestinae: Dicercini), 12 (list of species); HOLYŃSKI (2000): 1 (characters; as valid genus; key to subgenera; in Buprestinae: Buprestini: Psilopterina), Fig. 1 (distribution), Fig. 12 (cladogram); BELLAMY (2003): 40 (catalogue; as valid genus of Chrysochroinae: Poecilonotini); BELLAMY (2008): 601 (as subgenus of *Lamprodila*; catalogue; new combination; in Chrysochroinae: Poecilonotini).
Pagdenia Théry, 1934: 236 (characters; junior primary homonym). Type species: *Pagdenia plasoni* Théry, 1934, by original designation.
Pagdenia: THÉRY (1935): 143 (synonym of *Pagdeniella*); BELLAMY (1985): 420 (catalogue; synonym of *Pagdeniella*); BÍLÝ (1993b): 113 (synonym of *Pagdeniella*); BÍLÝ (1997): 10 (synonym of *Ovalisia*); HOLYŃSKI (2000): 5 (synonym of *Ovalisia*); BELLAMY (2003): 40 (catalogue; synonym of *Ovalisia*); BELLAMY (2008): 601 (catalogue; synonym of *Ovalisia*).
Pagdeniella Théry, 1935: 143 (new substitute name).
Pagdeniella: BELLAMY (1985): 420 (catalogue; in Buprestinae: Buprestini); HOLYŃSKI (1993): 14 (list), 30 (comment; in Buprestinae: Anthaxiini: Bubastina); BÍLÝ (1993a): 44, 45 (key; in Buprestinae: Buprestini); BÍLÝ (1993b): 112 (revision; characters; key to species; in Buprestinae: Buprestini); BELLAMY (1996): 215 (note; in Buprestinae: Buprestini: Thomassetiina), 219 (character state matrix), 220 (cladogram); BÍLÝ (1997): 10 (synonym of *Ovalisia*); HOLYŃSKI (2000): 5 (synonym of *Ovalisia*); BELLAMY (2003): 40 (catalogue; synonym of *Ovalisia*); BELLAMY (2008): 601 (catalogue; synonym of *Ovalisia*).

Remarks. The subgenus *Ovalisia* is a typical representative of the subtribe Poecilonotina and was revised by BÍLÝ (1993b, 1997). The list of studied specimens – see the chapter Material and methods. Species of the subgenus *Ovalisia* and the genera *Melobasina* and *Ulaikoilia* gen. nov. share nearly the same distribution. Moreover, the body shape of *Ulaikoilia jelineki* gen. nov., sp. nov. is very similar to some species of the subgenus *Ovalisia* (Fig. 98). Wing as in Fig. 124.

Revision of the subtribe *Nesotrinchina* subtrib. nov.

Subtribe *Nesotrinchina* Bílý, Kubáň & Volkovitsh, subtrib. nov.

(Figs. 54–97, 103–119, 121–123)

'*Nesotrinchus* generic group': VOLKOVITSH (2001): 55 (catalogue of higher taxa; in Poecilonotini); BELLAMY (2003): 41 (catalogue; in Poecilonotini); BELLAMY (2008): 624 (catalogue; in Poecilonotini).

Type genus. *Nesotrinchus* Obenberger, 1924 (present designation).

Diagnosis. See the key to subtribes (p. 736).

Genera included. *Melobasina* Kerremans, 1900, *Nesotrinchus* Obenberger, 1924, *Ulaikoilia* Bílý & Kubáň, gen. nov.

Key to genera of the subtribe *Nesotrinchina* subtrib. nov.

- 1(2) Body short and stout, elytra less than twice as long as wide (Fig. 103); supraantennal carinae strongly developed, frons between them moderately convex (Figs. 106, 109); distal antennomeres of male twice as wide as long (Figs. 105, 114); subhumeral elytral lobe strongly developed (Figs. 104, 115); latero-posterior pronotal depressions missing (Fig. 103); hind wings: anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} joined distally (Fig. 122); visible abdominal tergites strongly transverse, terminal tergite (T7) wider than long (Fig. 129); anal ventrite with obtuse lateral carinae and well-developed depression between them and lateral margins of ventrite (Figs. 108, 112); aedeagus short, slightly enlarged posteriorly (Fig. 113). *Ulaikoilia* Bílý & Kubáň, gen. nov.
- 2(1) Body slender, navicular, elytra 2.0–2.4 times as long as wide (Figs. 54, 59–60, 65, 71, 77, 79, 82, 90); supraantennal carinae missing or strongly developed, frons flat or deeply depressed (Figs. 56, 62, 68, 75, 86, 94, 110); distal antennomeres of male (in species for which male is known) 1.0–1.2 times as wide as long; subhumeral elytral lobe less developed, not prominent (Figs. 55, 61, 67, 72, 83, 91, 116); lateroposterior pronotal depressions well-developed (Figs. 54, 59, 66, 71, 82, 90); hind wings: anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free (Fig. 121, 123); visible abdominal tergites moderately transverse, terminal tergite (T7) longer than wide (Figs. 127–128); anal ventrite without longitudinal carinae and lateral depressions, only lateral spines of ventrite with obtuse carinae (Figs. 58, 64, 70, 76, 81, 87, 89, 95, 97); aedeagus (in species for which male is known) long, spindle-shaped (Figs. 117–119).
- 3(4) Elytral apex strongly caudiform (Figs. 71, 77, 79, 82, 90); supraantennal carinae strongly developed, frons between them deeply depressed (Figs. 75, 86, 94, 110); pronotum conical usually with prominent, more or less distinct knoll at midlength or anterior of posterior angles (Figs. 71, 77, 79, 82, 90); anal ventrite of female sharply trispinose (Figs. 76, 87, 95). *Nesotrinchus* Obenberger, 1924, stat. restit.
- 4(3) Elytra navicular, not caudiform (Figs. 54, 59, 66); supraantennal carinae missing, antennal sockets margined by sharp but not elevated edge, frons flat (Figs. 56, 62, 68); lateral pronotal margins nearly straight, moderately converging anteriorly (Figs. 54, 59, 66), lateral keels obtuse (Figs. 55, 61, 67); anal ventrite of female sharply trispinose (Figs. 58, 64) or median spine reduced into small tooth (Fig. 70). *Melobasina* Kerremans, 1900

Genus *Melobasina* Kerremans, 1900

(Figs. 54–70, 123, 128)

Melobasina Kerremans, 1900b: 68. Type species: *Melobasina apicalis* Kerremans, 1900, by monotypy.

Melobasina: KERREMANS (1903): 138 (key to genera), 144 (characters; in Buprestini: 'Buprestites'); OBENBERGER (1923): 17 (note); OBENBERGER (1930): 420 (catalogue; in Buprestini: 'Buprestites'); BELLAMY (1985): 420 (catalogue; in Buprestinae: Buprestini); HOLYŃSKI (1993): 13 (list), 27 (comment; in Buprestinae: Buprestini: Haplotrinchina); BELLAMY (1997): 76 (list; with *Nesotrinchus* in synonymy; in Buprestinae: Buprestini: Haplotrinchina); VOLKOVITSH (2001): 96 (note; in Chalcophorinae [now Chrysochroinae]: Poecilnotini: '*Nesotrinchus* generic group'); BELLAMY (2002): 62 (catalogue; with *Nesotrinchus* in synonymy; in Chalcophorinae: Psilopterini: Haplotrinchina); BELLAMY (2003): 41 (catalogue; with *Nesotrinchus* in synonymy; in Chrysochroinae: Poecilnotini: '*Nesotrinchus* generic group'); BELLAMY (2008): 624 (catalogue; with *Nesotrinchus* in synonymy; in Chrysochroinae: Poecilnotini: '*Nesotrinchus* generic group').

Diagnosis. This genus is characterised by navicular, not caudiform elytra (Figs. 54, 59, 66), missing supraantennal carinae, antennal sockets margined by sharp but not elevated edge, and flat frons (Figs. 56, 62, 68); lateral pronotal margins nearly straight, moderately converging anteriorly, lateral keels obtuse (Figs. 55, 61, 67); anal ventrite of female sharply trispinose (Figs. 58, 64) or median spine reduced into small tooth (Fig. 70); visible abdominal tergites moderately transverse, terminal tergite (T7) longer than wide; wing (Fig. 123) with anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free.

Remarks. BELLAMY (2002, 2003, 2008) misinterpreted the note of HOLYŃSKI (1993: 27) as a synonymisation of the genera *Nesotrinchus* and *Melobasina* (see above).

Key to species of *Melobasina* Kerremans, 1900

Note. *Melobasina fossicollis* (Kerremans, 1906) from the Solomon Islands (Shortland) is not included in the key; we could not locate any specimens and the original description is insufficient for a safe diagnosis.

- 1(2) Pronotum golden green with black disc; elytra blue with narrow, golden-green sutural stripe reaching elytral apex (Figs. 66); median spine of anal ventrite of female small, strongly reduced, much shorter than lateral spines (Fig. 70); 13.0 mm; Indonesian Moluccas (Halmahera). *M. suturalis* (Deyrolle, 1864)
- 2(1) Pronotum golden purple or blue without median, dark spot; elytra golden purple, violet or dark olive green, sometimes with darkened apical third, always without sutural stripe (Figs. 54, 59–60, 65); medial spine of anal ventrite of female well developed, as long as lateral spines (Fig. 64) or slightly longer (Fig. 58); eastern New Guinean islands (Bougainville, Tagula), Solomon Islands (Malaita, Guadalcanal and Shortland).
- 3(4) Pronotum dark blue, elytra violet with green tinge at humeral third (Fig. 54); median spine of anal ventrite of female somewhat longer than lateral spines (Fig. 58); 12.0 mm; Solomon Islands (Malaita). *M. solomonensis* (Théry, 1937)
- 4(3) Pronotum golden purple, elytra golden purple or dark olive green, sometimes with darkened (violet to black) apical third (Figs. 59–60, 65); median spine of anal ventrite of female as long as lateral spines (Fig. 64); 11.5–13.5 mm; Papua New Guinea (Bougainville, Tagula), Solomon Islands (Guadalcanal). *M. apicalis* Kerremans, 1900

***Melobasina apicalis* Kerremans, 1900**

(Figs. 59–65, 123, 128)

Melobasina apicalis Kerremans, 1900b: 69. Type locality: Papua New Guinea, Louisiade Archipelago, Tagula Island ('Ile Tugela').

Melobasina apicalis: KERREMANS (1903): 144 (list of species; distribution: 'Ile Tugela'), pl. 2, Fig. 2; OBENBERGER (1930): 420 (catalogue; distribution: 'Ins. Tugela'); BELLAMY (2002): 62 (catalogue); BELLAMY (2008): 625 (catalogue; distribution: Australasian: 'Tugela Island').

Nosotrinchus [sic!] *Ignitus* Théry, 1937: 647, **syn. nov.** Type locality: Papua New Guinea, Bismarck Archipelago, Bougainville province, Bougainville Island, Buin.

Nesotrinchus ignitus: BELLAMY (1987): 101 (note).

Melobasina ignita: BELLAMY (2008): 625 (catalogue; new combination; distribution: Australasian: Solomon Islands – mistake).

Misidentification. *Melobasina apicalis* Kerremans *sensu* BELLAMY (2003): 120, Fig. 185. The specimen is in fact a syntype of *Nesotrinchus simondsi*.

Type material examined. *Melobasina apicalis*: LECTOTYPE (designated here): ♀ (BMNH), PAPUA NEW GUINEA: LOUISIADE ARCHIPELAGO: 'Tugela [= Tagula Island], Sol[omon]. Is. ([C. M.] Wodford [leg.]) [p] / apicalis Kerr. Type. [h] / Type [p] [red-framed round label] / Melobasina Kerrem. [h] / M. apicalis Kerr. I. Tugela [h] / Kerremans. 1903–59. [p]'. Since the number of syntypes is unknown, we designate here the female specimen from BMNH as the lectotype to avoid any confusion in the future.

Melobasina ignita: HOLOTYPE (by monotypy): ♀ (NHMB, coll. G. Frey), PAPUA NEW GUINEA: BISMARCK ARCHIPELAGO: Bougainville Island: 'Buin Ile Salomon [h] / Nosotrinchus ignitus Thery [h] Type [p, red] / keine Type aber Paratype 1956 det. Kamp [h]'.
Additional material examined. SOLOMON ISLANDS: GUADALCANAL ISLAND: 'Wanderers Bay 84. 3 [h] / Solomon Islands, W Guadalcanal Isl., Wanderer Bay coast [h, Kubán's writing] / Melobasina fossicollis Kerr. det from descr. [h] B. Levey det. 196 [p] 8 [h]' (2 ♀♀, BMNH).

Remarks. BELLAMY (2008) formally transferred '*Nesotrinchus ignitus*' to the genus *Melobasina*. We have studied the types of *M. ignita* and *M. apicalis* and two females from Guadalcanal and failed to find any substantial differences and we consider them conspecific, *M. ignita* (Fig. 65) being a junior subjective synonym of *M. apicalis* (Fig. 59). HOLYŃSKI (1993: 27) most probably studied specimens from Guadalcanal deposited in BMNH and determined by B. Levey as '*Melobasina fossicollis* Kerr.' (see above).

Distribution. Papua New Guinea (Bismarck Archipelago: Bougainville Island; Louisiade Archipelago: Tagula Island), Solomon Islands (Guadalcanal Island).

***Melobasina fossicollis* (Kerremans, 1906)**

Exagistus fossicollis Kerremans, 1906: 415. Type locality: Solomon Islands: Shortland Island [Not Bismarck Archipelago!]

Melobasina fossicollis: OBENBERGER (1923): 17 (note; new combination); OBENBERGER (1930): 420 (catalogue; distribution: 'Archipel Bismarck' – mistake); HOLYŃSKI (1993): 27 (note); BELLAMY (2008): 625 (catalogue; distribution: Australasian: Papua New Guinea, Bismarck Archipelago – mistake).

Remarks. KERREMANS (1906) mentioned '2 exemplaires, des îles Shortland' (length 13 mm, width 4 mm); OBENBERGER (1923) transferred this species to the genus *Melobasina*, evidently based on the study of one of both syntypes and on illustration of *M. apicalis* in KERREMANS (1903). In the corresponding volume of Obenberger's catalogue (Obenberger 1930: 420) from the library of Obenberger (deposited in NMPC), we can find his handwritten note that this species is represented in his collection (deposited in NMPC) but we failed to find any

corresponding specimen in the collection. It is evident from the description that the type specimens came from the collection of H. E. R. von Bennigsen. A part of Bennigsen's collection (including material from New Guinea and Bismark Archipelago) was deposited in DEIC (HORN et al. 1990) and the rest was left in the collection of Bennigsen in Waitze an der Warthe (now Wiejce, by Warta river, Wojewódstwo Lubuskie, western Poland) but this species is not mentioned in the list of types deposited in DEIC (GAEDIKE 1985) and we failed to find any data about the fate of the rest of Bennigsen's collection (e.g. in GAEDIKE 1995). According to BELLAMY (2008), the type is deposited in MNHN but we did not find this type in MNHN or BMNH. It is also not mentioned in the list of types housed in IRSN. Given the original description and the type locality, we accept the transfer proposed by OBENBERGER (1923). We are also inclined to believe that *M. fossicollis* and *M. apicalis* are conspecific.

Distribution. Solomon Islands (Shortland Island).

Melobasina solomonensis (Théry, 1937)

(Figs. 54–58)

Nosotrinchus [sic!] *solomonensis* Théry, 1937: 38, pl. 3, Fig. 4. Type locality: Solomon Islands, Malaita Island, Auki.

Nosotrinchus solomonensis: THÉRY 1943: 646 (as *Nosotrinchus* [sic!]; list of species; distribution: Solomon Islands); BELLAMY (1987): 101 (note).

Melobasina solomonensis: BELLAMY (2008): 625 (catalogue; new combination; distribution: Australasian: Solomon Islands).

Type material. HOLOTYPE (by monotypy): ♀ (MCZC), SOLOMON ISLANDS: MALAITA ISLAND: 'Solomons: Malaita. Auki W. M. Mann [leg.] [p] / *Nosotrinchus* Salomoniensis [sic!] Thery [h] Type [p, red] / M.C.Z. Type [p] 22510 [h] [red label]'.

Remarks. BELLAMY (2008) formally transferred this species to the genus *Melobasina*. We have studied the types of *M. apicalis* and *M. ignita* and illustrations of *M. solomonensis* from the Database of insect primary types in the collection of the Museum of Comparative Zoology at Harvard University (Figs. 54–58) and found them to be very closely related but different, congeneric species.

Distribution. Solomon Islands (Malaita Island).

Melobasina suturalis (Deyrolle, 1864), comb. nov.

(Figs. 66–70)

Poecilnota suturalis Deyrolle, 1864: 58 (key) 64 (characters). Type locality: Indonesia, Maluku Islands, Halmahera island ('Gilolo').

Poecilnota suturalis: GEMMINGER & HAROLD (1869): 1376 (catalogue; distribution: 'Gilolo'); SAUNDERS (1871): 38 (catalogue; distribution: 'Gilolo'); KERREMANS (1892): 92 (catalogue; distribution: 'Gilolo').

Lampra suturalis: KERREMANS (1903): 135 (list of species; new combination; distribution: 'Gilolo'); OBENBERGER (1930): 356 (catalogue; distribution: 'Gilolo').

Lamprodila (Palmar) *suturalis*: BELLAMY (2008): 622 (catalogue; new combination; distribution: Indonesia: Halmahera, Kalimantan).

Type material examined. LECTOTYPE (designated here): sex not examined (MNHN, coll. Oberthür), INDONESIA: MALUKU ISLANDS: Halmahera Island: 'suturalis HDeyr. I. Gilolo [h]'. Since the number of syntypes is unknown, we designate here the specimen mentioned above as the lectotype to avoid any confusion in the future.

Additional material examined. INDONESIA: MALUKU ISLANDS: Halmahera Island (1 ♀, NMPC); Halmahera Island NW, 7 km SE of Jailolo, 200 m, 1°1'18"N 127°31'39"E, 26.–27.i.2006, UWP + clearing, A. Skale leg. (1 ♀, MNCA).

Remarks. BELLAMY (2008) formally transferred this species to the genus *Lamprodila* (subgenus *Palmar*) and mentioned it also from Kalimantan. We failed to trace any specimen or data from Kalimantan and suppose that its presence there is highly improbable. We find *M. suturalis* rather different from other species of the genus.

Distribution. Indonesia (Maluku Islands: Halmahera Island). ?Kalimantan.

Genus *Nesotrinchus* Obenberger, 1924, stat. restit.

(Figs. 71–97, 110, 116–119, 121, 127)

Nesotrinchus Obenberger, 1924: 12 (in Buprestini: Dicerctites), pl. 1, Fig. 2, pl. 2, Figs. 10–12, 14. Type species: *Nesotrinchus simondsi* Obenberger, 1924, by original designation.

Nesotrinchus: THÉRY (1925): 225 (as *Nosotrinchus* [sic!] – incorrect subsequent spelling; comment; synonym of *Haplotrinchus*); OBENBERGER (1926a): 97 (characters; comment; as valid genus); OBENBERGER (1930): 359 (catalogue; in Buprestini: Dicerctites); THÉRY (1937): 38, 39 (as *Nosotrinchus* [sic!]; characters); THÉRY (1943): 646 (as *Nosotrinchus* [sic!]; list of species; synonymy); BELLAMY (1985): 419 (catalogue; in Buprestinae: Dicerctini); BELLAMY (1986): 595 (catalogue; in Buprestinae: Dicerctini); BELLAMY (1987): 595 (comment; in Buprestinae: Dicerctini); HOLYŃSKI (1993): 27 (comment; as valid genus near to *Melobasina*, not as synonym of *Melobasina*; in Buprestinae: Buprestini: Haplotrinchina); BELLAMY (1997): 76 (as synonym of *Melobasina* – misinterpretation of HOLYŃSKI (1993: 27); in Buprestinae: Buprestini: Haplotrinchina); VOLKOVITSH (2001): 55 (catalogue; in Chalcophorinae [now Chrysochroinae]: Poecilonotini: ‘*Nesotrinchus* generic group’) 96 (characters); BELLAMY (2002): 62 (catalogue; as synonym of *Melobasina* – misinterpretation; in Buprestinae: Psilopterini: Haplotrinchina); BELLAMY (2003): 41 (catalogue; as synonym of *Melobasina* – misinterpretation; in Chrysochroinae: Poecilonotini: ‘*Nesotrinchus* generic group’); BELLAMY (2008): 624 (catalogue; as synonym of *Melobasina* – misinterpretation; in Chrysochroinae: Poecilonotini: ‘*Nesotrinchus* generic group’).

Diagnosis. The genus is characterised by a strongly caudiform elytral apex (Figs. 71, 77, 79, 82, 90); strongly developed supraantennal carinae with frons deeply depressed between them (Figs. 75, 86, 94, 110); pronotum conical and usually with prominent, more or less distinct lateral knoll at midlength or anterior of posterior angles (Figs. 71, 77, 79, 82, 90); anal ventrite of female sharply trispinose (Figs. 76, 87, 95); visible abdominal tergites moderately transverse, terminal tergite (T7) longer than wide; wing (Fig. 121) with anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free and aedeagi as in Figs. 117–119.

Remarks. BELLAMY (2002, 2003, 2008) formally transferred all *Nesotrinchus* species to the genus *Melobasina* due to a misinterpretation of the work by HOLYŃSKI (1993: 27) (see above). ‘*Buprestis wallisii* Montrouzier, 1855’ (type locality: Pacific Islands: Wallis Island) was placed in the most catalogues (KERREMANS 1903, OBENBERGER 1930, BELLAMY 2008) into the genus *Haplotrinchus* but in parallel also into the genus *Paracupta* Deyrolle, 1864 (now in Chrysochroinae: Chalcophorina) (KERREMANS 1903, OBENBERGER 1926b). As mentioned already by FAIRMAIRE (1881), this species is most probably similar to *N. caeruleipennis* (Fairmaire, 1877) due to its size (ca. 13.0 mm) and characters mentioned in the original description. We also suppose that this species is congeneric with other species of the genus *Nesotrinchus*; this genus forms a very compact taxon distributed only on the Pacific archipelagos of Fiji and Tonga and on the islands of Niue and Wallis.

Key to species of *Nesotrinchus* Obenberger, 1924

Note. *Nesotrinchus wallisii* (Montrouzier, 1855), distributed in the Pacific Islands (Wallis Island), is not included in the key because no specimens are available for study.

- 1(2) Lateral margins of pronotum with one prominent knoll at midlength and with another, much smaller knoll or gibbosity anteriorly posterior angles; elytral apex strongly caudiform; body nearly unicolorous: elytra black with slight blue lustre, head and pronotum with coppery-green lustre or whole body completely black (Figs. 71, 77, 79); aedeagus as in Fig. 119; 12.0–15.0 mm; Pacific Islands (Fiji). *N. australicus* (Kerremans, 1903)
- 2(1) Lateral margins of pronotum conical with small gibbosity anteriorly posterior angles; elytral apex less caudiform; body bi- or tricolorous: head and pronotum shining cupreous, elytra violet or violet with apical third shining blue (Figs. 82, 90).
- 3(4) Lateral pronotal margins nearly straight, with small but distinct gibbosity anteriorly posterior angles (Fig. 82) (see also Bellamy (1987): Fig. 1, as '*orientalis*'); pronotal disc with very fine punctation; elytral apices finely and shortly trispinose (Fig. 82) (see also Bellamy (1987): Fig. 1, as '*orientalis*'); lateral spines of anal ventrite less developed (Fig. 87); parameres of aedeagus parallel at posterior third (Fig. 117); 10.8–13.2 mm; Pacific Islands (Tonga, Niue). *N. caeruleipennis* (Fairmaire, 1877)
- 4(3) Lateral pronotal margins slightly convex with very small gibbosity anteriorly posterior angles (Fig. 90) (see also Bellamy (1987): Fig. 2, as '*caeruleipennis*'); pronotal disc with rougher punctation; elytral apices sharply and strongly trispinose (Fig. 90) (see also Bellamy (1987): Fig. 2, as '*caeruleipennis*'); lateral spines of anal ventrite longer and sharper (Fig. 95); parameres of aedeagus convex laterally (Fig. 118); 11.5–13.5 mm; Pacific Islands (Fiji). *N. thomsoni* Bílý & Kubáň, **nom. nov.**

Nesotrinchus australicus (Kerremans, 1903), **comb. nov.**

(Figs. 71–81, 119, 121, 127)

Haplotrinchus australicus Kerremans, 1903: 127. Type locality: Australia (without exact data).

Nesotrinchus simondsi Obenberger, 1924: 13, pl. 1, Fig. 2, pl. 2, Figs. 10–12, 14, **syn. nov.** Type locality: Fiji Islands, Ovalau Island.

Haplotrinchus simondsi: THÉRY (1925): 225 (characters; comment; synonym of *caeruleipennis* Thomson).

Nesotrinchus simondsi: OBENBERGER (1926a): 98 (characters; comment; valid species); OBENBERGER (1930): 359 (catalogue; distribution: Fiji); THÉRY (1943): 646 (list of species; in synonymy of *caeruleipennis* Fairmaire and *caeruleipennis* Thomson; distribution: Fiji); BELLAMY (1987): 101, 104 (comments); BELLAMY (1990): 91 (characters; comment; new locality for Fiji: Viti Levu Island).

Haplotrinchus australicus: OBENBERGER (1930): 358 (catalogue; distribution: Australia); BELLAMY (1997): 76 (note).

Nesotrinchus australicus: THÉRY (1943): 644 (comment; new combination), 646 (list of species; distribution: Australia); BELLAMY (1987): 101 (note), 104 (comment); HOLYŃSKI (1993): 27 (note).

Melobasina australica: BELLAMY (2002): 61 (as *australis* [sic!]; comment), 62 (as *austratica*; catalogue; new combination; distribution: Australia); BELLAMY (2008): 625 (catalogue; distribution: Australia).

Melobasina apicalis Kerremans *sensu* BELLAMY (2003): 120, Fig. 185 (mistake – in fact syntype of *simondsi*; list of genera).

Melobasina simondsi: BELLAMY (2008): 625 (catalogue; new combination; distribution incomplete: Oceania, Fiji: Viti Levu Island).

Type material examined. *Nesotrinchus australicus*: LECTOTYPE (designated here): ♀ (BMNH), ? AUSTRALIA: ‘Holo- type [p] [round, red-framed label] / Australie Tarnier. [h] / australicus Kerrem. Type. [h] / H. australicus Kerrem. Australie. [h] / Kerremans. 1903–59. [p]’. Since the number of syntypes is unknown, we designate here the female specimen from BMNH as the lectotype to avoid any confusion in the future.

Nesotrinchus simondsi: LECTOTYPE (designated here): 1 ♂ (BMNH), **FIJI: OVALAU ISLAND**: ‘Type [p] [round, red-framed label] / Syn- type [p] [round, blue-framed label] / Ovalau May/22 892 H.W.Simmonds. [h] Fiji. [p] / Pres. by Imp. Inst.Ent.Brit.Mus. 1931-283. [p] / Nesotrinchus n.g. Simondsi n.sp. Type [h] Det. D^r Obenberger [p] / Typus [p] [red label]’. PARALECTOTYPES (1 ♂ 2 ♀♀): **FIJI: OVALAU ISLAND**: ‘Syn- type [p] [round, blue-framed label] / 722 [p] v.1922. H.W.Simmonds. Ovalau [h] Fiji [p] May 22. H.W.S / Pres. by Imp. Inst.Ent.Brit.Mus. 1931-283. [p] / N. simondsi Obb. Paratype [h] det.K.G.Blair. [p] / Nesotrinchus australicus Kerr. [h] det.K.G.Blair. [p]’ (1 ♀, BMNH); ‘Fiji Islands. [p] Ovalau – v.1922 [h] H. W. Simmonds. [p] 892 [h] / Nesotrinchus n.g. Simondsi n.sp. Type [h] Det. D^r Obenberger [p] / Typus [p] [red label] / Mus. Nat. Pragae Inv. [p] 21992 [h] [orange label]’ (1 ♂, NMPC); same label data but ‘Nesotrinchus Simondsi m. Type’ and ‘Mus. Nat. Pragae Inv. [p] 21993 [h]’ (1 ♀, NMPC).

Other published records. **FIJI: VITI LEVU ISLAND**: Nukurua Forest, 60–130 m, 15.x.1979, forest, logged area, M, K, G. A. Kamah & S. L. Samuelson leg., Bishop Museum Acc. #1979. 260, 1 spec., BPBM (BELLAMY 1990, 2005; as *simondsi*).

Collecting circumstances. The specimen from Viti Levu Island was apparently collected in a logged forest area at 60–130 m a.s.l (BELLAMY 1990).

Remarks. We have studied the types of *Nesotrinchus australicus* and *N. simondsi* and failed to find any substantial differences. We regard them as conspecific, *N. simondsi* being a junior subjective synonym of *N. australicus*. The type locality of *N. australicus* ‘Australie’ is most probably an error.

Distribution. Pacific Islands: Fiji Islands (Ovalau Island, Viti Levu Island); ?Australia.

Nesotrinchus caeruleipennis (Fairmaire, 1877), comb. nov.

(Figs. 82–89, 110, 116–117)

Dicercomorpha caeruleipennis Fairmaire, 1877: 153. Type locality: Pacific Islands, Tonga: Vava’u Isl. (‘Vavao’).

Dicercomorpha caeruleipennis Fairmaire: FAIRMAIRE (1879): 97 (as *coeruleipennis* [sic!]; characters; distribution: Tonga: Vava’u Island (‘Vavao’); in Museum Godeffroy); FAIRMAIRE (1881): 265 (as *coeruleipennis* [sic!]; reference to THOMSON (1878) – mistake; with comment: probably identical with ‘*Buprestis wallisii* Montr.’; distribution: ‘à Samoa et aux îles Viti’); KERREMANS (1885): 135 (as *coeruleipennis* [sic!]; catalogue; as valid species; distribution: Vavao, Tonga; with a remark ‘= *Wallisi* Montr.? (sub nom. *Buprestis*?), Ann. Soc. Agr. Lyon, VII’); KERREMANS (1892): 85 (as *coeruleipennis* [sic!]; catalogue; as synonym of *wallisii*; reference to THOMSON (1878) – mistake; distribution: New Caledonia – mistake); KERREMANS (1900a): 294 (as *coeruleipennis* [sic!]; comment – as different from *wallisii*).

Haplotrinchus caeruleipennis (Fairmaire): KERREMANS (1903): 127 (as *coeruleipennis* [sic!]; new combination; list of species; as valid species; distribution: Fiji – mistake); OBENBERGER (1930): 358 (as *coeruleipennis* [sic!]; catalogue; as possible synonym of *caeruleipennis* Thomson; distribution: Fiji – mistake, Tonga: Vava’u Island).

Nesotrinchus caeruleipennis (Fairmaire): OBENBERGER (1936): 129 (as *coeruleipennis* [sic!]; comment; new combination; as possible form of *caeruleipennis* Thomson); THÉRY (1943): 644 (comment), 646 (list of species; as valid name; with synonyms *caeruleipennis* Thomson and *simondsi*; distribution: Fiji – mistake).

Nesotrinchus orientalis Bellamy, 1987: 101, **syn. nov.** Type locality: Pacific Islands, Tonga: Eua Island.

Nesotrinchus orientalis: BELLAMY (2003): 120, Fig. 186 (foto; list of genera).

Melobasina caeruleipennis (Fairmaire): BELLAMY (2008): 625 (catalogue; partim; as *coeruleipennis* [sic!]; distribution: Oceania, Tonga, erroneously Fiji: Ovalau Island, Vanua Levu Island, Viti Levu Island).

Melobasina orientalis: BELLAMY (2008): 625 (catalogue; distribution incomplete: Oceania, Tonga).

Misidentification. *Nesotrinchus caeruleipennis* (Fairmaire) sensu BELLAMY (1987, 1990, 2005, 2008) – in fact *N. caeruleipennis* (Thomson), is now *N. thomsoni* Bílý & Kubáň, nom. nov. (see below).

Type material examined. *Nesotrinchus caeruleipennis* (Fairmaire): LECTOTYPE (designated here): ♀ (BMNH), **TONGA: VAVA'U ISLAND:** 'Type [p] [round, red-framed label] / *Dicercomorpha caeruleipennis* Fairm. Vavau [h, Fairmaire's handwriting] / Fidji Fairm. [h, Kerremans handwriting] / *coeruleipennis* Fairmaire Type [h, Kerremans' handwriting] / *D. coeruleipennis* Fairmaire I. Fidji [h]'. Since the number of syntypes is unknown, we designate here the female specimen from BMNH as the lectotype to avoid any confusion in the future.

Nesotrinchus orientalis: PARATYPES: 7 ♂♂ 5 ♀♀ (BMNH), **TONGA: VAVA'U ISLAND:** 'Tonga: Vavau. iii.1925. G.H.E.Hopkins. [p] / Pres. by Imp. Inst. Ent. Brit. Mus. 1930–555. [p] / Paratype [p] *Nesotrinchus orientalis* ♂ [or ♀] [h] C. L. Bellamy [p] [yellow label]'; 3 ♂♂ 2 ♀♀ (BMNH), 'Neiafu Vavau Tonga 7. [or 8., or 11.] iii. [19]25 [h] S. Pacific Buxton & Hopkins [p] / Brit. Mus. 1928–25. [p] / Paratype [p] *Nesotrinchus orientalis* ♂ [or ♀] [h] C. L. Bellamy [p] [yellow label] / [one ♂ with: 'N. coeruleipennis Fairm. (var.) [h] det.K.G.Blair. [p]']; 1 ♂ (NMPC, ex coll. S. Bílý), 'Tonga, Vavau: Neiafu, 0–100m. March 1974 N.L.H. Krauss [p] / Paratype [p] *Nesotrinchus orientalis* ♂ [h] C. L. Bellamy [p] [yellow label]'.
Additional material examined. **TONGA: VAVA'U ISLAND:** 'Tonga: Vavau. iii.1925. G.H.E.Hopkins. [p] / *N. coeruleipennis* Fairm. [h] det. K.G.Blair. [p] / Pres. by Imp. Inst. Ent. Brit. Mus. 1930–555. [p] / *Nesotrinchus coeruleipennis* Frm. = *simondsi* Obb. (*Haplotrinchus*) [handwritten] A. Théry det. [p]' (1 ♂, NHMB, ex coll. G. Frey).

Other published records. Further type specimens of *N. orientalis* (BELLAMY 1987): **TONGA: EUA ISLAND:** Eua Isl., 21.viii.1975, J. C. Watt leg. (1 ♂ (holotype), 2 ♀♀, NZAC). **TONGATABU ISLAND:** Haamea, ii.1956, N. H. L. Krauss leg. (1 ♀). **VAVA'U ISLAND:** various data (24 spec.). **NEW ZEALAND: NIUE ISLAND:** various data (4 ♂♂ 4 ♀♀). Deposition of paratypes other than those from the Eua Island not specified.

Biology and collecting circumstances. Four paratypes of *Nesotrinchus orientalis* from Vava'u Island are labelled: 'Talau Forest, on tree trunks' and 13 paratypes from Vava'u Island (Neiafu) were collected at 0–100 m a.s.l. (BELLAMY 1987).

Remarks. We have studied the syntype of *N. caeruleipennis* (Fairmaire, 1877) and 21 paratypes of *N. orientalis* and failed to find any differences. We regard them as conspecific, *N. orientalis* being a junior subjective synonym of *N. caeruleipennis* (Fairmaire, 1877).

Distribution. Pacific Islands: Tonga (Eua Island, Tongatabu Island, Vava'u Island), Niue Island.

Nesotrinchus thomsoni Bílý & Kubáň, nom. nov.

(Figs. 90–97, 118)

Dicercomorpha caeruleipennis Thomson, 1878: 38 (junior primary homonym). Type locality: Pacific Islands, Fiji Islands ('Ins. Fidji', without exact data).

Dicercomorpha caeruleipennis Thomson: KERREMANS (1885): 134 (as *coeruleipennis* [sic!]; catalogue; as valid species; distribution: Fiji).

Haplotrinchus caeruleipennis (Thomson): THÉRY (1925): 225 (characters; comment; as valid species; synonym of *simondsi*; distribution: Fiji); OBENBERGER (1930): 358 (as *coeruleipennis* [sic!]; catalogue; as possible synonym of *caeruleipennis* Fairmaire; distribution: New Caledonia – mistake).

Nesotrinchus caeruleipennis (Thomson): OBENBERGER (1936): 129 (as *coeruleipennis* [sic!]; comment; new combination; as possible form of *caeruleipennis* Fairmaire); THÉRY (1943): 644 (with a comment: not in New Caledonia!); 646 (list of species; synonym of *caeruleipennis* Fairmaire and *simondsi*; distribution: Fiji); HOLYŃSKI (1993): 27 (note).

'*Nesotrinchus coeruleipennis* [sic!] (Fairmaire)' sensu BELLAMY (1987): 101 (misidentification; comment), 103–104, Figs. 2, 4, 6, 8, 10 (differential diagnosis).

'*Nesotrinchus coeruleipennis* [sic!] (Fairmaire)' sensu BELLAMY (1990): 91 (misidentification; comment; new locality: Fiji: Vanua Levu Island), 92 (differential diagnosis, compared with *simondsi* [now *australicus*]).

'*Melobasina caeruleipennis* [sic!] (Fairmaire)' sensu BELLAMY (2005): 5 (misidentification; catalogue; new localities: Fiji: Ovalau Island, Viti Levu Island; erroneously also Tonga).

'*Melobasina caeruleipennis* [sic!] (Fairmaire)' sensu BELLAMY (2008): 625 (partim; catalogue; misidentification; distribution Fiji: Ovalau Island, Vanua Levu Island, Viti Levu Island; erroneously also Tonga).

Melobasina caeruleipennis (Thomson): BELLAMY (2008): 625 (catalogue; as synonym of *caeruleipennis* Fairmaire).

Type material examined. LECTOTYPE (designated here): ♀ (MNHN), FIJI: 'Caeruleipennis (H. Deyr. M.s.s. Th. Type T. B. 38 I. Fidji / Th. Type [p] / Ex Musaeo James Thomson [p] / Type [p] [red label] / Muséum Paris 1952 Coll. R. Oberthür [p]'. Since the number of syntypes is unknown, we designate here the female specimen from MNHN as the lectotype to avoid any confusion in the future.

Additional material examined. FIJI: 'Fiji [h] / cum typo comparavit Sv. Bílý [p] 2006 [h]' (1 ♂, NMPC); 'Fidji. [h] / Collection de Bonneuil [p] / Chalybeipennis ED. I. Viti. [h] / caeruleipennis H.Deyr. = coeruleipennis Thoms. Fidji [h]' (1 ♀, BMNH); 'caeruleipennis (HDeyr.) [HDeyr. is scratched] Fairm. I. Fiji [h] / Ex Musaeo Mniszech [p] Muséum Paris 1952 Coll. R. Oberthür [p]' (1 ♀, MNHN).

Published records. FIJI: OVALAU ISLAND: no data, 1 spec., USPC (BELLAMY 2005, 2008; as *caeruleipennis* Fairmaire – misidentification). VANUA LEVU ISLAND: KOFUVULI, 26.x.1977, on *Agathis* log [Araucariaceae], BPBM, CLBC (BELLAMY 1990, 2005, 2008; as *caeruleipennis* Fairmaire – misidentification). VITI LEVU ISLAND: Colo-i-Suva, C.23, ex laulau leaves, 6.iii.1995, ex. resting on *Garcinia myrtifolia* leaves [Clusiaceae], 1 spec., FFDC; Navua Nakavu, ex. Mavota, 8.vi.1994, ex. *Gonystylus punctatus* [Thymelaeaceae], emerge in cage, 1 spec., FFDC (both data from BELLAMY 2005, 2008; as *caeruleipennis* Fairmaire – misidentification).

Biology and collecting circumstances. The specimen(s) from Vanua Levu were apparently collected on an *Agathis* log and the specimens from Viti Levu 'ex laulau leaves [...] ex. resting on *Garcinia myrtifolia* leaves' and 'ex. Mavota [= *Gonystylus*] [...] ex. *Gonystylus punctatus*, emerge in cage' (BELLAMY 1990, 2005, 2008).

Remarks. *Nesotrinchus caeruleipennis* (Thomson, 1878) is a junior primary and secondary homonym of *N. caeruleipennis* (Fairmaire, 1877). The taxon was erroneously treated as a junior subjective synonym of *N. caeruleipennis* (Fairmaire, 1877), moreover some authors (FAIRMAIRE 1881, KERREMANS 1892) mentioned the THOMSON's (1878) description only as a subsequent reference of *N. caeruleipennis* (Fairmaire, 1788). BELLAMY (1987) misidentified *N. caeruleipennis* (Fairmaire, 1877) and *N. caeruleipennis* (Thomson, 1878) and described specimens of the former species erroneously as *N. orientalis*. We propose a new substitute name, *N. thomsoni* Bílý & Kubáň, nom. nov., to remove the existing homonymy.

Distribution. Pacific Islands: Fiji Islands (Ovalau Island, Vanua Levu Island, Viti Levu Island).

Nesotrinchus wallisii (Montrouzier, 1855), comb. nov.

Buprestis wallisii Montrouzier, 1855: 11. Type locality: Pacific Islands, Wallis and Futuna (French): Wallis Island [not New Caledonia or Woodlark Island!].

Chalcophora wallisi [sic!]: GEMMINGER & HAROLD (1869): 1359 (catalogue; distribution: 'Polynesia').

'Doubtful genus' *wallisii*: SAUNDERS (1871): 137 (catalogue; genus doubtful; distribution: 'Polynesia').

Dicercomorpha wallisii: FAIRMAIRE (1881): 265 (comment – probably identical with '*coeruleipennis* [sic!] Fairm.' or '*coeruleipennis* [sic!] Thomson'; distribution: Wallis Island); KERREMANS (1885): 135 (as *wallisii* [sic!]; catalogue; as possible synonym of *caeruleipennis* Fairmaire); KERREMANS (1892): 85 (as *wallisii* [sic!]; catalogue; as valid name, with *caeruleipennis* Fairmaire and *caeruleipennis* Thomson as synonyms; distribution: New Caledonia – mistake); KERREMANS (1900a): 294 (as *wallisii* [sic!]; comment – different from *caeruleipennis* Fairmaire).

Paracupta wallisii: KERREMANS (1903): 84 (as *wallisii* [sic!]; list of species [see also *Haplotrinchus wallisii* in KERREMANS (1903: 127)]; distribution: 'Polynésie'); KERREMANS (1909): 109 (as *wallisii* [sic!]; in key as incertae sedis), 154 (as *wallisii* [sic!]; monograph; distribution: 'Iles Woodlark' – mistake); OBENBERGER (1926b): 125 (as *wallisii*

[sic!]; catalogue [see also *Haplotrinchus wallisii* in OBENBERGER (1930): 359]; distribution: Woodlark Island in Papua New Guinea – mistake); CARTER (1929): 301 (as *wallisii* [sic!]; list of species; distribution: Woodlark Island in Papua New Guinea – mistake).

Haplotrinchus wallisii: KERREMANS (1903): 127 (as *wallisii* [sic!]; list of species [see also *Paracupta wallisii* in KERREMANS (1903): 84]; distribution: New Caledonia – mistake); OBENBERGER (1930): 359 (as *wallisii* [sic!]; catalogue [see also *Paracupta wallisii* in OBENBERGER (1926b): 125]; distribution: New Caledonia – mistake); THÉRY (1943): 644 (as *wallisii* [sic!]; comment); BELLAMY (2008): 508 (catalogue; distribution: Woodlark Island in Papua New Guinea – mistake).

Remarks. We failed to find out any details on the type deposition but the type is not deposited in MNHN as mentioned by BELLAMY (2008). The type locality of *N. wallisii* is the Wallis Island and no other localities are known; its reported presence in New Caledonia and on the Woodlark Island (see above) is most probably an error. We agree with the concept of FAIRMAIRE (1881), who compared '*Buprestis wallisii* Montr.' with '*Dicercomorpha caeruleipennis* Fairm.', and attribute this species to the genus *Nesotrinchus*.

Distribution. Pacific Islands (French Wallis and Futuna: Wallis Island).

Genus *Ulaikoilia* Bílý & Kubáň, gen. nov.

(Figs. 103–109, 111–115, 122, 129)

Type species. *Ulaikoilia jelineki* Bílý & Kubáň, sp. nov. (present designation).

Diagnosis. Body medium-sized (9.8 mm), short, convex, metallic coloured, dorsal surface completely asetose (Fig. 103); ventral surface with microscopic, nearly indistinct pubescence; antennae and legs very short, anal ventrite of male strongly bispinose (Figs. 108, 112), elytra somewhat caudiform; hind wings with anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} joined distally (Fig. 122); visible abdominal tergites strongly transverse, terminal tergite (T7) wider than long (Fig. 129); tarsomeres with well-developed adhesive pads, body shape resembling that of *Lamprodila*, subgenus *Ovalisia*.

Description. Head relatively large, as wide as anterior pronotal margin, frontoclypeus deeply emarginate; frons flat with well-developed, sharp supraantennal carinae (Figs. 106, 109), vertex much wider than width of eye; sculpture of head consisting of sparse, large, simple punctures; eyes large, projecting beyond outline of head; antennae very short (Figs. 105, 114) barely reaching midlength of lateral pronotal margins when reclined, antennomeres 6–11 (in male) twice as wide as long.

Pronotum strongly convex, without lateroposterior depressions, bell-shaped, posterior margin with wide prescutellar lobe bearing small pit; pronotal sculpture consisting of sparse, simple punctures; lateral pronotal keels barely reaching midlength of pronotum, posterior pronotal angles sharp.

Scutellum twice as wide as long, obtusely pentagonal.

Elytra convex, slightly caudiform (Fig. 103), lateral margins sharply serrate in posterior half; apex of each elytron sharply trispinose, each elytron with 10 fine striae, composed of fine, simple, somewhat prolonged punctures, and shortened scutellar stria; humeral swellings small, basal transverse depression wide and deep, not reaching scutellum by far; epipleura missing, elytral margin with large, prominent subhumeral lobe (Figs. 104, 115).

Wing as in Fig. 122; anal veins $AA_4+AA_{3b}+AP_{1+2}$ and AP_{3a} joined distally.

Visible abdominal tergites strongly transverse, terminal tergite (T7) wider than long (Fig. 129).

Ventral surface with simple punctation, punctures finer and denser on ventrites than on remainder of ventral surface; anterior margin of prosternum slightly emarginate medially with deep, wide, transverse groove; prosternal process (Figs. 107, 111) wide and flat, with two deep grooves converging posteriorly; abdominal ventrites (except for anal ventrite) without lateral depressions, anal ventrite of male sharply bispinose (Figs. 108, 112).

Legs relatively short and stout, male metatibiae not modified; tarsal claws hook-shaped, slightly enlarged at base.

Aedeagus (Fig. 113) flattened, nearly subparallel, median lobe sharply pointed apically.

Etymology. The genus *Ulaikoilia* gen. nov. (feminine) is named in honour of Mr. Ulai Koil, one of the elder leaders of the Didipa clan from the Baitabag village and an outstanding natural history expert, whose leadership was instrumental in protecting rainforests on the lands of his Didipa clan as the Kau Wildlife area. His enthusiasm and expertise also guided the next generation of the Didipa clan in conservation and study of their rainforests.

Differential diagnosis. The genus *Ulaikoilia* gen. nov. belongs to the subtribe Nesotrinchina subtrib. nov. of the tribe Poecilonotini. Although *Ulaikoilia* gen. nov. resembles species of the subgenus *Ovalisia* of the genus *Lamprodila* by its body shape, the diagnostic characters (presence of well-developed supraantennal carinae, shape of pronotum, elytral apex and prosternal process, microsculpture of dorsal surface and missing elytral epipleura) show that it is related with the genus *Nesotrinchus*. The characters differentiating *Ulaikoilia* gen. nov. from related taxa are given in Table 1.

Ulaikoilia jelineki Bílý & Kubáň, sp. nov.

(Figs. 103–109, 111–115, 122, 129)

Type locality. Northeastern Papua New Guinea, Madang prov., Kau Wildlife area, Baitabag vill., 5°08'S 145°46'E, 50 m.

Type material. HOLOTYPE: ♂ (NMPC), 'Papua New Guinea, Madang prov., Kau Wildlife Area, Baitabag vill., 5°08'S 145°46'E, 50m, 1.–15.x.1999, local collector leg.'

Diagnosis. Medium-sized (9.8 mm), rather convex, stout, lustrous species (Fig. 103); both dorsal and ventral surface dark metallic blue, ventral surface with violet reflections; mandibles, antennae and tarsi black; dorsal surface completely asetose, ventral surface with extremely fine and sparse, nearly indistinct white pubescence; lateral grooves of prosternal process and basal portion of mesofemora with sparse white setae.

Description of the holotype (Fig. 103). Length 9.8 mm, width 3.8 mm.

Head large, as wide as anterior pronotal margin; frontoclypeus deeply emarginate, frons flat with sharp, well-developed supraantennal carinae (Figs. 106, 109), vertex flat, twice as wide as width of eye; eyes large, reniform projecting beyond outline of head; antennae (Figs. 105, 114) very short, reaching anterior third of lateral pronotal margins; scape short, pear-shaped, 1.6 times as long as wide, pedicel shortly cylindrical, about as long as wide; third antennomere slightly claviform, twice as long as wide; antennomere 4 triangular, 1.2 times as long

as wide, antennomere 5 obtusely triangular, as long as wide, antennomeres 6–11 trapezoid, twice as wide as long; sculpture of head consisting of sparse simple punctures, central portion of frons lustrous without punctation.

Pronotum strongly convex, 1.8 times as wide as long, bell-shaped, anterior pronotal margin straight, posterior margin bisinuose with wide medial prescutellar lobe; lateral margins slightly S-shaped, posterior pronotal angles sharp; anterior margin with fine groove present on each side and interrupted in middle, medial lobe of posterior margin with fine prescutellar pit; lateroposterior pronotal depressions missing; pronotal sculpture consisting of sparse, fine punctures on disc and in anterior angles and rough, dense punctures in posterior angles.

Scutellum slightly pentagonal, twice as wide as long.

Elytra convex, stout, 1.8 times as long as wide, slightly constricted at anterior third, widened at posterior third, with somewhat caudiform apices (Fig. 103); posterior half of elytral margins sharply serrate, apex of each elytron sharply trispinose, medial spine much closer to sutural spine than to outer spine; elytra distinctly striato-punctate, each elytron with small, shallow, irregular depression at posterior third; humeral swellings small but well-developed, basal transverse depression wide, deep but reaching only 3rd interstice; basal portion of 3rd interstice and apical portion of 4th interstice somewhat elevated; epipleura missing, subhumeral portion of elytral margin with large, prominent lobe (Figs. 104, 115).

Wing as in Fig. 122; anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} joined distally.

Visible abdominal tergites strongly transverse, terminal tergite (T7) wider than long (Fig. 129).

Ventral surface lustrous with rather dense, simple punctation, prosternal process with very sparse punctation; anterior margin of prosternum with deep and wide transverse groove, prosternal process (Figs. 107, 111) wide and flat, subparallel in anterior two thirds, strongly acuminate in posterior third with somewhat truncate apex and two deep oblique grooves composed of large punctures. Anal ventrite (Figs. 108, 112) strongly bispinose with deep lateral depressions.

Legs short and robust, protibiae slightly widened proximally, meso- and metatibiae somewhat enlarged at midlength; tarsi wide, much shorter than tibiae, with well-developed adhesive pads on tarsomeres 1–4. Tarsal claws sharp, hook-shaped, enlarged near base.

Aedeagus (Fig. 113) short and flat, slightly enlarged posteriorly, median lobe sharply pointed apically.

Female unknown.

Etymology. *Ulaikoilia jelineki* sp. nov. is named in honour of our friend and colleague Josef Jelínek, a leading specialist on the family Nitidulidae, on the occasion of his 70th birthday.

Biology and collecting circumstances. The type locality is in a mosaic of primary and secondary forests in the Kau Wildlife Area located about 15 km north of Madang in the foothills of the Adalbert mountain range. The holotype was collected by a local collector involved in the whole project; it was probably beaten from vegetation.

Distribution. So far known only from the type locality in the Madang province in the north-eastern part of Papua New Guinea.

Table 1. Character states of *Ulaikoilia* gen. nov., *Melobasina*, *Nesotrinchus* and *Lamprodila* (subgenera *Cinyrisia*, *Erialata*, *Mabomisia* and *Ovalisia*).

<i>Lamprodila</i>			
subgenus <i>Cinyrisia</i>	subgenus <i>Erialata</i>	subgenus <i>Mabomisia</i>	subgenus <i>Ovalisia</i>
Dorsal side dark bronze, elytra with small black interstitial spots (Fig. 44).	Dorsal side golden green, pronotum and elytra with large black spots (Fig. 40).	Dorsal side bright bronze, elytra with small black spots (Fig. 49).	Pronotum golden green or golden orange with black spots, elytra dark blue or violet (Fig. 98).
Body lustrous, elongate, navicular; elytra 2.0–2.2 times as long as wide, with fine apical serration, not caudiform, with short and indistinct pubescence (Figs. 44, 48).	Body matt, elongate, navicular; elytra 2.2 times as long as wide, with fine apical serration, not caudiform, with rather dense pubescence (Figs. 38, 40).	Body lustrous, elongate, navicular; elytra 2.2 times as long as wide with fine apical serration, not caudiform, with sparse and rather short lateral pubescence (Figs. 49, 53).	Body matt, stout and robust; elytra 1.7–1.8 times as long as wide, with very finely serrate and indistinctly caudiform apices, asetose (Figs. 98, 102).
Supraantennal carinae moderately developed, frons between them flat (Fig. 46).	Supraantennal carinae missing, frons flat (Figs. 36, 42).	Supraantennal carinae strongly developed, frons between them depressed (Fig. 51).	Supraantennal carinae strongly developed, frons between them depressed (Fig. 100).
Distal antennomeres of female 1.6 times as wide as long; male unknown.	Distal antennomeres of female as wide as long; male unknown.	Distal antennomeres of female 1.3 times as wide as long; male unknown.	Distal antennomeres of both sexes as wide as long.
Lateroposterior pronotal depressions poorly developed or missing (Fig. 44).	Lateroposterior pronotal depressions poorly developed or missing (Fig. 40).	Lateroposterior pronotal depressions poorly developed or missing (Fig. 49).	Lateroposterior pronotal depressions missing (Fig. 98).
Pronotum conical; pronotal sculpture not homogenous, consisting of irregular simple punctures (Fig. 44).	Pronotum slightly bell-shaped; pronotal sculpture homogenous, consisting of regular, pit-shaped punctures (Fig. 40).	Lateral pronotal margins subparallel in posterior half, nearly straight and converging in anterior half; pronotal sculpture consisting of irregular simple punctures (Fig. 49).	Pronotal margins widely rounded; pronotal sculpture very dense, nearly homogeneous, consisting of pit-shaped punctures; punctures on disc somewhat smaller and finer (Fig. 98).
Lateral pronotal carina well-developed, straight or slightly S-shaped (Fig. 45).	Lateral pronotal carina well-developed, straight or slightly S-shaped (Fig. 35).	Lateral pronotal carina well-developed, straight or slightly S-shaped (Fig. 50).	Lateral pronotal carina well-developed, straight or slightly S-shaped (Fig. 99).
Subhumeral elytral lobe poorly developed (Fig. 45).	Subhumeral elytral lobe poorly developed (Fig. 35).	Subhumeral elytral lobe poorly developed (Fig. 50).	Subhumeral elytral lobe less developed, not prominent (Fig. 99).
Wings: anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free.	Wings: anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free.	Wings: anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free.	Wings: anal veins $AA_4 + AA_{3b} + AP_{1+2}$ and AP_{3a} free (Fig. 124).
Anal ventrite with sharp lateral longitudinal carinae and wide, prolonged depressions between them and lateral margin of ventrite (Fig. 48).	Anal ventrite without longitudinal carinae and lateral depressions (Figs. 38, 39, 43).	Anal ventrite without longitudinal carinae and lateral depressions (Fig. 53).	Anal ventrite without longitudinal carinae and lateral depressions (Fig. 102).
Anal ventrite of female bispinose (Fig. 48).	Anal ventrite of female bispinose (Fig. 38) or (in some specimens) bispinose with poorly marked medial lobe (Figs. 39, 43).	Anal ventrite of female bispinose with obtuse medial lobe (Fig. 53).	Anal ventrite of female bispinose (Fig. 102).
Aedeagus: male unknown.	Aedeagus: male unknown.	Aedeagus: male unknown.	Aedeagus short, subparallel.
Distribution			
Peninsular Malaysia, Borneo, Sumatra	Peninsular Malaysia, Borneo	Borneo	Indonesia: Maluku: Morotai, Halmahera, Papua; Papua New Guinea: Kepulauan Aru, Tagula Is.; Solomon Islands: Russel Is.

<i>Melobasina</i>	<i>Nesotrinchus</i>	<i>Ulaicoilia</i> gen. nov.
Dorsal side purple, or elytra with violet apical third, or whole body dark blue with golden-orange lateral sides and golden-green sutural stripe (Figs. 54, 59–60, 65–66).	Dorsal side black with blue lustre or elytra dark violet and pronotum golden orange (Figs. 71, 82, 90).	Dorsal side deep blue (Fig. 103).
Body lustrous, navicular; elytra twice as long as wide, with rather rough apical serration, not caudiform, asetose (Figs. 54, 58, 59, 66).	Body lustrous, slender, navicular to wedge-shaped; elytra 2.1–2.4 times as long as wide, strongly serrate and strongly caudiform, asetose (Figs. 71, 82, 90, 97).	Body lustrous, stout and robust, elytra only 1.8 times as long as wide, slightly caudiform and roughly serrate posteriorly, asetose (Fig. 103, 108).
Supraantennal carinae missing, antennal sockets margined only by sharp but not elevated edge, frons flat (Figs. 56, 62, 68).	Supraantennal carinae strong, frons between them strongly depressed (Figs. 75, 86, 94, 110).	Supraantennal carinae strong, frons between them flat (Figs. 106, 109).
Distal antennomeres of female 1.0–1.2 times as wide as long; male unknown.	Distal antennomeres of both sexes 1.0–1.2 times as wide as long.	Distal antennomeres of male twice as wide as long (Figs. 105, 114); female unknown.
Lateroposterior pronotal depressions well developed (Figs. 54, 59, 60, 65, 66).	Lateroposterior pronotal depressions well developed (Figs. 71, 77, 82, 90).	Lateroposterior pronotal depressions missing (Fig. 103).
Lateral pronotal margins nearly straight and moderately converging anteriorly, or slightly bell-shaped; pronotal sculpture consisting of rough and dense punctures laterally and fine and sparse punctures medially (Figs. 54, 59, 60, 65, 66).	Pronotum conical or slightly bell-shaped with prominent knoll at midlength of lateral margin; pronotal sculpture irregular, consisting of simple punctures, medial portion of pronotum with very sparse punctation (Figs. 71, 82, 90).	Pronotum widely bell-shaped; pronotal sculpture consisting of sparse simple punctures, punctation somewhat finer and sparser on disc (Fig. 103).
Lateral pronotal carina poorly developed, obtuse (Figs. 61, 67).	Lateral pronotal carina missing, lateral margin somewhat swollen (Figs. 73, 84, 92).	Lateral pronotal carina poorly developed, obtuse (Fig. 104).
Subhumeral elytral lobe less developed, not prominent (Figs. 61, 67).	Subhumeral elytral lobe less developed, not prominent (Figs. 72, 83, 91, 116).	Subhumeral elytral lobe large, strongly developed (Figs. 104, 115).
Wings: anal veins $AA_4+AA_{3b}+AP_{1+2}$ and AP_{3a} free (Fig. 123).	Wings: anal veins $AA_4+AA_{3b}+AP_{1+2}$ and AP_{3a} free (Fig. 121).	Wings: anal veins $AA_4+AA_{3b}+AP_{1+2}$ and AP_{3a} fused distally (Fig. 122).
Abdominal tergites moderately transverse, terminal tergite (T7) longer than wide (Fig. 128).	Abdominal tergites moderately transverse, terminal tergite (T7) longer than wide (Fig. 127).	Abdominal tergites strongly transverse, terminal tergite (T7) wider than long (Fig. 129).
Anal ventrite without longitudinal carinae and lateral depressions; only lateral spines of ventrite with obtuse carinae (Figs. 58, 64, 70).	Anal ventrite without longitudinal carinae and lateral depressions; only lateral spines of ventrite with obtuse carinae (Figs. 76, 81, 87, 89, 95, 97).	Anal ventrite with obtuse lateral, longitudinal carinae and with well-developed depressions between them and lateral margins of ventrite (Figs. 108, 112).
Anal ventrite of female sharply trispinose (Figs. 58, 64) or bispinose with small medial tooth (Fig. 70).	Anal ventrite of female sharply trispinose (Figs. 76, 87, 95).	Female unknown.
Aedeagus: male unknown.	Aedeagus long, spindle-shaped (Figs. 117–119).	Aedeagus short, slightly enlarged posteriorly (Fig. 113).
Distribution		
Papua New Guinea (eastern islands): Tagula and Bougainville Isls.; Solomon Islands: Guadalcanal, Malaita and Shortland Isls.	Fiji, Tonga, Niue, Wallis Islands	Northeastern Papua New Guinea

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