

NMRLipids project

Open Collaboration to understand lipid systems in atomistic resolution

- We have mainly focused to find models that correctly capture lipid headgroup conformational ensemble and ion binding
- CHARMM36 has best conformational ensembles, but not correct^{1,4}. However, it captures many essential differences between PC, PE, PG and PS headgroups^{4b}.
- Cation binding affinity to PC, PG and PS headgroups can be improved by electronic continuum correction (ECC)^{5,6}.
- How about other lipids and ions? Membrane proteins? Other properties than headgroup? etc.

¹ <http://dx.doi.org/10.1021/acs.jpcb.5b04878> (NMRLipids I, 2015)

² <http://dx.doi.org/10.1039/C6CP04883H> (NMRLipids II, 2016)

³ <https://github.com/NMRLipids/NmrLipidsCholXray/blob/master/MANUSCRIPT/manuscript.pdf> (NMRLipids III, in progress)

⁴ <https://pubs.acs.org/doi/10.1021/acs.jpcb.9b06091> (NMRLipids IV, 2019)

^{4b} <https://github.com/NMRLipids/NMRLipidsIVPEandPG/blob/master/Manuscript/manuscriptPGPE.pdf> (NMRLipids IVb, in progress)

⁵ <http://dx.doi.org/10.1021/acs.jpcb.7b12510>

⁶ <http://dx.doi.org/10.1021/acs.jctc.9b00824>

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Some relevant lipid headgroups and their mixtures

	PC	PE	PG	PS	chol	CL	GM1	PIP	PA	DAG
PC	X ^{1,2}	X ^{4b}	X ^{4b}	X ⁴	X ³					
PE		X ^{4b}								
PG			X ^{4b}							
PS				X ⁴						
chol										
CL										
GM1										
PIP										
PA										
DAG										

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NMRlipids databank

Quality evaluated atomistic resolution MD simulations of biologically relevant lipid mixtures

- User/contributor feeds information of the data (location etc.)
- Databank **automatically** indexes the data and makes quality evaluation
- Databank content can be openly accessed and analysed

[illegible]

NMRlipids databank

expected applications

- Force field evaluation: *What is the best force field for my application?*
- Reference simulations: *For example, reference pure bilayer simulations for membrane-protein interaction studies.*
- Analysis of bilayer properties from large datasets: *For example, calculate P-N vector angle from all available PC and PG simulations.*
- Exercise and example for sharing simulation data: *“PDB” for simulations?*

NMRlipids databank

general properties

- Overlay databank: *NMRlipids databank contains indexed links to the data. The actual MD simulation data is currently in Zenodo, but could be in any stable location.*
- Quality evaluation: *NMRlipids databank contains a quality evaluation protocol that is applied to all contributed datasets. Also the quality evaluation results are also stored in the databank.*
- Analysis of the data: *NMRlipids databank enables flexible analysis of the content.*

NMRLipids databank

Current status

- Prototype:

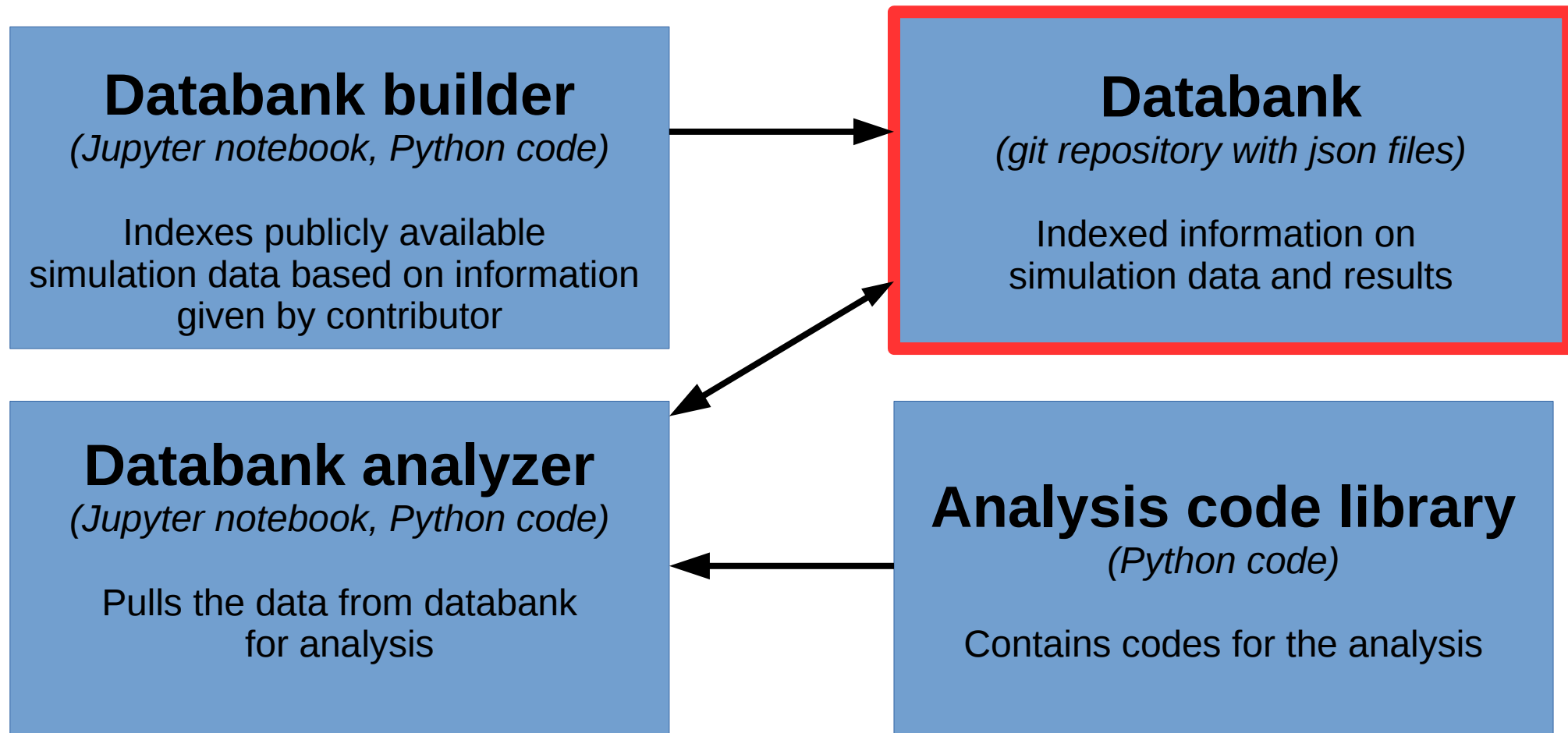
www.nmrlipids.fi

https://github.com/NMRLipids/MATCH/tree/master/Data/Lipid_Bilayers

- Indexing with folder names (not very flexible), adding data and analysis not very well documented
- New indexing system designed in the NMRLipids19 workshop in Berlin is now being implemented (currently mainly by Anne Kiirikki)

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New structure



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Databank builder

Current version (jupyter notebook):

<https://github.com/NMRLipids/NMRLipidsIVPEandPG/blob/master/scripts/AddData.ipynb>

- User gives information in the first shell:
 - DOI
 - def file for order parameter calculation
 - System name, software, force field name, source, and date
 - File names (xtc, tpr, gro or corresponding information)
 - Names of the molecules
- Program reads from tpr/gro file:
 - Amount of each molecule
 - Temperature
- This information is written into a json file that is stored into the databank

NMRLipids databank

New structure

Demo version with few datasets (json files, git):

<https://github.com/NMRLipids/NMRLipidsIVPEandPG/tree/master/Data/Simulations>

- Information from Databank Builder stored in dictionaries in json files.
For example see:

<https://github.com/NMRLipids/NMRLipidsIVPEandPG/blob/master/Data/Simulations/0d5/d1d/0d5d1dcb43e775faf4e53c4f9ff255a67481bd38/9b487701b24d3fad83991e311188b08d3d5ea768/README.json>

- Folder structure (see <https://github.com/NMRLipids/NMRLipidsVIpolarizableFFs/issues/3>):
'/asd/fgh/topology_checksum/trajectory_checksum/README.json',
where 'asd' and 'fgh' are the 1-3rd and 4-6th letters of the topology checksum.

NMRLipids databank

Databank analyzer

Current version (jupyter notebook):

<https://github.com/NMRLipids/NMRLipidsIVPEandPG/blob/master/scripts/readDATA.ipynb>

- Goes through the *Databank* and reads the dictionary files
- The data with selected lipids and atoms can be then analyzed calling functions in the *Analysis code library*

Urgent issues

- What should we store in the dictionary (readme.json)?
 - File names and locations, Force field and software, number of molecules, temperature
 - Something else?
- We need unique names for lipid molecules and the corresponding names in the simulation given by the user
 - CURRENT IDEA: We have a list of default names for molecules. If these do not correspond the simulation, contributor has to correct.
- We need unique naming convention for atoms within the molecules
 - For now, we are planning to use the mapping file (<http://nmrlipids.blogspot.com/2015/03/mapping-scheme-for-lipid-atom-names-for.html>)
 - Should we go more towards Sundaralingam notation?
 - Or should go for SMIRKS or similar ([dx.doi.org/10.26434/chemrxiv.8304578.v1](https://doi.org/10.26434/chemrxiv.8304578.v1))?

Other issues

- Temperature, pressure, thermostat, etc. parameters cannot be read using Mdanalysis or Mdtraj. For Gromacs, the solution is to use gmx dump, but other programs I do not know.
- In which extend we should save simulation parameters (thermostats, timestep, PME settings, etc.) in the dictionary?
- What properties will be automatically analyzed? Order parameters, Form factors, all dihedrals angles, something else?
- Which results will be stored in the databank? Only automatically analyzed or all?