Bio-hybrid implant, next generation of bio-engineered implant – A Review.

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Abstract

The lack of periodontal ligament (PDL) support in the implant bone junction is a matter of concern for implant treatment. The direct contact between bone to implant transmit forces directly to bone and reduces its functional efficiency. Several modifications in implant design and implant surface have been tried and tested. Recently bio-engineering and regenerative dentistry has thrown new lights in the modification of dental implant and to eliminate these drawbacks. Stem cell incorporated bio engineered implant also known as bio- hybrid implant has the potential to generate PDL and nerve cells along the implant bone junction. Pluripotent cells present on the bio-hybrid implant surface also have the ability of bone remodeling, wound healing and neuro transmission. This review article highlights recent status in the development of bio-hybrid implants along with its function and advantages.

Keywords: Bio-hybrid implant, ligaplants, bio-engineered PDL, stem cells, regenerative dentistry.

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Introduction

The history of the evolution of dental implants is a rich and fascinating travelogue through time. With advancement of technologies the modern implant treatment has also taken a quantum leap from inserting shell, gold tube to modern era oseointegrated titanium implants.

While dental implants are increasingly becoming the choice of replacement for missing teeth, the complications associated with them are progressively emerging too. Oseointegration is one the key factor for successful outcome of implant treatment is affected by several local and systemic factors.^[1] То overcome complication associated with oseointigration several modifications are made that can osseointegrate to bone faster and with a stronger bone-to-implant interface.^[2] Along with physical and chemical modification implant surface use of bio mimetic compound is also getting popular as a way to

increase bio-functionalization of dental implants.^[3]

In spite of all these modifications the absence of periodontal ligament (PDL) attachment around the implant hinders its biological function and efficiency.^[4] The periodontal ligament in natural teeth which acts as a viscoelastic "shock absorber" diminishes the magnitude of the occlusal force and distributes stress to the surrounding bone. Absence of PDL reduces the mobility of the implant and directly transfers the stress to the bone implant interface.^[5] Absence of proprioceptive fibers in PDL also reduces patients' functional efficiency.

With the advancement of regenerative dentistry, a novel fibrous tooth implant connection using dental follicle stem cells, bioengineered PDL has been developed to address these issues. While ligaplants implants using bioengineered PDL was introduced much earlier, bio-hybrid implant with stem cell technology is newer advancement of this field.

This review provides an insight into available techniques and biomaterials used in biohybrid implants with emphasis on the most recent outcomes and future avenues.

Current trends in Bio-engineered PDL Implant

Buser et al. (1990) 1st conceptualized bioengineered PDL and demonstrated PDL of the roots served as a source of cells which could populate the titanium implant surface during healing.^[6] Gault et al. in 2010 described the development and clinical application of a tissue-engineered ligament known as "ligaplants".^[7] In this process PDL cells obtained from teeth are prepared and cultivated in the bioreactor. Then single layer of these cells secured with biodegradable scaffolds are deposited around the titanium coated implant.^[8]

Histologic examination has shown soft-tissue space with dimension and structure similar to PDL space with abundant connective tissue fibers perpendicular to implant surface of ligaplants. Evidence of formation of osteocementum and neo vascularization around bone implant junction also been documented.^[9]

Apart from all these advantages ligaplants has shown high degree of implant rejection, limited mobility and no response to noxious stimulus.^[8,9]

Bio- hybrid implant

Bio- hybrid implants are implantable electronic arrays which harness stem cells. These implants not only stimulate the growth of damaged or missing tissue but also capable of interacting with nervous tissue and restore neural function (Figure 1).^[10] Bio-hybrid eye for the treatment of visual impairment patients and bio- hybrid cochlear implants for hearing loss is quite popular now a day. Osseointegrated bio-hybrid implants not only support mechanical loading but also connected to surrounding via connective tissue which maintains flexibility, mobility.^[11] Till now no experimental studies has been conducted on human but animal studies have shown promising results with generation of PDL fibers and nerve interface between bone and implant.

Oshima M et a.l (2014) developed 1st bio hybrid dental implant.^[11] They used HA coated implant enveloped with embryonic day (ED) 18.5 tooth germ-derived dental follicle tissue (ED18.5-DF) and transplanted into an adult mouse. Nakajima K et al. (2016) used HA & platinum coated titanium implant and enveloped it with rat PDL cells to develop a bio-hybrid implant.^[12] Lee D-J et al. (2017) used HA coated titanium implant and enveloped it with different cell type like immortalized human periodontal ligament (ihPDLs), immortalized human cells cementoblasts (ihCEMs), human umbilical vein endothelial cells (HUVECs) in multilayer formation.^[13]

A. Stem cell used in bio-hybrid implants:

Stem cells used in bio-hybrid implants can be obtained from extra oral tissues or oral tissues such as craniofacial bone, dental pulp, PDL, dental follicle, apical papilla, oral mucosa, gingiva, and periosteum.^[14] The dental stem cells (DSCs) are post-natal stem cells that have mesenchymal stem cells (MSC) like properties including the self-renewal and multilineage differentiation potential.^[15]

1. Dental pulp stem cells (DPSCs) These are multi potential cells with typical fibroblast-like morphology and dentinogenic, osteogenic, neurogenic, chondrogenic, and myogenic differentiation

potential.^[16,17] The human-DPSC cultures contain multipotent neural crest stem cell (NCSC) which can differentiate into neural crest cells . These cells express several bio markers like STRO-1, CD146, OCT4 which are mesenchymal and bone-marrow stem cell markers.^[15]

2. Periodontal ligament stem cells(PDLSCs)

These cells too show morphology similar to fibroblasts and can be differentiated into cementoblasts and osteoblasts cells.^[15] The rate of proliferation of these cells are higher than DPSCs. STRO-1, CD146, and tendon-specific transcription factor are the biological cues expressed by these cells.^[18]

3. Stem cells from human exfoliated deciduous teeth (SHED)

This mesenchymal progenitor stem cell was isolated by Muira et al (2003) from exfoliated deciduous teeth.^[19] The rate of proliferation and CFU of SHED is higher than DPSCs with early expression of biological markers like STRO-1 and CD146.^[15]

4. Tooth germ progenitor cells (TGPCs)

These are dental mesenchymal cells obtained from 3rd molar. TGPC express different bio markers like STRO-1 and CDs. They help in the expression of pluripotency-associated genes like nanog, oct4, sox2.^[20] multilineage TGPCs have differentiation capacity to differentiate into adipocytes. osteoblasts/odontoblasts, osteocytes, and neurons.^[15]

5. Dental follicle progenitor cells (DFPCs)

These cells are ectomesenchymal in origin and extracted from follicle of human third

molars.^[15] These cells express several bio markars like Notch1, STRO-1, and nestin.^[21] STRO-1-positive dentalfollicle stem cells (DFSCs) can differentiate into cementoblasts and can form cementum.^[22]

6. Gingival mesenchymal stem cells (GMSCs)

GMSCs obtained from gingival tissue and retro molar tissue exhibit clonogenicity, self-renewal, and multipotent differentiation capacity, and immunomodulatory properties.^[15,23] GMSCs express CDs and display positive signals for Oct4, Sox2, Nanog, Nestin, SSEA-4, and Stro-1. The main features of this type of cells are tissue regeneration and wound healing caused by increase telomerase activity.^[24]

7. Alveolar bone-derived mesenchymal stem cells (ABMSCs)

Matsubara et al. (2005) isolated these spindle shaped fibroblasts like stem cells.^[25] Similar to other stem cells these cells also have osteoblastic, chondrogenic and adipogenic differentiation potentials.^[15] These cells express the surface markers CD73, CD90, CD105, STRO-1and high level of alkaline phosphatase.^[5,15]

8. Adipose-derived stem cells (ADSCs) ADSCs have the same properties that of DSCs but the only advantage of these cells are they can be in higher amount easily.²⁶ They can be differentiate into chondrocytes, osteocytes, or myocytes and can be used areas with boney defect.^[27]

Other than these cells other verities of stem cells are also there like stem cells from the apical part of the human dental papilla (SCAP), Bone marrow-derived mesenchymal stem cells (BMSCs), Bloodderived stem cells.^[17]

B. Structure of bio-hybrid implant:

Implants functionalized with bio-engineered PDL and stem cells made up of 3 components – implant body, scaffold and stem cells. The function of the scaffold around the implant is to harbor stem cells and to regulate the action of the bio markers. Different materials used as scaffolds are as followed-

1. Natural Polymers-

3D-printed natural polymers like chitosan, collagen most commonly used as scaffolds.

2. Synthetic Polymers-

Different synthetic bio-degradable polymers like poly(ethylene glycol)b-poly(l-lactide-co-"-caprolactone) (PELCL), poly(lactic-co-glycolic) acid (PLGA), poly(caprolactone-coethyl ethylene phosphate) (PCLEEP) are used for fabrication of scaffolds. The verifying porosity (i.e., pore size of 150–500 nm) of these polymers helps to control release of cells and biological cues.^[28]

3. Bio-ceramics-

Hydroxyapatite nanoparticles, injectable mesoporous silica nanoparticles, bioactive glass nanoclusters are often used with polymers to fabricate scaffolds.²⁸

C. Fabrication of bio-hybrid implant

- Stem cells are extirpated from desired sites and are cultured in different culture medium like (Dulbecco's minimum Eagle's medium with 10% foetal bovine serum and 1% penicillin/streptomycin solution, Minimum essential medium alpha, endothelial growth media) with the help of bio degradable scaffold for stipulated days.^[11,13]
- 2. After harvesting different stem cells, cell sheets are detached from cultured medium. Each isolated tissues were wrapped around HA coated implant surface of approximately 50 mm in diameter.^[11]
- 3. Then these implants are placed onto a cell culture insert for 24 hours before transplantation. (Figure 2)

Discussion:

The main difference between the bio-hybrid implant and the ligaplants is that three dimensional cultured stem cells are used in bio-hybrid implants over two dimensional PDL cells from extracted teeth. The biohybrid implant not only helps to grow cementum, PDL, alveolar bone around implant but also helps to protect the implant from noxious stimuli generated by occlusal forces. Pluripotent stem cells in the cell sheet of bio-hybrid implant surface contains Hertwig's epithelial root sheath and epithelial cell rests of Malassez which secrets Wnt and regulates formation of signaling cementoblasts.^[29] The newly formed nerve cells at implant bone junction create microTENNS like structures. Neural bodies of the nerve cells are situated at one end of collagen hydrogel scaffold structure and their axonal projections grow unidirectionally toward the target tissue which forms a neural network that could act as a replacement for CNS reconstruction (Figure 3). [30,31] Oshima M et al. (2014) and Nakajima K et al. (2016) detected Anti-neurofilament (NF)immunoreactive nerve fibers and expression of c-Fos immunoreactive neuron in the PDL. The level of expression of these factors is equal to natural tooth which can be observed following noxious stimulation following 2 after orthodontic treatment.^[11,12] hours Functional analysis of bio-hybrid implant shows movement of implant similar to natural teeth with successful bone remodeling through generation of Colony-stimulating factor-1 (Csf-1) and osteocalcin (Ocn) on the compression and tension sides respectively.^[13]

More over bio-hybrid implants act as a reservoir of immune cells which helps to bone remodeling and regeneration of bone defect.^[11] Bio-hybrid implants provides platform onto which cells are seeded and also molecular factors incorporate (growth factors, guidance molecules, VEGF-alpha, insulin-like growth factor 1 etc) to stimulate and guide outgrowth of cells. These factors together known as secretomes enhance the migration, proliferation, and expression of osteogenic marker genes like RUNX2, SMAD5, RANKL.^[32] Whether fixed to the implant surface or delivered via microfluidic channels, these molecular factors offer an over traditional advantage cell transplantation by facilitating integration of cells to the intended targets.^[10] Tissue nonspecific alkaline phosphatase (TN-ALP), ectonucleotide

pyrophosphatase/phosphodiesterase 1 (ENNP1), integrin binding sialoprotein are among such factors which regulates phosphate level and mineralization of hard tissues.^[11]

These factors also work in epigenetic mechanism which includes DNA methylation and histone modifications. During osteoblastic differentiation of stem

cells increased methylation of the promoter of LIN28 facilitates osteogenesis by downregulation of Lin28 gene.^[33] DNA methyltransferases secreted by stem cells molecules are responsible for DNA methylation. This causes more condensed DNA structure, transcriptional repression, and gene silencing and regulates genes responsible for bone metabolism and expression of several factors like alkaline phosphatase, sclerosin, osteoprotegerin.^[19] Histone acetyl transferase, histone deacetylases are the enzymes responsible for histone modification which in turn causes osteoblast differentiation influenced by the action of osteocalcin and osteocalcin promoters.^[34]

Conclusion:

The concept of bioengineered PDL generated by stem cell has the potential to restore missing natural teeth using oral implants. This review provides an overview regarding different aspect of bio-hybrid implants. However, several questions like strength and longevity of the PDL attachment, functional efficiency of connective tissue is still not answered. Further human trials and long term clinical trials are needed to determine the scope of bio-hybrid implants.

References:

- 1. Gómez-de Diego R. Indications and contraindications of dental implants in medically compromised patients: update. Medicina oral, patologia oral y cirugia bucal. 2014 Sep;19(5):e483.
- Ting M, Jefferies SR, Xia W, Engqvist H, Suzuki JB. Classification and Effects of Implant Surface Modification on the Bone: Human Cell–Based In Vitro Studies. Journal of Oral Implantology. 2017 Feb;43(1):58-83.
- 3. Lutz R, Srour S, Nonhoff J, Weisel T, Damien CJ, Schlegel KA.

Biofunctionalization of titanium implants with a biomimetic active peptide (P-15) promotes early osseointegration. Clinical oral implants research. 2010 Jul;21(7):726-34.

- Bathla N, Sailo JL, Kapoor N, Nagpal A, Gupta R, Singla A. Ligaplants, the nextgeneration prosthodontic implants: A comprehensive review. Indian Journal of Dental Sciences. 2021 Apr 1;13(2):146-49.
- Shimono M, Ishikawa T, Ishikawa H, Matsuzaki H, Hashimoto S, Muramatsu T, Shima K, Matsuzaka KI, Inoue T. Regulatory mechanisms of periodontal regeneration. Microscopy research and technique. 2003 Apr 1;60(5):491-502.
- Saleem M, Kaushik M, Ghai A, Tomar N, Singh S. Ligaplants: A revolutionary concept in implant dentistry. Annals of Maxillofacial Surgery. 2020 Jan;10(1):195-99.
- Kiong AL, Arjunkumar R. Tissueengineered ligament: Implant constructs for tooth replacement (Ligaplants). Journal of Pharmaceutical Sciences and Research. 2014 Mar 1;6(3):158-62.
- Garg H, Deepa D. Bioengineered periodontal ligament: Ligaplants, a new dimension in the field of implant dentistry–Mini review. Journal of Oral Research and Review. 2018 Jul 1;10(2):92.
- Mathew A, Babu AS, Keepanasseril A. Biomimetic properties of engineered periodontal ligament/cementum in dental implants. Contemporary Clinical Dentistry. 2020 Oct 1;11(4):301.
- Rochford AE, Carnicer-Lombarte A, Curto VF, Malliaras GG, Barone DG. When bio meets technology: biohybrid neural interfaces. Advanced Materials. 2020 Apr;32(15):1903182.
- 11. Oshima M, Inoue K, Nakajima K, Tachikawa T, Yamazaki H, Isobe T, Sugawara A, Ogawa M, Tanaka C, Saito

M, Kasugai S. Functional tooth restoration by next-generation bio-hybrid implant as a bio-hybrid artificial organ replacement therapy. Scientific reports. 2014 Aug 13;4(1):1-0.

- Nakajima, K., Oshima, M., Yamamoto, N., Tanaka, C., Koitabashi, R., Inoue, T. and Tsuji, T., 2016. Development of a functional biohybrid implant formed from periodontal tissue utilizing bioengineering technology. Tissue Engineering .Part A, 22(17-18):.1108-15.
- Lee DJ, Lee JM, Kim EJ, Takata T, Abiko Y, Okano T, Green DW, Shimono M, Jung HS. Bio-implant as a novel restoration for tooth loss. Scientific reports. 2017 Aug 7;7(1):1-0.
- 14. Komada Y, Yamane T, Kadota D, Isono K, Takakura N, Hayashi SI, Yamazaki H. Origins and properties of dental, thymic, and bone marrow mesenchymal cells and their stem cells. PloS one. 2012 Nov 21;7(11):e46436.
- Chalisserry EP, Nam SY, Park SH, Anil S. Therapeutic potential of dental stem cells. Journal of tissue engineering. 2017 May 20;8:2041731417702531.
- Ikeda E, Tsuji T. Growing bioengineered teeth from single cells: potential for dental regenerative medicine. Expert opinion on biological therapy. 2008 Jun 1;8(6):735-44.
- 17. Hilkens P, Gervois P, Fanton Y, Vanormelingen J, Martens W, Struys T, Politis C, Lambrichts I, Bronckaers A. Effect of isolation methodology on stem cell properties and multilineage differentiation potential of human dental pulp stem cells. Cell and tissue research. 2013 Jul;353(1):65-78.
- Gay IC, Chen S, MacDougall M. Isolation and characterization of multipotent human periodontal ligament stem cells. Orthodontics & craniofacial research. 2007 Aug;10(3):149-60.

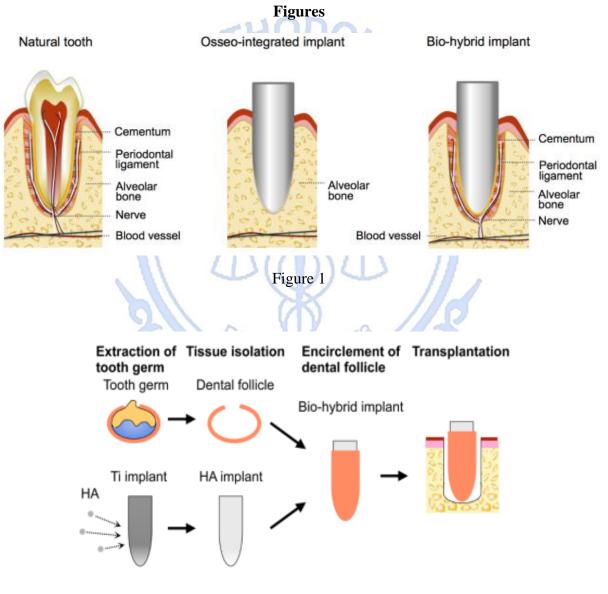
- 19. Miura M, Gronthos S, Zhao M, Lu B, Fisher LW, Robey PG, Shi S. SHED: stem cells from human exfoliated deciduous teeth. Proceedings of the National Academy of Sciences. 2003 May 13;100(10):5807-12.
- 20. Guven EP, Yalvac ME, Kayahan MB, Sunay H, SahIn F, Bayirli G. Human tooth germ stem cell response to calciumsilicate based endodontic cements. Journal of Applied Oral Science. 2013 Aug;21(4):351-7.
- 21. Morsczeck C, Götz W, Schierholz J, Zeilhofer F, Kühn U, Möhl C, Sippel C, Hoffmann KH. Isolation of precursor cells (PCs) from human dental follicle of wisdom teeth. Matrix biology. 2005 Apr 1;24(2):155-65.
- 22. Kémoun, P., Laurencin-Dalicieux, S., Rue, J., Farges, J.C., Gennero, I., Conte-F., Briand-Mesange, Auriol. F., Gadelorge, M., Arzate, H., Narayanan, A.S. and Brunel, G., 2007. Human dental follicle cells acquire cementoblast features under stimulation by BMP-2/-7 and enamel matrix derivatives (EMD) in and vitro. Cell tissue research. 329(2):283-94.
- 23. Zhang Q, Shi S, Liu Y, Uyanne J, Shi Y, derived from human gingiva are capable of immunomodulatory functions and ameliorate inflammation-related tissue destruction in experimental colitis. The Journal of Immunology. 2009 Dec 15:183(12):7787-98.
- 24. Tomar, G.B., Srivastava, R.K., Gupta, N., Barhanpurkar, A.P., Pote, S.T., Jhaveri, H.M., Mishra, G.C. and Wani, M.R., 2010. Human gingiva-derived mesenchymal stem cells are superior to bone marrow-derived mesenchymal stem cells for cell therapy in regenerative medicine. Biochemical and biophysical research communications, 393(3):377-83.

- 25. Matsubara T, Suardita K, Ishii M, Sugiyama M, Igarashi A, Oda R, Nishimura M, Saito M, Nakagawa K, Yamanaka K, Miyazaki K. Alveolar bone marrow as a cell source for regenerative medicine: differences between alveolar and iliac bone marrow stromal cells. Journal of Bone and Mineral Research. 2005 Mar;20(3):399-409.
- 26. Bressan E, Botticelli D, Sivolella S, Bengazi F, Guazzo R, Sbricoli L, Ricci S, Ferroni L, Gardin C, Velez JU, Zavan B. Adipose-derived stem cells as a tool for dental implant osseointegration: An experimental study in the dog. International journal of molecular and cellular medicine. 2015;4(4):197-203.
- 27. Baena RR, Rizzo S, Graziano A, Lupi SM. Bone regeneration in implant dentistry: Role of mesenchymal stem cells. A Textbook of Advanced Oral and Maxillofacial Surgery: Volume 3. 2016 Aug 31:3269.
- 28. Asa'ad F, Pelanyte G, Philip J, Dahlin C, Larsson L. The Role of Epigenetic Functionalization of Implants and Biomaterials in Osseointegration and Regeneration—A Bone Review. Molecules. 2020 Jan;25(24):5879-89.
- Shi S, Le AD. Mesenchymal stem cells 29. Iwasaki, K., Washio, K., Meinzer, W., Tsumanuma, Y., Yano, K. and Ishikawa, I., 2019. Application of cell-sheet engineering for new formation of cementum around dental implants. Heliyon, 5(6):1991-98.
 - 30. Struzyna LA, Harris JP, Katiyar KS, Chen HI, Cullen DK. Restoring nervous system structure and function using tissue engineered living scaffolds. Neural regeneration 2015 research. May;10(5):679-85.
 - 31. Adewole DO, Serruya MD, Wolf JA, Cullen DK. Bioactive neuroelectronic interfaces. Frontiers in neuroscience. 2019 Mar 29;13:269.

- Vrtačnik P, Marc J, Ostanek B. Epigenetic mechanisms in bone. Clinical Chemistry and Laboratory Medicine. 2014 May 1;52(5):589-608.
- 33. Dansranjavin T, Krehl S, Mueller T, Mueller LP, Schmoll HJ, Dammann RH. The role of promoter CpG methylation in the epigenetic control of stem cell related

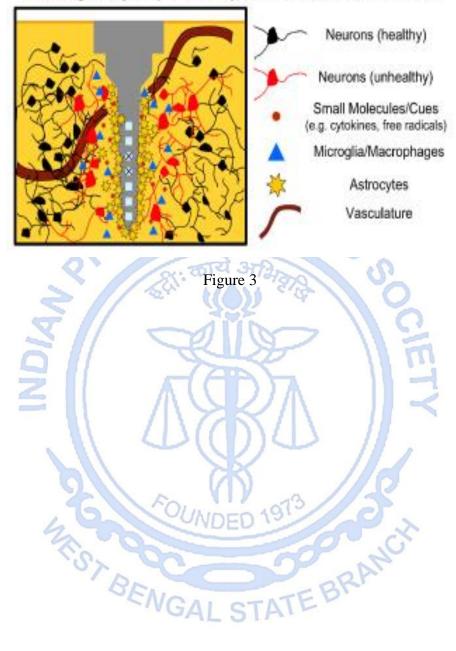
genes during differentiation. Cell cycle. 2009 Mar 15;8(6):916-24.

34. Kang JS, Alliston T, Delston R, Derynck R. Repression of Runx2 function by TGF-β through recruitment of class II histone deacetylases by Smad3. The EMBO journal. 2005 Jul 20;24(14):2543-55.





Bio-hybrid implant



The Foreign Body Response to Implanted Neuroelectronic Interfaces