



RCSI

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to better health

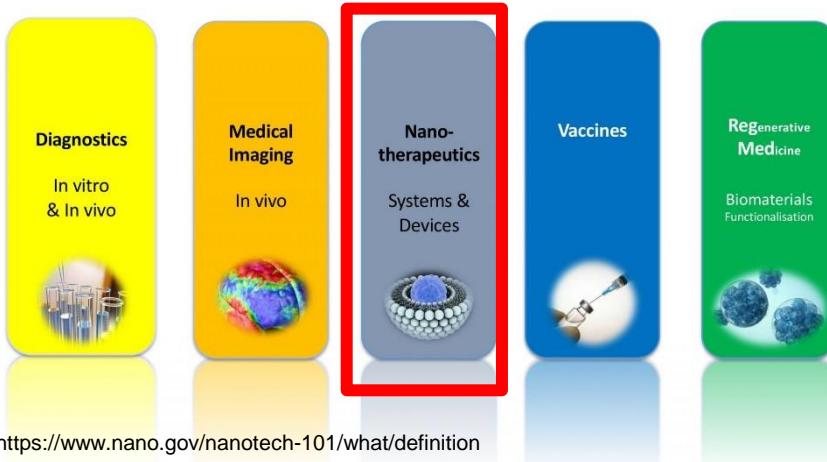
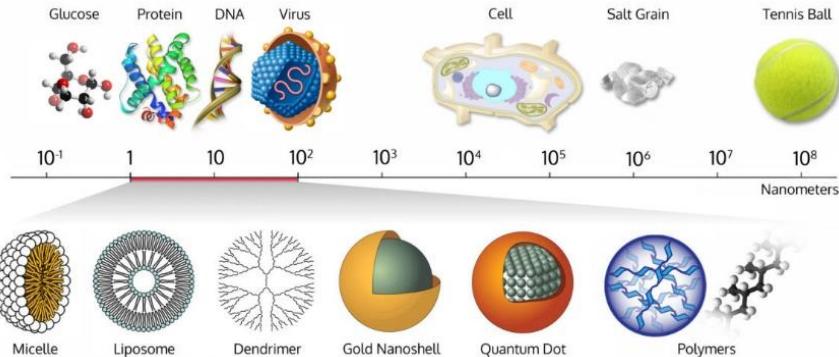
Nanoparticles via SET-LR-PISA

D. V. Tomasino
Supervisor A. Heise



Nanomedicine

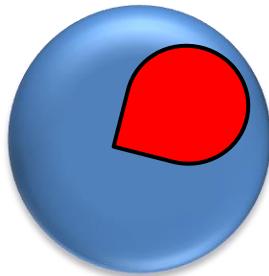
“Nanotechnology is the science conducted at the nanoscale, about 1 to 100 nm”



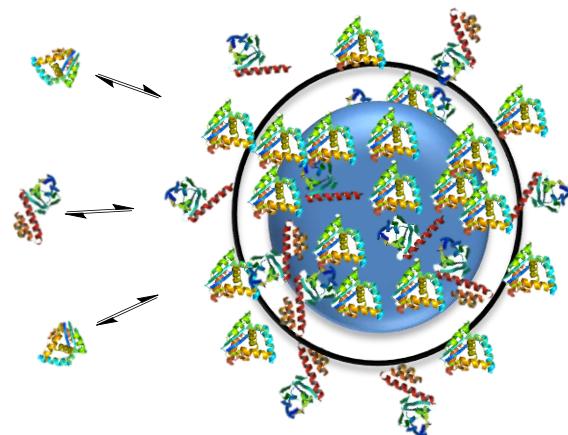
“Nanomedicine is the application of nanotechnology for medical purposes”

NPs in biological fluid

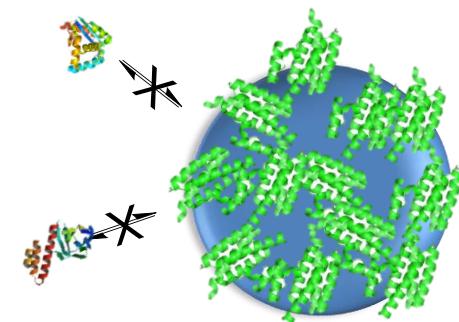
Polymeric NPs



Biological environment



Surface modification



Classified in terms of

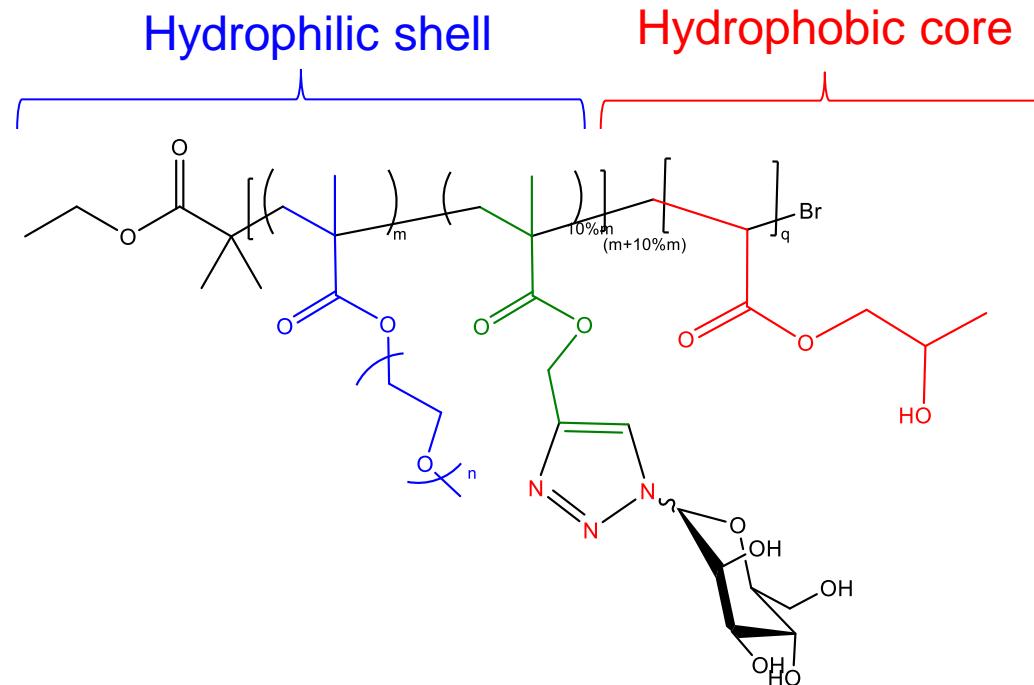
- Type of polymers
- Surface functionalisation
- morphology

- Formation of protein corona layer.
- New biological identity.
- Poor biodistribution.

- Creation of artificial corona layer.
- **Glycopolymers.**

Project goal

1. Develop a facile procedure for the synthesis of a glycosylated polymer nanoparticle using block copolymer self-assembly, specifically PISA technology.
2. Test the biological interaction of glycosylated polymer nanoparticles.



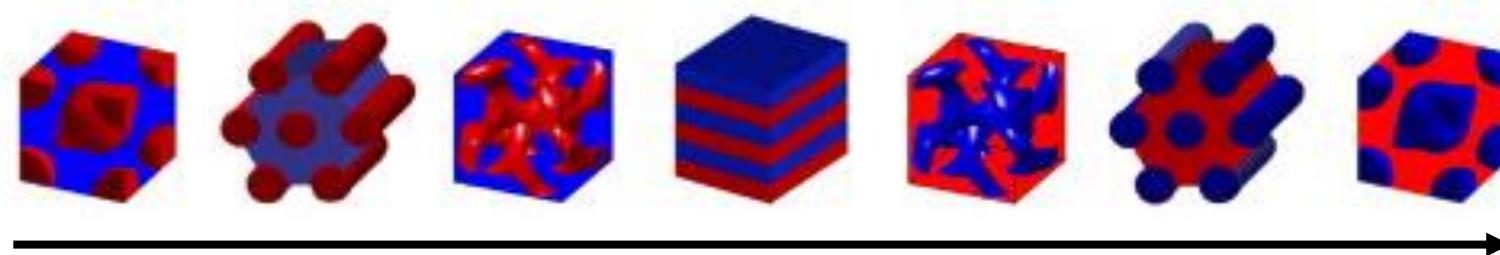
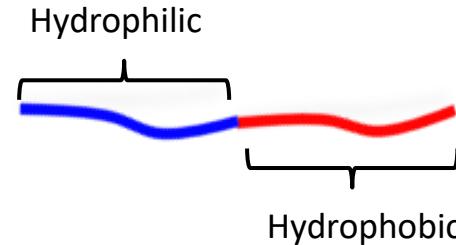
Amphiphilic block copolymer

Amphiphilic block copolymers comprise of discrete immiscible hydrophobic and hydrophilic blocks which undergo phase separation in aqueous conditions.

Microphase separation promoted by;

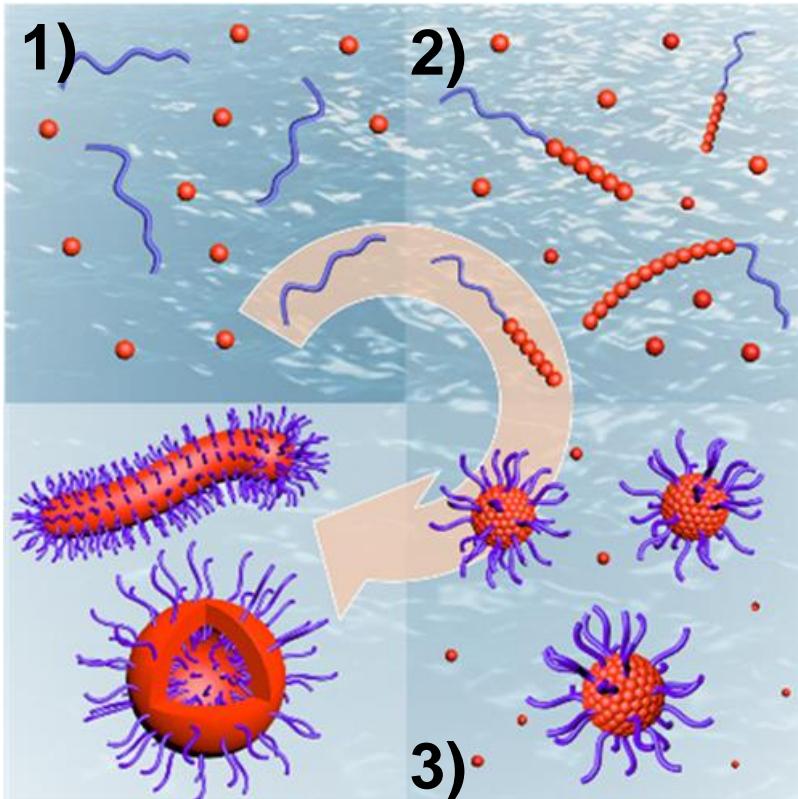
Hydrophilic repulsion → increase hydrophilic interactions with water

Hydrophobic attractions → decrease hydrocarbon water interface



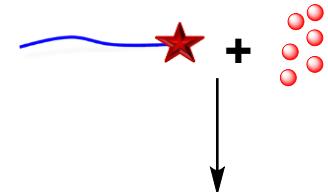
$$f_{red}$$

NPs synthesis by Polymerisation-induced self-assembly (PISA)

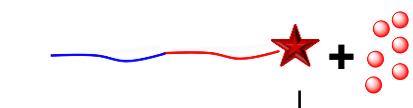


Carried out *via* controlled living radical polymerisation (i.e. RAFT, ATRP).

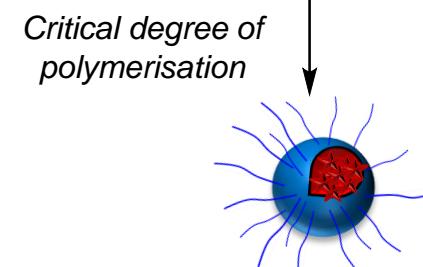
1) Synthesis of macro initiator (first block).



2) Chain extension.



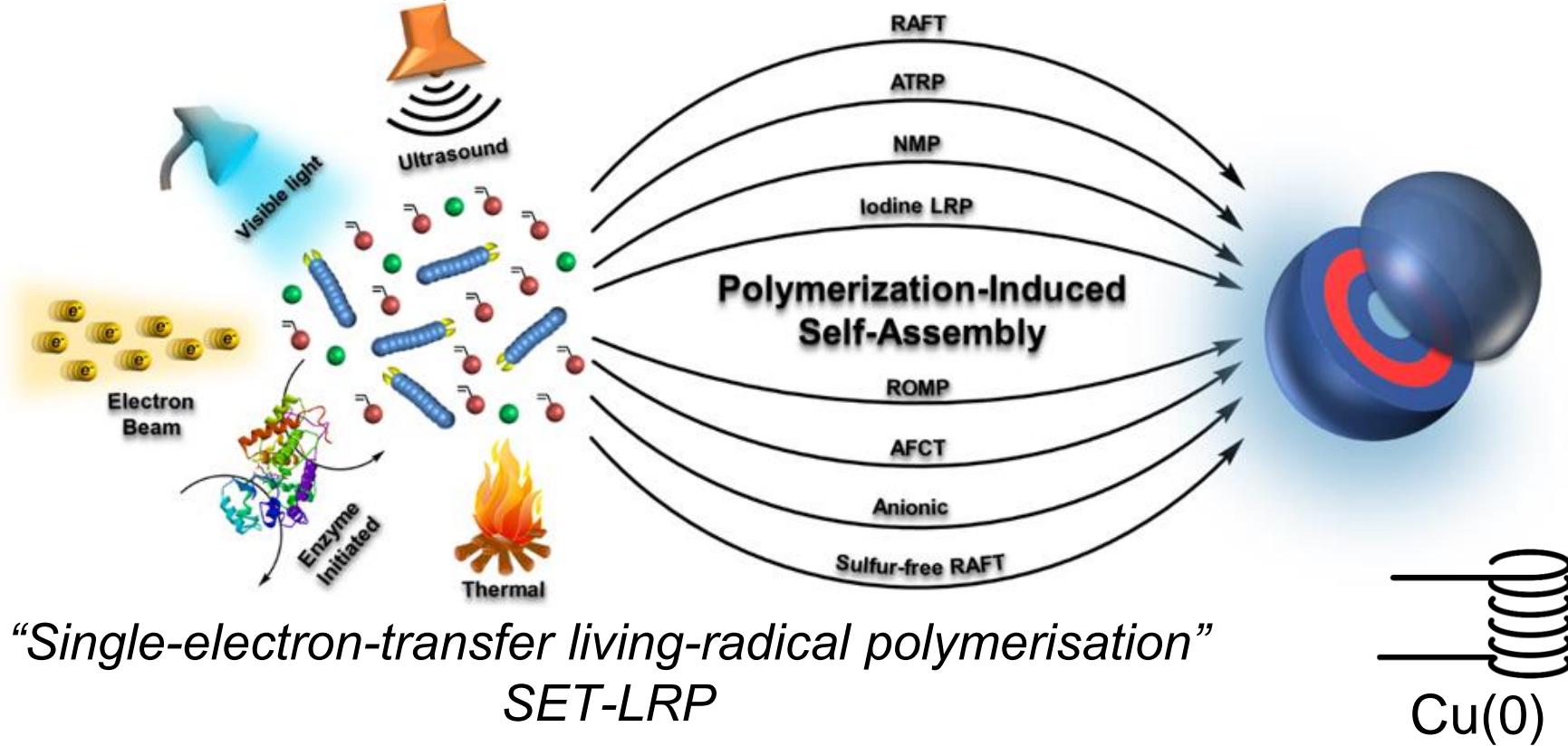
3) *In situ* self-assemble.



- Up to 50 w/w% solid concentration.
- Good candidate for industrial scale up.
- Nano-object of controlled size and morphology without further processing steps.

Polymerisation methods for PISA

All controlled radical polymerization methods suitable

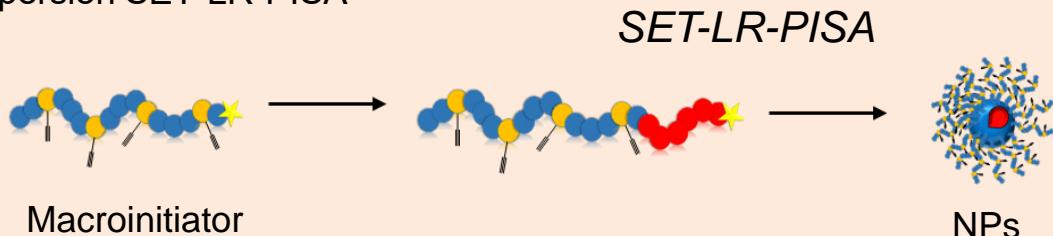


Project stages

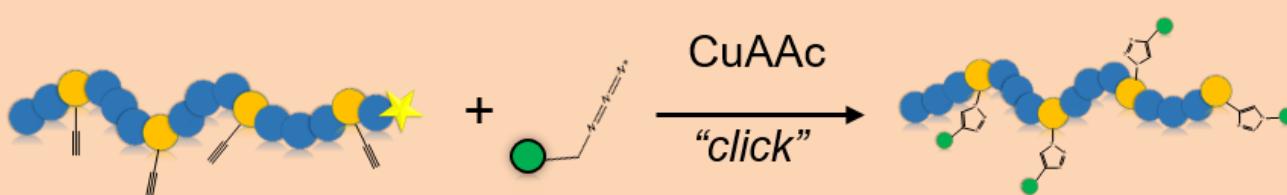
A) Preparation of NPs *via* aqueous dispersion SET-LR-PISA

Morphology evaluation:

- Core forming monomer concentration $[M]_0$
- Solvent

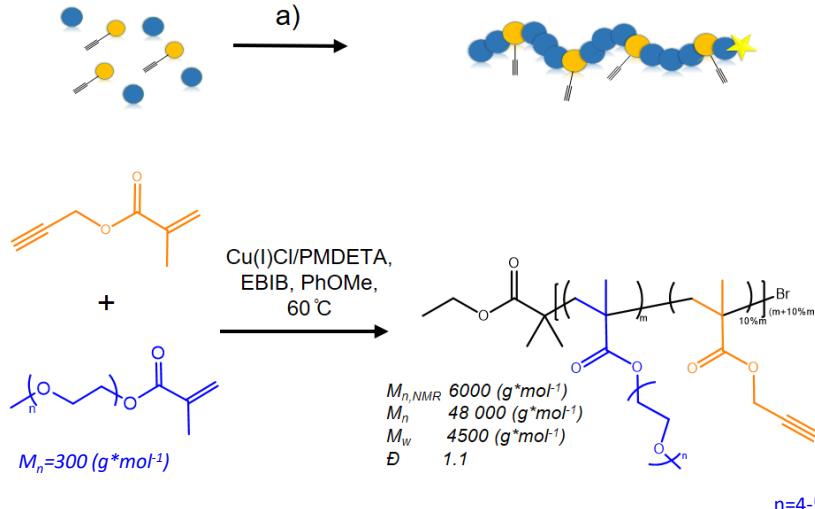


B) Macroinitiator functionalisation with mannosides via alkyne-azide copper catalysed cycloaddition:



Optimisation of NPs via aqueous dispersion SET-LR-PISA

SYNTHESIS MACROINITIATOR

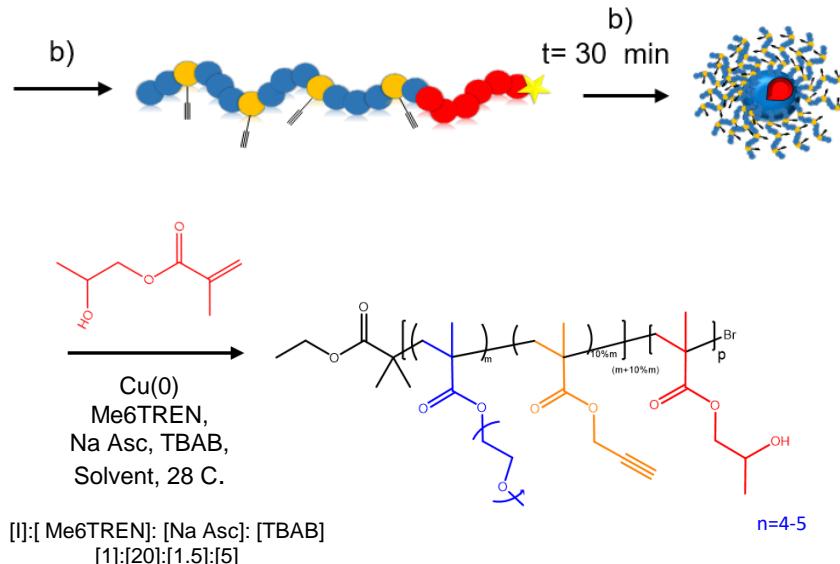


a) Conventional ATRP

- $\text{DP}_{\text{PEGMA}} = 20$
- $\text{DP}_{\text{PgMA}} = 5$



SET-LR-PISA



b) SET-LR-PISA

- $\text{DP}_{\text{HPMA}} = 150; 300$
- Solid content (SC)=10%

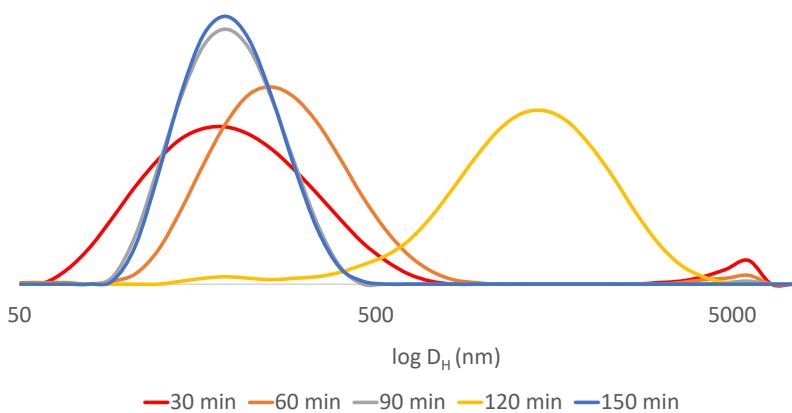
Followed by: DLS, TEM

Parameters:

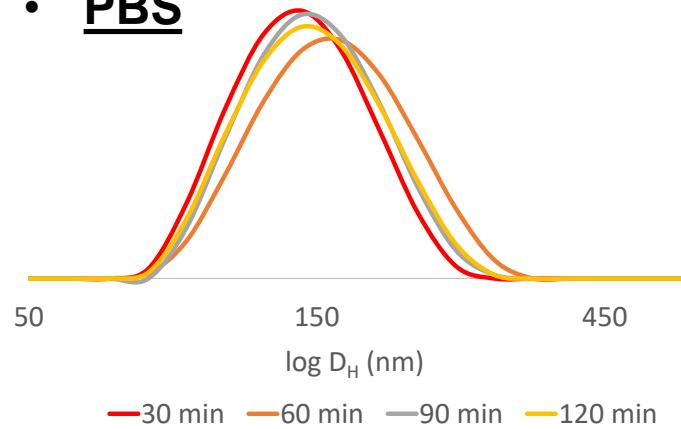
- Solvent (H_2O vs. PBS buffer)
- Monomer concentration

Influence of solvent: DP_{HPMA}=150 SC=10%

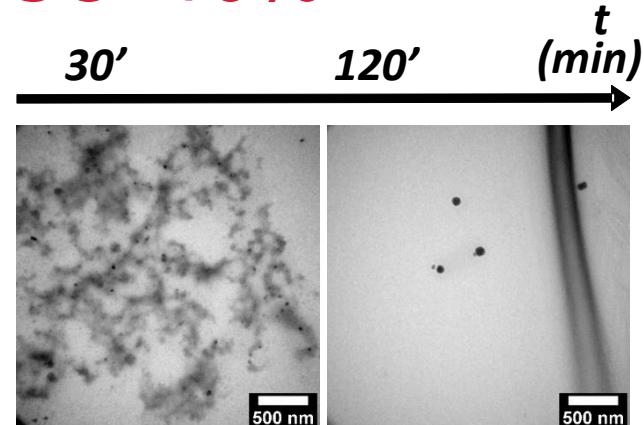
- H₂O



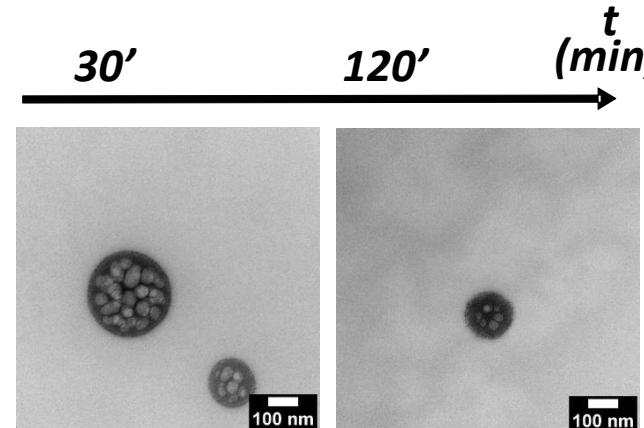
- PBS



Time (min)	D _H (nm)	PDI
30	186.6	0.33
60	254.2	0.20
90	185.6	0.11
120	1100	0.27
150	188.8	0.14

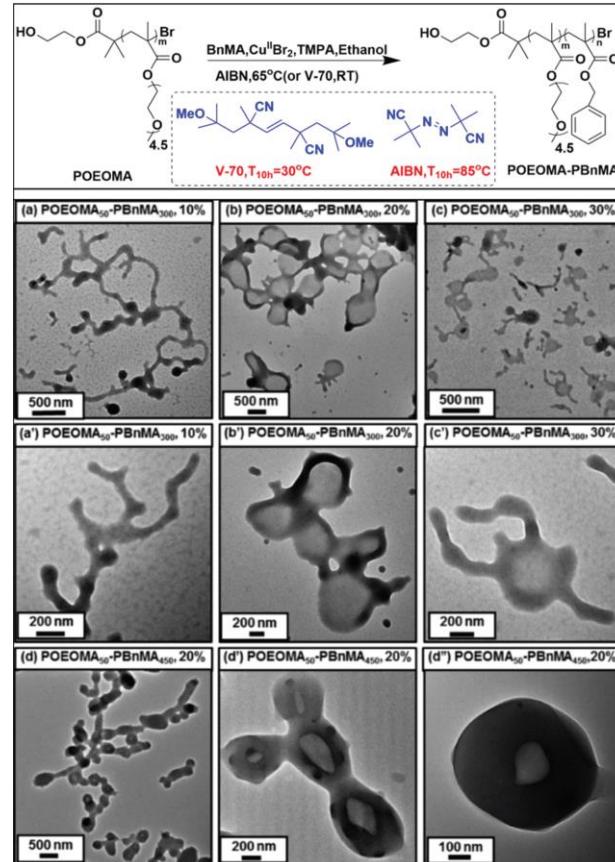
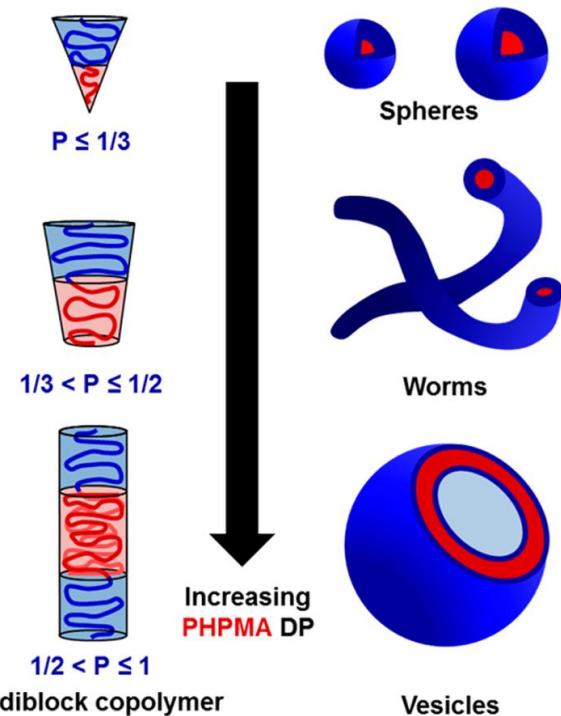


Time (min)	D _H (nm)	PDI
30	136.9	0.04
60	153.2	0.06
90	143.1	0.08
120	142.3	0.06



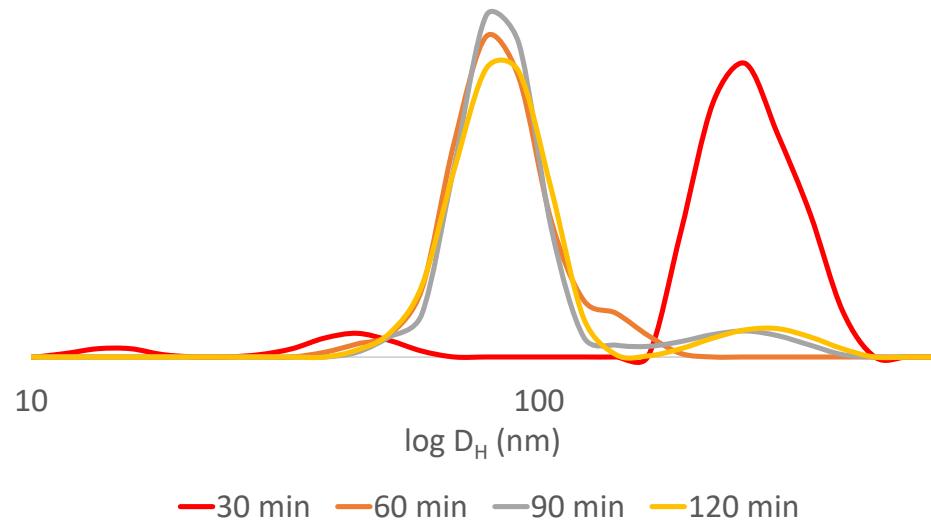
Influence of monomer concentration

- Traditional morphology evolution



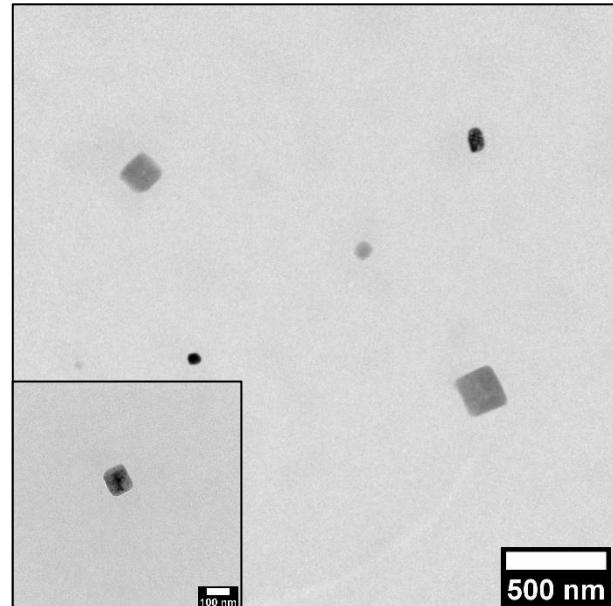
Influence of monomer concentration

- H_2O
- $\text{DP}_{\text{HPMA}} = 300$
- $\text{SC} = 10\%$



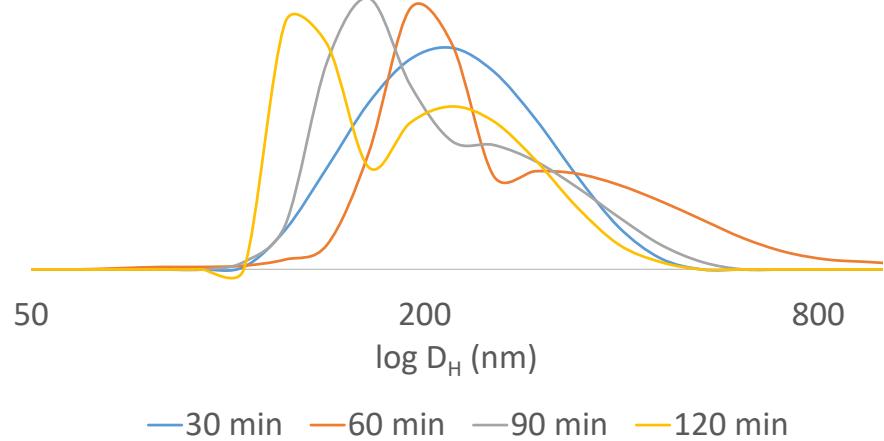
Time (min)	D_H (nm)	PDI
30	565.1	0.53
60	398.4	0.50
90	400.7	0.51
120	409.2	0.45

- 120 min

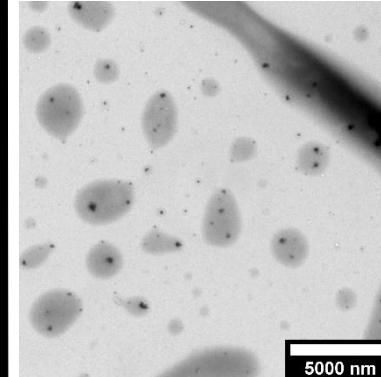


Influence of monomer concentration

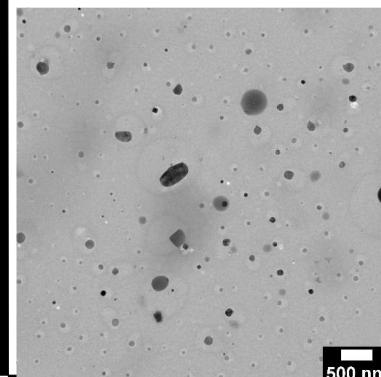
- PBS
- $\text{DP}_{\text{HPMA}} = 300$
- SC=10%



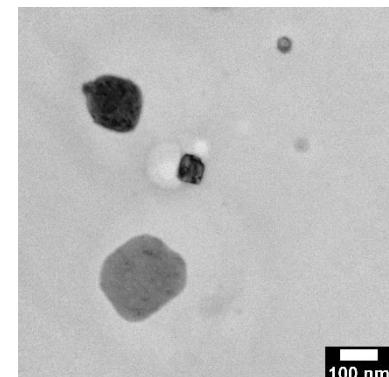
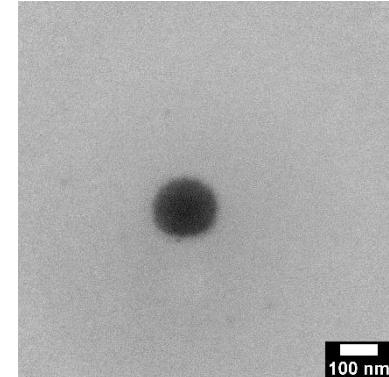
30'



120'



t
(min)

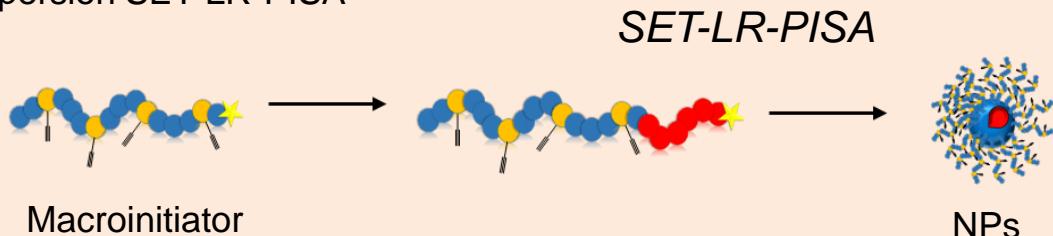


Project stages

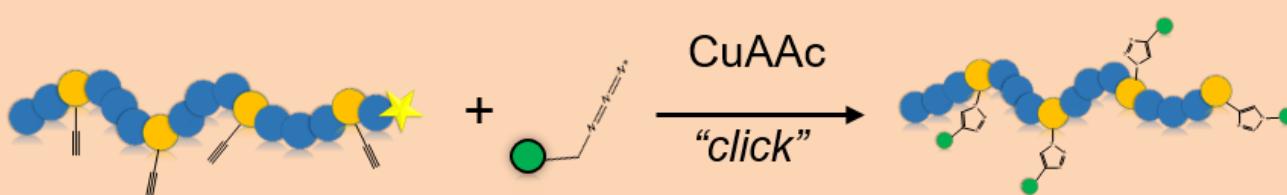
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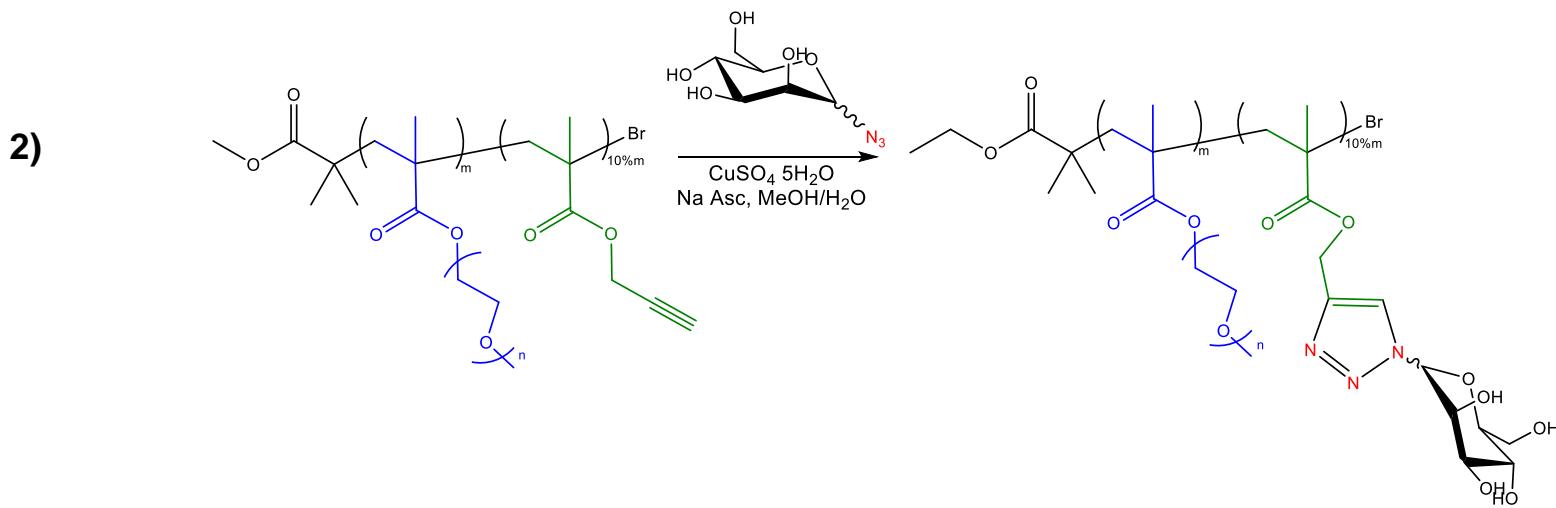
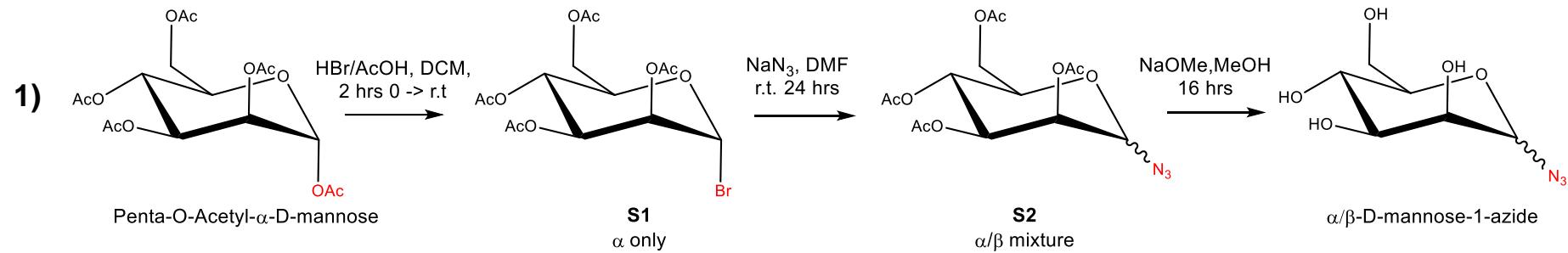
- Core forming monomer concentration $[M]_0$
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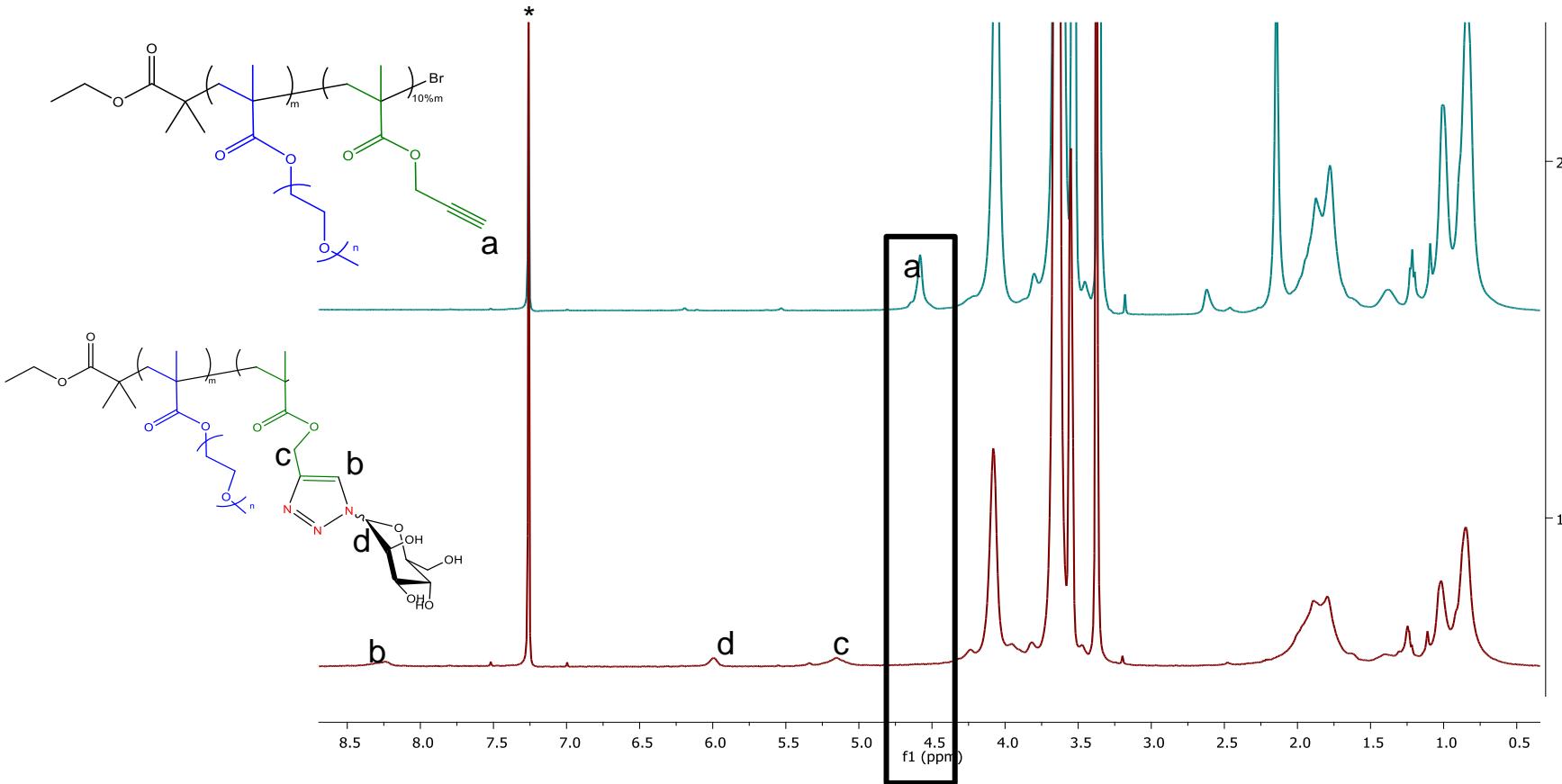
B) Macroinitiator functionalisation with mannosides via alkyne-azide copper catalysed cycloaddition:



Glycosylated macroinitiator for SET-LR-PISA system

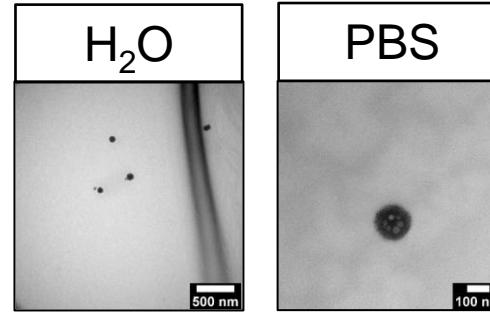


Glycosylated macroinitiator: racemic mixture



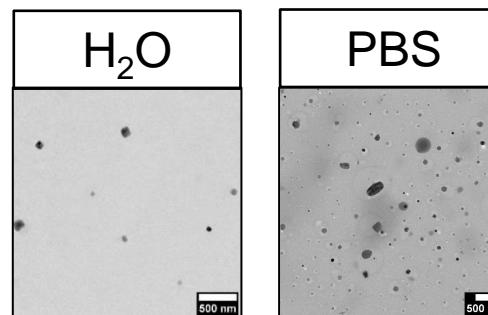
Conclusion

- ✓ Spherical morphology obtained when targeting $DP_{HPMA}=150$ after 120 min



- ✓ PBS provides more stable system

- ✓ Mixture of cubosomes and spherical NPs obtained when targeting $DP_{HPMA}=300$



- ✓ Macroinitiator suitable for glycosylation with mannosides



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**Thank you
for your
kind
attention**

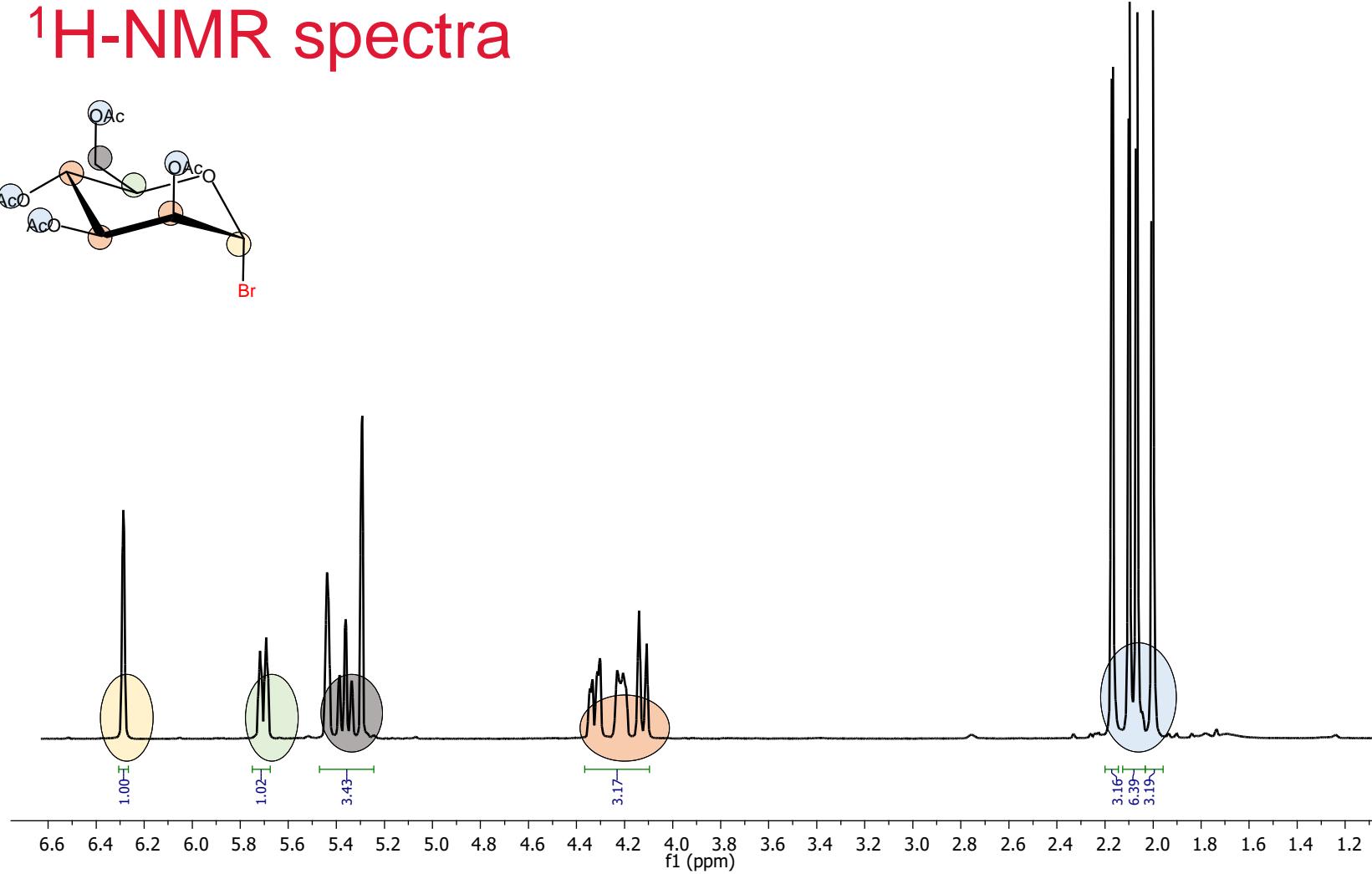
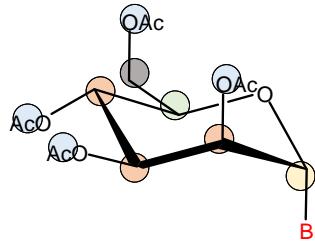




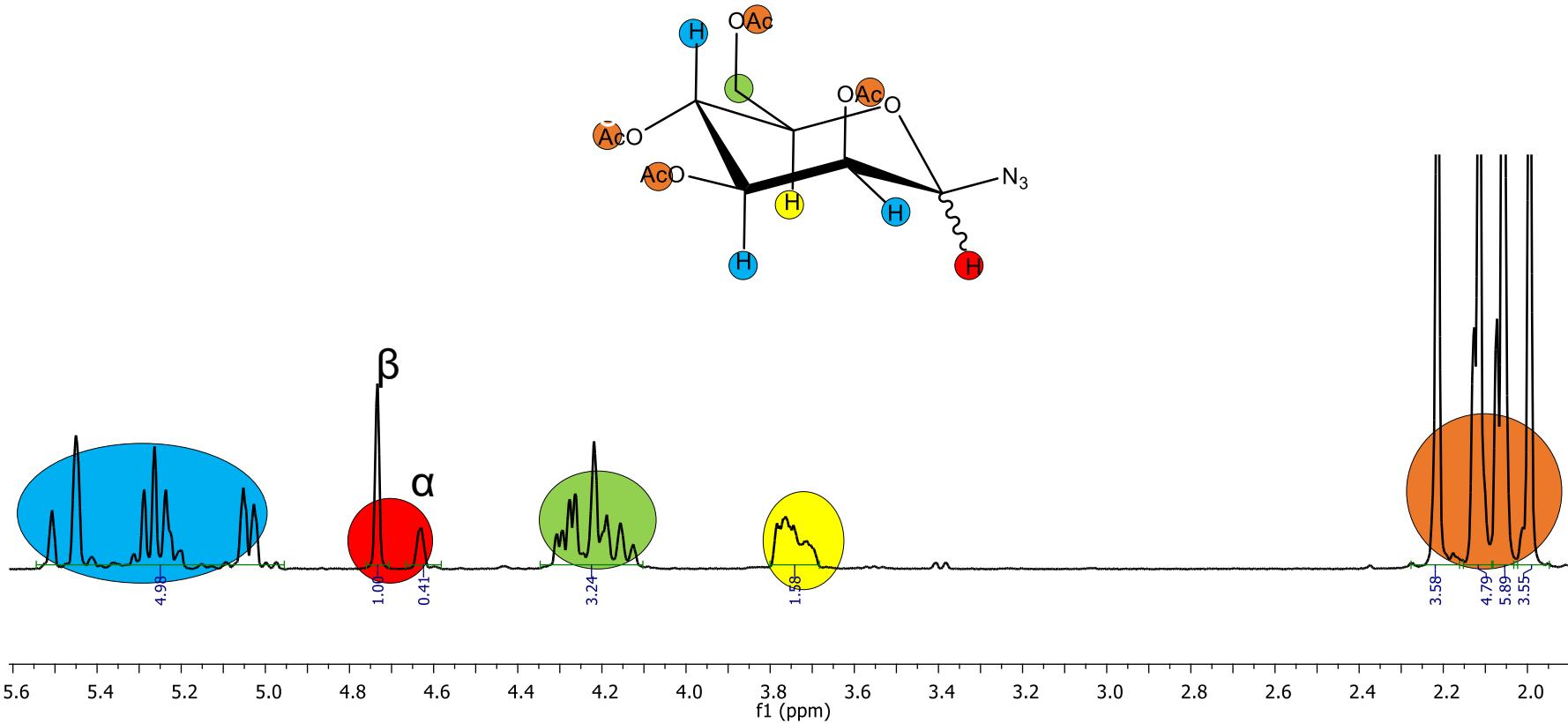
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Thank you

S1 ^1H -NMR spectra

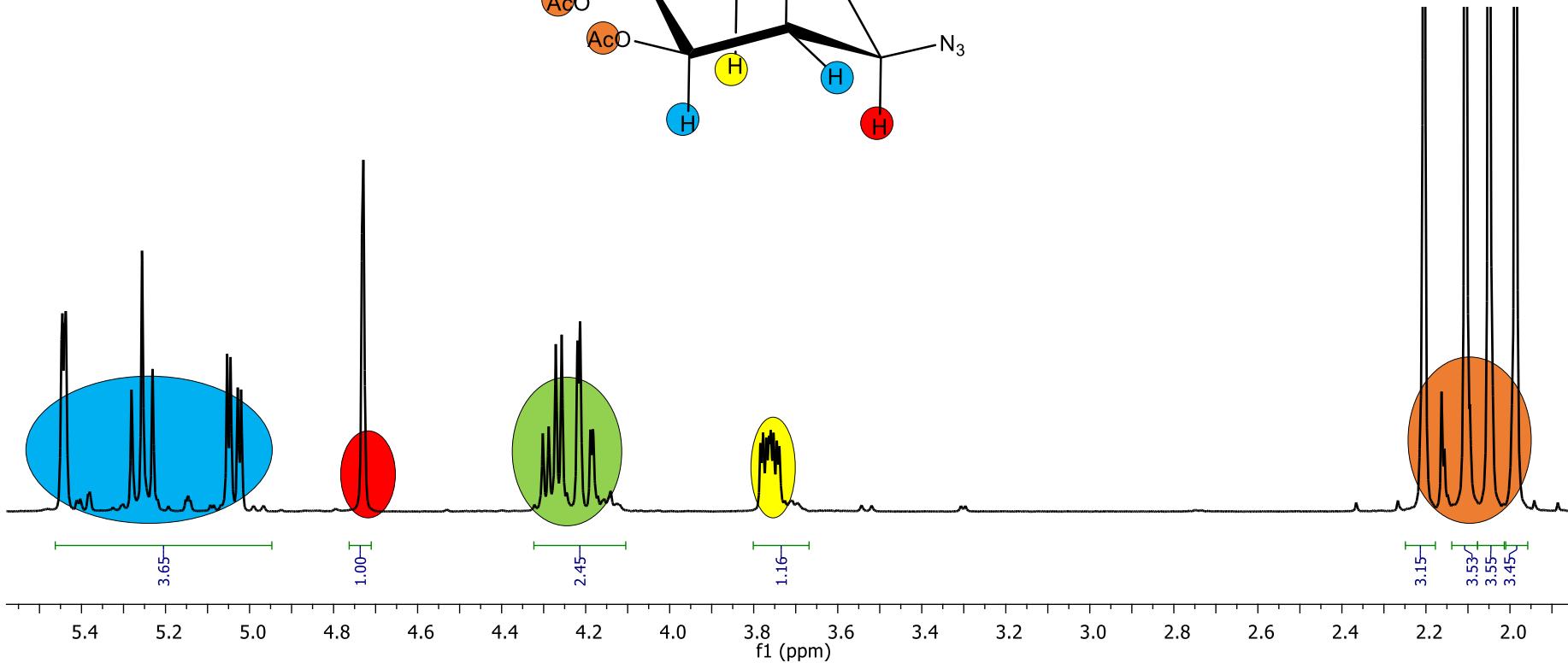
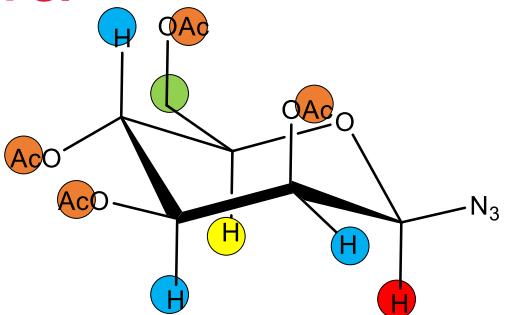


S2 ^1H -NMR spectra



S2 ^1H -NMR spectra

Reaction carried out at 70 °C



^1H -NMR spectra β -D-mannose-1-azide

