Assessing brain immune activation in psychiatric disorders: Clinical and

preclinical PET imaging studies of the 18-kDa Translocator Protein

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Abstract

Accumulating evidence from different lines of research suggests an involvement of the immune

system in the pathophysiology of several psychiatric disorders. During recent years, a series of PET

studies have been published using radioligands for the translocator protein (TSPO) to study microglia

activation in schizophrenia, bipolar I disorder, major depression, autism spectrum disorder and drug

abuse. The results have been somewhat conflicting, which could be due to differences both in

patient sample characteristics but also in PET methods. In particular, further work is needed to

address both methodological and biological sources of variability in TSPO levels, a process in which

the use of animal models and small animal PET systems can be a valuable tool. Given this

development, PET studies of immune activation have the potential to further increase our

understanding of disease mechanisms in psychiatric disorders, which is a requisite in the search for

new treatment approaches. Furthermore, molecular imaging could become an important clinical tool

for identifying specific subgroups of patients or disease stages that would benefit from treatment

targeting the immune system.

Key words

Psychiatry, PET, neuroinflammation, microglia, TSPO

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The role of the immune system in psychiatric disorders

The concept that the immune system may have a role in the pathophysiology of psychiatric disorders has been discussed for several decades. Early observations in support of this hypothesis include reports of psychotic symptoms in autoimmune and infectious diseases engaging the central nervous system [1, 2]. Furthermore, studies in both animals and humans showing that immune activation can induce depressive-like symptoms, as part of what is commonly referred to as the "sickness behaviour" syndrome [3, 4].

With regard to studies in psychiatric populations, initial epidemiological observations of increased incidence rates of schizophrenia for cohorts born after influenza epidemics [5] have been followed up by a series of studies showing that infections during gestation or in early life lead to an increased risk for developing psychotic disorders [6]. In addition, autoimmune diseases and infections have been associated with mood disorders [7]. In support of a role for genetically determined alterations in the immune system in schizophrenia, a recent large-scale genome-wide association study found that associations were enriched for genes related to immune function [8]. Effects of genes related to immune function have also been found for depression, bipolar disorder and autism [9, 10]. Furthermore, pharmacoepidemiological studies have suggested that anti-inflammatory drugs, such as non-steroidal anti-inflammatory drugs (NSAIDs), may lower the risk of psychiatric symptomatology[11].

Although the genetic and epidemiological data strongly suggest that immune function has a role in the development of psychiatric disease, an important question from a clinical perspective is if there are also ongoing brain immune disturbances in patients.

Post mortem studies have shown increases in microglia in patients with schizophrenia [12, 13], autism [14] and cocaine abuse [15]. However, the post-mortem results may differ from the brain immune situation *in vivo* and in general there is a large delay between disease onset and post mortem assessment. Studies on immune markers in blood, such as cytokines, have shown increased levels in both first-episode and chronic schizophrenia [16, 17] as well as in depression [18] and

autism [19]. Similarly, gene expression of immune-related proteins have shown to be altered in schizophrenia and bipolar disorder [20]. However, findings have generally been inconsistent, with different markers showing effects in different studies, and there is often a large degree of overlap between patients and control subjects. Importantly, there is little direct passage of cytokines between brain and periphery [21, 22] and therefore peripheral markers may not be representative of CNS processes. Studies in cerebrospinal fluid are scarce, but alterations in immune markers have been shown for schizophrenia [23, 24], bipolar disorder [25, 26] and at least in a subgroup of patients with depression [27]. Although these studies are encouraging, a large degree of inconsistency still remains. Furthermore, CSF analyses cannot provide any anatomical information, and there is thus a need for more direct measures of brain immune activation in psychiatric patients.

Measuring brain immune function in vivo: molecular imaging of TSPO

Molecular imaging is currently the only method to directly examine brain biomarkers at a molecular level *in vivo*. Of the available tools, positron emission tomography (PET) is the most accurate method, whereas single photon emission computed tomography (SPECT) has been less prevalent due to the lower sensitivity as well as limited temporal and spatial resolution as compared to PET [28]. Thus far, all PET studies assessing the immune system in psychiatric populations have targeted the 18 kDA translocator protein (TSPO), and this method is thus the focus of the present review.

TSPO, formerly known as the peripheral benzodiazepine receptor, is located in the outer mitochondrial membrane of microglia and, to some extent, astrocytes. Although its exact function remains unclear, TSPO has been described to be involved in many physiological processes, including cholesterol transport and steroid synthesis [29]. Importantly, microglia are considered to be the key cell type involved in CNS immune processes, as they can be activated by cytokines and additionally contribute further cytokine release [30]. Activation of resident microglia and infiltration of macrophages contributes to neuronal injury and synaptic damage [29], although neurotrophic and protective functions have also been observed [31, 32]. TSPO is a widely used marker for immune

activation in the CNS, since a change in phenotype and functional state of microglia is associated with increased TSPO expression [30].

The PET radioligand (R)-[11 C]PK11195 has been the most commonly used PET tracer to assess TSPO in the CNS. Several studies describe prolonged increased binding in patients with stroke and traumatic brain injury which can be seen as proof of concept studies [33, 34]. Since(R)-[11C]PK11195 shows a high level of non-specific binding, during the last decade significant efforts have been made to develop new TSPO radioligands. Those that have hitherto been applied to psychiatric conditions are [11C]DAA1106, [18F]FEPPA and [11C]PBR28 [35-40]. These second-generation ligands are considered to have improved signal-to-noise ratio compared to [11C]PK11195, based on a 7-60 fold higher affinity for TSPO and animal blocking studies showing up to 80-fold higher specific binding than [11C]PK11195 [36, 38, 40, 41]. A practical drawback with all these novel compounds is their sensitivity to a polymorphism in the gene encoding TSPO, which leads to three distinct binding classes: high-affinity binders (HABS) (49 % of the western population), mixed-affinity binders (MABS) (42%) and low-affinity binders (LABS) (9%) [13, 42]. As a consequence, up to ten percent of the population cannot be part of clinical studies due to LABS. In the remaining subjects the effects of TSPO polymorphism requires genotyping the subjects and controlling for genotype in the study design and analysis. Furthermore, second generation TSPO radioligands have thus far not been validated in preclinical models to the same extent.

Psychiatric disorders and TSPO PET imaging: clinical studies

We identified all PET studies that examined TSPO binding in psychiatric disorders (Table 1). Thus far, schizophrenia is the most studied condition with four articles, whereas only one to two studies have been published for mood disorders, autism spectrum disorder and drug abuse.

Schizophrenia

The first study on TSPO in schizophrenia used the radioligand (R)-[11 C]PK11195 in ten patients and ten age-matched healthy control subjects [43]. All patients were examined within five years of disease onset, with an average disease duration of 3.1 \pm 1.7 years. (R)-[11 C]PK11195 binding potential (BP_P) , as calculated with K1*k3/k2*k4) values were calculated using a two tissue compartment model (2TCM) with metabolite corrected plasma as input function. Patients had mild positive and negative symptoms at time of PET scanning as measured with the Positive and Negative Syndrome Scale (PANSS), with average scores of 12 \pm 3 and 14 \pm 4 for positive and negative symptoms respectively, and they were all on atypical antipsychotics. The authors observed an increase in (R)-[11 C]PK11195 binding potential in patients in total grey matter, which was the only region analysed (Table 1). No significant correlation between symptom severity and total grey matter BP $_P$ was found.

Doorduin et al. [44] used (R)-[11 C]PK11195 to examine seven patients with schizophrenia with active psychosis, defined by a score of 5 or more on 1 PANSS positive symptom item or a score of 4 on 2 items, in comparison to eight healthy volunteers. The disease duration ranged from 1 to 16 years (average 5.3 ± 5.6 years), with 1 to 4 experienced psychotic episodes. Patients had moderate symptoms at time of PET scanning, with average PANSS scores of 20 ± 3 , 17 ± 5 and 37 ± 7 for positive, negative and general subscales, respectively. All patients were using antipsychotics and the use of benzodiazepines was allowed for one to two weeks prior to PET examinations. Substance use and anti-inflammatory drugs were reported as exclusion criteria. (R)-[11 C]PK11195 binding potential (BP_{ND} defined as k3/k4) was quantified using 2TCM. In contrast to the study by van Berckel et al. [43], the (R)-[11 C]PK11195 binding potential of whole-brain grey matter was not found to be increased in

patients compared to control subjects. Multiple ROIs were examined (Table 1) and a significant increased BP_{ND} was found in the hippocampus of patients. To reduce the variation in the small sample size, the data was normalised to the whole-brain grey matter for statistical analysis which means that the results are not directly comparable to the other TSPO studies in schizophrenia.

Takano and colleagues [45] studied fourteen patients with chronic schizophrenia and fourteen age- and sex-matched controls with the second generation tracer [11]C]DAA1106. Patients had a long disease duration (18.8 ± 12.2 years). All patients were on antipsychotics, and benzodiazepines were allowed for more than one month before the start of the study. Substance and alcohol abuse were exclusion criteria. All cortical grey matter regions were assessed, as well as the striatum, and BP_{ND} as quantified using 2TCM was the main outcome measure. Patients had moderate symptoms at time of PET scanning as scored on PANSS (total score 78.6 ± 20.7). Although no significant differences in TSPO levels between patients and controls were reported, correlations were found between TSPO binding and disease duration as well as positive symptoms. However, TSPO genotype was not determined, and since in vitro studies show 4-fold differences in [11]C]DAA1106 affinity between MAB and HAB subjects [42], this significantly limits the interpretation of the results.

Using [18 F]FEPPA Kenk et al. [46] examined grey matter frontal and temporal ROIs, striatum, and white matter ROIs in a sample of 16 patients (10 HABs and 6 MABs) and 27 controls (19 HABs and 8 MABs)(Table 1). The disease duration was 14.8 ± 8.8 years and patients had moderate symptoms at time of PET scanning (PANSS total 70.2 ± 9.7). Patients were on treatment with either atypical or typical antipsychotics, and a minor proportion were also using antidepressants or anti-Parkinsonian drugs. Benzodiazepine use was not allowed except for clonazepam. Total distribution volumes (V_T) values were calculated using 2TCM with an arterial plasma input function. Since [18 F]FEPPA binding is also sensitive to TSPO genotype, this factor was included in the statistical

analysis. No significant differences in V_T values was found between patients and controls in either white or grey matter regions.

Clinical factors that might influence the study results are patient characteristics such as disease duration, symptom severity, and medication use. Both the two (R)-[11C]PK11195 studies that found an effect had patients with a shorter disease duration [43, 44], whereas no difference was been reported in patients with a longer disease duration [46]. This could indicate that microglia activation has a more prominent role in early disease phases. However, in order to answer this question longitudinal studies are necessary, ideally including also measurements in first-episode patients and high-risk individuals. In addition, patients had mild to moderate symptoms at time of PET scanning, and thus it is of interest to examine if more severe pathology shows a greater increase in TSPO binding. Importantly, benzodiazepine use was an exclusion criterium in all studies (except for clonazepam in one study [46]) since most benzodiazepines have affinity for the TSPO and can compete for binding with TSPO with the PET tracer [47]. However, a recent study found that this was mainly the case for the benzodiazepine diazepam in higher doses [48]. Further studies should be performed to confirm this observation, as alleviating this restriction could potentially improve recruitment of more severely ill patients.

One important limitation in all studies published thus far is that all patients were on treatment with antipsychotic drugs. Most antipsychotics, besides clozapine, tend to decrease TSPO expression [49]. Although evidence from in vivo human research is lacking, this suggests that there might be an underestimation of the signal. For instance, this could explain the negative findings in Kenk et al. [46]. Examination of drug-naive patients will be critical to address this issue.

Mood disorders

To date, two studies assessed TSPO in patients with major depressive disorder and one study examined bipolar I patients (Table 1).

In the study by Hannestad et al. $[50][^{11}C]PBR28$ was used to examine ten patients with major depression (MD) and ten healthy controls. The sample included seven HABs in both the patient group and control group respectively. Patients were allowed to take antidepressant medications if the dose had been stable for at least four weeks (although it was not reported how many patients were administered antidepressants at the time of PET). Laboratory signs of peripheral immune activation, as defined by elevated high sensitive C-reactive protein (CRP), was an exclusion criterium. Symptom severity at the time of examination was mild to moderate, with 19.7 ± 6.7 scores on the Montgomery- Åsberg Depression Rating Scale (MADRS, range 0-60; 0-6 normal, 7-19 mild, 20-34 moderate, >34 severe depression [51]). This represented a significant reduction from 25.6 ± 7.5 at screening. Groups were matched for TSPO genotype. [$^{11}C]PBR28 \ V_T$ values were calculated using 1TCM, 2TCM and the multilinear analysis MA1 (Ichise et al., 2002), using the metabolite corrected arterial plasma curve as input function. In addition, V_T corrected for plasma free fraction (V_T/f_P) was determined. No significant differences in binding was found in a selection of cortical ROIs, white matter and basal ganglia. Using V_T/f_P instead of V_T did not change these results.

Setiawan et al. [52] examined twenty MD patients (15 HABs, 5 MABs) and twenty healthy controls (14 HABs and 6 MABs) using [18 F]FEPPA. All patients were medication free for at least six weeks prior to the PET examination. Depressive symptoms were moderate to severe, with average scores of 20 \pm 3.8 on the 17-item Hamilton Depression Rating Scale (HDRS, range 0-52; 0-7= normal, 8-13 mild, 14-18 moderate, 19-22 severe, \geq 23 very severe depression [53]). [18 F]FEPPA V_T was calculated using 2TCM was the main outcome measure, and ROIs selected were prefrontal cortex, anterior cingulate cortex (ACC) and insula. Genotype was included as a factor in the statistical analysis. A global effect of diagnosis on [18 F]FEPPA V_T was shown, with higher values in patients in all regions examined. Furthermore, HDRS scores were positively correlated to TSPO V_T in the ACC, after correcting for genotype. Serum markers for neuroinflammation were measured, but there were no correlations to brain TSPO levels.

Haarman et al. [54] studied fourteen bipolar I patients compared to eleven healthy volunteers. All patients except one were euthymic, and the disease duration ranged from 2 to 37 years (average 25.6 ± 12.0 years). All patients had experienced multiple depressive and hypomanic or manic episodes. All except one patient were on mood stabilizing medication. (R)-[¹¹C]PK11195 BP_{ND} was quantified using 2TCM with plasma input function. Multiple brain regions were analysed, although hippocampus was selected as the main ROI, based on human and preclinical evidence suggesting that this region could be a particular focus of microglia activation. For the statistical analysis, binding in whole-brain grey matter was used as a covariate to reduce the between-subject variation. Whereas BP_{ND} in the whole-brain was not different between patients and healthy volunteers, higher values were observed in the right hippocampus of bipolar I patients.

The differences in clinical factors between the mood disorder studies makes it difficult to compare the PET outcome. In the two studies on depression, the study of Setiawan et al. [52] had a significantly larger sample size, and patients had more severe symptoms than in the study of Hannestad et al. [50], which might explain the difference in results. A further difference is the exclusion of signs of mild peripheral immune activation in the latter study. Since CRP has shown to be elevated in patients in depression [55], suggesting that peripheral immune activation is related to the disease mechanism, this could have led to a selection bias towards patients with relatively low levels of immune activation.

Furthermore, in two of the studies the patients were administered mood stabilizing medication [50, 54] whereas in the study by Setiawan [52] patients were medication free for six weeks. Most mood stabilizing medication tend to decrease TSPO expression [56, 57] which suggests an underestimation of the signal in the patients who were assigned to treatment, although this has to be confirmed in future studies with medication naïve patients.

Developmental disorders

To date, only one PET study has assessed TSPO binding in patients with developmental disorders. In the study by Suzuki et al. [58] (R)-[11 C]PK11195 was used to examine twenty individuals with autism spectrum disorder and twenty age- and intelligence quotient (IQ)-matched controls. Patients were young adults in an age range from 18.6 to 31.9 (average 23.3 ± 4.0) years, had IQ scores above 80 (average 95.9 ± 16.7), and did not receive any medication. Arterial blood was not sampled, instead (R)-[11 C]PK11195 BP_{ND} was quantified using a reference tissue approach. A normalised input curve based on the averaged TACs from the ROIs placed over the cerebellar cortices in the control group was used for both the control as the patient data analysis. In the ROI-based analysis an overall statistical effect was shown, and increased (R)-[11 C]PK11195 binding potential was reported in all examined regions, including (subparts of) the cerebellum, ACC, corpus callosum, and frontal, temporal, and parietal cortex (Table 1). These results were confirmed in a voxel-based analysis. There were no correlations between TSPO BP_{ND} and clinical symptoms levels. As discussed below, the reference tissue approach used in the study limits the conclusions that can be drawn.

Substance use disorders - and the potential confound of drug use

Two PET studies examined TSPO in substance use disorders (Table 1). Sekine et al. [59] used (R)-[11 C]PK11195 to examine twelve methamphetamine abusers in comparison to twelve healthy volunteers. The disease duration of the patients ranged from 1 to 12 years (average 6.8 \pm 3.9 years) however patients were abstinent on average almost two years prior to PET examinations. A modified version of the Drug Effect Rating Scale was used to assess the scale for methamphetamine craving. Scores were in a range from 1 (no craving) to 10 (most intense craving) with an average score of 4.9 \pm 3.4. PET data were analysed with a reference tissue approach, whereby averaged TACs from the frontal, parietal and occipital cortices from the control group was used for both the control as the patient data analysis. Significantly higher BP_{ND} values were found in methamphetamine abusers in all ROIs studied, with differences ranging from 3 to 15-fold. A significant negative correlation was found between (R)-[11 C]PK11195 BP_{ND} and the duration of methamphetamine abstinence in midbrain,

thalamus and striatum, suggesting that microglial function can be normalized by protracted abstinence.

In the study by Narendran et al. [60] fifteen chronic cocaine abusers (8 HABs, 8 MABs) and seventeen healthy controls (12 HABs, 5 MABs) were examined using [11 C]PBR28. Patients were on average 39.9 \pm 9.0 years old and had been smoking crack cocaine for an average of 17 \pm 7 years. Patients had a minimum of 2 weeks outpatient abstinence as monitored with urine toxicology and did not receive medication. V_T values were calculated in a standard manner, using 2TCM with an arterial plasma input function, and data was corrected for TSPO genotype. No differences in [11 C]PBR28 V_T was observed between patients and control subjects, as assessed in both cortical and subcortical brain regions.

Thus, these two studies differ markedly in their findings. However, apart from the fact that different drugs of abuse were studied, a direct comparison is precluded since there are major differences in both the clinical status of patients, such as time of abstinence, and methodology -in particular the use of a reference region in the study by Sekine et al. (see below for discussion).

For most psychiatric patients groups, there is an increased prevalence of substance use disorders, such as alcohol, cannabis and nicotine use [61]. For instance, cannabis is considered to be the most commonly used illegal drug of abuse among patients with bipolar disorder and schizophrenia [62, 63]. Although substance use disorders was an exclusion criteria in all studies, it cannot be excluded that patients had a higher recreational use of drugs of abuse. In most studies, actual drug use was assessed with urine analysis [43, 46, 50, 52, 59, 60]. However, only four of the studies reported on or controlled for smoking status, of which one study included only non-smokers [52], one study reported only three smokers in the patient group [50], one study allowed smoking and matched patients and controls for nicotine smoking status [60] and one study did not include subjects that fulfilled nicotine related DSM-IV criteria [59]. Importantly, *in vitro* studies have shown that cannabinoids and nicotine have a suppressant effect on the immune system [64, 65]. Although

these observations remains to be confirmed *in vivo*, the use of these common drugs of abuse could thus theoretically diminish differences in TSPO levels between patients and control subjects, which might in some cases contribute to negative findings. In contrast, if an effect of psychostimulants in TSPO levels even after long abstinence intervals can be confirmed, group differences in the use of this drug could confound the results in the other direction.

Preclinical studies of TSPO and psychiatric disorders

Animal models and microglia activation

As psychiatric disorders are complex human disorders of which the etiology and the exact pathophysiology are unknown, the three criteria commonly used for the validation of an animal model, i.e. face, construct and predictive validity [66], cannot be fulfilled by a single animal model. Animal models of psychiatric disorders do therefore only manifest certain aspects that can be translated to the human disease, i.e. endophenotypes, such as hyperlocomotion, anxiety or deficits in pre-pulse inhibition. With regard to the role of the immune systems in psychiatric disorders, many animal models have linked psychotic – and depressive-like behaviour to inflammatory processes. For instance, modelling effects of maternal immune activation on the risk of developing schizophrenia, viral infection of pregnant dams resulted in psychotic-like symptoms in the offspring, and also microglia were found to be activated [67, 68]. The bacterial endotoxin lipopolysaccharide (LPS) is often used to induce depressive-like behaviour in rodents, leading to immediate sickness behaviour followed by behavioural deficits that can be linked to depression [69, 70]. Importantly, LPS has been shown to induce an activation of microglia, lasting several days, as detected using immunohistochemistry [71]. Conversely, animal models for stress-induced depression have shown to induce both depressive behaviour and microglia activation [72].

Preclinical TSPO PET imaging

With the advent of small animal PET systems, it has become feasible to estimate TSPO levels in rats and mice, thus enabling a translational approach (Figure 1). Thus far only a few studies have assessed TPSO in animal models of psychiatric disorders. Herpes virus induced psychotic-like behaviour in rats was found to be accompanied by microglia activation, as shown with (R)-[11]C]PK11195 PET [73]. (R)-[11]C]PK11195 uptake was found to increase with the increased severity of psychotic-like behaviour, and psychotic-like behaviour and microglia activation could be reduced by antipsychotic treatment.

Dobos et al. [74] used (R)-[11]C]PK11195 PET to show microglia activation and depressive-like behaviour following intracerebral LPS injection in mice. Recently, LPS-induced systemic inflammation in baboons was shown to induce an activation of microglia, as shown with [11]C]PBR28 PET [75]. A robust increase in TSPO binding was observed at 1hr and 4hr after intravenous injection of LPS.

Depressive-like behaviour was not assessed in this study.

Methodological considerations

Taken together, the data thus far suggests a modest elevation of TSPO in psychiatric disorders. In general, the increase was found throughout the entire brain, although some studies reported a more focal elevation. However, almost half of the studies did not find a difference. Importantly, the results of the TSPO PET studies are in many instances difficult to compare, not only due to differences in patient characteristics as discussed above, but also since the methods differ in many aspects. In the following, we discuss some key methodological points of relevance for interpreting the findings.

The gold standard for TSPO PET quantification is to apply a 2TCM, using a metabolite corrected plasma input function to estimate the contribution of free and nonspecific binding to the brain signal. The outcome parameter of choice using this approach is V_T , since the direct estimation of BP for instance by calculating k3/k4, is less reliable. Therefore, to facilitate comparisons between the studies it would have been useful if the studies with BP as outcome parameter had additionally reported V_T values.

Plasma input models are considered to have a high variability, which may be due to methodological issues in measuring the metabolite-corrected plasma input curve. In order to reduce variability of plasma input models, several approaches have been proposed.

In two of the studies reviewed above, normalisation of regional values to whole brain binding was performed [44, 54]. Although this reduces the variance, the drawback with this approach is that it may underestimate true specific binding, especially in case of a global increase. Furthermore, the whole cortex cannot be analysed using this approach.

Reference tissue approaches, such as simplified reference tissue method (SRTM) [76] and Logan DVR [77] are in general preferred above plasma input models if a reference region can be identified, both to reduce noise but also from a practical perspective since arterial sampling can then be eliminated. This may be particularly important for psychiatric patient groups. One method is to use ratios of Standardised Uptake Values, which was recently reported to be more sensitive than V_T to detect TSPO elevations in Alzheimer's disease for [11 C]PBR28, with cerebellum as reference [78]. However, the use of this method assumes a very focal TSPO elevation, which might not be the case for most psychiatric disorders, and is sensitive to bias induced by difference in blood flow between regions and between groups.

In general, all reference tissue approaches are problematic in TSPO PET studies as there is no anatomical region in the brain devoid of TSPO. Whereas this protein was initially not thought to be present in the non-diseased brain, widespread (R)-[11C]PK11195 BP_{ND} increase was found in healthy aging [79]. In addition, specific binding to TSPO in healthy individuals was recently confirmed in a TSPO blocking study [80]. In the studies on autism and methamphetamine use by Suzuki et al. [58] and Sekine et al. [59], normalised average time activity curves (TACs) from the control group was used for both patient and control data, which is not according to convention for reference tissue approaches and creates further problems. In this case, group differences in radioligand delivery and

non-displaceable binding are not taken into account, which might generate artificially high BP_{ND} values in patients, and the results therefore have to be interpreted with caution.

An alternative approach to identify a reference "region" is to identify voxels with low specific binding based on the shape of the TAC using supervised cluster analysis. Several methods have been published for this purpose and it show reliable results in normal aging and after traumatic brain injury, both conditions leading to robust microglia activation [79, 81]. However, the validity of voxels devoid of TSPO can be questioned as for anatomical reference regions, and this method might underestimate the signal in comparison to other approaches [82]. Another method to eliminate the need for arterial blood sampling is to use image derived input function. By extracting and input function from PET voxels corresponding to blood vessels, this approach has been validated for tracers such as [11C]flumazenil and [11C]AZ10419369 [83]. Preliminary data has shown feasibility for this method to quantify [11C]PBR28 using HRRT [84]. So far neither of these approaches have been used to study psychiatric populations.

General comments and future directions

It is well established that immune activation generally has a negative effect on psychological well-being, and may contribute to both psychotic and depressive symptoms. A role for immune-related processes as part of specific, causative disease mechanisms in psychiatric disorders is at present more uncertain. For schizophrenia and autism, both genetic and epidemiological associations to immune function could reflect a role for immunological factors affecting neurodevelopment, which for instance is in line with recent observations suggesting that microglia are involved in synaptic pruning as well as providing trophic support for neuronal growth [32]. An alternative interpretation of these early effects is the "sensitization" theory, postulating that early immune challenges to a genetically vulnerable system leads to an increased inflammatory response later in life [85]. Even so, the changes in psychiatric disorders are likely to be more subtle than for classic CNS autoimmune disorders such as multiple sclerosis (MS) and systemic lupus erythematosus (SLE), as there are no

radiological signs and the changes in CSF and blood are less distinct. Since the immune system can in certain instances even be regarded as neuroprotective [31], immune dysregulation in psychiatric disorders may even be conceived as an imbalance rather than a "one-dimensional" immune response.

Given these proposed more subtle immune changes in psychiatric disorders, a major challenge for clinical TSPO PET studies is the large degree of variability even in healthy individuals. This is true irrespective of the radioligand and methods of quantification used. In the studies reviewed above, where variability was reported, the overall % covariance (COV) was around 35% for second generation radioligands across genotypes, and 25% for (R)-[11C]PK11195. With regard to interindividual variability, this has shown to be moderate to high for both (R)-[11C]PK11195 [86] and [11C]PBR28 [87, 88], whereas reproducibility has not been assessed for any other TSPO radioligands. Although part of this inter- and individual variability may be due to methodological noise as discussed above, there may also be biological explanations. For instance, in Collste et al. diurnal effects were shown on GM V_T values, indicating that part of the variability could be due to natural fluctuations in the immune system in health [88]. The variability in TSPO levels has important implications in terms of the sample samples needed, both for group comparisons and longitudinal studies. For instance, in the study on schizophrenia by Kenk et al. [46] based on mean and variability of VT values, it was calculated that 21 subjects per group was required to detect a 20% difference (alpha = .05 and power = 0.8), which can be compared to the 16% increase in [3H]PBR28 shown in a post-mortem study[13]. If similar effects are expected in other disorders, most studies performed thus far may have been underpowered. Sample size may be particularly critical for studies using second generation radioligands, since accounting for TSPO genotype in the analysis reduces statistical degrees of freedom. To enhance the sensitivity of TSPO PET, an important direction of research will be to not only reduce methodological noise by finding reliable alternatives to arterial sampling, but also to identify the physiological factors that may influence TSPO levels in brain.

One important limitation for PET TSPO studies in general is the lack of specificity of this marker. For instance, TSPO is present also in astrocytes, which could contribute to the signal [89, 90], and although both cell types contribute to immune response, the specific type of cell may have implications in terms of treatment. Furthermore, for the microglia population, TSPO cannot distinguish between pro-inflammatory (M1) or protective (M2) subpopulations. Thus, it will also be crucial to develop tracers for other, more specific microglia markers that are also sensitive to the functional phenotype. Importantly, PET data then needs to be combined with other measures of immune function in order to fully reflect the immunological status for each disease condition.

Although TSPO levels were compared to serum markers of inflammation in some studies reviewed above, ideally this comparison should be done using CFS samples which more closely reflect the state of the CNS immune system.

A crucial aim for research on the immune system in psychiatry is inform new treatment strategies. In schizophrenia, depression and autism, randomised controlled studies have shown that the addition of cyclo-oxygenase (COX)-2 inhibitors to antipsychotics and antidepressants treatment may lead to amelioration of symptoms [91–93], although negative results have also been reported [94]. In psychosis, both the COX inhibitor aspirin and the broad-spectrum tetracycline antibiotic minocyclin, which directly attenuates microglia activation, have shown to reduce symptoms [95, 96]. However, the immune hypothesis of psychiatric disorders would implicate a need for drugs specifically directed towards causative mechanisms. Molecular imaging is an important tool towards this goal, as it allows for non-invasive studies in both patients and animal models. Thus, immune markers can be followed over time, allowing for monitoring of disease progress and treatment effect. For instance, [111C]PBR28 PET was recently used to detect effects of immune-targeted treatment in Parkinson's disease [97]. Furthermore, due to the translational nature for PET, clinical studies can be supported by preclinical experiments, also allowing for validation with post-mortem techniques such as immunohistochemistry. For many psychiatric disorders where large clinical trials have suffered

from negative findings, these approaches could be crucial towards revitalising drug development programmes.

With increasing understanding of the biological processes as well as methodological refinement, PET studies of the immune system promises to be an important tool also for clinical use. An important task will be to aid diagnosis, and potentially identify specific subgroups of psychiatric patients in need for immune-targeting treatment. For instance, a proportion of patients with schizophrenia show auto-antibodies for neural receptors, suggesting that these patients may constitute a subgroups in specific need of treatment targeted against the immune system [98]. In depression, specific subgroups of patients was shown to respond to anti-TNF treatment [99]. Importantly, such PET markers may prove to be specific for particular pathophysiological mechanisms and symptom dimensions, rather than reflecting current disease classifications according to DSM.

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Compliance with Ethics Guidelines

T. F. van der Doef, J. Doorduin, B.N. M. van Berckel and S. Cervenka have nothing to disclose. This article does not contain any studies with human or animal subjects performed by the any of the authors.

Contributions

- T.F. van der Doef: Literature search and Review, Content Planning, Writing, Editing
- J. Doorduin: Literature search and Review, Content Planning, Writing, Editing
- B.N.M van Berckel: Literature search and Review, Content Planning, Writing, Editing
- S. Cervenka: Literature search and Review, Content Planning, Writing, Editing

References

- 1. Oommen KJ, Johnson PC, Ray CG (1982) Herpes simplex type 2 virus encephalitis presenting as psychosis. Am J Med 73:445–8.
- 2. Felgenhauer K (1990) Psychiatric disorders in the encephalitic form of multiple sclerosis. J Neurol 237:11–8.
- 3. Reichenberg A, Yirmiya R, Schuld A, et al. (2001) Cytokine-associated emotional and cognitive disturbances in humans. Arch Gen Psychiatry 58:445–452.
- 4. Dantzer R, O'Connor JC, Freund GG, et al. (2008) From inflammation to sickness and depression: when the immune system subjugates the brain. Nat Rev Neurosci 9:46–56.
- 5. Mednick SA, Machon RA, Huttunen MO, Bonnet D (1988) Adult Schizophrenia Following Prenatal Exposure to an Influenza Epidemic. Arch Gen Psychiatry 45:189.
- 6. Arias I, Sorlozano A, Villegas E, et al. (2012) Infectious agents associated with schizophrenia: a meta-analysis. Schizophr Res 136:128–36.
- 7. Benros ME, Waltoft BL, Nordentoft M, et al. (2013) Autoimmune diseases and severe infections as risk factors for mood disorders: a nationwide study. JAMA psychiatry 70:812–20.
- 8. Ripke S, Neale BM, Corvin A, et al. (2014) Biological insights from 108 schizophrenia-associated genetic loci. Nature.
- 9. Odell D, Maciulis A, Cutler A, et al. (2005) Confirmation of the association of the C4B null allelle in autism. Hum Immunol 66:140–5.
- 10. Consortium N and PAS of PG (2015) Psychiatric genome-wide association study analyses implicate neuronal, immune and histone pathways. Nat Neurosci 18:199–209.
- 11. Laan W, Selten J-P, Grobbee DE, et al. (2007) Non-steroidal anti-inflammatory drugs and the risk of psychosis. Eur Neuropsychopharmacol 17:309–311.
- 12. Radewicz K, Garey LJ, Gentleman SM, Reynolds R (2000) Increase in HLA-DR immunoreactive microglia in frontal and temporal cortex of chronic schizophrenics. J Neuropathol Exp Neurol 59:137–50.
- 13. Kreisl WC, Jenko KJ, Hines CS, et al. (2013) A genetic polymorphism for translocator protein 18 kDa affects both in vitro and in vivo radioligand binding in human brain to this putative biomarker of neuroinflammation. J Cereb Blood Flow Metab 33:53–8.

- 14. Morgan JT, Chana G, Pardo CA, et al. (2010) Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. Biol Psychiatry 68:368–76.
- 15. Little KY, Ramssen E, Welchko R, et al. (2009) Decreased brain dopamine cell numbers in human cocaine users. Psychiatry Res 168:173–180.
- 16. Upthegrove R, Manzanares-Teson N, Barnes NM (2014) Cytokine function in medication-naive first episode psychosis: A systematic review and meta-analysis. Schizophr Res 155:101–108.
- 17. Miller BJ, Buckley P, Seabolt W, et al. (2011) Meta-analysis of cytokine alterations in schizophrenia: clinical status and antipsychotic effects. Biol Psychiatry 70:663–71.
- 18. Dowlati Y, Herrmann N, Swardfager W, et al. (2010) A Meta-Analysis of Cytokines in Major Depression. Biol Psychiatry 67:446–457.
- 19. Mitchell RHB, Goldstein BI (2014) Inflammation in children and adolescents with neuropsychiatric disorders: A systematic review. J Am Acad Child Adolesc Psychiatry 53:274–296.
- 20. Drexhage RC, van der Heul-Nieuwenhuijsen L, Padmos RC, et al. (2010) Inflammatory gene expression in monocytes of patients with schizophrenia: overlap and difference with bipolar disorder. A study in naturalistically treated patients. Int J Neuropsychopharmacol 13:1369–81.
- 21. Bromander S, Anckarsäter R, Kristiansson M, et al. (2012) Changes in serum and cerebrospinal fluid cytokines in response to non-neurological surgery: an observational study. J Neuroinflammation 9:242.
- 22. Maier B, Laurer H-L, Rose S, et al. (2005) Physiological levels of pro- and antiinflammatory mediators in cerebrospinal fluid and plasma: a normative study. J Neurotrauma 22:822–35.
- 23. Soderlund J, Schroder J, Nordin C, et al. (2009) Activation of brain interleukin-1beta in schizophrenia. Mol Psychiatry 14:1069–71.
- 24. Schwieler L, Larsson MK, Skogh E, et al. (2014) Increased levels of IL-6 in the cerebrospinal fluid of patients with chronic schizophrenia significance for activation of the kynurenine pathway. J Psychiatry Neurosci 39:140126.
- 25. Jakobsson J, Bjerke M, Sahebi S, et al. (2015) Monocyte and microglial activation in patients with mood-stabilized bipolar disorder. J Psychiatry Neurosci 40:250–8.
- 26. Söderlund J, Olsson SK, Samuelsson M, et al. (2011) Elevation of cerebrospinal fluid interleukin-1ß in bipolar disorder. J Psychiatry Neurosci 36:114–8.

- 27. Young JJ, Bruno D, Pomara N (2014) A review of the relationship between proinflammatory cytokines and major depressive disorder. J Affect Disord 169C:15–20.
- 28. Rahmim A, Zaidi H (2008) PET versus SPECT: strengths, limitations and challenges. Nucl Med Commun 29:193–207.
- 29. Liu G-J, Middleton RJ, Hatty CR, et al. (2014) The 18 kDa Translocator Protein, Microglia and Neuroinflammation. Brain Pathol 24:631–653.
- 30. Venneti S, Lopresti BJ, Wiley CA (2013) Molecular imaging of microglia/macrophages in the brain. Glia 61:10–23.
- 31. Hohlfeld R, Kerschensteiner M, Meinl E (2007) Dual role of inflammation in CNS disease. Neurology 68:S58–63; discussion S91–6.
- 32. Paolicelli RC, Gross CT (2011) Microglia in development: linking brain wiring to brain environment. Neuron Glia Biol 7:77–83.
- 33. Cagnin A, Gerhard A, Banati RB (2002) In vivo imaging of neuroinflammation. Eur Neuropsychopharmacol 12:581–586.
- 34. Gerhard A, Schwarz J, Myers R, et al. (2005) Evolution of microglial activation in patients after ischemic stroke: A [11C](R)-PK11195 PET study. Neuroimage 24:591–595.
- 35. Fujita M, Imaizumi M, Zoghbi S, Fujimura Y (2008) Kinetic analysis in healthy humans of a novel positron emission tomography radioligand to image the peripheral benzodiazepine receptor, a potential biomarker for. Neuroimage 40:43–52.
- 36. Imaizumi M, Briard E, Zoghbi S, Gourley J (2008) Brain and whole-body imaging in nonhuman primates of PBR28, a promising PET radioligand for peripheral benzodiazepine receptors. Neuroimage 39:1289–1298.
- 37. Ikoma Y, Yasuno F, Ito H, et al. (2007) Quantitative analysis for estimating binding potential of the peripheral benzodiazepine receptor with [(11)C]DAA1106. J Cereb Blood Flow Metab 27:173–84.
- 38. Maeda J, Suhara T, Zhang M-R, et al. (2004) Novel peripheral benzodiazepine receptor ligand [11C]DAA1106 for PET: an imaging tool for glial cells in the brain. Synapse 52:283–91.
- 39. Rusjan PM, Wilson AA, Bloomfield PM, et al. (2011) Quantitation of translocator protein binding in human brain with the novel radioligand [18F]-FEPPA and positron emission tomography. J Cereb Blood Flow Metab 31:1807–16.

- 40. Wilson AA, Garcia A, Parkes J, et al. (2008) Radiosynthesis and initial evaluation of [18F]-FEPPA for PET imaging of peripheral benzodiazepine receptors. Nucl Med Biol 35:305–14.
- 41. Kreisl WC, Fujita M, Fujimura Y, et al. (2010) Comparison of [(11)C]-(R)-PK 11195 and [(11)C]PBR28, two radioligands for translocator protein (18 kDa) in human and monkey: Implications for positron emission tomographic imaging of this inflammation biomarker. Neuroimage 49:2924–2932.
- 42. Owen DRJ, Gunn RN, Rabiner EA, et al. (2011) Mixed-affinity binding in humans with 18-kDa translocator protein ligands. J Nucl Med 52:24–32.
- 43. Van Berckel BN, Bossong MG, Boellaard R, et al. (2008) Microglia Activation in Recent-Onset Schizophrenia: A Quantitative (R)-[11C]PK11195 Positron Emission Tomography Study. Biol Psychiatry 64:820–822.
- 44. Doorduin J, de Vries EFJ, Willemsen ATM, et al. (2009) Neuroinflammation in schizophrenia-related psychosis: a PET study. J Nucl Med 50:1801–7.
- 45. Takano A, Arakawa R, Ito H, et al. (2010) Peripheral benzodiazepine receptors in patients with chronic schizophrenia: a PET study with [11C]DAA1106. Int J Neuropsychopharmacol 13:943–950.
- 46. Kenk M, Selvanathan T, Rao N, et al. (2014) Imaging Neuroinflammation in Gray and White Matter in Schizophrenia: An In-Vivo PET Study With [18F]-FEPPA. Schizophr Bull 41:85–93.
- 47. Gavish M, Bachman I, Shoukrun R, et al. (1999) Enigma of the peripheral benzodiazepine receptor. Pharmacol Rev 51:629–650.
- 48. Kalk NJ, Owen DR, Tyacke RJ, et al. (2013) Are prescribed benzodiazepines likely to affect the availability of the 18 kDa translocator protein (TSPO) in PET studies? Synapse 67:909–12.
- 49. Danovich L, Veenman L, Leschiner S, et al. (2008) The influence of clozapine treatment and other antipsychotics on the 18 kDa translocator protein, formerly named the peripheral-type benzodiazepine receptor, and steroid production. Eur Neuropsychopharmacol 18:24–33.
- 50. Hannestad J, DellaGioia N, Gallezot JD, et al. (2013) The neuroinflammation marker translocator protein is not elevated in individuals with mild-to-moderate depression: A [11C]PBR28 PET study. Brain Behav Immun 33:131–138.
- 51. Montgomery SA, Asberg M (1979) A new depression scale designed to be sensitive to change. Br J Psychiatry 134:382–389.
- 52. Setiawan E, Wilson A a, Mizrahi R, et al. (2015) Role of Translocator Protein Density, a Marker of Neuroinflammation, in the Brain During Major Depressive Episodes. 72:1–8.

- 53. HAMILTON M (1960) A rating scale for depression. J Neurol Neurosurg Psychiatry 23:56–62.
- 54. Haarman BCMB, Riemersma-Van der Lek RF, de Groot JC, et al. (2014) Neuroinflammation in bipolar disorder - A [11C]-(R)-PK11195 positron emission tomography study. Brain Behav Immun 40:219–225.
- 55. Howren MB, Lamkin DM, Suls J (2009) Associations of depression with C-reactive protein, IL-1, and IL-6: a meta-analysis. Psychosom Med 71:171–186.
- 56. Haarman BBCM, Riemersma-Van der Lek RF, Burger H, et al. (2014) Relationship between clinical features and inflammation-related monocyte gene expression in bipolar disorder towards a better understanding of psychoimmunological interactions. Bipolar Disord 16:137–150.
- 57. Rybakowski JK (2000) Antiviral and immunomodulatory effect of lithium. Pharmacopsychiatry 33:159–164.
- 58. Suzuki K, Sugihara G, Ouchi Y, et al. (2013) Microglial activation in young adults with autism spectrum disorder. JAMA psychiatry 70:49–58.
- 59. Sekine Y, Ouchi Y, Sugihara G, et al. (2008) Methamphetamine causes microglial activation in the brains of human abusers. J Neurosci 28:5756–5761.
- 60. Narendran R, Lopresti BJ, Mason NS, et al. (2014) Cocaine Abuse in Humans Is Not Associated with Increased Microglial Activation: An 18-kDa Translocator Protein Positron Emission Tomography Imaging Study with [11C]PBR28. J Neurosci 34:9945–50.
- 61. Ziedonis D, Hitsman B, Beckham JC, et al. (2008) Tobacco use and cessation in psychiatric disorders: National Institute of Mental Health report. Nicotine Tob Res 10:1691–1715.
- 62. Bally N, Zullino D, Aubry JM (2014) Cannabis use and first manic episode. J Affect Disord 165:103–108.
- 63. Leweke FM, Koethe D (2008) Cannabis and psychiatric disorders: It is not only addiction. Addict Biol 13:264–275.
- 64. Guan Y-Z, Jin X-D, Guan L-X, et al. (2014) Nicotine Inhibits Microglial Proliferation and Is Neuroprotective in Global Ischemia Rats. Mol Neurobiol 51:1480–1488.
- 65. Suárez-Pinilla P, López-Gil J, Crespo-Facorro B (2014) Immune system: A possible nexus between cannabinoids and psychosis. Brain Behav Immun 40:269–282.
- 66. Van der Staay FJ, Arndt SS, Nordquist RE (2009) Evaluation of animal models of neurobehavioral disorders. Behav Brain Funct 5:11.

- 67. Mattei D, Djodari-Irani A, Hadar R, et al. (2014) Minocycline rescues decrease in neurogenesis, increase in microglia cytokines and deficits in sensorimotor gating in an animal model of schizophrenia. Brain Behav Immun 38:175–184.
- 68. Juckel G, Manitz MP, Brüne M, et al. (2011) Microglial activation in a neuroinflammational animal model of schizophrenia--a pilot study. Schizophr Res 131:96–100.
- 69. Fu X, Zunich SM, O'Connor JC, et al. (2010) Central administration of lipopolysaccharide induces depressive-like behavior in vivo and activates brain indoleamine 2,3 dioxygenase in murine organotypic hippocampal slice cultures. J Neuroinflammation 7:43.
- 70. O'Connor JC, Lawson MA, André C, et al. (2009) Lipopolysaccharide-induced depressive-like behavior is mediated by indoleamine 2,3-dioxygenase activation in mice. Mol Psychiatry 14:511–522.
- 71. Buttini M, Limonta S, Boddeke HWGM (1996) Peripheral administration of lipopolysaccharide induces activation of microglial cells in rat brain. Neurochem Int 29:25–35.
- 72. Reader BF, Jarrett BL, McKim DB, et al. (2015) Peripheral and central effects of repeated social defeat stress: monocyte trafficking, microglial activation, and anxiety. Neuroscience 289:429–42.
- 73. Doorduin J, Klein HC, Dierckx RAJO, de Vries EFJ (2010) HSV-1 induced behavioural changes and microglia cell activation are inhibited by antipsychotics. Eur J Nucl Med Mol Imaging 37:S387.
- 74. Dobos N, De Vries EFJ, Kema IP, et al. (2012) The role of indoleamine 2,3-dioxygenase in a mouse model of neuroinflammation-induced depression. J Alzheimer's Dis 28:905–915.
- 75. Hannestad J, Gallezot JD, Schafbauer T, et al. (2012) Endotoxin-induced systemic inflammation activates microglia: [11C]PBR28 positron emission tomography in nonhuman primates. Neuroimage 63:232–239.
- 76. Lammertsma AA, Hume SP (1996) Simplified reference tissue model for PET receptor studies. Neuroimage 4:153–158.
- 77. Logan J (2000) Graphical analysis of PET data applied to reversible and irreversible tracers. Nucl Med Biol 27:661–670.
- 78. Lyoo CH, Ikawa M, Liow J-S, et al. (2015) Cerebellum can serve as a pseudo-reference region in Alzheimer's disease to detect neuroinflammation measured with PET radioligand binding to translocator protein (TSPO). J Nucl Med 701–707.

- 79. Schuitemaker A, van der Doef TF, Boellaard R, et al. (2012) Microglial activation in healthy aging. Neurobiol Aging 33:1067–1072.
- 80. Owen DR, Guo Q, Kalk NJ, et al. (2014) Determination of [(11)C]PBR28 binding potential in vivo: a first human TSPO blocking study. J Cereb Blood Flow Metab 34:989–94.
- 81. Folkersma H, Boellaard R, Yaqub M, et al. (2011) Widespread and prolonged increase in (R)-(11)C-PK11195 binding after traumatic brain injury. J Nucl Med 52:1235–1239.
- 82. Su Z, Herholz K, Gerhard A, et al. (2013) [11C]-(R)PK11195 tracer kinetics in the brain of glioma patients and a comparison of two referencing approaches. Eur J Nucl Med Mol Imaging 40:1406–1419.
- 83. Schain M, Benjaminsson S, Varna K, et al. (2013) Arterial input function derived from pairwise correlations between PET-image voxels. 1058–1065.
- 84. Kanegawa N, Schain M, Collste K, et al. (2015) Towards less-invasive quantification of [11C]PBR28: Image-derived and population-based input functions. Int. Conf. Quantif. Brain Funct. with PET
- 85. Bilbo SD, Schwarz JM (2009) Early-life programming of later-life brain and behavior: a critical role for the immune system. Front Behav Neurosci 3:14.
- 86. Jučaite A, Cselényi Z, Arvidsson A, et al. (2012) Kinetic analysis and test-retest variability of the radioligand [11C](R)-PK11195 binding to TSPO in the human brain a PET study in control subjects. EJNMMI Res 2:15.
- 87. Park E, Gallezot J-D, Delgadillo A, et al. (2015) 11C-PBR28 imaging in multiple sclerosis patients and healthy controls: test-retest reproducibility and focal visualization of active white matter areas. Eur J Nucl Med Mol Imaging.
- 88. Collste K, Forsberg A, Varrone A, et al. Test-retest reproducibility of [11C]PBR28 binding to TSPO in healthy control subjects. Eur. J. Nucl. Med. Mol. Imaging
- 89. Lavisse S, Guillermier M, Herard A-S, et al. (2012) Reactive Astrocytes Overexpress TSPO and Are Detected by TSPO Positron Emission Tomography Imaging. J Neurosci 32:10809–10818.
- 90. Toth M, Little P, Arnberg F, et al. (2015) Acute neuroinflammation in a clinically relevant focal cortical ischemic stroke model in rat: longitudinal positron emission tomography and immunofluorescent tracking. Brain Struct Funct.
- 91. Asadabadi M, Mohammadi M-R, Ghanizadeh A, et al. (2013) Celecoxib as adjunctive treatment to risperidone in children with autistic disorder: a randomized, double-blind, placebo-controlled trial. Psychopharmacology (Berl) 225:51–9.

- 92. Sommer IE, de Witte L, Begemann M, Kahn RS (2012) Nonsteroidal anti-inflammatory drugs in schizophrenia: ready for practice or a good start? A meta-analysis. J Clin Psychiatry 73:414–9.
- 93. Abbasi S-H, Hosseini F, Modabbernia A, et al. (2012) Effect of celecoxib add-on treatment on symptoms and serum IL-6 concentrations in patients with major depressive disorder: randomized double-blind placebo-controlled study. J Affect Disord 141:308–14.
- 94. Nitta M, Kishimoto T, Müller N, et al. (2013) Adjunctive use of nonsteroidal antiinflammatory drugs for schizophrenia: a meta-analytic investigation of randomized controlled trials. Schizophr Bull 39:1230–41.
- 95. Oya K, Kishi T, Iwata N (2014) Efficacy and tolerability of minocycline augmentation therapy in schizophrenia: a systematic review and meta-analysis of randomized controlled trials. Hum Psychopharmacol 29:483–91.
- 96. Laan W, Grobbee DE, Selten J-P, et al. (2010) Adjuvant aspirin therapy reduces symptoms of schizophrenia spectrum disorders: results from a randomized, double-blind, placebo-controlled trial. J Clin Psychiatry 71:520–527.
- 97. Jucaite A, Svenningsson P, Rinne JO, et al. (2015) Effect of the myeloperoxidase inhibitor AZD3241 on microglia: a PET study in Parkinson's disease. Brain.
- 98. Deakin J, Lennox BR, Zandi MS (2014) Antibodies to the N-methyl-D-aspartate receptor and other synaptic proteins in psychosis. Biol Psychiatry 75:284–91.
- 99. Raison CL (2013) A Randomized Controlled Trial of the Tumor Necrosis Factor Antagonist Infliximab for Treatment-Resistant Depression: The Role of Baseline Inflammatory Biomarkers. JAMA Psychiatry 70:31-41

Figure legend

Fig 1 Cross-species TSPO imaging using [¹¹C]PBR28: SUV images of A) mouse B) rat and C) a healthy human subject. All PET images are overlaid over MR images. Courtesy of the PET centre, Karolinska Institutet