Otolith shape variations between artificially stocked and autochthonous pikeperch (*Sander lucioperca*)

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Abstract

11 Fish stocking is one of the most widespread and frequent management strategies in freshwater 12 systems. However, the contribution of stocked fish to the population is seldom investigated, and hence 13 the effectiveness of this strategy is virtually unknown for many populations. Understanding the 14 contribution of stocked fish into the population is crucial because it allows the disentanglement of the 15 confounding effects generated by allochthonous individuals into the estimation of survival and growth rates of the population. To discriminate between the allochthonous and autochthonous 16 17 individuals in a population, the shape of sagittae otoliths from pikeperch (Sander lucioperca) where 18 compared. Results indicated significant differences among stocked and non-stocked fish, with the 19 former having smaller and wider sagittae otoliths than the later. Our results suggest that this technique 20 can be used to discriminate the natal origin of fish in a much faster and cheaper way than commonly 21 used techniques.

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23 Key-words: fisheries, fish, freshwater, morphometry, stock discrimination

24 **1. Introduction**

25 Fish are extremely important components of aquatic environments. They provide essential ecosystem services to humans, including demand-derived ecosystem services such as the food resources and 26 27 recreational activities, as well as fundamental ecosystem services such as resilience and nutrient 28 cycling that ultimately affect water quality which is critical for humans (Holmlund and Hammer, 29 1999). With increasing threats to fish communities like habitat degradation, stock exploitation, pollution and climate change (Arthington et al., 2016; Gordon et al., 2018), many fish populations 30 31 require operational management to mitigate the detrimental effects of human activities (Agostinho et 32 al., 2008; Hansson et al., 1997; Welcomme and Bartley, 1998). Fish stocking (i.e. introduction of 33 allochthonous individuals into the ecosystem) is one of the most widespread, frequent and cost-34 efficient management strategy adopted by stakeholders in freshwater systems (Hunt et al., 2017), which usually spend relatively large part of their budget in this activity (Welcomme and Bartley, 35 36 1998).

Given that the rearing conditions are fundamentally different from the wild conditions, and the 37 homogeneity of behavioral and other biological traits of artificially bred fish, it is expected that the 38 39 survival rates of stocked fish in the wild may differ considerably from the autochthonous individuals (Adriaenssens and Johnsson, 2013). Consequently, the managed populations receive an influx of 40 41 individuals displaying unique set of characteristics that likely differ from the autochthonous fish, but 42 their performance in the wild is not known, nor investigated, and hence the effectiveness of this strategy is virtually unknown in many cases. This knowledge gap is not easily disentangled because 43 44 the discrimination between stocked and wild fish is not trivial. Different techniques can be used to mark stocked fish, including fin clipping, otolith staining and micro wire, VIE (Visual Implant 45 Elastomer) and PIT (Passive Integrated Transponder) tags, but the results are not always satisfactory 46 47 due to decreased capacity of mark detection, tag losses, differences detection and retention between sexes, influences on social behavioral and decreased survival rate of marked fish, biasing the stock 48

49 assessment (Buckley and Blankenship, 1990; Halačka et al., 2019; Jungwirth et al., 2019; Skalski et
50 al., 2009; Šmejkal et al., 2019; Svåsand et al., 1990; Taylor et al., 2005; Utter, 1998).

51 Fish otolith structures are considered natural tags (Bouchoucha et al., 2018; Campana, 1999; 52 Campana et al., 2000; Hamer et al., 2003; L'Abée-Lund and Jensen, 1993) and are excellent structures 53 to retrieve data on the chronologies of life history of fish. Otoliths grow continuously throughout 54 accretions of calcium carbonate (CaCO₃) aragonitic crystals on protein layers (Campana and Neilson, 55 1985), with the growth being acellular and metabolically inert, meaning that these natural tags are 56 neither reworked nor reabsorbed, even during times of starvation (Campana and Neilson, 1985). 57 Therefore the life history trajectory is preserved into the structure allowing researchers to trace back 58 important events on fish life. Otolith shapes are species-specific conservative and have been 59 successfully used in fisheries to discriminate fish stocks (Campana and Casselman, 1993; DeVries et 60 al., 2002; Paul et al., 2013; Schulz-Mirbach et al., 2008). The otolith morphometry is influenced by both genetics and the environment where the fish lives (Vignon and Morat, 2010), with individuals 61 from the same population but living in different localities having different otolith shapes (Cerna et 62 63 al., 2019; Costa et al., 2018; Pérez and Fabré, 2013). Therefore, otolith shape is a powerful tool to 64 identify spatial distribution and natal origin of fish, because it integrates multiple variables in one 65 conservative structure.

Pikeperch (*Sander lucioperca*) is a very popular sportfishing target in European inland waters, but the management of its population is often inadequate, leading to overexploitation which causes significant ecological and economic impacts (Saulamo and Thoresson, 2005; Specziár and Ero"s, 2015). Pikeperch reproduction and overwinter survival of the newborns are highly sensitive and dependent on various environmental conditions (Blabolil et al., 2016; Kokkonen et al., 2019; Lappalainen et al., 2009). Therefore, stocking became a recurrent measure to mitigate the impacts of poor management practices and natural recruitment failure (Specziár and Turcsányi, 2017). 73 Proper description of the contribution of stocked fish into wild populations is of paramount importance because it would allow researchers to have a deeper understanding of the population 74 75 dynamics, stocking dependence, and provide adjustments on growth and survival rates of fish cohorts 76 taking into consideration their natal origin. Despite being widely used to discriminate wild stocks 77 (Campana and Casselman, 1993), otolith shapes are seldom used to differentiate autochthonous and allochthonous individuals. In this context, the present study aimed at applying shape analysis to 78 79 discriminate between wild and stocked individuals of an ecologically and economically relevant 80 species (S. lucioperca) using an accurate and low-cost analytical tool.

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82 2. Material and Methods

Fish sampling were conducted separately for wild and stocked *S. lucioperca*. Wild individuals came
from the Lipno reservoir (Southwestern, Czechia), while stocked fish came from the rearing facilities
c.a. 150 km from the lake.

Young pikeperch were hatched at Pohořelice Fishery facilities and when they reach 4 cm (total length) they were transported to the shallow rearing pond (Velké Rozběhlo – area = 13.2 ha) (Fig. 1). Fish were kept under semi-natural conditions, being exposed to competition (intra and interspecific) and predators (intracohort cannibalism and terrestrial species – birds and small mammals) and have access to natural food represented mostly by plankton and later by young of the year stone moroco (*Pseudorasbora parva*). In the shallow rearing pond the pikeperch had the company of adult common carps (*Cyprinus carpio*) that were kept under relatively high densities (400 kg/ha).

Wild fish were sampled by European standard multimesh gillnets (CEN - Comité Européen de Normalisation, 2005) in last week of August 2018 in Lipno reservoir, whereas stocked fish were sampled during the fish stocking campaign in just before releasing to Lipno in mid October of the same year. Stocked fish were randomly sampled using a scoop net and immediately euthanized by thermal shock. The standard length (SL) of all fish were measured to the nearest 5mm using a fish 98 ruler. The sex of all individuals sampled in the current study was not determined because they were 99 all sexually immature. In the rearing facilities there was no separation between male and females at 100 the juvenile stage.

Only juvenile fish ranging from 190 to 245 mm of length (SL) were included into the analysis because we could only ascertain that wild fish was not stocked until this size (currently, managers do not stock fish smaller than this size in Lipno reservoir). The growth of wild individuals of the same cohort can vary significantly (van Densen and Vijverberg, 1982) and for that reason, we selected for the analysis only the wild fish with similar sizes of the stocked individuals (i.e. faster growers) to avoid bias due to significant differential growth rates between stocked and wild fish. Additionally, using only juvenile fish avoided bias due to sexual dimorphism on otolith shape (Vaux et al., 2019).

In laboratory the sagittae otoliths of *S. lucioperca* individuals were extracted from the cranial cavity using a scalpel and tweezers. Otoliths were then mechanically cleaned and air-dried before being photographed (15x magnification) under transmitted light using a camera (Optikam B3) mounted on a stereomicroscope (STM 800). Otoliths were positioned over a flat surface with the sulcus facing downwards (i.e. outer face up).

Contours of individual sagittae otoliths were digitally extracted from the photographs using the R 113 114 software (R Core Team, 2020) following the method described by Claude (2008). The standardization 115 of the otoliths positions was performed digitally by aligning them to a common center, orienting the otoliths to remove discrepancies in positioning and scaling to centroid size using the functions 116 coo center, coo rotate and coo slidedirection functions from the Momocs package (Bonhomme et 117 118 al., 2014). Due to otoliths shape irregularities, the outlines were smoothed to reduce the noise of digitization using the function coo smooth from the Momocs package (Bonhomme et al., 2014), 119 Otolith length (O_L), width (O_W), area (O_A) and perimeter (O_P) (Fig. 2) were computed using the 120 package rgeos (Bivand et al., 2019). Eight different shape indices (Burke et al., 2008; Leguá et al., 121 2013; Rashidabadi et al., 2020; Škeljo and Ferri, 2012) were computed for each individual otolith 122

123 (Table 1). The aspect ratio (SI_{AR}) between the otolith length and width expresses the otolith elongation. The circularity index (SI_{CI}), i.e., the ratio between the squared perimeter and the otolith 124 area indicates the circularity of the structure. The ellipticity index (SI_{EL}), i.e., the ratio between the 125 126 difference in otolith length and width and their sum expresses the similarity of the otolith shape to an ellipse. The format factor (SI_{FF}) is the inverse ratio of the squared perimeter of the structure to the 127 squared perimeter of a circle of the same surface. The rectangularity index (SIRE) is the ratio between 128 129 the area and the product of the length and width of the otolith. The roundness index (SIRO) is the ratio 130 between the actual area and the area of a circle of the same circumference. The convexity (SI_{CO}) is 131 the ratio between the perimeter of the convex hull fitted into the otolith outline and the perimeter of 132 the otolith expressing the otolith edge roughness. The solidity (SI_{SO}) is the ratio between the otolith 133 area and the area of the convex hull fitted on the otolith outline expressing the concavity of the otolith. SI_{CO} and SI_{SO} were computed using the functions *coo* convexity and *coo* solidity from the Momocs 134 package respectively (Bonhomme et al., 2014). The use of different shape indices in the same otolith 135 provides numerical evidence to describe spatial format of the structure (Tuset et al., 2003), and hence 136 137 is essential to a robust analysis on the 2D format of the calcified structures.

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140 2.1 Statistical analyses

The mean values, standard error and confidence intervals of the otolith descriptors and shape indices were obtained after data bootstrapping with 1000 bootstrap replicates of indices using the functions *boot* and *boot.ci* from the package *boot* (Canty, 2020). Bootstrapping assigns measures of accuracy to the sample estimates by randomly sampling and replacing data, and hence providing a more precise estimate of the values.

146 The correlations between each of the eight shape indices were tested for correlation (Pearson's 147 correlation) using the *findCorrelation* function in the *caret* package (Kuhn et al., 2020). The non148 correlated indices were then selected for the training of a resilient backpropagation with back tracing neural network using the *neuralnet* function from the *neuralnet* package (Fritsch et al., 2019). The 149 neural network is a type of supervised machine learning tool that is used to predict patterns among 150 151 multiple variables and that does not need to follow the parametric premises of traditional statistical tests (Beck, 2018). This method characterizes the relationships among variables using an arbitrary 152 number of parameters (hidden layers) that are chosen through interactive training with the algorithm, 153 154 and hence, this tool can be classified as a hyper-parameterized non-linear model that can fit a smooth 155 function to any dataset with minimal residual error (Hornik, 1991). The number of hidden layers were 156 selected based on the number of input variables in the model (2/3 of the size of input layers plus the 157 size of output layers). Machine learning tools outperform traditional statistical classifiers in otolith shape analysis and can improve the accuracy of fish stock discrimination studies (Smoliński et al., 158 2019). The relative importance of each variable was assessed by the Olden's algorithm (Olden and 159 Jackson, 2002) using the *olden* function of the *NeuralNetTools* package (Beck, 2018). The relative 160 importance of input variables in neural networks is expressed by the sum of the product of the raw 161 162 input-hidden, hidden-output connection weights, with the values being only interpreted based on the relative sign and magnitude between the explanatory variables (Olden and Jackson, 2002). The 163 dataset was randomly divided in two (training (60%) and testing (40%)) and the accuracy of the model 164 165 was assessed by comparing the observed values with the ones obtained by the model prediction using. The recall (i.e. sensitivity) and precision of the model were evaluated using the recall and precision 166 functions from the caret package (Kuhn et al., 2020). The neural network analysis were performed in 167 168 R version 3.5.1 (R Core Team, 2020).

Additionally, the 2D shapes of the pikeperch sagittae otoliths were analyzed using the Elliptical Fourier Analysis (*efourier* function) without the normalization of the coefficients. The number of harmonics used was defined by applying the *calibrate_harmonicpower_efourier* function. The Elliptical Fourier Analysis is commonly used to describe the fish otolith shapes with a high degree of precision and success in capturing the shape of the otoliths, even those with rough contours (Assis et
al., 2020; Gagliano and McCormick, 2004; Karahan et al., 2014; Lestrel, 1997; Mahé et al., 2019;

Mérigot et al., 2007; Rashidabadi et al., 2020; Rodgveller et al., 2017; Smoliński et al., 2019).
The first coefficients derived from the first harmonic were excluded from the analysis because the

outlines reconstructed from these coefficients are simple ellipses used to standardize each outline for 177 size, orientation and starting point (Crampton, 1995). The normality of the coefficients were tested 178 using the shapiro.test function from the stats package (R Core Team, 2020). The coefficients that did 179 180 not follow a normal distribution were excluded from the analysis. The allometric relationships between otolith length and each of the Fourier descriptors were tested using the ANCOVA test. The 181 182 descriptors that had a correlation between otolith length and the Fourier descriptors were excluded from the analysis to remove the effects of allometry in otolith shape. Otolith length was used instead 183 184 of fish length because it is less prone to errors (Campana and Casselman, 1993; Rodgveller et al., 2017). 185

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The remainder Fourier descriptors were used to compare the shape of the otoliths between autochthonous and allochthonous fish applying a multivariate analysis of variance (MANOVA) using the *MANOVA* function. A linear discriminant analysis – LDA (*LDA* function) was applied to test the correct classification of the otolith shapes based on the fish origin. The mean shape of the otoliths from stocked and wild fish was obtained using the *mshapes* function and the differences between the average shapes were obtained using the *coo_ruban* function. All analysis based on the Fourier transformation method were performed on the *Momocs* package in R (Bonhomme et al., 2014).

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195 **3. Results**

196 Overall, 158 otoliths were analysed (wild = 122, stocked = 36), with stocked fish having slightly 197 smaller body than wild individuals, with otolith area, length, width and perimeter being larger in the former group (Table 2). All shape indices but SI_{CI} were larger in the wild pikeperch, with the strongest
differences between stocked and wild pikeperch being recorded for SI_{CI}, SI_{CO}, SI_{FF} and SI_{SO}, whereas
SI_{AR}, SI_{EL}, SI_{RE} and SI_{RO} having less prominent differences (Table 3).

201 The correlation analysis among the shape indices plus O_P (otolith perimeter) indicated that SI_{AR}, SI_{CI}, 202 SIFF and SIRO were correlated and hence, OP, SICO, SIEL, SIRE and SISO were selected for the training of the neural network. The neural network used 4 hidden layers based on the selection of variables, 203 204 with only one hidden layer (H2) being positively linked to the output layer. The strongest positive 205 links were observed between SIEL and the fourth hidden layer (H4) and SIRE with the third hidden layer (H3), whereas the strongest negative links were observed between OP and the fourth H4 and 206 SI_{RE} and H4 (Fig. 3). The relative importance of the input variables shows that the most important 207 208 variables were O_P (423.93), SI_{EL} (-368.60), SI_{RE} (-228.40), SI_{CO} (-192.14) and SI_{SO} (48.86) 209 respectively. The neural network showed a global accuracy of 79.59%, with the accuracy of the 210 classification of wild fish being 94.87%, whereas for the stocked fish of 10.00%. The model precision 211 was 0.78 and the recall 0.80.

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The elliptical Fourier analysis was able to describe the outlines of *S. lucioperca* sagittae otoliths with 99.9% accuracy using 34 elliptical harmonics (136 Fourier descriptors). After the removal of the first coefficients of the first harmonics (4 Fourier descriptors), the non-normally distributed Fourier descriptors (30) and the descriptors with allometric influence (5), a total of 102 Fourier descriptors were used in the analysis. The MANOVA test revealed that the otolith shapes were significantly different between stocked and wild fish (F = 1.736, p < 0.05). Additionally, the LDA (linear discriminant analysis) was able to correctly classify 77.9% of the otoliths based on fish origin, with
89.7% accuracy on the wild fish group and 32.0% on the stocked group.

The statistical tests support the visual inspection of the average otolith outlines from stocked and wild fish, where it is possible to detect differences on shapes. The differences are especially pronounced in the otolith rostrum, which is shorter in the otoliths from wild pikeperch in comparison with the stocked individuals. The zone between the antirostrum and the postero-dorsal angle of in the dorsal portion of otoliths is also different, with higher roughness being recorded in the otoliths from the stocked fish and smoother contours in the wild pikeperch otoliths (Fig. 4).

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231 4. Discussion

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234 Our results indicated that autochthonous and allochthonous fish have dissimilar otolith shapes, suggesting that this feature can be applied in analytical tools to identify the natal origin of pikeperch 235 populations. In general, the otoliths from the stocked pikeperch were bigger, rougher and with a 236 longer rostrum compared to the wild individuals. Otolith shapes are influenced by many factors 237 (Mahé et al., 2019; Mille et al., 2016; Vignon, 2012; Vignon and Morat, 2010) and without 238 experimental studies it is difficult to disentangle the effects of multiple drivers. In this study we did 239 not attempted to investigate the drivers of otolith shape variation, but rather to test if there is any 240 241 signal of these drivers into the otolith shapes of fish of different natal origins. Using two different 242 methods and descriptors we were able to detect significant differences on the otolith shapes of 243 autochthonous and allochthonous individuals. The neural network applied to the shape indices spot differences between the groups, with the 5 variables being important to classify the natal origin of 244 pikeperch individuals. Overall, the neural network was slightly more accurate than the LDA model, 245 with the accuracy at classifying the wild individuals being higher in the former, whereas the accuracy 246

247 of stocked individuals being higher in the later model. The higher accuracy of machine learning methods over more traditional statistical tests observed in this study corroborates the finds of 248 249 Smoliński et al., (2019), but, in our case, the difference seems to be small and not true for all groups 250 (i.e. stocked fish). In both methods, the classification of wild individuals was much higher than the 251 stocked fish. It is a bit surprising that the accuracy of origin determination was higher with the wild fish group. The higher accuracy of the models in this group of fish might suggest that the sample size 252 253 is an important factor to improve the accuracy of the methods, given that the number of wild 254 individuals used in this study is higher than stocked fish, and hence, in future studies the sample sizes 255 should be larger whenever it is feasible. Interestingly all individuals in stocked group were kept in 256 much more homogeneous condition than the wild fish. Alternatively, this result might indicate that the rearing conditions lead to a higher variation on the otolith shape on S. lucioperca. Even though 257 258 the rearing conditions try to create homogeneous conditions for the fish, they do not allow the fish to freely explore a broad range of micro-habitats that could theoretically allow the fish to actively seek 259 the optimal conditions for its growth. Given that the stocked fish were reared in semi-intensive 260 261 conditions (shallow ponds), they can be more vulnerable to other types of stresses than wild individuals, which can cause shifts on the otolith shape (Fernandez-Jover and Sanchez-Jerez, 2015; 262 263 Koeberle et al., 2020; Vignon, 2018), because the later can perform vertical migrations to cope with 264 temperature shifts (Lehtonen, 1983; Lind, 1977; Saulamo and Lappalainen, 2007). Also, given that captive fish inhabited different ponds before reaching Lipno reservoir, their growth and otolith shape 265 could be influenced by these different environments in a non-linear way (i.e. individual variability on 266 growth in different habitats), which might be reflected by the slightly higher diversity on the otolith 267 shapes in this group of individuals. 268

Differences in the food items consumption can have a remarkable effect on otolith shape (Mille et al., 2016), and therefore can also be one of the factors influencing the observed differences between autochthonous and allochthonous fish, given that the food intake and food items are likely to be 272 dissimilar between the groups. Mille et al., (2016) stated that the food item is more relevant than food amount in affecting the otolith shape, therefore our results can indicate that fish reared in captivity, 273 274 even though in semi-intensive conditions, have different diet composition than wild fish. Wild fish 275 could have explored different habitats and areas of the reservoir throughout the year, adjusting their food item selection according to their needs (Huuskonen et al., 2019), while stocked fish did not have 276 available a diverse set of food types in the ponds. In the same time the diet of pikeperch of the given 277 size in the Lipno reservoir is also quite uniform (100% percid fish fry, Vašek et al., (2018)), which is 278 279 believed to be the optimal diet for young pikeperch (Dörner et al., 2007).

280 A recent study showed that otolith shape is more sensitive to water temperature than to food quality 281 (Mahé et al., 2019). This fact can also be behind the observed dissimilarities on otolith shapes between stocked and wild pikeperch in Lipno reservoir. The shallow depths of the rearing facilities compared 282 283 to the reservoir conditions might have constrained fish to a broader thermal range of conditions than in the reservoir (due to the impossibility of spatio-temporal migrations to cope with thermal 284 fluctuations). Given that pikeperch do move to find better living conditions throughout the year 285 286 (Huuskonen et al., 2019), the individuals in the wild might experienced narrower thermal ranges than stocked pikeperch, which might be reflected on the higher shape similarity among wild than stocked 287 288 group of fish. The water pH can be another variable behind the observed differences in otolith shape between stocked and wild pikeperch individuals, given that lower pH values are associated with 289 bigger and rougher otoliths (Coll-Lladó et al., 2018; Holmberg et al., 2019), which are the most 290 striking differences between the two groups in our study. Alternatively, higher food availability during 291 292 early life (likely higher for stocked fish) can also cause higher otolith roughness (Hüssy, 2008).

Reared fish have a higher prevalence of otolith shape and mineral anomalies (vaterites) that can cause important hearing impairments for the fish (Oxman et al., 2007; Reimer et al., 2016). The prevalence of these anomalies is linked to stressful conditions present in the rearing facilities, such as high population densities, temperature fluctuation, noise, vibration, diseases, poor water quality and 297 nutrition (Sweeting et al., 2004). In the present study, we did not detect the presence of vaterites in the stocked nor in the wild fish, but otolith shape still differed between these fish. Other studies found 298 299 differences in otolith shapes in areas closely associated to fish farms but with low population density 300 (Fernandez-Jover and Sanchez-Jerez, 2015), suggesting that water quality might be linked to changes 301 in otolith shapes. High population density can significantly affect fish growth and bias fisheries' assessments (Lorenzen and Enberg, 2002; Martino et al., 2019). Due to the tight connection between 302 303 somatic growth, food availability and otolith growth trajectory (Mahé et al., 2019; Vignon, 2012) it 304 is expected that population density can significantly influence the otolith shape as well, however the 305 role of population density dependence on otolith shape is yet to be scrutinized experimentally.

306 Many stock assessments studies neglect the share of stocked and wild fish in the population, which can create biases in the interpretation of the population parameters extracted for the whole 307 308 metapopulation. For instance, a higher survival and cumulative dominance of stocked fish in the 309 population could wrongly indicate an accelerated somatic growth rate and early maturation of fish in the population, given that fish in captivity often grow faster and mature earlier than wild individuals 310 311 (McDermott et al., 2011; Zupa et al., 2017), which is not possible to see in this study due to the selection of wild fish with comparable sizes with the stocked individuals to avoid the bias of the 312 presence of slower and much smaller fish in the wild group (van Densen and Vijverberg, 1982). This 313 314 option allow us to compare individuals with more similar conditions, because the slow growing pikeperch are in fact much smaller than the fast growing individuals in the studied area (Jůza et al., 315 2013), but refrain us from assessing the full spectrum of otolith shapes in the wild population. 316 317 Nevertheless, differences in fish growth could potentially bias the assessment of long-term patterns of a fish stock, masking the effects of other stressors like environmental carrying capacity, density 318 319 dependence, fishing pressure, shifts in management rules and climate change. In this regard, the 320 continuous monitoring of the contribution of stocked fish in the population could provide ways to 321 cope with the potential differences in conditions of the groups in the population.

322 In the present study, we did not perform any genetic analysis on the analyzed fish (from both autochthonous and allochthonous individuals), therefore the genetic influence on the pikeperch 323 324 otolith shape cannot be determined. On the other hand, this method might not be the best option to 325 discriminate the fish origin in this case, because the ecosystem is continuously stocked and pikeperch 326 natural recruitment is successful, and hence the autochthonous fish genetics might contain the genetic 327 material of the allochthonous population due to the breeding between fish of different origins. 328 Additionally, genetics can be outperformed by other methods in stock discrimination studies 329 (Marengo et al., 2017).

Fish sex is believed to influence the otolith shape (Campana and Casselman, 1993), but recent studies suggest that this can be a consequence of differences in the way of life between males and females (Parmentier et al., 2018; Vaux et al., 2019). In the present study, individuals of both sexes were pooled together because they were not adults yet and thus no behavioral sexual dimorphism would be expected. So the effects of sex on otolith shape in this study are unlikely.

It is important to highlight that in our study, we focused only in fish of similar sizes with confirmed 335 336 natal origins and we did not attempted to identify the natal origin fish of larger sizes and with 337 unknown origin because we do not know if the observed differences in otolith shape are still 338 noticeable in stocked individuals after a certain period of residence in the lake. To classify older fish 339 of unknown origin and already living into the lake it is important to conduct further studies to describe 340 the consequences of habitat shift on the otolith shape through ontogeny, possibly combining shape 341 analysis with other analytical tools. Otolith shape is successful in identifying the natal origin of fish 342 in stock discrimination studies (Brophy et al., 2016; Burke et al., 2008; Campana and Casselman, 343 1993; Libungan et al., 2015; Paul et al., 2013; Schade et al., 2019). Given that the otoliths are 344 conservative structures (Campana and Neilson, 1985), the shape signal of early ages will invariably be permanently record on the structure of older fish and detectable in stocked fish even after years 345

living in the wild (Negus, 1999; Vignon, 2018; Volk et al., 1999), and hence, otolith shape analysis is
being successfully used to differentiate mixed-stocks.

Many mixed-stock discrimination studies are performed in the marine areas where there is no artificial stocking being conducted (Burke et al., 2008; DeVries et al., 2002; Paul et al., 2013; Vignon and Morat, 2010). They are targeted to detect natal areas of stocks with complex metapopulation structure, while our study targets a freshwater species that is artificially stocked by managers and do not perform long distance migrations. The scientific literature is much scarcer in this type of conditions (but see Pereira et al., (2019)) and our results shows that the otolith shape analysis is a cost efficient tool that can be successfully applied in actively managed freshwater fish stocks.

355 Our results highlights that the detection of the natal origin of fish based on their otolith shape is 356 promising and the application of this method can provide a low cost and effective tool for managers to assess the efficiency of the stocking regime in their systems. Schade et al., (2019) compared the 357 efficiency of four common stock discrimination tools and found out that the otolith shape analysis 358 and stable oxygen analysis on the otolith nuclei were the most effective to distinguish the natal origin 359 360 of mixed-stocks. However, sometimes the combination of shape analysis with other tools are needed to differentiate the origin of fish in the population (Marengo et al., 2017; Schade et al., 2019). For 361 362 instance, the use of microelemental composition of fish otoliths can precisely indicate the origin of 363 the fish in freshwater (Avigliano et al., 2017; Lazartigues et al., 2017; Spurgeon et al., 2018), but its application is financially costly, whereas the shape analysis is much more affordable, especially if 364 365 used routinely. However, the combination of both tools could theoretically create the mechanisms to cut the costs in long-term, by developing a database of shape information that could be used to 366 determine the natal origin of the fish using the different classification tools. It is also crucial to 367 368 understand how the otolith growth trajectory is affected after the stocked fish is trans-located to the lake. The correct stock discrimination can uncover many hidden signals in fish population structure, 369 which in turn will allow scientists to better guide stakeholders to improved management plans. 370

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382 5. References

- Adriaenssens, B., Johnsson, J.I., 2013. Natural selection, plasticity and the emergence of a behavioural syndrome in the wild. Ecol. Lett. 16, 47–55. https://doi.org/10.1111/ele.12011
- Agostinho, A.A., Pelicice, F.M., Gomes, L.C., 2008. Dams and the fish fauna of the Neotropical region: impacts and management related to diversity and fisheries. Braz. J. Biol. 68, 1119–1132. https://doi.org/10.1590/S1519-69842008000500019
- Arthington, A.H., Dulvy, N.K., Gladstone, W., Winfield, I.J., 2016. Fish conservation in freshwater and marine realms: status, threats and management. Aquat. Conserv. Mar. Freshw. Ecosyst. 26, 838–857. https://doi.org/10.1002/aqc.2712
- Assis, I.O., da Silva, V.E.L., Souto-Vieira, D., Lozano, A.P., Volpedo, A.V., Fabré, N.N., 2020. Ecomorphological patterns in otoliths of tropical fishes: assessing trophic groups and depth strata preference by shape. Environ. Biol. Fishes 103, 349–361. https://doi.org/10.1007/s10641-020-00961-0
- Avigliano, E., Domanico, A., Sánchez, S., Volpedo, A.V., 2017. Otolith elemental fingerprint and scale and otolith morphometry in *Prochilodus lineatus* provide identification of natal nurseries. Fish. Res. 186, 1–10. https://doi.org/10.1016/j.fishres.2016.07.026
- Beck, M.W., 2018. NeuralNetTools: Visualization and Analysis Tools for Neural Networks. J. Stat. Softw. 85, 1–20. https://doi.org/10.18637/jss.v085.i11

383

- 384 Bivand, R., Rundel, C., Pebesma, E., Stuetz, R., Hufthammer, K.O., Giraudoux, P., Davis, M.,
- 385 Santilli, S., 2019. rgeos: Interface to Geometry Engine Open Source ('GEOS').
 - Blabolil, P., Ricard, D., Peterka, J., Říha, M., Jůza, T., Vašek, M., Prchalová, M., Čech, M., Muška, M., Seďa, J., Mrkvička, T., Boukal, D.S., Kubečka, J., 2016. Predicting asp and pikeperch recruitment in a riverine reservoir. Fish. Res., Ecology of Fish in Lakes and Reservoirs 173, 45–52. https://doi.org/10.1016/j.fishres.2015.08.003

386

- 387 Bonhomme, V., Picq, S., Gaucherel, C., Claude, J., 2014. Momocs: Outline Analysis Using R. J. Stat.
- 388 Softw. 56, 1–24. https://doi.org/10.18637/jss.v056.i13
 - Bouchoucha, M., Pécheyran, C., Gonzalez, J.L., Lenfant, P., Darnaude, A.M., 2018. Otolith fingerprints as natural tags to identify juvenile fish life in ports. Estuar. Coast. Shelf Sci. 212, 210–218. https://doi.org/10.1016/j.ecss.2018.07.008
 - Brophy, D., Haynes, P., Arrizabalaga, H., Fraile, I., Fromentin, J.M., Garibaldi, F., Katavic, I., Tinti,
 F., Karakulak, F.S., Macías, D., Busawon, D., Hanke, A., Kimoto, A., Sakai, O., Deguara, S.,
 Abid, N., Santos, M.N., 2016. Otolith shape variation provides a marker of stock origin for
 north Atlantic bluefin tuna (*Thunnus thynnus*). Mar. Freshw. Res. 67, 1023–1036.
 https://doi.org/10.1071/MF15086

- 390 Buckley, R.M., Blankenship, H.L., 1990. Internal extrinsic identification systems: Overview of
- 391 implanted wire tags, otolith marks, and parasites, in: Parker, N.C., Giorgi, A.E., Heidinger,

- 392 R.C., Jester, D.B., Prince, E.D., Winans, G.A. (Eds.), American Fisheries Society Symposium.
- Bethesda, Maryland, pp. 178–182.
 - Burke, N., Brophy, D., King, P.A., 2008. Otolith shape analysis: its application for discriminating between stocks of Irish Sea and Celtic Sea herring (*Clupea harengus*) in the Irish Sea. ICES J. Mar. Sci. 65, 1670–1675. https://doi.org/10.1093/icesjms/fsn177
 - Campana, S.E., 1999. Chemistry and composition of fish otoliths: pathways, mechanisms and applications. Mar. Ecol. Prog. Ser. 188, 263–297. https://doi.org/10.3354/meps188263

- 395 Campana, S.E., Casselman, J.M., 1993. Stock Discrimination Using Otolith Shape Analysis. Can. J.
- 396 Fish. Aquat. Sci. 50, 1062–1083. https://doi.org/10.1139/f93-123

- 398 Campana, S.E., Chouinard, G.A., Hanson, J.M., Fréchet, A., Brattey, J., 2000. Otolith elemental
- 399 fingerprints as biological tracers of fish stocks. Fish. Res. 46, 343–357.
- 400 https://doi.org/10.1016/S0165-7836(00)00158-2
 - Campana, S.E., Neilson, J.D., 1985. Microstructure of Fish Otoliths. Can. J. Fish. Aquat. Sci. 42, 1014–1032. https://doi.org/10.1139/f85-127
 - Canty, A., 2020. boot: Bootstrap Functions (Originally by Angelo Canty for S).
 - CEN Comité Européen de Normalisation, 2005. Water quality Sampling of fish with multi-mesh gillnets (CEN 14757). Brussels.
 - Cerna, F., Saavedra-Nievas, J.C., Plaza-Pasten, G., Niklitschek, E., Morales-Nin, B., 2019. Ontogenetic and intraspecific variability in otolith shape of anchoveta (*Engraulis ringens*) used to identify demographic units in the Pacific Southeast off Chile. Mar. Freshw. Res. 70, 1794–1804. https://doi.org/10.1071/MF18278
 - Claude, J., 2008. Morphometrics with R, Use R! Springer-Verlag, New York. https://doi.org/10.1007/978-0-387-77789-4
 - Coll-Lladó, C., Giebichenstein, J., Webb, P.B., Bridges, C.R., de la Serrana, D.G., 2018. Ocean acidification promotes otolith growth and calcite deposition in gilthead sea bream (*Sparus aurata*) larvae. Sci. Rep. 8, 8384. https://doi.org/10.1038/s41598-018-26026-y
 - Costa, R.M.R. da, Fabré, N.N., Amadio, S.A., Tuset, V.M., Costa, R.M.R. da, Fabré, N.N., Amadio, S.A., Tuset, V.M., 2018. Plasticity in the shape and growth pattern of asteriscus otolith of black prochilodus *Prochilodus nigricans* (Teleostei: Characiformes: Prochilodontidae) freshwater Neotropical migratory fish. Neotropical Ichthyol. 16. https://doi.org/10.1590/1982-0224-20180051
 - Crampton, J.S., 1995. Elliptic Fourier shape analysis of fossil bivalves: some practical considerations. Lethaia 28, 179–186. https://doi.org/10.1111/j.1502-3931.1995.tb01611.x
 - DeVries, D.A., Grimes, C.B., Prager, M.H., 2002. Using otolith shape analysis to distinguish eastern Gulf of Mexico and Atlantic Ocean stocks of king mackerel. Fish. Res. 57, 51–62. https://doi.org/10.1016/S0165-7836(01)00332-0

- Dörner, H., Hülsmann, S., Hölker, F., Skov, C., Wagner, A., 2007. Size-dependent predator-prey relationships between pikeperch and their prey fish. Ecol. Freshw. Fish 16, 307–314. https://doi.org/10.1111/j.1600-0633.2006.00223.x
- Fernandez-Jover, D., Sanchez-Jerez, P., 2015. Comparison of diet and otolith growth of juvenile wild fish communities at fish farms and natural habitats. ICES J. Mar. Sci. 72, 916–929. https://doi.org/10.1093/icesjms/fsu153
- Fritsch, S., Guenther, F., Wright, M.N., Suling, M., Mueller, S.M., 2019. neuralnet: Training of Neural Networks.
- Gagliano, M., McCormick, M.I., 2004. Feeding history influences otolith shape in tropical fish. Mar. Ecol. Prog. Ser. 278, 291–296. https://doi.org/10.3354/meps278291
- Gordon, T. a. C., Harding, H.R., Clever, F.K., Davidson, I.K., Davison, W., Montgomery, D.W., Weatherhead, R.C., Windsor, F.M., Armstrong, J.D., Bardonnet, A., Bergman, E., Britton, J.R., Côté, I.M., D'agostino, D., Greenberg, L.A., Harborne, A.R., Kahilainen, K.K., Metcalfe, N.B., Mills, S.C., Milner, N.J., Mittermayer, F.H., Montorio, L., Nedelec, S.L., Prokkola, J.M., Rutterford, L.A., Salvanes, A.G.V., Simpson, S.D., Vainikka, A., Pinnegar, J.K., Santos, E.M., 2018. Fishes in a changing world: learning from the past to promote sustainability of fish populations. J. Fish Biol. 92, 804–827. https://doi.org/10.1111/jfb.13546
- Halačka, K., Mareš, J., Vetešník, L., Blabolil, P., 2019. Application of the alizarin red S mass marking technique and its detection in stocked asp (*Leuciscus aspius*). Biologia (Bratisl.) 74, 1359–1362. https://doi.org/10.2478/s11756-019-00247-6
- Hamer, P.A., Jenkins, G.P., Gillanders, B.M., 2003. Otolith chemistry of juvenile snapper *Pagrus auratus* in Victorian waters: natural chemical tags and their temporal variation. Mar. Ecol. Prog. Ser. 263, 261–273. https://doi.org/10.3354/meps263261
- Hansson, S., Arrhenius, F., Nellbring, S., 1997. Benefits from fish stocking—experiences from stocking young-of-the-year pikeperch, Stizostedion lucioperca L. to a bay in the Baltic Sea. Fish. Res. 32, 123–132. https://doi.org/10.1016/S0165-7836(97)00050-7
- Holmberg, R.J., Wilcox-Freeburg, E., Rhyne, A.L., Tlusty, M.F., Stebbins, A., Jr, S.W.N., Honig, A., Johnston, A.E., Antonio, C.M.S., Bourque, B., Hannigan, R.E., 2019. Ocean acidification alters morphology of all otolith types in Clark's anemonefish (*Amphiprion clarkii*). PeerJ 7, e6152. https://doi.org/10.7717/peerj.6152
- Holmlund, C.M., Hammer, M., 1999. Ecosystem services generated by fish populations. Ecol. Econ. 29, 253–268. https://doi.org/10.1016/S0921-8009(99)00015-4
- Hornik, K., 1991. Approximation capabilities of multilayer feedforward networks. Neural Netw. 4, 251–257. https://doi.org/10.1016/0893-6080(91)90009-T
- Hunt, T.L., Scarborough, H., Giri, K., Douglas, J.W., Jones, P., 2017. Assessing the costeffectiveness of a fish stocking program in a culture-based recreational fishery. Fish. Res., Fisheries Enhancement 186, 468–477. https://doi.org/10.1016/j.fishres.2016.09.003
- Hüssy, K., 2008. Otolith shape in juvenile cod (*Gadus morhua*): Ontogenetic and environmental effects. J. Exp. Mar. Biol. Ecol. 364, 35–41. https://doi.org/10.1016/j.jembe.2008.06.026
- Huuskonen, H., Piironen, J., Syväranta, J., Eronen, R., Biasi, C., Kiiskinen, P., Kortet, R., Vainikka, A., 2019. Diet and movements of pikeperch (*Sander lucioperca*) in a large oligotrophic lake with an exceptionally high pikeperch yield. Ecol. Freshw. Fish 28, 533–543. https://doi.org/10.1111/eff.12495
- Jungwirth, A., Balzarini, V., Zöttl, M., Salzmann, A., Taborsky, M., Frommen, J.G., 2019. Longterm individual marking of small freshwater fish: the utility of Visual Implant Elastomer tags. Behav. Ecol. Sociobiol. 73, 49. https://doi.org/10.1007/s00265-019-2659-y
- Jůza, T., Mrkvička, T., Blabolil, P., Čech, M., Peterka, J., Vašek, M., Kubečka, J., 2013. Occurrence of age-0 year dwarf pikeperch Sander lucioperca in late summer – an overlooked phenomenon in reservoirs. J. Fish Biol. 83, 1444–1452. https://doi.org/10.1111/jfb.12229

Karahan, A., Borsa, P., Gucu, A.C., Kandemir, I., Ozkan, E., Orek, Y.A., Acan, S.C., Koban, E., Togan, I., 2014. Geometric morphometrics, Fourier analysis of otolith shape, and nuclear-DNA markers distinguish two anchovy species (*Engraulis* spp.) in the Eastern Mediterranean Sea. Fish. Res. 159, 45–55. https://doi.org/10.1016/j.fishres.2014.05.009

Koeberle, A.L., Arismendi, I., Crittenden, W., Leer, D., Noakes, D.L.G., 2020. Fluctuating asymmetry of adult Chinook Salmon (Oncorhynchus tshawytscha) otoliths from wild and hatchery origins. Aquat. Ecol. 54, 431–446. https://doi.org/10.1007/s10452-019-09733-0

401

402 Kokkonen, E., Mitikka, S., Huuskonen, H., Olin, M., Ruuhijärvi, J., Vainikka, A., 2019. Structural

403 equation models suggest that bottom-up processes override top-down processes in boreal

- 404 pikeperch (Sander lucioperca) lakes. Freshw. Biol. 64, 1054–1063.
- 405 https://doi.org/10.1111/fwb.13285
 - Kuhn, M., Wing, J., Weston, S., Williams, A., Keefer, C., Engelhardt, A., Cooper, T., Mayer, Z., Kenkel, B., R Core Team, Benesty, M., Lescarbeau, R., Ziem, A., Scrucca, L., Tang, Y., Candan, C., Hunt, T., 2020. caret: Classification and Regression Training.
 - L'Abée-Lund, J.H., Jensen, A.J., 1993. Otoliths as natural tags in the systematics of salmonids. Environ. Biol. Fishes 36, 389–393. https://doi.org/10.1007/BF00012418
 - Lappalainen, J., Milardi, M., Nyberg, K., Venäläinen, A., 2009. Effects of water temperature on year-class strengths and growth patterns of pikeperch (*Sander lucioperca* (L.)) in the brackish Baltic Sea. Aquat. Ecol. 43, 181–191. https://doi.org/10.1007/s10452-007-9150-y
 - Lazartigues, A., Girard, C., Brodeur, P., Lecomte, F., Mingelbier, M., Sirois, P., 2017. Otolith microchemistry to identify sources of larval yellow perch in a fluvial lake: an approach towards freshwater fish management. Dispersal Early Life Hist. Fish 01, 474–487. https://doi.org/10.1139/cjfas-2016-0289@cjfas-ddel/issue01
 - Leguá, J., Plaza, G., Pérez, D., Arkhipkin, A., 2013. Otolith shape analysis as a tool for stock identification of the southern blue whiting, *Micromesistius australis*. Lat. Am. J. Aquat. Res.
 - Lehtonen, H., 1983. Stocks of pike-perch (*Stizostedion lucioperca* L.) and their management in the Archipelago Sea and the Gulf of Finland. Finn. Fish. Res. 5, 1–16.
 - Lestrel, P.E., 1997. Fourier Descriptors and Their Applications in Biology. Cambridge University Press.
 - Libungan, L.A., Óskarsson, G.J., Slotte, A., Jacobsen, J.A., Pálsson, S., 2015. Otolith shape: a population marker for Atlantic herring *Clupea harengus*. J. Fish Biol. 86, 1377–1395. https://doi.org/10.1111/jfb.12647
 - Lind, E.A., 1977. A Review of Pikeperch (*Stizostedion lucioperca*), Eurasian Perch (*Perca fluviatilis*), and Ruff (*Gymnocephalus cernua*) in Finland. J. Fish. Res. Board Can. 34, 1684–1695. https://doi.org/10.1139/f77-233
 - Lorenzen, K., Enberg, K., 2002. Density-dependent growth as a key mechanism in the regulation of fish populations: evidence from among-population comparisons. Proc. R. Soc. Lond. B Biol. Sci. 269, 49–54. https://doi.org/10.1098/rspb.2001.1853
 - Mahé, K., Gourtay, C., Defruit, G.B., Chantre, C., de Pontual, H., Amara, R., Claireaux, G., Audet, C., Zambonino-Infante, J.L., Ernande, B., 2019. Do environmental conditions (temperature and food composition) affect otolith shape during fish early-juvenile phase? An experimental approach applied to European Seabass (*Dicentrarchus labrax*). J. Exp. Mar. Biol. Ecol. 521, 151239. https://doi.org/10.1016/j.jembe.2019.151239

- Marengo, M., Baudouin, M., Viret, A., Laporte, M., Berrebi, P., Vignon, M., Marchand, B., Durieux, E.D.H., 2017. Combining microsatellite, otolith shape and parasites community analyses as a holistic approach to assess population structure of *Dentex dentex*. J. Sea Res. 128, 1–14. https://doi.org/10.1016/j.seares.2017.07.003
- Martino, J.C., Fowler, A.J., Doubleday, Z.A., Grammer, G.L., Gillanders, B.M., 2019. Using otolith chronologies to understand long-term trends and extrinsic drivers of growth in fisheries. Ecosphere 10, e02553. https://doi.org/10.1002/ecs2.2553
- McDermott, S.F., Cooper, D.W., Guthridge, J.L., Spies, I.B., Canino, M.F., Woods, P., Hillgruber, N., 2011. Effects of Maternal Growth on Fecundity and Egg Quality of Wild and Captive Atka Mackerel. Mar. Coast. Fish. 3, 324–335. https://doi.org/10.1080/19425120.2011.608592
- Mérigot, B., Letourneur, Y., Lecomte-Finiger, R., 2007. Characterization of local populations of the common sole *Solea solea* (Pisces, Soleidae) in the NW Mediterranean through otolith morphometrics and shape analysis. Mar. Biol. 151, 997–1008. https://doi.org/10.1007/s00227-006-0549-0
- Mille, T., Mahé, K., Cachera, M., Villanueva, M.C., Pontual, H. de, Ernande, B., 2016. Diet is correlated with otolith shape in marine fish. Mar. Ecol. Prog. Ser. 555, 167–184. https://doi.org/10.3354/meps11784
- Negus, M.T., 1999. Thermal Marking of Otoliths in Lake Trout Sac Fry. North Am. J. Fish. Manag. 19, 127–140. https://doi.org/10.1577/1548-8675(1999)019<0127:TMOOIL>2.0.CO;2
- Olden, J.D., Jackson, D.A., 2002. Illuminating the "black box": a randomization approach for understanding variable contributions in artificial neural networks. Ecol. Model. 154, 135–150. https://doi.org/10.1016/S0304-3800(02)00064-9
- Oxman, D.S., Barnett-Johnson, R., Smith, M.E., Coffin, A., Miller, D.L., Josephson, R., Popper, A.N., 2007. The effect of vaterite deposition on sound reception, otolith morphology, and inner ear sensory epithelia in hatchery-reared Chinook salmon (*Oncorhynchus tshawytscha*). Can. J. Fish. Aquat. Sci. 64, 1469–1478. https://doi.org/10.1139/f07-106
- Parmentier, E., Boistel, R., Bahri, M.A., Plenevaux, A., Schwarzhans, W., 2018. Sexual dimorphism in the sonic system and otolith morphology of *Neobythites gilli* (Ophidiiformes). J. Zool. 305, 274–280. https://doi.org/10.1111/jzo.12561

- 407 Paul, K., Oeberst, R., Hammer, C., 2013. Evaluation of otolith shape analysis as a tool for
- 408 discriminating adults of Baltic cod stocks. J. Appl. Ichthyol. 29, 743–750.
- 409 https://doi.org/10.1111/jai.12145
 - Pereira, L.A., Santos, R.V., Hauser, M., Duponchelle, F., Carvajal, F., Pecheyran, C., Bérail, S., Pouilly, M., 2019. Commercial traceability of *Arapaima* spp. fisheries in the Amazon basin: can biogeochemical tags be useful? Biogeosciences 16, 1781–1797. https://doi.org/10.5194/bg-16-1781-2019
 - Pérez, A., Fabré, N.N., 2013. Spatial population structure of the Neotropical tiger catfish *Pseudoplatystoma metaense*: skull and otolith shape variation. J. Fish Biol. 82, 1453–1468. https://doi.org/10.1111/jfb.12046
 - R Core Team, 2020. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
 - Rashidabadi, F., Abdoli, A., Tajbakhsh, F., Nejat, F., Avigliano, E., 2020. Unravelling the stock structure of the Persian brown trout by otolith and scale shape. J. Fish Biol. 96, 307–315. https://doi.org/10.1111/jfb.14170

Reimer, T., Dempster, T., Warren-Myers, F., Jensen, A.J., Swearer, S.E., 2016. High prevalence of vaterite in sagittal otoliths causes hearing impairment in farmed fish. Sci. Rep. 6, 25249. https://doi.org/10.1038/srep25249

- Rodgveller, C.J., Hutchinson, C.E., Harris, J.P., Vulstek, S.C., Iii, C.M.G., 2017. Otolith shape variability and associated body growth differences in giant grenadier, *Albatrossia pectoralis*. PLOS ONE 12, e0180020. https://doi.org/10.1371/journal.pone.0180020
- 410
- 411
- 412 Saulamo, K., Lappalainen, J., 2007. Effects of abiotic factors on movements of pikeperch during pre-
- 413 spawning and spawning season in a Baltic archipelago. Hydrobiologia 579, 271–277.
- 414 https://doi.org/10.1007/s10750-006-0410-y
 - Saulamo, K., Thoresson, G., 2005. Management of Pikeperch Migrating over Management Areas in a Baltic Archipelago Area. AMBIO J. Hum. Environ. 34, 120–124. https://doi.org/10.1579/0044-7447-34.2.120
 - Schade, F.M., Weist, P., Krumme, U., 2019. Evaluation of four stock discrimination methods to assign individuals from mixed-stock fisheries using genetically validated baseline samples. Mar. Ecol. Prog. Ser. 627, 125–139. https://doi.org/10.3354/meps13061
 - Schulz-Mirbach, T., Stransky, C., Schlickeisen, J., Reichenbacher, B., 2008. Differences in otolith morphologies between surface- and cave-dwelling populations of *Poecilia mexicana* (Teleostei, Poeciliidae) reflect adaptations to life in an extreme habitat. Evol. Ecol. 10, 537–558.
 - Skalski, J.R., Buchanan, R.A., Griswold, J., 2009. Review of Marking Methods and Release-Recapture Designs for Estimating the Survival of Very Small Fish: Examples from the Assessment of Salmonid Fry Survival. Rev. Fish. Sci. 17, 391–401. https://doi.org/10.1080/10641260902752199
 - Škeljo, F., Ferri, J., 2012. The use of otolith shape and morphometry for identification and sizeestimation of five wrasse species in predator-prey studies. J. Appl. Ichthyol. 28, 524–530. https://doi.org/10.1111/j.1439-0426.2011.01925.x
 - Šmejkal, M., Blabolil, P., Bartoň, D., Duras, J., Vejřík, L., Sajdlová, Z., Kočvara, L., Kubečka, J., 2019. Sex-specific probability of PIT tag retention in a cyprinid fish. Fish. Res. 219, 105325. https://doi.org/10.1016/j.fishres.2019.105325
 - Smoliński, S., Schade, F.M., Berg, F., 2019. Assessing the performance of statistical classifiers to discriminate fish stocks using Fourier analysis of otolith shape. Can. J. Fish. Aquat. Sci. 77, 674–683. https://doi.org/10.1139/cjfas-2019-0251
 - Specziár, A., Ero"s, T., 2015. Freshwater resources and fisheries in Hungary, in: Freshwater Fisheries Ecology. John Wiley & Sons, Ltd, pp. 196–200. https://doi.org/10.1002/9781118394380.ch15
 - Specziár, A., Turcsányi, B., 2017. Management of pikeperch stocking in Lake Balaton: effect of season, area, fish size and method of release on the rate and distribution of recaptures. Knowl. Manag. Aquat. Ecosyst. 52. https://doi.org/10.1051/kmae/2017047
 - Spurgeon, J.J., Pegg, M.A., Halden, N.M., 2018. Mixed-origins of channel catfish in a large-river tributary. Fish. Res. 198, 195–202. https://doi.org/10.1016/j.fishres.2017.09.001

Svåsand, T., Jørstad, K.E., Kristiansen, T.S., 1990. Enhancement studies of coastal cod in western Norway. Part I. Recruitment of wild and reared cod to a local spawning stock. ICES J. Mar. Sci. 47, 5–12. https://doi.org/10.1093/icesjms/47.1.5

Sweeting, R.M., Beamish, R.J., Neville, C.M., 2004. Crystalline otoliths in teleosts: Comparisons between hatchery and wild coho salmon (*Oncorhynchus kisutch*) in the Strait of Georgia. Rev. Fish Biol. Fish. 14, 361–369. https://doi.org/10.1007/s11160-005-3793-3

415

- 416
- 417 Taylor, M.D., Fielder, D.S., Suthers, I.M., 2005. Batch marking of otoliths and fin spines to assess
- 418 the stock enhancement of Argyrosomus japonicus. J. Fish Biol. 66, 1149–1162.
- 419 https://doi.org/10.1111/j.0022-1112.2005.00678.x
 - Tuset, V.M., Lozano, I.J., González, J.A., Pertusa, J.F., García-Díaz, M.M., 2003. Shape indices to identify regional differences in otolith morphology of comber, *Serranus cabrilla* (L., 1758).
 J. Appl. Ichthyol. 19, 88–93. https://doi.org/10.1046/j.1439-0426.2003.00344.x

Utter, F., 1998. Genetic Problems of Hatchery-Reared Progeny Released into the Wild, and how to Deal with them. Bull. Mar. Sci. 62, 623–640.

van Densen, W.L.T., Vijverberg, J., 1982. The relations between 0+ fish density, zooplankton size and the vulnerability of pikeperch, *Stizostedion lucioperca*, to angling in the Frisian lakes, in: Gulati, R.D., Parma, D.S. (Eds.), Studies on Lake Vechten and Tjeukemeer, The Netherlands: 25th Anniversary of the Limnological Institute of the Royal Netherlands Academy of Art and Sciences, Developments in Hydrobiology. Springer Netherlands, Dordrecht, pp. 321–336. https://doi.org/10.1007/978-94-009-8015-0_22

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- 422 Kubečka, J., Peterka, J., 2018. Stable isotopes and gut contents indicate differential resource
- 423 use by coexisting asp (*Leuciscus aspius*) and pikeperch (*Sander lucioperca*). Ecol. Freshw.
- 424 Fish 27, 1054–1065. https://doi.org/10.1111/eff.12414
 - Vaux, F., Rasmuson, L.K., Kautzi, L.A., Rankin, P.S., Blume, M.T.O., Lawrence, K.A., Bohn, S., O'Malley, K.G., 2019. Sex matters: Otolith shape and genomic variation in deacon rockfish (*Sebastes diaconus*). Ecol. Evol. 9, 13153–13173. https://doi.org/10.1002/ece3.5763

- 426 Vignon, M., 2018. Short-term stress for long-lasting otolith morphology brief embryological stress
- 427 disturbance can reorient otolith ontogenetic trajectory. Can. J. Fish. Aquat. Sci. 75, 1713–
- 428 1722. https://doi.org/10.1139/cjfas-2017-0110

- Vignon, M., 2012. Ontogenetic trajectories of otolith shape during shift in habitat use: Interaction between otolith growth and environment. J. Exp. Mar. Biol. Ecol. 420–421, 26–32. https://doi.org/10.1016/j.jembe.2012.03.021
- Vignon, M., Morat, F., 2010. Environmental and genetic determinant of otolith shape revealed by a non-indigenous tropical fish. Mar. Ecol. Prog. Ser. 411, 231–241. https://doi.org/10.3354/meps08651
- Volk, E.C., Schroder, S.L., Grimm, J.J., 1999. Otolith thermal marking. Fish. Res. 43, 205–219. https://doi.org/10.1016/S0165-7836(99)00073-9
- Welcomme, R.L., Bartley, D.M., 1998. Current approaches to the enhancement of fisheries. Fish. Manag. Ecol. 5, 351–382. https://doi.org/10.1046/j.1365-2400.1998.550351.x
- Zupa, R., Rodríguez, C., Mylonas, C.C., Rosenfeld, H., Fakriadis, I., Papadaki, M., Pérez, J.A., Pousis, C., Basilone, G., Corriero, A., 2017. Comparative Study of Reproductive Development in Wild and Captive-Reared Greater Amberjack *Seriola dumerili* (Risso, 1810). PLOS ONE 12, e0169645. https://doi.org/10.1371/journal.pone.0169645





Fig. 1. Map showing the origin of the pikeperch (*Sander lucioperca*) individuals used in this study.
Reservoir = full symbol, rearing facilities = open symbols. Blue lines represent the largest riverine
systems in the country.



438 Fig. 2. Measurements extracted from the pikeperch (Sander lucioperca) sagittae otoliths and the

439 landmark used to standardize the otolith positions.



Fig. 3. Neural network interpretation diagram showing the relationships between the otolith shape indices and the fish origin. The widths of the connections represent the strength of the relationship, with red color representing the negative weight connections and blue color representing the positive connection weights. Only the non-correlated shape indices were used in the analysis (i.e. roundness index (SI_{RO}), rectangularity index (SI_{RE}), format factor index (SI_{FF}) and solidity index (SI_{SO})). Letters inside the circles indicate the layer type in the neural network (I = input layer, H = hidden layer, O = output layer).

449



452 Fig. 4. Mean shape of the sagittae otolith from stocked (top left corner) and wild (bottom left corner)

453 pikeperch (Sander lucioperca). Differences in the otolith average shape of stocked and wild pikeperch

454 (right side) is expressed in color heatmap scale (small difference = light yellow, high difference =

455 dark red). Otolith outlines were scaled to allow better visual comparison between groups.

456 **Table 1.** Shape indexes (SI) computed for the pikeperch (*Sander lucioperca*) otoliths. $O_A =$ otolith 457 area, $O_{AC} =$ otolith convex hull area, $O_L =$ otolith length, $O_P =$ otolith perimeter, $O_{PC} =$ otolith convex 458 hull perimeter, $O_W =$ otolith width.

Shape index	Index acronym	Formula
Aspect ratio	SI _{AR}	$SI_{AR} = O_L / O_W$
Circularity	SI _{CI}	$SI_{CI} = O_P^2 / O_A$
Convexity	SI _{CO}	$SI_{CO} = O_{PC} / O_P$
Ellipicity	SI _{EL}	$\mathrm{SI}_{\mathrm{EL}} = (O_L - O_W) / (O_L + O_W)$
Format factor	SI _{FF}	$\mathrm{SI}_{\mathrm{FF}} = 4 \pi \cdot O_A / O_p^2$
Rectangularity	SI _{RE}	$SI_{RE} = O_A / (O_L \cdot O_W)$
Roundness	SI _{RO}	$\mathrm{SI}_{\mathrm{RO}} = 4 \cdot O_A / \pi \cdot O_P^2$
Solidity	SI _{SO}	$SI_{SO} = O_A / O_{AC}$

460 **Table 2.** Comparisons of fish standard length (SL) and the morphometrical measurements taken from 461 the otoliths of the stocked and wild pikeperch individuals (*Sander lucioperca*) from Lipno lake 462 (Czechia). O_A = otolith area, O_L = otolith length, O_P = otolith perimeter, O_W = otolith width, SE = 463 standard error, CI = confidence interval (5% – 95%), Range = (minimum – maximum).

			Stocked				Wild	
	Mean	SE	CI	Range	Mean	SE	CI	Range
OA	8.28	0.74	6.83 - 9.73	1.39 – 17.34	7.91	0.39	7.19 - 8.68	1.13 – 15.23
(mm ²)								
O _L (mm)	4.97	0.25	4.46 - 5.45	1.93 – 7.3	4.83	0.14	4.56 - 5.09	1.79 – 7.43
O _P (mm)	14.62	0.83	12.98 – 16.25	5.27 - 24.48	13.88	0.4	13.04 -14.67	4.66 - 21.21
O _W (mm)	2.29	0.11	2.1 - 2.5	1.01 – 3.6	2.23	0.05	2.12 - 2.33	0.91 - 3.29
SL (mm)	208.46	2.29	204.4 - 213.3	195 – 245	217.32	1.36	214.6 - 219.9	190 - 245

465	Table 3. Average values for the shape indexes computed for the otoliths of stocked and wild pikeperch
466	(Sander lucioperca) individuals from the Lipno lake (Czechia). SI_{AR} = aspect ratio index, SI_{CI} =
467	circularity index, SI_{CO} = convexity index, SI_{EL} = ellipicity index, SI_{FF} = format factor index, SI_{RE} =
468	rectangularity index, SI_{RO} = roundness index, SI_{SO} = solidity index, SE = standard error, CI =
469	confidence interval $(5\% - 95\%)$, Range = (minimum – maximum).

Otolith shape index		Stoo	cked		Wild				
	Mean	SE	CI	Range	Mean	SE	CI	Range	
SI _{AR}	2.139	0.037	2.070	1.858	2.142	0.017	2.109	1.889	
								2.823	
SI _{CI}	20.014	0.36	19.30	16.643	19.349	0.15	19.06	16.011	
			20.76	24.824			 19.63	23.627	
SI _{CO}	0.903	0.008	0.887	0.777	0.924	0.003	0.918	0.841	
				0.962			0.930	 0.989	
SI_{EL}	0.361	0.007	0.348	0.300	0.362	0.003	0.356	0.308	
			0.375	_ 0.463			 0.367		
SI _{FF}	0.633	0.011	0.611	0.506	0.653	0.005	0.644	0.532	
			 0.656	0.755			_ 0.664		
SI _{RE}	0.683	0.004	0.676	0.653	0.687	0.002	0.683	0.630	
			_ 0.691	0.728			 0.692		
SI _{RO}	0.409	0.006	0.398	0.334	0.411	0.003	0.405	0.328	
			 0.420						
SI _{SO}	0.950	0.002	0.944	0.918	0.954	0.001	0.952	0.924	
			0.955	0.974			0.957	0.981	