

Computational biomolecular simulation workflows with **BioExcel building blocks**

BioExcel Webinar
10/09/2020

Adam Hospital Gasch
Institute for Research in Biomedicine (IRB) – Barcelona - Spain

Partners



Funding



Index

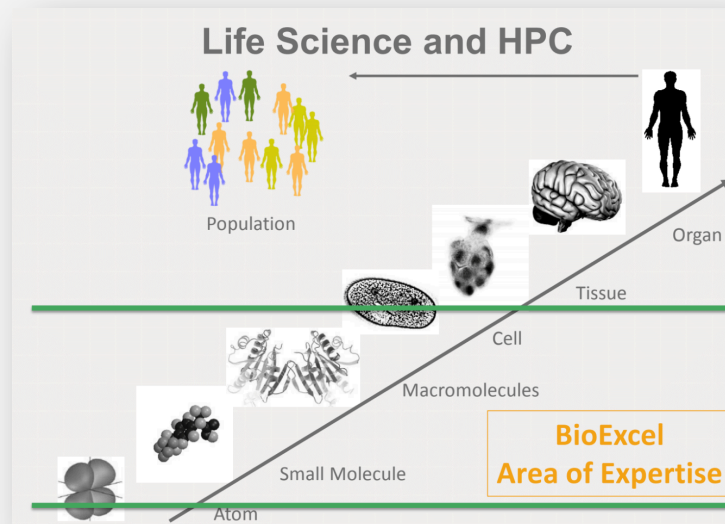
- **BioExcel Center of Excellence (CoE)**
- **Biomolecular workflows**
- **BioExcel Building Blocks software library (BioBB)**
- **Biomolecular workflows using BioBB**
 - **Demonstration** workflows (Jupyter Notebooks)
 - **Pre-exascale HPC** workflows (PyCOMPSs)
 - ✧ **Example: COVID-19** pre-exascale workflows

BioExcel Center of Excellence (CoE)



Centre of Excellence for Computational Biomolecular Research

A central hub for biomolecular modelling and simulations



EMBL-EBI



MANCHESTER 1824

The University of Manchester

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Ian Harrow Consulting



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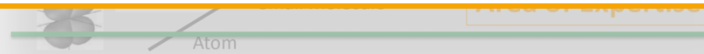
Barcelona Supercomputing Center
Centro Nacional de Supercomputación



Universiteit Utrecht

Enabling better science by:

- Improving the **performance** and **functionality** of key applications
- Providing **support** to **non-experts** and **advanced** users
- Developing **user-friendly computational workflows**



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Molecular Dynamics:

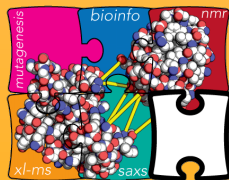
GROMACS
FAST. FLEXIBLE. FREE.



Free energy calculations:

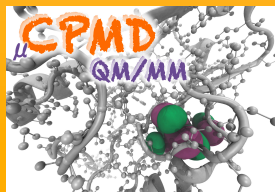
pmx: generate hybrid protein structure and topology
Computational Biomolecular Dynamics Group

Protein-Protein Docking:



ADDOCK
High-Ambiguity Driven Docking

QM / MM:



<https://bioexcel.eu/>



- *Webinars*
- *Training events*
- *Conferences*
- *Workshops*
- *Industry visits*
- *Forum*
(ask.bioexcel.eu)

Key Applications

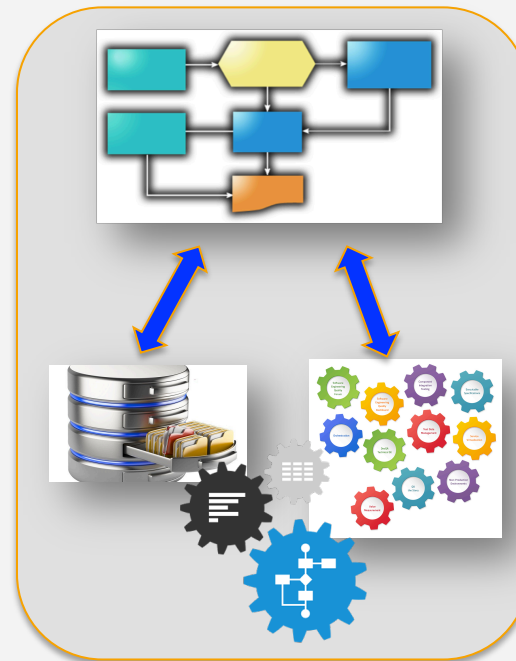
Support

- Design, deploy and make available solution-oriented biomolecular workflows.

- Excellence in **Usability**.



- Ease of use
- Availability
- Reproducibility
- Multi-platform
- Multi-infrastructure

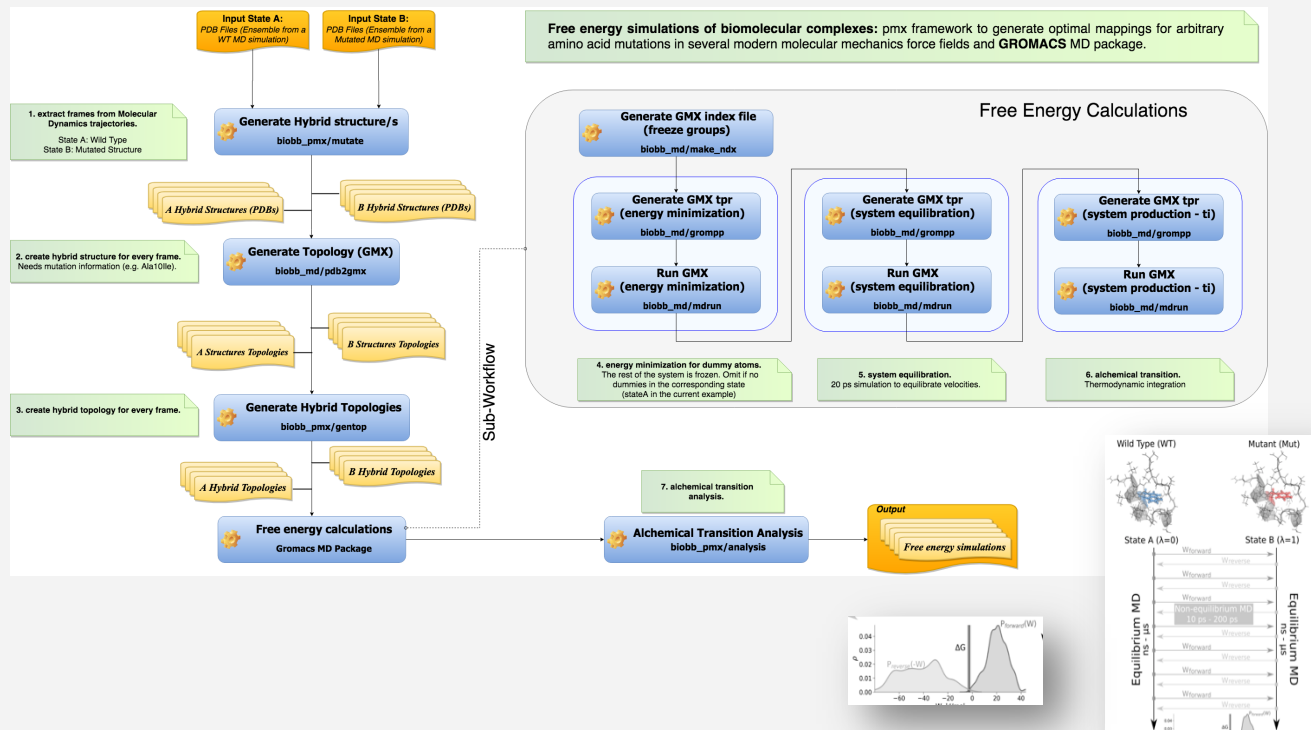


Use case-driven

Final deployment



Biomolecular workflows



- **Biomolecular simulation workflows** are usually built from a number of **tools** performing different **tasks**.
 - *Molecular Structure File format conversions*
 - *Structure Modelling*
 - *Molecular Dynamics*
 - *Quantum Mechanics*
 - *QM / MM*
 - *Trajectory analyses*
 - *Docking*
 - *Free energy*
 - *Ligand parameterization*
 - *Cheminformatics*
 - *Data analytics*



GROMACS
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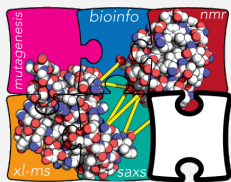


Amber18

AmberTools19

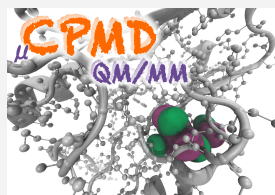
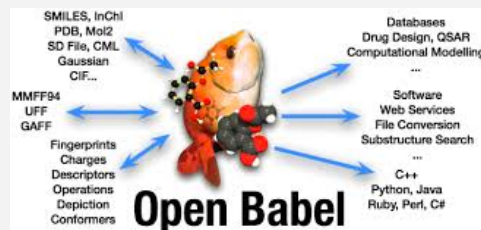
NAMD
Scalable Molecular Dynamics

VMD
Visual Molecular Dynamics

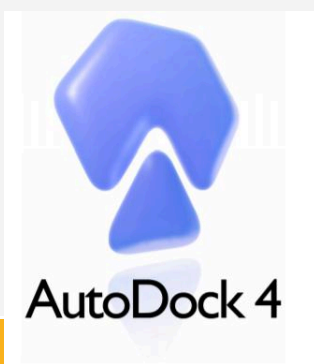


ADDock
High-Ambiguity Driven Docking

ACPYPE



pmx: generate hybrid protein structure and topology
Computational Biomolecular Dynamics Group



Modeller

Program for Comparative Protein Structure Modelling by Satisfaction of Spatial Restraints



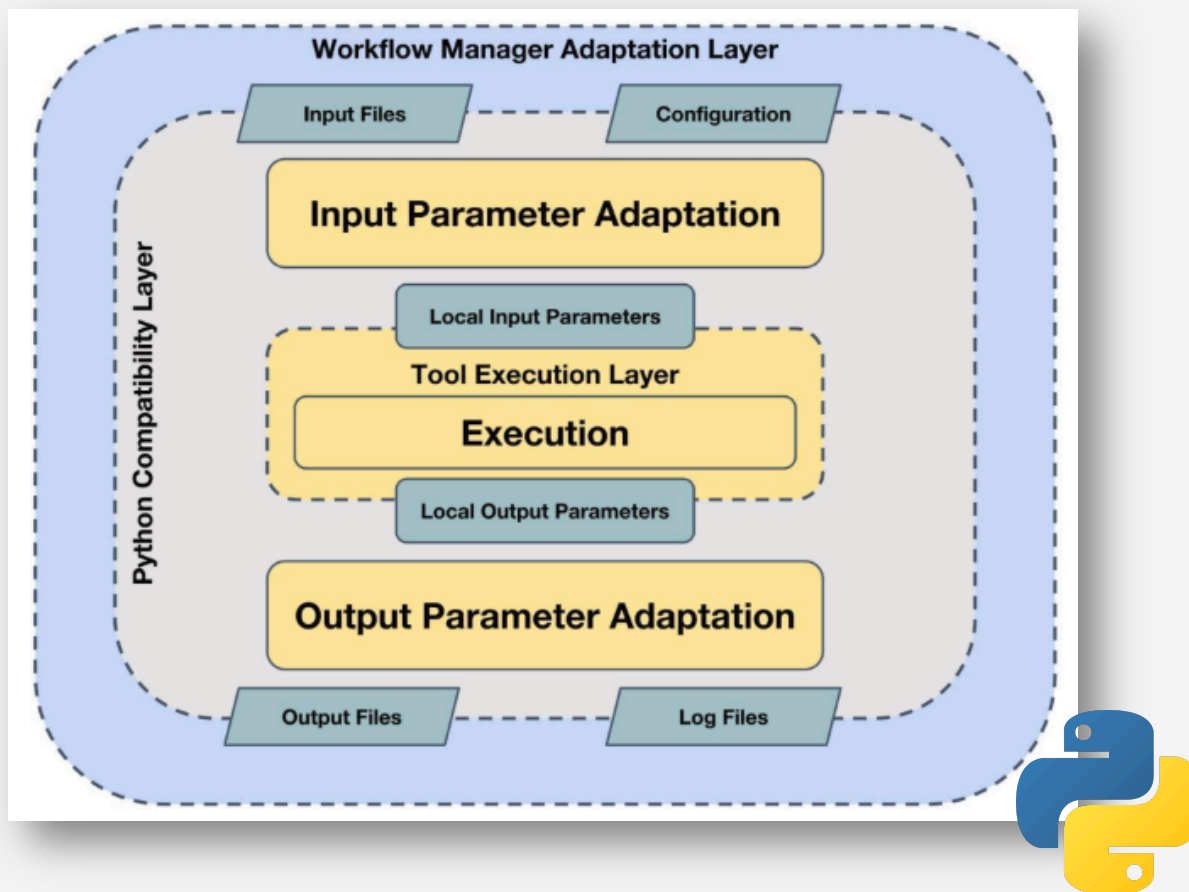
Shell script:

- *Step 1*
 - *Step 2*
- ↓
- *Step n*

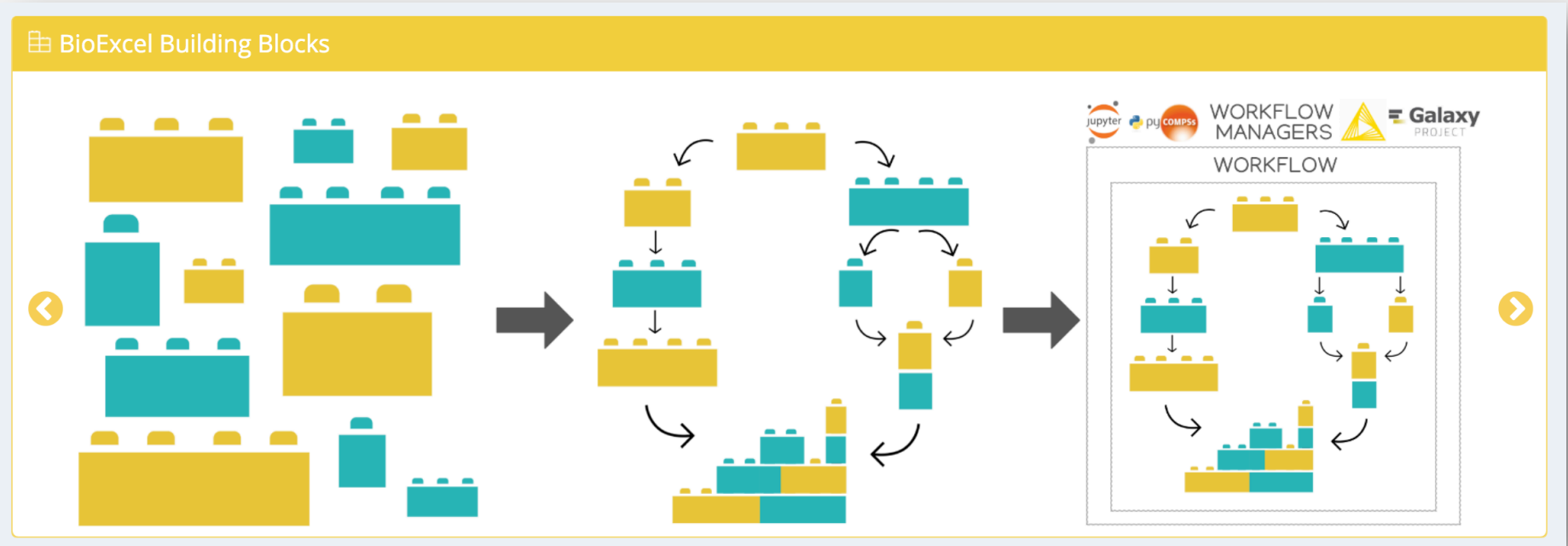


<http://mmb.irbbarcelona.org/biobb/>

BioExcel Building Blocks



<http://mmb.irbbarcelona.org/biobb/>



1) *Building Blocks*

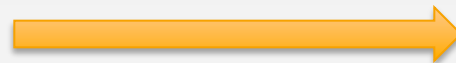
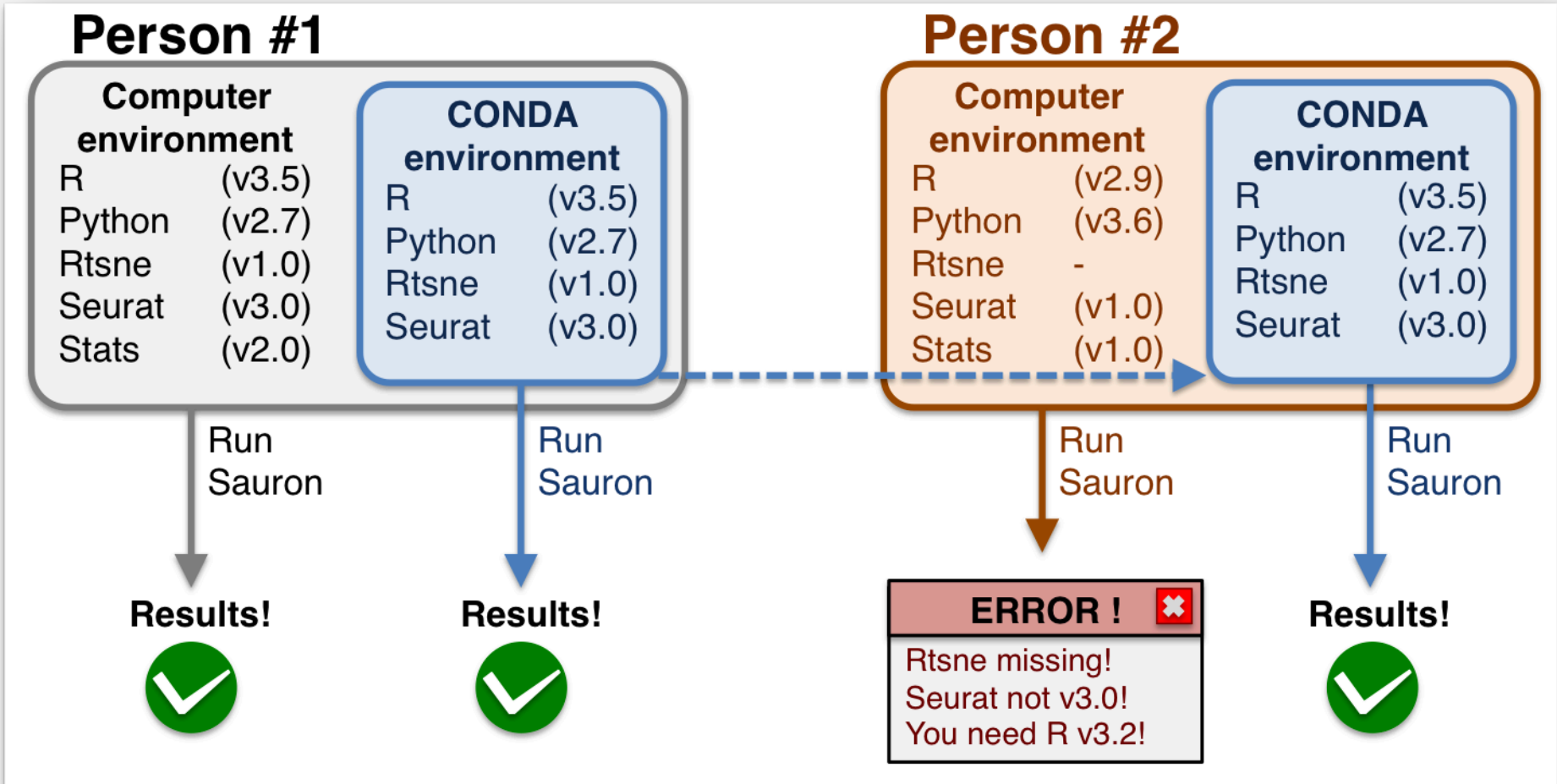
2) *Workflows*

3) *Workflow Managers*



F_{indable} A_{ccessible} I_{nteroperable} R_{eusable}



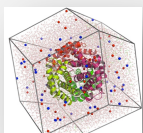


Reproducibility



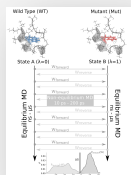
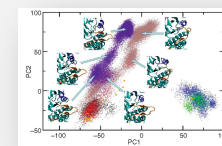
- **biobb_common**: base package required to use the biobb library

- **biobb_io**: collection to fetch data from biological databases



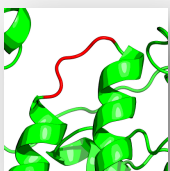
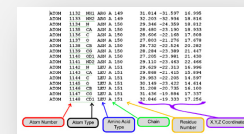
- **biobb_md**: collection to perform Molecular Dynamics simulations

- **biobb_analysis**: collection to perform analysis over MD simulations



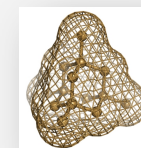
- **biobb_pmx**: collection to perform free energy calculations

- **biobb_structure_utils**: collection to modify or extract information from a PDB



- **biobb_model**: collection to check and model 3D structures

- **biobb_chemistry**: cheminformatics analyses and format conversions



















































- **biobb_ml**: machine learning algorithms (scikit learn, tensorflow)

☰ SOURCE AND DOCS FOR BIOEXCEL BUILDING BLOCKS

<http://mmb.irbbarcelona.org/biobb/availability/source>

Search by text

Search by keywords

Package	Description	Python	ReadTheDocs	Bioconda	Docker	Singularity	Version
 biobb_analysis	Biobb_analysis is the Biobb module collection to perform analysis of molecular dynamics simulations.						3.0.1
 biobb_chemistry	Biobb_chemistry is the Biobb module collection to perform chemistry over molecular dynamics simulations.						3.0.1
 biobb_common	Biobb_common is the base package required to use the biobb packages						3.0.0
 biobb_io	Biobb_io is the Biobb module collection to fetch data to be consumed by the rest of the Biobb building blocks.						3.0.0
 biobb_md	Biobb_md is the Biobb module collection to perform molecular dynamics simulations.						3.0.0
 biobb_model	Biobb_model is the Biobb module collection to check and model 3d structures, create mutations or reconstruct missing atoms.						3.0.0
 biobb_pmx	Biobb_pmx is the Biobb module collection to perform PMX executions.						2.0.2
 biobb_structure_utils	Biobb_structure_utils is the Biobb module collection to modify or extract information from a PDB structure file.						3.0.0

SOURCE AND DOCS FOR BIOEXCEL BUILDING BLOCKS

<http://mmb.irbbarcelona.org/biobb/availability/source>

Search by text

Search by keywords

Package	Description	Python	ReadTheDocs	Bioconda	Docker	Singularity	Version
biobb_analysis	Biobb_analysis is the Biobb module collection to perform analysis of molecular dynamics simulations.						3.0.1

conda install biobb_analysis

Building block	Wrapped tool	Description
GMXCluster	gmx cluster	Creates cluster structures from a given GROMACS compatible trajectory.
GMXRms	gmx rms	Performs a Root Mean Square deviation (RMSd) analysis from a given GROMACS compatible trajectory.
GMXRgyr	gmx gyrate	Computes the radius of gyration (Rgyr) of a molecule about the x-, y- and z-axes, as a function of time, from a given GROMACS compatible trajectory.
GMXEnergy	gmx energy	Extracts energy components from a given GROMACS energy file.
GMXImage	gmx trjconv	Corrects periodicity (image) from a given GROMACS compatible trajectory file.
GMXTrjconvStr	gmx trjconv	Converts between GROMACS compatible structure file formats and/or extracts a selection of atoms.
GMXTrjconvStrEns	gmx trjconv	Extracts an ensemble of frames containing a selection of atoms from GROMACS compatible trajectory files.
GMXTrjconvTrj	gmx trjconv	Converts between GROMACS compatible trajectory file formats and/or extracts a selection of atoms.
CpptrajAverage	Ambertools cpptraj	Calculates a structure average of a given cpptraj compatible trajectory.
CpptrajBfactor	Ambertools cpptraj	Calculates the Bfactor fluctuations of a given cpptraj compatible trajectory.
CpptrajRms	Ambertools cpptraj	Calculates the Root Mean Square deviation (RMSd) of a given cpptraj compatible trajectory.

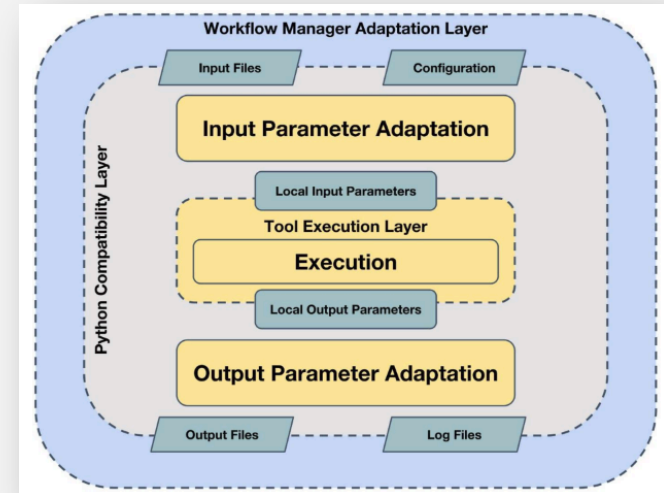
AmberTools19

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- Import Module
- Define:
 - inputs & output paths
 - properties dictionary
- Launch building block

Inputs Paths
Outputs Paths
Properties

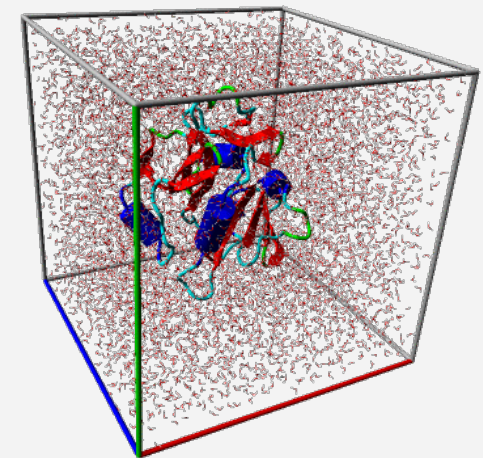


```
# Editconf: Create solvent box
# Import module
from biobb_md.gromacs.editconf import Editconf
```

```
# Create prop dict and inputs/outputs
output_editconf_gro = pdbCode+'_editconf.gro'

prop = {
    'box_type': 'cubic',
    'distance_to_molecule': 1.0
}
```

```
#Create and launch bb
Editconf(input_gro_path=output_pdb2gmx_gro,
         output_gro_path=output_editconf_gro,
         properties=prop).launch()
```

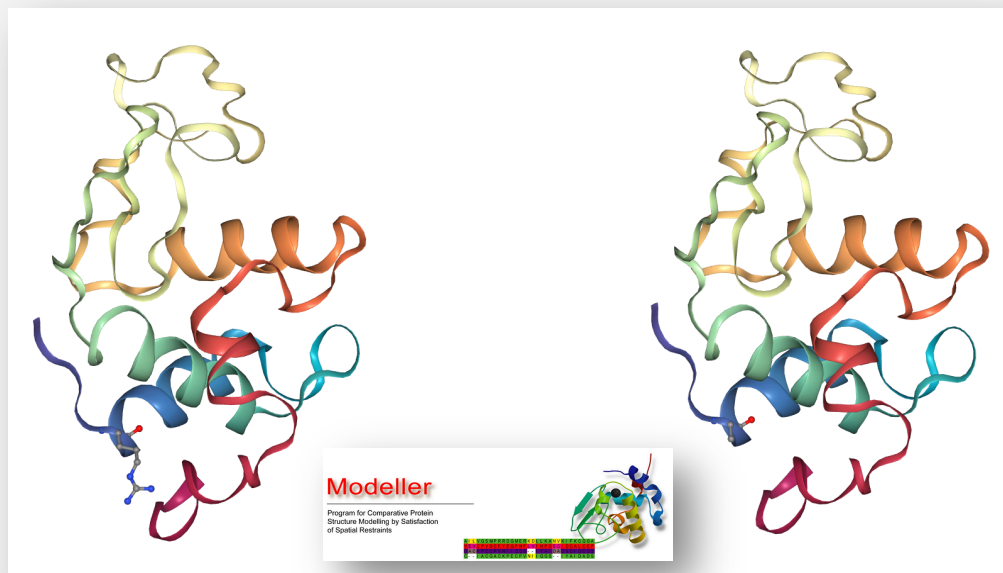


```
# Mutate Residue from PDB
# Import module
from biobb_model.model.mutate import Mutate

# Create prop dict and inputs/outputs
mutated_pdb = pdbCode + '_mutated.pdb'

prop = {
    'mutation_list': 'A:Arg5Ala'
}

# Create and launch bb
Mutate(input_pdb_path=fixed_pdb,
       output_pdb_path=mutated_pdb,
       properties=prop).launch()
```



```
from biobb_analysis.gromacs.gmx_cluster import GMXcluster

#traj = 'stateA_ins.xtc'
#top = 'stateA.tpr'

#traj = 'lnql_bound.0-200.imaged.rot.10.xtc' # 700MB !!
#top = 'lnql_bound.dry.pdb'

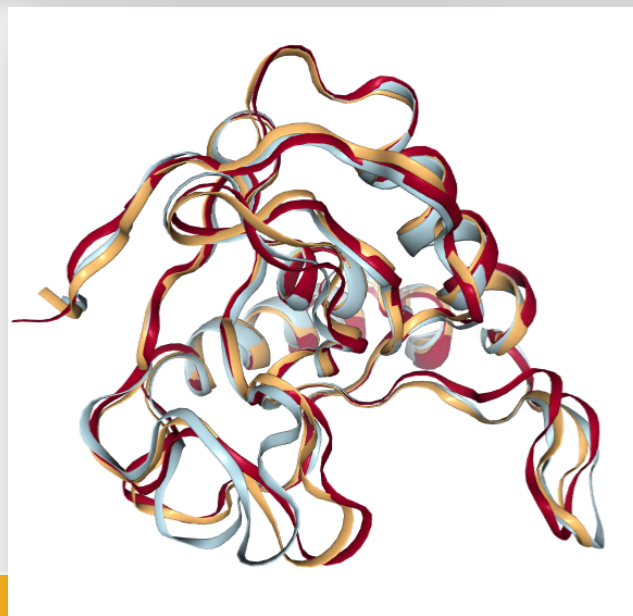
traj = 'lnql_G39R.0-50.imaged.rot.10.xtc'
top = 'lnql_G39R.dry.pdb'

out_ensemble = 'ensemble.pdb'

prop = {
    'fit_selection': 'Protein',
    'output_selection': 'Protein'
}

GMXcluster(
    input_structure_path=top,
    input_traj_path=traj,
    output_pdb_path=out_ensemble,
    properties = prop
).launch()
```

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```
In [ ]: # Ligand: Download ligand structure from MMB PDB mirror REST API (http://mmb.irbbarcelona.org/api/)
# Import module
from biobb_io.api.ligand import Ligand
from biobb_io.api.pdb import Pdb

# Create prop dict and inputs/outputs
input_structure = ligandCode + '.pdb'

prop = {
    'ligand_code' : 'IBP'
}

#Create and launch bb
Ligand(output_pdb_path=input_structure,
        properties=prop).launch()
```



```
In [ ]: from biobb_chemistry.babelm.babel_add_hydrogens import BabelAddHydrogens

ligandFile_wH = 'IBP.H.pdb'

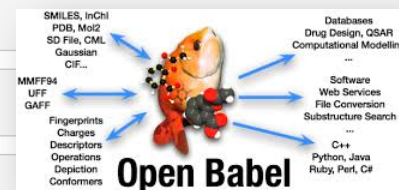
BabelAddHydrogens(input_path=input_structure,
                  output_path=ligandFile_wH,
                  ).launch()
```

```
In [ ]: view = nglview.show_file(ligandFile_wH)
view
```

```
In [ ]: from biobb_chemistry.babelm.babel_minimize import BabelMinimize

ligandFile_wH_min = 'IBP.H.min.pdb'

BabelMinimize(input_path=ligandFile_wH,
               output_path=ligandFile_wH_min,
               properties=prop).launch()
```



```
In [ ]: view = nglview.show_file(ligandFile_wH_min)
view
```

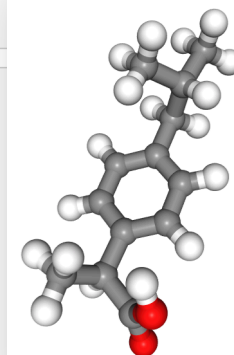
```
In [ ]: from biobb_chemistry.acpype.acpype_params_gmx import AcpypeParamsGMX


output_acpype_gro = 'IBP_params.gro'
output_acpype_itp = 'IBP_params.itp'
output_acpype_top = 'IBP_params.top'
output_acpype = 'IBP_params'

prop = {
    'basename' : 'IBP'
}

AcpypeParamsGMX(input_path=ligandFile_wH_min,
                 output_path_gro=output_acpype_gro,
                 output_path_itp=output_acpype_itp,
                 output_path_top=output_acpype_top,
                 properties=prop).launch()
```

ACPYPE





latest

- Introduction & installation
- API Documentation
 - acypype package
 - Submodules
 - acypype.acypype_params_ac module
 - acypype.acypype_params_cns module
 - acypype.acypype_params_gmx module
 - acypype.acypype_params_gmx_opls module
 - babelm package
 - ambertools package
- Command Line Documentation
- Changelog

acypype.acypype_params_gmx module

Module containing the Acypype class and the command line interface.

`class acypype.acypype_params_gmx.AcypypeParamsGMX(input_path, output_path_gro, output_path_itp, output_path_top, properties=None, **kwargs)` [\[source\]](#)

Bases: `object`

Small molecule parameterization for GROMACS MD package. Wrapper for the Acypype module. Generation of topologies for GROMACS. Acypype is a tool based in Python to use Antechamber to generate topologies for chemical compounds and to interface with others python applications like CCPN or ARIA. [Visit the official page.](#)

- Parameters:
- **input_path** (*str*) - Path to the input file. File type: input. [Sample file](#). Accepted formats: pdb, mdl, mol2.
 - **output_path_gro** (*str*) - Path to the GRO output file. File type: output. [Sample file](#). Accepted formats: gro.
 - **output_path_itp** (*str*) - Path to the ITP output file. File type: output. [Sample file](#). Accepted formats: itp.
 - **output_path_top** (*str*) - Path to the TOP output file. File type: output. [Sample file](#). Accepted formats: top.
 - **properties** (*dic*) -
 - **basename** (*str*) - ("BBB") A basename for the project (folder and output files).
 - **charge** (*int*) - (0) Net molecular charge, for gas default is 0.
 - **acypype_path** (*str*) - ("acypype") Path to the acypype executable binary.
 - **remove_tmp** (*bool*) - (True) [WF property] Remove temporal files.
 - **restart** (*bool*) - (False) [WF property] Do not execute if output files exist.
 - **container_path** (*str*) - (None) Container path definition.
 - **container_image** (*str*) - ('mmbirb/acypype:latest') Container image definition.
 - **container_volume_path** (*str*) - ('/tmp') Container volume path definition.
 - **container_working_dir** (*str*) - (None) Container working directory definition.
 - **container_user_id** (*str*) - (None) Container user_id definition.
 - **container_shell_path** (*str*) - ('/bin/bash') Path to default shell inside the container.




MENU ▾

SCIENTIFIC DATA 

Article | [Open Access](#) | Published: 10 September 2019

BioExcel Building Blocks, a software library for interoperable biomolecular simulation workflows

Pau Andrio, Adam Hospital, Javier Conejero, Luis Jordá, Marc Del Pino, Laia Codo, Stian Soiland-Reyes, Carole Goble, Daniele Lezzi, Rosa M. Badia, Modesto Orozco & Josep Ll. Gelpi 

Scientific Data **6**, Article number: 169 (2019) | [Cite this article](#)

799 Accesses | **4** Altmetric | [Metrics](#)

Innovation Radar

 Search by keyword...



Exploring

Innovations actively exploring value creation opportunities.

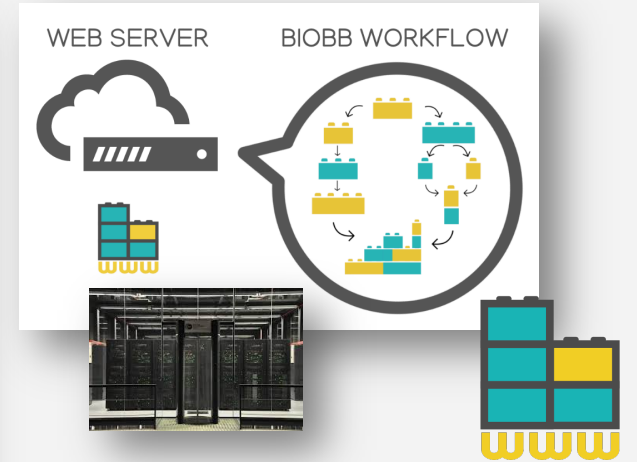
The Innovation Radar is a European Commission initiative to identify high potential innovations and innovators in EU-funded research and innovation framework programmes ([tell me more](#))

LIBRARY VERSATILITY TUTORIALS

Common Workflow Language

Protein MD Setup REST API

Command-line Workflows



Tools

search tools

pdb cluster: get zip from mmb.irbbarcelona.org

Grompp: Gromacs preprocessor. Reads topology file to create TPR atomic description

Pdb2gmx: creates a ZIP Gromacs topology from a given PDB file.

Solvate: creates a Gromacs topology file adding solvent molecules to a given input.

Ndx2restop: creates a Gromacs topology applying the force restrains

RMS: performs a RMSD analysis.

Make NDX: creates a Gromacs index file (NDX).

Pdb2gmx: creates a ZIP Gromacs topology from a given PDB file. (Galaxy Version 0.1.5)

☆ Favorite Options

Output GRO name

myPdb2gmx.gro

Name for the Output GRO. Format: [output].gro

Output ZIP name

myPdb2gmx.zip

Name for the Output TOP topology. Format: [output].zip

Input PDB file

No pdb dataset available.

Select your input PDB file. Format: [input].pdb

Take tool settings:

by default

Select where tool settings are to be read from

MD_pdb2gmx

GROMACS pdb2gmx. Create a GROMACS topology from a PDB file

Dialog Options

Force Field:
Force Field (Default: AMBER99SB-ILDN), Options: AMBER03, AMBER94, AMBER96, AMBER99, AMBER99SB, AMBER99SB-ILDN, AMBERGS, CHARMM27, GROMOS96 43a1, GROMOS96 43a2, GROMOS96 45a5, GROMOS96 53a5, GROMOS96 53a6, GROMOS96 54a7.

Water Type:
Water Type (Default: SPC/E), Options: TIP3P, TIP4P, TIP4P-Ew, TIP5P, SPC, SPC/E, None.

Ignore Hydrogens:
Ignore Hydrogens

Ports

```

KNIME Console
WARN PythonService PythonService is not initialized
WARN PythonService Could not start python service.
WARN Create File Name Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Create File Name 1:5:9 Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Create File Name 1:6:9 Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Create File Name 1:7:9 Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Create File Name 1:8:9 Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Create File Name 1:10:9 Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Create File Name 1:11:9 Selected directory /private/var/folders/7f/@xkgf3d971b981k.f
WARN Proteins Viewer 2:19 Column with molecules missing on port proteins
    
```



1) Demonstration Workflows (Jupyter Notebooks)

Increasing usability

- Making biomolecular simulation tools interoperable
- Making biomolecular simulation workflows accessible, easy to build and use

```
In [21]: # Downloading desired PDB file
# Import module
from bioexcelsuite.pdb import Pdb

# Create properties dict and inputs/outputs
downloaded_pdb = PdbCode('pdb')
prop = {
    'pdb_code': 'pdbCode'
}

# Create and launch job
PdbCodeJob(download_pdb, downloaded_pdb,
            properties=prop).launch()

2020-06-01 14:51:58,250 [MainThread] [INFO] Downloading 1aki from https://files.rcsb.org/download/1aki.pdb
2020-06-01 14:51:58,302 [MainThread] [INFO] Writing pdb to /home/joyce/bioexcelsuite/notebooks/1aki.pdb
2020-06-01 14:51:58,504 [MainThread] [INFO] Filtering lines NOT starting with one of these words: ['ATOM', 'MODE
L', 'HEADER']
```

Visualizing 3D structure
Visualizing the downloaded/given PDB structure using NGL:

```
In [31]: # Show protein
view = ngview.show_structure_file(download_pdb)
view.add_representation(repr_type='ball-and-stick', selection='all')
view._remote_call('setSize', target='Widget', args=['', '600px'])
view
```

2) Pre-exascale Workflows (PyCOMPSs)

Enabling scalability

Making biomolecular simulation workflows exascale ready



Biomolecular workflows using biobb

Demonstration Workflows



Increasing usability

Making biomolecular simulation tools interoperable

Making biomolecular simulation workflows accessible, easy to build and use

```
In [2]: # Downloading desired PDB file
# Import module
from biobb_lo_apl.pdb import Pdb

# Create properties dict and inputs/outputs
downloaded_pdb = pdbCode + ".pdb"
prop = {
    'pdb_code': pdbCode
}

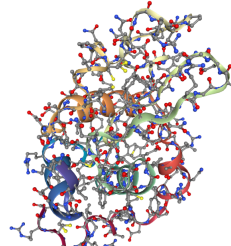


# Create and launch bb
Pdb(output_pdb_path=downloaded_pdb,
    properties=prop).launch()

2020-06-01 14:51:58,256 [MainThread] [INFO] Downloading lakt from: https://files.rcsb.org/download/lakt.pdb
2020-06-01 14:51:58,502 [MainThread] [INFO] Writing pdb to /home/jorjey/biobb_wf_md_setup/notebooks/LAKT.pdb
2020-06-01 14:51:58,504 [MainThread] [INFO] Filtering lines not starting with one of these words: ['ATOM', 'HETM', 'TER', 'ENDMDG']
```

Visualizing 3D structure

Visualizing the downloaded/given PDB structure using NGL:

```
In [3]: # Show protein
view = ngview.show_structure_file(download_pdb)
view.add_representation(repr_type="ball+stick", selection="all")
view.remove_call('rotation', target='Widget', args=('', 600px))
view
```

In general:

- Fantastic tool for **training**
- Inspect **Intermediate** results
- **Interactively** modify parameters
- Possibility to run it in **myBinder**

In particular (biobbs):

- Be familiar with **biobb syntax**
- Learn how to build workflows (**tutorials**)
- **Package** workflow (**Conda**)

```
In [2]: # Downloading desired PDB file
# Import module
from biobb_io.api.pdb import Pdb

# Create properties dict and inputs/outputs
downloaded_pdb = pdbCode+'.pdb'
prop = {
    'pdb_code': pdbCode
}

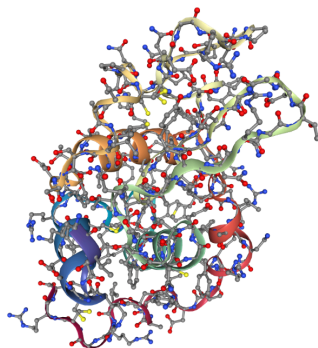
#Create and launch bb
Pdb(output_pdb_path=downloaded_pdb,
    properties=prop).launch()

2020-06-01 14:51:58,256 [MainThread ] [INFO ] Downloading: laki from: https://files.rcsb.org/download/laki.pdb
2020-06-01 14:51:58,502 [MainThread ] [INFO ] Writing pdb to: /home/jovyan/biobb_wf_md_setup/notebooks/LAKI.pdb
2020-06-01 14:51:58,504 [MainThread ] [INFO ] Filtering lines NOT starting with one of these words: ['ATOM', 'MODE
L', 'ENDMDL']
```

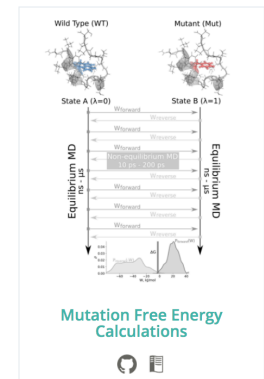
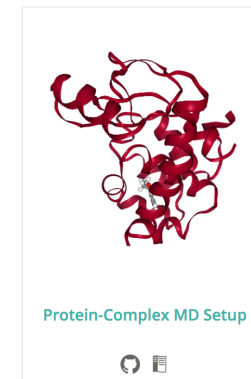
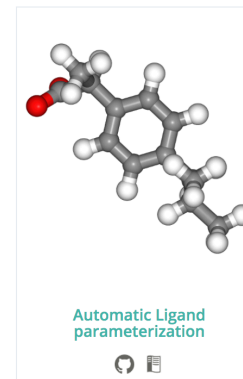
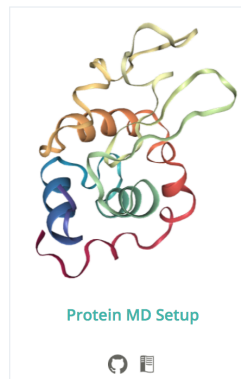
Visualizing 3D structure

Visualizing the downloaded/given PDB structure using NGL:

```
In [3]: # Show protein
view = nglview.show_structure_file(download_pdb)
view.add_representation(repr_type='ball+stick', selection='all')
view._remote_call('setSize', target='Widget', args=['', '600px'])
view
```



DEMONSTRATION WORKFLOWS TUTORIALS





Protein MD Setup tutorial using BioExcel Building Blocks (biobb)

Based on the official GROMACS tutorial: <http://www.mdtutorials.com/gmx/lysozyme/index.html>

This tutorial aims to illustrate the process of **setting up a simulation system** containing a **protein**, step by step, using the **BioExcel Building Blocks library (biobb)**. The particular example used is the **Lysozyme** protein (PDB code 1AKI).

Settings

Biobb modules used

- [biobb_io](#): Tools to fetch biomolecular data from public databases.
- [biobb_model](#): Tools to model macromolecular structures.
- [biobb_md](#): Tools to setup and run Molecular Dynamics simulations.
- [biobb_analysis](#): Tools to analyse Molecular Dynamics trajectories.

Auxiliar libraries used

- [nb_conda_kernels](#): Enables a Jupyter Notebook or JupyterLab application in one conda environment to access kernels for Python, R, and other languages found in other environments.
- [nglview](#): Jupyter/IPython widget to interactively view molecular structures and trajectories in notebooks.
- [ipywidgets](#): Interactive HTML widgets for Jupyter notebooks and the IPython kernel.
- [plotly](#): Python interactive graphing library integrated in Jupyter notebooks.
- [simpletraj](#): Lightweight coordinate-only trajectory reader based on code from GROMACS, MDAnalysis and VMD.

Create protein system topology

Building GROMACS topology corresponding to the protein structure.

Force field used in this tutorial is [amber99sb-ildn](#): AMBER parm99 force field with corrections on backbone (sb) and side-chain torsion potentials (ildn).

Water molecules type used in this tutorial is [spc/e](#).

Adding hydrogen atoms if missing. Automatically identifying disulfide bridges.

Generating two output files:

- GROMACS structure (gro file)
- GROMACS topology ZIP compressed file containing:
 - GROMACS topology top file (top file)
 - GROMACS position restraint file/s (itp file/s)



Building Blocks used:

- [Pdb2gmx](#) from `biobb_md.gromacs.pdb2gmx`

```
In [14]: # Create system topology
# Import module
from biobb_md.gromacs.pdb2gmx import Pdb2gmx

# Create inputs/outputs
output_pdb2gmx_gro = pdbCode+'_pdb2gmx.gro'
output_pdb2gmx_top_zip = pdbCode+'_pdb2gmx_top.zip'

# Create and launch bb
Pdb2gmx(input_pdb_path=fixed_pdb,
        output_gro_path=output_pdb2gmx_gro,
        output_top_zip_path=output_pdb2gmx_top_zip).launch()

2019-12-03 15:49:16,485 [MainThread ] [INFO ] GROMACS Pdb2gmx 20191 version detected
2019-12-03 15:49:16,491 [MainThread ] [INFO ] Not using any container
2019-12-03 15:49:17,750 [MainThread ] [INFO ] gmx -nobackup -nocopyright pdb2gmx -f lAKI_fixed.pdb -o lAKI_pdb2gmx.gro -p p2g.top -water spce -ff amber99sb-ildn -i posre.itp

2019-12-03 15:49:17,756 [MainThread ] [INFO ] Exit code 0

2019-12-03 15:49:17,759 [MainThread ] [INFO ]
Using the Amber99sb-ildn force field in directory amber99sb-ildn.ff

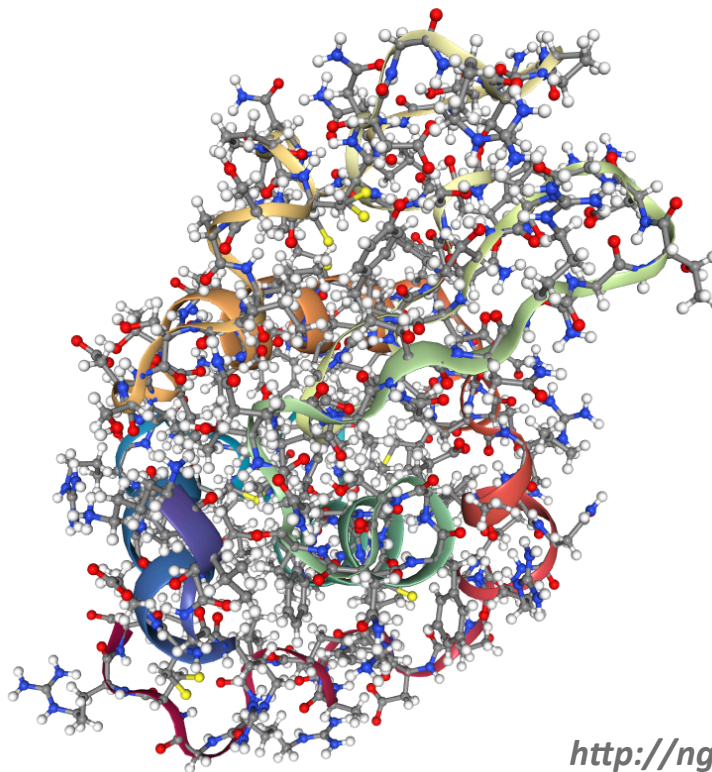
going to rename amber99sb-ildn.ff/aminoacids.r2b
going to rename amber99sb-ildn.ff/dna.r2b
going to rename amber99sb-ildn.ff/rna.r2b
Reading lAKI_fixed.pdb...
Read '', 1001 atoms
Analyzing pdb file
Splitting chemical chains based on TER records or chain id changing.
There are 1 chains and 0 blocks of water and 129 residues with 1001 atoms

chain #res #atoms
```

Visualizing 3D structure

Visualizing the generated **GRO structure** using **NGL**. Note that **hydrogen atoms** were added to the structure by the **pdb2gmx GROMACS tool** when generating the **topology**.

```
In [12]: # Show protein
struct_file = nglview.FileStructure(output_pdb2gmx_gro)
view = nglview.show_file(struct_file)
view.add_representation(repr_type='ball+stick', selection='all')
view._remote_call('setSize', target='Widget', args=['', '600px'])
view
```



<http://nglviewer.org/nglview/latest/>

```

import plotly
import plotly.graph_objs as go

#Read data from file and filter energy values higher than 1000 Kj/mol^-1
with open(output_min_ene_xvg, 'r') as energy_file:
    x,y = map(
        list,
        zip(*[
            (float(line.split()[0]),float(line.split()[1]))
            for line in energy_file
            if not line.startswith(("#", "@"))
            if float(line.split()[1]) < 1000
        ])
    )

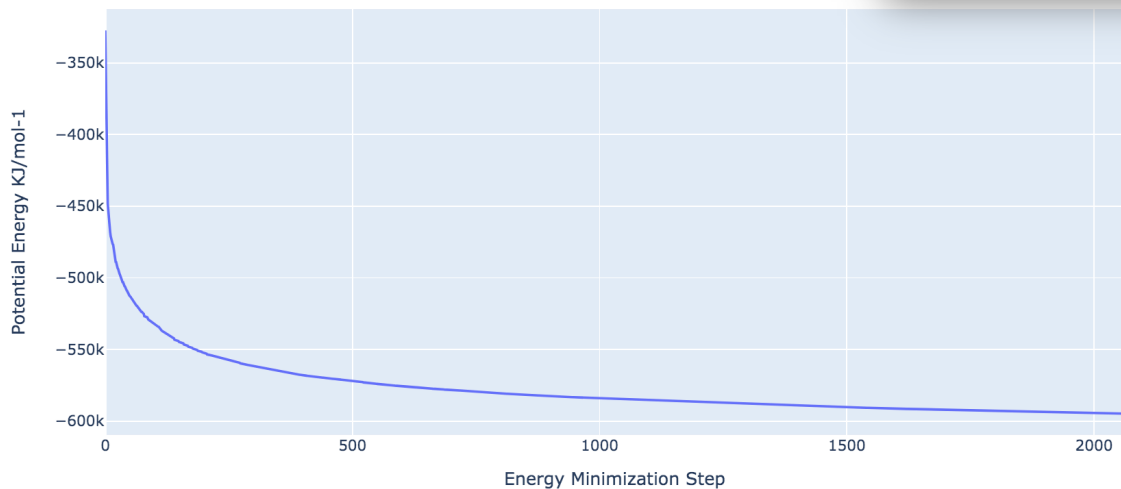
plotly.offline.init_notebook_mode(connected=True)

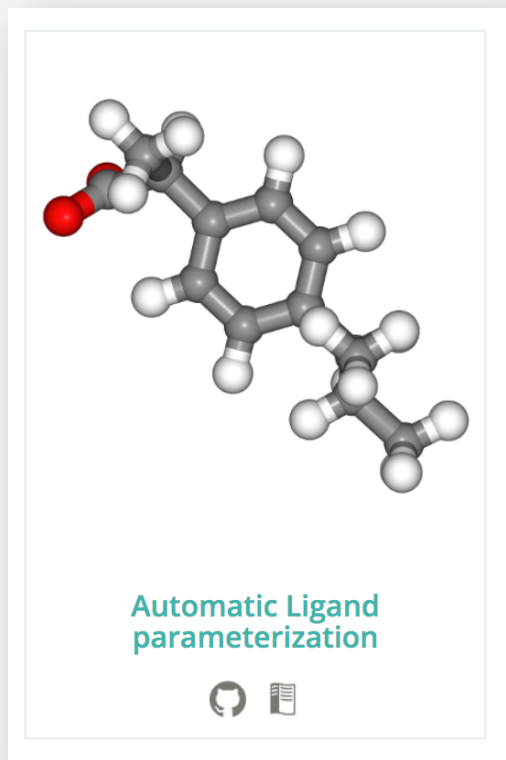
fig = {
    "data": [go.Scatter(x=x, y=y)],
    "layout": go.Layout(title="Energy Minimization",
        xaxis=dict(title = "Energy Minimization Step"),
        yaxis=dict(title = "Potential Energy KJ/mol-1")
    )
}

plotly.offline.iplot(fig)


```

Energy Minimization





Branch: master ▾ [biobb_wf_ligand_parameterization](#) / [conda_env](#) / [environment.yml](#)

 **gbayarri** Removing python as a environment dependency


1 contributor

11 lines (11 sloc) | 211 Bytes

```

1  name: biobb_ligand_parameterization_tutorial
2  channels:
3    - conda-forge
4    - bioconda
5  dependencies:
6    - biobb_common==3.0.0
7    - biobb_io==3.0.0
8    - biobb_chemistry==3.0.1
9    - nb_conda_kernels
10   - nglview
11   - conda

```



Conda Installation and Launch

```

git clone https://github.com/bioexcel/biobb_wf_ligand_parameterization.git
cd biobb_wf_ligand_parameterization
conda env create -f conda_env/environment.yml
conda activate biobb_ligand_parameterization_tutorial
jupyter-nbextension enable --py --user widgetsnbextension
jupyter-nbextension enable --py --user nglview
jupyter-notebook biobb_wf_ligand_parameterization/notebooks/biobb_ligand_parameterization_tutorial.ipynb

```




Turn a Git repo into a collection of interactive notebooks

Have a repository full of Jupyter notebooks? With Binder, open those notebooks in an executable environment, making your code immediately reproducible by anyone, anywhere.

Build and launch a repository


GitHub repository name or URL

GitHub

Git branch, tag, or commit

Path to a notebook file (optional)

Copy the URL below and share your Binder with others:

Copy the text below, then paste into your README to show a binder badge: 

Branch: **master** | [biobb_wf_ligand_parameterization](#) / [conda_env](#) / [environment.yml](#)

 **gbayarri** Removing python as a environment dependency

1 contributor

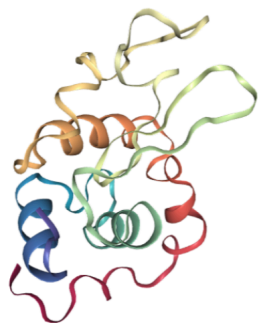
11 lines (11 sloc) | 211 Bytes

```

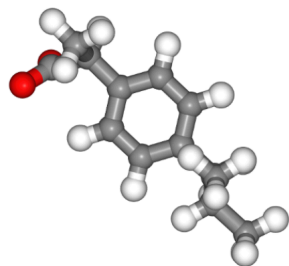
1 name: biobb_ligand_parameterization_tutorial
2 channels:
3   - conda-forge
4   - bioconda
5 dependencies:
6   - biobb_common==3.0.0
7   - biobb_io==3.0.0
8   - biobb_chemistry==3.0.1
9   - nb_conda_kernels
10  - nglview
11  - conda
  
```



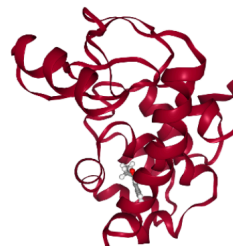
DEMONSTRATION WORKFLOWS TUTORIALS



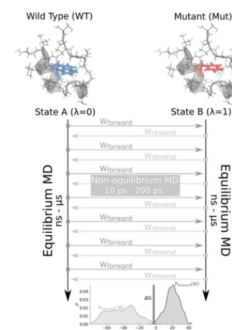
Protein MD Setup



Automatic Ligand parameterization



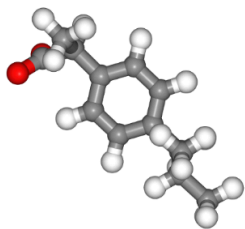
Protein-Complex MD Setup



Mutation Free Energy Calculations



AUTOMATIC LIGAND PARAMETERIZATION



This tutorial aims to illustrate the process of ligand parameterization for a small molecule, step by step, using the BioExcel Building Blocks library (biobb). The particular example used is the Ibuprofen small compound (3-letter code IBP, Drugbank code DB01050), a non-steroidal anti-inflammatory drug (NSAID) derived from propionic acid and it is considered the first of the propionics.

[Execute in binder](#)

[View tutorial](#)

[Open Github repository](#)

[Open documentation](#)



Biomolecular workflows using biobb HPC Workflows

Enabling scalability

Making biomolecular
simulation workflows
exascale ready



**200 * 192 cores(4 nodes)
38,400 cores**

jupyter biobb_MDsetup_tutorial Last Checkpoint: 12/03/2019 (unsaved changes) ✓

File Edit View Insert Cell Kernel Widgets Help

New Notebook ▶ ↑ ↓ ▶ Run ■ ↺ ▶ Markdown ▾

Open...

Make a Copy...

Save as...

Rename...

Save and Checkpoint

Revert to Checkpoint ▶

Print Preview

Download as ▶

Trusted Notebook

Close and Halt

AsciiDoc (.asciidoc)

HTML (.html)

LaTeX (.tex)

Markdown (.md)

Notebook (.ipynb)

PDF via LaTeX (.pdf)

reST (.rst)

Python (.py)

Reveal.js slides (.slides.html)

hing PDB structure

loading **PDB structure** with the **protein molecule** from the RCSB PDB database. Alternatively, a **PDB file** can be used as starting structure.

In [8]: `# Down`
`# Imp`
`from`
`# Cre`
`downl`
`prop = {`

biobb_MDsetup_tutorial.py

```
(biobb_MDsetup_tutorial) Adams-MacBook-Pro:Python hospital$ python biobb_MDsetup_tutorial.py
/anaconda3/envs/biobb_MDsetup_tutorial/lib/python3.6/site-packages/biobb_common/tools/file_utils.py:327: UserWarning: Warning: filter is not a recognized property. The most similar property is: filter_error_property, close_property))
2019-12-11 08:26:44,183 [MainThread ] [INFO ] Downloading: 1aki from: https://files.rcsb.org/download/1aki.pdb
2019-12-11 08:26:44,891 [MainThread ] [INFO ] Writing pdb to: /Users/hospital/biobb_tutorials/biobb_wf_md_setup/biobb_wf_md_setup/Python/1AKI.pdb
2019-12-11 08:26:44,891 [MainThread ] [INFO ] Filtering lines NOT starting with one of these words: ['ATOM']
2019-12-11 08:26:45,386 [MainThread ] [INFO ] check_structure -i 1AKI.pdb -o 1AKI_fixed.pdb --force_save fixside --fix ALL
2019-12-11 08:26:45,387 [MainThread ] [INFO ] Exit code 0
```

Disadvantages:

- Graphical cells not showing.
- Losing interactivity

Advantages:

- Gaining High Throughput (automation, repetition)

Problem:

- Modify parameters for a certain step
→ modify the Python script



Workflow script

- Building blocks
- Python code
- Loops / conditionals
- Global log
- Output folders hierarchy



Workflow parameters

- Steps Inputs / Outputs
- Steps Dependencies
- Steps Properties
- Workflow inputs
- Workflow parameters



Workflow parameters

```

31 global_log.info("step1_pdb: Download the initial Structure")
32 Pdb(**global_paths["step1_pdb"], properties=global_prop["step1_pdb"]).launch()
33
34 global_log.info("step2_fixsidechain: Modeling the missing heavy atoms in the structure side chains")
35 FixSideChain(**global_paths["step2_fixsidechain"], properties=global_prop["step2_fixsidechain"]).launch()
36
37 global_log.info("step3_pdb2gmx: Generate the topology")
38 Pdb2gmx(**global_paths["step3_pdb2gmx"], properties=global_prop["step3_pdb2gmx"]).launch()
39
40 global_log.info("step4_editconf: Create the solvent box")
41 Editconf(**global_paths["step4_editconf"], properties=global_prop["step4_editconf"]).launch()
42
43 global_log.info("step5_solvate: Fill the solvent box with water molecules")
44 Solvate(**global_paths["step5_solvate"], properties=global_prop["step5_solvate"]).launch()
45
46 global_log.info("step6_grompp_genion: Preprocess ion generation")
47 Grompp(**global_paths["step6_grompp_genion"], properties=global_prop["step6_grompp_genion"]).launch()
48
49 global_log.info("step7_genion: Ion generation")
50 Genion(**global_paths["step7_genion"], properties=global_prop["step7_genion"]).launch()
51
52 global_log.info("step8_grompp_min: Preprocess energy minimization")
53 Grompp(**global_paths["step8_grompp_min"], properties=global_prop["step8_grompp_min"]).launch()
54
55 global_log.info("step9_mdrun_min: Execute energy minimization")
56 Mdrun(**global_paths["step9_mdrun_min"], properties=global_prop["step9_mdrun_min"]).launch()
57
58 global_log.info("step10_energy_min: Compute potential energy during minimization")
59 GMXEnergy(**global_paths["step10_energy_min"], properties=global_prop["step10_energy_min"]).launch()

```

```

1 # Example of a YAML configuration file for a BioExcel building blocks workflow
2
3 working_dir_path: md_tutorial # Folder to write i/o files of the workflow steps
4 can_write_console_log: False # Verbose writing of log information
5 restart: False # Skip steps already performed
6 remove_tmp: True
7
8 step1_pdb:
9   paths:
10    output_pdb_path: structure.pdb
11   properties:
12    pdb_code: 1aki
13
14 step2_fixsidechain:
15   paths:
16    input_pdb_path: dependency/step1_pdb/output_pdb_path
17    output_pdb_path: fixsidechain.pdb
18
19 step3_pdb2gmx:
20   paths:
21    input_pdb_path: dependency/step2_fixsidechain/output_pdb_path
22    output_gro_path: pdb2gmx.gro
23    output_top_zip_path: pdb2gmx_top.zip
24
25 step4_editconf:
26   paths:
27    input_gro_path: dependency/step3_pdb2gmx/output_gro_path
28    output_gro_path: editconf.gro
29
30 step5_solvate:
31   paths:
32    input_solute_gro_path: dependency/step4_editconf/output_gro_path
33    output_gro_path: solvate.gro
34    input_top_zip_path: dependency/step3_pdb2gmx/output_top_zip_path
35    output_top_zip_path: solvate_top.zip
36

```

Workflow script



```

24 # Loading the biobb configuration reader
25 conf = settings.ConfReader(sys.argv[1])
26
27 global_log, _ = fu.get_logs(path=conf.get_working_dir_path())
28 global_prop = conf.get_prop_dic(global_log=global_log)
29 global_paths = conf.get_paths_dic()
30
31 global_log.info("step1_pdb: Download the initial Structure")
32 Pdb(**global_paths["step1_pdb"], properties=global_prop["step1_pdb"]).launch()
33
34 global_log.info("step2_fixsidechain: Modeling the missing heavy atoms in the structure side chains")
35 FixSideChain(**global_paths["step2_fixsidechain"], properties=global_prop["step2_fixsidechain"]).launch()
36
37 for mutation in conf.properties['mutations_list']:
38
39     mut_paths = conf.get_paths_dic(mutation)
40     mut_prop = conf.get_prop_dic(mutation, global_log=global_log)
41
42     mut_paths['step3_mutate']['input_pdb_path'] = global_paths['step2_fixsidechain']['output_pdb_path']
43
44     global_log.info("step3_mutate: Modeling a particular residue mutation")
45     Mutate(**mut_paths["step3_mutate"], properties=mut_prop["step3_mutate"]).launch()
46
47     global_log.info("step4_pdb2gmx: Generate the topology")
48     Pdb2gmx(**mut_paths["step4_pdb2gmx"], properties=mut_prop["step4_pdb2gmx"]).launch()

```



```

7
8 mutations_list: ["A:Arg5Ala", "A:Arg5Gly", "A:Arg5Lys"]
9
10 step1_pdb:
11   paths:
12     output_pdb_path: structure.pdb
13   properties:
14     pdb_code: 1aki

```

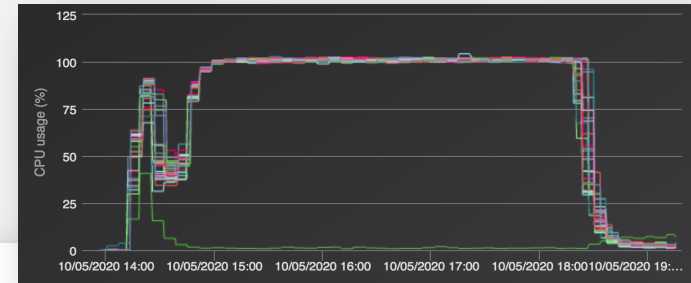
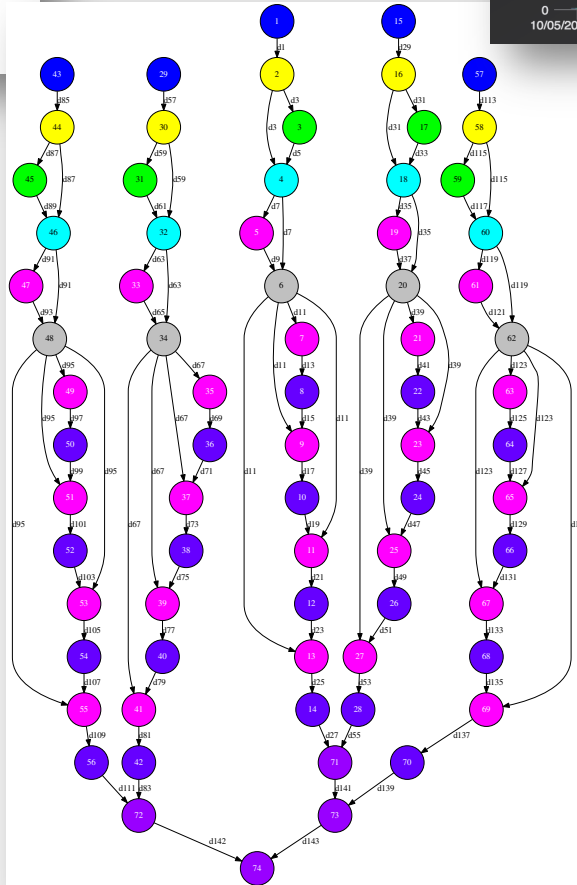



```
(biobb_Protein-Complex_MDsetup_tutorial) Adams-MacBook-Pro:Yaml hospital$ ls
biobb_MDsetup_tutorial-AlaScan.py  biobb_MDsetup_tutorial-lite.yaml  biobb_MDsetup_tutorial.py
biobb_MDsetup_tutorial-AlaScan.yaml  biobb_MDsetup_tutorial-mut.py      biobb_MDsetup_tutorial.yaml
biobb_MDsetup_tutorial-lite.py      biobb_MDsetup_tutorial-mut.yaml    md_tutorial_mut
(biobb_Protein-Complex_MDsetup_tutorial) Adams-MacBook-Pro:Yaml hospital$ ls -lrht md_tutorial_mut/
total 64
-rw-r--r--  1 hospital  staff    0B Dec 11 22:43 log.err
drwxr-xr-x  5 hospital  staff  170B Dec 11 22:43 step2_fixsidechain
drwxr-xr-x  5 hospital  staff  170B Dec 11 22:43 step1_pdb
dr-xr-xr-x 24 hospital  staff  816B Dec 11 22:52 A:Arg5Ala
dr-xr-xr-x 24 hospital  staff  816B Dec 11 23:01 A:Arg5Gly
dr-xr-xr-x 24 hospital  staff  816B Dec 11 23:11 A:Arg5Lys
-rw-r--r--  1 hospital  staff   30K Dec 11 23:16 log.out
(biobb_Protein-Complex_MDsetup_tutorial) Adams-MacBook-Pro:Yaml hospital$ ls -rlht md_tutorial_mut/A\:Arg5Ala/
total 0
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:43 step5_editconf
dr-xr-xr-x  6 hospital  staff  204B Dec 11 22:43 step4_pdb2gmx
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:43 step3_mutate
dr-xr-xr-x  6 hospital  staff  204B Dec 11 22:43 step8_genion
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:43 step7_grompp_genion
dr-xr-xr-x  6 hospital  staff  204B Dec 11 22:43 step6_solvate
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:43 step9_grompp_min
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:45 step12_grompp_nvt
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:45 step11_energy_min
dr-xr-xr-x  8 hospital  staff  272B Dec 11 22:45 step10_mdrun_min
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:47 step14_energy_nvt
dr-xr-xr-x  9 hospital  staff  306B Dec 11 22:47 step13_mdrun_nvt
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:47 step15_grompp_npt
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:50 step18_grompp_md
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:50 step17_energy_npt
dr-xr-xr-x  9 hospital  staff  306B Dec 11 22:50 step16_mdrun_npt
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:52 step21_rmsexp
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:52 step20_rmsfirst
dr-xr-xr-x  9 hospital  staff  306B Dec 11 22:52 step19_mdrun_md
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:52 step24_dry
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:52 step23_image
dr-xr-xr-x  5 hospital  staff  170B Dec 11 22:52 step22_rgyr
```

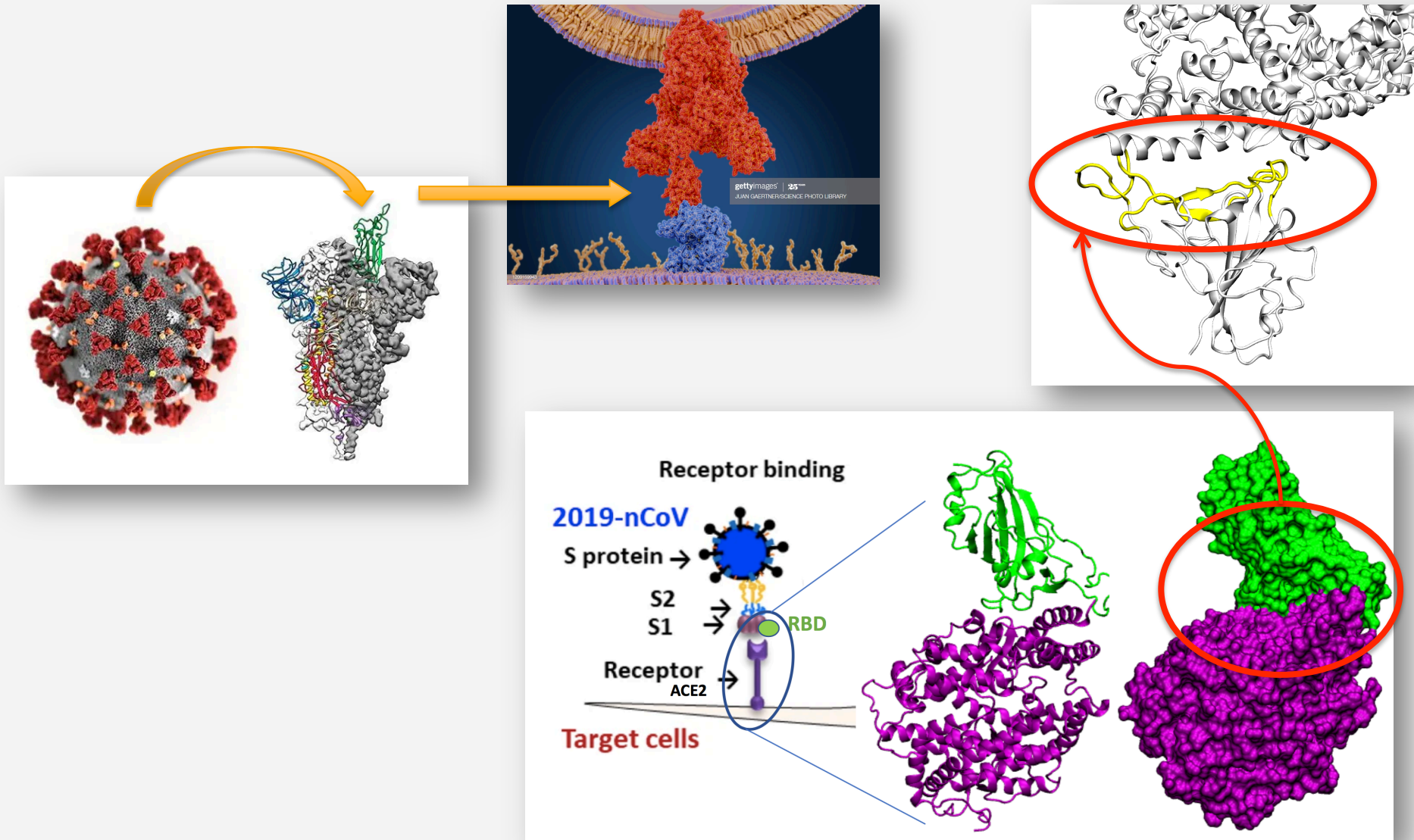
```
print 'step2: mmbuniprot -- Get mutations'
mmbuniprot = uniprot.MmbVariants(prop['pdb_code'])
mutations = mmbuniprot.fetch_variants()

for mut in mutations:
    mut_path = cdir(wd, mut)

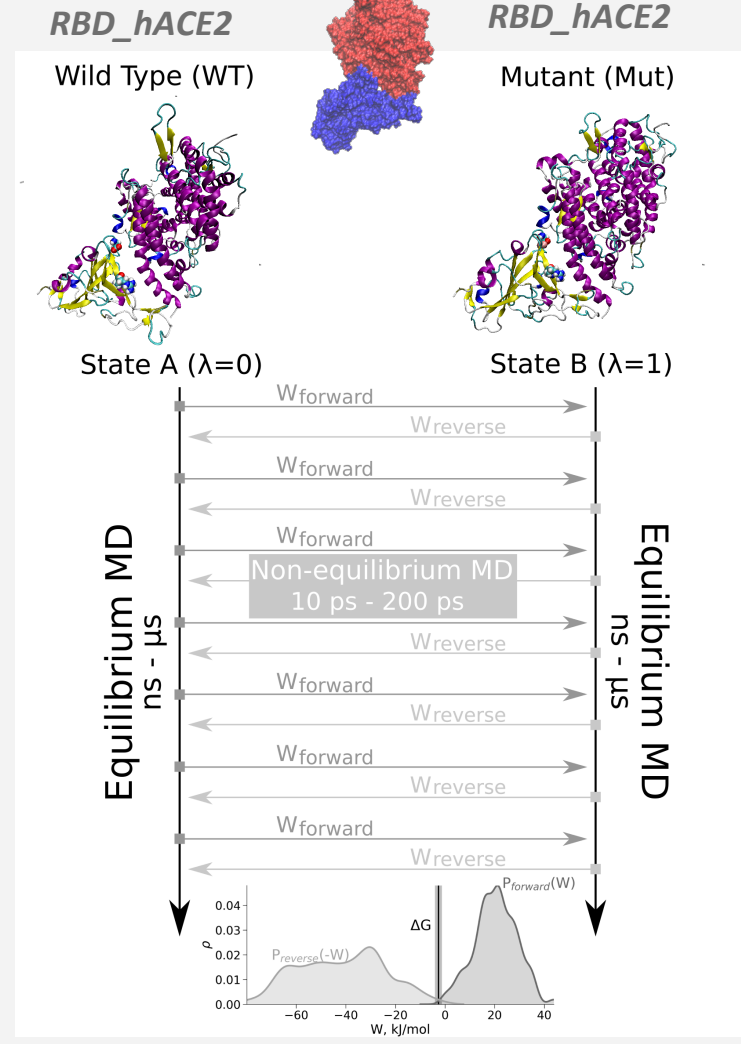
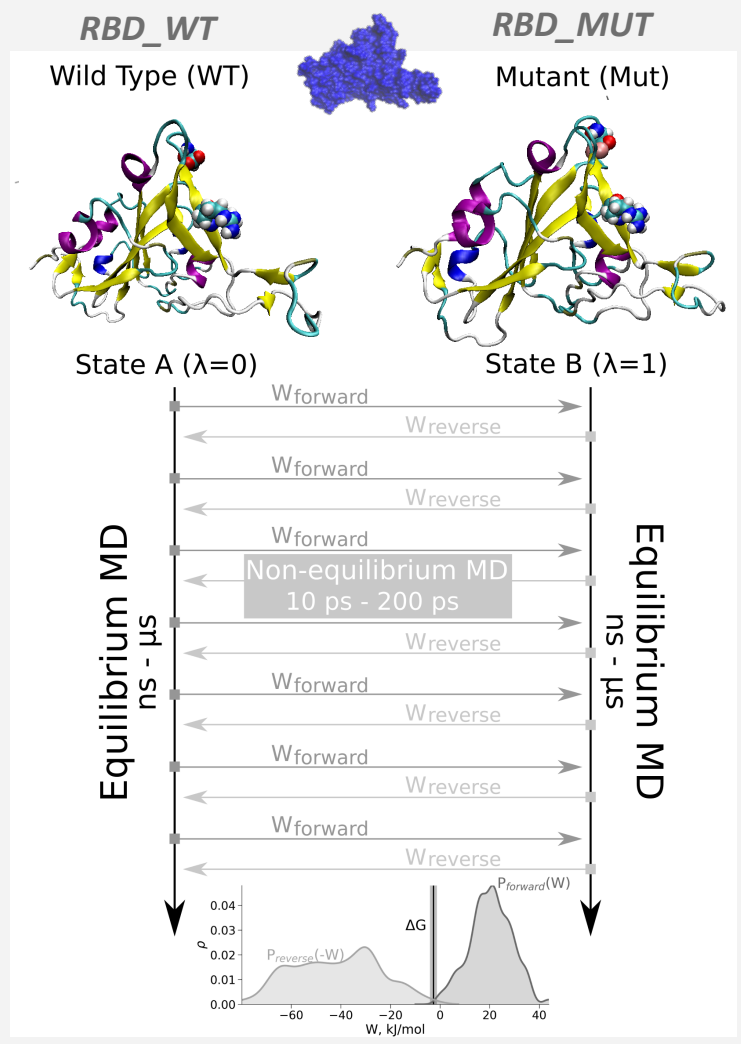
print 'step3: scw -- Model mutation'
scw_path = cdir(mut_path, 'step3_scw')
scw_pdb = opj(scw_path, prop['mutated_pdb'])
scw = scwr1.Scwr14(mmbpdb_pdb, scw_pdb, mut, scwr1_path=scwr1_path)
scw_pdb2 = scw.launchPyCOMPSs()
```

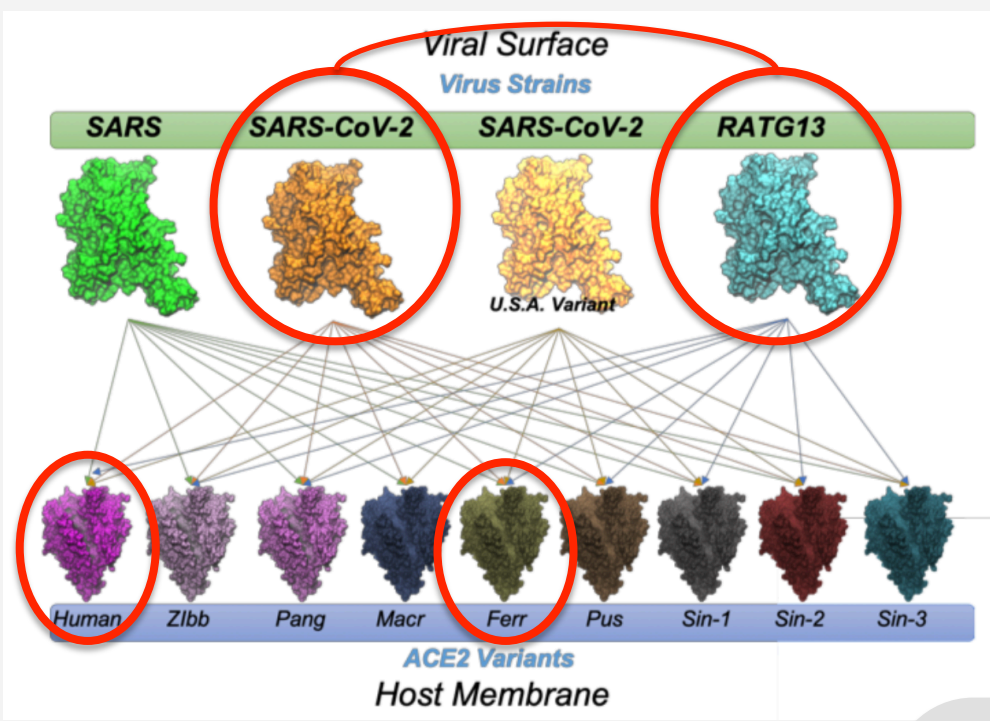


Example: pre-exascale COVID-19 biobb workflow



Alchemical free energy calculations of relative protein binding free energy difference





Big Data Study:
 PRACE computational power
 Massive amount of data

Impact of mutations in binding affinity

Molecular Dynamics simulation data

- For each mutation:
 - MD Simulations (RBD + ACE2 + Complex)
 - Free energy calculations



- **Fast-growth Thermodynamic Integration**
- **1000 independent short MD simulations** (500 forward + 500 reverse)
- **GROMACS + pmx**
- **Extremely parallelizable**



pmx: generate hybrid protein structure and topology
 Computational Biomolecular Dynamics Group

```
bsc@lg1:~/PMX> python mdlaunchCV.py -h
```

Workflow to **model, setup** and **run MD simulations** for a set of **mutations**.

Usage: mdlaunchCV.py [-h]

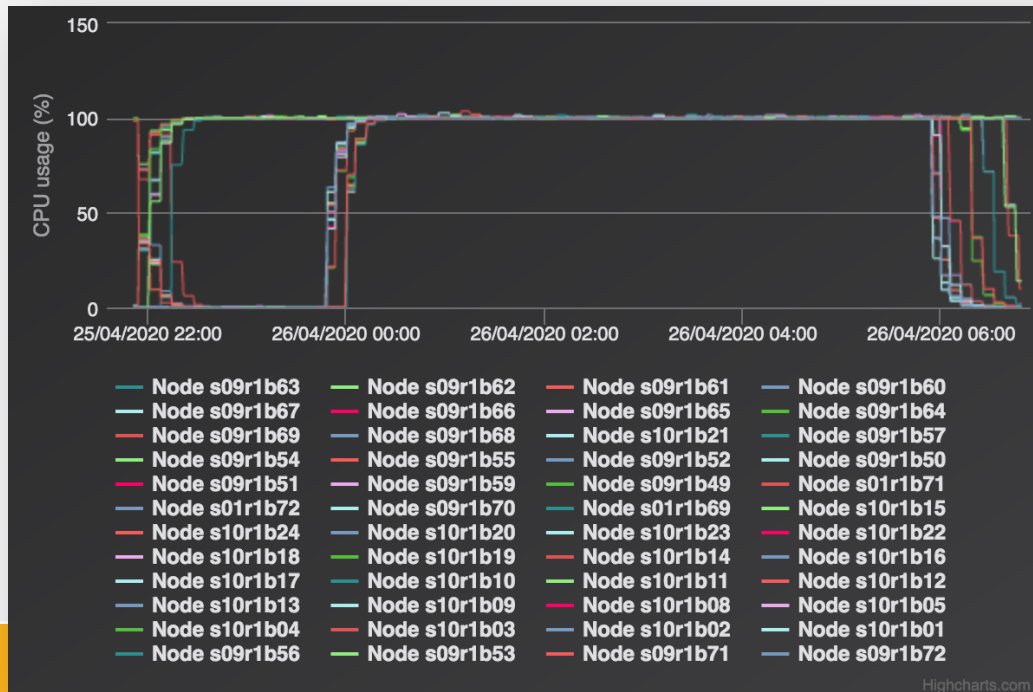
-m MUTATION -wt WT_STRUCTURE

[-q QUEUE] [-t TIME] [-nn NUM_NODES]

[-c CUMULATIVE] [-mpi MPI_NODES]

[-o OUTPUT_DIR] [-jn JOB_NAME] [-l MD_LENGTH]

```
python mdlaunchCV.py
-m WT+B:F756S
-c False
-wt complex.pdb
-l 10000
-nn 8
-mpi 4
-o COMPLEX_MUTS
-q bsc_ls
-t 2800
-jn mds_F756S_covid
```



48 MareNostrum nodes
2,304 cores → 1 job

12 mutations
10ns-length MDs
GROMACS 4 nodes MPI

Time: 8h

```
bsc@lg1:~/PMX> python pmxlaunchCV.py -h
```

Free Energy estimation upon amino acid modification with fast growth Thermodynamic Integration using GROMACS and pmx tools.

Usage: `pmxlaunchCV.py [-h]`

`-m MUTATION`

`-wt_top WT_TOPOLOGY -wt_trj WT_TRAJECTORY`

`-mut_top MUT_TOPOLOGY -mut_trj MUT_TRAJECTORY`

`[-q QUEUE] [-t TIME] [-nn NUM_NODES] [-fe FE_LENGTH]`

`[-nf NUM_FRAMES] [-prn PMX_RESNUM]`

```
python pmxlaunchCV.py
```

```
-m Lys8Arg
```

```
-wt_top wt.tpr
```

```
-wt_trj wt.xtc
```

```
-mut_top mut.tpr
```

```
-mut_trj mut.xtc
```

```
-nf 500
```

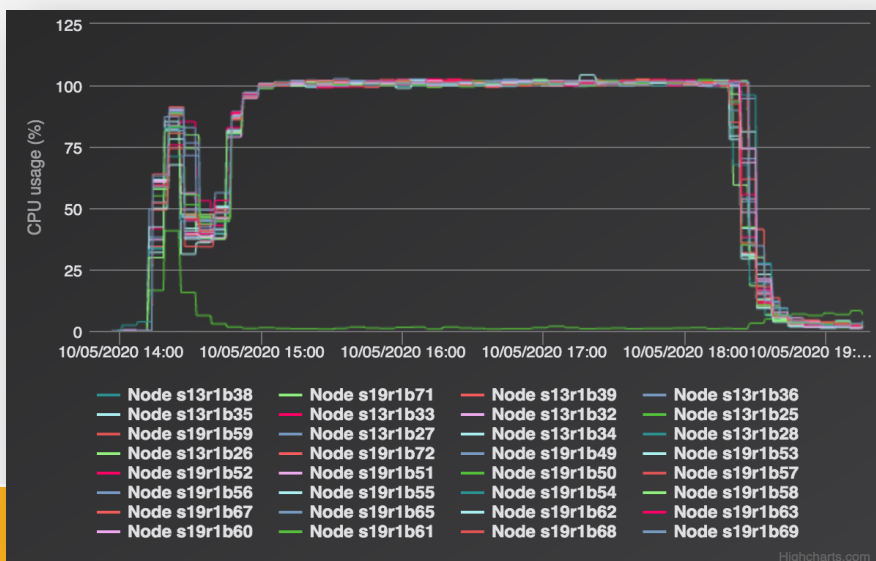
```
-q bsc_ls
```

```
-jn aceK8R_covid
```

```
-o Lys8Arg
```

```
-nn 16
```

```
-t 2800
```



32 MareNostrum nodes
1,536 cores → 1 job

1 mutation (RBD-ACE2)
1000 short TI MDs (50ps)

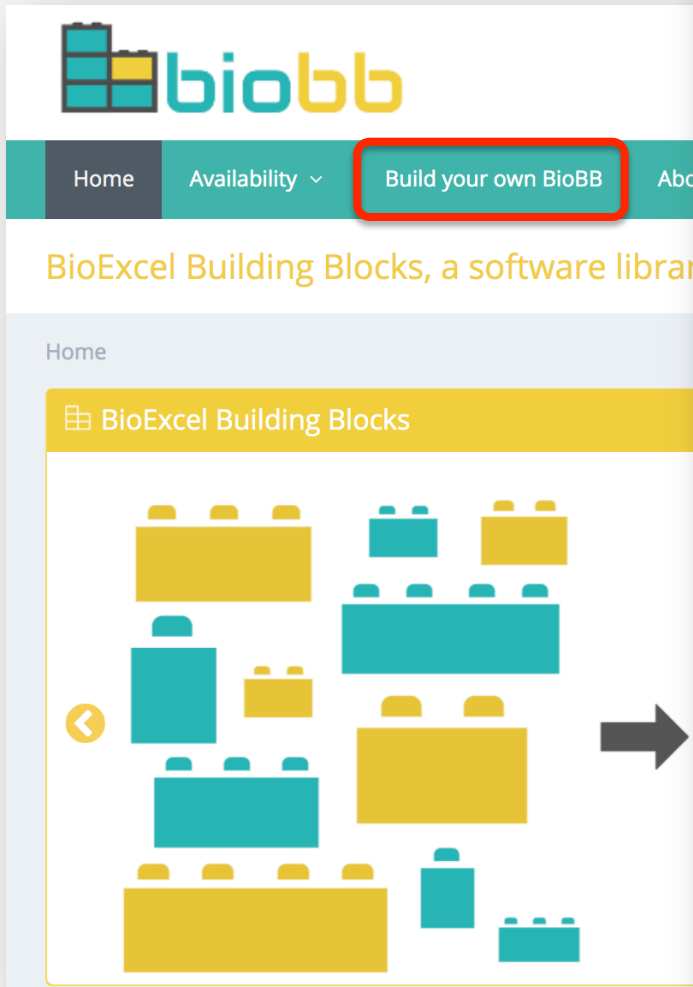
500 forward

+

500 reverse

Time: 5h

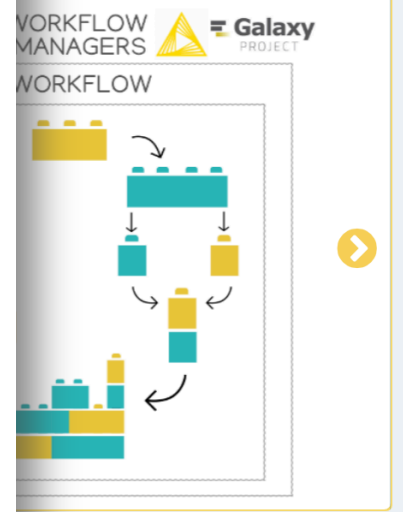
- **BioExcel Building Blocks** software library offers a new layer of interoperability on biomolecular software tools.
- **Biomolecular workflows**, portable and reproducible, can be easily built using the library.
- The additional **workflow manager compatibility layer** allows **workflows** built with the library to be launched and controlled with **different frameworks** and in **different infrastructures**:
 - **Demonstration** workflows (Jupyter Notebooks)
 - **Pre-exascale HPC** workflows (PyCOMPSs)



Bonus Track!

celona.org/biobb/

orkflows





Remote BioExcel Winter School on Biomolecular Simulations

📅 November 30, 2020 👤 Michelle Mendonca



📺 VIRTUAL TRAININGS

Direct interaction with trainers!

Virtual Trainings are training events done in a virtual or simulated environment, with the learner and the instructor being in separate locations. They are designed to simulate the traditional classroom or learning experience. Similarly to the webinars series, **BioExcel Virtual Training series** for computational biomolecular research also cover a broad range of topics related to the biomolecular simulation field. The first **Virtual Trainings** for the PDBe Knowledge Base (PDBe-KB) and for the **BioExcel Building Blocks** have already been run, but new sessions are being planned: **PDBe-KB**, **biobb**

- Training #1, 09/12/2019 - 12/12/2019: **biobb_VT** - **biobb_VT_wfs** - **biobb_VT_ex** - **biobb_VT_cli** - **examples**



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<p>Protein MD Setup</p>	<p>Automatic Ligand parameterization</p>	<p>Protein-Complex MD Setup</p>	<p>Mutation Free Energy Calculations</p>
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2020 Remote BioExcel Summer School on Biomolecular Simulations

3	<p>23:54</p>	<p>Basis of molecular dynamics simulations - Part 1 BioExcel Center of Excellence for Computational Biomolecular Research</p>
4	<p>28:06</p>	<p>Basis of molecular dynamics simulations - Part 2 BioExcel Center of Excellence for Computational Biomolecular Research</p>
5	<p>41:25</p>	<p>BioExcel Building Blocks - Part 1 BioExcel Center of Excellence for Computational Biomolecular Research</p>
6	<p>33:30</p>	<p>BioExcel Building Blocks - Part 2 BioExcel Center of Excellence for Computational Biomolecular Research</p>

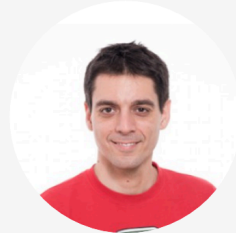
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<http://mmb.irbbarcelona.org/biobb/>

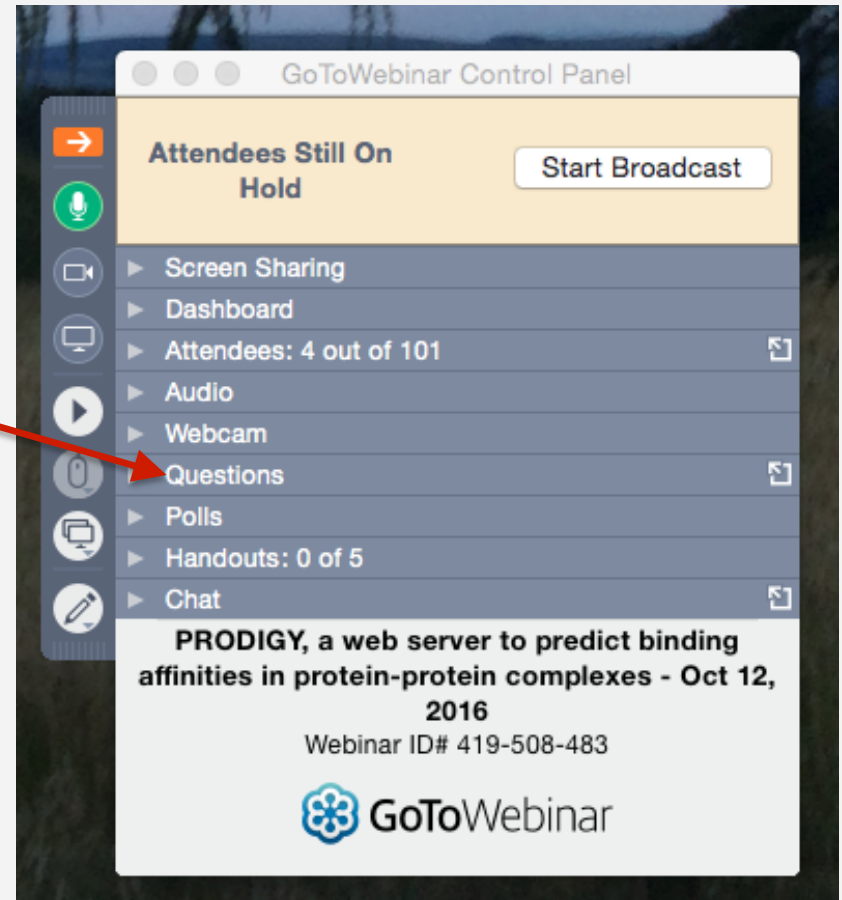
adam.hospital@irbbarcelona.org



Please use the **Questions** function in **GoToWebinar** application

- If you *don't have audio*, please mention that in the question.

Any other questions or points to discuss after the live webinar? Join the discussions at <http://ask.bioexcel.eu>





Special edition Student
Webinar: Summer School 2020
Edition (2020-09-15)

Molywood: streamlining
the design and rendering
of molecular movies

by Miłosz Wieczór
(2020-10-22)



See <https://bioexcel.eu/>