



Article

Functionalized Periodic Mesoporous Organosilica Nanoparticles for Loading and Delivery of Suramin

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Abstract: Suramin (SUR) is a known drug for treating parasitic infections though research studies and some clinical trials have shown its applicability for a plethora of other diseases. Herein we report on a novel SUR nanocarrier for the drug delivery to cells. We synthesized periodic mesoporous organosilica nanoparticles with spherical morphology, having mean diameter of 240 nm and high surface area (778 m²/g). The material's surface is modified with an amine-containing organic moiety *N*-[3-(Trimethoxysilyl)propyl]ethylenediamine (DA), followed by surface attachment with the drug. The rate of SUR release in physiological condition was low, though in vitro experiments on MRC-5 cell line demonstrate effective delivery of the drug to the cells and low toxicity of the materials without the adsorbed drug. These results are promising for opening new treatment strategies with SUR-bearing nanocarriers, with high efficiency and low adverse effects on healthy tissues.

Keywords: drug delivery; nanocarrier; porous nanoparticles; cancer treatment

1. Introduction

Originally developed for the treatment of parasitic infections such as trypanosomiasis and onchocerciasis, the drug Suramin (SUR) has been known since 1920s. [1]. However, the research studies increasingly reveal new biological targets for SUR, opening up its application for the treatment of different diseases such as AIDS [2], ZIKA virus [3], human enteroviruses [4], and cancer [5,6] while novel small, phase I/II clinical trial even revealed the potential of low doses of SUR for the treatment of Autism [7]. Hence, it is of substantial interest to develop safe SUR nanocarriers for achieving controlled and targeted drug delivery and thus increasing its therapeutic efficacy with inhibiting the occurrence of side effects.

SUR nanocarriers were previously constructed by entrapping the drug within surfactant-silica hybrid nanoparticles in situ, i.e., during the synthesis procedure of the material, and demonstrated for effective anti-angiogenic application [8]. Herein, we investigate the potential of pre-synthesized periodic mesoporous organosilica (PMO) nanomaterial to act as a platform for surface adsorption and delivery of SUR. PMO nanoparticles are distinguished as highly promising for drug delivery and other applications [9–13], predominantly due to their high surface area, possibilities for versatile surface modifications, stimuli-responsive degradability and high biocompatibility. Previously, we demonstrated the potential of PMO nanoparticles for efficient cancer therapy and diagnostics [14,15], as well as for skin protection from UV irradiation [16]. Mesoporous silica nanoparticles (MSN) are more studied structural analogues of PMO for cancer therapy and plethora of other applications [17–21]. MSN and PMO have the same silica surface though the inner composition of PMO particles is composed of organic bridges, while MSN is made entirely of SiO₂ framework, which may offer

significant benefits for PMO in terms of biocompatibility, biodegradability, photoluminescence and other novel properties [22]. Herein, we constructed ethylene-bridged PMO nanoparticles (PMOBTE) and functionalized their surface with *N*-[3-(trimethoxysilyl)propyl]-ethylenediamine in order to favor the interaction of the positively charged amine moieties in aqueous environment with the negatively charged sulfonate groups from the SUR molecule and enhance the drug loading. The efficacy of the drug loaded material for delivery of SUR was further investigated by *in vitro* experiments on human fetal lung fibroblast MRC-5 cell line. This cell line was chosen to demonstrate the proof of concept for drug delivery to cancer cells but also the previous studies evidence that these healthy fibroblast cells promote cancer cell motility and invasiveness [23], which identifies them as a potential target for battling cancer, if the treatment is localized to the tumor microenvironment.

2. Results and Discussion

Scanning electron micrograph (SEM) image of PMOBTE is shown on Figure 1a. The material consists of spherical nanoparticles with diameters in the range 110–380 nm, though the most nanoparticles are of the size 240 ± 30 nm. Nitrogen sorption analysis revealed that the material possesses high Brunauer–Emmett–Teller (BET) surface area ($778 \text{ m}^2/\text{g}$) with Barrett–Joyner–Halenda (BJH) pore determination indicating microporous nature of the nanoparticles (<2 nm) with a shoulder in the mesoporous region (2.8 nm). The material has a wormhole porous structure, as evidenced by the transmission electron micrograph (TEM) image (inset on Figure 1a).

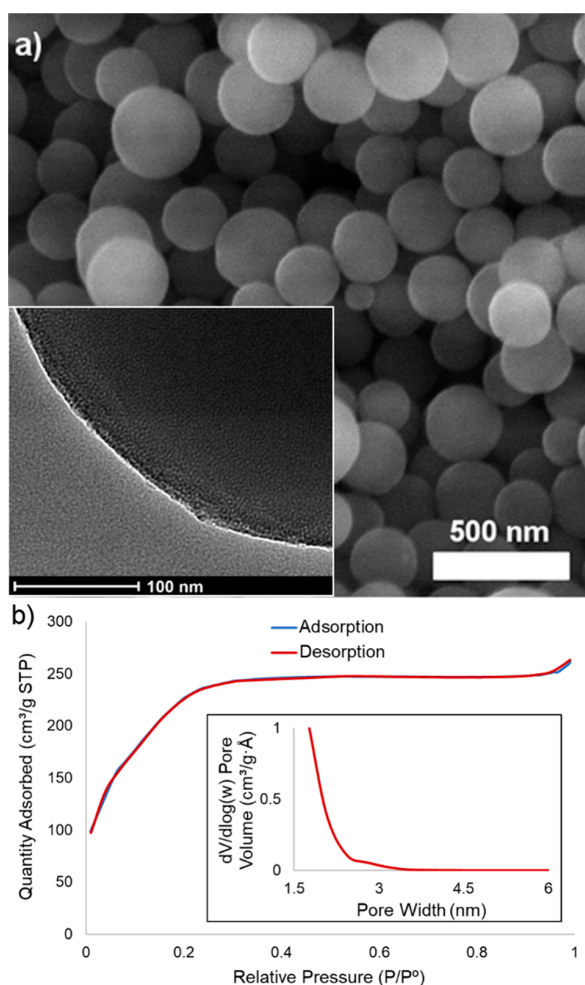


Figure 1. Characterization of ethylene-bridged PMO nanoparticles (PMOBTE) material with: (a) scanning electron micrograph and transmission electron micrograph (inset) (b) nitrogen adsorption–desorption isotherm and Barrett–Joyner–Halenda (BJH) pore size distribution plot (inset).

The material was functionalized with *N*-[3-(trimethoxysilyl)propyl]ethylenediamine (DA) followed by adsorption of SUR on its surface. Functionalized material was loaded with the drug SUR through electrostatic interaction between the positively charged amine groups on the PMO surface and the negatively charged sulfonate groups of SUR. The amount of adsorbed SUR was determined by comparison of starting and the final concentration of the drug during the loading procedure. This calculated amount was 83 μg of SUR per milligram of PMOBTE or 8.3 wt %. The infrared spectroscopy (Figure 2a) confirmed the structure of PMOBTE, the presence of surface functionalized DA and successful loading of SUR. All the spectra contain the peaks at 1412 cm^{-1} (CH_2 δ) and at 1272 cm^{-1} ($\text{CH}_2\text{-Si}$ δ) vibrations of the bridging ethylene moiety within the framework. In the range $1000\text{--}1200\text{ cm}^{-1}$ all the spectra show peaks from different Si-O vibrations within the nanoparticles or on its surface. Upon surface functionalization with DA, the appearance of strong band centered at 1655 cm^{-1} is evident, which is characteristic for δ NH_2 and NH vibrations from the functionalized moiety. The broader peak of weaker intensity at 1631 cm^{-1} in the spectrum of PMOBTE is typical for the surface-adsorbed water. In the spectrum of the SUR (Figure 2b)-adsorbed material (SUR@DA-PMOBTE) SUR S=O valence vibration is evident as an additional peak in the heavily overlapping area $1000\text{--}1200\text{ cm}^{-1}$, centered at 1136 cm^{-1} while the appearance of amide band of SUR is also noticeable in the spectrum of SUR@DA-PMOBTE, centered at 1542 cm^{-1} .

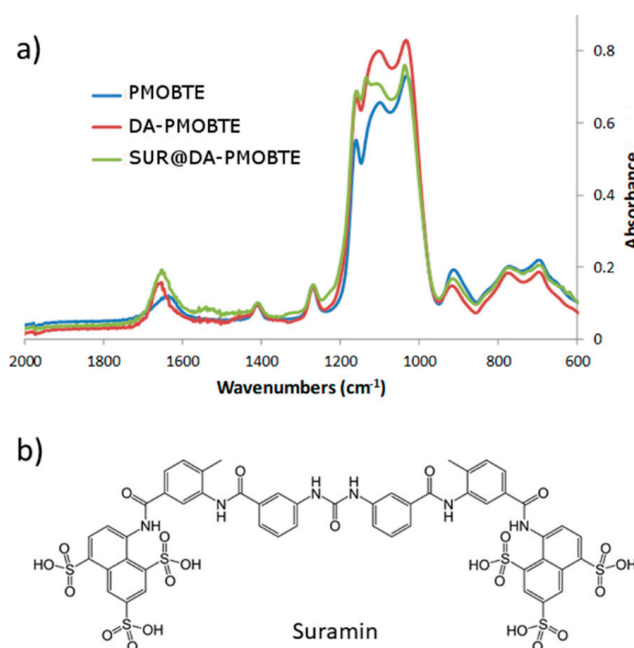


Figure 2. (a) Infrared spectra of the synthesized materials and (b) the molecular structure of Suramin (SUR).

The structure of the synthesized materials was further confirmed by thermogravimetric (TGA) analyses (Figure 3). All three materials show significant weight loss below $100\text{ }^\circ\text{C}$ due to surface-adsorbed water. The material PMOBTE shows a single sharp loss of mass centered at $462\text{ }^\circ\text{C}$, which can be ascribed to the bridging ethylene group. In case of DA-PMOBTE, more complex weight loss pattern is observed, clearly evidencing successful functionalization of PMOBTE. The weight loss centered at $176\text{ }^\circ\text{C}$ can be attributed to chemically bonded water molecules to the surface, i.e., possibly hydroxyl ions, attached to the amine groups of the functionalized DA moiety. The difference in weight loss above $250\text{ }^\circ\text{C}$ between DA-PMOBTE and PMOBTE indicates the amount of grafted DA on the surface (14.7 wt %). Significant loss of weight for both DA-PMOBTE and SUR@DA-PMO is observed above $300\text{ }^\circ\text{C}$, with two major bands, observed in the Derivative weight plot, centered at 361 and $540\text{ }^\circ\text{C}$ for DA-PMOBTE and at 358 and $530\text{ }^\circ\text{C}$ for SUR@DA-PMOBTE. The shift of these bands as well as the change in their relative intensity indicates the successful attachment of SUR to the

surface. Furthermore, in case of SUR@DA-PMOBTE the weight loss at 176 °C is less pronounced when compared to the analysis of DA-PMOBTE, as SUR replaced the hydroxyl groups as the counterion for amine groups of the functionalized DA, while a new weak band in the Derivative weight plot appears at 300 °C.

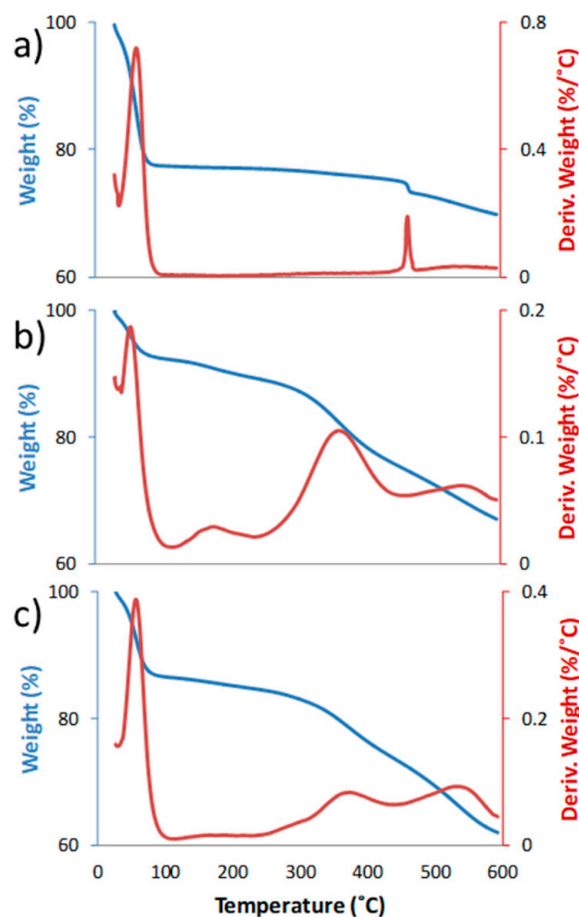


Figure 3. Thermogravimetric analysis (TGA) of PMOBTE (a), *N*-[3-(Trimethoxysilyl)propyl] ethylenediamine (DA)-PMOBTE (b) and SUR@DA-PMOBTE (c).

The drug release was measured from a stirred suspension (1 mg/mL) in PBS buffer (pH 7.4). As can be seen from Figure 4a the release of the drug from the surface was rather slow, which continued at a constant rate for the duration of the experiment (7 days). In these experimental conditions, with the concentration of material in suspension 1 mg/mL, the average rate of drug release from the material is 1.9 µg of SUR per day, per mg of SUR@DA-PMOBTE. The total release amount of SUR after 7 days was 17% of the total amount loaded on the material. The slow drug release also suggests that the drug is adsorbed on the entire surface of the material (both on external surface and on the surface inside the pores). In case of drug adsorption on the external surface the release is typically fast as all adsorbed molecules are easily available for the exchange with the bulk water molecules and PBS ions. In case of the drug adsorbed inside the mesopores, the slow diffusion rate of water going inside and of the drug going outside of the pores significantly lowers the rate of drug release, particularly in case of the material like PMOBTE, which has mostly narrow micropores of irregular, wormlike structure. Previous reports demonstrated that decreasing the pore size lowers the rate of drug release for mesoporous silica nanomaterials [24,25].

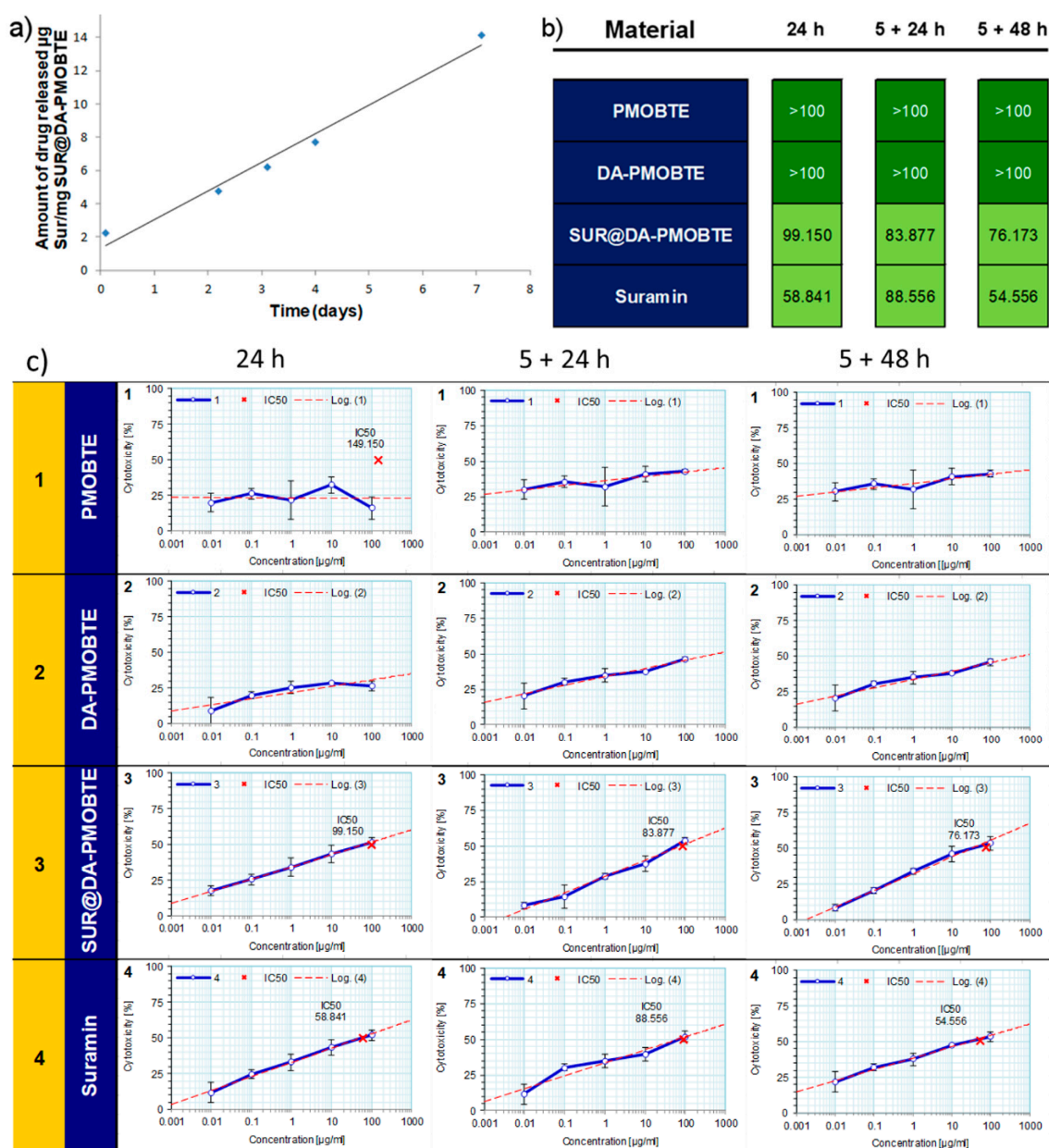


Figure 4. (a) Release kinetics of SUR from SUR@DA-PMOBTE in PBS buffer; (b) the half maximal inhibitory concentration (IC₅₀) values (expressed in µg/mL) for the MRC-5 cells treated with the synthesized materials and solution of SUR. The suspension of the materials or drug solution were in contact with the cells for 24 h, or for 5 h, followed by 24 h or 48 h of growth in the fresh medium, as indicated; (c) Cytotoxicity results for the treatment of MRC-5 cells with prepared materials.

Cell viability experiment on MRC-5 cells (Figure 4b) demonstrated that both materials without SUR (PMOBTE and DA-PMOBTE) showed high biocompatibility with the half maximal inhibitory concentration (IC₅₀) values above 100 µg/mL. SUR loaded materials SUR@DA-PMOBTE showed increased capability to cause cell death and shows IC₅₀ values comparable the free SUR, even though the portion of SUR in SUR@DA-PMOBTE material is only 8.3 wt %. This result showcases that DA-PMOBTE as a drug carrier increases the uptake of SUR by the cells and thus increases the treatment efficacy by the loaded drug. Hence, using DA-PMOBTE as a drug carrier would lower amounts of the drug necessary for the therapeutic activity and hence would lower the probability for adverse effects on healthy tissues.

3. Materials and Methods

3.1. Methodology

Scanning electron microscopy (SEM) images were obtained on the Tescan MIRA3 XMU FESEM (Tescan, Brno, Czech Republic), with the electron energies of 20 kV in a high vacuum. The samples were coated with thin layer of Au using a standard sputtering technique. Nitrogen adsorption desorption measurements were conducted on ASAP 2020 (Micromeritics, Norcross, GA, USA) instrument after degassing samples for 6 hours at 105 °C. The TG/DTA analyses were performed simultaneously (30–800 °C range) on an SDT Q600 TGA/DSC instrument (TA Instruments, New Castle, DE, USA). The heating rates were 20 °C/min using less than 10 mg sample mass. The furnace atmosphere consisted of air at a flow rate of 100 cm³/min. UV/VIS absorbance spectra were recorded on Shimadzu 1800 UV/Vis spectrophotometer (Shimadzu, Kyoto, Japan). FTIR spectra have been obtained by the transmission KBr pellet technique using Thermo-Nicolet Nexus 670 instrument (Thermo Fisher Scientific, Waltham, MA, USA). All reagents were purchased (Sigma-Aldrich, St. Louis, MO, USA) and used as received.

3.2. Synthesis of PMOBTE Nanomaterial

Cetyltrimethylammonium bromide (CTAB, 320 mg) was dissolved in a mixture of ethanol (25.6 mL) and water (28.8 mL). Solution of ammonium hydroxide (25%, 0.4 mL) was added and the mixture was magnetically stirred at room temperature. The organosilica reagent (1,2-bis(triethoxysilyl)ethane, 0.578 mL) was then added dropwise and the reaction mixture continuously stirred for 24 h at 700 rpm. After this time acetone was added to coagulate the nanoparticles and to facilitate the separation by centrifugation. The product was centrifuged at 11,000 rpm, washed twice with water, once with acetone and dried at 80 °C. CTAB was then washed by sonication (twice) in solution of NH₄NO₃ in ethanol (6 g/L) and further washing with ethanol twice and once with acetone.

3.3. Synthesis of DA-PMOBTE Nanomaterial

PMOBTE material (90 mg) was suspended in dry dimethylformamide (DMF, 20 mL), functionalized with *N*-[3-(Trimethoxysilyl)propyl]ethylenediamine (DA, 0.5 mL) by refluxing the mixture at 110 °C over night. The material was then washed twice with water and acetone and dried at 80 °C.

3.4. Synthesis of SUR@DA-PMOBTE Nanomaterial

SUR (SUR, 5.1 mg) was dissolved in 10 mL of ethanol and 1 mL of PBS buffer (pH 7.4) was added. DA-PMOBTE (60 mg) was then added to the solution and stirred for 24 h at room temperature. The material was then isolated by centrifugation at 11,000 rpm and washed twice with ethanol.

The supernatants were collected and evaporated. The residue was dissolved in 20 mL of PBS buffer and the amount of left-over SUR was determined by UV/VIS absorbance at 237 nm, from the previously prepared standard curve. The difference between the starting and residue amounts of SUR gives the amount of loaded SUR, calculated as 83 µg of SUR per mg of SUR@DA-PMOBTE material.

3.5. Drug Release Experiment

For determining the drug release kinetics, the suspension of the SUR@DA-PMOBTE in PBS buffer was prepared (1 mg/mL) and the amount of released SUR was monitored for the period of 7 days. The measurements were conducted on supernatants, after removal of the material by centrifugation at 11,000 rpm from aliquots of the sample solution, by measuring the UV/VIS absorbance at 237 nm and calculating the amount of released drug from the previously prepared standard curve.

3.6. Cell Line Experiments

Experimental procedure was performed using human lung fibroblasts (MRC-5 ATCC CCL 171). Cell lines were cultured in Dulbecco's modified Eagle's medium with 4.5 g/L glucose (DMEM, Sigma-Aldrich, St. Louis, MO, USA), 10% FBS (fetal bovine serum, Sigma) and containing antibiotic/antimycotic solution (Sigma). The cell lines were grown in thermostat at 37 °C with 100% humidity and 5% CO₂ (Heraeus, Hanau, Germany). The cells were passaged twice a week; those used in the experiments were in the logarithmic phase of growth between the third and the tenth passage. The number of cells and their viability were determined using the dye exclusion test with 0.1% trypan blue (DET, data not shown) [26].

3.7. MTT Assay

Growth inhibition was evaluated by tetrazolium colorimetric MTT assay (Sigma-Aldrich, St. Louis, MO, USA). The assay is based on the cleavage of the tetrazolium salt 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT), to formazan by mitochondrial dehydrogenases in viable cells [27]. Exponentially growing cells were harvested, counted by trypan blue and plated into 96-well microtiter plates (Costar) at optimal seeding density of 5×10^3 cells per well to assure logarithmic growth rate throughout the assay period. Viable cells were plated in a volume of 90 µL per well, and preincubated only in DMEM at 37 °C for 24 h to allow cell stabilization prior to the addition of substances. Tested substances dissolved in DMEM medium were added to all wells (10 µL/well) except to the control ones and microplates were incubated for 5 and 24 h. After 5 h the suspensions were removed, the cells were washed with PBS buffer and fresh aliquots of DMEM medium were added and incubated for 24 and 48 h. Three hours before the end of incubation period 10 µL of MTT solution was added to all wells. MTT was dissolved in medium at 5 mg/mL and filtered to sterilize and remove a small amount of insoluble residue present in some batches of MTT. Acid-isopropanol (100 µL of 0.04 N HCl in isopropanol) solution was added to all wells and mixed thoroughly to dissolve the dark blue crystals. After a few minutes at room temperature to ensure that all crystals were dissolved, the plates were read on a spectrophotometer plate reader (Multiscan MCC340, Labsystems Diagnostics Oy, Vantaa, Finland at 540/690 nm. The wells without cells containing complete medium and MTT only acted as blank. Inhibition of growth was expressed as a percent of a control and cytotoxicity was calculated according to the formula: $(1 - OD_{\text{test}}/OD_{\text{control}}) \times 100$. The substance potency was expressed as the IC₅₀ (50% inhibitory concentration).

3.8. Data Analysis

Two independent experiments were set out with quadruplicate wells for each concentration of the compound. IC₅₀ value defines the dose of compound that inhibits cell growth by 50%. The IC₅₀ of compounds was determined by Median effect analysis.

4. Conclusions

We successfully synthesized periodic mesoporous organosilica nanoparticles with spherical morphology, mean diameter of 240 nm and high surface area (778 m²/g). The material's surface was functionalized with propyl-ethylenediamine groups and SUR was successfully adsorbed to the surface in the amount of 8.3 wt %. The rate of release of SUR in PBS was low, with only 17% of the loaded drug released after 7 days. However, in vitro experiments on MRC-5 cell line demonstrated high toxicity of the drug loaded material and low toxicity of the materials without the adsorbed drug. These results may open the more efficient treatment possibilities with SUR nanocarriers, with lower adverse effects on healthy tissues.

Author Contributions: Conceptualization, N.Ž.K.; methodology, N.Ž.K. and V.K.; investigation, S.D. and V.K.; writing—original draft preparation, N.Ž.K. and V.K.; writing—review and editing, N.Ž.K.; supervision, N.Ž.K. and D.J.; project administration, D.J.; funding acquisition, D.J.

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Conflicts of Interest: The authors declare no conflict of interest.

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