



NAFlex: A web server for the study of Nucleic Acids Flexibility

BioExcel Webinar, 20/07/2017

Federica Battistini, IRB-Barcelona



Nucleic Acids SEQUENCE

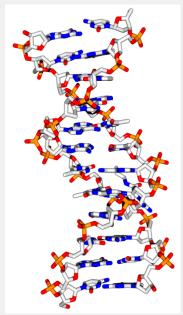




Nucleic Acids SEQUENCE



STRUCTURE



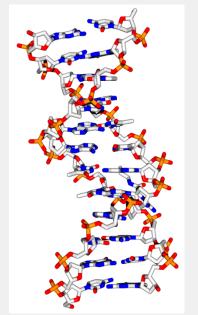
Structure depends on the sequence, not ideal B-DNA Each base pair characterized by flexibility and geometry



NA SEQUENCE

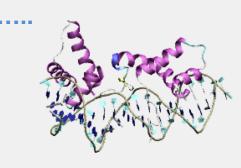


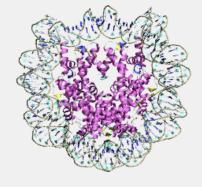
STRUCTURE



FUNCTION

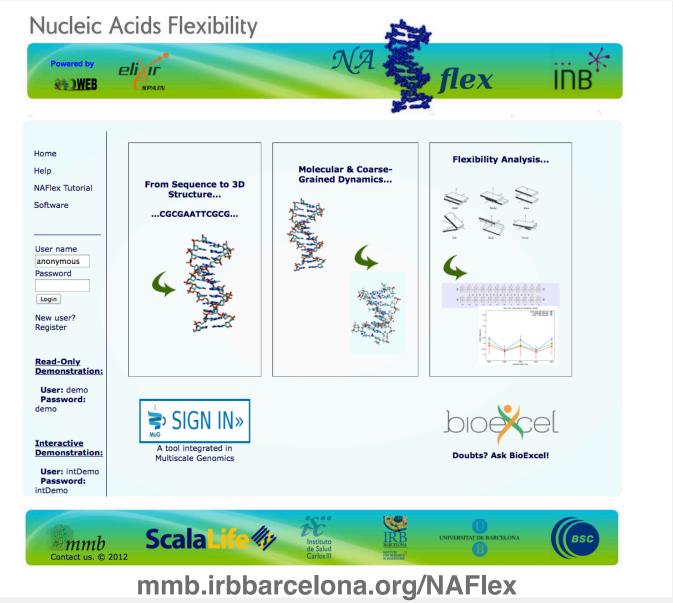
Protein Recognition
Protein-DNA binding
Genome organization
Expression control





Structure depends on the sequence, not ideal B-DNA Each base pair characterized by flexibility and geometry







Home General Help Getting started Structure checking Ligand checking Workspace Icons Software Related Links References Input From Sequence From Structure From Trajectory From Saved Project Simulation **Engines** Atomistic MD Simulations Coarse-Grained Simulations **Analysis Tools** Standard Cartesian Analysis **Nucleic Acids** Flexibility **Tutorials** Atomistic MD Setup tutorial **NAFlex Analysis** tutorial Coarse-Grained

Analysis tutorial

Nucleic Acids Flexibility

NA

flex

Input

From Sequence

From Structure

From Trajectory

From Saved Project



NAFlex Input Options

NAFlex offers four main entry options:

- Simulation (Single Structure):
 - Starts from a structure (PDB format), setup and run a MD simulation.
- Analysis (MD Trajectory):

Starts from a trajectory (usual MD trajectory formats), analyse, get information or convert between MD trajectory formats.

• DNA/RNA Simulation from Sequence:

Starts from a sequence, obtain a 3D structure from a nucleic acid sequence.

Upload past NAFlex Project:

Upload a previously stored NAFlex project.

NAFlex Project Entrance Page					
Project Title					
Description (optional)					
Input Type	Simulation (Single structure)				
Next Cancel	Analysis (MD Trajectory) Upload past NAFlex project DNA/RNA Simulation From Sequence				





NAFlex Simulation Engines

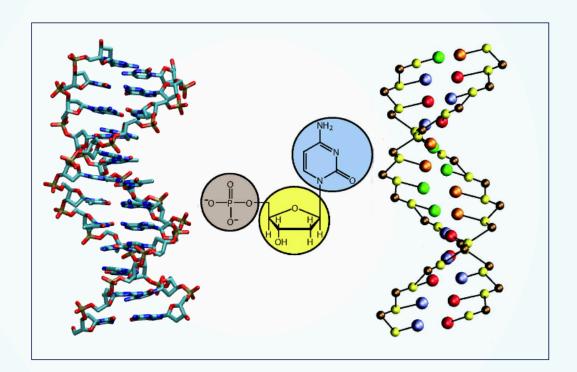
NAFlex offers a variety of simulation tools obtain dynamic information of nucleic acids. Basically the server has two main kind of **Simulation Engines**:

• Atomistic Molecular Dynamics Simulations:

Molecular Dynamics Simulations with fully atomistic representation, in which all atoms (usually including the solvent's) are considered explicitly.

• Coarse-Grained Simulations:

Coarse-Grained Simulations, where collections of atoms (number of them depending on the desired resolution) are represented by a few spherical beads connected by springs. **NAFlex** implements a couple of Coarse-grained algorithms, one at Nucleotide-Base Level (*Mesoscopic Elastic Model*) and another one at M x Base-Pair Level (*Worm-Like Chain Model*).

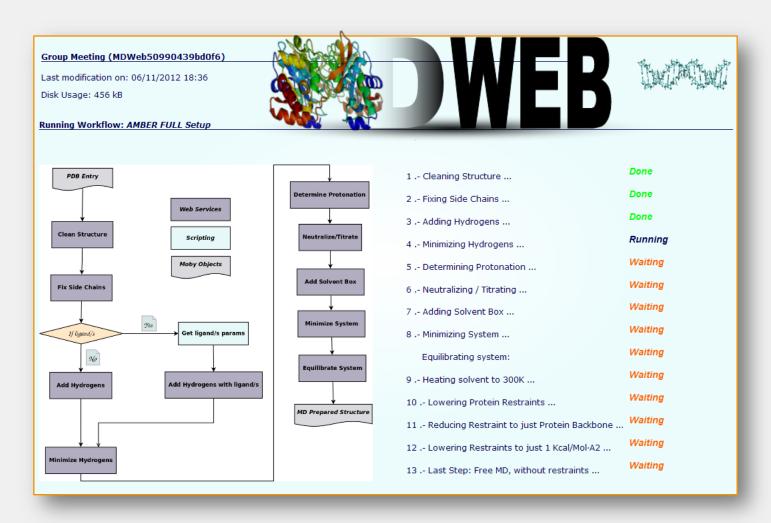




Atomistic Molecular vs Coarse-Grained Representation (J.Chem.Phys. 126, 2007)



Nucleic Acids MD (by MDWeb)



Hospital et al, Bioinformatics 2012, 28(9): 1278-1279



NAFlex Simulation Engines

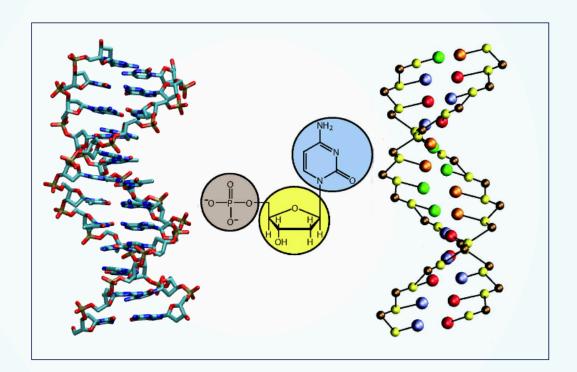
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Atomistic Molecular vs Coarse-Grained Representation (J.Chem.Phys. 126, 2007)

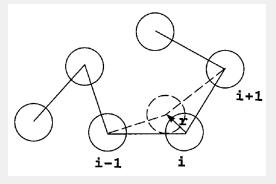


NAFlex Coarse-Grained Dynamics



A Metropolis Montecarlo algorithm associated to DNA helical parameters is used to obtain coarse-grained DNA dynamics.

Goñi et al, Bioinformatics 2008, 24: 1731-1732



Worm-Like Chain (WLC), Montecarlo Algorithm.

1 bead for each 4 base-pair steps.

Debye Hückel Equation to take into account implicit solvent and ionic concentration (Salt-dependent).

Uniform charge of DNA.

DNA stretching, bending and torsion.



Jian et al, JCPhysics 1997, 136: 168-179



Structure checking

Ligand checking

Workspace

Icons

Software

Related Links

References

Input

From Sequence

From Structure

From Trajectory

From Saved Project

Simulation Engines

Atomistic MD Simulations

Coarse-Grained Simulations

Analysis Tools

Standard Cartesian Analysis

Nucleic Acids Flexibility

Tutorials

Atomistic MD Setup tutorial

NAFlex Analysis tutorial



NA SEQUENCE-DEPENDENT PHYSICAL PROPERTIES



NAFlex Analysis Tools

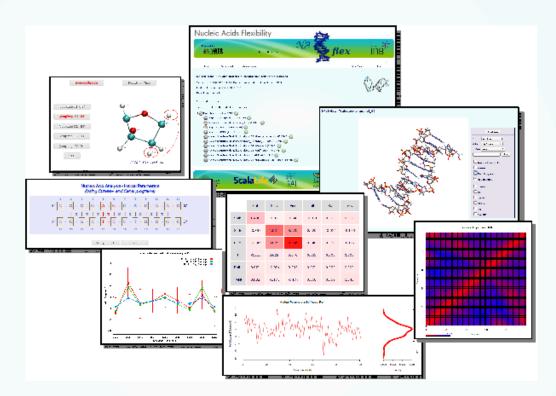
NAFlex offers a variety of analysis packages for mining nucleic acids trajectories. They can be grouped in two main families:

• Standard Cartesian Analysis:

Basic trajectory cartesian analysis such as Root Mean Square Deviation (RMSd), RMSd per Nucleotide, Radius of Gyration (RGyr), Bfactors, etc.

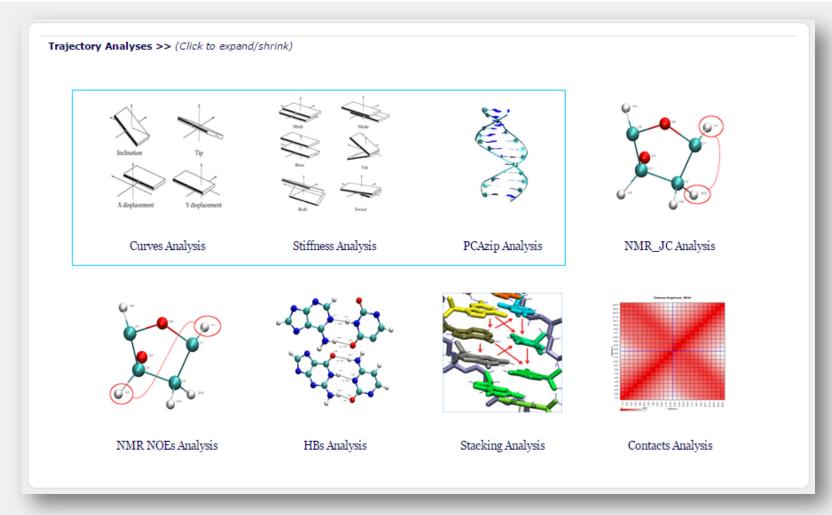
• Nucleic Acids Flexibility:

Set of different Flexibility analysis of Nucleic Acids: Helical Parameters, Principal Components (PCA), HB/Stacking energies, NMR-observables, etc.



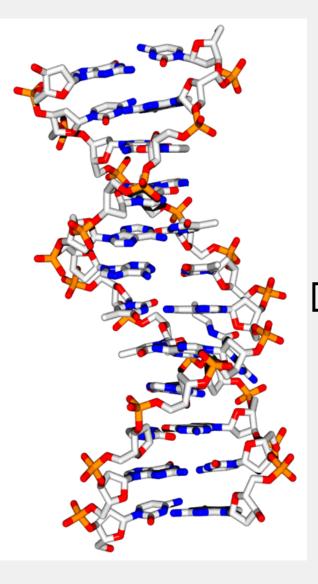










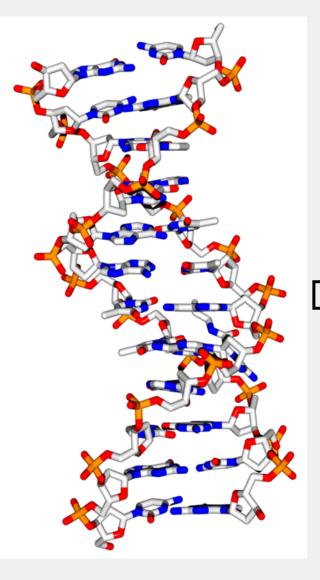


- Helical Parameters
- Stiffness Constants
- Principal Component (PCA)
- Nuclear Magnetic Resonance
 Observables (J-Couplings, NOE's)



- Canonical Hydrogen Bond (HB) Analysis
- Atom Pairs Distances
- Backbone Analysis
- HB/Stacking Energies
- Distance Contact maps



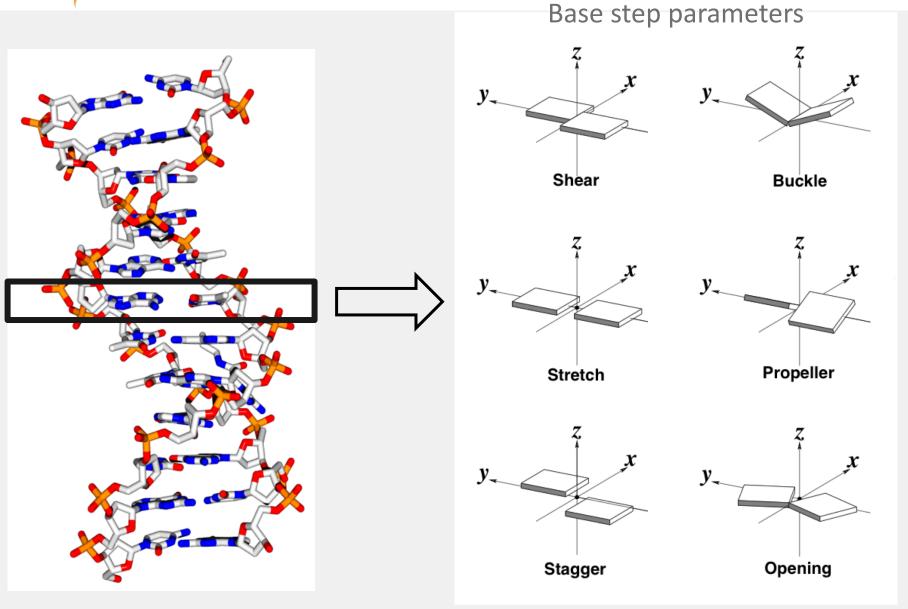


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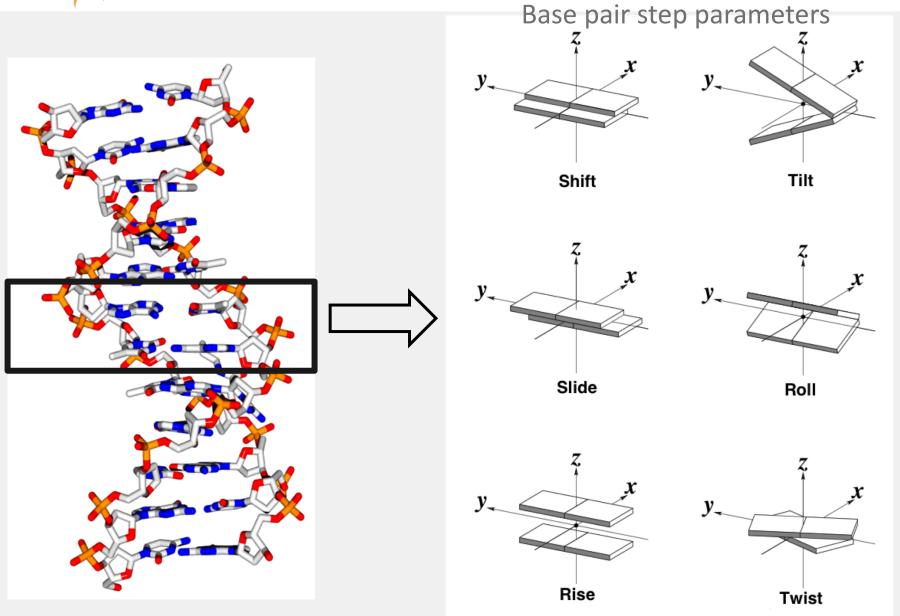


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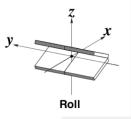






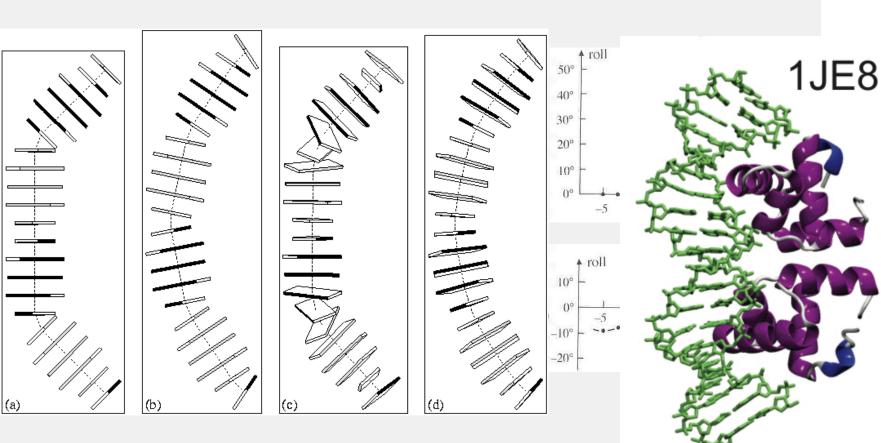


Roll angle and Bending



step

number

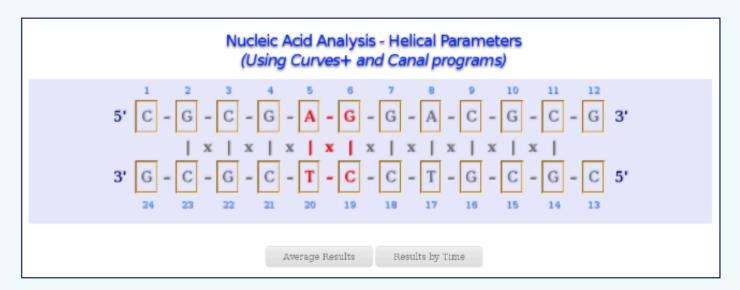


Calladine & Drew



Helical Parameters

When analysing **Nucleic Acid Helical Parameters**, a graphical representation of the two sequence strands will appear on the screen. This representation allows a user-friendly selection of **nucleotides**, **base pairs** and **base pair steps** (tetramers), just clicking at the corresponding region of the sequence. For example, to select a base pair, user may click at the | symbol between nucleotides (from different strands) involved in the interesting base-pair, to select a base pair step, at the **x** symbol between nucleotides involved in the interesting tetramer, and just over a nucleotide one-letter code to select the corresponding nucleotide.

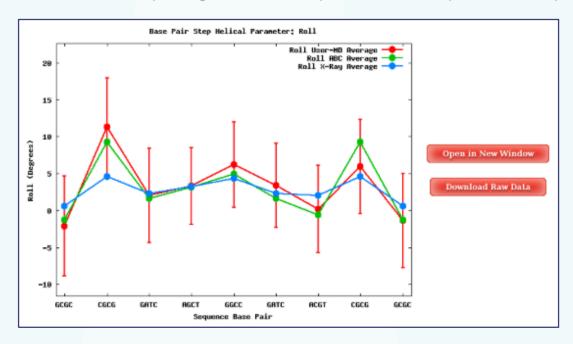






The **Average Results** section contains a set of plots generated after computing average values for all the trajectory snapshots. Plots and their associated raw data can be downloaded form the link provided.

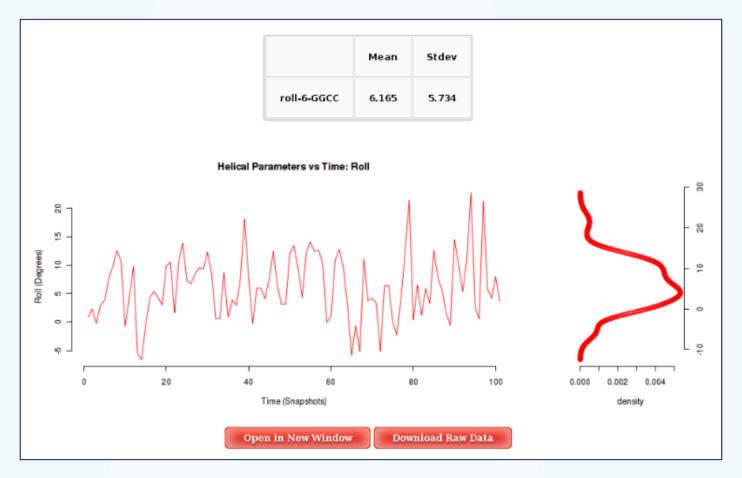
When available, *published data* corresponding to the selected parameter set are plotted for comparison.







Plots generated contain the corresponding **histogram** attached, and an associated table with calculated **mean** and **standard deviation** values.





Base pair step Stiffness Constants

$$(x-x_o)$$

Hooke's Law:
$$\Delta E = \sum k(x - x_o)^2$$

$$\Theta = E(\Delta X)^{-2} = k_B T \subset^{-1} = \begin{pmatrix} k_w & k_{wr} & k_{wt} & k_{ws} & k_{wl} & k_{wf} \\ k_{wr} & k_r & k_{rt} & k_{rs} & k_{rl} & k_{rf} \\ k_{wt} & k_{rt} & k_t & k_{st} & k_{tl} & k_{tf} \\ k_{ws} & k_{rs} & k_{st} & k_s & k_{ls} & k_{lf} \\ k_{wl} & k_{rl} & k_{tl} & k_{ls} & k_{l} & k_{lf} \\ k_{wf} & k_{rf} & k_{tf} & k_{lf} & k_{lf} & k_{f} \end{pmatrix}$$



Stiffness Constants

When analysing **Nucleic Acid Stiffness Constants**, the resulting web page will show again the graphical representation of the sequence strands (see previous section).

The **Stiffness Constants** analysis is divided in two main sections:

- Average Results
- Results by Time

The **Average Results** section contains a set of **Stiffness Matrices** (determined by the inversion of the covariance matrix in helical space) for each sequence **Base Pair Step**.

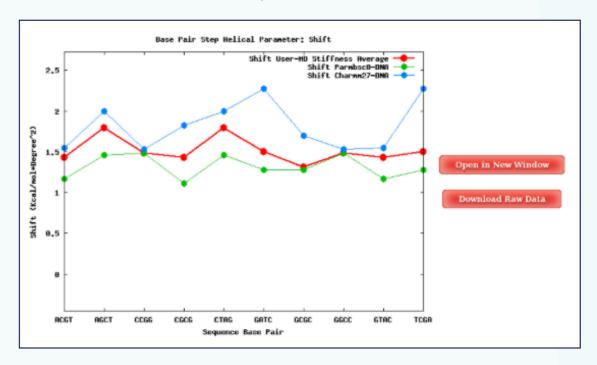
	Shift	Slide	Rise	Tilt	Roll	Twist
Shift	1.430	-0.396	-0.045	-0.011	0.021	0.082
Slide	-0.397	4.931	3.029	0.029	-0.024	-0.176
Rise	-0.045	3.029	8.066	0.040	0.032	-0.177
Tilt	-0.011	0.029	0.040	0.033	0.001	0.006
Roll	0.021	-0.024	0.032	0.001	0.020	0.009
Twist	0.082	-0.176	-0.177	0.006	0.009	0.041
alues in k	cal/(mol*Ų)				
Download Raw Data						





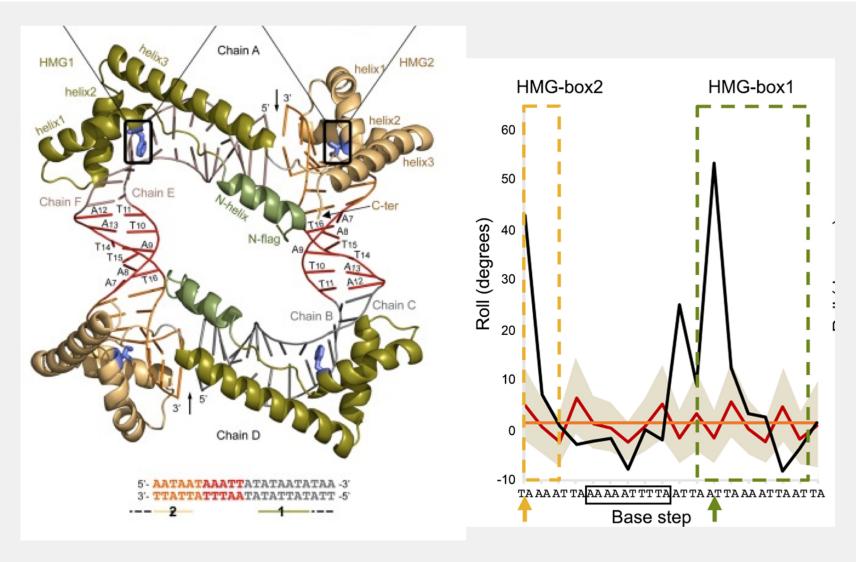
A set of average plots generated computing average values for all the **Stiffness Constants** associated to a **Base Pair Step** (average of the diagonal elements of the **Stifness Matrices**) are also offered.

When available, published data corresponding to the selected parameter are plotted for comparison. Plots and its associated raw data can be downloaded from the link provided.



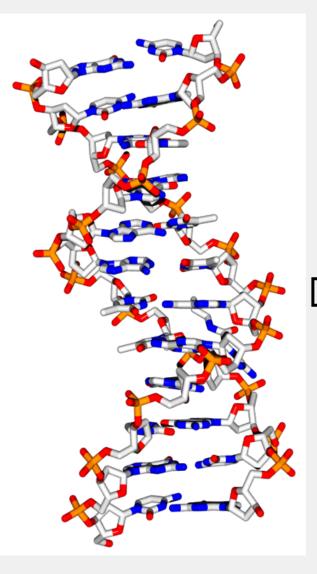






DNA structure directs positioning of the mitochondrial genome packaging protein Abf2p, Chakraborty, NAR (2017)





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- Principal Component (PCA)
- Nuclear Magnetic Resonance
 Observables (J-Couplings, NOE's)

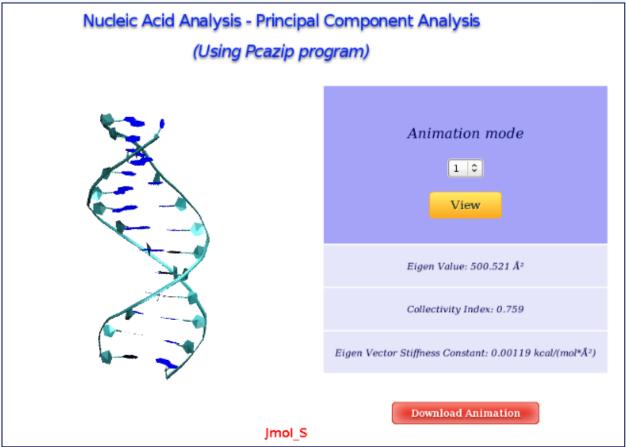


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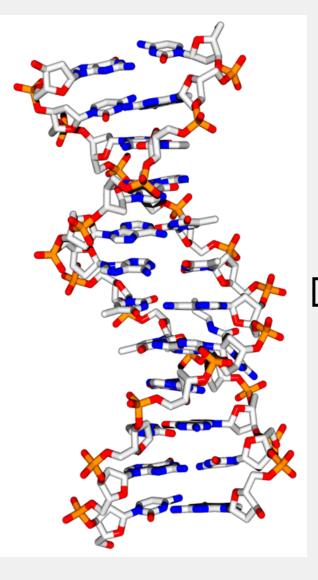
Principal Component Analysis

The **Principal Component Analysis** graphical interface offers the possibility of studying the real movements of the structure through the projections of the trajectory onto the different essential modes. An interactive **JMol applet** shows these movements, allowing user to translate, rotate and in general manipulate the visualization. The first 10 animation modes are offered for visualization and download. Associated values as **eigenvalues**, **collectivity indexes** and **eigenvector stiffness constants** are also shown.









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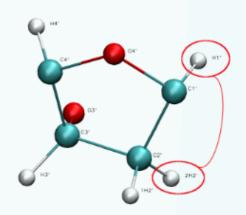


NAFlex Nucleic Acids Flexibility Analysis: Nuclear Magnetic Resonance Observables

J-Couplings

Vicinal ¹H-¹H Coupling Constants (³J-Couplings) are scalar couplings between protons located three bonds away (H-C-C-H).

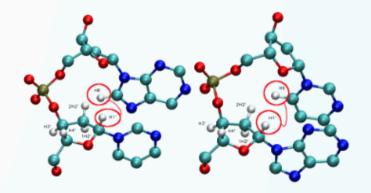
Vicinal J-Couplings are empirically correlated with the dihedral angle (H-C-C-H) through the so-called **Karplus equation**.



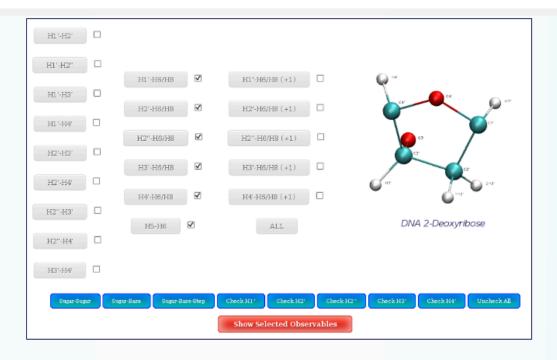
NOEs

Nuclear Overhauser Effect is the transfer of magnetization from one nuclear spin to another via cross-relaxation. The intensity of NOE cross-peaks between two particular protons depends on their relative distance (Inoe = $1/d^6$).

Proton-proton distances derived from NOEs are the most useful NMR parameters for structure elucidation.







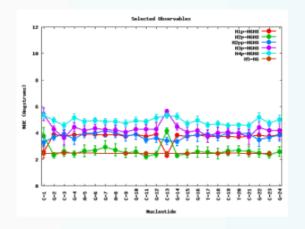
A set of buttons to automatically check interesting proton pairs are offered:

By proton pair position:

- Sugar-Sugar: Intra-Sugar proton pairs.
- Sugar-Base: Sugar-Base proton pairs.
- Sugar-Base Step: Sugar-Base Step proton pairs.

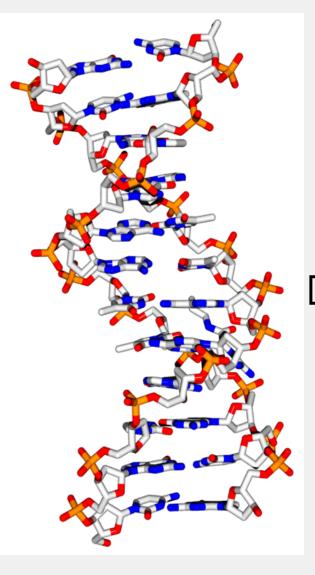
By specific proton:

- H1': All proton pairs involving H1'
- H2' All proton pairs involving H2'
- H3' All proton pairs involving H3'
- . H4' All proton pairs involving H4'
- H2" All proton pairs involving H2" (Only in DNA case)









- Helical Parameters
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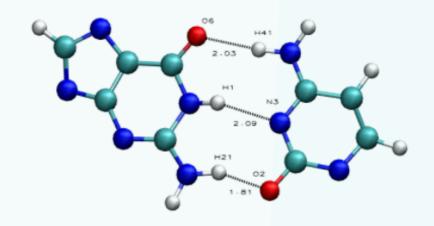
Canonical Hydrogen Bond Analysis

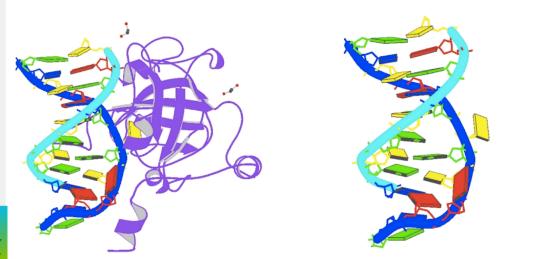
Purine-Pyrimidine HBs:

- Adenine N1 Thymine/Uracil H3
- Adenine H61 Thymine/Uracil O4

Pyrimidine-Purine HBs:

- Guanine O6 Cytosine H41
- Guanine H1 Cytosine N3
- Guanine H21 Cytosine O2







PDB ID 2ZO1

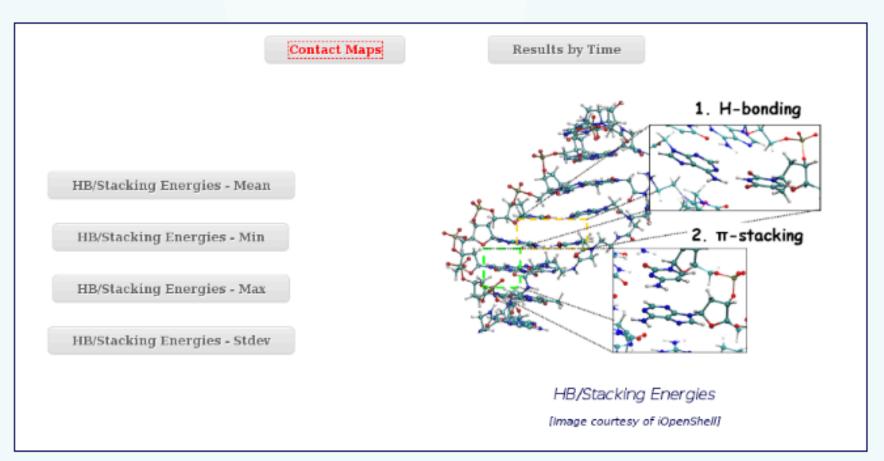


flex

Stored structures							
Click on structure title to deploy the toolbox.							
⊟ 🙉 Base trajectory (38.8 MB) 🕢							
☐ @Dry Trajectory_01 (4.9 MB)							
Select the desired operation.							
Title: _12 Comment:							
Nucleic Structure Flexibility Analysis ?							
Analysis: Atom Pairs Distances							
Above Daile (Davids or Newsborn Code, accounts 20015)							
Atom Pair (Residue Number@Atom Code, example: 2@H5)							
Atom Pair 1: Atom 1: Add another atom pair							
4							
Atom Pair 2: Atom 1: Atom 2: Remove							
ok cancel							
OK CHICE!							



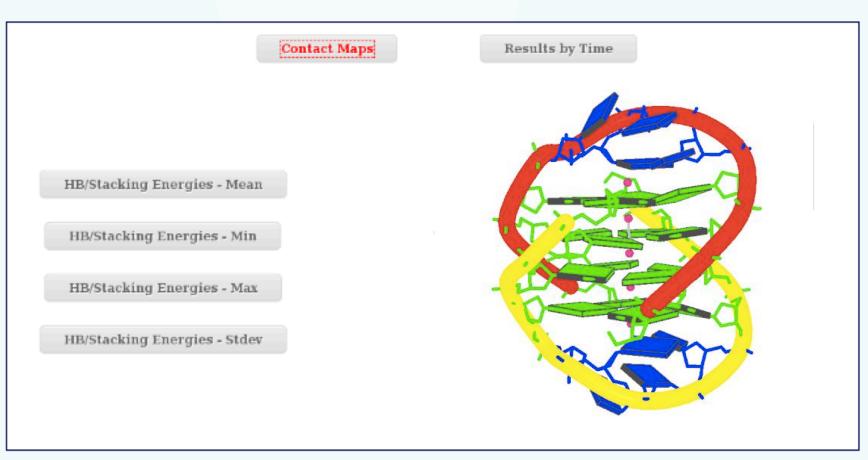
HB/Stacking Energies







HB/Stacking Energies







Backbone Torsions

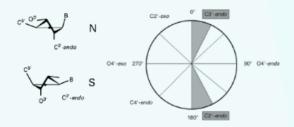
The three major elements of flexibility in the backbone are:

Sugar Puckering

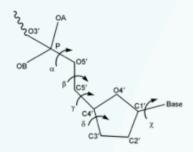
Sugar Puckering annotation is done by dividing the pseudorotational circle in four equivalent sections:

North: 315:45°
East: 45:135°
South: 135:225°
West: 225:315°

These four conformations are those dominating sugar conformational space, in agreement with all available experimental data.



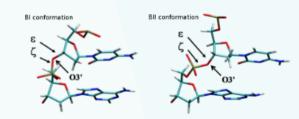
Canonical Alpha-Gamma



Rotations around α/γ torsions generate non-canonical local conformations leading to a reduced twist and they have been reported as being important in the formation of several protein-DNA complexes.

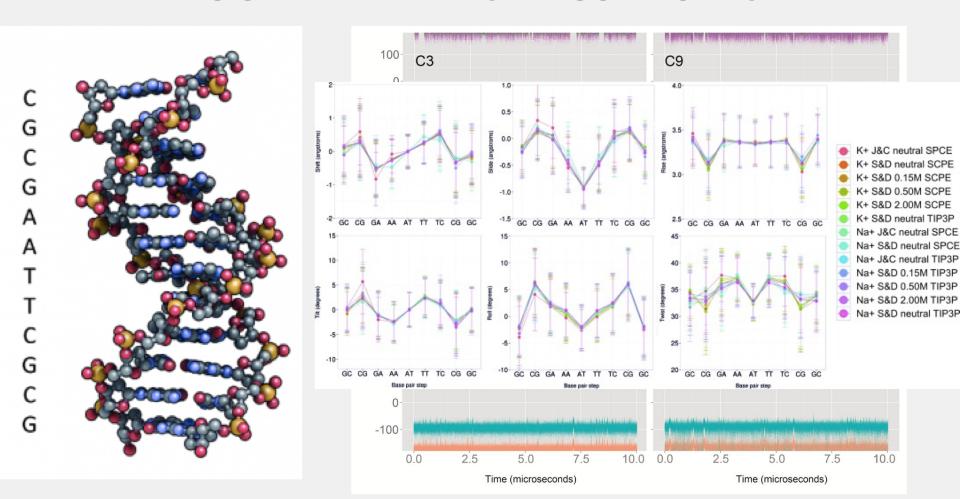
BI-BII Population

The concerted rotation around ζ/ϵ torsions generates two major conformers: **BI and BII**, which are experimentally known to co-exist in a ratio around **80%:20% (BI:BII)** in B-DNA.





ANALYSIS DNA DREW DICKERSON DODECAMER

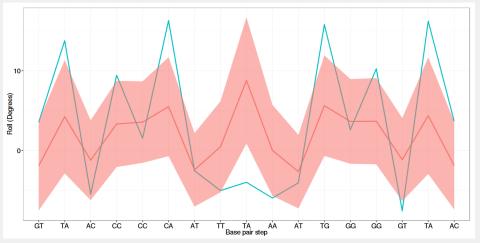


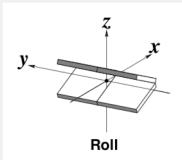
Long-timescale dynamics of the Drew Dickerson dodecamer, DANS PD, NAR (2016).

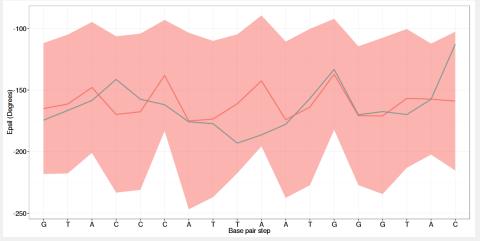


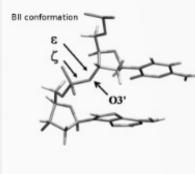
ANALYSIS DNA: PROTEIN-BOUND VS UNBOUND













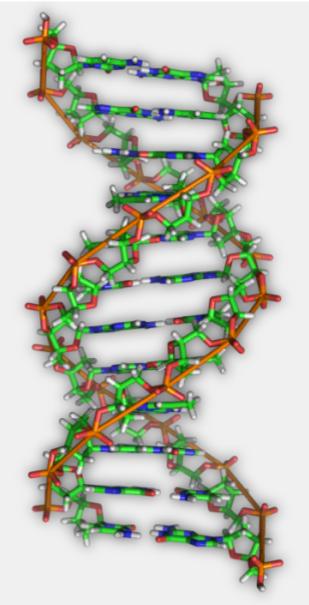
Software

NAFlex is powered by an Apache 2. web server with PHP 5. and MySQL 5.0.51. Calculations are redirected to a 8 core Intel(R) Xeon(R) CPU @ 2.67GHz - 16GB RAM cluster managed by a Sun Grid Engine batch manager.

List of external software used in NAFlex operations:

Program	Description	Package & Version
Curves+, Canal	Analysis of Nucleic Acids Structure and Flexibility	Curves+ 2.0
Open Babel	Chemical toolbox designed to work with many languages of chemical data.	Open Babel 2.1.1
R Statistics	Statistical Computing and Graphics software.	R 2.15.0
BLAST	Basic Local Alignment Search Tool	BLAST 2.2.17
CMIP	Classical Molecular Interaction Potential	CMIP 2.5.4
Gnuplot	Plotting tool	Gnuplot 4.2 patchlevel 2
Grace	Plotting tool	Grace 5.1.21
GROMACS	Molecular Dynamics Simulator	GROMACS 4.5.5
JMol	Molecular Graphics Viewer	JMol 10.00.46
MobyLite PerlAPI	BioMoby Perl API	MobyLite PerlAPI 1.0
NAMD	Molecular Dynamics Simulator	NAMD 2.8
PCAsuite	Trajectory compression tool	PCASuite 1.1
PropKa	Prediction of protein pKa values	Propka 2.0
Ptraj, cpptraj	Structure and dynamic analysis of trajectories	Ambertools 14
tgatoppm, pnmcrop, pnmtopng	Image management	Netpbm 10.0
Tleap	MD preparation program	Ambertools 14
VMD	Molecular Graphics Viewer	VMD 1.8.5





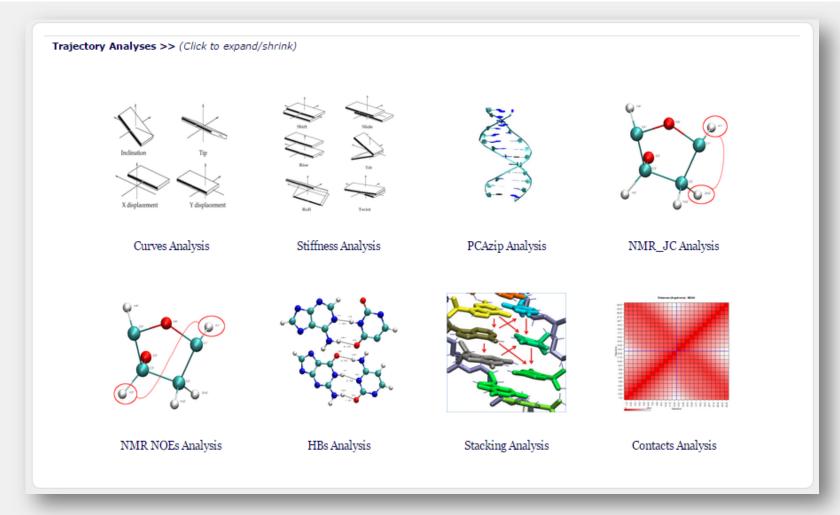


BIGNASim

A BigData approach to efficiently manage large nucleic acids simulation data











Multiscale complex Genomics







WP2

Outreach, training, exploitation

Community engagement, dissemination of results, training of VRE users, longterm sustainaibility and exploitation.



Multi-scale genome browser

Allowing for the first time to connect 1D to 3D/4D genomic data and to navigate across all the resolutions of DNA in the cell through a graphical user interface.

WP5

Computational infrastructure supporting the VRE services

Compatible with Cloud and HPC. Interoperable with Data Infrastructure. Integrates security and User Support.















http://www.multiscalegenomics.eu/

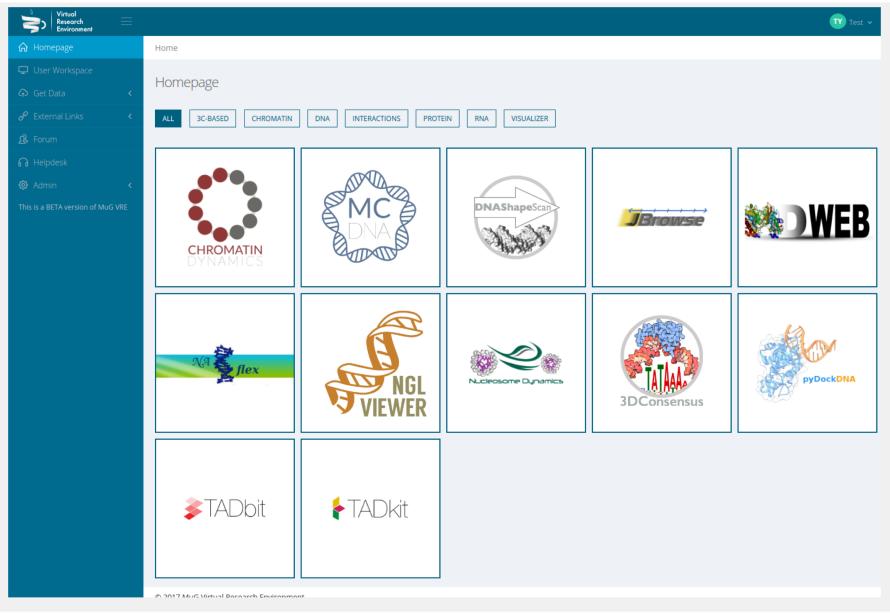


MuG community - interest group

Environment

3D/4D Genomics







Acknowledgements



Prof. Modesto Orozco Prof. Josep Lluís Gelpí Adam Hospital





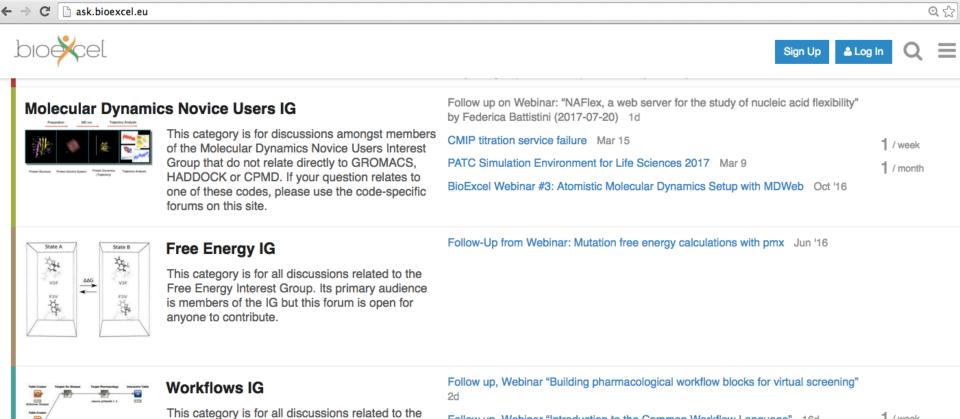








Bioexcel forum (ask.bioexcel.eu)



1 / week

2 / month

Globus" Nov 16

Workflows Interest Group. Its primary audience is members of the IG but this forum is open for

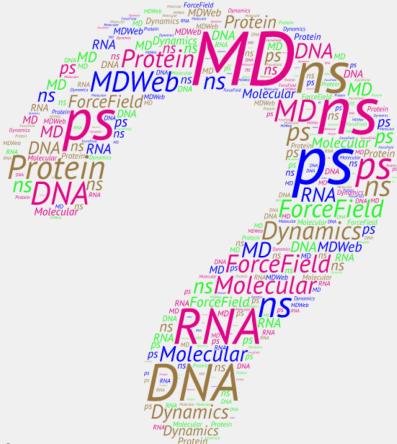
anyone to contribute.

Follow up, Webinar "Introduction to the Common Workflow Language" 16d

Musings on GROMACS and workflow description integration Nov '16

Follow up on "Large-scale analytical workflows on the cloud using Galaxy and





federica.battistini@irbbarcelona.org Ask.bioexcel.eu

