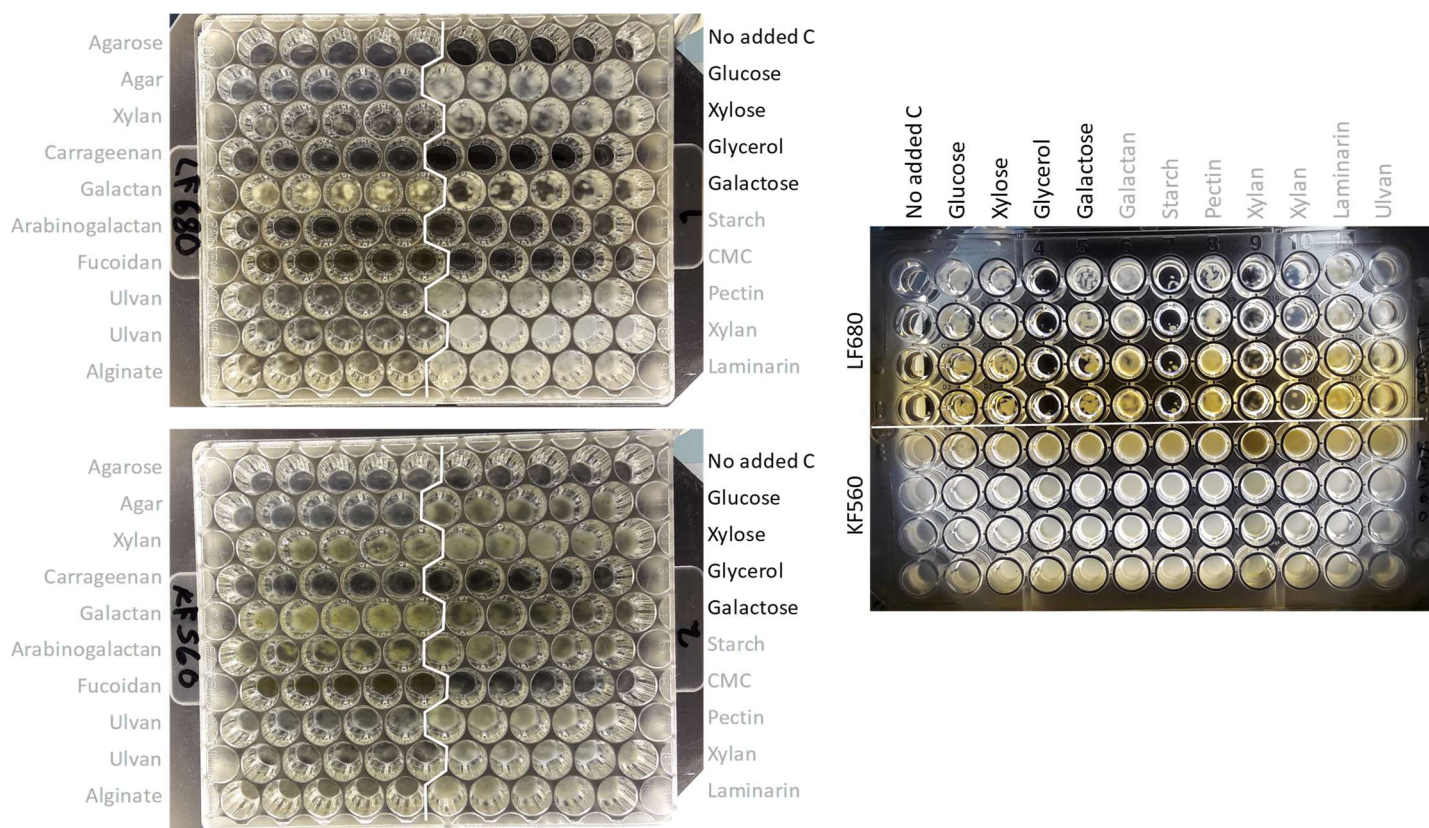


Data Figure 1. Growth of *P. chrysogenum* KF657 (upper) and *D. salina* LF304 (lower) in 100-well Honeycomb plates (right) and a 96-well microtiter plate after ca. 7 days incubation. The defined medium was based on that described by Verduyn et al. (1992), modified by adding 28 g L⁻¹ Tropic Marin® salt and buffered with PIPPS buffer to an initial pH of 4.25, and replacement of glucose with 2 g L⁻¹ of each carbohydrate indicated in the figure. In the 100-well Honeycomb plates, one strain was inoculated per plate, with medium containing each carbohydrate in 5 neighbouring wells. In the 96-well plates, 2 strains were inoculated per plate, with 8 neighbouring wells containing the same carbohydrate, 4 wells inoculated with each strain. Note that *D. salina* LF304 produced pigmented hyphae which are not readily visible against a dark background (smaller photo shows 96-well plate photographed against white background). Data for substrates indicated in black are included in data files Tamminen_et_al_2020_BioScreen_growth.xlsx and Tamminen_et_al_2020_Cytomat_growth.xlsx and are discussed in the associated journal article. Note also that stripes of yellow colouring in the photographs are derived from the photographic conditions, not mycelial colour.



Data Figure 2. Growth of *A. cruciatus* LF680 (upper) and *H. varia* KF560 (lower) in 100-well Honeycomb plates (right) and a 96-well microtiter plate after *ca.* 8 days incubation. The defined medium was based on that described by Verduyn et al. (1992), modified by adding 28 g L⁻¹ Tropic Marin® salt and buffered with PIPPS buffer to an initial pH of 4.25, and replacement of glucose with 2 g L⁻¹ of each carbohydrate indicated in the figure. In the 100-well Honeycomb plates, one strain was inoculated per plate, with medium containing each carbohydrate in 5 neighbouring wells. In the 96-well plates, 2 strains were inoculated per plate, with 8 neighbouring wells containing the same carbohydrate, 4 wells inoculated with each strain. Data for substrates indicated in black are included in data files Tamminen_et_al_2020_BioScreen_growth.xlsx and Tamminen_et_al_2020_Cytomat_growth.xlsx and are discussed in the associated journal article. Note also that stripes of yellow colouring which cross the whole photograph are derived from the photographic conditions, not mycelial colour.

Verduyn C, Postma E, Scheffers WA, van Dijken JP. Effect of benzoic acid on metabolic fluxes in yeasts: a continuous-culture study on the regulation of respiration and alcoholic fermentation. *Yeast*. 1992;8: 501–517.



Data Figure 3. Enlargement of wells containing *P. chrysogenum* KF657 in a 100-well Honeycomb plate (left), showing a tendency to grow near the walls of the well leaving a clearer area in the middle of the well, and *A. cruciatus* LF680 (upper) and *H. varia* KF560 (lower) in a 96-well microtiter plate (right), showing the tendency of *A. cruciatus* LF680 to form pellets in the centre of the well, while *H. varia* KF560 produced dispersed growth. (See also above.)



Data Figure 4. Example of condensation on the lid of a 96-well microtiter plate.