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A new species, new geographical records, and taxonomic notes in Oreodera Audinet-Serville, 1835 (Coleoptera: Cerambycidae: Lamiinae)

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A new species, new geographical records, and taxonomic notes in *Oreodera* Audinet-Serville, 1835 (Coleoptera: Cerambycidae: Lamiinae)

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Abstract. The Bolivian *Oreodera pergeri* Wappes and Santos-Silva (Coleoptera: Cerambycidae: Lamiinae) is described as new. *Oreodera howdeni* Monné and Fragoso, 1988 is newly recorded for the state of Chiapas in Mexico and for Belize, and the main differences between the sexes are reported. Other new records reported are: *Oreodera boucheri* Néouze and Tavakilian, 2010 for Peru; *O. vulgata* Monné and Fragoso, 1988 for Paraguarí Department in Paraguay; and *O. stictica* Monné and Fragoso, 1988 for Bolivia (Santa Cruz). *Oreodera sororcula* Martins and Monné, 1993 is illustrated, and notes on color, morphological variations, and correction in the published sex of a paratype provided.

Key words. Acanthoderini, Neotropical region, taxonomy.

Introduction

Oreodera Audinet-Serville, 1835 was described for O. glauca (Linnaeus, 1758), O. cinerea Audinet-Serville, 1835, and O. pubicornis Audinet-Serville, 1835. Subsequently, Thomson (1864), designated Cerambyx glaucus, now Oreodera glauca glauca, as the type species. The genus is the largest in the tribe Acanthoderini with 116 species and 2 subspecies (Monné 2018). Oreodera is also one of the most widely distributed Acanthoderini genera in the Americas being found from Mexico to southern South America (including West Indies). Species are geographically distributed (Monné 2018) as follows: four plus one subspecies are found only in Mexico; four found in Mexico and Central America; one found from Mexico through Central America, the West Indies and throughout Brazil; 12 are found only in Central America; eight found in Central America and South America; 86 only in South America; one only in West Indies; and one subspecies restricted to Jamaica.

Primary contributors as new species authors include Monné and Fragoso (20 species, plus one synonym), Bates (16 species, plus four synonyms), Néouze and Tavakilian (or vice versa, 17 species) and Martins et al. (19 species, nine (plus one synonym) of those only with Galileo). Additions to the genus continue to happen regularly with some 63 species (54 per cent of the total) being added in the last 40 years and 35 species (30 per cent of the total) so far in the 21st century.

Currently, diagnostics for nearly all genera of Acanthoderini are problematic, with the limits among them unclear or ill-defined. However, *Oreodera* can usually be defined as having the following combination of features: Frons transverse; genae short, about half of length of lower eye lobe; antennae longer than body, more so in male; antennomere III distinctly longer than scape; prothorax transverse, with distinct lateral tubercles; pronotum tuberculate; prosternal and mesoventral processes wide, the latter distinctly wider than mesocoxal cavities; and femora pedunculate-clavate.

Materials and Methods

Photographs were taken in the MZSP with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65mm f/2.8 1–5× macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken in "mm" using a measuring ocular Hensoldt/Wetzlar - Mess 10 in the Leica MZ6 stereomicroscope, also used in the study of the specimens. All specimens examined were adults.

The acronyms used in the text are as follows:

ACMT American Coleoptera Museum (James Wappes), San Antonio, Texas, USA

AMNH American Museum of Natural History, New York, New York, USA

FSCA Florida State Collection of Arthropods, Gainesville, Florida, USA

FWSC Fred W. Skillman collection, Pearce, Arizona, USA

MNRJ Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil

MZSP Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil

RFMC Roy F. Morris Collection, Lakeland, Florida, USA

Taxonomy

Oreodera pergeri Wappes and Santos-Silva, new species (Fig. 1–5)

Description. Male (Fig. 1–4). Integument mostly dark-brown, almost black in some areas; mouthparts dark reddish-brown, except yellowish-brown areas on palpomeres; anteclypeus and labrum fulvous; antennomeres dark reddish-brown except distal area dark-brown (lighter on III–IV); central area of gulamentum dark-reddish brown; central area of ventral surface of thorax dark reddish-brown; pro- and mesocoxae dark reddish-brown.

Head. Finely, densely punctate; with yellowish-white pubescence partially obscuring integument, whiter close to clypeus centrally; with a few long, erect brown setae laterally. Vertex and area behind eyes with dense yellowish-brown pubescence obscuring integument, and a few long, erect brown setae close to eyes, except: narrow area along median groove glabrous; central area between upper eye lobes glabrous; wide, triangular central area close to prothorax with white pubescence partially obscuring integument; area behind lower eye lobes with longitudinal glabrous area on center of pubescent area and glabrous close to prothorax. Genae with dense yellowish-brown pubescence close to eyes, with a few long, erect brown setae interspersed, most of distal half glabrous. Antennal tubercles with sculpturing as on frons except narrow apex smooth; with yellowish-brown pubescence nearly obscuring integument, with pubescence more yellowish-white frontally. Median groove distinct from clypeus to prothoracic margin. Most of gulamentum nearly smooth, with slightly distinct transverse stria toward anterior area, wide posterior area glabrous, depressed, very finely punctate, with short yellowish-white pubescence not obscuring integument anteriorly. Wide central area of postclypeus with decumbent white pubescence close to from (more yellowish-brown in sides of this area), with short, bristly white pubescence adjacent to the anteclypeus, glabrous laterally; with a few long, erect dark setae. Labrum coplanar with anteclypeus posteriorly, inclined anteriorly; with short, decumbent yellowish-white pubescence close to clypeus, gradually sparser, yellowish-brown pubescent toward apex, with a few long, erect dark setae interspersed; apex with fringe of golden pubescence. Distance between upper eye lobes 0.23 times length of scape; in frontal view, distance between lower eye lobes 0.56 times length of scape. Antennae 3.1 times elytral length, attaining elytral apex at distal end of antennomere V. Scape with inner and ventral sides distinctly widened from near base to about basal quarter, nearly parallel-sided in remaining area; with yellowish-white pubescence partially obscuring integument, more yellowish-brown in some areas, with long, erect, thick dark setae interspersed ventrally. Pedicel and antennomeres with yellowish-white pubescence not obscuring integument (whiter depending on light intensity); pedicel and antennomeres III-V with long, erect, thick dark setae ventrally, gradually shorter, sparser toward V; antennomeres VI-X with long, erect dark setae at apex; antennomere XI curved inward, more so distally. Antennal formula (ratio) based on antennomere III: scape = 0.57; pedicel = 0.13; IV = 0.93; V = 0.89; VI = 0.87; VII = 0.82; VIII = 0.80; IX = 0.74; X = 0.73; XI = 0.73.

Thorax. Prothorax distinctly wider than long, with large, blunt lateral tubercle. Pronotum with three tubercles, one conical, large, distinctly elevated in each side of anterior half, another elongate, less elevated, placed centrally in posterior half; with transverse row of coarse punctures near anterior margin, inverted V-shaped row of coarse punctures in anterior half centrally, wide U-shaped row of coarse punctures in posterior half centrally, coarse punctures on lateral tubercles of prothorax, and transverse row of coarse punctures near posterior margin (coarser than other punctures); remaining surface smooth; pubescence mostly light yellowish-brown, partially obscuring integument, except pubescence brown on part of lateroanterior tubercles, irregular white pubescent band laterally, from anterior margin to near middle, and irregular areas with dark yellowish-brown pubescence laterally. Sides of prothorax with row of coarse punctures near anterior and posterior margins; remaining surface smooth; with yellowish-brown pubescence nearly obscuring integument. Ventral surface of thorax with dense yellowish-white pubescence centrally, more yellowish-brown laterally. Scutellum dense yellowish-white pubescent, more pale-yellow toward apex, except sides of anterior 2/3 with brownish pubescence.

Elytra. Coarsely punctate in basal third, gradually sparser toward midlength, then smooth toward apex; apex obliquely truncated; basal 2/3 with dense white pubescence, not obscuring punctures, including small white pubescent macula on each side of scutellum, basal seventh also with yellowish-brown pubescence laterally (darker, slightly sparser in inclined area), subcircular brownish pubescent spot near suture in anterior quarter, another similar pubescent spot after middle, slightly more distant from suture than the former, large lateral macula with brownish pubescence before middle, not obscuring integument, narrowly continuing along epipleural margin to about posterior third, then distinctly widened into another large brownish pubescent macula; posterior third with yellowish-brown pubescence, distinctly sparser than white pubescence, marbled with white pubescence, except lateral brownish pubescent macula (somewhat C-shaped, with projection in anterior arm, and posterior arm forming elliptical band with sparser, slightly darker pubescence).

Legs. Femora with white pubescence partially obscuring integument, some areas lighter yellowish-brown. Tibiae with white pubescence in anterior 2/3, sparser, bristly, yellowish-brown and brown in posterior third (distinctly darker on dorsal surface of mesotibiae); protibiae with sparse, long, erect, thick black setae in ventral surface of posterior third; metatibiae with sparse, long, erect, thick black setae dorsally and laterally. Tarsi distinctly long, especially pro- and mesotarsi; dorsal surface of tarsomere I with white pubescence in basal 2/3, distinctly sparser, dark-brown in distal third; laterobasal areas of tarsomere III with white pubescence, and remaining surface with sparse dark-brown pubescence; tarsomere III nearly entirely covered with sparse brownish pubescence; basal half of tarsomere V with sparse white pubescence marbled with brownish pubescence, and remaining surface with dark-brown pubescence.

Abdomen. Ventrites with dense pale-yellow pubescence (whiter on central area of I–II), yellower at apex of I–IV; dorsal surface of ventrite V lacking central sulcus; apex of ventrite V concave centrally.

Female (Fig. 5). Differs from male by the shorter antennae (2.3 times elytral length, reaching elytral apex at apex of antennomere VI), antennomere XI not curved, and abdominal ventrite V longitudinally sulcate in anterior half centrally.

Variation. Labrum brown; ventral surface of thorax entirely dark-brown; pro- and mesocoxae dark-brown; antennae gradually yellowish-brown on lighter area of distal segments; pubescence on labrum nearly entirely yellowish-brown; erect dark setae on labrum more abundant; white pubescent area of elytra reaching or not the scutellum, but always projected forward centrally.

 $\begin{array}{l} \textbf{Dimensions (mm), holotype/paratypes male/paratypes female.} & \textbf{Total length, } 15.25/14.85-15.05/13.70-15.00; \textbf{prothoracic length, } 2.70/2.60-2.65/2.20-2.45; \textbf{anterior prothoracic width, } 3.90/3.80-3.90/3.50-3.80; \textbf{basal prothoracic width, } 3.90/3.80-4.10/3.50-3.80; \textbf{widest prothoracic width, } 4.90/4.75-4.85/4.00-4.50; \textbf{humeral width, } 5.80/5.60-5.85/5.10-5.55; \textbf{elytral length, } 11.10/10.60-10.70/10.05-11.20. \end{array}$

Type material. Holotype male – BOLIVIA, *Santa Cruz*: 4 km N Bermejo (Refugio Los Volcanes; 1045–1350 m; 18°06′S / 63°36′W), 17–24.X.2014, Wappes and Morris col. (FSCA, formerly ACMT).

Paratypes – BOLIVIA, Santa Cruz: Potrerillo del Guendá (Snake farm; 17°40′S / 63°27′W; 370-400 m), 1 male, 16.X.2011, Skillman and Wappes col. (MZSP); 4 km N Bermejo (Refugio Los Volcanes; 1000 m; 18°06′S / 63°36′W), 1 male, 4–8.X.2007, J. Wappes and R. Morris col. (ACMT); Florida (4 km N Bermejo; Refugio Los Volcanes; 18°06′S / 63°36′W; 1000–1200 m), 1 female, 25–29.X.2011, Wappes and Skillman col. (ACMT); 1 female, 28.X.2011, Skillman and Wappes col. (FWSC); 1 female, 5.XII.2013, Skillman and Wappes col. (FWSC); 1 male, 10.XII.2015, Skillman, Wappes and Kuckartz col. (FWSC); 1 female, 12.XII.2015, Skillman, Wappes and Kuckartz col. (FWSC); 3.7 km SSE Buena Vista (Hotel Flora and Fauna; 430 m; blacklight trap; tropical transition forest), 1 male, 5–15.XI.2001, M.C. Thomas and B.K. Dozier col. (FSCA); vicinity of Buena Vista (Flora and Fauna Hotel), 1 male, 27–31.X.2002, Morris and Wappes col. (RFMC); Potrerillo del Guendá (Reserva Natural aka Snake farm; 400 m; 17°40′15″S / 63°27′26″W), 1 female, 13–17.XI.2012, Bettella, Bonaso and Romero col. (MZSP); 1 male, 13–17.XI.2014, Wappes and Morris col. (ACMT).

Etymology. We are pleased to name this species for Robert Perger, La Paz, Bolivia for his enjoyable and informative consultations and published taxonomic contributions to the Coleoptera fauna of Bolivia.

Remarks. Oreodera pergeri new species is similar to O. albicans Monné and Fragoso, 1988, but differs as follows: antennomere III in male distinctly shorter than humeral width; second dark macula on dorsal surface of the elytra near suture small and subcircular in both sexes; antennomere IV shorter than III in male; femora shorter, with femoral club thicker in both sexes. In O. albicans, the antennomere III in male is about as long as or slightly shorter than humeral width, second dark macula on dorsal surface of the elytra near suture, slender and elongated in both sexes, antennomere IV in male as long as or slightly longer than III, femora longer with femoral club slenderer in both sexes. It differs from O. albata Villiers, 1971, O. granulifera Bates, 1872, and O. basiradiata Tippmann, 1960 (see photograph of the holotypes at Bezark 2019), by the elytral punctures finer and denser (coarser and sparser in those species); male also differs from that of O. basiradiata by the longer antennae (shorter in male of O. basiradiata); male differs from that of O. granulifera by the antennomere III distinctly shorter than humeral width (almost as long as humeral width in O. granulifera), and by the protibiae not granulated ventrally (granulated in O. granulifera), and lacking tubercles on basal area of the elytra in both sexes (present in O. granulifera). Oreodera pergeri differs from O. leucostigma Monné and Fragoso, 1988 (see photograph of the holotype at Bezark 2019), especially by the pronotum with sparse punctures in wide central area (noticeably abundant in O. leucostigma). It differs from O. semialba Bates, 1874 (see photograph of the holotype at Bezark 2019), by the humerus not granulated (granulated in O. semialba), and elytra lacking tubercles in basal third (tuberculate in *O. semialba*).

Little is known of the natural history of this species. Adults are most commonly encountered at MV/UV lights, or otherwise at night, crawling or mating on recently felled tree trunks.

Oreodera howdeni Monné and Fragoso, 1988 (Fig. 6–7)

Oreodera howdeni Monné and Fragoso 1988: 818; Hovore 1989: 255 (distr.); Chemsak et al. 1992: 131 (checklist); Monné and Giesbert 1994: 236 (checklist); Monné 1994: 28 (cat.); Julio et al. 2000: 45 (holotype); Martínez 2000: 94 (distr.); Monné 2005: 191 (cat.); Monné and Hovore 2006: 207 (checklist); Swift et al. 2010: 46 (distr.); Monné and Monné 2016: 55 (holotype); Monné 2018: 264 (cat.).

This species was originally described based on 3 females from Colombia. Subsequently, Hovore (1989) recorded it for Costa Rica, and Monné and Giesbert (1994) for Mexico (Veracruz). The holotype female (see photograph at Bezark 2019), and a paratype female were destroyed by the 2018 fire that devastated the MNRJ. According to the original description a third paratype female is deposited in Carleton University, Ottawa, Canada. The male of *O. howdeni* is illustrated (Fig. 6) and commented on for the first time here. It differs from the female by the longer antennae (2.15 times elytral length, reaching elytral apex at basal third of antennomere VII), and has the last abdominal ventrite flattened and not centrally sulcate. In the female, the antennae are 1.85 times elytral length, reaching elytral apex at basal third of antennomere VIII, and the last abdominal ventrite is convex with a shallow longitudinal, central sulcus in basal half.

Material examined. MEXICO, *Chiapas* (**new state record**): Palenque (picking old logs), 1 female, 16.VI.2009, Skillman and Hildebrant col. (FWSC). BELIZE (**new country record**), *Toledo*: BARC San Pedro Columbia (16°16′43″N / 88°57′49″W; on dead tree trunks at night), 1 male, 23.IX.2004, P.W. Kovarik col. (ACMT).

Oreodera boucheri Néouze and Tavakilian, 2010 (Fig. 8)

Oreodera boucheri Néouze and Tavakilian 2010: 59; Tavakilian and Néouze 2011: 70; Morvan and Roguet 2013: 15 (distr.); Monné 2018: 257 (cat.).

This species was originally described based on both sexes and known only from French Guiana. Here, although far from French Guiana, it is recorded from Peru near the western border of the Amazone Biome.

Material examined. PERU (new country record), San Martin: Escalera Lodge (435 m; Tarapoto), 1 female, 9–12.X.2012, J.B. Heppner col. (FSCA).

Oreodera vulgata Monné and Fragoso, 1988 (Fig. 9)

Oreodera vulgata Monné and Fragoso 1988: 827; Monné 1994: 32 (cat.); Monné and Giesbert 1994: 236 (checklist); Julio et al. 2000: 46 (holotype); Monné 2005: 196 (cat.); Monné and Hovore 2006: 208 (checklist); Monné et al. 2010: 246 (distr.); Monné et al. 2012: 48 (distr.); Nascimento et al. 2016: 558 (distr.); Monné and Monné 2016: 56 (holotype); Monné et al. 2016: 21 (distr.); Nascimento et al. 2017: 89 (distr.); Monné and Monné 2017: 234 (distr.); Monné 2018: 273 (cat.).

This species was described based on a series of both sexes, from Brazil (Bahia, Goiás, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina) and Paraguay (Ñeembucú). Unfortunately, the holotype and paratypes (83) were destroyed in the 2018, MNRJ fire. The recently collected male specimen listed below allows presentation of a new illustration of the species and represents a new department record for Paraguay.

Material examined. PARAGUAY, *Paraguarí* (**new department record**): Cerro Acahay (Finca R. Garcete; 325 m; 25°52′S / 57°10′W), 1 male, 03.XII.2012, David Brzoska col. (ACMT).

Oreodera sororcula Martins and Monné, 1993 (Fig. 10–17)

Oreodera sororcula Martins and Monné 1993: 135; Monné 1994: 32 (cat.); Monné and Giesbert 1994: 236 (checklist); Monné 2005: 195 (cat.); Morvan and Morati 2006: 37 (distr.); Monné and Hovore 2006: 208 (checklist); Touroult et al. 2010: 31; Tavakilian and Néouze 2011: 83 (hosts); Machado et al. 2012: 192 (hosts); Giuglaris 2012: 63 (distr.); Morvan and Roguet 2013: 16 (distr.); Santos-Silva and Botero 2016: 427 (distr.); Monné 2018: 271 (cat.).

This species was originally described based on 2 males (holotype from Venezuela, AMNH; paratype from Guyana, MZSP) and 1 female (from Guyana, AMNH). The paratype belonging to the MZSP collection was reported as being a female but is a male. However, when describing the dimensions of the specimens, Martins and Monné (1993) correctly provided the measurements of two males and one female.

Morvan and Morati (2006) were the first to report this species in French Guiana. Although the paratype female from Guyana was not examined, the holotype and paratype (Fig. 10-13) males are distinctly lighter than specimens from French Guiana (specimens also examined through photographs, Fig. 14–17). In addition to the difference in color, the anterolateral pronotal tubercles are somewhat smaller and more distinctly conical in the holotype and paratype male than in the specimens from French Guiana. However, as no other morphological features were found, it is probable that the differences in the French Guiana specimens are simply local variants.

Material examined. GUYANE: upper Mazaruni River, paratype male, IX–X.1938, A. S. Pinkus col. (MZSP). FRENCH GUIANA: Kaw road (D-6), PK 38 (Amazone Nat. Lodge; Montagne de Kaw; 4°33′N / 52°11′W; 970 ft; MV/light), 1 female, 11–23.VIII.2017, J. Wappes and R. Morris col. (ACMT).

Oreodera stictica Monné and Fragoso, 1988

(Fig. 18–21)

Oreodera stictica Monné and Fragoso 1988: 814; Monné 1994: 32 (cat.); Monné and Giesbert 1994: 236 (checklist); Julio et al. 2000: 46 (holotype); Monné 2005: 195 (cat.); Monné and Hovore 2006: 208 (checklist); Monné and Monné 2016: 56 (holotype); Monné 2018: 271 (cat.).

This species was described based on three males from Brazil (Mato Grosso) which were subsequently destroyed in the 2018 MNRJ fire. Fortunately, the holotype was illustrated in the original description (Fig. 20) and photographed by Steven W. Lingafelter in 2003, although, by then it had become seriously contaminated by fungi (Fig. 21), which dramatically altered its appearance rendering it nearly unrecognizable.

Although not mentioned in the original description, the metatibiae are distinctly widened from near base, making the basal third wider than usual among *Oreodera* species.

Since the original description, the species has only been mentioned in catalogs and checklists. Often, species occurring in the Brazilian state of Mato Grosso also occur in Bolivia which is the case here.

Material examined. BOLIVIA (new country record), Santa Cruz: Potrerillo del Guendá (17°40.26′S / 65°24.44′W), 2 males, 9-28.XI.2006, B.K. Dozier and F. and J. Romero col. (ACMT); (Snake farm; 17°40′S / 63°27′W; 370–400 m), 1 female, 3.XII.2012, Skillman and Wappes col. (FWSC); 1 male, 15–22.XI.2011, Bettella, Bonaso and Romero col. (MZSP); Buena Vista (410 m), 1 female, 29.X.1999, Porter and Stange col. (FSCA); Huaico (430 m; 17°40′S / 63°24′W), 1 female, 21.XI.2013, Skillman and Wappes col. (FWSC); Huaico (near Potrerillo del Guendá; 430 m; MV/UV light; 63°26′S / 17°40′W), 1 male, 21.XI.2013, Wappes and Skillman col. (ACMT); 1 male, 1 female, 27–29.XI.2013, Wappes and Kuckartz col. (ACMT).

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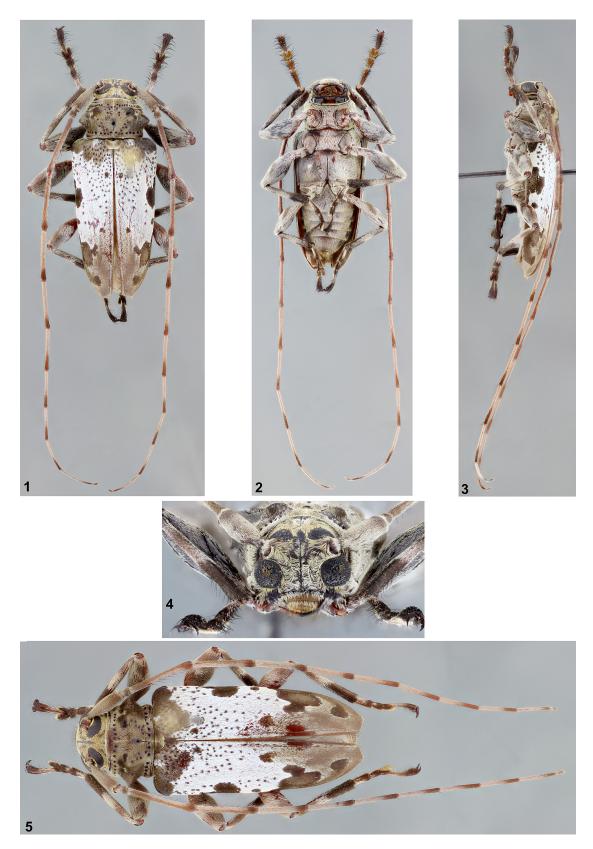
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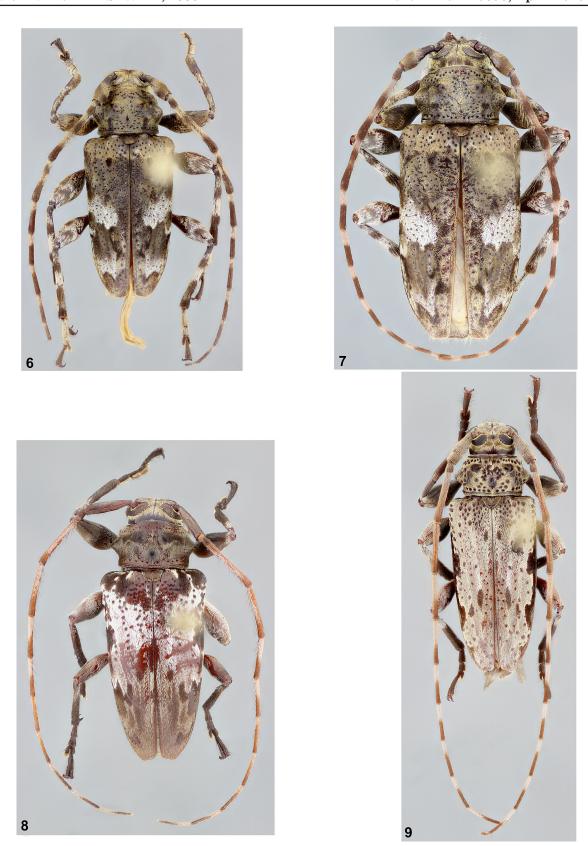
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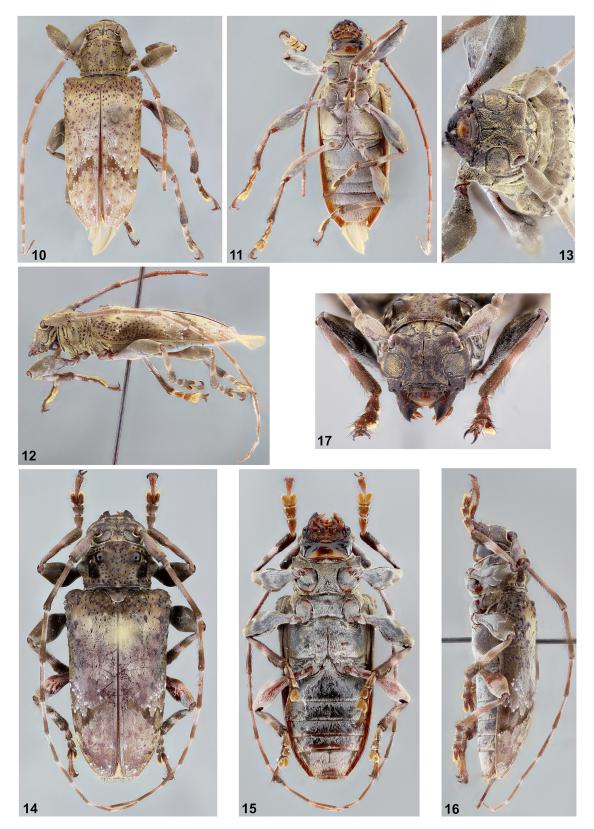
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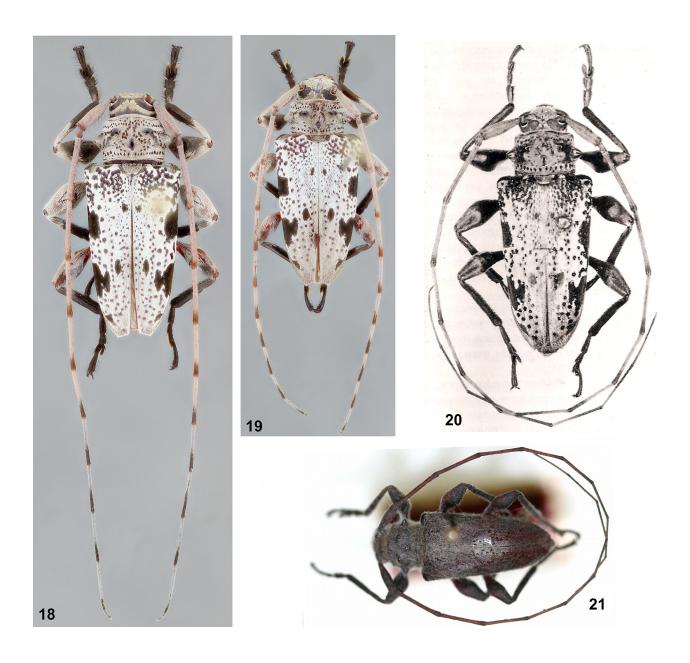
Figures 1–5. Oreodera pergeri. 1–4) Holotype male. 1) Dorsal habitus. 2) Ventral habitus. 3) Lateral habitus. 4) Head, frontal view. 5) Paratype female, dorsal habitus.



Figures 6–9. Oreodera species. **6–7)** Oreodera howdeni. **6)** Dorsal habitus, male. **7)** Dorsal habitus, female. **8)** Oreodera boucheri, dorsal habitus, female. **9)** Oreodera vulgata, dorsal habitus, male.



Figures 10–17. Oreodera sororcula. 10–13) Paratype male. 10) Dorsal habitus. 11) Ventral habitus. 12) Lateral habitus. 13) Head, frontal view. 14–17) Female. 14) Dorsal habitus. 15) Ventral habitus. 16) Lateral habitus. 17) Head, frontal view.



Figures 18–21. Oreodera stictica. **18)** Dorsal habitus, male. **19)** Dorsal habitus, female. **20)** Holotype male, from Monné and Fragoso (1988). **21)** Holotype male, dorsal habitus.