

New Spectrophotometric Methods for the Quantification of an Anti-Peptic Ulcer Drug in Bulk and Tablets

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Abstract

Two simple, economical and reproducible fundamental and derivative UV spectrophotometric methods were developed and validated for determination of Famotidine in bulk and dosage form. Famotidine showed maximum absorption at 281 nm in phosphate buffer pH 7.5 while it has 282 nm as its absorption maxima in borate buffer pH 9.0. The linearity was determined in the concentration range of 30-80 µg/mL (r^2 as 0.9993 & 0.9987) and 10-60 µg/mL (r^2 as 0.9991 & 0.9995) for the fundamental and derivative methods in phosphate and borate buffers. The developed methods were validated as per ICH guidelines. Recovery studies gave satisfactory results indicating that none of the major additives/excipients interfered with the assay method. This method may be useful for routine laboratory analysis of famotidine.

Keywords: Borate buffer pH 9.0, derivative, famotidine, fundamental, phosphate buffer pH 7.5, UV spectrophotometric, validation

INTRODUCTION

Famotidine is a histamine H₂ receptor antagonist that inhibits stomach acid production [1]. It is commonly used in the treatment of peptic ulcer disease and gastroesophageal reflux disease. Chemically Famotidine (Fig. 1) is 3-[[2-(diaminomethylideneamino)-1, 3-thiazol-4-yl]methylsulfanyl]-N'-sulfamoylpropanimidamide with a molecular

weight of 337.44 g/mol. It is very slightly soluble in water. Literature review reveals analytical methods like UV spectroscopy [2-10], colorimetry [11-18], spectrofluorimetry [19], HPLC [20-30], HPTLC [31], flow injection analysis [32, 33] and electrochemical analysis [34-36]. An attempt has been made to develop a simple, sensitive and economical UV spectroscopic method that can be used in routine analysis.

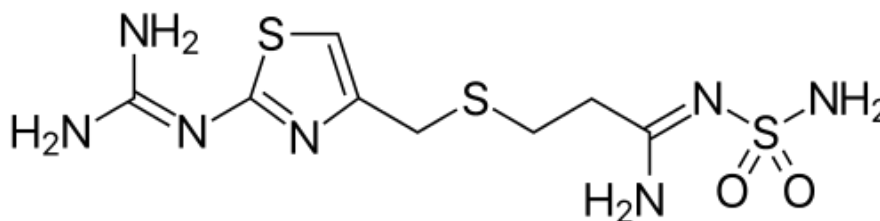


Figure 1: Chemical structure.

EXPERIMENTAL

Instrumentation

Double beam UV-Visible Spectrophotometer (UV-1800) Shimadzu

(Japan) connected to computer located with software UV probe was employed with spectral band width of 1nm and wavelength accuracy of 0.3 nm with a pair

of 10 mm path length matched quartz cells. For scanning, the wavelength range selected was 400 nm to 200 nm with medium scanning speed. All weights were taken using electronic balance (Shimadzu, Japan). All Experiments were performed at room temperature.

Chemicals and Reagents

Pure sample of Famotidine was supplied by Dr. Reddy's laboratories, Hyderabad and the commercial tablets were purchased from the local pharmacy. Potassium di hydrogen orthophosphate, boric acid and sodium hydroxide (AR grade) were procured from Qualigens and distilled water was used throughout the study.

Preparation of Phosphate Buffer pH 7.5

Phosphate buffer pH 7.5 was prepared by dissolving 45.36 gm of potassium di hydrogen orthophosphate in sufficient water to produce 1000 mL.

Preparation of Borate Buffer pH 9.0

Borate buffer pH 9.0 was prepared by dissolving 6.20 gm of boric acid in 500ml of water, pH was adjusted to 9.0 with sodium hydroxide and diluted with water to produce 1000ml.

Preparation of Stock (1000 µg/mL) and Working Standard (100 µg/mL) Solutions

Accurately weighed about 25.0 mg of Famotidine was weighed and transferred to clean and dry volumetric flask, dissolved in methanol and made up the volume to 25 mL with the same solvent. 2.5 mL of the above solution was transferred to different 10 mL volumetric flasks and diluted with phosphate buffer pH 7.5 and borate buffer pH 9.0 separately. This solution was used for making series of dilutions for calibration curve. All solutions were freshly prepared before analysis.

Preparation of Sample Solution (from tablets)

20 tablets of Famotidine (Famocid) were weighed, powdered and weight of powder equivalent to 10 mg of Famotidine was

taken into a 10mL volumetric flask, dissolved in methanol, sonicated for 15 min and volume was made up to the mark with the same solvent and filtered. A solution of 100 µg/mL was prepared using phosphate buffer pH 7.5 and borate buffer pH 9.0 separately. The above solutions were suitably diluted to the required concentrations with phosphate buffer pH 7.5 and borate buffer pH 9.0.

Fundamental UV Spectrophotometric Method (D⁰)

The drug solutions were scanned in the UV range (200-400nm) and the absorption spectra were recorded against the reagent blank. The absorbance was measured at 281 nm in phosphate buffer pH 7.5 and at 282 nm in borate buffer pH 9.0.

First Order Derivative UV Spectrophotometric Method (D¹)

The spectra of all the drug solutions obtained in the zero order method were derivatized into first order spectra using the UV probe software and the derivative absorbance was measured at corresponding maxima and minima. The amplitude was calculated in the range of 269.23 nm to 302.2 nm in phosphate buffer pH 7.5 and 271.23 nm to 304 nm in borate buffer pH 9.0.

Validation

The methods were validated [37] as per International Conference of Harmonization (ICH) guidelines for linearity, precision and accuracy.

Linearity

Aliquots of working standard solutions were suitably diluted with phosphate buffer pH 7.5 and borate buffer pH 9.0 and the absorbance of each solution was measured as per the method. Linearity plots were constructed for concentration v/s absorbance in D⁰ and concentration v/s amplitude in D¹ methods.

Precision

The precision of the method was studied in terms of repeatability and intermediate precision. Replicate sample solutions of

20, 40 and 60 µg/mL famotidine were prepared in both the buffers and analysed for D^0 and D^1 on the same day and on different days. The absorbance, derivative absorbance was measured from which the assay and % RSD was calculated.

Accuracy

Accuracy was studied by standard addition method at three different levels (50, 100 and 150 %). The recovery and % RSD were calculated.

Assay

20 tablets of Famotidine (Famocid) were used to prepare the sample solutions. A solution of 100 µg/mL was prepared using phosphate buffer pH 7.5 and borate buffer pH 9.0 as per the above procedure. These solutions were suitably diluted to the required concentrations with phosphate buffer pH 7.5 and borate buffer pH 9.0 and assay was performed.

RESULTS AND DISCUSSION

Two simple and sensitive fundamental and first order derivative UV spectroscopic methods have been developed in phosphate buffer pH 7.5 (method A) and borate buffer pH 9.0 (method B). Several buffers and solvents have been used in the method optimization and the best optical characteristics were obtained with the selected phosphate buffer pH 7.5 and borate buffer pH 9.0. These methods were

validated as per the ICH guidelines and the results are discussed below.

Linearity

The linearity was evaluated using different concentrations of standard solution. The Beer-Lambert's law was obeyed in the concentration range of 30 – 80 µg/mL in phosphate buffer pH 7.5 (D^0 & D^1) and 10–60 µg/mL in borate buffer pH 9.0 (D^0 & D^1) as confirmed from the correlation coefficients. The corresponding spectra and linearity plots are given in Fig. 2a–2d, Fig. 3a–3d and the data is tabulated in Table 1.

Precision

Precision was studied in terms of intraday and interday precision. The results of the precision indicate that the methods are reliable. The % RSD for assay in D^0 and D^1 was found to be 0.11-0.75 (intraday) and 0.05-1.02 (inter day) in both the buffers. The results are given in the Table 2a and 2b.

Accuracy

Accuracy was evaluated by standard addition method and the percent recovery calculated at 50, 100 and 150% levels of a pre analysed formulation solution were obtained in the range of 99.2–101.3%. The % RSD was found to be 0.15–1.08. The results are given in Table 3.

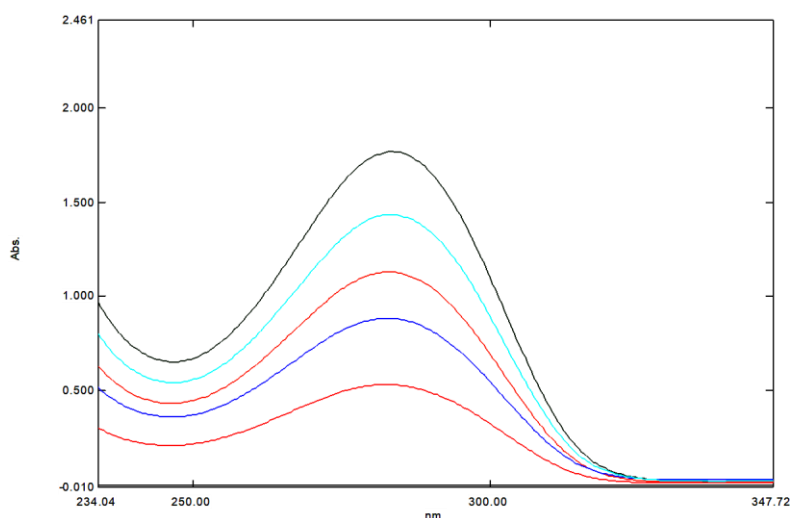


Figure 2a: D^0 spectra in phosphate pH 7.5.

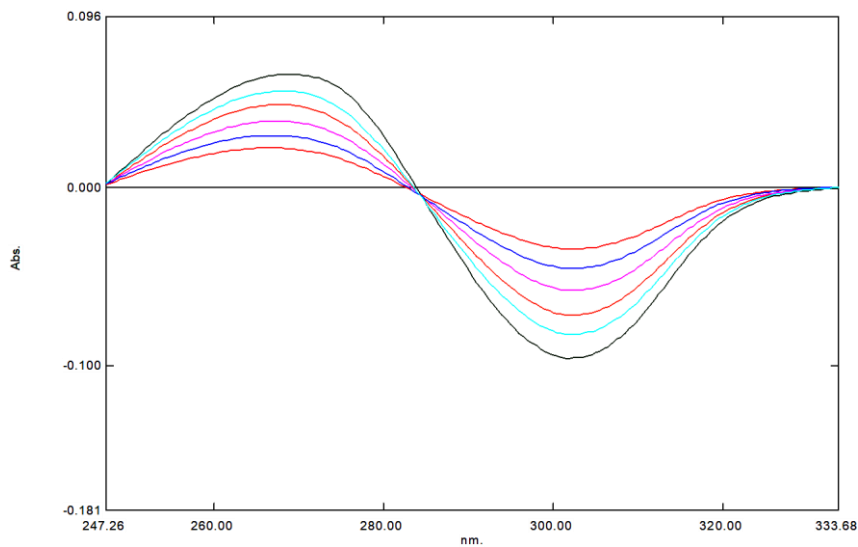


Figure 2b: D^1 spectra in phosphate pH 7.5.

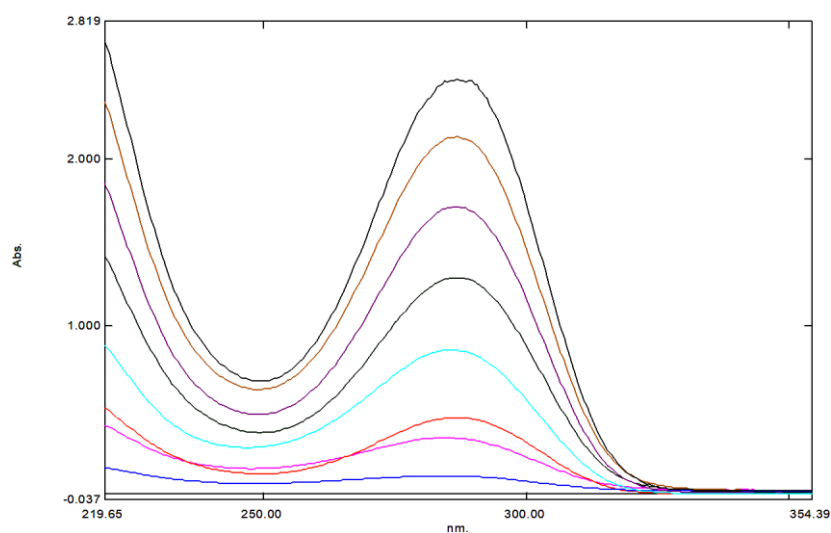


Figure 2c: D^0 spectra in borate pH 9.0.

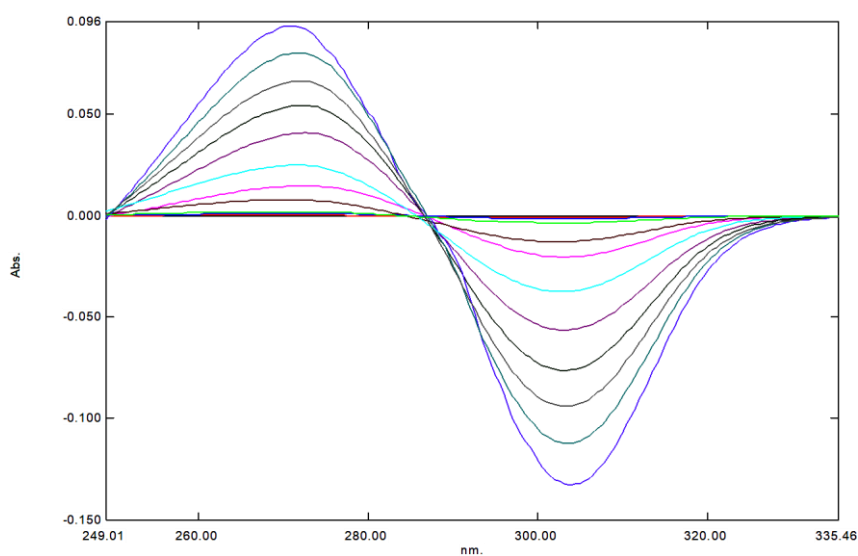


Figure 2d: D^1 spectra in borate pH 9.0.

Table 1: Data for linearity.

Phosphate Buffer pH 7.5			Borate Buffer pH 9.0		
Conc. (µg/mL)	Absorbance	Amplitude	Conc. (µg/mL)	Absorbance	Amplitude
30	0.8845	0.056	10	0.4514	0.034
40	1.1328	0.075	20	0.8555	0.062
50	1.4317	0.095	30	1.2826	0.097
60	1.7627	0.118	40	1.7096	0.131
70	2.0230	0.138	50	2.129	0.161
80	2.3534	0.159	60	2.4721	0.193

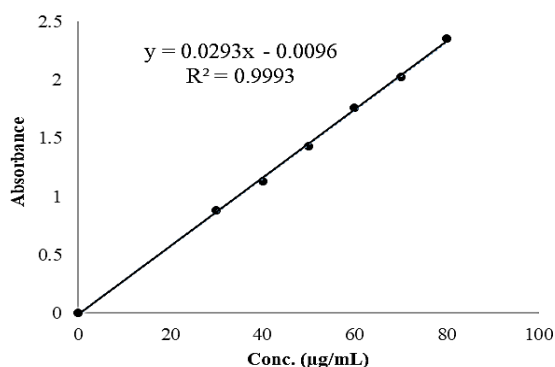


Figure 3a: Linearity plot (D^0), phosphate 7.5.

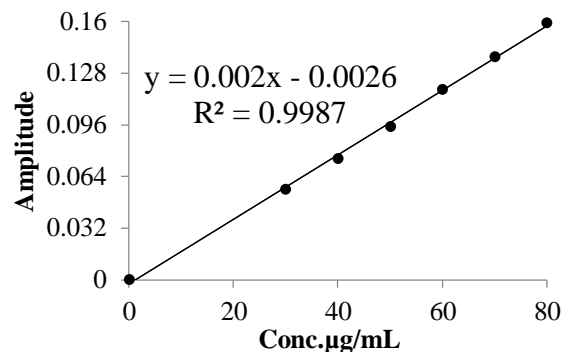


Figure 3b: Linearity plot (D^1), phosphate 3.5.

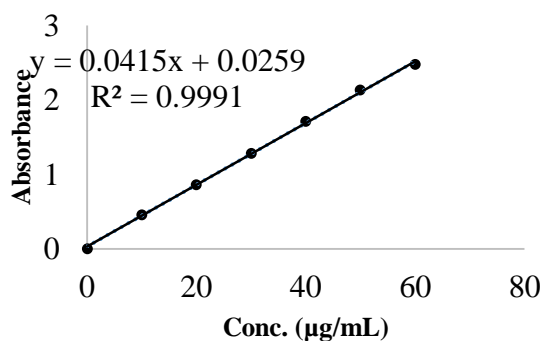


Figure 3c: Linearity plot (D^0), borate 9.0.

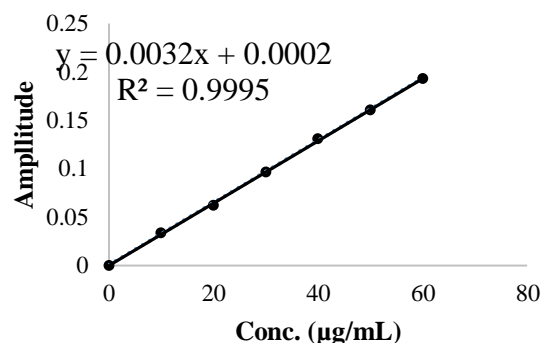


Figure 3d: Linearity plot (D^1), borate 9.0.

Table 2a: Precision data in phosphate buffer pH 7.5.

Conc. (µg/mL)	Intraday Precision		Interday Precision	
	D^0	D^1	D^0	D^1
	*Assay (% w/w) ± SD, % RSD			
20	102.17 ± 0.76, 0.75	101.3 ± 0.57, 0.56	103.6 ± 0.61, 0.61	101.1 ± 1.04, 1.02
40	103.9 ± 0.36, 0.34	101.5 ± 0.46, 0.45	104.7 ± 0.18, 0.18	101.3 ± 0.15, 0.15
60	103.7 ± 0.45, 0.38	101.3 ± 0.57, 0.56	104.1 ± 0.90, 0.86	101.5 ± 0.5, 0.49

*Mean of three determinations

Table 2b: Precision data in borate buffer pH 9.0.

Conc. (µg/mL)	Intraday Precision		Interday Precision	
	D^0	D^1	D^0	D^1
	*Assay (% w/w) ± SD, % RSD			
20	101.6 ± 0.76, 0.75	99.5 ± 0.40, 0.40	102.2 ± 0.8, 0.76	99.7 ± 0.40, 0.40
40	103.1 ± 0.12, 0.11	101.5 ± 0.75, 0.73	104.0 ± 0.11, 0.11	100.5 ± 0.45, 0.42
60	103.7 ± 0.23, 0.22	99.64 ± 1.13, 1.14	104.1 ± 0.05, 0.05	99.4 ± 0.96, 0.96

*Mean of three determinations.

Table 3: Recovery studies.

Level (%)	Phosphate buffer pH 7.5		Borate buffer pH 9.0	
	D ⁰	D ¹	D ⁰	D ¹
	*Recovery ± SD, % RSD			
50	99.4 ± 0.15, 0.15	100.2 ± 0.57, 0.57	99.2 ± 0.45, 0.45	99.7 ± 0.23, 0.23
100	100.2 ± 0.92, 0.91	101.1 ± 0.57, 0.52	100.7 ± 0.72, 0.71	101.3 ± 0.95, 0.96
150	101.2 ± 0.69, 0.68	101.3 ± 0.57, 0.56	101.4 ± 0.44, 0.44	100.9 ± 1.08, 1.08

*Mean of three determinations

Assay

The developed methods were also applied for the determination of Famotidine in commercial tablets. There was no

interference from the excipients as observed from the assay as stated against the label claim and the results are given in Table 4.

Table 4: Assay in tablets.

Method	Brand	Label claim (mg)	*Amount obtained (mg)	*Assay (% w/w) ± SD	
Phosphate buffer pH 7.5	FAMOCID	40.0	D ⁰	39.9	99.75 ± 0.82
			D ¹	40.2	100.5 ± 0.76
Borate buffer pH 9.0			D ⁰	40.5	101.25 ± 0.83
			D ¹	39.7	99.25 ± 1.04

*Mean of three determinations

Table 5: Summary of optical and validation parameters.

Parameters	Phosphate buffer pH 7.5		Borate buffer pH 9.0	
	D ⁰	D ¹	D ⁰	D ¹
Range (µg/mL)	30 – 80	30 – 80	10 – 60	10 – 60
Regression equation	y = 0.0293x - 0.0096	y = 0.002x - 0.0026	y = 0.0415x + 0.0259	y = 0.0032x + 0.0002
Correlation coefficient (r ²)	0.9993	0.9987	0.9991	0.9995
Intraday precision (% RSD)	0.34-0.75	0.45-0.56	0.11-0.75	0.40-1.14
Inter day precision (% RSD)	0.18 - 0.86	0.15-1.02	0.05-0.76	0.40-0.96
Accuracy (% RSD)	0.15 – 0.91	0.52-0.57	0.44 – 0.71	0.23-1.08
Sandell's sensitivity (µg/cm ² /0.001)	0.034	-	0.022	-
Molar Absorptivity (L. mol. ⁻¹ cm ⁻¹)	9793.3	-	14464.4	-
LOD (µg/mL)	1.235	2.83	0.872	0.309
LOQ (µg/mL)	4.081	9.37	2.879	1.021

CONCLUSION

Two simple and economical UV spectrophotometric methods were proposed for the determination of famotidine with reasonable precision and accuracy. Validation parameters justify this method for application to quantification of Famotidine in pure and dosage form. Moreover, the methods are free from interference by common additives and excipients making them specific for the assay and evaluation of Famotidine in

pharmaceutical dosage form.

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REFERENCES

1. Indian Pharmacopoeia (2014), Government of India, Ministry of

- Health & Family Welfare, The Indian Pharmacopoeia Commission, Ghaziabad, Volume 2.
- Li H, Xuan J (1993), "UV-spectrophotometry of famotidine tablets", *Zhongguo Yiyao gangye Zazhi*, Volume 24, pp. 319–321.
 - Mohite MT, Shet SN, Shaikh S, Vaidya VR, Karodi RS (2010), "Analytical method development of famotidine USP in bulk and single component formulation", *IJRAP*, Volume 1, Issue 2, pp. 475–479.
 - Okram ZD, Kanakapura B, Pavagada JR et al. (2011), "Simple and sensitive UV spectrophotometric methods for determination of famotidine in tablet formulations", *Farmacia*, Volume 59, Issue 5, pp. 647–657.
 - Yogita BW, Dipak DP, Joshi NS, Bari SB (2013), "Development and validation of difference spectrophotometric method for the estimation of famotidine in bulk and pharmaceutical dosage form", *Int. J. Drug Dev. & Res.*, Volume 5, Issue 2, pp. 272–277.
 - Erum Z, Sohail H, Hafiz Mi et al. (2014), "Development and validation of a UV spectrophotometric method for determination of famotidine in suspensions", *Fuuast J. Biol.*, Volume 4, Issue 1, pp. 7–12.
 - Safila N, Farya Z (2014), "UV spectrophotometric assay of famotidine formulations", *American Journal of Pharmacy and Pharmacology*, Volume 1, Issue 3, pp. 28–31.
 - Vijaya Bhaskara RT, Rambabu C, Sowjanya RN et al. (2014), "Assay of famotidine in API and dosage forms by UV direct and UV derivative spectrophotometric methods", *Der Pharmacia Sinica.*, Volume 5, Issue 1, pp. 57–65.
 - Ranganath MK, Narendra RK (2014), "Method development and validation for the estimation of famotidine in pure and tablet dosage form by derivative UV spectroscopy", *Rajiv Gandhi University of Health Sciences Journal of Pharmaceutical Sciences*, Volume 4, Issue 1, pp. 17–21.
 - Ritesh K, Amrishi C, Pawan KG (2017), "Development and validation of ultraviolet spectrophotometric method for quantitative estimation of famotidine in bulk and tablet dosage form", *Asian J Pharm Clin Res.*, Volume 10, Issue 8, pp. 381–385.
 - Agarwal YK, Shivaramachandra K, Singh GN et al. (1992), "Spectrophotometric determination of famotidine in pharmaceutical preparations", *J Pharm Biomed Anal.*; Volume 10, Issue 7, pp. 521–523. [https://doi.org/10.1016/0731-7085\(92\)80074-W](https://doi.org/10.1016/0731-7085(92)80074-W)
 - Kamath BV, Shivram K, Vangani S (1992), "Spectrophotometry determination of famotidine by charge transfer complexation", *Anal. Let.*, Volume 25, Issue 12, pp. 2239–2247.
 - Abuzuhri AZ, Shubiartah RM, Badah GM (1999), "Extractional spectrophotometric determination of famotidine in pharmaceutical formulations", *J. Pharm. Biomed. Anal.*, Volume 21, Issue 2, pp. 459–465.
 - Sastry CSP, Chintalapati R (2000), "Two simple visible spectrophotometric methods for the assay of famotidine in bulk drug form and formulation", *East Pharm.*, Volume 43, pp. 159–161.
 - Rahman N, Kashif M (2003), "Kinetic spectrophotometric determination of Famotidine in commercial dosage forms", *Analytical Sciences*, Volume 19, Issue 6, pp. 907.
 - Rahman N, Kashif M (2003), "Application of ninhydrin to spectrophotometric determination of famotidine in drug formulation", *IL Farmaco*, Volume 58, Issue 10, pp. 1045–1050. <https://doi.org/10.1016/S0014->

- 827X(03)00184-8.
17. Darwish IA, Hussein SA, Mohmoud AM et al. (2007), "Sensitive indirect spectrophotometric method for determination of H₂-receptor antagonists in pharmaceutical formulations", *Int. J. Biomed. Sci.*, Volume 3, Issue 2, pp. 123–103.
 18. Kanakapura B, Okram Z (2011), "Spectrophotometric determination of famotidine using sulphonphthalein dyes", *Química Nova.*, Volume 4, Issue 5, pp. 735–742. <http://dx.doi.org/10.1590/S0100-40422011000500002>
 19. Walash MI, Sharaf-El-Din, MK, El-Sayed MM et al. (2009), "Spectrofluorimetric determination of famotidine in pharmaceutical preparations and biological fluids, Application to stability studies", *J Fluoresc*, Volume 19, pp. 333–344.
 20. Biffar SE, Mazzo DJ (1986), "Reversed-phase determination of famotidine, potential degradates, and preservatives in pharmaceutical formulations by high-performance liquid chromatography using silica as a stationary phase", *J. Chromatogr.*, Volume 363, Issue 2, pp. 243–249.
 21. Mutaz A, Sheikh S, Hanan AN, Adnan BA (1989), "High pressure liquid chromatographic analysis and dissolution of famotidine in tablet formulation", *Anal Lett.*, Volume 22, Issue 11, 12, pp. 2501–2505.
 22. Cakir B, Tosun AV, Sahin MF (1997), "Quantitative high-performance liquid chromatography analysis of Famotidine in Pharmaceutical dosage forms", *Pharmaceutical Science*, Volume 3, Issue 10, pp. 493–495.
 23. Tahboub YR, Zaater MF, Najib NM (1998), "Reversed phase liquid chromatography method for the determination of Famotidine in serum", *Quin Anal*, Volume 17, pp. 117–120.
 24. Zendelovska D, Stafilov T (2003), "High-performance liquid chromatographic determination of famotidine in human plasma using solid-phase column extraction", *J. Serb. Chem. Soc.*, Volume 68, Issue 11, pp. 883–892.
 25. Helali N, Darghouth F, Monser L (2004), "RP-HPLC determination of famotidine and its potential impurities in pharmaceuticals", *Chromatographia.*, Volume 60, Issue 7, 8, pp. 455–460.
 26. Zarghi A, Shapaati A, Foroutan SM et al. (2005), "Development of a rapid HPLC method for determination of famotidine in human plasma using a monolithic column", *J. Pharm. Biomed. Anal.*, Volume 39, Issue 3, 4, pp. 677–680.
 27. Helali N, Monser L (2008), "Stability indicating method for famotidine in pharmaceuticals using porous graphite column", *Journal of Serparation Sciences*, Volume 31, Issue 2, pp. 276–282.
 28. Dowling G, Gallo P, Fabbrocino S et al. (2008), "Determination of famotidine in human plasma and urine by high-performance liquid chromatography", *Food Addit. Conta.*, Volume 25, Issue 12, pp. 1497–1508.
 29. Vamsi Krishna M, Madhavi G, Rama Prasad LA et al. (2010), "Impurity profiling of famotidine in bulk drugs and pharmaceutical formulations by RP-HPLC method using ion pairing agent", *Der Pharm. Lett.*, Volume 2, Issue 3, pp. 1–11.
 30. Reddy TVB, Ramu G, Lakshmana Rao PV et al. (2012), "Development and validation of stability indicating reverse phase liquid chromatography method for assay of famotidine in bulk and formulations", *Rasayan J.Chem.*, Volume 5, Issue 2, pp. 250–255.
 31. Campbell AN, Sherma J (2003), "Determination of famotidine in acid reduction tablets by HPTLC and video densitometry of fluorescence quenched

- zones”, *J Liquid Chromatogr Relat Technol.*, Volume 26, Issue 16, pp. 2719–2722.
<https://doi.org/10.1081/JLC-120024542>
32. Tzanavaras PD, Verdoukas A, Balloma T (2006), “Optimization and validation of a dissolution test for famotidine tablets using flow injection analysis”, *J. Pharm. Biomed. Anal.*, Volume 41, Issue 2, pp. 437–441.
 33. Tang YH, Wang NN, Xiong XY et al. (2007), “A new sensitive flow-injection chemiluminescence method for the determination of H₂-receptor antagonists”, *J Biolumin Chemilumin.*; Volume 22, Issue 4, pp. 343–348. DOI: 10.1002/bio.969
 34. Squella JA, Valencia J, Lenus I et al. (1989), “Polarographic determination of famotidine in dosage forms”, *J Assoc off Anal Chem*, Volume 72, pp. 549–551.
 35. Ayad MM, Shalaby A, Abdullatif HE et al. (2002), “Potentiometric determination of famotidine in pharmaceutical formulations”, *J Pharm Biomed Anal.*, Volume 29, pp. 247–254.
 36. Ibrahim WM, Sharaf-El-Din MK, El-Sayed MM (2005), “Polarographic determination of famotidine through complexation with Nickel(II) Chloride”, *J Chin Chem Soc.*, Volume 52, pp. 927–935.
 37. (Nov 2005), “ICH Harmonized Tripartite Guidelines, Validation of Analytical Procedures: Text and Methodology Q2 (R1)”.