



Type of the Paper (Article)

## Extraction and Thermogravimetric Characterization of Lignin Phenolic Polymers from Date Seeds by Mild Alkaline Solutions

Shadia Sirry

<sup>1</sup>Chemistry Department, Faculty of Science, Taibah University, P.O. Box 30002, Saudi Arabia  
E-Mail: smrry90@hotmail.com;  
Tel.: +966502008141

Received: 27/02/2019

/Accepted: 27/07/2019

**Abstract:** Date seeds are abundant lignocellulosic waste materials contain lignin phenolic polymers that act as antioxidants in food, cosmetic and pharmaceutical industries. Lignin have been extracted by several methods including alkaline medium. In the present work, optimum alkaline conditions for extraction of and de-polymerization of lignin from date seeds into low molecular weight phenolic compounds were studied. Date seeds surface were characterized before and after extraction by FTIR and thermal analysis (TGA and DTG). Total phenolic content, TPC and antioxidant capacity of extracts were evaluated spectrophotometrically. Maximum TPC was  $124 \pm 8.78 \mu\text{g GAE/mL}$  extract and optimum antioxidant capacity was determined as DPPH free radical inhibition ( $I = 76.56\%$ ). The optimum values of TPC and DPPH inhibition were attained at 0.005- 0.010M  $\text{Na}_2\text{CO}_3$ .

**Keywords:** extraction, antioxidant, DPPH, total phenolic, date seeds, TGA

### I. Introduction

Date seed, DS of fruit of *Phoenix dactylifera* L. is an abundant by-product material (about 7 million tones) and it is popular in the Middle East. DS are lignocellulosic compounds that composed of 23%, 20 and 55% of lignin,  $\alpha$  cellulose and hemicellulose, respectively [1] and it also contains antioxidants and phenolic compounds [2]. Phenolic compounds exist as free or bind with cell wall component so their extraction depends on matrix components in plant [3].

Several workers have reported antioxidant capacity and total Phenolic content of several varieties of DS from different countries after extractions by methanol, ethanol, water, acetone, formic and dimethyl sulfoxide [4-8]. Alkaline treatment of biomass cause of solubility of hemicellulose and lignin parts [9].

Lignin is a biopolymer of polyphenolic contain p-coumaryl, coniferyl, and sinapyl alcohol, which linked together through carbon-carbon or ether bonds. Lignin have been characterized by Fourier Transform infrared, FTIR and thermogravimetric, TGA analysis [10-11]. Lignin's phenolic compounds were known as natural antioxidant compounds [12] to scavenge free radicals and protect humans from oxidation damage. They have been used in food, cosmetic and pharmaceutical industries [13]. The antioxidant potential of lignin depends on the type of lignocellulose material and extraction solvents [14]. Organic, aqueous acidic or alkaline solvents have accomplished for extraction of lignin. Acidic extraction was reported as less effective to bound phenolic compounds and alkaline extraction was recommended [15,16]. Alkaline extraction of lignin from cell wall (delignification) is as a result of

hydrolysis of both ester and ether bonds of lignin and hemicellulose and converting into high amount of lower molecular mass of phenolic compound [17, 18]. Delignification of lignocellulose in alkaline medium have been reported by several investigators [19-23] Alkaline extraction were carried out by hydroxide ion or carbonate ion however strongly alkaline solution may lead to decrease of lignin extraction [24] and lignin extracted in carbonate more than in hydroxide ion [17]. Some researchers reported that phenolic compounds might be converted into non-oxidative quinones in strong alkaline medium [24-25]. The ability of extracted phenolic compounds as free radical scavenger has been characterized using diphenyl picrylhydrazyl (DPPH) method

This work was aimed to study the effect of mild alkaline carbonate concentration on extraction of antioxidant and total phenolic content of extracted date stone as well as characterization of DS surface after extraction by analyzing residual solid DS by FTIR and TGA methods. Antioxidant capacity was deduced by diphenylpicrylhydrazyl (DPPH) radical scavenging method however, total phenolic content, TPC was determined by Folin–Ciocalteu reagent.

## II. Experimental Section

### II.1. Materials

Date stone, DS was separated from date fruits (Sokary type), washed with distilled water, grounded and sieved to 814micron. Sodium carbonate, 2, 2-diphenyl-1-picrylhydrazyl (DPPH), gallic acid, Folin–Ciocalteu reagent and methanol were obtained from Sigma-Aldrich or chemical pure company.

### II.2. Extraction and characterization method

Six different concentrations of  $\text{Na}_2\text{CO}_3$  solutions ( $1 \times 10^{-4}$ -0.05M) were prepared and shaken with 5 gram of DS for four hours in water bath shaker (JSSB-30T) at 150 rpm then filtered. The modified stones were characterized by Fourier Transforms infrared (FTIR-8400S SHIMADZU) and thermogravimetric (SDT 600). Filtrates were analyzed for antioxidant potential by DPPH radical scavenging and total phenolic content (TPC) methods.

### II.3. Free Radical Scavenging Method

DPPH radical scavenging assay was utilized for determination of antioxidant potential by following the inhibition of strong absorption band at 517 nm by UV-Visible spectrophotometer (Thermo Scientific™ GENESYS™ 10S)[26]. 2mL of DPPH (0.2mM in methanol) was added to 0.1 ml of each DS extract solution and incubated in dark for 15 min. The absorbance was recorded at 517nm and the percentage of inhibition of DPPH radical activity, % I was calculated according to the following equation:

$$\%I = \frac{(A_o - A_1) \times 100}{A_o} \quad (1)$$

Where  $A_o$  is absorbance of control,  $A_1$  is the absorbance of control plus extract.

### II.4. Measurement of the total phenolic contents

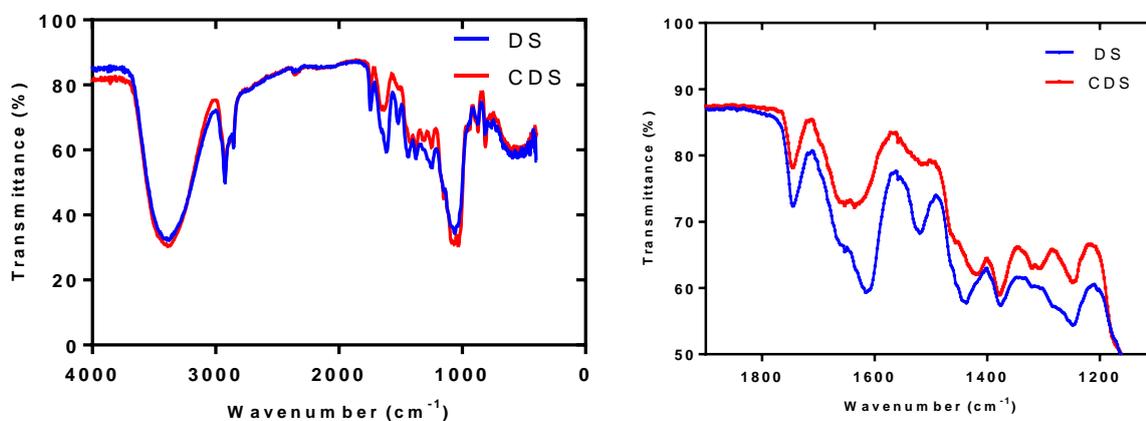
The amount of total phenolic compounds, TPC of CDS extracts were determined using Folin–Ciocalteu reagent, FC with respect to gallic acid standard [27]. FC reagent react with TPC in alkaline medium to give blue color that proportional with the amount of TPC. In 10 mL volumetric flask 2mLof extract, 2.5 mL of diluted FC reagent ( 10%) and 2mL of  $\text{Na}_2\text{CO}_3$  were mixed were kept in water bath at 50°C for 5minutes. The mixture was analyzed spectrophotometrically at 760nm and TFC was calculated as gallic acid equivalent, GAE by comparing with standard curve of gallic acid.

## III. Results and Discussion

DS was extracted with variable concentration of  $\text{Na}_2\text{CO}_3$  solutions and the remaining solid DS were characterized with and TGA.

### III.1. FTIR characterization

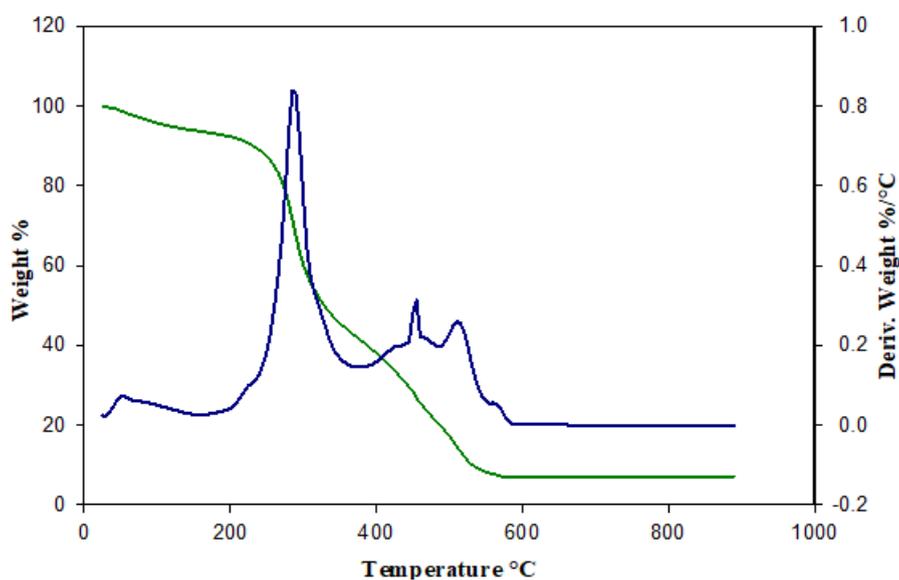
FTIR spectrum of raw DS and CDS (Fig.1) show characteristic bands of lignin aromatic skeletal  $1600$ ,  $1491$ ,  $1525$ , and  $1446 \text{ cm}^{-1}$  [28-31] and ether bonds  $1246$  and  $1033 \text{ cm}^{-1}$ . In CDS, bands have lower intensity which indicates the extraction of lignin.



**Figure 1.** IR characterization of DS and CDS

### III.2. TGA characterization

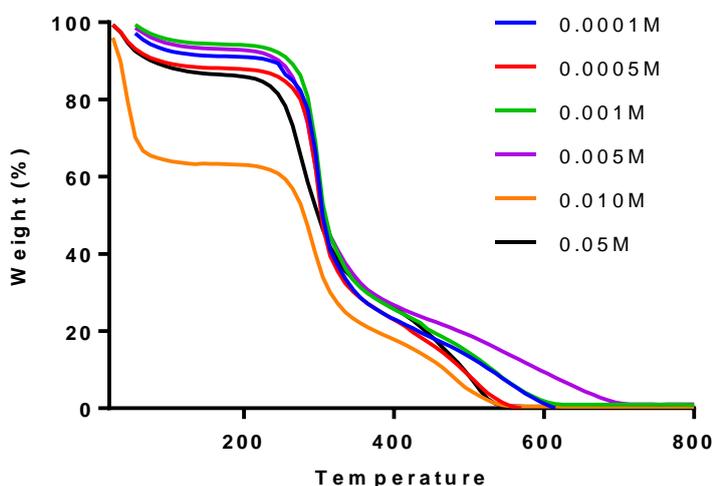
Thermogravimetric analysis for raw and modified of DS or carbonate modified DS (CDS) were performed at a heating rate of  $10 \text{ }^\circ\text{C}$  per minute. The dose of around  $2.5$ -  $4.70 \text{ mg}$  of each were heated at  $25$  up to  $900 \text{ }^\circ\text{C}$



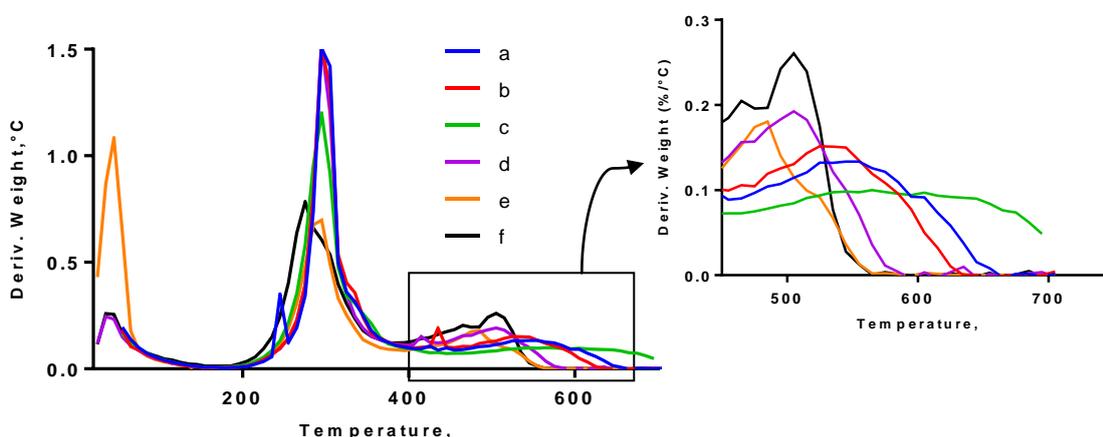
**Fig.2.** TGA and DTG analysis of raw DS

In Fig.2 there are three characteristic TGA&DTG peaks of untreated DS at 275,465 and 505°C representing hemicellulose, cellulose and lignin peaks, respectively. Delaying of degradation of lignin is due to cleavage of interunit linkage of polymer structure producing phenolic monomers then decomposing of aromatic ring of lignin above 500 °C [32]. The initial mass loss which started at 50°C correspond to water and volatile components and the second mass loss was started at 200°C to 300°C corresponding to hemicellulose degradation\*. The third mass loss was due to lignin degradation at 490-550 °C.

In case of CDS samples the position of DTG peaks of lignin (Fig.3) were varied from DS according to stabilization of lignin and re-dissociation again on varying  $\text{Na}_2\text{CO}_3$  concentration [33].



**Fig.3** TGA analysis of CDS at variable concentrations of  $\text{Na}_2\text{CO}_3$



**Fig4.** Comparison of DTG of CDS samples at variable  $\text{Na}_2\text{CO}_3$  concentration

Increasing alkaline concentration shift DTG band to lower temperature except of  $\text{Na}_2\text{CO}_3$  0.001M concentration, which indicates the increasing solubility of lignin, by increasing  $\text{Na}_2\text{CO}_3$  concentrations. Maximum extraction of lignin may be at 0.01M of  $\text{Na}_2\text{CO}_3$  because it show DTG peak at lowest temperature and lower mass loss in TGA. Utilizing of 0.001M  $\text{Na}_2\text{CO}_3$  for DS extraction may increase stability of DS's lignin and retard its extraction. This may be due to formation of larger macromolecule [31]

### III.3. Antioxidant capacity

Antioxidant capacity of CDS extracts were estimated by DPPH free radical scavenging capacity (table 1)

**Table 1. Effect of  $[Na_2CO_3]$  on total antioxidant capacities, % I of CDS extracts**

| $[Na_2CO_3], M$          | 0.0001 | 0.0005 | 0.0010 | 0.0050 | 0.0100 | 0.0500 |
|--------------------------|--------|--------|--------|--------|--------|--------|
| %I                       | 5.47   | 10.45  | 8.24   | 76.56  | 72.91  | 8.13   |
| $\pm$ Standard Deviation | 1.11   | 0.64   | 0.83   | 0.63   | 1.03   | 1.43   |

Generally, at higher concentration of  $Na_2CO_3$  the antioxidant capacity of CDS extract increase by increasing carbonate concentration lead to hydrolysis of lignocellulosic ester bond, which enhance lignin extraction. Lignin extract are contain phenolic hydroxyl and ortho- methoxy substituted aromatic ring that responsible of lignin antioxidants. In case of utilizing 0.001M of  $Na_2CO_3$  the antioxidant capacity of CDS, extracts slightly decrease. This may be due to formation of other stable lignin type which concordant with DTG results. Largest antioxidant capacities ( $I \cong 77-73\%$ ) were at 0.005 and 0.01M carbonate extracts, which correspond to pH 9.3 and 9.8. However, utilizing 0.05M concentration of  $Na_2CO_3$  lead to diminishing antioxidant capacity from 77% to 9% due to the increase of extraction of hemicellulose, which suppress antioxidant capacity. Hemicellulose are carbohydrates which have ability to linked to phenolic groups by hydrogen bonding and interfere in antioxidant properties [34, 35]

#### III.4. Total phenolic content

In alkaline medium Lignin are hydrolyzed to monomer lower molecular weight phenolic compounds. Phenolic compounds have been considered as antioxidants by free radical scavenging and metal chelating [36-37]. TPC was determined as  $\mu g$  gallic acid equivalent (GAE) per mL of extract from calibration curve of standard gallic acid which have an equation  $y = 0.0091x + 0.0135$  and correlation factor  $R^2 = 0.998$ . Total phenolic content of CDS extract increase linearly with increasing carbonate concentration and reach maximum value at 0.01M thereafter it decrease (table 2). Increasing of TPC are due to de-polymerization of lignin and formation of monomer phenolic compounds however, decreasing of TPC at 0.05M  $Na_2CO_3$  are due to re-polymerization of highly reactive phenolic compounds [38, 23].

**Table 2. Total phenolic content of CDS extract at variable concentration of  $Na_2CO_3$**

| $[Na_2CO_3], M$          | 0.0001 | 0.0005 | 0.0010 | 0.0050  | 0.0100  | 0.0500 |
|--------------------------|--------|--------|--------|---------|---------|--------|
| $\mu g$ GAE/mL           | 63.556 | 50.185 | 72.222 | 106.370 | 124.040 | 83.889 |
| $\pm$ Standard Deviation | 1.56   | 1.50   | 4.12   | 1.74    | 8.78    | 0.44   |

From above results, it is noticed that the optimum alkaline condition for lignin total antioxidant extraction of DS are  $Na_2CO_3$  concentration are 0.05-0.01M which give lowest temperature of lignin DTG band and highest antioxidant capacity, %I and TPC.

#### IV. Conclusion

Mild alkaline carbonate are optimum condition for extraction of maximum amount of lignin antioxidants of abundantly by-product date seeds which is important in cosmetic and pharmaceutical industrials.

#### V. References

- [1] Briones, Rodrigo, et al. "Polyol production by chemical modification of date seeds." *Industrial Crops and Products* 34.1 (2011): 1035-1040.
- [2] Al-Farsi, Mohamed Ali, and Chang Yong Lee. "Optimization of phenolics and dietary fibre extraction from date seeds." *Food Chemistry* 108.3 (2008): 977-985.
- [3] González, Mónica, and Venerando González. "Sample preparation of tropical and subtropical fruit biowastes to determine antioxidant phytochemicals." *Analytical Methods* 2.12 (2010): 1842-1866.
- [4] Baliga, Manjeshwar Shrinath, et al. "A review of the chemistry and pharmacology of the date fruits (*Phoenix dactylifera* L.)." *Food research international* 44.7 (2011): 1812-1822.
- [5] Ardekani, Mohammad Reza Shams, et al. "Comparison of antioxidant activity and total phenol contents of some date seed varieties from Iran." *Iranian journal of pharmaceutical research: IJPR* 9.2 (2010): 141.
- [6] Ammar, Abdalla SM, and Ramadan AA Habiba. "Phenolic content and antioxidant activity of date seeds." *J Agric Vet Sci Qassim Univ* 3 (2010): 3-8.
- [7] Khan, Shah Alam, et al. "In vitro inhibitory effects on  $\alpha$ -glucosidase and  $\alpha$ -amylase level and antioxidant potential of seeds of *Phoenix dactylifera* L." *Asian Pacific Journal of Tropical Biomedicine* 6.4 (2016): 322-329.
- [8] Alem, Chakib, et al. "Phytochemical compositions and antioxidant capacity of three date (*Phoenix dactylifera* L.) seeds varieties grown in the South East Morocco." *Journal of the Saudi Society of Agricultural Sciences* 16.4 (2017): 350-357.
- [9] Sun, Shaolong, et al. "One-step process based on the order of hydrothermal and alkaline treatment for producing lignin with high yield and antioxidant activity." *Industrial Crops and Products* 119 (2018): 260-266.
- [10] Yu, Jie, et al. "Cellulose, xylan and lignin interactions during pyrolysis of lignocellulosic biomass." *Fuel* 191 (2017): 140-149.
- [11] Erdtman, Holger. "Lignins: Occurrence, formation, structure and reactions, KV Sarkanen and CH Ludwig, Eds., John Wiley & Sons, Inc., New York, 1971. 916 pp. \$35.00." *Journal of Polymer Science Part B: Polymer Letters* 10.3 (1972): 228-230.
- [12] Mahmood, Zahed, et al. "Lignin as Natural Antioxidant Capacity." *Lignin-Trends and Applications*. InTech, 2018.
- [13] Meliana, Y., and A. H. Setiawan. "Antioxidant activity of lignin phenolic compounds as by-product of pretreatment process of bioethanol production from empty fruits palm bunch." *AIP Conference Proceedings*. Vol. 1712. No. 1. AIP Publishing, 2016.
- [14] Ugartondo, Vanessa, Montserrat Mitjans, and María Pilar Vinardell. "Comparative antioxidant and cytotoxic effects of lignins from different sources." *Bioresource technology* 99.14 (2008): 6683-6687.
- [15] Oreopoulou, A., Papavassilopoulou, E., Bardouki, H., Vamvakias, M., Bimpilas, A., & Oreopoulou, V. (2018). Antioxidant recovery from hydrodistillation residues of selected Lamiaceae species by alkaline extraction. *Journal of applied research on medicinal and aromatic plants*, 8, 83-89.
- [16] Boussetta, Nadia, et al. "Valorization of oilseed residues: extraction of polyphenols from flaxseed hulls by pulsed electric fields." *Industrial Crops and Products* 52 (2014): 347-353.
- [17] Durot, Nathalie, François Gaudard, and Bernard Kurek. "The unmasking of lignin structures in wheat straw by alkali." *Phytochemistry* 63.5 (2003): 617-623.
- [18] García, Juan C., et al. "Search for optimum conditions of wheat straw hemicelluloses cold alkaline extraction process." *Biochemical engineering journal* 71 (2013): 127-133.
- [19] Gupta, Rajesh, and Y. Y. Lee. "Investigation of biomass degradation mechanism in pretreatment of switchgrass by aqueous ammonia and sodium hydroxide." *Bioresource technology* 101.21 (2010): 8185-8191.
- [20] Jiang, Bo, et al. "Structural elucidation and antioxidant activity of lignin isolated from rice straw and alkali-oxygen black liquor." *International journal of biological macromolecules* 116 (2018): 513-519.
- [21] Sun, Shaolong, et al. "One-step process based on the order of hydrothermal and alkaline treatment for producing lignin with high yield and antioxidant activity." *Industrial Crops and Products* 119 (2018): 260-266.
- [22] Li, Zhili, and Yuanyuan Ge. "Antioxidant activities of lignin extracted from sugarcane bagasse via different chemical procedures." *International journal of biological macromolecules* 51.5 (2012): 1116-1120.
- [23] Fernández-Rodríguez, Javier, et al. "Lignin depolymerization for phenolic monomers production by sustainable processes." *Journal of energy chemistry* 26.4 (2017): 622-631.
- [24] Yen, Gow-Chin, and Chien-Ya Hung. "Effects of alkaline and heat treatment on antioxidative activity and total phenolics of extracts from *Hsian-tsao* (*Mesona procumbens* Hemsl.)." *Food Research International* 33.6 (2000): 487-492.
- [25] Brooks, R. J., Bunton, C. A., & Hellyer, J. M. (1973). Oxidative hydrolysis of p-hydroxyphenyl phosphates. *The Journal of Organic Chemistry*, 38(12), 2151-2156.
- [26] Brand-Williams, Wendy, Marie-Elisabeth Cuvelier, and C. L. W. T. Berset. "Use of a free radical method to evaluate antioxidant activity." *LWT-Food science and Technology* 28.1 (1995): 25-30.

- [27] Singleton, Vernon L., Rudolf Orthofer, and Rosa M. Lamuela-Raventós. "[14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent." *Methods in enzymology*. Vol. 299. Academic press, 1999. 152-178.
- [28] Li, Zhili, and Yuanyuan Ge. "Antioxidant activities of lignin extracted from sugarcane bagasse via different chemical procedures." *International journal of biological macromolecules* 51.5 (2012): 1116-1120.
- [29] Tejado, Ayo, et al. "Physico-chemical characterization of lignins from different sources for use in phenol-formaldehyde resin synthesis." *Bioresource Technology* 98.8 (2007): 1655-1663.
- [30] Alriols, M. González, et al. "Agricultural palm oil tree residues as raw material for cellulose, lignin and hemicelluloses production by ethylene glycol pulping process." *Chemical Engineering Journal* 148.1 (2009): 106-114. García, Araceli, et al. "Study of the antioxidant capacity of Miscanthus sinensis lignins." *Process Biochemistry* 45.6 (2010): 935-940.
- [31] García, Araceli, et al. "Study of the antioxidant capacity of Miscanthus sinensis lignins." *Process Biochemistry* 45.6 (2010): 935-940.
- [32] Norgren, Magnus, and Håkan Edlund. "Lignin: Recent advances and emerging applications." *Current Opinion in Colloid & Interface Science* 19.5 (2014): 409-416.
- [33] Ndazi, Bwire Sturmius, Christian W. Nyahumwa, and Joseph Tesha. "Chemical and thermal stability of rice husks against alkali treatment." *BioResources* 3.4 (2008): 1267-1277.
- [34] Dizhbite, Tatiana, et al. "Characterization of the radical scavenging activity of lignins—natural antioxidants." *Bioresource Technology* 95.3 (2004): 309-317.
- [35] Scalbert, Augustin, et al. "Dietary polyphenols and the prevention of diseases." *Critical reviews in food science and nutrition* 45.4 (2005): 287-306.
- [36] Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10), 7313-7352.
- [37] Barman, D. N., Haque, M. A., Kang, T. H., Kim, M. K., Kim, J., Kim, H., & Yun, H. D. (2012). Alkali pretreatment of wheat straw (*Triticum aestivum*) at boiling temperature for producing a bioethanol precursor. *Bioscience, biotechnology, and biochemistry*, 76(12), 2201-2207.
- [38] Roberts, V. M., Stein, V., Reiner, T., Lemonidou, A., Li, X., & Lercher, J. A. (2011). Towards quantitative catalytic lignin depolymerization. *Chemistry—A European Journal*, 17(21), 5939-5948.