

# A clone resource of *Magnaporthe oryzae* effectors that share sequence and structural similarities across host-specific lineages

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**We describe a clone resource of 195 effectors of the blast fungus *Magnaporthe oryzae*. These clones are freely available as Golden Gate compatible entry plasmids. Our aim is to provide the community with an open source effector clone library to be used in a variety of functional studies.**

## Introduction

Plant pathogens secrete effectors that play central roles in subjugating plants for colonization. Effectors typically have signal peptides, and occasionally carry conserved folds and motifs (Lo Presti *et al.*, 2015; Franceschetti *et al.*, 2017). *Magnaporthe oryzae* (Syn. *Pyricularia oryzae*) is an important plant pathogen that is able to infect around 50 species of both wild and cultivated grasses including important cereals of the Poaceae family. *M. oryzae* is mostly known to cause rice blast but can also cause disease on other crops such as barley, wheat, foxtail millet, and finger millet. The global population of *Magnaporthe* is composed of genetically differentiated lineages which, in some cases, still exhibit a measurable degree of gene flow (Gladieux *et al.*, 2018). Fungal isolates from each of those lineages show a preference for a specific host and also encode distinct repertoires of effector genes (Yoshida *et al.*, 2016).

The first genomic sequence of *Magnaporthe oryzae* was released in 2005 for the lab strain 70-15 and allowed to predict a large set of secreted proteins such as enzymes involved in secondary metabolism and virulence-associated factors including putative effectors (Dean *et al.*, 2005). Recently an increasing number of genome sequences of isolates from different lineages have become available, allowing the research community to perform comparative genomic studies (Chiapello *et al.*, 2015; Yoshida *et al.*, 2016).

Many of the validated effectors of *M. oryzae* are known as the MAX (M*agnaporthe* AVRs and ToxB like) effectors. These effectors, while showing little primary sequence similarity, share a conserved structural fold composed of 6  $\beta$ -sheets alternating in an anti-parallel manner (de Guillen *et al.*, 2015). The MAX family has been largely expanded in *Magnaporthe* as those effectors account for 5-10% of the effector repertoire and for 50% of the already cloned effectors of *Magnaporthe* (de Guillen *et al.*, 2015). Indeed, the identification of structural motifs enables more sensitive predictions of effectors from pathogen genomes compared to sequence similarity searches (Franceschetti *et al.*, 2017).

The aim of this project was to computationally identify a set of *M. oryzae* effectors from the main host-specific lineages and develop an open access clone resource for functional analyses.

## Results and Discussion

**Bioinformatics.** Here, we describe a bioresource of 195 predicted *M. oryzae* effectors that we assembled from available genome sequences. To develop this resource, we initially analysed secretomes of *M. oryzae* isolates infecting different hosts such as rice, wheat, finger millet, foxtail millet, oat and some species of the *Digitaria* genus using the following computational pipeline. First, we identified proteins showing sequence similarity to known *M. oryzae* effectors with an avirulence activity (i.e. detected by plant immune receptors), using BLAST and a library of 21 previously characterized effectors. Second, we performed a search for remote relationships using a Hidden Markov Model (HMM) trained with multiple, structure-based alignments of *M. oryzae* MAX effectors (de Guillen *et al.*, 2015) using the HMMER program (Finn *et al.*, 2011). Next, we grouped candidates obtained by both approaches to form a non-redundant list of putative effectors and functionally annotated them using two methods. The first is a classical Gene Ontology (GO) search using the Blast2GO program (Conesa *et al.*, 2005). The second is a BLASTP similarity search against a custom-made database (the Darwin database) which contains more than 2,600,000 predicted proteins of 137 different eukaryotic organisms. We removed candidates with functional domain annotation as most *M. oryzae* effectors are proteins with no predicted function. Finally, we also removed candidates with poor gene model predictions and very low sequence similarity to *M. oryzae* effectors. The combination of these approaches resulted in 194 effector candidates that we selected for gene synthesis as a clone resource. We also added a homolog of the well-studied *M. oryzae* effector AVR-Pik of the *Lolium perenne* isolate PGKY to this library.

**Golden Gate system.** The Golden Gate cloning system enables rapid and high throughput assembly of multiple sequence modules, such as promotor, terminator or tags into a common vector (Patron *et al.*, 2015). This cloning strategy is ideal for wide dissemination of cloning material by using a universal code for cloning. We synthesized the 195 *M. oryzae* effectors in a Golden Gate compatible fashion to enable the transfer of these genes into a variety of vectors for various applications such as yeast two-hybrid assays, heterologous protein expression, and fungal transformation (see for example the fungal transformation vectors described by Pennington *et al.*, 2017, 2018).

**Effector clone resource.** We synthesized the coding sequences corresponding to the mature proteins (without the signal peptide) of 195 candidates in the Golden Gate compatible vector pUC57-Kan. Each coding sequence was flanked by *BsaI* restriction enzyme sites and relevant overhang sequences to ensure optimal Golden Gate reactions. The coding sequences were manually codon-optimised for expression in *Saccharomyces cerevisiae*, *Nicotiana benthamiana* and *Escherichia coli* (when it was not possible to optimise for all three organisms, priority was given to *S. cerevisiae* and *N. benthamiana*). Table S1 describes the 195 candidates and includes information such as sequence similarity to *M. oryzae* effectors, results of the HMM search, SignalP scores, functional annotation and host specificity of the strain in which the effector candidate was identified. Table 1 summarizes the different candidates identified, their host specificity and the rationale behind their selection whether it was based on sequence or structural similarities. Fig 1 shows an overview of the Golden Gate system and a generic schematic representation of the pUC57-Kan vector insertion site. Additional supplementary files include the DNA and amino acid sequences of the selected genes.

**Clone distribution.** Level 0 modules are available via Addgene (addgene.com). Catalog numbers are provided in Table S1.



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## Supplementary material

**Table S1. Characteristics of the selected *Magnaporthe oryzae* candidate effectors.** Addgene catalog number: number under which each effector candidate can be found in the Addgene catalog. match\_blast\_AVR: *M. oryzae* effector against which the best blast hit is found. Evaluate\_hmm: evaluate found with the HMM search. score\_hmm: score found with the HMM search. Score\_signalP: score from signalP search (Petersen *et al.*, 2011). signalP\_HMM\_score: combined score from signalP and tmhmm searches (Krogh *et al.*, 2001). hit\_blast\_Darwin\_without\_magnaporthe: best blast hit against the Darwin database after removing the proteins from *Magnaporthe spp.* GO: Gene Ontology attributed by the Blast2GO program. Host: host of the isolate from which the gene sequence originates. Specificity: hosts of all the isolates that express this protein. Present\_in: other isolates in which this protein can be found. Validated\_effector: YES: this protein is an already-known *M. oryzae* effector.

**Supplementary file 1. Protein sequences of the selected *Magnaporthe oryzae* candidate effectors in FASTA format.**

**Supplementary file 2. Codon-optimised nucleotide sequences of the *Magnaporthe oryzae* candidate effectors (with overhang sequences) in FASTA format.**

**Supplementary folder 1. Nucleotide sequences of the *Magnaporthe oryzae* candidate effectors in the pUC57-Kan vector (Genbank format).**

**Supplementary folder 2. Plasmid maps.**