

1
2
3
4
5
6

7A rapid method for the chromatographic analysis of
8volatile organic compounds in exhaled breath of tobacco
9cigarette and electronic cigarette smokers

10

11Esther Marco and Joan O. Grimalt*

12Institute of Environmental Assessment and Water Research (IDÆA). Spanish Council
13for Scientific Research (CSIC). Jordi Girona, 18. 08034-Barcelona. Catalonia. Spain

14

15

16

17

18

19

20

21

22*Author for correspondence. Phone: +34934006118. Fax +34932045904. E-mail:

23joan.grimalt@idaea.csic.es

24

25 **Abstract**

26

27 A method for the rapid analysis of volatile organic compounds (VOCs) in smoke from
28 tobacco and electronic cigarettes and in exhaled breath of users of these smoking
29 systems has been developed. Both disposable and rechargeable e-cigarettes were
30 considered. Smoke or breath were collected in Bio-VOCs. VOCs were then desorbed in
31 Tenax cartridges which were subsequently analyzed by thermal desorption coupled to
32 gas chromatography-mass spectrometry. The method provides consistent results when
33 comparing the VOC compositions from cigarette smoke and the equivalent exhaled
34 breath of the smokers. The differences in composition of these two sample types are
35 useful to ascertain which compounds are retained in the respiratory system after tobacco
36 cigarette or e-cigarette smoking.

37 Strong differences were observed in the VOC composition of tobacco cigarette
38 smoke and exhaled breath when comparing with those of e-cigarette smoking. The
39 former involved transfers of a much larger burden of organic compounds into smokers,
40 including benzene, toluene, naphthalene and other pollutants of general concern. e-
41 Cigarettes led to strong absorptions of propylene glycol and glycerin in the users of
42 these systems. Tobacco cigarettes were also those showing highest concentration
43 differences between nicotine concentrations in smoke and exhaled breath. The results
44 from disposable e-cigarettes were very similar to those from rechargeable e-cigarettes.

45

461. Introduction

47

48 Electronic cigarettes (e-cigarettes) are designed to transfer mixtures of air and vapors
49 into the respiratory system [1-3]. They use plastic or metal cylinders that contain
50 electronic vaporization systems, a battery, in some cases, a charger, electronic controls
51 and, optionally, replaceable cartridges. Different humectants, e.g. propylene glycol or
52 glycerin, flavorings and nicotine at various concentrations are generally contained in the
53 cartridges. They can be disposable (Type 1 e-cigarette) or rechargeable (Type 2 e-
54 cigarette). Concern has been raised for the compounds incorporated into smokers as
55 consequence of e-cigarette vaping.

56 Exhaled breath, namely the alveolar breath [4], may provide significant clues on
57 the compounds that are retained in humans as consequence of this activity. Studies on
58 VOCs in exhaled breath from e-cigarette smokers have been developed using solid
59 phase microextraction inside a breath collection device [5] or exposure chambers which
60 are subsequently sampled by absorption into solid phase sorption tubes. These tubes are
61 then analyzed by desorption into gas chromatography coupled to mass spectrometry
62 (GC-MS) [6]. In other cases, the absorption cartridge has been installed at the outlet of a
63 smoking machine and the retained compounds are eluted with CS₂ and methanol for
64 subsequent analysis by GC-MS [7].

65 In the present study, we describe a simplified method using a Bio-VOCs exhaled
66 air sampler developed by the UK Health and Safety Laboratory (Markes International
67 Ltd, Llantrisant, UK) for the comparison of the smoke generated by Type 1 and Type 2
68 e-cigarettes, tobacco cigarettes and the exhaled breath after vaping or smoking. This
69 device has been used in the analysis of both exhaled alveolar air and mouth air [8-15].
70 Now, we are using BIO-VOCs for a rapid method of characterization of the volatile
71 organic constituents in tobacco cigarettes and e-cigarettes. Blend type American tobacco
72 cigarettes with filters (length 83 mm, length of filter 23 mm, diameter 8 mm) were used
73 as test examples. Cigarettes with low nicotine content (0.6 mg), low tar (8 mg) and low
74 carbon monoxide (9 mg) were chosen. The compounds analyzed in the present study
75 were mostly in the gas phase. The results add to the current knowledge of exposure of
76 smokers to organic compounds that so far have been mostly characterized in particulate
77 phase transfer processes [16-23].

78

79

802. Experimental section

812.1. Sampling cartridges

82

83 Volatile organic compounds were concentrated by sorption into stainless steel sorbent
84 cartridges (89 mm long 0.64 cm outer diameter) packed with 200 mg of Tenax TA 35/60
85 mesh (Markes International Ltd, Pontyclun, UK). The sorbent cartridges were
86 preconditioned using helium (5N grade; 100 ml/min) at 320°C for 2 hours and then at
87 335°C for 30 min. In later conditioning cycles these cartridges were reconditioned at
88 335°C for 20 minutes with the same flow carrier gas. Once cleaned, the cartridges were
89 sealed with brass Swagelock storage endcaps fitted with PTFE ferrules and stored in
90 solvent-free clean environments.

91

922.2. Sampling

93

94 Exhaled breath was sampled with a Bio-VOC system 30 min after tobacco cigarette or
95 e-cigarette smoking. To avoid metabolic differences all volunteers were asked to smoke
96 with the tobacco cigarettes and Type 1 and 2 e-cigarettes considered in this study.
97 People inspired and expired deeply three times, then retained the breath for 20 s and
98 blew into the Bio-VOC body through a disposable cardboard mouthpiece at their
99 highest capacity. The air remaining in the Bio-VOC was transferred into the sorbent
100 cartridge by pushing a screw-in plunger through the Bio-VOC body. This procedure was
101 repeated five times in each smoking test and all exhaled VOCs were accumulated in the
102 same cartridge. Thus, a total volume of 750 mL of exhaled breath was collected.

103 Tobacco cigarette and e-cigarette smoke were sampled by connecting the mouth
104 outlets to the Bio-VOC outlet. The screw-in plunger was used to pull smoke into the
105 Bio-VOC cylinder. Then, the tobacco cigarette or e-cigarettes were removed and the
106 cartridge was connected to the Bio-VOC outlet and the screw-in plunger was used to
107 push the smoke present in the Bio-VOC into the cartridge which sorbed the VOCs from
108 the sample. The sampled volume with this procedure was 150 mL.

109 Indoor ambient air was also sampled for comparison using this device. The
110 procedure was the same as that used for tobacco cigarette and e-cigarette smoke but
111 without connecting any of those devices to the sorbent cartridge. In this case the
112 procedure was repeated four times and a total volume of 600 mL was collected.

113

1142.3. Transfer of the VOC into the GC-MS

115

116 VOCs trapped in the sorbent cartridges were transferred with helium (5N grade; no inlet
117 split flow) to a thermal desorption (TD) instrument equipped with a Unity Series 2
118 Thermal Desorber and an Ultra 50:50 Multi-tube Auto-sampler (Markes International
119 Ltd). The compounds were desorbed from the cartridges at 300°C for 5 min (desorption
120 flow 40 mL/min) and re-concentrated in a graphitized carbon sorbent cold trap (U-
121 T11GPC-2S for General Purpose; Markes International Ltd) cooled at -20°C. This cold
122 trap was heated to 300°C over 5 min while passing a helium flow of 7.5 ml/min (split
123 flow 6 ml/min) for VOC transfer to an uncoated and deactivated fused-silica capillary
124 transfer line of 1 m length (internal and outer diameters 0.25 and 0.35 mm, respectively)
125 heated at 200°C. Total split ratio was 5:1.

126 For the Type 2 e-cigarette analyses, inlet split flow during cartridge desorption
127 was 50 mL/min and desorption trap conditions operated at a carrier helium flow of 28.5
128 mL/min and an outlet split flow of 27 mL/min. Total split ratio was 95:1.

129

1302.4. GC-MS operational conditions

131

132 The transfer line introduced the compounds into a Gas Chromatograph 7890 (GC;
133 Agilent Technologies Inc., Santa Clara, CA) coupled to a Mass Spectrometer 5975C
134 Inert XL MSD. The GC was equipped with a DB-5MS UI capillary column (length 60
135 m; internal diameter 0.32 mm; film thickness 1 μm; Agilent J&W GC Columns). Helium
136 (5N grade) was the carrier gas at a flow of 1.5 ml/min (constant flow mode). The GC
137 oven temperature program started at 40°C (holding time 10 min) then it increased to
138 150°C at 5°C/min and to 210°C at 15°C/min (final holding time 10 min).

139 A transfer line heated to 280°C carried the compounds from the GC to the MS.
140 The MS source and quadrupole temperatures were 230°C and 150°C, respectively. The
141 MS operated in electron impact mode. The detector was full scanned between 30-380
142 amu.

143

1442.5. Compound identification and quantification

145

146 VOCs were identified based on retention times and library identification of the mass
147 spectrum from each chromatographic peak (NIST2009, Mass Spectral Search Program,
148 version 2.0f). Quantification was performed by the external standard method.

149 Calibration curves encompassed nine calibration solutions in methanol (Merck
150 KGaA, Darmstadt, Germany) at different concentration in the range between 0.5 and
151 200 µg/ml. They were prepared from commercial solutions: UST Modified Gasoline
152 Range Organics (1000 µg/ml in methanol; Supelco, Inc. Bellefonte, PA, USA), FIA
153 Paraffin Standard (Accustandard Inc., New Haven, CT), and the individual standards: 2-
154 methylbutane, 1-pentene, cis-2-pentene, trans-2-pentene and 4-methyl-1-pentene, all
155 grade GC Standard (Sigma-Aldrich Co., St. Louis, Mo).

156 A Calibration Solution Loading Ring (CSLR™, Markes International Ltd.,
157 Llantrisant, UK) was used to introduce the calibration solution into clean sorbent
158 cartridges which allowed controlled vaporization and purging of the solvent (carrier gas
159 flow at 50 ml/min during 3 min). The different standard solutions were directly
160 introduced into the cartridges which were subsequently analyzed in the TD-GC-MS.
161 This allowed the determination of linear concentration ranges and limits of detection.
162 Recoveries were determined by introduction of standard solutions into the Bio-VOCs
163 heated at 50°C. Repetitiveness was also determined by sequential analysis of standards
164 introduced into the Bio-VOCs.

165

166

1673. Results and discussion

1683.1. Exhaled breath and air concentrations.

169

170 The gas chromatograms corresponding to indoor air from a building of Barcelona and
171 exhaled breath of volunteers present in this indoor environment without smoking are
172 compared in Fig. 1. Compound identification is reported in Table 1. Acetone and
173 isoprene were the main compounds in exhaled breath. These are two endogenous
174 compounds usually present in this type of sample. Both chromatograms also had some
175 common peaks such as benzene, toluene, styrene, benzaldehyde, δ-limonene, decanal,
176 nonanoic acid, and a siloxane series. Benzene and toluene may constitute trace amounts
177 of vehicular exhaust in the area. The siloxane series may represent some background
178 input of the analytical system. The other compounds may reflect a relationship between
179 in-door atmospheric VOCs and exhaled breath of residents in this environment.

1813.2. Smoke from tobacco cigarettes and e-cigarettes

183 Representative chromatograms of the VOC in the smoke composition of tobacco
184 cigarettes and Type 1 and Type 2 e-cigarettes are shown in Fig. 2. As expected a strong
185 contrast was observed between tobacco cigarette and e-cigarette smoke. The former
186 contained a wealth of compounds including nicotine and related products such as
187 nicotyrine, 7-methyl-1H-indole, myosmine, isonicotine. The occurrence of myosmine,
188 isonicotine and nicotyrine together with nicotine in tobacco cigarette smoke has been
189 reported in previous studies [24, 25]. 2,5-dimethylfuran is another compound
190 characteristic of tobacco cigarette smoke that has been proposed as a specific marker
191 [25-29]. In the present study, this compound was present in the chromatogram of the
192 tobacco cigarette smoke and absent in those of the e-cigarette smoke (Fig. 2; Table 1).

193 Besides these specific compounds several aromatic compounds such as benzene,
194 toluene, xylenes, ethylbenzene and styrene were also found in the chromatogram of
195 tobacco cigarette smoke (Fig.2). These compounds are not specific for tobacco cigarette
196 smoke, as several of them are found in the BTEX mixtures associated to traffic
197 emissions. However, as documented elsewhere [25-27, 30-31], benzene, a known
198 carcinogen, is common in tobacco cigarette smoke. In this respect, the relative
199 proportion of benzene and toluene in the samples described in this study, 44% and 56%,
200 respectively, is in agreement with the relative proportion of these compounds measured
201 in other tobacco smoke cigarettes measured with other sampling methods, 43% and
202 57%, respectively [31].

203 Other compounds commonly related with traffic emissions were also present in
204 the tobacco cigarette smoke chromatogram, e.g. n-heptane, n-octane, 1-ethyl-2-
205 methylbenzene, 1-ethyl-3-methylbenzene and naphthalene. The occurrence of these
206 compounds in tobacco cigarette smoke has also been reported [25, 27].

207 In addition to these VOCs, many polar compounds were also represented in the
208 tobacco cigarette smoke chromatogram, e.g. ethanol, acetone, acetic acid, butane-2,3-
209 dione, methyl ethyl ketone, methylfuran, isovaleraldehyde, pyridine, methylpyridine,
210 benzaldehyde, phenol, benzonitrile, acetophenone. These compounds have also been
211 found in tobacco cigarette smoke in previous studies [7, 25-26, 30, 32]. Some aldehydes
212 such as crotonaldehyde are also identified with this method. This compound has also

213been found in tobacco cigarette smoke in analyses using the dinitrophenylhydrazine
214method [33].

215 Chromatographic peaks for several unsaturated compounds were also found in
216the tobacco cigarette smoke sample, such as buta-1,3-diene, isoprene, hex-1-ene, hep-1-
217ene and δ -limonene. Several of them are known natural products that can also be found
218in many plant species. Their presence in tobacco cigarette smoke is consistent with
219previous studies [7, 25, 27, 30].

220 The analytical approach of the present study has been designed for the
221identification and quantification of the volatile compounds. However, some compounds
222found in the present study (Table 1) have also been identified in the particulate phase in
223analytical methods specifically designed for the compounds present in this phase, e.g.
224acetic acid, crotonaldehyde, n-heptane, phenol, δ -limonene, benzoic acid, hydroquinone,
225nicotine, 7-methyl-1H-indole, myosmine and nicotine [23]. These compounds are
226generally polar and formed by pyrolysis or distillation of the tobacco components under
227the high temperature conditions of smoking. Condensation processes lead to their
228distribution between the gas and particulate phases.

229 In contrast, the smoke of the e-cigarettes was mainly composed of propylene
230glycol and glycerin which is consistent with the product description of the
231manufacturers (note that the chromatographic peaks are overloaded). In addition the
232smoke of the e-cigarettes contained nicotine and related products such as miosmine and
233nicotyrine. The smoke of Type 2 e-cigarette also contained vanillin and ethyl vanillin
234which were likely added as a flavor.

235

2363.3 Exhaled breath from tobacco cigarette and e-cigarette users

237

238Representative chromatograms of the VOCs in the exhaled breath of tobacco cigarette
239and Type 1 and Type 2 e-cigarette users are shown in Fig. 2. The chromatogram of
240exhaled breath of a tobacco cigarette smoker showed a simplified mixture of the
241compounds found in the previously described smoke of these cigarettes (Figure 2)
242indicating that most of the original smoke components were retained in the lungs. Thus,
243the relative intensity of most of the higher molecular weight VOCs, those of higher
244chromatographic retention time, decreased significantly. However, some compounds
245that are specific of tobacco cigarette smoke such as nicotine, nicotyrine and 2,5-
246dimethylfuran were found in the exhaled breath. Their occurrence in the VOC

247composition can be used to indicate the exposure of the individuals to tobacco smoke
248compounds. Other VOCs such as benzene, toluene or δ -limonene were less specific of
249tobacco cigarette smoke but they still were dominant peaks in the exhaled breath
250chromatograms of the tobacco cigarette smokers. Isoprene was the most abundant
251exhaled breath peak. As mentioned above, this is an endogenous compound.

252 In the exhaled breath of the e-cigarette smokers the chromatographic peaks of
253propylene glycol and glycerin were absent indicating that they remained in the
254respiratory system of the smokers. Comparison of both original e-cigarette smoke and
255exhaled breath of the e-cigarette smokers also showed a strong decrease of the peaks
256corresponding to nicotine and related compounds. On the other hand, two main peaks in
257the chromatograms from exhaled breath were those corresponding to acetone and
258isoprene which likely represent endogenous sources. In addition, benzene, toluene and
2592,5-dimethylfuran were also found. These peaks were below limit of detection in the e-
260cigarette smoke vapors. Their occurrence in exhaled breath could reflect past exposures
261of the volunteers.

262

2633.4. Figures of merit

264

265Linear concentration ranges over three magnitudes of concentration were found for
266most compounds analyzed (Table 2). In some cases, e.g. n-hexane, naphthalene, these
267ranges were about 200. The limits of detection ranged between 0.05 and 0.65 ng. The
268transformation of these limits into concentration values ($\mu\text{g}/\text{m}^3$) must be done by
269reference to the sampled volume that depends on the number of Bio-VOC replicates
270(N). Thus, amount detection limit (ng) is equivalent to concentration detection limit
271($\mu\text{g}/\text{m}^3$) when multiplying the former by $1000/(150*N)$. The number of replicates in the
272analyses is indicated in section 2.2. The highest limits, e.g. toluene (0.65 ng), were due
273to background atmospheric levels by use of this compound in nearby labs. Repeatability
274(residual standard deviation of ten measurements) ranged between 5.9 and 23% which is
275consistent with previous measurements with Bio-VOCs in other studies [11]. Recoveries
276of standards introduced into the Bio-VOCs and analyzed as described in the
277experimental section ranged between 92 and 114% (Table 2).

278

2793.4. Quantitative differences

280

281The concentrations of some representative VOCs found in the tobacco cigarette and e-
282cigarette smoke and in the exhaled breath of the smokers are shown in Table 3.
283Concentrations of the same compounds in ambient indoor air and in volunteers
284breathing this air without smoking are shown for comparison. Tobacco cigarette smoke
285provided the samples containing highest concentrations of all compounds analyzed.
286Besides nicotine (1300 $\mu\text{g}/\text{m}^3$) it contained benzene, toluene, xylenes, ethylbenzene and
287naphthalene in high abundance (1100, 1400, 1500, 660 and 240 $\mu\text{g}/\text{m}^3$, respectively) as
288well as other compounds such as isoprene (2700 $\mu\text{g}/\text{m}^3$), pent-1-ene (700 $\mu\text{g}/\text{m}^3$), n-
289pentane (1200 $\mu\text{g}/\text{m}^3$), n-hexane (975 $\mu\text{g}/\text{m}^3$), n-heptane (1400 $\mu\text{g}/\text{m}^3$) and others. This
290composition was in strong contrast with that of smoke from the e-cigarettes in which all
291these compounds were virtually absent except nicotine (710-720 $\mu\text{g}/\text{m}^3$). Propylene
292glycol and glycerin were not found in the indoor air sample.

293 In principle, the compositions of exhaled breath reflected the differences of the
294cigarette smoke compositions (Table 3). Thus, tobacco cigarette smoke was the one with
295highest nicotine concentration and the highest content of this compound was found in
296the exhaled breath after tobacco cigarette smoking. In the cases shown in the present
297study, the differences in nicotine concentration between smoke and exhaled breath were
298highest for tobacco cigarettes, indicating that this was the smoking system with the
299highest nicotine transfer.

300 Isoprene is an endogenous compound and similar concentrations should be
301expected in all exhaled breath samples. However, it was found between 47-87 $\mu\text{g}/\text{m}^3$ in
302the e-cigarette smokers and 670 $\mu\text{g}/\text{m}^3$ in the tobacco cigarette smokers (Table 3). The
303high concentration of this compound in this volunteer may respond to a combination of
304non-absorbed compound from the tobacco cigarette smoke and generation of high yield
305of this compound after tobacco cigarette smoking. In fact, the exhaled breath of the
306tobacco cigarette smoker shows higher concentrations of all above mentioned
307compounds, including benzene, toluene, xylenes, ethylbenzene and naphthalene, than in
308the other exhaled breath samples.

309

310

3114. Conclusions

312

313The analysis of VOCs in smoke from tobacco cigarette and e-cigarettes and in the
314exhaled breath of users of these smoking systems can be performed by collection with

315Bio-VOC, absorption in Tenax cartridges and analysis by TD-GC-MS. This method
316provides consistent results when comparing the composition of VOCs in cigarette
317smoke and exhaled breath of the smokers. It also allows the discrimination between
318endogenous and exogenous compounds and compounds reflecting past exposures to
319pollutants or tobacco smoke. Comparison of the concentrations between smoke and
320equivalent exhaled breath of the smokers illustrated the incorporation of higher burdens
321of VOCs in the tobacco cigarette smokers than in the e-cigarette smokers.

322

323

324**Acknowledgements**

325

326We thank the Spanish Ministry of Economy and Competitiveness for the Expo-Cov
327(CTM2012-39298) fellowship. Financial support from the EU projects HEALS (FP7-
328ENV-2013- 603946) and CROME (LIFE12 ENV/GR/001040) is acknowledged.

329

330

331**References**

332

333[1] M. Williams, T. Talbot, Variability among electronic cigarettes in the pressure drop,
334 airflow rate and aerosol production, *Nicotine Tob. Res.* 13 (2011) 1276-1283.

335[2] J.-F. Etter, C. Bullin, Electronic cigarette: users profile, utilization, satisfaction and
336 perceived efficacy, *Addiction* 106 (2011) 2017-2018.

337[3] D. Cressey, Regulation stacks up for e-cigarettes, *Nature* 501 (2013) 473.

338[4] V. Vereb, A.M. Dietrich, B. Alfeeli, M. Agah, The possibilities will take your breath
339 away: breath analysis for assessing environmental exposure, *Environ. Sci. Technol.*
340 45 (2011) 8167-8175.

341[5] A.N. Martin, G.R. Farquac, A.D. Jones, M. Frank, Human breath analysis: method
342 for sample collection and reduction of localized background effects,. *Anal.*
343 *Bioanal. Chem.* 396 (2010) 739-750.

344[6] J. Czogala, M.L. Goniewicz, B. Fidelius, W. Zielinska-Danch, M.J. Travers, A.
345 Sobezak, Secondhand exposure to vapors from electronic cigarettes, *Nicotine Tob.*
346 *Res.* 16 (2014) 655-662.

347[7] S. Uchiyama, T. Tomizawa, Y. Inaba, N. Kunugita, Simultaneous determination of
348 volatile organic compounds and carbonyls in mainstream cigarette smoking using a

349 sorbent cartridge followed by two-step elution, *J. Chromatogr. A* 1314 (2013) 31-
350 37.

351[8] S. van den Velde, M. Quirynen, P. van Hee, D. van Steenberghe, Differences
352 between alveolar air and mouth air, *Anal. Chem.* 79 (2007) 3425-3429.

353[9] J.W.H. Biesterbos, G. Beckmann, R.B.M. Anzion, A.M.J. Ragas, F.G.M. Russel,
354 P.T.J. Scheepers, Sensitive method for quantification of
355 octamethylcyclotetrasiloxane (D4) and decamethylcyclopentasiloxane (D5) in end-
356 exhaled air by thermal desorption gas chromatography mass spectrometry, *Anal.*
357 *Chem.* 86 (2004) 5794-5799.

358[10] M.K. Das, S.C. Bishwal, A. Das, D. Dabral, A. Varshney, V.K. Badireddy, R.
359 Nanda, Investigation of gender-specific exhaled breath volatome in humans by
360 GCxGC-TOF-MS, *Anal. Chem.* 86 (2014) 1229-1237.

361[11] D. Dyne, J. Cocker, H.K. Wilson, A novel device for capturing breath samples for
362 solvent analysis, *Sci. total Environ.* 199 (1997) 83-89.

363[12] K. Jones, M. Meldrum, E. Baird, S. Cottrell, P. Kaur, N. Plant, D. Dyne, J. Cocker,
364 Biological monitoring for trimethylbenzene exposure: a human volunteer study and
365 a practical example in the workplace, *Ann. Occup. Hyg.* 50 (2006) 593-598.

366[13] A. Hryniuk, B.M. Ross, Detection of acetone and isoprene in human breath using a
367 combination of thermal desorption and selected ion flow tube mass spectrometry,
368 *Int. J. Mass Spectrom.* 285 (2009) 26-30.

369[14] J. Kwak, M. Fan, S.W. Harshman, C.E. Garrison, V.L. Dershem, J.B. Phillips, C.C.
370 Grigsby, D.K. Ott, Evaluation of Bio-VOC sampler for analysis of volatile organic
371 compounds in exhaled breath, *Metabolites* 4 (2014) 879-888.

372[15] P.T.J. Scheepers, J. Konings, G. Demirel, E.O. Gaga, R. Anzion, P.G.M. Peer, T.
373 Dogeroglu, S. Ornektekin, W. van Doorn, Determination of exposure to benzene,
374 toluene and xylenes in Turkish primary school children by analysis of breath and
375 by environmental passive sampling, *Sci. total Environ.* 408 (2010) 4863-4870.

376[16] J.J. McAughey, D.A. Knight, A. Black, C.J. Dickens, Environmental tobacco
377 smoke retention in humans from measurements of exhaled smoke compounds, *Inh.*
378 *Toxicol.* 6 (1994) 615-631.

379[17] R.R. Baker, M. Dixon, The retention of tobacco smoke constituents in the human
380 respiratory tract, *Inh. Toxicol.* 18 (2006) 255-294.

381[18] A.K. Armitage, M. Dixon, B.E. Frost, D.C. Mariner, N.M. Sinclair, The effect of
382 inhalation volume and breath-hold duration on the retention of nicotine and

383 solanesol in the human respiratory tract and on subsequent plasma nicotine
384 concentrations during cigarette smoking, *Beit. Tabak Int.* 21 (2004) 240-249.

385[19] S.C. Moldoveanu, W.M. Coleman III, J.M. Wilkins, Determination of polycyclic
386 aromatic hydrocarbons in exhaled cigarette smoke. *Beit. Tabak Int.* 23 (2008) 85-
387 97.

388[20] S.C. Moldoveanu, W.M. Coleman III, A pilot study to assess solanesol levels in
389 exhaled cigarette smoke, *Beit. Tabak Int.* 23 (2008) 144-152.

390[21] S.C. Moldoveanu, W.M. Coleman III, The influence of a humectant on the retention
391 by humans of solanesol from cigarette smoke (Part 1, propylene glycol), *Beit.*
392 *Tabak Int.* 23 (2008) 153-159.

393[22] S.C. Moldoveanu, W.M. Coleman III, The influence of a humectant on the retention
394 by humans of solanesol from cigarette smoke (Part 2, glycerin), *Beit. Tabak Int.* 23
395 (2009) 377-383.

396[23] S.C. Moldoveanu, F.K. St. Charles, Differences in the chemical composition of the
397 particulate phase of inhaled and exhaled cigarette mainstream smoke, *Beit. Tabak*
398 *Int.* 22 (2007) 290-302.

399[24] J. Cai, B. Liu, Q. Su, Fast analysis of nicotine related alkaloids in tobacco and
400 cigarette smoke by megabore capillary gas chromatography, *J. Chromatogr. A* 1017
401 (2003) 187-193.

402[25] S.-O. Baek, R.A. Jenkins, Characterization of trace organic compounds associated
403 with aged and diluted sidestream tobacco smoke in a controlled atmosphere-
404 volatile organic compounds and polycyclic aromatic hydrocarbons, *Atmos.*
405 *Environ.* 38 (2004) 6583-6599.

406[26] K.S. Pandey, K.-H. Kim, A review of environmental tobacco smoke and its
407 determination in air. *Trends Anal. Chem.* 29 (2010) 804-819.

408[27] S.M. Charles, C. Lia, S.A. Batterman, C. Godwin, VOC and particulate emissions
409 from commercial cigarettes: Analysis of 2,5-DMF as an ETS tracer, *Environ. Sci.*
410 *Technol.* 42 (2008) 1324-1331.

411[28] S.M. Charles, S.A. Batterman, C. Jia, Composition and emissions of VOCs in
412 main- and side-stream smoke of research cigarettes, *Atmos. Environ.* 41 (2007)
413 5371-5384.

414[29] M. Alonso, J.M. Sanchez, Analytical challenges in breath analysis and its
415 application to exposure monitoring, *Trends Anal. Chem.* 44 (2013) 78-89.

- 416[30] G.M. Polzin, R.E. Kosa-Maines, D.L. Ashley, C.H. Watson, Analysis of volatile
417 organic compounds in mainstream cigarette smoke, *Environ. Sci. Technol.* 41
418 (2007) 1297-1302.
- 419[31] S. Moldoveanu, W. Coleman III, J. Wilkins, Determination of benzene and toluene
420 in exhaled cigarette smoke, *Beit. Tabak Int.* 23 (2008) 107-114.
- 421[32] S.K. Pandey, K.-H. Kim, Determination of hazardous VOCs and nicotine released
422 from mainstream smoke by the combination of the SPME and GC-MS methods,
423 *The Scientific World J.* 10 (2010) 1318-1329.
- 424[33] S. Moldoveanu, W. Coleman III, J. Wilkins, Determination of carbonyl compounds
425in exhaled cigarette smoke, *Beit. Tabak Int.* 22 (2007) 346-357.

426

427 **FIGURE CAPTIONS**

428

429 Figure 1. Chromatograms showing the composition of volatile organic compounds in air
430 from an in-door environment and exhaled breath of a volunteer present in this
431 environment without smoking.

432

433 Figure 2. Chromatograms showing the composition of volatile organic compounds in
434 smoke from tobacco cigarettes, Type 1 and Type 2 e-cigarettes and in exhaled
435 breath of smokers. To avoid metabolic differences all displayed exhaled breath
436 chromatograms correspond to the same volunteer.

437

438