

Biosorption of Heavy Metal Arsenic (AS) and Growth Promoting Efficacy of Marine Macroalga, *Ulva fasciata* (L.) on *Vigna radiata*(L.) Seedlings

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ABSTRACT

The impact of heavy metal Sodium arsenate on the growth, yield and biochemical characteristics of *Vignaradiata* (L.) was determined. The addition of high concentrations of heavy metal resulted in decrease in the growth, biochemical and yield characters of *Vignaradiata*. The marine macroalga, *Ulvafasciata* (L.) as liquid fertilizer and algal biomass was screened for its heavy metal biosorption and growth promoting effect. The supplementation with *Ulvafasciata* extract increases the percentage of growth, yield and biochemical content of *Vignaradiata* (L.) compared with control treated with Sodium arsenate alone. Maximum increase in percentage of growth and yield parameters were observed at 20% Liquid Fertilizer and 15g algal biomass of *U.fasciata* treated seedling. The results led to confirm that marine macroalgae could be used to remove the toxicity of heavy metal in polluted environment for sustainable agriculture. It is an economic, cost-effective and safe alternate to existing commercial adsorbents of heavy metal.

1. Introduction

The industrialisation coupled with the technological advances, had led to inadvertent discharge of toxic compounds which adversely affected the ecosystem. The heavy metals are among the toxic compounds which are harmful to the biological systems and do not undergo biodegradation. Metal pollutants can easily enter into the food chain if heavy metal contaminated soil is used for production of food crops resulting in reducing farm productivity (Gosavi *et al.*, 2004). Arsenic is one among the heavy metal that pollutes the environment and has become a global environmental threat. It inhibits plant growth and yield. The World Health Organisation (WHO) and Environmental Protective Agency (EPA) have declared that inorganic arsenic is a human carcinogen. The health of children and adults exposed to high amount of arsenic are affected (Leo and Irudayaraj, 2010). The arsenic contamination also poses a major threat to the marine flora and fauna; the consumption of arsenic contaminated fish causes various ill effects. Many remediation techniques have developed to control and remove the metal pollutants from the environment. But those techniques are highly expensive and environmental disruptive (Gonzaga *et al.*, 2006). The application of biotechnology in controlling and removing metal pollution has been paid much attention. The process of biosorption is a new alternative method, which utilizes certain natural of biological origin, including bacteria, fungi, yeast and algae which possess metal sequestering property (Wang and Chen, 2009) and which have advantages of being cheap, highly efficient and environmentally friendly (Farooq *et al.*, 2010; Srivastava and Majumder 2008; Abdel – Aty *et al.*, 2013).

Marine macroalgae or seaweed are large group of marine benthic algae grown abundantly in the shallow sea

water, estuaries and backwater. Numerous studies revealed their wide range of beneficial effects on crop improvement and yield and elevated resistance to biotic and abiotic stress (Norrie and Keathley, 2006). Besides eliciting a growth promoting effect on plants, macroalgae has been proven as a potent biosorption agent (Davis *et al.*, 2000; Vijayaraghavan *et al.*, 2004; Hashim *et al.*, 2004). Hassan *et al.*, 2004 evaluated metals like Cd, Hg and Pb uptake capacities of green marine macroalgae viz; *Cladophora fascicularis*, *Ulva lactuca* from contaminated solution which was attributed to their uniform cell size and number of metal binding sites on their cell wall (Ramelow *et al.*, 1992). These sites include carboxyl groups, polysaccharides and sulfhydryl groups (Yu *et al.*, 2001). Further, biosorption approach using macroalgae would be technically feasible and economically attractive.

The normal growth and metabolism of plants requires the presence of heavy metals as they have a very important role in the biosynthesis of some enzymes and growth hormones (Nanson and McElroy, 1963). However, their excess supply hinders the plant growth and metabolism of many plant species as reported earlier by Foy *et al.* (1978). The present study attempts to find out the effect of various concentrations of Sodium arsenate on the Green Gram (*Vignaradiata* (L.)). The biosorption capacity of different concentrations of Liquid Fertilizer and algal biomass of *Ulva fasciata* (L.) on the growth yield and biochemical content of Sodium arsenate treated *Vignaradiata* (L.) seedlings was also studied.

2. Materials and methods

Seed weed collection and preparation of Seaweed Liquid Fertilizer (SLF)

The macroalgae, *Ulva fasciata* (L.) was collected from Manavalakurichi coast (8°27'N Latitude and 77°18'E Longitude) of Kanyakumari district during early morning low tide period. It was washed thoroughly, allowed to shade dry and finely powdered using mixer grinder. The powder was sieved and preserved (algal biomass) in polythene bags for metal biosorption studies and determination of plant growth promoting efficacy.

Seaweed Liquid Fertilizer (SLF) was prepared from the dry algal powder as per procedure of Thangam and Rani (2006). The algal powder mixed with distilled water in the ratio of 1:20 (w/v) and autoclaved for 30 min. The hot extracts obtained was filtered through cheese cloth and centrifuged at 10,000 rpm for 15 minutes. The Supernatant was dried at 60°C for 48 hrs. The dried macroalgal extracts considered as 100% seaweed liquid fertilizer and stored at 4°C. From this, different concentrations viz., 1, 5, 10, 15, 20 and 25% of SLF were prepared using double distilled water.

Preparation of Heavy Metal Solution

The heavy metal, Arsenic (As) stock solution (100 ppm) was prepared by dissolving analytical grade sodium arsenate in deionised water. From the stock solution, different concentrations viz., 1, 5, 10, 15, 20 and 25 ppm were prepared by dilution using distilled water.

Experimental Setup

The viable seeds of *Vignaradiata* (L.) were obtained from Tamil Nadu Agricultural University, Pechiparai. The seeds were subjected to viability test; those with 90% viability were selected for the experimental studies.

The planting medium was prepared by mixing garden soil, sand and cow dung in the ratio of 1:2:1. The prepared medium was taken in mud pots of size 30 x 33 cm and is filled off about

two-third of its height. Ten viable seeds of *Vignaradiata* (L.) were sown to a depth of 1.5 cm in each pot. The seeds were watered daily to germinate. After seven days, the seedling of *Vignaradiata* (L.) WILCZEK were treated with various concentrations of Sodium arsenate i.e., 1, 5, 10, 15, 20 and 25 ppm. After seven days of treatment, various morphometric and biochemical characters were analysed. In the other set, 20 ppm sodium arsenate treated seedlings were applied with various concentrations of SLF and algal biomass of *Ulva fasciata* (L.). After seven days of the treatment, morphometric, biochemical and yield parameters were recorded.

The morphometric characters such as root and shoot length, leaf area, number of lateral roots, fresh weight, dry weight, fruit length and number of fruits were measured. The biochemical characters were analysed by the standard methods – Chlorophyll content as per Wellburn and Lichtenthaler (1984), Protein content as per Lowry *et al.* (1951), amino acid content as per Jayaraman, (1981) and total soluble sugar content based on Dubois *et al.* (1951). The experiments were carried out in three replicas.

3. Result and discussion

Effect of sodium arsenate on the growth and yield of *Vignaradiata* (L.)

The addition of increased concentrations of sodium arsenate resulted in gradual decrease in growth and yield of *Vignaradiata* (L.) seedlings. Highest reduction in growth and yield was obtained at 20 ppm concentration of sodium arsenate. The percentage reduction in root and shoot length of sodium arsenate (20 ppm) treated seedlings was 63.6 and 48.5 respectively. There was a drastic reduction in the number of lateral roots (56.8%), leaf area (76.5%), fresh weight (74.2%) and the number and length of fruits (76.9 and 50% respectively) compared with the control treated with water alone (Table 1).

Table -1 Effect of heavy metal Sodium arsenate on the growth and yield parameters of *Vignaradiata* (L.)

Parameters	Control (water)	Concentration of Sodium arsenate (ppm)					% decrease (compared to control)
		1	5	10	15	20	
Root length (cm)	8.25 ± 0.26	4.0 ± 0.70	3.5 ± 0.41	3.2 ± 0.32	3.1 ± 0.25	3.0 ± 0.15	63.6
Shoot length (cm)	16.5 ± 0.18	13.6 ± 0.2	12.5 ± 0.4	11.4 ± 0.6	10.6 ± 0.8	8.5 ± 0.2	48.5
Number of lateral roots	18.5 ± 0.3	15 ± 0.2	14 ± 0.3	12 ± 0.5	10 ± 0.4	8 ± 0.3	56.8
Leaf area (m ²)	17 ± 0.3	14.8 ± 0.5	14.5 ± 0.8	13.1 ± 0.4	10.4 ± 0.2	7.8 ± 0.7	54.1
Fresh weight (gm)	19.1 ± 0.54	17.6 ± 0.20	13.5 ± 0.1	12.9 ± 0.1	7.1 ± 0.04	4.49 ± 0.03	76.5
Dry weight (gm)	6.2 ± 0.92	6.01 ± 0.7	4.5 ± 0.5	4.2 ± 0.4	2.05 ± 0.3	1.63 ± 0.05	74.2
Number of fruits	6.5 ± 0.8	5.2 ± 0.2	4.3 ± 0.09	4.0 ± 0.2	2.1 ± 0.6	1.5 ± 0.01	76.9
Length of fruits (cm)	10 ± 0.03	8 ± 0.9	7 ± 0.4	6.5 ± 0.7	6.5 ± 0.2	5 ± 0.02	50

The findings were in accordance with the study conducted by Upadhyaya *et al.* (2014) on the effect of Sodium arsenate on mung bean seedlings. The uptake of metal occurs primarily through the roots which inhibit the root and shoot growth. The metal uptake also reduces the fresh and dry weight of seedlings treated with heavy metal sodium arsenate (Arudini *et al.*, 1996). Similarly, the reduction of leaf area in response to sodium arsenate treatment was also related to

accumulation of arsenic in leaves. The results coincided with the finding of Pandey and Pathak (2006) in which some of the plants under arsenic stress showed constriction of leaves with appearance of burning spots in to leaf apex. The previous findings of Abedinet *et al.* (2002) and Dhankher *et al.* (2006) also support the present findings.

Similarly, there were variations in the biochemical content of *Vignaradiata* (L.) seedlings treated with Sodium

arsenate. The sodium arsenate (20 ppm) significantly reduced the total chlorophyll (58.8%) and chlorophyll content (40.9%) and slight reduction noted for chlorophyll b (16.3%) content. Similarly, the protein, amino acid and starch content was reduced to 32.3, 66.7 and 53.4 percentage respectively compared to the control. The decrease in the contents of chlorophyll, protein, amino acid and starch of *Vignaradiata* (L.) treated with sodium arsenate could be due to the

generation of reactive oxygen species like superoxide and hydroxyl radicals and hydrogen peroxide that have potential to damage nucleic acid and amino acid involved in the biosynthetic pathway of chlorophyll synthesis (Abbas *et al.*, 2018). Srivastava *et al.* (2013) also reported a similar decline of chlorophyll a, chlorophyll b and total chlorophyll content in *Hydrilla verticillata* at higher doses of arsenic treatment (Table 2).

Table - 2 Effect of heavy metal, Sodium arsenate on the biochemical content of *Vignaradiata* (L.)

Parameters	Control	Concentration of Sodium arsenate (ppm)					% decrease (compared to control)
		1	5	10	15	20	
Chlorophyll a ($\mu\text{g/g}$)	3.5 \pm 0.041	3.32 \pm 0.054	3.06 \pm 0.036	2.84 \pm 0.047	2.42 \pm 0.286	2.07 \pm 0.027	40.9
Chlorophyll b ($\mu\text{g/g}$)	1.236 \pm 0.024	1.207 \pm 0.042	1.144 \pm 0.026	1.134 \pm 0.072	1.112 \pm 0.05	1.035 \pm 0.017	16.3
Total Chlorophyll ($\mu\text{g/g}$)	5.85 \pm 0.062	5.37 \pm 0.062	4.49 \pm 0.049	4.17 \pm 0.012	3.53 \pm 0.021	2.41 \pm 0.026	58.8
Protein (mg/g)	15.50 \pm 0.85	14.10 \pm 0.661	13.3 \pm 0.051	12.7 \pm 0.028	11.6 \pm 0.017	10.5 \pm 0.06	32.3
Amino acid (mg/g)	6.0 \pm 0.1	5.5 \pm 0.75	4.5 \pm 0.26	3.5 \pm 0.17	2.3 \pm 0.71	2.0 \pm 0.68	66.7
Starch (mg/g)	5.8 \pm 0.02	4.0 \pm 0.5	3.8 \pm 0.12	3.5 \pm 0.06	2.9 \pm 0.18	2.7 \pm 0.20	53.4

Determination of biosorption efficacy of SLF from *Ulva fasciata*

Studies on the biosorption efficacy of heavy metal Sodium arsenate by macroalgae, *Ulva fasciata* (L.) revealed that at seaweed liquid fertilizer (SLF) from *Ulva fasciata* at all the studied concentrations viz., 1, 5, 10, 15, and 20 % enhanced

growth in terms of root length (40%), number of lateral roots (61.7%), shoot length (40.5%), leaf area (33.6%), fresh weight (257.5%), dry weight (324.5%), number of fruits (172.2%) and length of fruits (68.8%) of *Vignaradiata* (L.) compared with the control i.e., the seedlings treated with 20 ppm sodium arsenate (Table 3).

Table 3. Cumulative effect of SLF of *U. fasciata* (L.) and heavy metal Sodium arsenate (20 ppm) on the growth and yield parameters of *Vignaradiata* (L.) seedlings.

Parameters	Untreated	Concentrations of SLF (%) and 20 ppm Sodium arsenate						% increase (compared to control)
		Control (20 ppm sodium arsenate)	1	5	10	15	20	
Root length (cm)	11.2 \pm 0.2	7.5 \pm 0.4	7.85 \pm 0.10	8.1 \pm 0.40	9.15 \pm 0.55	9.75 \pm 0.62	10.50 \pm 0.74	40.0
Shoot length (cm)	25.5 \pm 0.84	15.8 \pm 0.25	15.8 \pm 0.08	16.1 \pm 0.14	19 \pm 0.49	22.1 \pm 0.55	22.2 \pm 0.70	40.5
Number of lateral roots	32.5 \pm 0.65	8 \pm 0.3	18 \pm 0.53	25.5 \pm 0.42	27 \pm 0.81	28.5 \pm 0.34	29.1 \pm 0.85	61.7
Leaf area (m ²)	17 \pm 0.3	13.1 \pm 0.7	13.7 \pm 0.2	14.2 \pm 0.5	15.8 \pm 0.2	16.5 \pm 0.1	17.5 \pm 0.32	33.6
Fresh weight (gm)	19.1 \pm 0.54	4.49 \pm 0.25	7.05 \pm 0.21	13.03 \pm 0.17	14.3 \pm 0.28	15.9 \pm 0.12	16.05 \pm 0.07	257.5
Dry weight (gm)	6.5 \pm 0.92	1.63 \pm 0.05	2.05 \pm 0.12	5.1 \pm 0.15	5.13 \pm 0.28	6.7 \pm 0.25	6.92 \pm 0.71	324.5
Number of fruits	1.5 \pm 0.01	1.8 \pm 0.2	2 \pm 0.7	3.1 \pm 0.2	4.2 \pm 0.1	5.1 \pm 0.2	4.9 \pm 0.7	172.2
Length of fruits (cm)	5 \pm 0.02	8 \pm 0.5	8.2 \pm 0.2	9.5 \pm 0.3	10.2 \pm 0.5	10.5 \pm 0.7	10.37 \pm 0.5	68.8

The lowest root length was recorded with sodium arsenate (20 ppm) treated seedlings. The root length of the plants treated with different concentrations of liquid fertilizer of *Ulva fasciata* (L.) was comparatively more. The root length was recorded the highest (90.5% increase) compared with the control for the seedlings treated with 20% SLF of *Ulva fasciata* (L.). The studies by Revathiet *al.*, (2013) on the phytotoxic effect of chromium and EDTA on growth of *Sesbaniagrandiflora* L. supported the findings. Their results indicated that the increase in concentrations of chromium, EDTA, even at very low concentration were effective in phytoextraction of chromium. The phyto remediation efficacy of

macroalgae is due to the biosorption of heavy metal because of the large surface area and metal binding sites on their cell wall. They also have the functional groups like carboxyl, amino acid, polysaccharides and sulfhydryl groups which are present on the cell wall of macroalgae (Yu *et al.* 1999). The present findings coincided with the findings of Selvarajet *al.*, (2010) on the application of seaweed *Gracilariacorticata* at different concentrations with 6mM nickel chloride treated *Vignaradiata* (L.) seedling. Their studies indicated the stress relieving effect of macro algae, on the nickel chloride treated seedling of *Vignaradiata* (L.).

In the present study, the stress imposed by Sodium arsenate treatment to the seedlings of *Vignaradiata*(L.) led to reduction in the quantity and yield. But the yield was enhanced when the seedlings were treated with 20% SLF of *Ulva fasciata*(L.). It was similar to the findings of Sridhar and Rengasamy(2002). They suggested that the phytoremediation activity of macroalgal extracts was due to the presence of macro and micro-elements as well as plant growth regulators like cytokinin in it.

The treatment with seaweed liquid fertilizer of *Ulva fasciata* (L.) also helped in enhancing the content of Total chlorophyll,

Chlorophyll a, Chlorophyll b and Carotenoids (Table 4). The findings are indicative of the fact that of SLF even at the lowest concentration controlled the mobility of heavy metal sodium arsenate as was evident from the growth, yield and increased biochemical contents when compared with the control. The observations were in par with that made by Azmat and Askari(2015) in mercury (Hg) stressed plants and Selvaraj et al(2010) in Nickel chloride treated plants of *Vignaradiata* (L.) These results clearly indicated that addition of Liquid fertilizer from macroalgae, *U.fasciata* reduced the toxic effect of sodium arsenate and promoted the growth and biochemical content of *Vignaradiata* (L.).

Table-4. Cumulative effect of SLF of *U. fasciata*(L.) and heavy metal Sodium arsenate (20ppm) on the biochemical content of *Vignaradiata*(L.)seedlings

Parameters	Control (20ppm Sodium arsenate)	Concentrations of SLF(%) and 20ppm Sodium arsenate (As)					% increase (compared to control)
		1	5	10	15	20	
Chlorophyll a ($\mu\text{g/gm}$)	2.07 \pm 0.07	3.44 \pm 0.041	3.18 \pm 0.050	3.13 \pm 0.030	2.91 \pm 0.028	2.92 \pm 0.026	111.0
Chlorophyll b ($\mu\text{g/gm}$)	1.035 \pm 0.17	1.23 \pm 0.050	1.45 \pm 0.065	2.99 \pm 0.024	2.18 \pm 0.954	2.11 \pm 0.047	195.0
Total Chlorophyll ($\mu\text{g/gm}$)	2.41 \pm 0.026	2.45 \pm 0.048	3.17 \pm 0.050	4.11 \pm 0.054	5.66 \pm 0.061	4.01 \pm 0.039	131.02
Protein(mg/gm)	10.5 \pm 0.06	21.1 \pm 0.443	23.4 \pm 0.581	20.5 \pm 0.482	18.3 \pm 0.391	15.2 \pm 0.221	122.9
Amino acid (mg/gm)	2.0 \pm 0.68	5.2 \pm 0.52	6.1 \pm 0.89	6.5 \pm 0.75	6.4 \pm 0.67	5.5 \pm 0.63	225.0
Starch (mg/gm)	2.7 \pm 0.61	6.4 \pm 0.30	7.2 \pm 0.52	8.9 \pm 0.78	8.8 \pm 0.91	7.4 \pm 0.61	174.1

The treatment with algal biomass of *Ulva fasciata*(L.) to the sodium arsenate (20ppm) treated seedlings also showed a similar trend. A drastic increase in the growth and yield was noted with the algal biomass treated seedlings than control. It is because of the presence of some growth promoting substances such as

IAA, IBA, Gibberellins, Cytokinins, micronutrients and macronutrients in algal biomass which affect the cellular metabolism in heavy metal treated seedling leading to enhanced the growth and crop yield (Durand et al 2003, Ordoget al., 2004).

Table-5. Cumulative effect of algal biomass of *U. fasciata*(L.) and Sodium arsenate (20 ppm) on the growth and the yield parameters of *Vignaradiata* (L.)seedlings

Parameters	control (20ppm Sodium arsenate)	Quantity of algal biomass (gm) and 20ppm Sodium arsenate					Maximum increased in % over control
		1	5	10	15	20	
Root length(cm)	4.5 \pm 0.12	8.7 \pm 0.05	9.35 \pm 0.32	10.95 \pm 0.64	12.4 \pm 0.75	13.9 \pm 0.80	108.8
Shoot length(cm)	8.5 \pm 0.12	10.4 \pm 0.10	15.2 \pm 0.17	18.75 \pm 0.35	22.65 \pm 0.67	24.2 \pm 0.73	166.4
Number of lateral roots	8.1 \pm 0.21	11.5 \pm 0.07	18 \pm 0.42	21 \pm 0.59	27.5 \pm 0.71	33.5 \pm 0.85	116.04
Leaf area(m ²)	13.1 \pm 0.7	13.7 \pm 0.9	14.5 \pm 0.12	15.9 \pm 0.32	16.2 \pm 0.48	17.5 \pm 0.55	31.29
Fresh weight(gm)	4.5 \pm 0.12	5.23 \pm 0.24	7.4 \pm 0.15	12.96 \pm 0.28	21.06 \pm 0.17	12.9 \pm 0.81	212.4
Dry weight(gm)	2.8 \pm 0.31	3 \pm 0.25	4.52 \pm 0.17	5.61 \pm 0.15	7.4 \pm 0.22	4.23 \pm 0.21	164.2
Number of fruits	1.5 \pm 0.2	2.5 \pm 0.3	3.1 \pm 0.2	3.5 \pm 0.3	4.1 \pm 0.1	3.2 \pm 0.5	173.3
Length of fruits(cm)	5 \pm 0.21	6.4 \pm 0.2	7.7 \pm 0.1	8.8 \pm 0.2	9.8 \pm 0.3	9.1 \pm 0.2	96.0

Algal biomass of *Ulva fasciata* (L.) at different quantities treated with sodium arsenate (20ppm) on *Vignaradiata* seedlings improved the pigment and biochemical characters viz., protein, sugar, amino acid and starch in *Vignaradiata*(L.). It was similar to the findings of Sevugaperumalet al., (2012). According to their studies on, the bioadsorption of algal

biomass of *Padina commersonni* on the *Vignaradiata* (L.) pretreated with 6mM concentration of aluminium chloride, increased the level of chlorophyll, protein, sugar, amino acid and starch compared with the control i.e., treated with heavy metal alone. Again these results were coincided with the findings of Jayakumar and Ramasubramanian (2009). Their

results also confirmed that the addition of dry algal biomass reduce the toxic effect of sodium arsenate and promote the

growth and biochemical content of *Vignaradiata*(L) Wilcekseedlings.

Table -6. Cumulative effect of algal biomass of *U. fasciata* and Sodium arsenate (20ppm) on the biochemical content of *Vignaradiata* (L.) seedlings

Parameters	Control (20ppm Sodium arsenate)	Weight of Algal biomass (gm) and 20 ppm Sodium arsenate					Maximum increase in % over control
		1	5	10	15	20	
Chlorophyll a ($\mu\text{g/gm}$)	2.07 \pm 0.027	3.01 \pm 0.024	3.2 \pm 0.062	3.91 \pm 0.067	4.15 \pm 0.072	3.34 \pm 0.059	100.5
Chlorophyll b ($\mu\text{g/gm}$)	1.035 \pm 0.017	1.24 \pm 0.082	1.18 \pm 0.016	1.26 \pm 0.018	1.35 \pm 0.019	1.18 \pm 0.015	30.4
Total Chlorophyll ($\mu\text{g/gm}$)	2.41 \pm 0.026	3.14 \pm 0.027	3.27 \pm 0.038	4.79 \pm 0.036	5.62 \pm 0.048	5.2 \pm 0.042	133.1
Protein(mg/gm)	10.5 \pm 0.06	12 \pm 0.373	12.6 \pm 0.328	13.7 \pm 0.278	15 \pm 0.451	14.5 \pm 0.331	47.6
Amino acid (mg/gm)	2.0 \pm 0.68	3.1 \pm 0.69	4.5 \pm 0.72	5.6 \pm 0.75	6.4 \pm 0.79	6.1 \pm 0.72	155.0
Starch (mg/gm)	2.7 \pm 0.20	5.4 \pm 0.19	6.0 \pm 0.25	7.2 \pm 0.24	9.2 \pm 0.81	8.1 \pm 0.80	151.8

The above results clearly showed that the application of algal biomass of *Ulva fasciata*(L.) on the heavy metal Sodium arsenate treated seedlings reduce the toxic effect of arsenic and enhance the growth and productivity of *Vignaradiata*(L.).

The results of present study indicate that the application of SLF and algal biomass of *Ulva fasciata*(L.) reduce the toxic effect of heavy metal Arsenic and promote the growth and yield of *Vignaradiata* (L.) WILCZEK. Thus, macroalgae or seaweed are used as cost effective, eco-friendly safe alternative to

existing commercial adsorbent for heavy metal stresses plants and polluted environment.

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References

1. Abbas, G., Mustaza, B., Bibi, I., Shahid, M., Niazi, N.K., Khan M.I., Amjad, M., Hussain, M. and Natasha. 2018. Arsenic Uptake, Toxicity, Detoxification and Speciation in Plants: Physiological, Biochemical, and Molecular Aspects: *Int. J. Environ. Res. Public Health*. 15(59): 1-45.
2. Abdel-Aty, A.M. Ammar, N.S., Abdel Ghafar, H.H. and Ali, R.K. 2013. Biosorption of cadmium and lead from aqueous solution by fresh water alga *Anabaena* & *Phaerica* biomass. *Journal of Advanced Research* 4(4), 367-374
3. Abedin M.J., Feldmann J., Mecharg A.A. 2002. Uptake kinetics of arsenic species in rice plants. *Plant Physiol*. 128. 1120 – 11210. 1104/pp. 010733 [PMC free article] [PubMed] [Cross Ref]
4. Aruduini. I., God Bold, D.L. and Onnis, A. 1996. Influence of copper on root growth and morphology of *Pinus plnea L. and Pinus pinaster* Ant seedlings. *Tree physiology*, 15; 411 – 415.
5. Azmat, R. and Askari, S. 2015. Improvement in the bioenergetics system of plants under Hg Stress environment via seaweeds *Pak. J.Bot.*, 47(3): 851-858.
6. Davis T. A., Volesky, B. and Vieira, R.H.S.F. 2000. *Sargassum* seaweed as Biosorbent for Heavy Metals. *Wat.Res.* 34(17): 4270-4278.
7. Dhankher, O.P., Rosen, B.P., Mckinney, E.C. and Meagher, R.B. 2006. Hyperaccumulation of Arsenic in the shoots of *Arabidopsis* Silenced for Arsenate Reductase (ACR 2) *Proceedings of the National Academy of Sciences of the United States of America*, 103, 541-5418.
8. Dubois M., Gilles, K., Hamilton, J. K., Rebers, P. A. and Smith, F. 1951. A colorimetric method for the determination of sugar. *Nature*, 168: 167.
9. Durand, N., Briand, X. and Meyer, C. 2003. The effect of marine bioactive substances (NPRO) and exogenous cytokinins on nitrate reductase activity in *Arabidopsis thaliana*, *Plant physiol* 119: 489 – 493.
10. Farooq, V., Kozinski, J.A., Khan, M.A. and Athar, M. 2010. Biosorption of heavy metal ions using wheat based biosorbents- A review of the recent literature *Bioresource Technology*, 101(14), 5043-5053.
11. Foy, C. D., Chaney, R. L., and White M. C. 1978. The Physiology of Metal Toxicity in Plants. *Ann. Rev. Plant Physiol*. 29: 11-56.
12. Gonzaga MIS, Santos JAG, Ma LQ. 2006. Arsenic phytoextraction and hyperaccumulation by fern species. *Sci Agric*. 63(1): 90–101.
13. Gosavi, K., Sammut, J., Gifford, S. and Jankooski, J. 2004. Macroalgal bio – monitors of trace metal contamination in acid sulphate soil aquaculture. ponds. *Sci. Total Environ.*, 25-39
14. Hashim, M. A and Chu, K.H. 2004. Biosorption of cadmium by brown, green and red seaweeds. *chem. Eng. J.*, 97, 249-255.
15. Hassan Z., Ali, S., Rizwan, M., Ibrahim, M., Nafees, M. and Waseem, M. 2004. Role of Bioremediation Agents (Bacteria, Fungi and Algae) in Alleviating Heavy Metal Toxicity (In: Probiotics in Agroecosystem, Kumar et al.) Springer, pg. 527.
16. Jayakumar, S and Ramasubramanian. V (2009), Bioremoval of chromium using seaweeds as biosorbents. *J. Basic and App Biol*, 3 (s and 4): 121 – 128.
17. Jayaraman, J. 1981. Laboratory manual in Biochemistry, Wiley-Eastern Company Limited, Madras, pp. 1 – 65.
18. Leo, S.C and Irudayaraj, P. 2010. Studies on heavy metal (Arsenic) tolerance in a mangrove fern *Acrostichum aureum* (L). (Peteridaceae). *J. of Basic and Applied Bio.*, 4 (3): 143-152.

19. Lowry, O. H., Rosenbrough, N. J., Farr, A. L. and Randall, R. J., 1951. Protein Measurement with the Folin Phenol Reagent. *J. Biol. Chem.*, 193, pp. 265-275.
20. Nanson A. and McElroy W.D. 1963. Modes of action of the essential mineral elements in: Steward F.C. (eds.) *Plant Physiology: A treatise* Vol.3. Academic Press, New York, pp 451-521.
21. Norrie, J. and Keathley, J.P. 2006. Benefits of *Ascophyllum nodosum* marine plant extract application to 'Thompson seedless' grape production. (Proceedings of the xth International Symposium on plant Bioregulators in Fruit Production, 2005. *Acta Hort.* 727:243-247.
22. Ordog, v., Stirk, W.A., Lenobel, R., Bancirova, M. and Vanstanden, J. 2004. Screening microalgae for some potentially useful agricultural and pharmaceutical secondary metabolites. *Journal of Applied Phycology* 16 309-3014.
23. Pandey, N and Pathak, G.C. 2006. Nickel alters antioxidative defence water status in green gram. *Indian J. Plant Physiol* 11(2): 113 – 118.
24. Ramelow, G.J., Fralick, D and Zhao, Y.F. 1992. Factors affecting the uptake of aqueous metal ions by dried seaweed biomass. *Microbios.*, 72 (291) : 81-93.
25. Revathi, K., Sudha, P., Vinayagam, N.B. and Maximash, A., Rosed, H. 2013. Comparative studies on the effect of antioxidant properties of *Brassica Juncea* and *Sorghum vulgare* when exposed to heavy metal chromium. *Der pharmacia letter* 5 (4) : 84 – 87.
26. Selvaraj K. Jeyaprakash R and Ramasubramanian.V., 2010. Impact of Nickel on growth and biochemical characteristics of *Vignaradiata* (L) Wilczelze and amelioration of the stress by the seaweed treatment. *J. Basic and App. Biol.*, 4(3): 181 – 187.
27. Selvaraj, S., Kitano, H., Fujinaga, Y. Ohga, H., Yoneda, M., Yamaguchi, A.. 2010. Molecular characterization, tissue distribution, and mRNA expression profiles of two kiss genes in the adult male and female chub mackerel. (*Scomber japonicus*) during different gonadal stages. *Gen. comp. Endocrinol.* 169, 28-38. 10.1016.
28. Sevugaperumal, R., Selvaraj, K. and Ramasubramanian, V. 2012. Removal of aluminium by *Padina* as bioadsorbent. *Int. S. Bi & Phar. Res* 3(94): 610-615. ISSN: 0976 3651.
29. Sridhar, S. and Rengasamy, R. 2002. Effect of seaweed liquid fertilizer obtained from *Ulvalactuca* on the biomass, pigments and protein content of *spirulina platensis*. *Seaweed Res. Utilin.* 24: 145 – 149.
30. Srivastava, N.K. and Majumder, C.B. 2008. Novel biofiltration on methods for the treatment of heavy metals from industrial wastewater. *Journal of Hazardous Materials*. 151(1): 1-8.
31. Thangam, R.T and Rani, S.M.V. 2006. Effect of Seaweed Liquid Fertilizers on photosynthetic pigments of Sorghum bicolor (L) Moench. *Seaweed Res. Utilin.*, 28; 81-84.
32. Upadhyaya, H. Shome, S. Roy, D. and Bhattacharya, M.K. 2014. Arsenic Induced changes in Growth and Physiological Responses in *Vignaradiata* seedling: Effect of Curcumin interaction. *American Journal of Plant Sciences*, 5, 3609-3618.
33. Vijayaraghavan K., Joseph R. J., Palanivelu, K and Manickam V. 2004. Copper removal from aqueous solution by marine green alga *Ulva reticulata*. *Electronic Journal. Biotechnol.*, 7 (1) :61-71.
34. Wang, J. and Chen, C. 2009. Biosorbents for heavy metals removal and their future. *Biotechnology Advances* 27, 195-226.
35. Wellburn, A.R., Lichtenthaler, H. 1984. Formulae and program to determine total carotenoids and chlorophyll a and b of leaf extracts in different solvents. In: *Advances in Photosynthesis Research* (C. Sybesma, ed.), The Hague, Netherlands, vol. 2: 9-12.
36. Yu, Q., Kaewasm, P and Yin, P. 2001. Removal of heavy metal ions from waste water by using biosorbent. *Chinese J. Chemical Eng.* 9: 133-136.
37. Yu, Q., Matheickal, J.T., Yin, P., Kaewarn, P. 1999. Heavy metal uptake capacities of common marine macro algal biomass. *water Res.* 33:1534-7.