

Bayesian estimation of predator diet composition from fatty acids and stable isotopes

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Abstract

Quantitative analysis of stable isotopes (SI) and, more recently, fatty acid (FA) profiles are useful and complementary tools for estimating the relative contribution of different prey items in the diet of a predator.

The combination of these two approaches, however, has thus far been limited and qualitative. We propose a mixing model for FA profiles that follows the Bayesian machinery employed in state-of-the-art mixing models for SI. This framework provides both point estimates and probability distributions for individual and population level diet proportions. Where fat content and conversion coefficients are available, they can be used to improve diet estimates. This model can be explicitly integrated with analogous models for SI to increase resolution and clarify predator-prey relationships. We apply our model to simulated data, demonstrating feasibility and model performance, and re-analyse an experimental dataset to illustrate modeling strategies and applications to real fatty acid profiles. Our methods are provided as an open source software package for the statistical computing environment R.

Keywords Stable isotope analysis, quantitative fatty acid analysis, QFASA, lipid profile, diet analysis, Bayesian mixing model, fatty acid signature, dietary marker

1 Introduction

Quantitative estimates of an animals diet are a critical component of predator-prey studies, ecosystem models, and ecosystem-based management.

Existing methods of estimating diet proportions all have strengths and weaknesses (**bowen'methods'2012**). Traditional stomach content and fecal matter analysis represent a brief snapshot of diet at a particularly place and time and can be invasive, time-consuming, and potentially biased by differential rates of digestion of prey or ingestion of identifiable prey parts (**bowen'methods'2012**). Chemical markers such as stable isotopes (SI) and fatty acids (FA; often called fatty acid signatures or profiles) solve some of these problems. For example, both approaches integrate diet composition over an extended time period - typically weeks to months, depending on tissue turnover rates (**tucker'convergence'2008**). These advantages have led to rapid growth in the use of chemical markers in diet studies (**elsdon'unraveling'2010; williams'using'2010; kelly'fatty'2011; bowen'methods'2012**). However, chemical dietary markers generally lack the specificity of traditional stomach content analysis. In particular, several prey species often have similar isotopic signatures. More recent studies have sought greater dietary resolution through the use of SI of other elements in addition to carbon and nitrogen (**belicka'stable'2012**), compound specific SI ratios (**budge'tracing'2008; jack'individual'2011**), or a combination of stomach content analysis and SI or FA (**pethybridge'seasonal'2012**). The use of SI and FA in combination also holds great promise; however, studies that have used both chemical markers have been qualitative ([**e.g.; guest'trophic'2009**) or based on positive correlation of results from both methods (**tucker'convergence'2008**).

Analysis tools for SI data have become very sophisticated in recent years,

48 starting with the development of general Bayesian analysis tools for estimating
 49 diet proportions, and leading to customized (hierarchical) models for
 50 individual applications (**moore'incorporating'2008**;
 51 **hopkins'estimating'2012**; **parnell'bayesian'2012**). The latter models
 52 can, for instance, estimate dietary differences of geographically distinct
 53 populations (**semmens'quantifying'2009**), accommodate temporal changes
 54 in diets or estimate the effect of covariates (e.g., age, size, sex) on diet
 55 proportions (**parnell'bayesian'2012**). While these models provide a
 56 considerable step towards ecologically relevant models in diet studies, the
 57 underlying SI data is limited in the resolution that it can provide. Since
 58 typically only 2-3 SI are measured, the contrast that is achievable from such a
 59 low number of variables is necessarily limited, especially when the number of
 60 potential prey items increases (**phillips'source'2003**;
 61 **ward'quantitative'2011**). Optimally aggregating prey items into prey
 62 groups may circumvent this problem (**ward'quantitative'2011**), but may
 63 also be less satisfactory in complex food webs.

64 FA data can, in theory, provide considerably more resolution compared to SI
 65 data, due to large number of potential FA that can be measured. Furthermore,
 66 **blanchard'inference'2011** developed a Bayesian model for diet inference
 67 from FA (furthering the development of Bayesian mixing models for
 68 compositional data by **billheimer'compositional'2001**), showing that
 69 model based inferences of predator diets from FA are achievable. Nevertheless,
 70 studies employing FA remain either qualitative in their estimates of prey
 71 proportions in predator diets, or use Quantitative Fatty Acid Signature

72 Analysis (**iverson'quantitative'2004**) to obtain quantitative estimates of
73 diet proportions.

74 QFASA is the only available (i.e., off the shelf) method thus far for use with
75 FA data, and, in contrast to recent (Bayesian) SI and FA mixing models, relies
76 on a distance metric rather than a model based formulation to estimate the
77 most likely diet proportions. This framework provided the first quantitative
78 approach to estimating diet proportions using FA and it has already seen
79 widespread use, particularly in studies of marine mammals
80 (**bowen'methods'2012**) and seabirds (**williams'using'2010**).

81 Nevertheless, QFASA has a number of limitations. Since it is not based on a
82 probabilistic model, it is difficult to estimate uncertainty associated with
83 estimated diet proportions (**blanchard'inference'2011**). The absence of an
84 explicit model also makes it impossible to build ecological mechanisms (e.g.,
85 covariates of consumed diets) directly into the model. Furthermore,
86 uncertainty about conversion coefficients representing enrichment and
87 preferential uptake of FA cannot be considered, yet small changes in these
88 coefficients can lead to differences in inferred diet proportions
89 (**wang'validating'2010**).

90 Given the discrepancy in methods applied to SI and FA data, it is perhaps not
91 surprising that their joint application has commonly relied on qualitative
92 comparisons. Because both markers integrate diet composition over often
93 comparable time-scales, however, an explicit integration of these data types
94 could provide substantial benefits. While FA data could mitigate the
95 resolution problem in SI data, SI data could provide increased resolution and

clarify predator-prey relationships, the knowledge of which is usually a pre-requisite for FA data. For example, for many non-modified fatty acids, FA alone cannot discriminate between the case of two species which share a common diet and the situation in which one of these species eats the other. In either case, the two species may have similar FA. The addition of a stable isotope with trophic fractionation (e.g., ^{15}N), however, can readily distinguish predation from dietary overlap.

Here, we develop a mixing model for FA data based on a probabilistic model whose parameters are estimated using Bayesian methods, and explicitly integrate SI in the estimation of diet proportions. Using both simulated and published data, we demonstrate the suitability of this model for FA analysis and highlight the potential benefit of explicit integration with SI data to increase the precision of diet estimates.

2 Methods

2.1 A Bayesian mixing model for fatty acid profiles

Bayesian models for SI data are commonly based on the assumption that SI ratios are normally distributed. This assumption cannot be made for FA data, since for most methods of analysis, the concentration of individual FA is normalized to the total lipid content of the sample. Thus, the FA are a collection of proportions (referred to as a composition), which lie between 0 and 1, and are constrained to sum to 1. A common solution to this problem,

117 however, is to consider transformations that make the data approximately
 118 normal (**budge'studying'2006**). To construct our model, we considered the
 119 additive log ratio transformation (**aitchison'convex'1999**), also called alr
 120 transformation, such that

$$y_{i,s} = alr(\phi_{i,s}) = \log \left(\frac{\phi_{i,s,1 \dots p-1}}{\phi_{i,s,p}} \right) \quad (1)$$

121 where $\phi_{i,s}$ is the p -variate fatty acid composition of individual i of prey species
 122 s , with a total of n potential prey species considered. Note that in the
 123 following we often drop the subscript for FA, e.g., $\phi_{i,s}$ and $y_{i,s}$ are thus p and
 124 $p - 1$ dimensional vectors, respectively. We assumed that the distribution of y
 125 is multivariate normal, with species specific mean μ_s and covariance matrix
 126 Σ_s , or $y_{i,s} \sim N(\mu_s, \Sigma_s)$. A vaguely informative prior on μ_s and Σ_s allows for
 127 uncertainty in prey distributions to propagate to estimates of diet proportions
 128 (**ward'including'2010**).

129 Each predator j consumes a proportion π_j of each prey source, and analogous
 130 to stable isotope mixing models, predator FA are then a linear combination of
 131 prey FA, normalized to sum to one. Since predators may selectively assimilate
 132 or metabolize FA (**iverson'quantitative'2004; budge'studying'2006;**
 133 **rosen'effects'2012**), we specify prey-specific conversion coefficients
 134 $\kappa_s = \kappa_{s,1} \dots \kappa_{s,p}$ for each of the p FA (**rosen'effects'2012**). Furthermore, the
 135 n prey species likely have different fat content Φ that will affect the total
 136 amount of FA assimilated from each prey species by the predator. The

137 expected FA of predator τ_j is then a linear combination of the prey FA,
 138 modified by conversion coefficients for each fatty acid p and fat content for
 139 each prey i :

$$t_j \sim N(\text{alr}(\tau_j), \Sigma_\tau) \quad (2)$$

$$\tau_j = C \left\{ \sum_s^n (\pi_{j,s} \Phi_s) (\kappa_s \otimes \phi_{j,s}) \right\} \quad (3)$$

140 Here, C is the closure operation which normalizes the FA to sum to one and \otimes
 141 is the outer (element wise) product. $\phi_{s,j}$ is the FA of prey items of species s
 142 consumed by predator j . Similarly to **parnell'bayesian'2012**, we thus
 143 assumed that individual predators do not necessarily feed on 'average' prey
 144 items, but rather consume prey items with signatures drawn from the
 145 estimated prey distribution. We formulate predator signatures t as draws from
 146 a normal distribution after transformation. We further assumed that Φ and κ
 147 are log-normally and gamma distributed, respectively, around known mean
 148 and variance values (estimated or calculated from controlled diet experiments,
 149 see below). The closure operation in Equation 2 (i.e., the sum-to-one
 150 constraint on the FA) leads to κ being determined in terms of relative uptake
 151 of FA (i.e., up to a multiplicative constant), and implicitly makes the
 152 multivariate prior distribution over all κ a Dirichlet distribution. The same
 153 logic applies to Φ , and in both cases we opted for formulations that can be
 154 readily parametrized from priors studies or published values (e.g., sample
 155 means and variances from experiments).

156 The diet proportions π of predators are the main focus of investigation in diet
 157 studies. These may be modeled at the (statistical) population level (thus
 158 dropping the subscript j in Equation 2) or at the individual level, as suggested
 159 in Equation 2. In the latter case, individual predator FA can be modeled as
 160 draws from a population level distribution of predator diet proportions.
 161 Recent approaches to stable isotope mixing have focused on transformations of
 162 the diet proportion vector π to get around the problems associated with the
 163 compositional nature of the diet proportions in such a hierarchical setup, and
 164 we follow this approach in our model. The diet proportions are transformed
 165 using clr transformations (**semmens'quantifying'2009**), such that the
 166 support of is the real line rather than the interval $[0;1]$, and we then assume
 167 that $clr(\pi_j) \sim N(\Pi, \Sigma_\Pi)$, where Π is the vector of mean (population level) diet
 168 proportions. It is then possible to model diet proportions as function of
 169 covariates, such as size, sex, or region (**parnell'bayesian'2012**). While this
 170 approach is appealing, it adds to computation time needed to estimate model
 171 parameters, and correlates with generally slower convergence. We therefore
 172 use a vague Dirichlet prior on the proportions when convenient (i.e., when we
 173 estimate only population level parameters).
 174 Depending on the amount of samples for prey and predators, it may be
 175 necessary to use informative priors for Σ_s and Σ_τ . Both were given
 176 inverse-Wishart priors, and since both are co-variances of transformed data, it
 177 is not straightforward to formulate default priors for these parameters. We
 178 have found that in practice manual adjustment of these priors is often needed
 179 to be able to achieve convergence and mixing (efficient exploration of the

posterior distribution by the sampling algorithm) of the Markov Chain Monte Carlo (MCMC) employed to estimate model parameters. This is especially true when there are few source and/or predator samples. The package allows for high level adjustment of these parameters through the specification of the order of magnitude of the diagonal of each covariance matrix.

2.2 Joint diet estimation from FA and SI

There are at least three potential benefits of integrating FA and SI data: i) increased information to discriminate among sources, ii) the potential of SI to resolve predator prey relationships due to trophic enrichment of SI, and iii) the potential reduction in estimation error due to a larger body of research on fractionation coefficients for stable isotopes as opposed to conversion coefficients in FA. Integrating the two complimentary types of data in a single model to estimate diet proportions may thus considerably improve estimates of diet proportions over estimation from either data-source alone.

Our model for FA is conceptually similar to recent models proposed for SI data, and integration of FA and SI data into a single model is straightforward in the present setting. We assume that the vector of SI signatures of sampled prey items q follow a multivariate normal distribution, such that $y_{q,s}^{SI} \sim N(\mu_s^{SI}, \Sigma_s^{SI})$, where the superscript SI denotes that these are stable isotope signatures. Predator SI signatures are again a linear combination of prey SI, this time modified by additive fractionation coefficients γ . Fractionation may, in turn, depend on prey isotope concentrations

202 (**hussey'rescaling'2014; caut'variation'2009**). In our model, we assume
 203 additive fractionation, and suggest that concentration dependence is taken
 204 into account when specifying distributions for prey and SI specific
 205 fractionation coefficients γ_s (see examples below). The expected SI signature
 206 for predator r is then

$$t_r^{SI} = \sum_s^n \pi_{r,s} (y_{q,r} + \gamma_s) \quad (4)$$

$$clr(\pi_r) \sim N(\Pi, \Sigma_\Pi) \quad (5)$$

$$\gamma_{s,SI} \sim N(\nu_{SI}, \sigma_{SI}) \quad (6)$$

207 Note that the different subscripts to the FA model imply that there is no need
 208 to have SI and FA from the same prey or predator samples, as long as we can
 209 assume that the prey samples are drawn from the same statistical population
 210 as those for FA, and that individual diet proportions of predators are drawn
 211 from the same population distribution of diet proportions.

212 The exact formulation of the integration of SI and FA depends on the
 213 assumptions that one is comfortable with in a given setting: identical dietary
 214 proportions may be appropriate if diets (and hence SI and FA) are thought to
 215 be stable, or if both chemical tracers are thought to integrate over similar
 216 time-scales. If the time scales of these two elements are thought to be different
 217 (e.g., for different tissue types), individual diet proportions may be more
 218 appropriate, and may be drawn from an overall population distribution of diet

219 proportions.

220 An R (**R'core'2014**) package (called fastinR) implementing methods
221 outlined here, along with simulated examples and the analysis of experimental
222 data described further below, is available on the open source repository
223 github.com/philipp-neubauer/fastinR. Models implemented in the package
224 include the above-mentioned formulations for population level diet estimates,
225 individual diet estimates as well as linear model (regression and ANOVA)
226 formulations for diet proportions, all available for SI and FA individually or as
227 combined models (see below). Model parameters were estimated using MCMC
228 methods implemented in JAGS (**plummer'jags'2003**), called from R
229 through higher level functions in the fastinR package that allow for data input,
230 inspection and manipulation.

231 **2.3 Simulation studies**

232 We initially explored the feasibility and performance of our model setup in a
233 range of simulations, which are illustrated (including code) in supplemental
234 information S1. Simulations were also used to explore sensitivities of inferred
235 diet proportions to the source configuration and diet evenness in a series of
236 simulation experiments. We hypothesized that estimated diet proportions are
237 sensitive to diet source separation in FA space, co-linearity in FA space
238 (**blanchard'inference'2011**) and diet makeup (e.g., specialist versus
239 generalist diets). Further details and simulation results can be found in
240 supplemental information S2.

241 **2.4 Selecting fatty acids for analysis: an ordination**

242 **approach**

243 A potentially large number of FA are available from analysis methods such as
244 gas-chromatography. A common practice is to simply set a threshold and keep
245 the most abundant FA for analysis. This practice may, however, discard
246 potential useful information, and a more judicious approach is to retain FA
247 based on the among diet source variability that they explain.

248 **wang'validating'2010** used a method by which they tested the QFASA
249 method on a series of subsets to determine the subset that gave the best
250 accuracy. Although feasible, such a method is prohibitive with fully Bayesian
251 models, which can take a long time to run with a realistic dataset.

252 Here, we propose a variable selection method based on constrained ordination,
253 which considers the contribution of individual fatty acids to axes separating
254 diet sources. Based on this contribution relative to the overall separation, the
255 user can choose FA that contribute most to source separation. This procedure
256 is intended to reduce computation time (and dimensionality) of the models,
257 while retaining as much accuracy in diet estimates as possible. Further details
258 about the procedure are given in supplemental information S3.

259 2.5 Application: estimating predator diets in a 260 controlled experiment

261 To illustrate the potential of the models presented above, we analysed data
262 from an experimental study by **stowasser'experimental'2006**, which
263 investigated changes in squid FA and SI as a function of diet treatments. The
264 treatments consisted of exclusive fish and crustacean diets, as well as switched
265 and mixed diets, with the former switching diets from fish (henceforth SF,
266 $n=4$) to crustacean (SC, $n=5$) after 15 days of the 30 day experiment.

267 In order to apply our model, we first estimated conversion coefficients of FA
268 and fractionation in SI, using squid from the 30 day diet treatments feeding
269 exclusively crustacean and fish diets. The model for estimation of SI
270 fractionation followed the model in **hussey'rescaling'2014**, thus accounting
271 for diet $\delta^{15}N$ and $\delta^{13}C$, and used their results as priors for fractionation
272 parameters for $\delta^{15}N$, and results from **caut'variation'2009** to construct
273 priors for $\delta^{13}C$. Estimation of FA conversion coefficients used (2) with
274 proportions assumed known from feeding trials. Computational details on the
275 estimation of conversion coefficients and fractionation are given in
276 supplemental information S4.

277 In our diet analysis, we analyzed samples from the switched diet treatments,
278 and used both SI and FA to investigate if our models allow us to infer diet
279 proportions in either treatments. We subset the data to use only switched diet
280 squid that were analysed for FA and SI after at least 10 days under the
281 respective treatment. We only had overlapping SI and FA for the SC

282 treatment squid, and we therefore started by analyzing this treatment in
283 isolation to demonstrate that both SI and FA can resolve diet proportions, and
284 to demonstrate the benefit of using the two tracers in a joint model. We then
285 analyzed the SF treatment squid, for which we only had 3 specimen with FA
286 and 1 specimen with SI. The markers available for this treatment did not
287 overlap for any of the sampled squid.

288 We lastly estimated individual diet proportions in the SC treatment. To
289 demonstrate how the model based approach to diet estimation can be use to
290 answer ecologically relevant questions about predator diets, we also analyzed
291 SF and SC treatment squid together in a linear model setup that investigated
292 treatment differences explicitly. The linear model used treatment dummy
293 variables to estimate individual intercepts for each treatment and prey
294 combination, and allows us to estimate, conditional on the data and priors,
295 whether squid in either one treatment group consumed significantly more of
296 any one prey type.

297 FA analyses used data obtained by analyzing digestive glad tissue, which is
298 thought to rapidly assimilate dietary FA in relatively unmodified proportions
299 relative to the original diet. SI were analyzed from muscle tissue since we had
300 more individuals sampled for SI from this tissue, which may be more prone to
301 fractionation and slower turnover than digestive glad tissue. In the original
302 study, a total of 25 FA were reported. Here, we selected FA using ordination
303 methods described above. For estimation of model parameters, priors for prey
304 and predator specific variances were adjusted manually to give reasonable
305 behaviour in the MCMC algorithm. The analyses are detailed in supplemental

306 information S5.

307 **3 Results**

308 **3.1 Simulation studies**

309 Simulated test cases suggested that our model can estimate diet proportions
310 from both SI and FA (supplemental information S1), with accuracy depending
311 mainly on source separation and diet evenness (supplemental information S2).
312 For very uneven diet proportions, such as in the feeding trials analyzed in the
313 squid example, we found the choice of posterior means as point estimate for
314 diet proportions inevitably introduced error at the margins of the 0-1 interval
315 when compared to true simulated diet proportions.

316 Models with low accuracy conversion coefficients (with prior mean for all FA
317 set to 1 and large prior variance) also performed substantially worse than
318 models with accurately specified coefficients when comparing point estimates
319 of diet proportions to simulated diet proportions (supplemental information
320 S2), showing decreasing accuracy with increasing variance among simulated
321 convergence coefficients.

322 **3.2 Squid diet experiments**

323 Dimension reduction by NMDS on FA of squid and their potential prey
324 suggested that crustacean diets were readily distinguishable from fish diets
325 (??a). For fish diet items, however, no single fish species could be clearly