

Concordance and Diagnostic Accuracy of [¹¹C]PIB PET and Cerebrospinal Fluid Biomarkers in a Sample of Patients with Mild Cognitive Impairment and Alzheimer's Disease

Antoine Leuzy^a, Stephen F. Carter^{a,b}, Konstantinos Chiotis^a, Ove Almkvist^{a,c}, Anders Wall^d and Agneta Nordberg^{a,e,*}

^aDepartment NVS, Centre for Alzheimer Research, Division of Translational Alzheimer Neurobiology, Karolinska Institutet, Huddinge, Sweden

^bWolfson Molecular Imaging Centre, University of Manchester, Manchester, United Kingdom

^cDepartment of Psychology, Stockholm University, Stockholm, Sweden

^dSection of Nuclear Medicine and PET, Department of Radiology, Oncology and Radiation Sciences, Uppsala University, Uppsala, Sweden

^eDepartment of Geriatric Medicine, Karolinska University Hospital Huddinge, Stockholm, Sweden

Accepted 12 January 2015

Abstract.

Background: Alzheimer's disease (AD) pathology can be quantified *in vivo* using cerebrospinal fluid (CSF) levels of amyloid- β_{1-42} ($A\beta_{1-42}$), total-tau (t-tau), and phosphorylated tau (p-tau_{181p}), as well as with positron emission tomography (PET) using [¹¹C]Pittsburgh compound-B ([¹¹C]PIB). Studies assessing concordance between these measures, however, have provided conflicting results. Moreover, it has been proposed that [¹¹C]PIB PET may be of great clinical utility in terms of identifying patients with mild cognitive impairment (MCI) who will progress to the dementia phase of AD.

Objective: To determine concordance and classification accuracy of CSF biomarkers and [¹¹C]PIB PET in a cohort of patients with MCI and AD.

Methods: 68 patients (MCI, $n = 33$; AD, $n = 35$) underwent [¹¹C]PIB PET and CSF sampling. Cutoffs of >1.41 ([¹¹C]PIB), <450 pg/mL—and a more lenient cutoff of 550 pg/mL—($A\beta_{1-42}$), <6.5 ($A\beta_{1-42}/p\text{-tau}_{181p}$), and 1.14 ($A\beta_{1-42}/t\text{-tau}$), were used to determine concordance. Logistic regression was used to determine classification accuracy with respect to stable MCI (sMCI) versus MCI who progressed to AD (pMCI).

Results: Concordance between [¹¹C]PIB and $A\beta_{1-42}$ was highest for sMCI (67%), followed by AD (60%) and pMCI (33%). Agreement was increased across groups using $A\beta_{1-42} < 550$ pg/mL, or $A\beta_{1-42}$ to tau ratios. Logistic regression showed that classification accuracy of [¹¹C]PIB, between sMCI and pMCI, was superior to $A\beta_{1-42}$ (73% versus 58%), $A\beta_{1-42}/t\text{-tau}$ (63%), and $A\beta_{1-42}/p\text{-tau}_{181p}$ (65%).

*Correspondence to: Agneta Nordberg, MD, PhD, Division of Translational Alzheimer Neurobiology, Karolinska University Hospital Huddinge, Novum 5th Floor, Blickagången 6 S-141 57 Stockholm, Sweden 141 57. Tel.: +46 8 585 854 67; Fax: +46 8 585 854 70; E-mail: Agneta.k.nordberg@ki.se.

Conclusion: In the present study, [¹¹C]PIB proved a better predictor of progression to AD in patients with MCI, relative to CSF measures of A β ₁₋₄₂ or A β ₁₋₄₂/tau. Discordance between PET and CSF markers for A β ₁₋₄₂ suggests they cannot be used interchangeably, as is currently the case.

Keywords: [¹¹C]PIB, Alzheimer's disease, amyloid, cerebrospinal fluid, mild cognitive impairment, positron emission tomography, tau

INTRODUCTION

While at present incurable, intensive efforts to develop disease-modifying drugs to counteract the progression of Alzheimer's disease (AD) are ongoing. Given the increasing consensus that such drugs must be administered early on in the disease course if they are to prove effective, the further development and validation of diagnostic tools capable of accurately identifying AD pathophysiology at an early stage has become an important area of research. In addition to aiding in the selection of patients appropriate for inclusion in clinical trials of disease modifying drugs, effective diagnostic tools will also be required in routine clinical practice to identify patients with mild cognitive impairment (MCI) due to AD so as to provide treatment should these drugs be approved for widespread use.

On the basis of the current hypothetical model of dynamic biomarkers for AD [1], the aggregation and deposition of the 42 amino acid variant of amyloid- β (A β ₁₋₄₂) is thought to set in motion various neurodegenerative processes that result in cognitive impairment, and, ultimately, dementia. Indeed, biomarker studies in sporadic AD have shown that decreased cerebrospinal fluid (CSF) levels of A β ₁₋₄₂ is a very early change [2, 3], with studies in familial AD suggesting that these reductions precede expected symptom onset by 25 years [4]. In addition to low A β ₁₋₄₂, increased CSF levels of total tau (t-tau) and tau phosphorylated at threonine 181 (p-tau_{181p}) are typically seen in AD [5-7]. In this respect, a previous study reported that ratios of CSF A β ₁₋₄₂ to t-tau (A β ₁₋₄₂/t-tau) and p-tau_{181p} (A β ₁₋₄₂/p-tau_{181p}) outperformed any single analyte in discriminating patients with and without cortical amyloid deposition [8], suggesting that a combinatorial approach is best when evaluating CSF biomarkers.

In addition to CSF measurements, cerebral amyloid deposition can be determined using positron emission tomography (PET) and amyloid-binding radiotracers, such as [¹¹C]-labelled Pittsburgh compound

B ([¹¹C]PIB). The most widely studied PET amyloid ligand to date, [¹¹C]PIB uptake has been shown to be significantly increased in familial AD mutation carriers 15 years prior to expected symptom onset [4], and strongly associated with region-matched, quantitative analyses of A β in postmortem AD tissue [9, 10]. In MCI, [¹¹C]PIB uptake follows a bimodal distribution, with patients showing uptake values that overlap with those seen in cognitively normal controls (i.e., below the threshold required for [¹¹C]PIB positivity) or that correspond to those seen in AD [11].

Previous studies using both continuous and dichotomous correlations have shown the information obtained using [¹¹C]PIB PET and CSF A β ₁₋₄₂ to be in strong agreement [12-14], pointing to convergent validity of both types of biomarkers [15, 16]. A recent study, however, suggests that [¹¹C]PIB may be more sensitive than the concentration of CSF A β ₁₋₄₂ with respect to the detection of cerebral amyloidosis in MCI [17]. In line with this report, a study addressing the diagnostic performance of CSF A β ₁₋₄₂ and PET [¹⁸F]florbetapir—a radiofluorinated amyloid tracer exhibiting high agreement with [¹¹C]PIB [18]—showed that while the diagnostic accuracy of both measures were similar, PET had greater specificity owing to discordance between measures [19]. Importantly, PET and CSF amyloid biomarkers are considered clinically equivalent on the basis of the recently revised diagnostic criteria for MCI and dementia due to AD [20, 21].

Using a memory clinic sample of patients with MCI and AD, the present study investigated 1) concordance between CSF A β ₁₋₄₂ (both alone, and combined with tau) and [¹¹C]PIB PET; and 2) classification accuracy of [¹¹C]PIB PET and CSF A β ₁₋₄₂ (both alone, and combined with tau) with respect to stable MCI (sMCI) versus MCI who progressed to AD dementia (pMCI). It was hypothesized that [¹¹C]PIB PET and CSF measures of A β ₁₋₄₂, A β ₁₋₄₂/t-tau, and A β ₁₋₄₂/p-tau_{181p} would exhibit high concordance, and that PET would prove superior to CSF measures in the case of discerning sMCI from pMCI.

114 METHODS

115 Patients

116 Thirty-three MCI and 35 AD patients were
117 recruited from the Department of Geriatric Medicine,
118 Karolinska University Hospital Huddinge, Stockholm,
119 Sweden. All patients underwent a comprehensive
120 routine assessment procedure, including a physical
121 examination, evaluation of neurological and psy-
122 chiatric status, blood (including apolipoprotein E
123 (APOE) genotyping), serum and urine analysis, struc-
124 tural imaging, CSF sampling, and neuropsychological
125 assessment. MCI patients met the Petersen criteria
126 [22, 23], with the diagnosis of probable AD based
127 on the National Institute of Neurological and Commu-
128 nication Disorders, Alzheimer’s Disease and Related
129 Disorders Association (NINCDS-ADRDA) criteria
130 [24]. In all cases, diagnosis was issued via a con-
131 sensus based committee approach, which included
132 neurologists, clinical neuropsychologists, and special-
133 ist nurses.

134 MCI patients underwent regular clinical follow-ups,
135 with 12 progressing to AD dementia (pMCI). Clas-
136 sification of MCI patients into sMCI and pMCI took
137 into account findings from CSF, but not [¹¹C]PIB-PET.
138 8 out of 31 MCI received treatment with acetyl-
139 cholinesterase (AChE) inhibitors. In the AD group, 30
140 received treatment with AChE inhibitors—including
141 14 with phenserine—with 5 subjects receiving no phar-
142 macological treatment. All patients and their caregivers
143 provided written informed consent to participate in the
144 investigation, which was conducted according to the
145 declaration of Helsinki and subsequent revisions. Eth-
146 ical approval was obtained from the regional human
147 ethics committee of Stockholm and the Faculty of
148 Medicine and Radiation, Hazard Ethics Committee of
149 Uppsala University Hospital, Sweden.

150 Neuropsychological assessment

151 A routine clinical neuropsychological assessment
152 was performed on all participants. Global cognition
153 was assessed using the Mini-Mental State Examina-
154 tion (MMSE) and a composite measure including the
155 Information and Similarities subscales of the Wech-
156 sler Adult Intelligence Scale (language), Block design
157 and Rey Osterrieth copy (visuospatial), Digit Span
158 and Corsi Span (working memory), the Digit Symbol
159 and Trail Making Tests A and B (attention/executive
160 domains). In addition, episodic memory was assessed
161 using the Rey Auditory Verbal Learning Test learning

162 and retention, as well as the Rey Osterrieth reten-
163 tion. Patients’ raw neuropsychological test scores were
164 transformed into z-scores with respect to a reference
165 group of healthy elderly controls from Karolinska Uni-
166 versity Hospital, Huddinge, Sweden [25]. Applying a
167 cutoff of -1.5 SD to this composite episodic memory
168 score, 12 out of 21 MCI patients were considered as
169 amnesic MCI.

PET imaging

170 PET investigations were performed at Upp-
171 sala PET Centre on ECAT EXACT HR+scanners
172 (Siemens/CTI) or a Discovery ST PET/CT scanner
173 (GE). The orbito-meatal line was used to center the
174 head of the participants. PET data was acquired in
175 3D mode yielding a 155-157 mm field of view. The
176 [¹¹C]PIB-PET data acquisitions consisted of 24 frames
177 (4 × 30, 9 × 60, 3 × 180, and 8 × 300 s), acquired over
178 60 min. A late 40–60 min [¹¹C]PIB summation image
179 was created and used for subsequent image analysis.
180 The mean injected dose was 295 ± 69.5 MBq.

181 All emission data were reconstructed with filtered
182 back projection (FBP) using a 4 mm Hanning filter,
183 resulting in a transaxial spatial resolution of 5 mm
184 in the FOV. The matrix included 128 × 128 pixels,
185 with a zoom factor of 2.5. All reconstructed frames
186 were re-aligned to correct for between frame patient
187 motion.

PET data analysis

188 Individual 40–60 min integral [¹¹C]PIB images
189 were nonlinearly spatially normalized to a population-
190 based [¹¹C]PIB template (see Nordberg et al. [26] for
191 a detailed description of the [¹¹C]PIB template) using
192 the normalize function in SPM5 (Functional Imaging
193 Laboratory, Wellcome Department of Imaging Neu-
194 roscience, UCL, London, UK). Spatially normalized
195 images were then resampled using a 23-region grey
196 matter atlas, created in parallel to the [¹¹C]PIB tem-
197 plate [26]. Subsequently, standardized uptake value
198 ratio (SUV_R) images were calculated by normal-
199 izing the activity within frontal, temporal, parietal,
200 occipital, parahippocampal, anterior, and posterior
201 cingulate regions to mean cerebellar grey matter
202 (vermis excluded) activity. Using the upper 95% con-
203 fidence limit of 1.41 from a previously characterized
204 population of normally distributed healthy controls
205 [26], patients were subdivided into [¹¹C]PIB ‘posi-
206 tive’ ([¹¹C]PIB+), and [¹¹C]PIB ‘negative’ ([¹¹C]
207 PIB-).
208

CSF measurements

CSF samples were obtained via lumbar puncture (LP) from 67 patients (all MCI and 34 of 35 AD). LP was performed under non-fasting conditions, between 8 and 11 a.m., with a total of 10 mL of CSF collected. After discarding the first 0.5 mL, samples were centrifuged at $1500 \times g$ (3000–4000 rpm) for 10 min at $+4^{\circ}\text{C}$. Samples were then stored at -80°C in 1 ml portions pending biochemical analysis, without being thawed or refrozen. Levels of $\text{A}\beta_{1-42}$, t-tau, and p-tau_{181p} were determined using commercially available sandwich ELISAs (Innogenetics, Ghent, Belgium) [27–29]. For the MCI group, levels of $\text{A}\beta_{1-42}$ and t-tau were obtained for all subjects, with p-tau_{181p} available for 26 of 33 subjects. For the AD group, tau levels (total and phosphorylated) were unavailable for 2 and 10 subjects, respectively. Cutoff values of <450 pg/mL ($\text{A}\beta_{1-42}$), >400 pg/mL (t-tau), and >80 pg/mL (p-tau_{181p}) were adopted based on receiver operating characteristic analyses previously conducted by the department of Clinical Chemistry, Karolinska University Hospital Huddinge [30, 31]. In addition, we applied a more lenient cutoff of <550 pg/mL for $\text{A}\beta_{1-42}$ since preliminary findings from the Biomarkers for Alzheimer's and Parkinson's disease (BIOMARKAPD) initiative [32] suggest this to be an optimal cutoff (unpublished data). Cutoffs of 1.14 for $\text{A}\beta_{1-42}/\text{t-tau}$ and 6.5 for $\text{A}\beta_{1-42}/\text{p-tau}_{181p}$ were taken from the literature [17, 33].

Statistics

Statistical analyses were conducted using SPSS version 20.0 (IBM Corp, Armonk, NY), with a $p \leq 0.05$ used to indicate statistical significance. Patient characteristics were compared using analysis of variance (ANOVA), Kruskal-Wallis ANOVA, and χ^2 . *Post-hoc* *t*-tests and were performed where appropriate.

Concordance between [¹¹C]PIB PET and CSF $\text{A}\beta_{1-42}$ was defined as the proportion of subjects positive or negative for both (i.e., [¹¹C]PIB+ and CSF $\text{A}\beta_{1-42} < 450$ pg/mL or [¹¹C]PIB- and CSF $\text{A}\beta_{1-42} > 450$ pg/mL). Concordance between [¹¹C]PIB and CSF t-tau and p-tau_{181p} was determined in the same fashion, using the cutoffs outlined above. Discordance between [¹¹C]PIB PET and CSF biomarkers was defined as the proportion of cases exhibiting only one abnormal biomarker, with the other being above its respective cutoff. Logistic regression among MCI was used to assess the classification accuracy of [¹¹C]PIB

PET and CSF biomarkers, with progression to dementia due to AD as the dependent variable.

RESULTS

Patient characteristics according to diagnostic group are shown in Tables 1 and 2. Groups did not differ with respect to age, education, gender, or frequency of the APOE $\epsilon 4$ allele. As expected, AD patients had lower MMSE relative to sMCI and pMCI ($p < 0.001$), with pMCI patients showing lower scores relative to sMCI ($p < 0.05$). Using the composite neuropsychological score for global cognition, performance of pMCI patients was below that of sMCI ($p < 0.05$)—as was the case with AD ($p < 0.001$)—with no differences found between pMCI and AD. Using the composite score for episodic memory, performance of pMCI patients was worse than that of sMCI patients ($p < 0.01$), with AD patients performing worse than both sMCI ($p < 0.001$) and pMCI patients (0.01). Regarding use of AChE inhibitors, prescription rates were higher in the AD group (relative to sMCI, $p < 0.001$, pMCI $p < 0.01$). No group differences were found for the period of time between PET and CSF assessments. With respect to the classification of sMCI versus pMCI, clinical follow-up times were longer in sMCI ($p < 0.001$).

Global [¹¹C]PIB PET SUV_R was higher in AD, relative to both sMCI ($p < 0.001$) and pMCI ($p < 0.05$), with levels in pMCI greater than sMCI ($p < 0.01$). In addition, regional analyses showed significant differences in [¹¹C]PIB between sMCI and pMCI in the temporal lobe ($p = 0.05$), the anterior cingulate ($p < 0.001$), the posterior cingulate ($p < 0.001$), the frontal cortex ($p < 0.01$), the parietal cortex ($p < 0.01$), as well as the parahippocampus and insula ($p < 0.01$) Relative to AD, [¹¹C]PIB retention in pMCI differed only in the frontal cortex ($p < 0.05$; see Supplementary Table 1). CSF levels of $\text{A}\beta_{1-42}$ were lower in AD and pMCI,

Table 1
Subject demographics

	sMCI	pMCI	AD
<i>n</i>	21	12	35
Age, years	63.52 ± 8.23	62.33 ± 6.96	67.12 ± 8.82
Education, years	13.10 ± 3.24	13.58 ± 3.40	11.79 ± 3.79
Gender, f	10 (48%)	9 (75%)	21 (60%)

Data are presented as mean ± standard deviation, or as *n* (%). sMCI, stable mild cognitive impairment (i.e., those who had not progressed to dementia at clinical follow-up); pMCI, progressive mild cognitive impairment (i.e., those who had developed to dementia of the Alzheimer's type at clinical follow-up); AD, dementia due to Alzheimer's disease; f, female. Differences between groups were assessed using ANOVA (age, education) and χ^2 (gender).

Table 2
Neuropsychological, clinical, and imaging data according to diagnostic group

	sMCI	pMCI	AD
<i>n</i>	21	12	35
MMSE	28.43 ± 1.32	27.08 ± 1.62 ^{a*}	23.51 ± 3.72 ^{a,b,***}
Global cognition	-0.43 (1.26)	-1.22 (1.84) ^{a***}	-1.72 (2.89) ^{a,b,***}
Episodic memory	-0.59 (1.07)	-1.46 (0.66) ^{a***}	-2.01 (2.50) ^{a,b,***}
Amnesic subtype	8 (38%)	4 (33%)	—
APOE ε4 frequency	12 (57%)	10 (83%)	28 (80%)
AChE inhibitors	4 (19%)	4 (33%)	30 (86%) ^{a***,b**}
Global [¹¹ C]PIB PET SUV _R	1.38 ± 0.3	1.68 ± 0.16 ^{a**}	1.82 ± 0.24 ^{a***,b*}
CSF Aβ ₁₋₄₂	656.20 ± 275.59	490.92 ± 100.37 ^{a*}	439.83 ± 157.02 ^{a**}
CSF t-tau	383.67 ± 196.78	459.67 ± 157.14	563.06 ± 251.03 ^{†,a**}
CSF p-tau	65.26 ± 25.26 [‡]	68.72 ± 18.54 [†]	88.31 ± 32.88 ^{¶,a*}
CSF Aβ ₁₋₄₂ /t-tau	2.26 ± 1.49	1.17 ± 0.39 ^{a**}	0.92 ± 0.54 ^{†,a**}
CSF Aβ ₁₋₄₂ /p-tau _{181p}	12.52 ± 8.07	7.49 ± 2.09 ^{a*}	5.90 ± 2.87 ^{a**}
Time PET to CSF, months	12.49 ± 21.44	5.22 ± 2.99	9.01 ± 9.30
Time PET to FU, months	44.46 ± 22.61 ^{b***}	11.92 ± 12.77	—
Time CSF to FU, months	49.85 ± 27.70 ^{b***}	15.95 ± 11.81	—
% positive [¹¹ C]PIB PET	6 (29%)	12 (100%) ^{a***}	35 (100%) ^{a***}
% positive Aβ ₁₋₄₂	5 (24%)	4 (33%) ^{a*}	21 (60%) ^{a*}
% positive CSF t-tau	7 (33%)	8 (67%) ^{a*}	23 (68%) ^{†,a*}
% positive CSF p-tau _{181p}	4 (27%) [‡]	3 (27%) [†]	14 (54%) ^{¶,a*}
% positive Aβ ₁₋₄₂ /t-tau	6 (29%)	6 (50%) ^{a**}	24 (67%) ^{†,a**}
% positive Aβ ₁₋₄₂ /p-tau _{181p}	3 (20%) [‡]	5 (45%) ^{†,a*}	19 (73%) ^{¶,a**}

Data are presented as mean ± standard deviation, or as *n* (%). sMCI, stable mild cognitive impairment (i.e., those who had not progressed to dementia at clinical follow-up); pMCI, progressive mild cognitive impairment (i.e., those who had developed to dementia of the Alzheimer's type at clinical follow-up); AD, dementia due to Alzheimer's disease; MMSE, Mini-Mental State Examination; Global cognition, composite of the Information and Similarities subscales of the Wechsler Adult Intelligence Scale (language), Block design and Rey Osterrieth copy (visuospatial), Digit Span and Corsi Span (working memory), the Digit Symbol and Trail Making Tests A and B (attention/executive domains); Episodic memory, composite of the Rey Auditory Verbal Learning Test learning and retention, and the Rey Osterrieth retention. Amnesic subtype, amnesic MCI; - Does not apply; AChE inhibitors, acetylcholinesterase inhibitors; Global [¹¹C]PIB PET SUV_R, cutoff >1.41; CSF Aβ₁₋₄₂, cutoff <450 pg/mL; CSF t-tau, cutoff >400 pg/mL; CSF p-tau, cutoff >80 pg/mL; CSF Aβ₁₋₄₂/t-tau, cutoff <1.14; CSF Aβ₁₋₄₂/p-tau, cutoff <6.5; Time PET to FU, time between [¹¹C]PIB PET and clinical follow-up, in months; Time CSF to FU, time between CSF sampling and clinical follow-up, in months. This period can also be considered representative of the total follow-up time since CSF sampling preceded baseline diagnosis by a short, but variable, time interval; - Since the focus of clinical follow-ups in the present study was to determine whether patients with MCI had progressed to dementia due to AD, follow-up times for AD are not reported. Differences between groups were assessed using ANOVA (MMSE, Global cognition, Episodic memory, Global [¹¹C]PIB PET SUV_R, CSF Aβ₁₋₄₂, t-tau, p-tau, Aβ₁₋₄₂/t-tau, Aβ₁₋₄₂/p-tau, Time PET to CSF, Time PET to FU, Time CSF to FU, months) and χ² (Amnesic subtype, APOE ε4 frequency, % positive [¹¹C]PIB PET, CSF Aβ₁₋₄₂, t-tau, p-tau, Aβ₁₋₄₂/t-tau, Aβ₁₋₄₂/p-tau). ^arelative to sMCI, ^brelative to pMCI, ^crelative to AD. **p* < 0.05 ***p* < 0.01 ****p* < 0.001.

294 relative to sMCI (*p* < 0.001 and 0.05, respectively).
 295 Relative to sMCI, CSF levels of total and phospho-
 296 rylated tau were higher in AD (*p* < 0.01 and 0.05,
 297 respectively). Using the ratio of Aβ₁₋₄₂ to t-tau, pMCI
 298 and AD patients showed lower ratio values compared
 299 to sMCI (*p* < 0.01), with the same pattern observed
 300 for Aβ₁₋₄₂/p-tau_{181p} (*p* < 0.05 and 0.01, respectively).
 301 Compared to sMCI, [¹¹C]PIB PET was more often
 302 found to be abnormal in pMCI and AD (*p* < 0.001), as
 303 was the case for CSF Aβ₁₋₄₂ (*p* < 0.05), t-tau (*p* < 0.05),
 304 Aβ₁₋₄₂/t-tau (*p* < 0.01), and CSF Aβ₁₋₄₂/p-tau_{181p}
 305 (*p* < 0.01). Stable and pMCI did not differ in levels of
 306 p-tau_{181p}. Relative to sMCI, levels of p-tau_{181p} were
 307 greater in AD (*p* = 0.05).

308 Across groups, concordance between [¹¹C]PIB PET
 309 and CSF Aβ₁₋₄₂ was 57% (see Table 3, Fig. 1).
 310 Using a higher, more lenient cutoff of <550 pg/mL
 311 for CSF Aβ₁₋₄₂—or a combination of Aβ₁₋₄₂ and
 312 tau (both total and phosphorylated)—concordance
 313 reached 79%, 69%, and 70%, respectively. Within
 314 groups, concordance between [¹¹C]PIB PET and CSF
 315 Aβ₁₋₄₂ was highest for sMCI (67%), followed by AD
 316 (60%), and pMCI (33%). Though the use of this more
 317 lenient cutoff did not increase concordance within the
 318 sMCI group, agreement increased to 83% and 86% in
 319 pMCI and AD, respectively. Using Aβ₁₋₄₂/t-tau and
 320 Aβ₁₋₄₂/p-tau_{181p}, concordance was highest for sMCI
 321 (81% and 80%), followed by AD (71% and 77%), and

Table 3
Concordance between [¹¹C]PIB PET SUV_R and CSF biomarkers

[¹¹ C]PIB PET SUV _R	All	sMCI	pMCI	AD
CSF Aβ ₁₋₄₂ <450	57%	67%	33%	60%
CSF Aβ ₁₋₄₂ <550	79%	67%	83%	86%
CSF t-tau	70% [†]	76%	67%	68% [†]
CSF p-tau _{181p}	58% [§]	87% [‡]	27% [†]	54% [¶]
CSF Aβ ₁₋₄₂ /t-tau	69% [†]	81%	50%	71% [†]
CSF Aβ ₁₋₄₂ /p-tau _{181p}	70% [§]	80% [‡]	45% [†]	77% [¶]

All, all groups. sMCI, stable mild cognitive impairment (i.e. those who had not progressed to dementia at clinical follow-up); pMCI, progressive mild cognitive impairment (i.e. those who had developed to dementia of the Alzheimer's type at clinical follow-up). AD, dementia due to Alzheimer's disease. [¹¹C]PIB PET SUV_R, based on a global cutoff of >1.41. CSF Aβ₁₋₄₂ <450, CSF cut-off of <450 pg/mL. CSF Aβ₁₋₄₂ <550, CSF cut-off of <550 pg/mL. CSF t-tau, CSF cut-off of >400 pg/mL. CSF p-tau_{181p}, CSF cut-off based on >80 pg/mL. CSF Aβ₁₋₄₂/t-tau, CSF cut-off based on <1.14. CSF Aβ₁₋₄₂/p-tau_{181p}, CSF cut-off based on <6.5. [†] data for 1 subject missing. [‡] data for 6 subjects missing. [¶] data for 9 subject missing. [§] data for 16 subjects missing.

322 pMCI (50% and 45%). In terms of discordance, PET
323 was more often positive than CSF Aβ₁₋₄₂, using both
324 450 pg/mL and 550 pg/mL cutoffs (see Figs. 2 and 3).

325 Logistic regression among MCI subjects showed
326 that the classification accuracy of [¹¹C]PIB-PET was

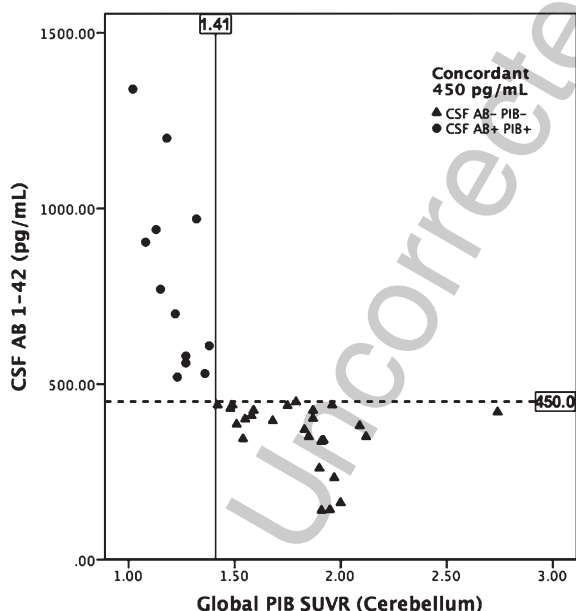


Fig. 1. Concordance between [¹¹C]PIB PET and CSF Aβ₁₋₄₂, using a CSF cutoff of 450 pg/mL. Scatterplot showing concordance between [¹¹C]PIB PET and CSF Aβ₁₋₄₂ (cut-off <450 pg/mL). Triangles indicate [¹¹C]PIB PET-, CSF Aβ₁₋₄₂- subjects; circles indicate [¹¹C]PIB PET+, CSF Aβ₁₋₄₂+ subjects. Horizontal solid line indicates cut-off for abnormal [¹¹C]PIB PET (SUVR >1.41); horizontal dashed line indicates cut-off for abnormal CSF Aβ₁₋₄₂ (<450 pg/mL).

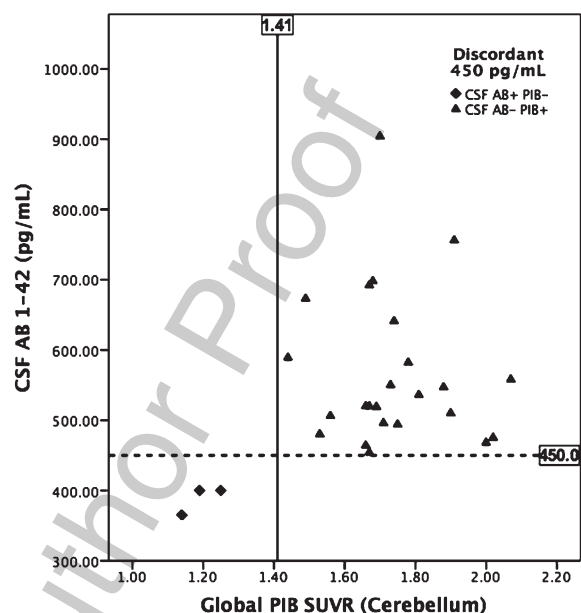


Fig. 2. Concordance between [¹¹C]PIB PET and CSF Aβ₁₋₄₂, using a CSF cutoff of 450 pg/mL. Scatterplot showing discordance between [¹¹C]PIB PET and CSF Aβ₁₋₄₂ (cut-off <450 pg/mL). Diamonds indicate [¹¹C]PIB PET-, CSF Aβ₁₋₄₂+ subjects; triangles indicate [¹¹C]PIB PET+, CSF Aβ₁₋₄₂- subjects. Horizontal solid line indicates cut-off for abnormal [¹¹C]PIB PET (SUVR >1.41); horizontal dashed line indicates cut-off for abnormal CSF Aβ₁₋₄₂ (<450 pg/mL).

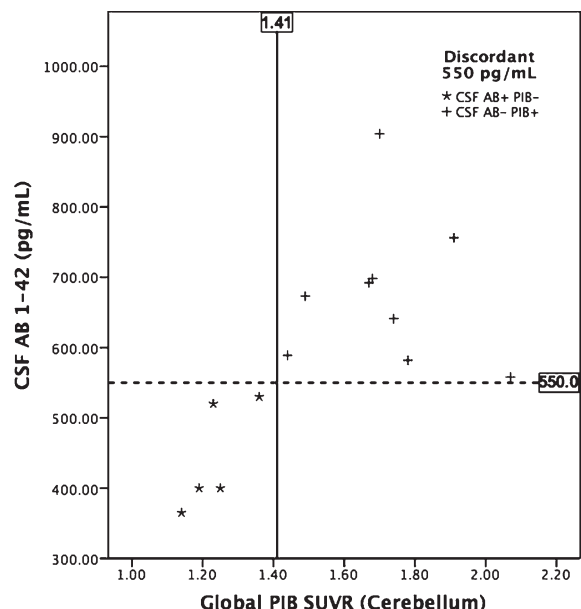


Fig. 3. Concordance between [¹¹C]PIB PET and CSF Aβ₁₋₄₂, using a CSF cutoff of 550 pg/mL. Scatterplot showing discordance between [¹¹C]PIB PET and CSF Aβ₁₋₄₂ (cut-off <550 pg/mL). Stars indicate [¹¹C]PIB PET-, CSF Aβ₁₋₄₂+ subjects; crosses indicate [¹¹C]PIB PET+, CSF Aβ₁₋₄₂- subjects. Horizontal solid line indicates cut-off for abnormal [¹¹C]PIB PET (SUVR >1.41); horizontal dashed line indicates cut-off for abnormal CSF Aβ₁₋₄₂ (<550 pg/mL).

Table 4

Logistic regression derived classification accuracies for [¹¹C]PIB PET and CSF biomarkers, both alone and with use of demographic and clinical covariates

[¹¹ C]PIB PET SUV _R	Accuracy (alone)	<i>p</i>	Accuracy (covariates)	Model ^a	<i>p</i> ^b
Global	73%	**	79%	**	NS
Frontal	73%	*	82%	*	**
Anterior cingulate	73%	**	82%	***	*
Posterior cingulate	79%	**	82%	**	*
Insula	67%	**	79%	**	NS
Parietal	67%	**	70%	**	NS
Temporal	58%	*	82%	*	NS
Caudate nucleus	64%	*	85%	**	*
Putamen	64%	**	88%	**	*
Nucleus accumbens	70%	**	85%	**	*
Hippocampus	64%	NS	79%	*	NS
Parahippocampal	75%	*	72%	*	NS
CSF					
Aβ ₁₋₄₂	58%	NS	82%	**	NS
t-tau	58%	*	82%	*	NS
p-tau _{181p}	54%	NS	70%	NS	NS
Aβ ₁₋₄₂ /t-tau	63%	NS	83%	**	NS
Aβ ₁₋₄₂ /p-tau _{181p}	64%	NS	77%	*	NS

NS, not significant. sMCI, stable mild cognitive impairment (i.e., those who had not progressed to dementia at clinical follow-up); pMCI, progressive mild cognitive impairment (i.e., those who had developed to dementia of the Alzheimer's type at clinical follow-up); AD, dementia due to Alzheimer's disease. ^asignificance level of the model. ^bsignificance level of the variable within the model. Covariates: age, gender, education, MMSE, and APOE ε4 genotype. **p* < 0.05, ***p* < 0.01, ****p* < 0.001.

superior to Aβ₁₋₄₂ (73% versus 58%), t-tau (58%), p-tau_{181p} (54%), Aβ₁₋₄₂/t-tau (63%), and Aβ₁₋₄₂/p-tau_{181p} (65%) (see Table 4). Using regional [¹¹C]PIB uptake, classification accuracy was highest using the posterior cingulate (79%) and parahippocampal gyrus (75%), with additional regions proving similar to or inferior to global [¹¹C]PIB. Inclusion of age, gender, education, MMSE, and APOE ε4 genotype as covariates generally increased classification accuracy of PET (using both global and regional values) and CSF. While the regression models were considered statistically significant when taken together—save for p-tau_{181p}—only [¹¹C]PIB values within the frontal cortex, the cingulate gyrus, the caudate, and the nucleus accumbens within the models themselves cross the threshold for significance (*p* < 0.05).

Characteristics of patients showing discordance between amyloid biomarkers are presented in Table 5. Though raising the cutoff to 550 pg/mL within the group discordant at 450 pg/mL increased agreement, 21% (14 out of 68) of subjects remained discordant. Among discordant subjects, 80% had a positive [¹¹C]PIB scan and normal CSF Aβ₁₋₄₂. Concordant and discordant subjects differed only with

respect to age (concordant 450 pg/mL > discordant 450 pg/mL, *p* < 0.05) and global cognition (concordant 450 pg/mL > discordant 550 pg/mL, *p* < 0.01). As a whole, discordant subjects were within one standard deviation (SD) of the 450 pg/mL cut-off (mean ± SD: 563.54 ± 129.54). While the predominant diagnosis in the group discordant at 450 pg/mL was AD, in those discordant at 550 pg/mL it was sMCI.

DISCUSSION

The present study showed that global and regional [¹¹C]PIB PET proved superior to CSF Aβ₁₋₄₂—both alone, and combined with tau—in terms of differentiating cases of stable MCI from those who progressed to dementia due to AD. Though the classification accuracies of [¹¹C]PIB and CSF measures were comparable following the addition of clinical and demographic covariates, only frontal, cingulate gyrus, caudate nucleus, and putaminal [¹¹C]PIB values retained statistical significance. In this respect, previous work has shown that MCI who progressed to AD had higher [¹¹C]PIB retention in these regions, when compared with MCI who did not progress [34]. We reproduced this finding in the present study, with pMCI subjects showing higher [¹¹C]PIB values within these regions, relative to sMCI subjects. This finding suggests that regional analysis—including, in particular, striatal components—may prove superior to the use of a global [¹¹C]PIB with respect to predicting progression toward AD dementia in patients with MCI.

As expected, the proportion of subjects showing abnormal [¹¹C]PIB and CSF Aβ₁₋₄₂ was greater in pMCI and AD. Using 450 pg/mL as a cutoff, agreement between CSF Aβ₁₋₄₂ and [¹¹C]PIB PET was inferior to that using Aβ₁₋₄₂ to tau ratio values across all groups, with the reverse being true in pMCI and AD when using the more lenient cutoff of 550 pg/mL. In sMCI, agreement between Aβ₁₋₄₂ and [¹¹C]PIB PET was unchanged by cutoff levels, with agreement between [¹¹C]PIB and Aβ₁₋₄₂/tau ratios higher relative to pMCI and AD. These concordance findings reflect the fact that nearly half of the sMCI subjects were amyloid negative, with the syndrome likely the result of non-AD pathology. Comparisons between concordant and discordant subjects proved unremarkable, with only subjects concordant and discordant at 450 pg/mL differing in age.

Our findings using the more lenient Aβ₁₋₄₂ cutoff are in line with previous studies showing high concordance between CSF Aβ₁₋₄₂ and [¹¹C]PIB PET [13, 15, 31, 35–37]. Moreover, our results lend support to

Table 5
 Characteristics of patients showing discordance between [¹¹C]PIB PET and CSF A β ₁₋₄₂

	Concordant 450 pg/mL	Discordant 450pg/mL	Discordant 550 pg/mL
Total	39 (57%)	27 (40%)	14 (21%)
sMCI	14 (36%)	7 (26%)	7 (50%)
pMCI	4 (10%)	8 (30%)	2 (14%)
Amnesic subtype	8 (44%)	6 (40%)	3 (33%)
AD	21 (54%)	12 (44%)	5 (36%)
Age, years	66.82 \pm 8.90 ^{a*}	62.59 \pm 7.57	65.00 \pm 7.16
Education	12.10 \pm 3.78	13.14 \pm 3.17	12.71 \pm 3.14
Gender, f	20 (51%)	18 (67%)	6 (60%)
MMSE	25.58 \pm 3.50	25.34 \pm 3.83	26.21 \pm 3.95
Global cognition	1.15 \pm 2.71	-0.94 \pm 2.10	-0.64 \pm 1.72 ^{b**}
Episodic memory	-1.09 \pm 1.70	-1.23 \pm 1.49	-1.06 \pm 1.49
APOE ϵ 4 frequency	27 (69%)	21 (78%)	11 (79%)
AChE inhibitors	22 (54%)	17 (63%)	8 (57%)
Global [¹¹ C]PIB PET SUV _R	1.64 \pm 0.37	1.68 \pm 0.23	1.55 \pm 0.29
CSF A β ₁₋₄₂	494.10 \pm 265.56	548.03 \pm 117.89	593.42 \pm 149.35
CSF t-tau	463.82 \pm 237.76	525.93 \pm 241.70	488.07 \pm 277.15
CSF p-tau _{181p}	69.72 \pm 25.62 [‡]	84.23 \pm 27.18 [‡]	76.78 \pm 33.67 [‡]
CSF A β ₁₋₄₂ /t-tau	1.45 \pm 1.25	1.31 \pm 0.87	1.67 \pm 1.17
CSF A β ₁₋₄₂ /p-tau _{181p}	8.89 \pm 7.01 [§]	7.38 \pm 2.73 [‡]	10.10 \pm 4.31 [‡]
Time PET to CSF, months	10.36 \pm 15.16	8.63 \pm 12.19	15.17 \pm 24.14

Data are presented as mean \pm standard deviation, mean \pm standard deviation (z-scores), or as n (%). sMCI, stable mild cognitive impairment (i.e. those who had not progressed to dementia at clinical follow-up); pMCI, progressive mild cognitive impairment (i.e., those who had developed to dementia of the Alzheimer's type at clinical follow-up); AD, dementia due to Alzheimer's disease; f, female; MMSE, mini-mental state examination; Global cognition, composite of the Information and Similarities subscales of the Wechsler Adult Intelligence Scale (language), Block design and Rey Osterrieth copy (visuospatial), Digit Span and Corsi Span (working memory), the Digit Symbol and Trail Making Tests A and B (attention/executive domains); Episodic memory, composite of the Rey Auditory Verbal Learning Test learning and retention, and the Rey Osterrieth retention. APOE ϵ 4, apolipoprotein ϵ 4 allele; AChE inhibitors, acetylcholinesterase inhibitor. Global [¹¹C]PIB PET SUV_R, cutoff >1.41; CSF A β ₁₋₄₂, cutoff <450 pg/mL; CSF t-tau, cutoff >400 pg/mL; CSF p-tau, cutoff >80 pg/mL; CSF A β ₁₋₄₂/t-tau, cutoff <1.14; CSF A β ₁₋₄₂/p-tau, cutoff <6.5. Differences between groups were assessed using ANOVA (MMSE, Global cognition, Episodic memory, Global [¹¹C]PIB PET SUV_R, CSF A β ₁₋₄₂, t-tau, p-tau, A β ₁₋₄₂/t-tau, A β ₁₋₄₂/p-tau, Time PET to CSF, months) and χ^2 (Amnesic subtype, APOE ϵ 4 frequency, % positive [¹¹C]PIB PET, CSF A β ₁₋₄₂, t-tau, p-tau, A β ₁₋₄₂/t-tau, A β ₁₋₄₂/p-tau).[‡] data for 5 subjects missing. [§] data for 6 subjects missing. data for 10 subjects missing. ^arelative to subjects discordant at 450 pg/mL. ^brelative to subjects concordant at 450 pg/mL. * p < 0.05 ** p < 0.01.

401 previous work showing that A β ₁₋₄₂/tau ratios are superior to these CSF markers individually with respect to predicting progression to AD dementia in patients with MCI [37]. While the majority of subjects discordant for A β ₁₋₄₂ were within one SD of the threshold for abnormality, the fact that discordant cases were primarily [¹¹C]PIB+ conflicts with the current hypothetical model of AD biomarkers [1], in which concentrations of A β ₁₋₄₂ are thought to cross the threshold for abnormality prior to positive findings using amyloid PET.

411 Given the sizeable body of evidence showing decreased levels of CSF A β ₁₋₄₂ in MCI and AD [5–8, 14, 38–41], our finding of high levels of A β ₁₋₄₂ (i.e., those discordant at 550 pg/mL) in [¹¹C]PIB+ subjects may represent a methodological artifact tied to preanalytical or assay-related factors. Specifically, increased concentrations of CSF A β ₁₋₄₂ have been shown to

418 relate to the time interval between CSF collection and centrifugation, as well as to dilution and buffer factors [39]. The effects of differing calibrator peptides and antibodies within immunoassay kits—as well as batch-to-batch variation—[42] likewise cannot be ruled out since the CSF samples utilized in this study were collected over a relatively long period of time. Indeed, this explanatory stance seems reasonable since similar factors were purported to be at play in the only study to date reporting increased levels of CSF A β ₁₋₄₂ in MCI and AD [43]. Finally, insufficient quality and quantity of sleep have been shown to augment CSF levels of A β ₁₋₄₂ [44], an important consideration given the high prevalence of sleep disorders in AD [45]. As such, sleep insufficiency may have contributed to the discordance between [¹¹C]PIB and CSF A β ₁₋₄₂, though this was not assessed.

418
419
420
421
422
423
424
425
426
427
428
429
430
431
432
433
434

Though [¹¹C]PIB possesses high affinity for fibrillary A β found in cored plaques and cerebral amyloid angiopathy, it binds only weakly to amorphous A β plaques [46, 47]. As such, [¹¹C]PIB may prove unable to detect variants of AD characterized by the predominance of diffuse (nonfibrillar) plaques [48]. In support of this hypothesis, low levels of CSF A β ₁₋₄₂ have been reported in a case of sporadic AD with negative [¹¹C]PIB PET and an abundance of diffuse, but not neuritic (fibrillary), plaques at postmortem, two years following the CSF and [¹¹C]PIB assessments [48]. The low CSF A β ₁₋₄₂ in the absence of [¹¹C]PIB positivity observed in a subset of MCI patients in the present study may therefore reflect the aggregation of A β in diffuse ([¹¹C]PIB-negative) plaques, or the accumulation of oligomeric forms prior to substantial fibrillary ([¹¹C]PIB-positive) A β deposition [49]. Though recent work suggests that [¹¹C]PIB uptake increases over time in some patients with MCI [50], PET studies were conducted after CSF assessment in the present study, suggesting that [¹¹C]PIB-negativity in this subgroup is not simply a reflection of lower disease burden. Finally, though four out of five of the [¹¹C]PIB-/CSF A β ₁₋₄₂₊ subjects were classified as nonamnestic—a subtype typically thought to represent the prodromal phase of non-AD dementias—longitudinal studies highlight a significant rate of progression to AD dementia within this population [51–54]. Further clinical follow-up of these patients will be required to address these possibilities.

Certain methodological aspects, however, limit interpretation of the present findings. In addition to a relatively small sample size, we were unable to obtain measurements for p-tau_{181p} in a number of subjects as this measurement was not routinely done in older CSF samples, with repeat analysis of the original samples an impossibility. Another limitation of this study was the use of literature-derived cutoffs for A β to tau ratios, given the large between center variability reported for CSF measurements [42]. The temporal dissociation between CSF and PET assessments stands as another potential limitation, as ideally the two procedures would have been separated by as short an interval as possible. Several studies have shown, however, that changes in CSF levels of A β ₁₋₄₂, t-tau, and p-tau_{181p} are minimal or absent during both the progression from MCI to AD dementia [28], and during the course of dementia due to AD [54–56]. Taken together, these results suggest that pathologic CSF levels are most likely reached during the preclinical, asymptomatic phase of the disease [28, 55–57]. Even assuming increased levels of fibrillary ([¹¹C]PIB-

positive) amyloid in our MCI subjects [50]—in line with the recent findings mentioned above—between CSF and PET assessments, the fact that CSF sampling was conducted before PET suggests that the discordance observed between CSF A β ₁₋₄₂ and [¹¹C]PIB PET is unlikely to be due the delay between the two assessments.

Despite these caveats, our results suggest that relative to CSF measures of A β ₁₋₄₂ or A β ₁₋₄₂/tau, [¹¹C]PIB PET may prove a better predictor of progression to AD in patients with MCI. Moreover, our findings suggest that evaluation of [¹¹C]PIB-binding in AD signature regions may provide additional diagnostic information relative to global cortical to cerebellar binding. Finally, though the use of a more lenient cutoff greatly increased agreement between CSF A β ₁₋₄₂ and [¹¹C]PIB PET, continued discordance in a subset of MCI patients suggests that these two biomarkers cannot be used interchangeably, as is currently the case.

ACKNOWLEDGMENTS

This study was financially supported by, the Swedish Research Council (project 05817), Swedish Brain Power, the regional agreement on medical training and clinical research (ALF) between Stockholm County Council, The Strategic Research Program in Neuroscience at Karolinska Institutet, Knut and Alice Wallenberg Foundation, the foundation for Old Servants, Gun and Bertil Stohnes Foundation, KI foundations, The Swedish Brain Foundation, the Alzheimer Foundation in Sweden, EU FW6 network of excellence program DiMI (<http://www.dimi.eu>) and within the design of the EU FW7 large scale integrating project INMiND (<http://www.uni-muenster.de/INMiND>), the JPND Project BIOMARKAPD. The authors are grateful to Dr. Jian Fransén for the processing of the [¹¹C]PIB imaging data.

Authors' disclosures available online (<http://j-alz.com/manuscript-disclosures/14-2952r1>).

SUPPLEMENTARY MATERIAL

The supplementary material is available in the electronic version of this article: <http://dx.doi.org/10.3233/JAD-142952>.

REFERENCES

- [1] Jack CR Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, Shaw LM, Vemuri P, Wiste HJ, Weigand SD, Lesnick TG, Pankratz VS, Donohue MC, Trojanowski JQ

- (2013) Tracking pathophysiological processes in Alzheimer's disease: An updated hypothetical model of dynamic biomarkers. *Lancet Neurol* **12**, 207-216.
- [2] Skoog I, Davidsson P, Aevansson O, Vanderstichele H, Vanmechelen E, Blennow K (2003) Cerebrospinal fluid beta-amyloid 42 is reduced before the onset of sporadic dementia: A population-based study in 85-year-olds. *Dement Geriatr Cogn Disord* **15**, 169-176.
- [3] Gustafson DR, Skoog I, Rosengren L, Zetterberg H, Blennow K (2007) Cerebrospinal fluid beta-amyloid 1-42 concentration may predict cognitive decline in older women. *J Neurol Neurosurg Psychiatry* **78**, 461-464.
- [4] Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC, Marcus DS, Cairns NJ, Xie X, Blazey TM, Holtzman DM, Santacruz A, Buckles V, Oliver A, Moulder K, Aisen PS, Ghetti B, Klunk WE, McDade E, Martins RN, Masters CL, Mayeux R, Ringman JM, Rossor MN, Schofield PR, Sperling RA, Salloway S, Morris JC, Dominantly Inherited Alzheimer N (2012) Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N Engl J Med* **367**, 795-804.
- [5] Blennow K, Hampel H (2003) CSF markers for incipient Alzheimer's disease. *Lancet Neurol* **2**, 605-613.
- [6] Clark CM, Xie S, Chittams J, Ewbank D, Peskind E, Galasko D, Morris JC, McKeel DW Jr, Farlow M, Weitlauf SL, Quinn J, Kaye J, Knopman D, Arai H, Doody RS, DeCarli C, Leight S, Lee VM, Trojanowski JQ (2003) Cerebrospinal fluid tau and beta-amyloid: How well do these biomarkers reflect autopsy-confirmed dementia diagnoses? *Arch Neurol* **60**, 1696-1702.
- [7] de Leon MJ, DeSanti S, Zinkowski R, Mehta PD, Pratico D, Segal S, Rusinek H, Li J, Tsui W, Saint Louis LA, Clark CM, Tarshish C, Li Y, Lair L, Javier E, Rich K, Lesbre P, Mosconi L, Reisberg B, Sadowski M, DeBernadis JF, Kerkman DJ, Hampel H, Wahlund LO, Davies P (2006) Longitudinal CSF and MRI biomarkers improve the diagnosis of mild cognitive impairment. *Neurobiol Aging* **27**, 394-401.
- [8] Fagan AM, Shaw LM, Xiong C, Vanderstichele H, Mintun MA, Trojanowski JQ, Coart E, Morris JC, Holtzman DM (2011) Comparison of analytical platforms for cerebrospinal fluid measures of beta-amyloid 1-42, total tau, and p-tau181 for identifying Alzheimer disease amyloid plaque pathology. *Arch Neurol* **68**, 1137-1144.
- [9] Driscoll I, Troncoso JC, Rudow G, Sojkova J, Pletnikova O, Zhou Y, Kraut MA, Ferrucci L, Mathis CA, Klunk WE, O'Brien RJ, Davatzikos C, Wong DF, Resnick SM (2012) Correspondence between *in vivo* (11)C-PiB-PET amyloid imaging and postmortem, region-matched assessment of plaques. *Acta Neuropathol* **124**, 823-831.
- [10] Kadir A, Marutle A, Gonzalez D, Scholl M, Almkvist O, Mousavi M, Mustafiz T, Darreh-Shori T, Nennesmo I, Nordberg A (2011) Positron emission tomography imaging and clinical progression in relation to molecular pathology in the first Pittsburgh Compound B positron emission tomography patient with Alzheimer's disease. *Brain* **134**, 301-317.
- [11] Kemppainen NM, Aalto S, Wilson IA, Nagren K, Helin S, Bruck A, Oikonen V, Kailajarvi M, Scheinin M, Viitanen M, Parkkola R, Rinne JO (2007) PET amyloid ligand [¹¹C]PIB uptake is increased in mild cognitive impairment. *Neurology* **68**, 1603-1606.
- [12] Forsberg A, Engler H, Almkvist O, Blomquist G, Hagman G, Wall A, Ringheim A, Langstrom B, Nordberg A (2008) PET imaging of amyloid deposition in patients with mild cognitive impairment. *Neurobiol Aging* **29**, 1456-1465.
- [13] Fagan AM, Mintun MA, Mach RH, Lee SY, Dence CS, Shah AR, LaRossa GN, Spinner ML, Klunk WE, Mathis CA, DeKosky ST, Morris JC, Holtzman DM (2006) Inverse relation between *in vivo* amyloid imaging load and cerebrospinal fluid Abeta42 in humans. *Ann Neurol* **59**, 512-519.
- [14] Fagan AM, Roe CM, Xiong C, Mintun MA, Morris JC, Holtzman DM (2007) Cerebrospinal fluid tau/beta-amyloid(42) ratio as a prediction of cognitive decline in nondemented older adults. *Arch Neurol* **64**, 343-349.
- [15] Jagust WJ, Landau SM, Shaw LM, Trojanowski JQ, Koeppe RA, Reiman EM, Foster NL, Petersen RC, Weiner MW, Price JC, Mathis CA, Alzheimer's Disease Neuroimaging I (2009) Relationships between biomarkers in aging and dementia. *Neurology* **73**, 1193-1199.
- [16] Zwan M, van Harten A, Ossenkuppe R, Bouwman F, Teunissen C, Adriaanse S, Lammertsma A, Scheltens P, van Berckel B, van der Flier W (2014) Concordance between cerebrospinal fluid biomarkers and [¹¹C]PIB PET in a memory clinic cohort. *J Alzheimers Dis* **41**, 801-807.
- [17] Koivunen J, Pirttila T, Kemppainen N, Aalto S, Herukka SK, Jauhianen AM, Hanninen T, Hallikainen M, Nagren K, Rinne JO, Soininen H (2008) PET amyloid ligand [¹¹C]PIB uptake and cerebrospinal fluid beta-amyloid in mild cognitive impairment. *Dement Geriatr Cogn Disord* **26**, 378-383.
- [18] Wolk DA, Zhang Z, Boudhar S, Clark CM, Pontecorvo MJ, Arnold SE (2012) Amyloid imaging in Alzheimer's disease: Comparison of florbetapir and Pittsburgh compound-B positron emission tomography. *J Neurol Neurosurg Psychiatry* **83**, 923-926.
- [19] Mattsson N, Insel PS, Landau S, Jagust W, Donohue M, Shaw LM, Trojanowski JQ, Zetterberg H, Blennow K, Weiner M, the Alzheimer's Disease Neuroimaging, Initiative (2014) Diagnostic accuracy of CSF Aβ42 and florbetapir PET for Alzheimer's disease. *Ann Clin Transl Neurol* **1**, 534-543.
- [20] McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR Jr, Kawas CH, Klunk WE, Koroshetz WJ, Manly JJ, Mayeux R, Mohs RC, Morris JC, Rossor MN, Scheltens P, Carrillo MC, Thies B, Weintraub S, Phelps CH (2011) The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* **7**, 263-269.
- [21] Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, Gamst A, Holtzman DM, Jagust WJ, Petersen RC, Snyder PJ, Carrillo MC, Thies B, Phelps CH (2011) The diagnosis of mild cognitive impairment due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* **7**, 27-279.
- [22] Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E (1999) Mild cognitive impairment: Clinical characterization and outcome. *Arch Neurol* **56**, 303-308.
- [23] Winblad B, Palmer K, Kivipelto M, Jelic V, Fratiglioni L, Wahlund LO, Nordberg A, Backman L, Albert M, Almkvist O, Arai H, Basun H, Blennow K, de Leon M, DeCarli C, Erkinjuntti T, Giacobini E, Graff C, Hardy J, Jack C, Jorm A, Ritchie K, van Duijn C, Visser P, Petersen RC (2004) Mild cognitive impairment—beyond controversies, towards a consensus: Report of the International Working Group on Mild Cognitive Impairment. *J Intern Med* **256**, 240-246.
- [24] McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM (1984) Clinical diagnosis of Alzheimer's disease: Report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* **34**, 939-944.

- 663 [25] Bergman I, Blomberg M, Almkvist O (2007) The importance of impaired physical health and age in normal cognitive aging. *Scand J Psychol* **48**, 115-125. 728
- 664
- 665 [26] Nordberg A, Carter SF, Rinne J, Drzezga A, Brooks DJ, Vandenberghe R, Perani D, Forsberg A, Langstrom B, Scheinin N, Karrasch M, Nagren K, Grimmer T, Miederer I, Edison P, Okello A, Van Laere K, Nelissen N, Vandenbulcke M, Garibotto V, Almkvist O, Kalbe E, Hinz R, Herholz K (2013) A European multicentre PET study of fibrillar amyloid in Alzheimer's disease. *Eur J Nucl Med Mol Imaging* **40**, 104-114. 729
- 666
- 667
- 668
- 669
- 670
- 671
- 672
- 673
- 674 [27] Blennow K, Wallin A, Agren H, Spenger C, Siegfried J, Vanmechelen E (1995) Tau protein in cerebrospinal fluid: A biochemical marker for axonal degeneration in Alzheimer disease? *Mol Chem Neuropathol* **26**, 231-245. 730
- 675
- 676
- 677
- 678 [28] Andreassen N, Minthon L, Vanmechelen E, Vanderstichele H, Davidsson P, Winblad B, Blennow K (1999) Cerebrospinal fluid tau and Abeta42 as predictors of development of Alzheimer's disease in patients with mild cognitive impairment. *Neurosci Lett* **273**, 5-8. 731
- 679
- 680
- 681
- 682
- 683 [29] Vanmechelen E, Vanderstichele H, Davidsson P, Van Kerschaver E, Van Der Perre B, Sjogren M, Andreassen N, Blennow K (2000) Quantification of tau phosphorylated at threonine 181 in human cerebrospinal fluid: A sandwich ELISA with a synthetic phosphopeptide for standardization. *Neurosci Lett* **285**, 49-52. 732
- 684
- 685
- 686
- 687
- 688
- 689 [30] Carter SF, Scholl M, Almkvist O, Wall A, Engler H, Langstrom B, Nordberg A (2012) Evidence for astrogliosis in prodromal Alzheimer disease provided by 11C-deuterium-L-deprenyl: A multitracers PET paradigm combining 11C-Pittsburgh compound B and 18F-FDG. *J Nucl Med* **53**, 37-46. 733
- 690
- 691
- 692
- 693
- 694
- 695 [31] Forsberg A, Almkvist O, Engler H, Wall A, Langstrom B, Nordberg A (2010) High PIB retention in Alzheimer's disease is an early event with complex relationship with CSF biomarkers and functional parameters. *Curr Alzheimer Res* **7**, 56-66. 734
- 696
- 697
- 698
- 699
- 700 [32] Biomarkers for Alzheimer's and Parkinson's disease, <http://biomarkapd.org/what-is-biomarkapd/abstract-3/>. Last Updated November 2014, Accessed November 1, 2014. 735
- 701
- 702
- 703 [33] Vos S, van Rossum I, Burns L, Knol D, Scheltens P, Soinenen H, Wahlund LO, Hampel H, Tsolaki M, Minthon L, Handels R, L'Italien G, van der Flier W, Aalten P, Teunissen C, Barkhof F, Blennow K, Wolz R, Rueckert D, Verhey F, Visser PJ (2012) Test sequence of CSF and MRI biomarkers for prediction of AD in subjects with MCI. *Neurobiol Aging* **33**, 2272-2281. 736
- 704
- 705
- 706
- 707
- 708
- 709 [34] Koivunen J, Scheinin N, Virta JR, Aalto S, Vahlberg T, Nagren K, Helin S, Parkkola R, Viitanen M, Rinne JO (2011) Amyloid PET imaging in patients with mild cognitive impairment: A 2-year follow-up study. *Neurology* **76**, 1085-1090. 737
- 710
- 711
- 712
- 713 [35] Grimmer T, Riemenschneider M, Forstl H, Henriksen G, Klunk WE, Mathis CA, Shiga T, Wester HJ, Kurz A, Drzezga A (2009) Beta amyloid in Alzheimer's disease: Increased deposition in brain is reflected in reduced concentration in cerebrospinal fluid. *Biol Psychiatry* **65**, 927-934. 738
- 714
- 715
- 716
- 717
- 718 [36] Tolboom N, van der Flier WM, Yaqub M, Boellaard R, Verwey NA, Blankenstein MA, Windhorst AD, Scheltens P, Lammertsma AA, van Berckel BN (2009) Relationship of cerebrospinal fluid markers to 11C-PiB and 18F-FDDNP binding. *J Nucl Med* **50**, 1464-1470. 739
- 719
- 720
- 721
- 722
- 723 [37] Degerman Gunnarsson M, Lindau M, Wall A, Blennow K, Dareh-Shori T, Basu S, Nordberg A, Larsson A, Lannfelt L, Basun H, Kilander L (2010) Pittsburgh compound-B and Alzheimer's disease biomarkers in CSF, plasma and urine: An exploratory study. *Dement Geriatr Cogn Disord* **29**, 204-212. 740
- 724
- 725
- 726
- 727
- 728 [38] Hansson O, Zetterberg H, Buchhave P, Londos E, Blennow K, Minthon L (2006) Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: A follow-up study. *Lancet Neurol* **5**, 228-234. 741
- 729
- 730
- 731 [39] Galasko D, Chang L, Motter R, Clark CM, Kaye J, Knopman D, Thomas R, Kholodenko D, Schenk D, Lieberburg I, Miller B, Green R, Basherad R, Kertiles L, Boss MA, Seubert P (1998) High cerebrospinal fluid tau and low amyloid beta42 levels in the clinical diagnosis of Alzheimer disease and relation to apolipoprotein E genotype. *Arch Neurol* **55**, 937-945. 742
- 732
- 733
- 734
- 735
- 736
- 737
- 738
- 739 [40] Shaw LM, Vanderstichele H, Knapik-Czajka M, Clark CM, Aisen PS, Petersen RC, Blennow K, Soares H, Simon A, Lewczuk P, Dean R, Siemers E, Potter W, Lee VM, Trojanowski JQ, Alzheimer's Disease Neuroimaging I (2009) Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. *Ann Neurol* **65**, 403-413. 743
- 740
- 741
- 742
- 743
- 744
- 745
- 746 [41] Bjerke M, Portelius E, Minthon L, Wallin A, Anckarsater H, Anckarsater R, Andreassen N, Zetterberg H, Andreasson U, Blennow K (2010) Confounding factors influencing amyloid Beta concentration in cerebrospinal fluid. *Int J Alzheimers Dis* **2010**, pii: 986310. 746
- 747
- 748
- 749
- 750
- 751 [42] Mattsson N, Andreasson U, Persson S, Arai H, Batish SD, Bernardini S, Bocchio-Chiavetto L, Blankenstein MA, Carrillo MC, Chalbot S, Coart E, Chiasserini D, Cutler N, Dahlfors G, Duller S, Fagan AM, Forlenza O, Frisoni GB, Galasko D, Galimberti D, Hampel H, Handberg A, Heneka MT, Herskovits AZ, Herukka SK, Holtzman DM, Humpel C, Hyman BT, Iqbal K, Jucker M, Kaeser SA, Kaiser E, Kapaki E, Kidd D, Klivenyi P, Knudsen CS, Kummer MP, Lui J, Llado A, Lewczuk P, Li QX, Martins R, Masters C, McAuliffe J, Mercken M, Moghekar A, Molinuevo JL, Montine TJ, Nowatzke W, O'Brien R, Otto M, Paraskevas GP, Parnetti L, Petersen RC, Prvulovic D, de Reus HP, Rissman RA, Scarpini E, Stefani A, Soinenen H, Schroder J, Shaw LM, Skinningsrud A, Skrogstad B, Spreer A, Talib L, Teunissen C, Trojanowski JQ, Tumani H, Umek RM, Van Broeck B, Vanderstichele H, Vecsei L, Verbeek MM, Windisch M, Zhang J, Zetterberg H, Blennow K. (2011) The Alzheimer's Association external quality control program for cerebrospinal fluid biomarkers. *Alzheimers Dement* **7**, 386-395 e386. 749
- 752
- 753
- 754
- 755
- 756
- 757
- 758
- 759
- 760
- 761
- 762
- 763 [43] Bouwman FH, van der Flier WM, Schoonenboom NS, van Elk EJ, Kok A, Rijmen F, Blankenstein MA, Scheltens P (2007) Longitudinal changes of CSF biomarkers in memory clinic patients. *Neurology* **69**, 1006-1011. 764
- 764
- 765
- 766
- 767
- 768
- 769
- 770 [44] Ooms S, Overeem S, Besse K, Rikkert MO, Verbeek M, Claassen JA (2014) Effect of 1 night of total sleep deprivation on cerebrospinal fluid beta-amyloid 42 in healthy middle-aged men: A randomized clinical trial. *JAMA Neurol* **71**, 971-977. 770
- 771
- 772
- 773
- 774
- 775
- 776
- 777
- 778
- 779 [45] Vitiello MV, Borson S (2001) Sleep disturbances in patients with Alzheimer's disease: Epidemiology, pathophysiology and treatment. *CNS Drugs* **15**, 777-796. 779
- 780
- 781
- 782 [46] Ikonomic MD, Klunk WE, Abrahamson EE, Mathis CA, Price JC, Tsopelas ND, Lopresti BJ, Ziolkowski S, Bi W, Paljug WR, Debnath ML, Hope CE, Isanski BA, Hamilton RL, DeKosky ST (2008) Post-mortem correlates of *in vivo* PiB-PET amyloid imaging in a typical case of Alzheimer's disease. *Brain* **131**, 1630-1645. 782
- 783
- 784
- 785
- 786
- 787
- 788 [47] Backsai BJ, Frosch MP, Freeman SH, Raymond SB, Augustinack JC, Johnson KA, Irizarry MC, Klunk WE, Mathis CA, Dekosky ST, Greenberg SM, Hyman BT, Growdon JH (2007) Molecular imaging with Pittsburgh Compound B confirmed at autopsy: A case report. *Arch Neurol* **64**, 431-434. 788
- 789
- 790
- 791
- 792

- 793 [48] Cairns NJ, Ikonovic MD, Benzinger T, Storandt M, Fagan 816
 794 AM, Shah AR, Reinwald LT, Carter D, Felton A, Holtzman 817
 795 DM, Mintun MA, Klunk WE, Morris JC (2009) Absence of 818
 796 Pittsburgh compound B detection of cerebral amyloid beta 819
 797 in a patient with clinical, cognitive, and cerebrospinal fluid 820
 798 markers of Alzheimer disease: A case report. *Arch Neurol* **66**, 821
 799 1557-1562. 822
- 800 [49] Blennow K, Zetterberg H, Fagan AM (2012) Fluid biomark- 823
 801 ers in Alzheimer disease. *Cold Spring Harb Perspect Med* **2**, 824
 802 a006221. 825
- 803 [50] Kemppainen NM, Scheinin NM, Koivunen J, Johansson J, 826
 804 Toivonen JT, Nagren K, Rokka J, Karrasch M, Parkkola R, 827
 805 Rinne JO (2014) Five-year follow-up of 11C-PIB uptake in 828
 806 Alzheimer's disease and MCI. *Eur J Nucl Med Mol Imaging* 829
 807 **41**, 283-289. 830
- 808 [51] Busse A, Hensel A, Guhne U, Angermeyer MC, Riedel-Heller 831
 809 SG (2006) Mild cognitive impairment: Long-term course of 832
 810 four clinical subtypes. *Neurology* **67**, 2176-2185. 833
- 811 [52] Fischer P, Jungwirth S, Zehetmayer S, Weissgram S, 834
 812 Hoenigschnabl S, Gelpi E, Krampla W, Tragl KH (2007) 835
 813 Conversion from subtypes of mild cognitive impairment to 836
 814 Alzheimer dementia. *Neurology* **68**, 288-291. 837
- 815 [53] Rountree SD, Waring SC, Chan WC, Lupo PJ, Darby 838
 EJ, Doody RS (2007) Importance of subtle amnestic and
 nonamnestic deficits in mild cognitive impairment: Prognosis
 and conversion to dementia. *Dement Geriatr Cogn Disord* **24**,
 476-482.
- [54] Lopez OL, Kuller LH, Becker JT, Dulberg C, Sweet RA, Gach
 HM, Dekosky ST (2007) Incidence of dementia in mild cog-
 nitive impairment in the cardiovascular health study cognition
 study. *Arch Neurol* **64**, 416-420.
- [55] Zetterberg H, Pedersen M, Lind K, Svensson M, Rolstad
 S, Eckerstrom C, Syversen S, Mattsson UB, Ysander C,
 Mattsson N, Nordlund A, Vanderstichele H, Vanmeche-
 len E, Jonsson M, Edman A, Blennow K, Wallin A
 (2007) Intra-individual stability of CSF biomarkers for
 Alzheimer's disease over two years. *J Alzheimers Dis* **12**,
 255-260.
- [56] Blennow K, Zetterberg H, Minthon L, Lannfelt L, Strid S,
 Annas P, Basun H, Andreasen N (2007) Longitudinal stability
 of CSF biomarkers in Alzheimer's disease. *Neurosci Lett* **419**,
 18-22.
- [57] Mattsson N, Portelius E, Rolstad S, Gustavsson M, Andreas-
 son U, Stridsberg M, Wallin A, Blennow K, Zetterberg H
 (2012) Longitudinal cerebrospinal fluid biomarkers over four
 years in mild cognitive impairment. *J Alzheimers Dis* **30**,
 767-778.

Uncorrected Author Proof