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An overview of genera and subgenera of the *Asura* / *Miltochrista* generic complex (Lepidoptera, Erebidae, Arctiinae). Part 1. *Barsine* Walker, 1854 *sensu lato*, *Asura* Walker, 1854 and related genera, with descriptions of twenty new genera, ten new subgenera and a check list of taxa of the *Asura* / *Miltochrista* generic complex

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Abstract

Lithosiini genera of the *Asura* / *Miltochrista* generic complex related to *Barsine* Walker, 1854 *sensu lato* and *Asura* Walker, 1854 are overviewed. *Barsine* is considered to be a group having such an autapomorphic feature as a basal saccular process of valva only. Many species without this process are separated to the diverse and species-rich genus *Ammatho* **stat. nov.**, which is subdivided here into eight subgenera including *Idopterum* Hampson, 1894 downgraded here to a subgenus level, and six new subgenera: *Ammathella* Volynkin, **subgen. nov.**, *Composine* Volynkin, **subgen. nov.**, *Striatella* Volynkin & Huang, **subgen. nov.**, *Conicornuta* Volynkin, **subgen. nov.**, *Delineatia* Volynkin & Huang, **subgen. nov.** and *Rugosine* Volynkin, **subgen. nov.** A number of groups of species considered previously by various authors as members of *Barsine* are erected here to 20 new genera and four subgenera: *Ovipennis* (*Barsipennis*) Volynkin, **subgen. nov.**, *Ovipennis* (*Coccinigripennis*) Volynkin & Huang, **subgen. nov.**, *Barsura* (*Tenebrasura*) Volynkin, **subgen. nov.**, *Argentosine* Volynkin, **gen. nov.**, *Esmasura* Volynkin & Huang, **gen. nov.**, *Matsumursine* Volynkin & Huang, **gen. nov.**, *Floridasura* Volynkin, **gen. nov.**, *Fossia* Volynkin, Ivanova & Huang, **gen. nov.**, *Wittasura* Volynkin, **gen. nov.**, *Disparsine* Volynkin, **gen. nov.**, *Moorasura* Volynkin & Huang, **gen. nov.**, *Sarbine* Volynkin, **gen. nov.**, *Sarbine* (*Processine*) Volynkin, **subgen. nov.**, *Hampsonascia* Volynkin, **gen. nov.**, *Cernysura* Volynkin, **gen. nov.**, *Barsilene* Volynkin & Huang, **gen. nov.**, *Nanarsine* Volynkin, **gen. nov.**, *Amphisine* Volynkin, **gen. nov.**, *Karolia* Volynkin, **gen. nov.**, *Niveutane* Volynkin, **gen. nov.**, *Rubrindiania* Volynkin & Huang, **gen. nov.**, *Barsaurea* Volynkin & Huang, **gen. nov.**, *Integrivalvia* Volynkin & Huang, **gen. nov.** and *Aberrasine* Volynkin & Huang, **gen. nov.** The genus *Nebulene* Volynkin & Černý is downgraded to a subspecies of *Ovipennis*. The genus *Eutane* Walker, 1854 is downgraded to a subspecies of *Asura*. The genera *Miltasura* Roepke, 1946 and *Gymnasura* Hampson, 1900 are synonymised here with *Cyme* Felder, 1861 and *Asura* respectively. The genera *Asuropsis* Matsumura, 1927, *Neasuroides* Matsumura, 1927 and *Asuridoides* Daniel, 1951 are synonymised with *Miltochrista* Hübner, [1819]. The genus *Allochrista* Roepke, 1946 is synonymised with the subgenus *Thyrgorina* Walker, [1865] of the genus *Lemyra* Walker, 1856 (tribe Arctiini) with establishing a new combination *Lemyra* (*Thyrgorina*) *toxopei* (Roepke, 1946), **comb. nov.** Other six new synonyms are established: *Barsine pardalis* (Mell, 1922) = *Barsine miranda* Kishida & Wang, 2017, **syn. nov.**, *Barsine striata striata* (Bremer & Grey, 1852) = *Miltochrista quelparta* Okamota,

1924, **syn. nov.**, *Floridasura tricolor* (Wileman, 1910) = *Barsine coccinea* Moore, 1886, **syn. nov.**, *Disasuridia metaphaea* (Hampson, 1900) = *Disasuridia flava* Fang, 1991, **syn. nov.**, *Aberrasine aberrans aberrans* (Butler, 1877) = *Miltochrista decussata* (Moore, 1877), **syn. nov.** and *Cabarda nigripuncta* (Wileman & South, 1919) = *Asura lunilinea* Schaus, 1922, **syn. nov.** In addition, it is stated that *Miltochrista quadrifasciata* Rothschild, 1913 described from New Guinea and currently belonging to the genus *Cyme* (a junior synonym of *Cyme sexualis* (Felder, 1864)) is an invalid name, being a secondary junior homonym (**homonym nov.**) of *Cyme quadrifasciata* (Rothschild, 1913), **comb. nov.** described from Sulawesi. A full check-list of members of the *Asura* / *Miltochrista* generic complex with 370 new combinations is present.

Key words: Lithosiini, Nudariina, new genus, new subgenus, new status, new synonym, new homonym, new combination.

Introduction

The *Asura* / *Miltochrista* generic complex is a diverse and very species-rich group within the subtribe Nudariina of the tribe Lithosiini. It is widespread in Afrotropical, Oriental, eastern Palaearctic and Australian Regions with only species known from western Palaearctic. According to the latest data, the generic complex comprised 43 genera: *Barsine* Walker, 1854, *Cyme* Felder, 1861, *Miltasura* Roepke, 1946, *Idopterum* Hampson, 1894, *Ovipennis* Hampson, 1900, *Nebulene* Volynkin & Černý, 2018, *Barsochrista* N. Singh & Kirti, 2016, *Asuridia* Hampson, 1900, *Barsura* Volynkin, Dubatolov & Kishida, 2017, *Pseudobarsine* N. Singh & Kirti, 2016, *Nepita* Moore, [1860], *Eutane* Walker, 1854, *Asura* Walker, 1854, *Gymnasura* Hampson, 1900, *Graptasura* Hampson, 1900, *Xanthetis* Hampson, 1900, *Disasuridia* Fang, 1991, *Quadrasura* Holloway, 2001, *Melanaema* Butler, 1877, *Longarista* Volynkin, 2019, *Indiania* Kirti, Joshi & N. Singh, 2014, *Albarrania* Bolotov, Spitsyn & Kondakov, 2019, *Arctelene* N. Singh Kirti & Gill, 2008, *Parvuspina* N. Singh, Kirti & Datta, 2019, *Sesapa* Walker, 1854 with subgenera *Sesapa* and *Nipponasura* Inoue, 1965, *Chrysasura* Hampson, 1914, *Trichocerosia* Hampson, 1900, *Chiretolpis* Watson, 1980, *Micronyctemera* de Vos & van Mastrigt, 2007, *Symmetrodes* Meyrick, 1886, *Miltochrista* Hübner, [1819], *Asuropsis* Matsumura, 1927, *Neasuroides* Matsumura, 1927, *Asuridoides* Daniel, 1951, *Cabarda* Walker, 1863, *Gurna* Swinhoe, 1892, *Adites* Moore, [1882], *Pseudoadites* N. Singh & Kirti, 2016, *Afrasura* Durante, 2009, *Palaeugoa* Durante, 2012, *Tumicla* Wallengren, 1863, *Parafrasura* Durante, 2012 and *Allochrista* Roepke, 1946. The taxonomy of this group still remains poorly studied and many new taxa are still awaiting their discovery and description. Numerous new species and genera of the *Asura* / *Miltochrista* generic complex have been described during last two decades (Fang 2000; Holloway 2001; Kaleka 2003; de Vos & Mastrigt 2007; Kühne 2007; Durant 2008; 2009; 2012; Kirti & Gill 2008; 2009; Černý & Pinratana 2009; Bucsek 2012; 2014; Dubatolov *et al*. 2012; Dubatolov & Bucsek 2013; 2014; Wu & Fu 2013; Wu *et al.* 2013; Kirti *et al.* 2013; 2014; Kirti & Singh 2015, 2016; Černý 2016; Volynkin & Bucsek 2016; Volynkin & Černý 2016a; 2016b; 2016c; 2017a; 2017b; 2017c; 2017d; 2018a; 2018b; 2018c; 2019a; 2019b); Joshi *et al.* 2017; 2018; Kishida & Wang 2017; Volynkin 2017a; 2018b; 2017c; 2017d; 2018a; 2018b; 2019a; 2019b; 2019c; Volynkin & László 2018a; 2018b; 2019; Volynkin *et al.* 2018a; 2018b; 2019a; 2019b; 2019c; 2019d; Spitsyn & Bolotov 2018; Spitsyn *et al.* 2018; Huang *et al.* 2018; Bayarsaikhan *et al.* 2019; Bolotov *et al.* 2019; Singh *et al.* 2019; Wu 2019). Despite this fact, some genera of the group, especially *Barsine* and *Miltochrista*, remain polyphyletic and comprising a large amount of unrelated species. On the other hand, until now, some genera of the generic complex were unstudied and their genitalia structures were unknown. During our studies, we examined male and female genitalia of type species of described genera and many other species of the generic complex as well. The accumulated data allowed us to revise the generic and subgeneric structure of the complex, separating groups of related species united by autapomorphic features or/and combinations of common genital features.

The present paper is a first step in ordering the *Asura* / *Miltochrista* generic complex devoted to the genus *Barsine* in wide sense and its relatives including *Asura*. Here we consider the genus *Barsine* to be comprising species having such an autapomorphic feature as a basal saccular process of valva only. A lot of species without this process are separated to the diverse and species-rich genus *Ammatho* **stat. nov.**, which is subdivided here into eight subgenera including *Idopterum* downgraded here to a subgenus level. Many groups of species considered previously by various authors as members of *Barsine* are erected here to separated genera. The genus *Nebulene* Volynkin & Černý is downgraded to a subspecies of *Ovipennis*. The

genus *Eutane* Walker, 1854 is downgraded to a subspecies of *Asura*. The genera *Miltasura* and *Gymnasura* are synonymised here with *Cyme* and *Asura* respectively. In addition, the "*Eutane*" *nivea* species-group is erected to the new genus *Niveutane* Volynkin, **gen. nov.**

The genus *Miltochrista* and its relatives, namely *Arctelene* N. Singh Kirti & Gill, 2008, *Parvuspina* N. Singh, Kirti & Datta, 2019, *Sesapa* Walker, 1854, *Chrysasura* Hampson, 1914, *Trichocerosia* Hampson, 1900, *Chiretolpis* Watson, 1980, *Micronyctemera* de Vos & van Mastrigt, 2007, *Symmetrodes* Meyrick, 1886, *Cabarda* Walker, 1863, *Gurna* Swinhoe, 1892, *Adites* Moore, [1882], *Pseudoadites* N. Singh & Kirti, 2016 and *Afrasura* Durante, 2009 are not included to this paper. The taxonomy of this group will be revised in the next paper on the *Asura* / *Miltochrista* generic complex (Volynkin, in prep.). However, it is necessary to provide some notes on this complex in the present paper. In their book, Kirti & Singh (2016) synonymised the genus *Lyclene* Moore, [1860] with *Miltochrista* and established new combinations for all Indian taxa. Later, in several papers devoted to descriptions of various new species of *Miltochrista*, some other species previously considered as members of *Lyclene* were transferred to *Miltochrista* (Joshi *et al.* 2017; Volynkin 2017; Singh *et al.* 2019). Nevertheless, a full check-list of the genus *Miltochrista* was never published and many taxa still remain to be associated with *Lyclene* without establishing of new combinations under *Miltochrista*. In the present paper we provide a full check-list of members of the *Asura* / *Miltochrista* generic complex and establish 370 new combinations. Unfortunately, some taxa of species rank remain unstudied by us therefore their current generic placement is provisional and will be a matter of forthcoming researches. The genera *Asuropsis*, *Neasuroides* and *Asuridoides* are synonymised here with *Miltochrista*.

The genus *Allochrista* included earlier to the *Asura* / *Miltochrista* generic complex deserves a special mention. It has been described for the new species *Allochrista toxopei* Roepke, 1946 from Sulawesi Island. Unfortunately, we did not examine specimens of this species, but its original description (Roepke 1946) contains a good drawing of its male genitalia, which structure clearly corresponds to that of the subgenus *Thyrgorina* Walker, [1865] of the genus *Lemyra* Walker, 1856 (tribe Arctiini). Thus, here we establish a new synonymy and a new combination: *Lemyra* Walker, 1856 = *Allochrista* Roepke, 1946, **syn. nov.**, *Lemyra* (*Thyrgorina*) *toxopei* (Roepke, 1946), **comb. nov.**

Material and methods

Abbreviations used are as follows: CAV = private collection of Anton Volynkin (Barnaul, Russia); CKC = private collection of Karel Černý (Innsbruck, Austria); HT = holotype; MWM/ZSM = Museum Witt / Zoologische Staatssammlung, München (Munich, Germany); NHMUK = Natural History Museum, London (United Kingdom, formerly British Museum of Natural History, BMNH); PT = paratype; ST = syntype; ZFMK = Zoological Research Museum Alexander Koenig (Zoologisches Forschungsmuseum Alexander Koenig, Bonn, Germany); ZMA = Naturalis Biodiversity Center (former Zoölogisch Museum van Amsterdam, Leiden, The Netherlands); ZMB = Museum of Natural History, Berlin (Museum für Naturkunde, Berlin, Germany).

The genitalia were dissected and mounted in euparal on glass slides. Photos of imagoes were taken using the camera Nikon D3100/AF-S Nikkor, 18–55 mm. Photos of the genitalia were taken by the same camera attached to a microscope with an LM-scope adapter. All pictures were processed using the Adobe Photoshop CC 2018® software.

An overview of *Barsine* Walker, 1854 *sensu lato*, *Asura* Walker, 1854 and their relatives

Genus ***Barsine*** Walker, 1854
(Figs 1, 2, 104, 160)

Barsine Walker, 1854, *List of the specimens of lepidopterous insects in the collection of the British Museum* **2**: 546 (Type species: *Barsine defecta* Walker, 1854, by subsequent designation by Kirby (1892)).

= *Castabala* Walker, 1864, *List of the specimens of lepidopterous insects in the collection of the British Museum* **31**: 270 (Type species: *Castabala roseata* Walker, 1864, by monotypy).

= *Mahavira* Moore, 1878, *Proceedings of the general meetings for scientific business of the Zoological Society of London* **1878**: 11 (Type species: *Mahavira flavicollis* Moore, 1878, by monotypy).

Diagnosis. Medium-sized moths with yellow, orange, red or white ground colour and pattern consisting of black elements on veins and reddish spots between veins. In some species yellow forms without red spots are known. In *B. defecta* all females have no red spots. In some populations of some other taxa (*B. gratissima* and *B. orientalis bigamica*) such yellow coloured females are also common. In some species (*B. perpallida*, *B. andromeda* and members of the *B. flavicollis* species-group) the wing pattern is completely reduced. The main autapomorphic feature of the male genitalia is the presence of a long and robust basal saccular process. In addition, the male genitalia are characterised by the combination of the following features: (1) the weakly sclerotised, more or less X-shaped juxta consisting of two lateral parts connected by a weak membrane; (2) the presence of a robust medial costal process, which is usually broadened basally, apically curved and weakly setose; (3) the narrow distal section of costa protruding distally beyond the medial process and sometimes having a small distal process; (4) the presence of a distal membranous lobe of valva having a broad and elongated base; (5) the sacculus being distally narrow and having a robust distal process being well-separated from the valva surface; in many species this process has an additional dorsal lobe subbasally; (6) the aedeagus vesica having several well-separated diverticula bearing fields of small stout cornuti or robust granulation.

Distribution. The genus is widespread from India and Nepal through Mainland China and Indochina to Sundaland including the Philippine island of Palawan. Some species are known from Russian Far East, Korean Peninsula, Japan and the islands of Taiwan and Hainan.

Number of species. The genus comprises 63 valid species and 7 subspecies.

Genus ***Cyme*** Felder, 1861
(Figs 3, 4, 105, 161)

Cyme Felder, 1861, *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe* **43** (1): 36. (Type species: *Cyme reticulata* Felder, 1861, by subsequent designation by Kirby (1892)).

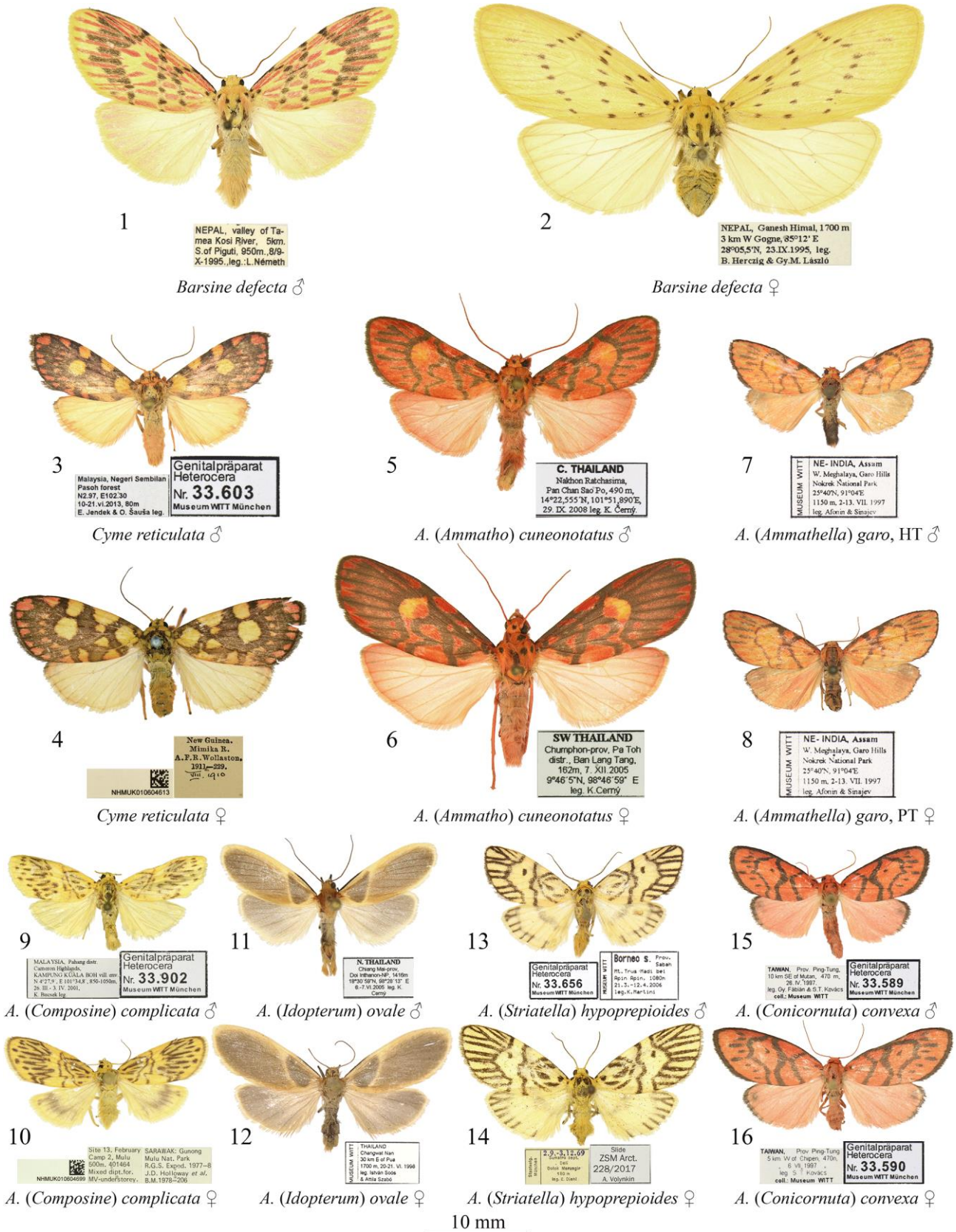
= *Pallene* Walker, 1854, *List of the specimens of lepidopterous insects in the collection of the British Museum* **2**: 542 (Type species: *Pallene structa* Walker, 1854, by subsequent designation by Kirby (1892)), nec. *Pallene* Dejean, 1821 (Coleoptera), **syn. rev.**

= *Prinasura* Hampson, 1903, *The Annals and magazine of natural history* (7) **11** (64): 349 (Type species: *Prinasura pyrrhopsamma* Hampson, 1903 (by monotypy), a junior synonym of *Pallene structa* Walker, 1854), **syn. rev.**

= *Miltasura* Roepke, 1946, *Tijdschrift voor entomologie* **87**: 85 (Type species: *Miltasura celebensis* Roepke, 1946, by monotypy), **syn. nov.**

Remarks. 1. The genus *Miltasura* was described for a new species *Miltasura celebensis* Roepke, 1946 (the male genitalia are illustrated by Roepke (1946)) being closely related to "*Barsine*" *euprepioides* widespread in mainland Asia and Sunda Shelf. The Javan "*Asura*" *septemmaculata* (Heylaerts, 1891) belongs to this species-group also. Holloway (2001) considered *euprepioides* as a member of *Barsine*, but its male (short costa without distal process) and female genitalia structure are clearly matching to those of *Cyme*. Therefore here we synonymise *Miltasura* with *Cyme* and transfer "*Barsine*" *euprepioides* and "*Asura*" *septemmaculata* to the genus *Cyme*. **2.** Rothschild (1913) described *Asura quadrifasciata* Rothschild, 1913 (page 210) from Sulawesi and *Miltochrista quadrifasciata* Rothschild, 1913 (page 214) from New Guinea. The second one was synonymised with *Asura cancellata* Pagenstecher, 1900 by Hampson (1900) but restored to the species status by Strand (1922). Later, de Vos (2019) transferred both taxa to the genus *Cyme* and considered to be junior synonyms of *Cyme sexualis*. The study of the male genital structures of the second, Sulawesian taxon proved its belonging to the genus *Cyme* also. Here we consider the New Guinean taxon to be a new junior secondary homonym of the Sulawesian one.

Diagnosis. Medium-sized or small moths with robust body and reticulate forewing pattern. The male genitalia of the genus are similar to those of the nominate subgenus of *Ammatho*, but differ by the absence of a distal costal process and the short costa not extending the medial costal process, a feature considering here to be autapomorphic. The female genitalia are surprisingly very similar to those of *Barsine* due to the evenly sclerotised ductus bursae with a strongly wrinkled posterior section, the sclerotised appendix bursae and the corpus bursae without spinules, but differ by the presence of a medial concavity of ostium bursae, which is absent in *Barsine*.



Figures 1–16. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: adults. Depositories of specimens: 1–3, 7–9, 12–16 – MWM/ZSM; 4, 10 – NHMUK; 5, 6, 11– CKC.

Distribution. The genus is distributed from mainland Asia through the Sunda Shelf, the Philippines and the Maluku Islands to New Guinea and Australia. The genus is most diverse in New Guinea with surrounding islands and Australia and poorly represent in mainland Asia, Greater Sunda Islands and the Philippines.

Number of species. The genus comprises 37 valid species and 2 subspecies.

Genus ***Ammatho*** Walker, 1855, **stat. nov.**
(Figs 5–20, 106–114, 162–169)

Ammatho Walker, 1855, *List of the specimens of lepidopterous insects in the collection of the British Museum* **3**: 759 (Type species: *Ammatho cuneonotatus* Walker, 1855, by subsequent designation by Kirby (1892).

Diagnosis. Members of the genus are significantly various in size, wing colouration and pattern, and in genitalia structures as well. However, the genus is defined here by the combination of the following genital features. Male genitalia: (1) medial costal process is present and well-developed; (2) costa is protruding beyond the medial costal process and forms a distal process (absent in the subgenus *Composine*); (3) distal membranous lobe of valva is present, well-developed (only in the subgenus *Delineatia* it is relatively small); (4) aedeagus vesica with a number of short diverticula bearing fields of granulation, numerous short but robust cornuti or, very rarely, short spinules (some species of *Striatella*). Female genitalia: (5) ductus bursae consists of a large and heavily sclerotised antrum and a short membranous anterior section (the latter is absent in *Rugosine*); (6) corpus bursae has dense spinulose scobination (that is very weak in *Composine*) and lacks spines, robust denticles or sclerotised plates; (7) appendix bursae is well-developed, situated postero-laterally, directed laterally or anteriorly, membranous, sometimes with spinulose scobination basally (fully scobinated in *Conicornuta*).

Distribution. The genus is widely distributed in the Oriental tropics, reaching the Maluku Islands in the east. Some species are known from China also.

Subgenus *Ammatho* Walker, 1855, **stat. nov.**
(Figs 5, 6, 106, 162)

Diagnosis. Species of the nominate subgenus are the largest within the genus and have forewing pattern similar to that of some species of *Cyme* (especially, the *C. euprepioides* species-group). Male genitalia are characterised by the combination of the following important characters: (1) medial costal process is large, elongated; (2) distal costal process is relatively small, thorn-like; (3) distal membranous lobe of valva is large, elongated; (4) juxta is trapezoidal, consisting of two crest-like sclerotised plates connected by a membrane (similar to that of *Cyme*); (5) aedeagus vesica has a large ventral diverticulum bearing several small subdiverticula. In some species aedeagus bears a cluster of dentation distally. Female genitalia are characterised by the large and weakly scobinated basally appendix bursae directed anteriorly or latero-anteriorly.

Distribution. *Ammatho* is the most widespread subgenus known from Sri Lanka and India through Indochina and Sundaland to the Maluku Islands; also present in the Philippines, but absent in the Palaearctic Region.

Number of species. The nominate subgenus comprises 14 valid species.

Subgenus *Ammathella* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:162E865C-26AA-48D3-9056-4205B4D2B6D5
(Figs 7, 8, 107, 163)

Type species: *Barsine garo* Volynkin, 2018.

Etymology. The diminutive suffix ‘ella’ refers to the relatively small size of subgenus members in comparison with the nominate sudgenus. Gender feminine.

Diagnosis. Members of the new subgenus are most similar externally to members of the genus *Moorasura* and not *Ammatho s. str*. Their male genitalia clearly show all key features of the genus, especially *Ammatho s. str*., but are characterised by the following features unique within the genus: (1)

medial costal process is weakly sclerotised, short, strongly broadened basally and slightly swollen (whereas in *Ammatho s. str*. that process is elongated, flattened and heavily sclerotised); (2) distal costal process small and apically rounded (that is thorn-like in *Ammatho s. str*.). In addition, juxta of *Ammathella* bears a well sclerotised medial process (whereas in *Ammatho s. str*. the juxta consists of two crest-like sclerotised plates connected by a membrane. Among other members of the genus *Ammatho* the similar process is present in some species of the subgenus *Striatella* (the *zebrina* species-group) only. Female genitalia are similar to those of *Ammatho s. str*. but have lateral ostial ligula, the character unique within the generic complex. Appendix bursae is smaller than in *Ammatho s. str*. and directed latero-inwards, whereas in *Ammatho s. str*. that is large and directed anteriorly.

Distribution. The subgenus is known from northeastern India and North Myanmar.

Number of species. Up to date, two valid species of *Ammathella* are known (Volynkin 2018a).

Subgenus *Composine* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:56FA9BFA-67EE-4DB2-88BC-4F531E6B5BAB
(Figs 9, 10, 108, 164)

Type species: *Sesapa complicata* Butler, 1877.

Etymology. The subgenus name is a combination of the species name *complicata* and the generic name *Barsine*. Gender feminine.

Diagnosis. The only known species of the subgenus is most similar to members of the subgenus *Barsipennis* of the genus *Ovipennis* and some *Miltochrista* species as well. However, according to the male and female genitalia structures, it definitely belongs to the genus *Ammatho*. The male genitalia of *Composine* are most similar to those of the subgenus *Striatella*, but differ by the absence of a distal costal process, the feature unique within the genus *Ammatho*, but found in some other genera. The female genitalia of *Composine* differ from those of other subgenera by the absence of spinulose scobination of corpus and appendix bursae and the presence of a band-like segmented sclerotised band covered with small denticles (similar structures are found in the genus *Ovipennis*).

Distribution. Sundaland.

Number of species. The subgenus is monobasic.

Subgenus *Idopterum* Hampson, 1894, **stat. nov.**
(Figs 11, 12, 109, 165)

Idopterum Hampson, 1894, *The Fauna of British India, including Ceylon and Burma (Moths)* **2**: 103 (Type species: *Idopterum ovale* Hampson, 1894, by original designation).

Diagnosis. The type species of the subgenus has an unusual monotonous brown wing colouration with yellow suffusion, while pattern elements common within the generic complex are not developed. The ground plan of its male genital capsule and aedeagus vesica are similar to those of the subgenus *Striatella*, but in *Idopterum* the distal costal process is extremely long, and the juxta has swollen and shortly dentate lateral lobes. The latter feature is unique within the generic complex, but female genitalia of *Idopterum* have no significant differences from those of the subgenera *Striatella* and *Delineatia* therefore here we consider *Idopterum* to be a subgenus of *Ammatho* and not a separated genus.

Distribution. Indochina, Java and the Bismarck Archipelago.

Number of species. Besides the type species, three other taxa were originally described as members of *Idopterum* and had an unclear generic placement. *Idopterum milani* Černý, 2009 is transferred herein to the genus *Barsilene*. Two other species, *A.* (*Idopterum) admirabilis* (Schaus, 1922), **comb. nov.** and *A.* (*Idopterum) novaepommeraniae* (Strand, 1922), **comb. nov.** remain unstudied by us and their subgeneric and generic placement is provisional and needs further clarification.

Subgenus *Striatella* Volynkin & Huang, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:4C0B635A-B6DF-4516-828F-E0C5A8CECD3A
(Figs 13, 14, 110, 166)

Type species: *Lithosia hypoprepioides* Walker, 1862.

Etymology. 'Striatus' means 'striped' in Latin; the diminutive suffix 'ella' refers to the small size of subgenus members. Gender feminine.

Diagnosis. Small or, rarely, medium-sized moths. Most members of the subgenus have whitish or yellowish colouration with blackish 'miltochristoid' pattern. The male and female genitalia of the new subgenus are very similar to those of *Ammatho s. str.* However, in the male genitalia of *Striatella*, the juxta is entire, shield- or plate-like, whereas that of *Ammatho s. str.* is trapezoidal, consisting of two crest-like sclerotised plates connected by a membrane. In addition, in *Striatella* the distal membranous lobe of valva is shorter and not so well separated than that of *Ammatho s. str.*, and the aedeagus vesica has no large ventral diverticulum bearing several small subdiverticula, which is characteristic for *Ammatho s. str.* The female genitalia of *Striatella* differ from those of *Ammatho s. str.* by the more robust spinulose scobination of corpus and appendix bursae. In addition, in most species of *Striatella* a signum bursae is present, while in *Ammatho s. str.* it is absent.

Distribution. The subgenus *Striatella* is widely distributed from Nepal and northeastern India through southern Mainland China, Hainan and Taiwan Islands, Indochina and Greater Sundas to the Philippines and the island of Sulawesi. It is absent from South India, Sri Lanka, and the Lesser Sundas east of Bali.

Number of species. Currently the subgenus comprises 55 species and one subspecies. Most species of the subgenus have been reviewed by Volynkin *et al.* (2019a) as members of 'the *Barsine hypoprepioides* species-group' and its allies. Here we additionally include several other species to the subgenus and exclude *'Barsine' salakia*, which is placed into the separated subgenus due to its unique male and female genitalia structures.

Subgenus *Conicornuta* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:ACC697A5-1369-42D6-9428-104D4F89079B
(Figs 15, 16, 111, 167)

Type species: *Miltochrista convexa* Wileman, 1910.

Etymology. The subgenus name refers to its large conical cornuti in aedeagus vesica. Gender feminine.

Diagnosis. Small moths with red colouration and black pattern. Externally, the only known species of the subgenus is similar to species of the subgenus *Delineatia* and the genera *Integrivalvia* and *Albarrania*, but, according to its male and female genitalia structures, clearly belongs to the genus *Ammatho*. Its male genital capsule is very similar to that of *Striatella*, but, however, is characterised by the presence of a short medial saccular process, a feature making *Conicornuta* similar to the genus *Melanaema* and the nominate subgenus of *Sarbine*. Nevertheless, valva of *Conicornuta* has a large medial costal process directed ventrally, whereas in *Melanaema* and *Sarbine* (*Sarbine*) that is absent. In addition, the vesica of *Conicornuta* bears 2–3 large conical cornuti, whereas in *Melanaema* and *Sarbine* (*Sarbine*) vesici bear clusters of numerous short but robust cornuti, and/or fields of granulation and patches of small spinules. This feature also separates *Conicornuta* from all other subgenera of *Ammatho*. The female genitalia of *Conicornuta* are dissimilar to those of *Melanaema* and are clearly similar to other subgenera of *Ammatho*, but differ from them by the presence of a lateral sclerotised protrusion of the posterior section of corpus bursae being densely covered with robust spinules, a feature considering here to be autapomorphic.

Distribution. The only known species of the subgenus is an endemic of Taiwan Island.

Number of species. The subgenus is monobasic.

Subgenus *Delineatia* Volynkin & Huang, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:3431D68D-C1EA-4F21-8C3C-52CB9A2B0462
(Figs 17, 18, 112, 113, 168)

Type species: *Hypoprepia delineata* Walker, 1854.

Etymology. The subgenus name is derived from its type species name. Gender feminine.

Diagnosis. Medium-sized moths with red colouration and black pattern. Externally, members of the subgenus are similar to species of the subgenus *Conicornuta* and the genera *Integrivalvia* and *Albarrania*. Male genitalia are characterised by (1) the densely serrulate anellus; (2) the small medial costal process at very long and narrow, lobe-like base (an autapomorphic feature); (3) very large and heavily sclerotised distal

costal process bearing several denticles or short prominences; (4) very small distal membranous lobe of valva; (5) strongly setose sacculus; (6) narrow vesica with short diverticula bearing clusters of very small spine-like cornuti or granulation. Female genitalia structure is typical for the genus *Ammatho* and is characterised by (1) very broad but narrow postvaginal plate; (2) cup-like antrum and very short anterior membranous section of corpus bursae; (3) weakly scobinated posterior section of corpus bursae; (4) membranous anterior section of corpus bursae without signa.

Distribution. Northern Indochina, Mainland China and Taiwan Island.

Number of species. The genus comprises two valid species reviewed by Volynkin & Černý (2017a).

Subgenus *Rugosine* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:8832D267-4272-435F-8DCB-72F1248D877B
(Figs 19, 20, 114, 169)

Type species: *Miltochrista salakia* Schaus, 1922.

Etymology. The subgenus name is a combination of the word 'rugose' (a reference to the strongly rugose ductus bursae of the type species) and the generic name *Barsine*. Gender feminine.

Diagnosis. Small moths with white colouration and black pattern. Externally the only known species of the subgenus is similar to numerous whitish specimens of the subgenus *Striatella*, but clearly differs from them by its male and female genitalia structure. The male genital capsule is similar to that of *Striatella*, but the two following unique features are characteristic for the subgenus: (1) the manica bears two very large trigonal lobes densely covered with numerous spinules; and (2) the medial costal process is heavily sclerotised, plate-like, rectangular. The female genitalia structure is unique for the generic complex and is characterised by the very broad, heavily sclerotised and rugose ductus bursae, whose rugose sclerotisation asymmetrically protrudes to the right side of the posterior section of corpus bursae. Signum bursae is present. Appendix bursae is small, membranous.

Distribution. Sumatra, Java and Bali Islands.

Number of species. The subgenus is monobasic.

Genus ***Ovipennis*** Hampson, 1900
(Figs 21–26, 115–117, 170–173)

Ovipennis Hampson, 1900, *Catalogue of Lepidoptera Phalaenae in the British Museum* **2**: 410 (Type species: *Nudaria dudgeoni* Elwes, 1890, by original designation).

Diagnosis. Small moths with diverse colouration and pattern. The male genitalia are similar to those of *Ammatho* (especially, the subgenus *Striatella*), but in *Ovipennis* aedeagus vesica has elongated diverticula with bunches of spinules, whereas in *Ammatho* vesici bear clusters of numerous small cornuti or separated large conical cornuti. Medial costal process is usually well developed, but reduced (up to absence) in the subgenus *Coccinigripennis*. Female genitalia are characterised by (1) the evenly sclerotised ductus bursae (in *Ammatho* that is separated into antrum and membranous anterior section) and (2) the presence of rows of robust spinules or heavily sclerotised plates which may be dentate (absent in *Ammatho*).

Distribution. The genus is widespread from South India through Himalaya to China and northern Indochina.

Subgenus *Ovipennis* Hampson, 1900
(Figs 21, 22, 115, 170)

Diagnosis. Small moths with pattern expressed as large spots and longitudinal stripes. The male genital capsule is very similar to that of *Ammatho* (*Striatella*), but distal costal process is lobe-like broadened, whereas in *Ammatho* (*Striatella*) that is thorn-like. In some species aedeagus carina bears a small cluster of small but robust spinules. Aedeagus vesica with elongated diverticula bearing bunches of large spine-like cornuti. Female genitalia are similar to those of *Miltochrista*, with the posterior section of corpus bursae bearing clusters of numerous robust spine-like cornuti posteriorly.

Distribution. The subgenus is distributed from Himalaya through northern Myanmar to southern China and northern Indochina.

Number of species. The nominate subgenus comprises seven valid species.

Subgenus *Barsipennis* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:2EC2C0E7-8CD0-4DC7-91C0-25BDE20C701B
(Figs 23, 24, 116, 171, 172)

Type species: *Barsine joshii* Volynkin & Černý, 2019.

Etymology. The subgenus name is a combination of the generic names *Barsine* and *Ovipennis*. Gender feminine.

Diagnosis. Small moths with yellow colouration and black 'miltochristoid' pattern. The male genital capsule is similar to that of the nominate subspecies, but distal costal process is developed as a very short rounded protrusion (*O. phaeodonta*, *O. joshii* and *O. meyi*) or absent (*O. mophi* and *O. arrigera*). The aedeagus vesica structure is similar to that of the nominate subspecies, but diverticula bear bunches of spinules and not spine-like cornuti. Female genitalia are characterised by the presence of sclerotised plates in corpus bursae which may be dentate.

Distribution. The subgenus is distributed from Himalaya through northern Myanmar to northern Indochina.

Number of species. The subgenus comprises five valid species reviewed by Volynkin & Černý (2019a) as 'the *Barsine phaeodonta* species-group'.

Subgenus *Nebulene* Volynkin & Černý, 2018, **stat. nov.**
(Figs 25, 26, 117, 174)

Nebulene Volynkin & Černý, 2018, *Zootaxa* **4394** (2): 220 (Type species: *Setina nebulosa* Moore, 1878, by original designation).

Diagnosis. Small moths, the male antennae are bipectinate, the female antennae are finely ciliate. Forewing pattern consists of a trigonal antemedial and broad postmedial shades connected at dorsum, a wavy and thin medial line, and a discal spot. The male genital capsule is very similar to that of *Barsipennis*, but the medial costal process is more robust. In aedeagus vesica bears one–two large, well separated blade-like cornuti, whereas in *Barsipennis* patches of spinules are present only. Such a robust medial costal process and large blade-like cornuti are present in male genitalia of several other genera of the *Asura* / *Miltochrista* generic complex, but never in combination as in *Nebulene*. The male genitalia of *Nebulene* also resemble those of *Barsura*, but differ by the absence of a distal costal process, absence of a carinal plate, and presence of well separated, long, narrow, but robust cornuti. In the female genitalia of *Nebulene* the ductus bursae is weakly membranous, a feature also found in the *Asura* / *Miltochrista* generic complex in the genus *Barsochrista* only. Nevertheless, in *Nebulene* the appendix bursae is situated postero-laterally and the posterior section of corpus bursae bears heavily sclerotised and dentate plates. The latter feature is also occurring in the subgenus *Barsipennis* only.

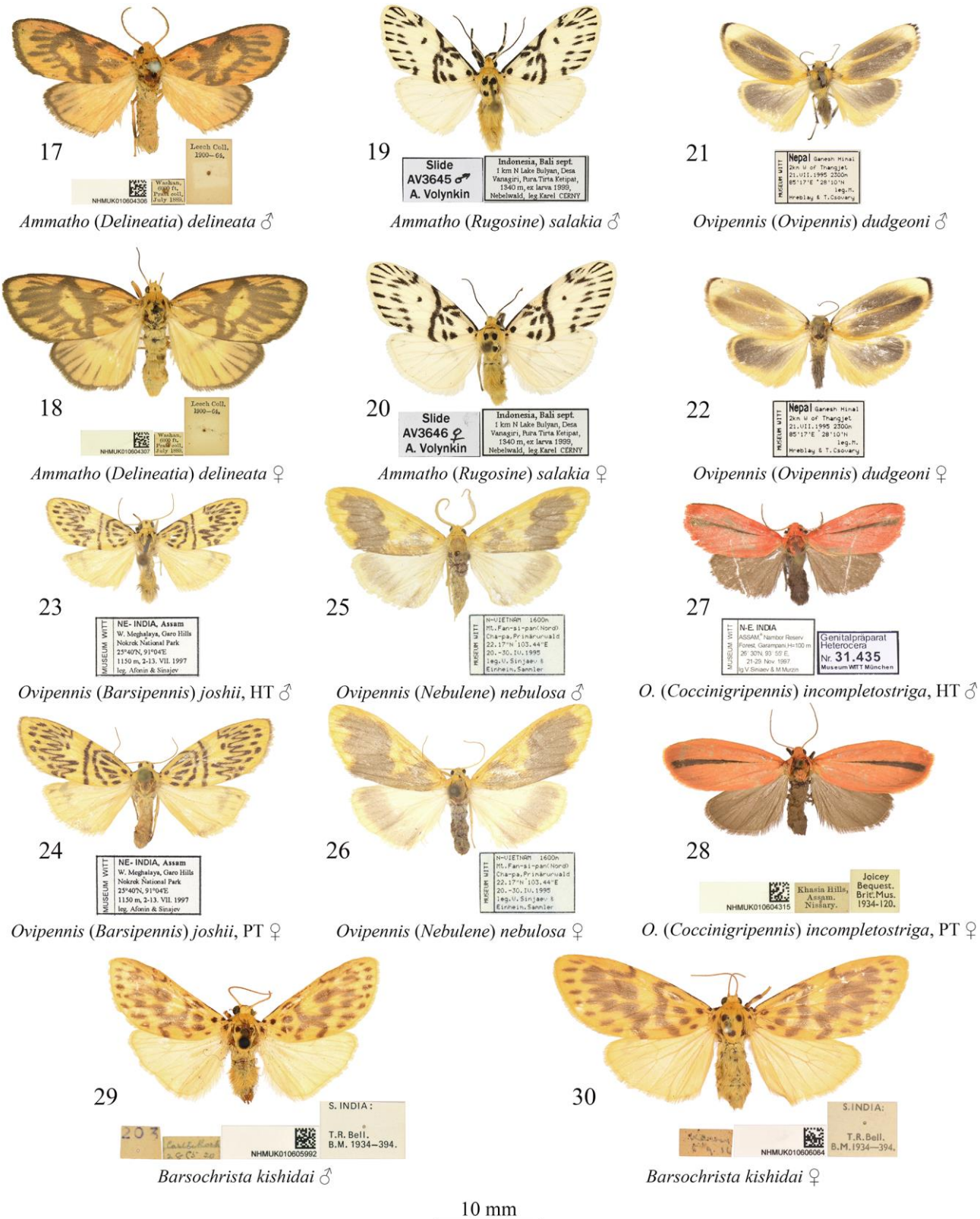
Distribution. The genus is distributed in Himalaya, southwestern China and Indochina.

Number of species. The genus comprises two valid species reviewed by Volynkin & Černý (2018c).

Subgenus *Coccinigripennis* Volynkin & Huang, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:6D3F1640-01E9-40D3-B0C1-B1B3C3C8CE85
(Figs 27, 28, 118, 173)

Type species: *Barsine incompletostriga* Volynkin & Cerny, 2017.

Etymology. The subgenus name is a combination of the Latin word 'coccinigrum' meaning 'crimson and black' and the generic name *Ovipennis*. Gender feminine.



Figures 17–30. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: adults. Depositories of specimens: 17, 18, 28–30 – NHMUK; 19, 20 – CKC; 21–27 – MWM/ZSM.

Diagnosis. Small moths with outwardly rounded forewing having crimson ground colour and black pattern represented as a longitudinal strip or a broad shade. In male genital capsule, valva is elongated, with setose sacculus and small distal costal process. The subgenus differs from other subgenera by its medial

costal process reduced to a small tubercle or denticle (the *O. anomala*, *O. fansipana* and a part of *O. miloslavae* species-groups) or absent (most species of the *miloslavae* species-group). Female genitalia of the subgenus are characterised by the presence of band-like clusters of numerous short but robust spinules (except for *O. anomala* and *O. fansipana* having corpus bursae evenly covered with numerous spinules). Signum bursae is present in all species.

Distribution. The subgenus is distributed from Himalaya through northern Myanmar to southern China and northern Indochina.

Number of species. The subgenus comprises six valid species reviewed by Volynkin & Černý (2017b) as the '*Barsine anomala*', '*Barsine fansipana*' and '*Barsine miloslavae*' species-groups.

Genus ***Barsochrista*** N. Singh & Kirti, 2016
(Figs 29, 30, 119, 175)

Barsochrista N. Singh & Kirti, *in* Kirti & N. Singh, 2016, *Arctiid Moths of India* **2**: 2017 (Type species: *Lyclene kishidai* Kirti & Gill, 2009, by original designation).

Diagnosis. Medium-sized moths with dark yellow forewing ground colour and pattern consisting of brown spots and wavy and irregularly dentate transverse lines and elliptical discal spot. Hindwings yellow, without pattern. The male genitalia of *Barsochrista* are similar to those of *Ammatho* and *Ovipennis*, but are characterised by such an autapomorphic feature as the voluminous, tubercle-like and heavily sclerotised medial costal process (whereas in *Ammatho* and *Ovipennis* that is flattened, lobe-, stick- or thorn-like. The voluminous tubercle-like medial costal process is also present in *Esmasura*, but it is weakly sclerotised and densely setose. In addition, the aedeagus vesica of *Barsochrista* is more or less round and bears two bunches of long and robust cornuti, whereas in *Ammatho* vesica bears broad clusters of small cornuti or granulation; in *Ovipennis*, bunches of spines are also present, but they are positioned on elongated diverticula. Female genitalia of *Barsochrista* are most similar to those of *Nebulene* due to the very short and membranous ductus bursae, but differ by the presence of clusters of numerous robust spinules in the posterior section of corpus bursae (that bears sclerotised dentate plates in *Nebulene*) and the large membranous appendix bursae originating from the anterior section of corpus bursae. The latter feature is also known in the genus *Asuridia* only.

Distribution. Sri Lanka and southern India.

Number of species. The genus comprises two valid species reviewed by Volynkin & Ivanova (2017).

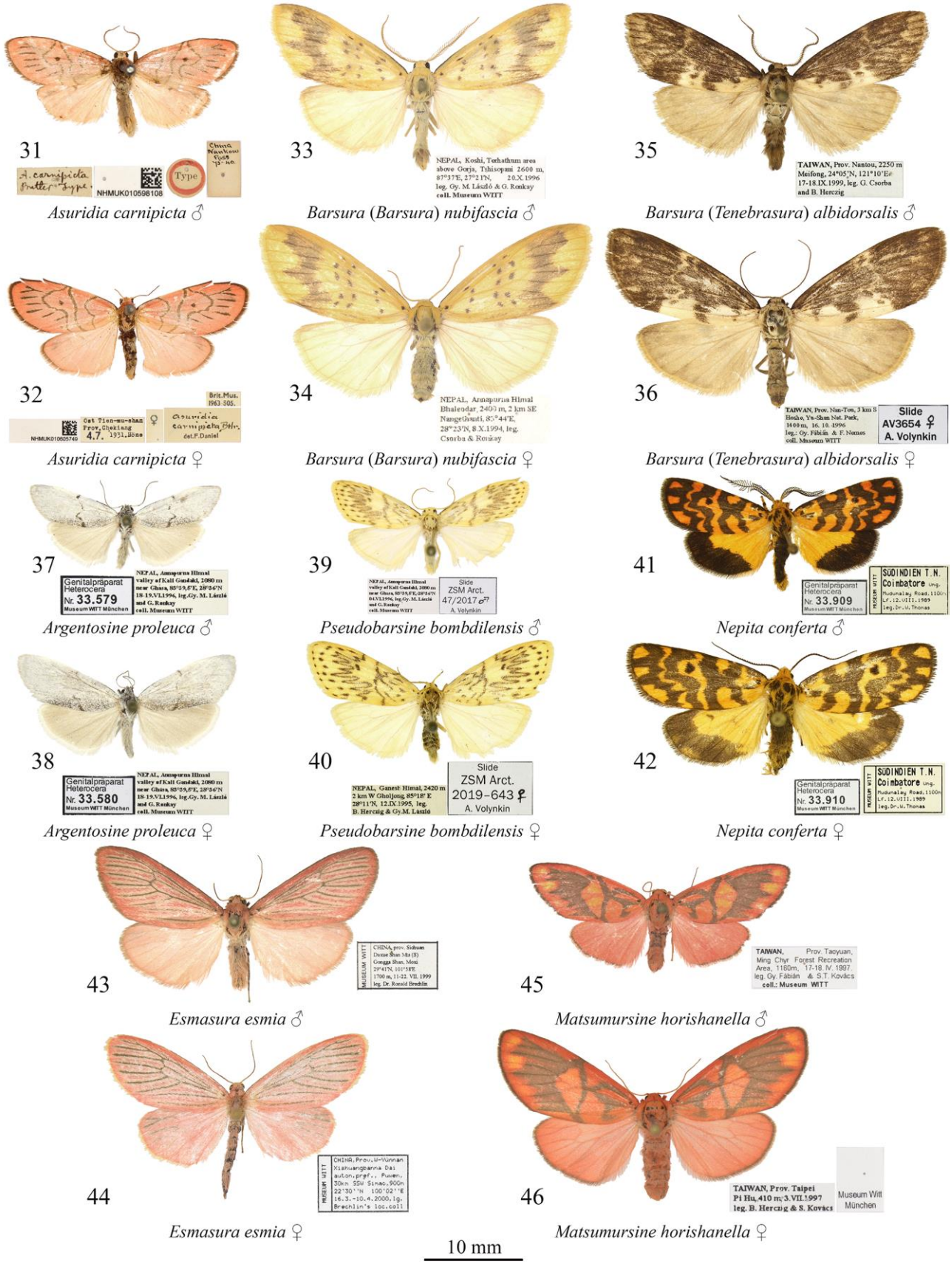
Genus ***Asuridia*** Hampson, 1900
(Figs 31, 32, 120, 176)

Asuridia Hampson, 1900, *Catalogue of Lepidoptera Phalaenae in the British Museum* **2**: 412 (Type species: *Ammatho carnipicta* Butler, 1877, by original designation).

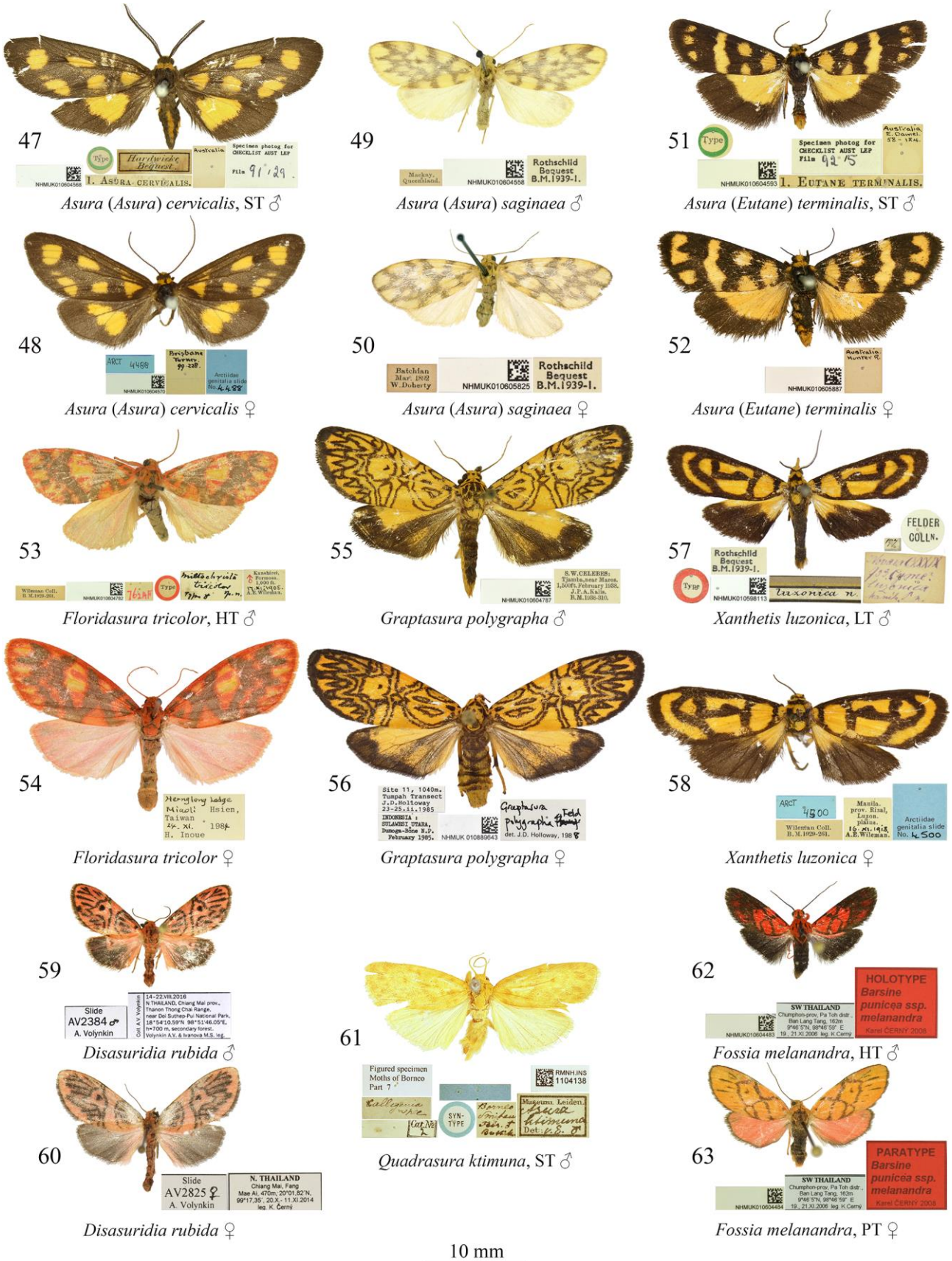
Diagnosis. Small moths with reddish colouration and black pattern. The male genital capsule of *Asuridia* is most similar to that of *Ammatho*, but is characterised by the densely setose medial costal process and sacculus. In aedeagus vesica, the large hook-like curved membranous diverticulum is characteristic. Female genitalia are dissimilar to those of *Ammatho* and resemble those of *Barsochrista* due to the appendix bursae originating from the anterior section of corpus bursae, but in *Asuridia* that is weakly scobinated and directed anteriorly. In addition, for female genitalia of *Asuridia* the following features are characteristic: (1) the antevaginal plate is large and strongly rugose (absent in *Barsochrista* and *Ammatho*); (2) the ductus bursae is asymmetrically sclerotised (dissimilar to *Barsochrista* and *Ammatho*, but this feature is represent in *Barsura* also).

Distribution. Himalaya, Mainland China, Taiwan and northern Indochina.

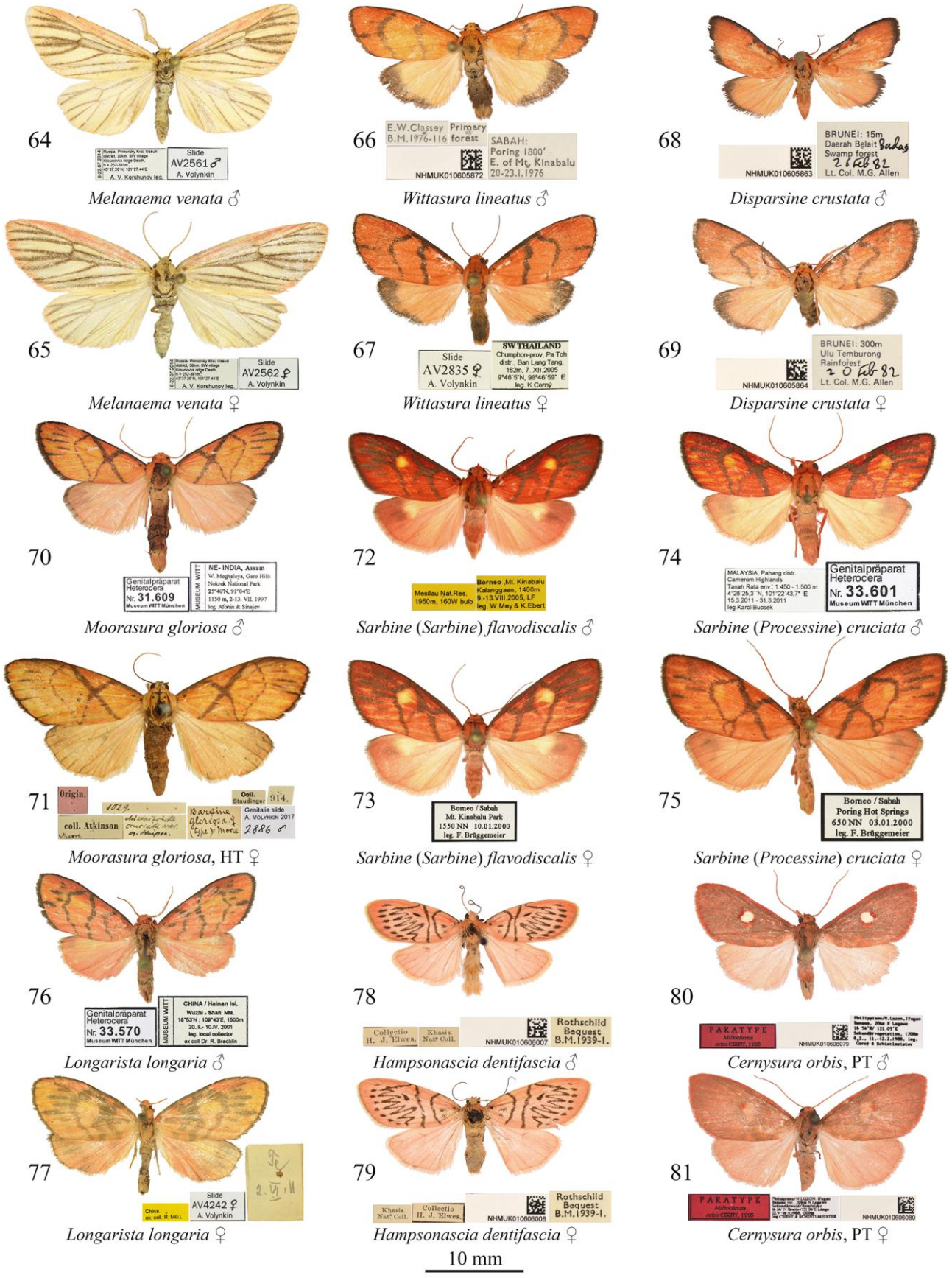
Number of species. The genus comprises 11 valid species. Species distributed in New Guinea and surrounding islands formerly treated as members of *Asuridia* by de Vos (2019) are obviously unrelated to *Asuridia* in fact and provisionally transferred here to the genus *Integrivalvia*.



Figures 31–46. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: adults. Depositories of specimens: 31, 32 – NHMUK; 33–46 – MWM/ZSM.



Figures 47–63. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: adults. Depositories of specimens: 47–58, 62, 63 – NHMUK; 59 – CAV; 60 – CKC; 61 – ZMA (photo by R. de Vos).



Figures 64–81. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: adults. Depositories of specimens: 64, 65 – CAV; 66, 68, 69, 78–81 – NHMUK; 67, 73, 75 – CKC; 70, 74, 76 – MWM/ZSM; 71, 72, 77 – ZMB.

Genus ***Barsura*** Volynkin, Dubatolov & Kishida, 2017
(Figs 33–36, 121, 122, 177, 178)

Barsura Volynkin, Dubatolov & Kishida, 2017, *Zootaxa* **4299** (1): 55 (Type species: *Barsine nubifascia* Walker, [1865], by original designation).

Diagnosis. Medium-sized moths, the male antennae are bipectinate, the female antennae are ciliate. The male genitalia of *Barsura* are similar to those of *Ammatho*, but aedeagus of *Barsura* bears a well separated, robust and strongly dentate carinal plate (absent in *Ammatho*). The female genitalia are characterised by the combination of the following features: (1) the antevaginal plate is present, rugose; (2) the ductus bursae is asymmetric, with more elongated left anterior part; (3) posterior section of corpus bursae with heavily scobinated lateral enlargements, while the anterior section is weakly scobinated and very small (nearly reduced in *Umbrasura*).

Distribution. The genus is distributed from Himalaya through southern Mainland China and northern Vietnam to Taiwan.

Subgenus *Barsura* Volynkin, Dubatolov & Kishida, 2017
(Figs 33, 34, 121, 177)

Diagnosis. The male genitalia of the subgenus are characterised by the large distal costal process, narrow aedeagus and narrow vesica with ventral subbasal diverticulum and clusters of small cornuti and bunches of spinules. In female genitalia lateral enlargements of the posterior section of corpus bursae are heavily sclerotised, pocket-like; anterior, weakly scobinated section of corpus bursae is well-developed.

Distribution. The subgenus is distributed from Himalaya through southern Mainland China to northern Vietnam, but absent in Taiwan.

Number of species. The nominate subgenus comprises 11 species arranged into two species-groups (Volynkin *et al.* 2017; Volynkin 2017a; 2018).

Subgenus *Tenebrasura* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:489E372E-2F88-44D3-8C7C-A9A23244F647
(Figs 35, 36, 122, 178)

Type species: *Asura albidorsalis* Wileman, 1914.

Etymology. The subgenus name is a combination of the word ‘tenebrosus’ meaning ‘dark’ in Latin and the generic name *Asura*. Gender feminine.

Diagnosis. Male genital capsule of the new subgenus differs from that of the nominate subgenus by the costa protruding beyond the distal process to the distal lobe of valva (in *Barsura* it ends with the process), and the distal costal process being much smaller, thorn-shaped. In female genitalia, the antevaginal plate is present and short, similar to that of the *B. simplifascia* species-group, and the ductus bursae is broader and more massive than that of *Barsura*, but asymmetrical also. However, unlike the nominate subgenus, in *Tenebrasura* the posterior section of corpus bursae is broadened and heavily scobinated, its left lateral enlargement is short but heavily sclerotised; a right enlargement is not developed; the anterior, weakly scobinated section of corpus bursae is broad, but very short, barely noticeable.

Distribution. Endemic of Taiwan Island.

Number of species. The subgenus is monobasic.

Genus ***Argentosine*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:0320A5A4-4BCE-4177-9F05-E7E3EF885286
(Figs 37, 38, 123, 179)

Type species: *Miltochrista proleuca* Hampson, 1900.

Etymology. The genus name is a combination of the word 'argenteus' meaning 'silver' in Latin and the generic name *Barsine*. The name refers to the silver grey ground colour of the genus' type species. Gender feminine.

Diagnosis. Externally, the type species of the genus more resembles some members of the subtribe Lithosiina than Nudariina due to its narrow silver grey forewings with reduced pattern, but its male and female genitalia are typical for the *Asura* / *Miltochrista* generic complex. The male aedeagus vesica is very similar to that of the genus *Ammatho* by a number of small but robust cornuti typical for the *Barsine* genus group, but the genital capsule is similar to that of some *Miltochrista* species due to the absence of a medial costal process. The combination of such features as the absence of a medial costal process and the presence of small but robust cornuti only is characteristic for the genus *Amphisine* also (especially, *A. asaphes* and its allies), but in *Argentosine* the distal saccular process is well-developed and elongated, the distal lobe of valva is present and large, and the costa is not protruding beyond them, whereas in *Amphisine* the distal saccular process is reduced to a small protrusion, the distal lobe of valva is absent and the costa forms a strongly elongated narrow distal process directed along the valva axis. The female genitalia of *Argentosine* are similar to those of *Ammatho*, but can be easily recognised by the broad area of rugose sclerotisation occupying the posterior end and left side of corpus bursae and the base of appendix bursae. The lateral position of appendix bursae, which is directed laterally, is also characteristic.

Distribution. Nepal and northeastern India.

Number of species. The genus is monobasic.

Genus ***Pseudobarsine*** N. Singh & Kirti, 2016
(Figs 39, 40, 124, 180)

Pseudobarsine N. Singh & Kirti, *in* Kirti & N. Singh 2016, *Arctiid Moths of India* **2**: 85 (Type species: *Pseudobarsine bombdilensis* N. Singh & Kirti, 2016, by original designation).

Diagnosis. Small moths with yellow colouration and blackish 'miltochristoid' pattern. Both species of the genus are very similar externally to some *Miltochrista* species. The male genital capsule is also similar to that of some *Miltochrista* groups due to the absence of both costal processes, but is characterised by the presence of such an autapomorphic feature as a swollen setose short process at base of a distal saccular process. The aedeagus vesica has several diverticula, some of which bear clusters of small spinules. The female genitalia can be recognised by the following two autapomorphic features: (1) the weakly sclerotised but densely spinulose antevaginal plate; and (2) the shortly dentate postvaginal plate. In addition, the corpus bursae densely spinulose, the appendix bursae short, membranous, situated postero-laterally.

Distribution. Nepal, northern and northeastern India, northwestern Myanmar.

Number of species. The genus comprises two valid species.

Genus ***Nepita*** Moore, [1860]
(Figs 41, 42, 125, 181)

Nepita Moore, *in* Horsfield & Moore, [1860], *A catalogue of the lepidopterous insects in the Museum of Natural History at the East-India House* **2**: 302, a replacement name for *Pitane* Walker, 1854.

= *Pitane* Walker, 1854, *List of the specimens of lepidopterous insects in the collection of the British Museum* **2**: 531 (Type species: *Pitane conferta* Walker, 1854, by subsequent designation by Hampson (1900)), nec. Walker, 1854 (Arctiini).

Diagnosis. Medium-sized moths; male antennae strongly bipectinate, female antennae serrate; wing pattern is similar to that of some *Cyme* and *Ammatho*. In the male genitalia, the tegumen is broad; the juxta with a deep medial basal concavity; the valva is short and medially broadened, nearly elliptical. The combination of such features as the presence of a robust, thorn-shaped medial costal process and the absence of distal costal and saccular processes and distal membranous lobe of valva is characteristic for the genus. Aedeagus is large, and the vesica structure is similar to that of *Barsine* and *Ammatho s. str*. In the female genitalia, the large and heavily sclerotised antrum and the short membranous anterior section of corpus

bursae are similar to those of *Ammatho s. str*., but the corpus and appendix bursae are completely membranous, only an elliptical signum bursae is present.

Distribution. Sri Lanka and India.

Number of species. The genus is monobasic.

Genus ***Esmasura*** Volynkin & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:89360C04-B569-4C9F-95E9-04AF1FC68B1E
(Figs 43, 44, 126, 182)

Type species: *Miltochrista esmia* Swinhoe, 1893.

Etymology. The genus name is a combination of its type species' name *esmia* and the generic name *Asura*. Gender feminine.

Diagnosis. Medium-sized moths with pale red body and wing colouration and pattern consisting of blackish suffusion along veins only. Antennae of both sexes ciliate. In the male genitalia, the valva lacks distal costal and saccular processes, like in *Nepita*, but the medial costal process is small, tubercle-like and setose as well as the area around it (an autapomorphic feature; similar tubercle-like process is known in *Barsochrista* only, but in the latter that is heavily sclerotised and not setose). In addition, in *Esmasura* the sacculus is broader and reaching the valva apex, where it connects with costa, whereas in *Nepita* the sacculus is narrower and shorter, not reaching the valva apex. Aedeagus is elongated and narrow, nearly straight; vesica is short, nearly elliptical, with two clusters of numerous various sized small and very small thorn-like cornuti. In the female genitalia, the ductus bursae is subdivided into the heavily sclerotised trapezoidal antrum and the moderately sclerotised anterior section. The anterior section of corpus bursae is elliptical, membranous, with a weakly dentate signum medially. The posterior section of corpus bursae is narrow, moderately sclerotised and weakly rugose. The appendix bursae originates from the posterior section of corpus bursae and is evenly moderately sclerotised and weakly rugose also. Such an appendix bursae structure is unique among the generic complex.

Distribution. Northern Myanmar, China and northern Vietnam.

Number of species. The genus is monobasic.

Genus ***Matsumursine*** Volynkin & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:F1E01D56-B96D-44B6-9CF1-803F6C9D6DFA
(Figs 45, 46, 127, 183)

Type species: *Asura horishanella* Matsumura, 1927.

Etymology. The genus name is a combination of the surname Matsumura, an author of the type species, and the generic name *Barsine*. Gender feminine.

Diagnosis. Small moths with red colouration and dark grey forewing pattern and diffuse yellow spots. Antennae of both sexes are ciliate. In the male genital capsule, the anellus is weakly sclerotised, valva is strongly narrowed apically, the medial costal process is robust, the costa is not reaching the valva apex, the distal lobe of valva small, the distal saccular process is robust. The main autapomorphic feature in male genitalia is the valva costa structure: in *Matsumursine*, the costa has two processes medially: the medial one directed ventrally (the feature common for the generic complex) and the dorsal one, which is broad, short, swollen and weakly sclerotised in the type species and robust and thorn-like but situated more basally in *M. speciosa*. The aedeagus is narrow, with an elongated coecum and a short thorn-like carinal process. Aedeagus vesica short, nearly globular, with cluster of very small spinules or spine-like cornuti. In the female genitalia, the combination of the following features is characteristic for the genus: (1) the postvaginal plate is present, strongly broadened (similar to that of *Ammatho* (*Delineatia*), but much longer and broader); (2) the ductus bursae is asymmetrical, curved laterally, its long and protruding right side is heavily sclerotised, while the left side is very short and weakly sclerotised (autapomorphic feature; similar asymmetrical ductus bursae is known in *Barsura* only, but in the latter genus it is longer and not curved, and is combined with broad sclerotisation and scobination of corpus bursae, whereas in *Matsumursine* the corpus

bursae is membranous with weak granulation only); (3) posterior section of corpus bursae narrow, twisted, membranous; (4) anterior section of corpus bursae is broadened, sack-like, membranous, with a field of very weak granulation and a small signum; (5) appendix bursae is narrow, finger-like, membranous, situated postero-laterally.

Distribution. Mainland China and Taiwan Island.

Number of species. The genus comprises two valid species.

Genus ***Asura*** Walker, 1854

Asura Walker, 1854, *List of the specimens of lepidopterous insects in the collection of the British Museum* **2**: 484 (Type species: *Asura cervicalis* Walker, 1854, by subsequent designation by Kirby (1892)).

Diagnosis. Medium-sized or small moths with various colouration and pattern. Male antennae bipectinate or ciliate, female antennae ciliate. The male genitalia of the genus are characterised by the combination of the following features: (1) the medial costal process is present; (2) distal membranous lobe of valva is present, well-developed; (3) distal saccular process is present, robust; (4) aedeagus robust, straight, relatively short; (5) aedeagus vesica with several short diverticula bearing clusters of numerous spinules. In the female genitalia, the ductus bursae is dorso-ventrally flattened, sclerotised and not narrowed anteriorly, and antrum is absent.

Distribution. The genus is widespread from mainland Asia to New Guinea and Australia.

Subgenus *Asura* Walker, 1854
(Figs 47–50, 128, 129, 184, 185)

Asura Walker, 1854, *List of the specimens of lepidopterous insects in the collection of the British Museum* **2**: 484 (Type species: *Asura cervicalis* Walker, 1854, by subsequent designation by Kirby (1892)).

= *Stonia* Walker, [1865], *List of the specimens of lepidopterous insects in the collection of the British Museum* **31**: 187 (Type species: *Stonia bipars* Walker, [1865], by monotypy).

= *Gymnasura* Hampson, 1900, *Catalogue of Lepidoptera Phalaenae in the British Museum* **2**: 425 (Type species: *Calligenia saginaea* Turner, 1899, by original designation), **syn. nov.**

Remark. Male and female genitalia of the type species of *Asura* and *Gymnasura* have no significant differences therefore here we synonymise *Gymnasura* with *Asura*. The bilobate distal saccular process of *Asura saginaea* is also characteristic for some other species (e.g., *A. toxodes*), but in *A. fulguritis*, which is definitely closely related to *A. saginaea* and *A. toxodes*, the distal saccular process is unilobate therefore this feature cannot be considered as generic (see that of *Barsine* also).

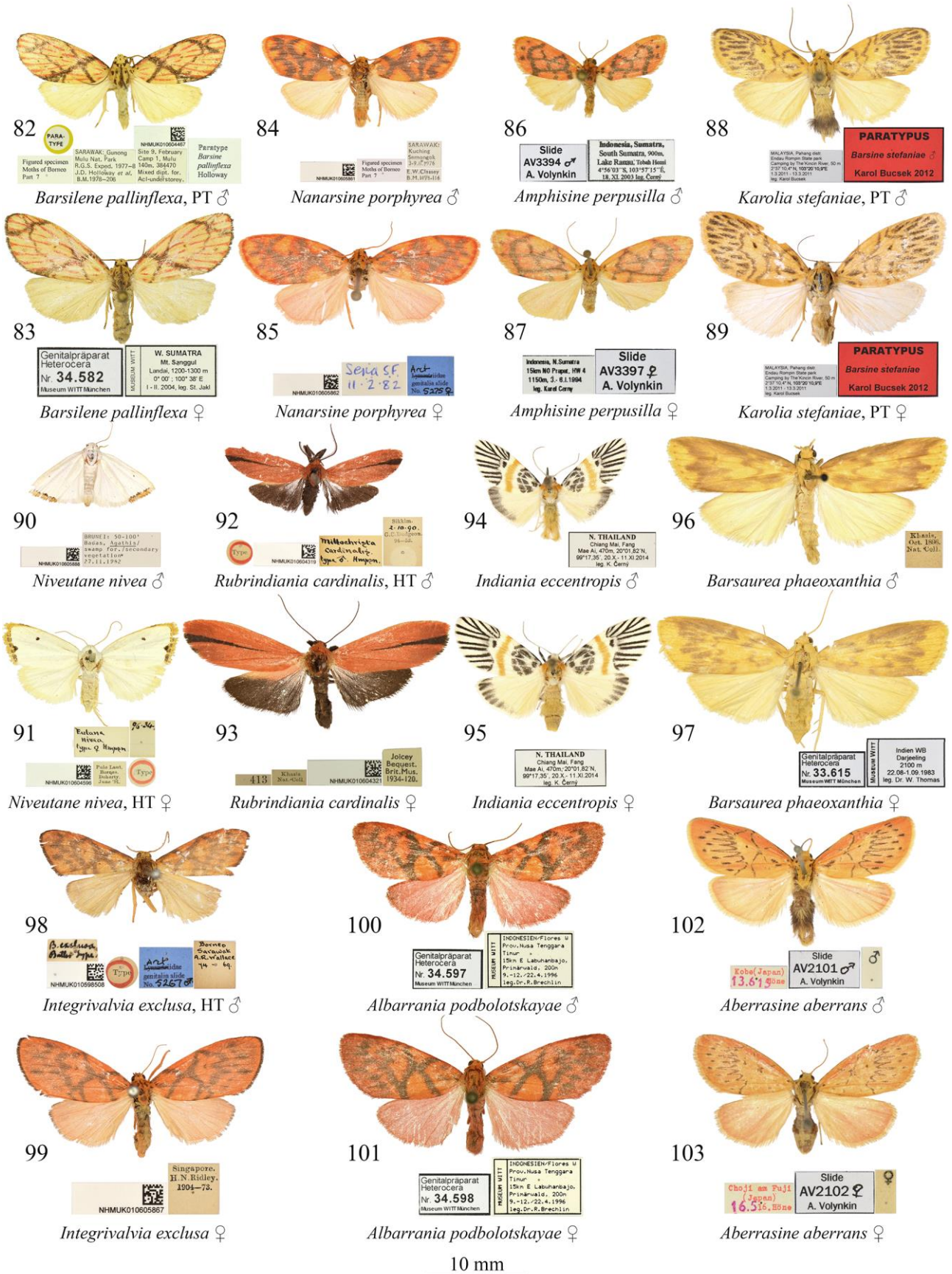
Diagnosis. The male genitalia of the nominate subgenus are characterised by the small, weakly setose medial costal process. Costa sometimes have a small round distal process. Anellus of some species bears bunches of spinules. Distal saccular process is bilobate in some species. The female genitalia are characterised by the moderately sclerotised ductus bursae (sometimes with a membranous anterior section) and weakly sclerotised appendix bursae.

Distribution. Most species are distributed in Australia, New Guinea and surrounding islands. Some species are known from northeastern India, Indochina, Sundaland and the Philippines.

Number of species. Here we associate with the subgenus 59 valid species and 2 subspecies, but generic placement of some species is still unclear and needs clarification. Some species formerly associated with *Gymnasura* are provisionally transferred here to the genus *Ovipennis s. str.*, but their placement needs further clarification also.

Subgenus *Eutane* Walker, 1854, **stat. nov.**
(Figs 51, 52, 130, 186)

Eutane Walker, 1854, *List of the specimens of lepidopterous insects in the collection of the British Museum* **2**: 531 (Type species: *Eutane terminalis* Walker, 1854, by monotypy).



Figures 82–103. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: adults. Depositories of specimens: 82, 84, 85, 90–93, 98, 99 – NHMUK; 83, 88, 89, 97, 100, 101 – MWM/ZSM; 86, 87, 94, 95 – CKC; 96, 102, 103 – ZFMK.

Diagnosis. The male genitalia of the two subgenera are very similar, but in *Eutane* the valva costa is shorter and the medial costal process is much larger than those of *Asura*. The main difference between *Eutane* and *Asura* is in the female genitalia structure: (1) in *Eutane* the ductus bursae is rugose anteriorly, whereas in *Asura* that is evenly sclerotised along its length and not rugose; (2) the corpus bursae of *Eutane* bears a heavily sclerotised and swollen area laterally, whereas in *Asura* such a sclerotisation is absent, but spinulose scobination or rows of spinules are usually present; (3) the appendix bursae of *Eutane* is membranous, whereas in *Asura* that is weakly sclerotised.

Distribution. Australia and New Guinea.

Number of species. The subgenus currently comprises four valid species. "*Eutane*" *nivea* and its relatives are clearly not congeneric to the type species of *Eutane* and erected below to the separated genus *Niveutane* related to *Nanarsine* and *Amphisine*.

Genus ***Floridasura*** Volynkin, **gen. nov.**

https://zoobank.org/urn:lsid:zoobank.org:act:269D7507-E9CA-4094-833B-1C55B2E1C204

(Figs 53, 54, 131, 187)

Type species: *Miltochrista tricolor* Wileman, 1910.

Etymology. The genus name is a combination of the Latin word 'floridus' meaning 'colourful' and the generic name *Asura*. The genus' name refers to the colourful forewing of its type species. Gender feminine.

Diagnosis. Small moths with red body and wing ground colour. Forewing pattern diffuse, dark grey in combination with yellow spots. Antennae of both sexes ciliate. The male genitalia of the new genus resemble those of *Asura*, but can be easily distinguished by the combination of the following features: (1) anellus is heavily sclerotised, with two bunches of spinules dorso-basally and two bunches consisting of two spinules apically (an autapomorphic feature); (2) costa reaches the valva apex and forms two processes: subapical and apical ones separated by a broad but shallow concavity; (3) aedeagus long, narrow, S-like curved, similar to that of *Graptasura*; (4) vesica with several short diverticula bearing clusters of weak granulation and small bunches of spinules, similar to that of *Graptasura* also. The female genitalia differ clearly from those of *Asura* by the following features: (1) the presence of a well-developed antrum (absent in *Asura*); (2) the ductus bursae narrowed anteriorly; (3) the absence of scobination or spinules in corpus bursae (usually present in the nominate subspecies of *Asura*), except for a small weakly dentate posterior plate at appendix bursae base; (4) the presence of a row-shaped series of several elongated signa bursae (in *Asura* there is one signum only or signa are absent); (5) the appendix bursae is membranous (that is more or less sclerotised in the nominate subspecies of *Asura* and membranous in *Eutane*). The antrum structure of *Floridasura* is unique for the generic complex: it has broad postero-lateral lobes and a deep medial ventral concavity bounded anteriorly by a semilunar dentate plate.

Distribution. The genus is widespread from northeastern India through Myanmar, Thailand, Laos, Cambodia and southern Mainland China to Vietnam, Hainan and Taiwan Island.

Number of species. The genus is monobasic.

Genus ***Graptasura*** Hampson, 1900

(Figs 55, 56, 132, 188)

Graptasura Hampson, 1900, *Catalogue of Lepidoptera Phalaenae in the British Museum* **2**: 421 (Type species: *Cyme polygrapha* Felder, 1874, by original designation).

Diagnosis. Medium-sized or small moths with elongated wings, yellow colouration and black pattern. Antennae of both sexes ciliate. Male genitalia capsule is similar to that of the related *Asura*, but differs clearly by the absence of a medial costal process. The aedeagus structure is very similar to that of *Floridasura* and, compared to that of *Asura*, displays the following differences: in *Graptasura* the aedeagus tube is conspicuously more elongated and narrower, its apex with elongated band-like extensions protruding to the vesica base; the vesica of *Graptasura* has a characteristic finger-like dorsal diverticulum and several smaller diverticula, some of which bear narrow bunches of very small spinules, whereas in *Asura* vesica may

have various shape and diverticula configuration, but always bears large clusters (bunches or band-like ones) of longer and more robust spinules. The female genitalia of *Graptasura* are most similar to those of *Floridasura*, but differ by the antrum structure: in *Graptasura* the postero-lateral lobes are short and narrow, the medial concavity is broad and shallow, the medial dentate plate is absent, and the anterior section bears two small tubercle-like processes. In addition, *Graptasura* lacks a sclerotised posterior plate at appendix bursae base, which is present in *Floridasura*.

Distribution. Malay Peninsula, Borneo and Sulawesi Islands.

Number of species. The genus comprises four valid species.

Genus ***Xanthetis*** Hampson, 1900
(Figs 57, 58, 133, 189)

Xanthetis Hampson, 1900, *Catalogue of Lepidoptera Phalaenae in the British Museum* **2**: 414 (Type species: *Cyme luzonica* Felder, 1875, by original designation).

Diagnosis. Small moths with narrow and elongated wings and black and yellow colouration. Male antennae strongly ciliate; female antennae weakly ciliate. Male genital capsule of the genus is very characteristic and can be easily distinguished from those of other genera of the generic complex by the combination of the following features: (1) the uncus is plate-like dorso-ventrally flattened, apically rounded, setose (among members of the generic complex, the similar flattened uncus is known in the genus *Disasuridia* only, but in the latter the uncus is not setose and has a short pointed tip); (2) the tegumen broad and sclerotised; (3) the valva costa is strongly broadened distally and not reaching the valva apex; (4) the medial costal process originates from the distal broad section of costa; (5) the distal saccular process massive, well separated from the valva apex. The main autapomorphic feature of the genus is the unique aedeagus structure, which has (1) a large stick-like process anteriorly of the ductus ejaculatorius; (2) the massive conical coecum directed ventrally and being perpendicular to the aedeagus axis; (3) the distal section of aedeagus tube is longitudinally rugose and bears numerous robust spines apically. All the listed features of aedeagus are unique within the generic complex. In addition, the aedeagus vesica is narrow, tubular and bears no cornuti or spines but granulation only. Similar vesica ornamentation is known in *Karolia* and *Disasuridia* only, but in both *Karolia* and *Disasuridia* the vesici are broadened and have short lateral diverticula. The female genitalia are characterised by the presence of rugose and serrulate lateral subostial lobes, the very short ductus bursae and the small corpus bursae being heavily scobinated posteriorly and medially.

Distribution. The Philippines and northern Maluku Archipelago.

Number of species. The genus comprises two valid species. The taxon originally described as *Xanthetis luzonica pardalina* (Rothschild, 1936) is obviously related to *Chrysasura leopardina* and *C. metahyala* and is transferred here to the genus *Chrysasura*.

Genus ***Disasuridia*** Fang, 1991
(Figs 59, 60, 134, 190)

Disasuridia Fang, 1991, *Acta Entomologica Sinica* **34** (3): 356, 360 (Type species: *Disasuridia rubida* Fang, 1991, by original designation).

Diagnosis. Small moths with reddish or yellow coloration and thin black pattern. Members of *Disasuridia* are small moths being very similar superficially to members of the genus *Asuridia*. However, male and female genitalia of the genus are unmistakably recognisable. For the male genitalia the following features are characteristic: (1) the uncus is plate-like, dorso-ventrally flattened, with a short pointed tip; (2) the tegumen with short dentate lateral subapical lobes (an autapomorphic feature; lateral lobes of tegumen are also present in *Quadrasura*, but in *Quadrasura* they are very large and not dentate); (3) the apical half of tegumen fused, broad; (4) the valva is short and broad, nearly elliptical; (5) the medial costal process is massive, trigonal or thorn-like, its tip protrudes beyond the dorsal margin of sacculus; (6) the costa is heavily sclerotised, distal costal process is present, short, with a blunt tip; (7) the sacculus is narrow but heavily sclerotised, its dorsal

surface is serrulate; (8) the distal saccular process is absent, the distal section of sacculus is robust, curved dorsad and fused with the costa inwardly of the distal lobe; (9) the distal membranous lobe of valva is present, small, well separated from the sacculus and costa; (10) the aedeagus short, with two massive processes, of which the middle one is dentate (an autapomorphic feature); (11) the aedeagus vesica is short, elliptical, with two fields of weak granulation only. In the female genitalia, (1) the 7th and 8th sternites are fused; (2) the ostium bursae is narrow, situated at middle of the 8th sternite and is surrounded by four broad folds (an autapomorphic feature); (3) corpus bursae elliptical, with strong spinulose scobination, which is more robust posteriorly; (4) signum bursae is present; (5) the corpus bursae has a poster-lateral heavily sclerotised trigonal lateral process; such processes are known from the genus *Chrysasura* and some members of the genus *Afrasura* only.

Distribution. Nepal, northern and northeastern India, China and northern Indochina.

Number of species. The genus comprises seven valid species.

Genus ***Quadrasura*** Holloway, 2001
(Figs 61, 135)

Quadrasura Holloway, 2001, *Moths of Borneo* **7**: 365 (Type species: *Asura ktimuna* van Eecke, 1920, by original designation).

Diagnosis. Small moths with yellow colouration and diffuse pattern. The type species of the genus resembles externally some members of the genus *Miltochrista*. The male genitalia are characterised by (1) the presence of large lateral subapical lobes of tegumen (am autapomorphic feature); (2) the most part of tegumen being fused (an autapomorphic feature); (3) the vinculum having two short lateral processes and a shallow medial concavity; (4) the narrow valva with short costa having a short trigonal medial process but no distal process; (5) the presence of a well-developed distal saccular process; (6) the broad aedeagus with the vesica being tubular basally and comma-like curved distally, weakly granulated and bearing a small, blunt, plate-like cornutus subbasally.

Female is unknown.

Distribution. Endemic of Borneo Island.

Number of species. The genus is monobasic.

Genus ***Fossia*** Volynkin, Ivanova & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:0EC71ED7-A2E6-4DD0-AE59-83694CFFAB6E
(Figs 62, 63, 136, 191)

Type species: *Barsine melanandra* Černý, 2009.

Etymology. The generic name is dedicated to the memory of the A.V. Volynkin and M.S. Ivanova's friend, Mr Grigory G. Foss. Gender feminine.

Diagnosis. Small moths with a strongly developed sexual dimorphism, which is unique within the generic complex (only the genus *Disparsine* is sexually dimorphic also, but its dimorphism is slight and developed as the special male forewing structure having a slightly deformed costa and a row of androconial scales in the cell). Males of *Fossia* are significantly smaller than females, have narrower fore- and hindwings, more diffuse and darker forewing pattern and hindwing being almost fully black (only a subbasal area along costa is reddish, whereas in females hindwing is as forewing ground colour with a narrow blackish outer margin). Forewing colouration is red or orange, pattern is blackish. Antennae of both sexes ciliate. The male genitalia of *Fossia* remotely resemble those of *Ammatho* (*Delineatia*), but are easily recognisable by the combination of the following features: (1) the anellus is membranous (that is densely serrulate in *Delineatia*); (2) the juxta is heavily sclerotized and has two weakly serrulate apical lobes (an autapomorphic feature; a heavily sclerotised juxta is also known from the genus *Longarista*, but in *Longarista* the juxta is strongly swollen and evenly dentate, while in *Fossia* only apical processes are serrulate; in *Ammatho* (*Idopterum*) the juxta also has swollen serrulate lobes, but they are situated laterally, and, in addition, the medial part of juxta is short and weakly sclerotised, while in *Fossia* the juxta is elongated, evenly sclerotised and its processes are

situated apically); (3) the medial costal process is long and narrow, setose, directed distally (an autapomorphic feature); (4) distal section of costa is dentate, similar to that of *Delineatia*; (5) a distal membranous lobe is absent; (6) the sacculus is narrow, with a robust, thorn-like, apically pointed medial process directed dorsally (similar well-developed medial saccular processes are also known from the genera *Melanaema*, *Laminasura* and *Sarbine*, but they are not so heavily sclerotised); (7) the distal saccular process is present as a small blunt protrusion; (8) the aedeagus is small, narrow and straight; (9) the aedeagus vesica structure is uniform within the genus and is characterised by the presence of a large membranous ventral diverticulum and a globular medial dorsal diverticulum covered with numerous very small cornuti or robust granules. The female genitalia structure of *Fossia* is unique within the generic complex and is characterised by the following features: (1) the presence of a heavily sclerotised bilobate antevaginal plate fused with the 8th sternite and having a very narrow ostium bursae in its middle part (an autapomorphic feature); (2) the antrum is present, very short and narrow, but heavily sclerotised and fused with the antevaginal plate (an autapomorphic feature); (3) the anterior section of ductus bursae is very short and narrow, tube-like, membranous; (4) corpus bursae is elliptical, weakly scobinated, with one elliptical signum bursae medially.

Distribution. The genus is widespread in Himalaya, western and southern China and Indochina.

Number of species. The genus comprises five valid species and three subspecies and was reviewed as 'the *Barsine punicea* species-group' by Volynkin *et al.* (2018b).

Genus ***Melanaema*** Butler, 1877
(Figs 64, 65, 137, 192)

Melanaema Butler, 1877, *The Annals and magazine of natural history* (4) **20** (119): 397 (Type species: *Melanaema venata* Butler, 1877, by monotypy).

Diagnosis. Medium-sized moths with reddish and yellowish wing colouration and reduced pattern consisting of a blackish suffusion on veins only. Male antennae are bipectinate; female antennae are ciliate. The male genitalia resemble those of *Ammatho* due to the well-developed distal costal and saccular processes, distal lobe of valva and the presence of numerous short cornuti in aedeagus vesica, but differ clearly by the absence of a medial costal process and the presence of the medial saccular process (in *Ammatho* that is present in the subgenus *Conicornuta* only, but in the latter it is very short). The female genitalia are characterised by the following features: (1) the antrum having two large lateral lobes separated by a deep and narrow medial ventral concavity (similar to that of *Moorasura*); (2) the narrowed posterior section of corpus bursae having two sclerotised and serrulate areas protruding to the basal section of appendix bursae; (3) the elliptical anterior section of corpus bursae bearing weak spinulose scobination and a round signum; (4) the large membranous appendix bursae directed posteriorly.

Distribution. The type species of the genus is distributed in southern Russia Far East, eastern China, Korean Peninsula and Japan.

Number of species. Here we consider the genus to be monobasic. New Guinean *Melanaema sanguinea* Hampson, 1900 is obviously not congeneric with *M. venata* and is provisionally placed here into the genus *Integrivalvia*.

Genus ***Wittasura*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:864E7B41-ACD4-4298-91CC-5E7AEB61B8C0
(Figs 66, 67, 138, 194)

Type species: *Ammatho lineatus* Walker, 1855.

Etymology. The generic name is a combination of the surname Witt and the generic name *Asura*. The genus is dedicated to the memory of Dr Thomas J. Witt, a famous lepidopterist and a founder of the Museum Witt Munich having one of the largest collections of Heterocera in the world. Gender feminine.

Diagnosis. Small moths with orange colouration and blackish pattern consisting of transverse lines and longitudinal strokes. The male genitalia of *Wittasura* are very characteristic and can be easily recognised by the complex of the following features: (1) the vinculum is rectangular, weakly sclerotised, with flimsy

coremata apically (an autapomorphic feature); (2) a distal membranous lobe of valva is absent; (3) the valva apex is heavily sclerotised, strongly curved dorsally, lobe-like with two thorn-like processes (an autapomorphic feature); (4) the medial costal process is short, thorn-like; (5) the sacculus is narrow but heavily sclerotised, distally fused with the costa, bears a two thorn-like processes a distal and a subdistal ones (an autapomorphic feature); (6) the aedeagus is robust, slightly curved; (7) the aedeagus vesica with several various shaped diverticula ornamented with fields of various sized granulation, and an elongated robust cornutus, which is segmented plate-like. The female genitalia of *Wittasura* most resemble those of the genus *Aberrasine* due to the spinulose corpus bursae and the appendix bursae being curved basally, directed anteriorly and having a heavily sclerotised basal section, but in *Wittasura* the ductus has a well-developed, strongly broadened antrum, which is absent in *Aberrasine*.

Distribution. The genus is distributed in Sundaland, the Philippines and Sulawesi Island.

Number of species. The genus comprises three valid species and one subspecies.

Genus ***Disparsine*** Volynkin, **gen. nov.**

https://zoobank.org/urn:lsid:zoobank.org:act:FA5D6D1B-ABE5-43A4-837C-516D0D387CA0

(Figs 68, 69, 139, 193)

Type species: *Asura crustata* Talbot, 1926.

Etymology. The generic name is a combination of the Latin word ‘dispar’ meaning ‘unpaired’ and the generic name *Barsine*. The name refers to the sexual dimorphism of the genus. Gender feminine.

Diagnosis. Small moths with orange colouration, blackish pattern and slightly expressed sexual dimorphism: males have the forewing costa being wavy deformed and having a longitudinal row of androconial scales in the cell. In addition, in males the forewing pattern is reduced. Antennae of both sexes are ciliate. In the male genitalia, the valva is curved, blade-shaped, solid, without distal costal and distal saccular processes, similar to valvae of the genera *Tumicla* and the recently described *Parvuspina*, but also has the medial costal process being narrow, stick-like and situated subbasally, and the very small denticle-like medial saccular process. Such a combination of valva features is unique for the generic complex. The aedeagus is large, straight; the vesica structure is unspecific and is similar to that of *Ammatho*: it has several short diverticula ornamented with fields of various sized granulation and a cluster of short, stout, various sized cornuti medially. The female genitalia are characterised by the combination of the following features: (1) the antrum is present, has a deep medial concavity and is strongly connected to the 7th abdominal sternite; (2) the ductus bursae is dorso-ventrally flattened and evenly sclerotised; (3) the corpus bursae is broad, strongly spinulose scobinated; (4) the appendix bursae is small, conical, situated postero-dorsally and directed posteriorly. The female genitalia of *Disparsine* are similar to those of *Albarrania*, but in the latter genus the antrum is connected to the 7th sternite by a membrane, and the appendix bursae is larger and more heavily sclerotised.

Distribution. The genus is widespread in Sundaland and Sulawesi Island.

Number of species. The genus comprises two valid species.

Genus ***Moorasura*** Volynkin & Huang, **gen. nov.**

https://zoobank.org/urn:lsid:zoobank.org:act:C50226BA-D073-49F2-80A2-94D819FA2C59

(Figs 70, 71, 140, 195)

Type species: *Barsine gloriosa* Moore, 1878.

Etymology. The generic name is a combination of the Frederic Moore’s surname, who is an author of its type species, and the generic name *Asura*. Gender feminine.

Diagnosis. Medium-sized moths with orange colouration and dark brown or blackish pattern similar to that of *Ammatho* (*Ammathella*). Antennae of both sexes are ciliate. The male genitalia of *Moorasura* resemble those of *Ammatho* due to the presence of distal costal and distal saccular processes, a distal membranous lobe of valva, and the aedeagus vesica having fields of granulation and clusters of small cornuti or spinules. However, unlike *Ammatho*, in *Moorasura* the medial costal process is reduced, very small and

situated at the dorsal valva margin, while in *Ammatho* it is situated more inwardly. The sacculus of *Moorasura* has a small and weak medial process, similar to that of *Ammatho* (*Conicornuta*). In addition, in *Moorasura* the tegumen is conspicuously curved ventrad, the feature found in *Longarista* also. The main autapomorphic feature of the genus is a swollen basal section of uncus (reaching its extreme form in *B. inflexa*, see Volynkin & Černý (2019b)). The female genitalia of *Moorasura* resemble those of *Ammatho*, but in *Moorasura* the ductus bursae is narrow and the appendix bursae is membranous and directed posteriorly.

Distribution. The genus is widespread in Sundaland and Sulawesi Island.

Number of species. The genus comprises six valid species reviewed by Volynkin & Černý (2019b) as 'the *Barsine inflexa* species-group'.

Genus ***Sarbine*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:B4EDCA7A-52A0-467C-A978-0EDBE54DDEA1
(Figs 72–75, 141, 142, 196, 197)

Type species: *Miltochrista flavodiscalis* Talbot, 1926.

Etymology. The generic name is an anagram of the name *Barsine*. Gender feminine.

Diagnosis. Small or medium-sized moths with reddish or orange colouration and pattern resembling that of *Moorasura* and some *Ammatho* and consisting of blackish brown transverse lines and longitudinal strokes and yellow spots. The male genitalia are similar to those of *Moorasura* but differ by the uncus being evenly narrow (that is basally swollen in *Moorasura*), the tegumen being not curved ventrally, the absence of a medial costal process (that is present in *Moorasura*, but small), and the well-developed and heavily sclerotised medial saccular process, which is weak and very short in all *Moorasura* species. In addition, in *Sarbine* the costa has a short medial protrusion directed dorsally. The aedeagus vesica of *Sarbine* has several diverticula ornamented with clusters of granulation, short but stout cornuti, bunches of small spinules or separated large spike-like cornuti. The female genitalia of *Sarbine* differ from those of *Moorasura* by the absence of an antrum.

Remark. The genus was reviewed by Volynkin & Černý (2019b) as 'the *Barsine flavodiscalis* species-group'.

Subgenus *Sarbine* Volynkin, **subgen. nov.**
(Figs 72, 73, 141, 196)

Diagnosis. The male genitalia of the nominate subgenus are characterised by the large medial costal protrusion directed dorsally, the short, but prominent distal costal process, the sacculus being not setose dorsally, the short trigonal medial saccular process situated medially, the not developed distal saccular process, the curved aedeagus and the vesica ornamented with clusters of strong granulation, short but stout cornuti and bunches of small spinules. The female genitalia are characterised by the ductus bursae being sclerotised medially and posteriorly and membranous anteriorly, the corpus bursae densely covered with spinulose scobination, and the membranous appendix bursae situated postero-laterally and directed posteriorly.

Distribution. The nominate subgenus is distributed in Indochina and Borneo.

Number of species. The genus comprises two valid species.

Subgenus *Processine* Volynkin, **subgen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:B000A9EE-AC9B-406D-849F-864AF7F9BBC8
(Figs 74, 75, 142, 197)

Type species: *Hypoprepia cruciata* Walker, 1862.

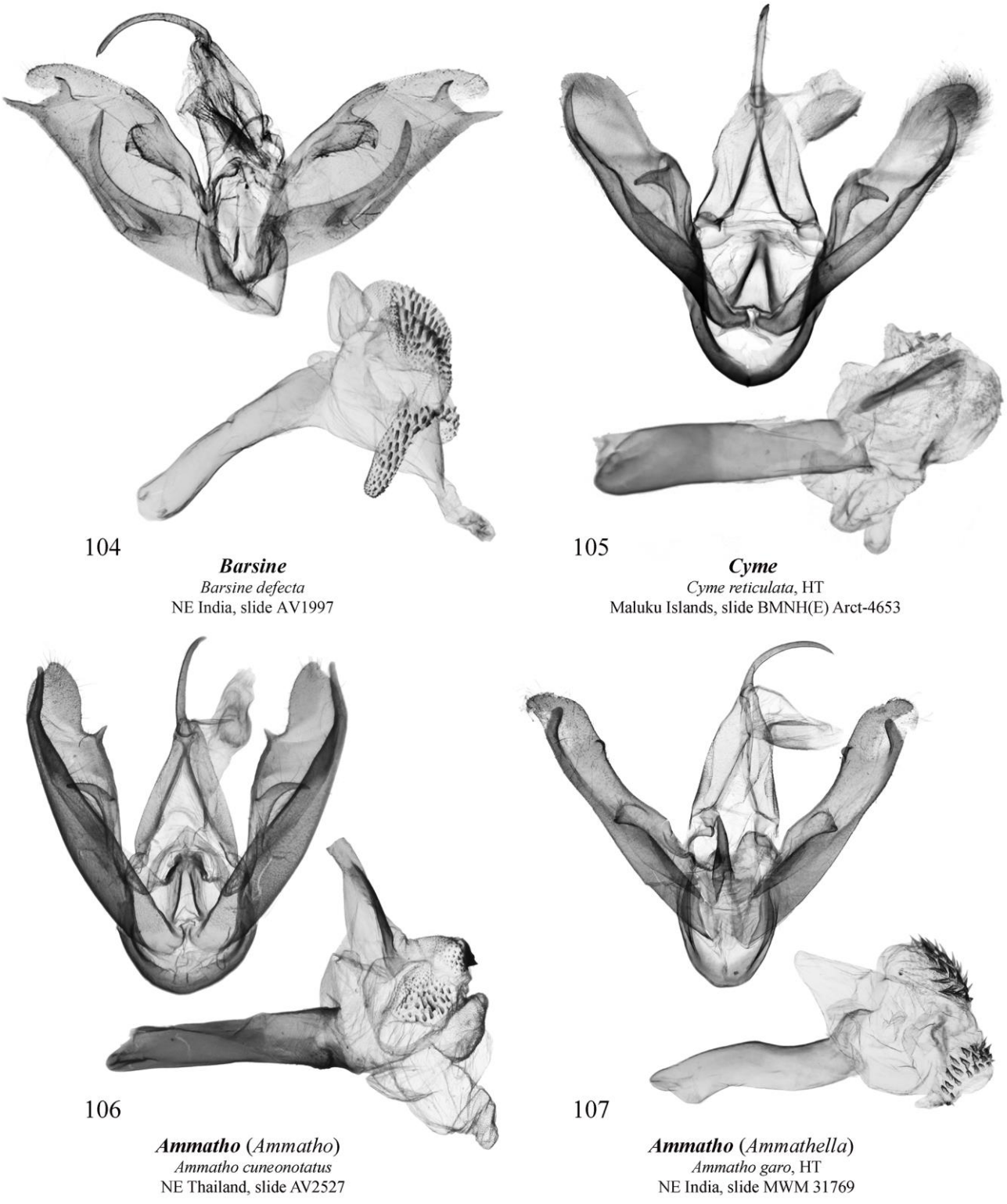
Etymology. The subgeneric name is a combination of the word 'process' and the generic name *Barsine*. The name refers to the well-developed, extremely elongated medial saccular process. Gender feminine.

Diagnosis. The male genitalia of the subgenus *Processine* are characterised by the small medial costal protrusion, the not developed distal costal process, the setose sacculus, the well-developed trigonal distal saccular process, the very long, narrow and curved medial saccular process situated subapically, the straight

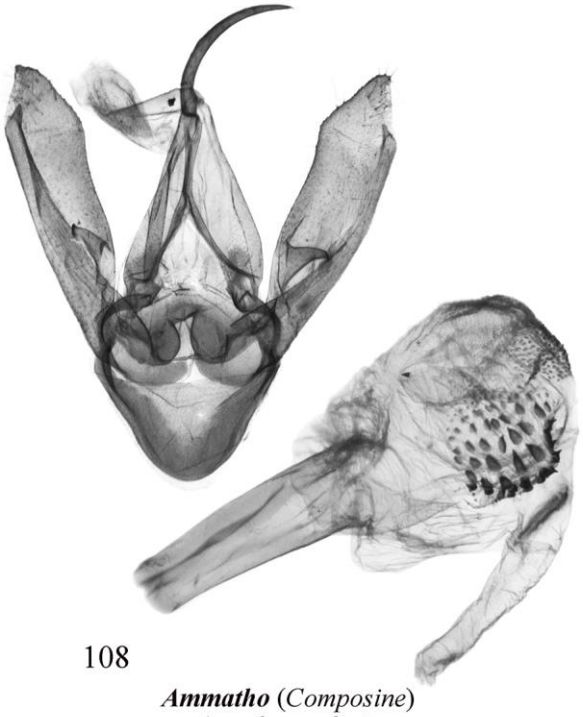
aedeagus, and the membranous vesica having only one field of weak granulation and a very long robust spine on a dentate base.

Distribution. The subgenus is distributed in Sundaland.

Number of species. The genus comprises two valid species.



Figures 104–107. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: male genitalia.

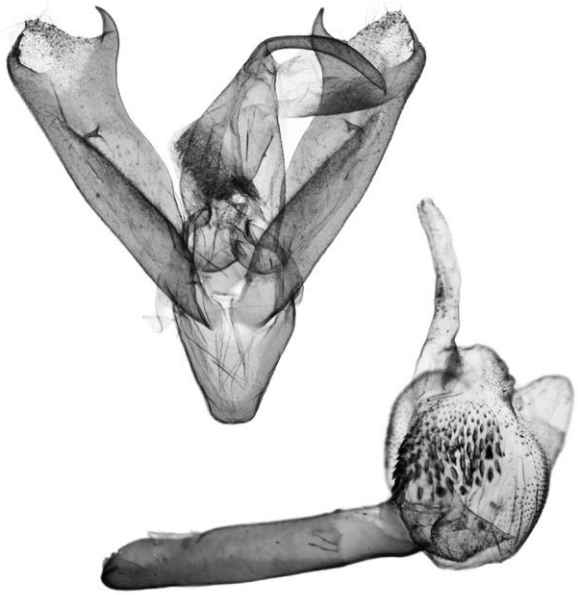


108

Ammatho (*Composine*)
Ammatho complicata
Malay Peninsula, slide MWM 33902

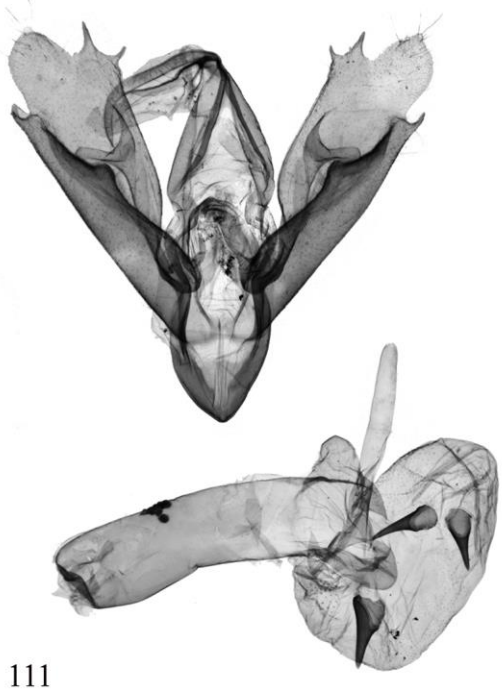
109

Ammatho (*Idopterum*)
Ammatho ovale
N Thailand, Nan prov., slide MWM 31381



110

Ammatho (*Striatella*)
Ammatho hypoprepioides
Borneo, slide MWM 33657



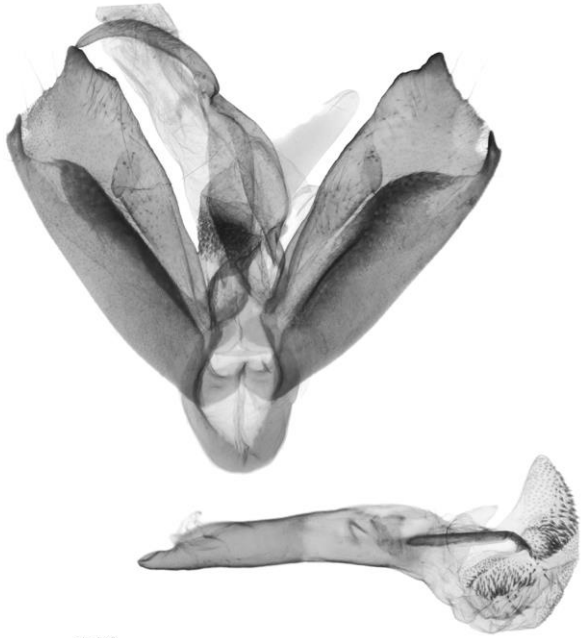
111

Ammatho (*Conicornuta*)
Ammatho convexa
Taiwan, slide Arct-5271

Figures 108–111. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: male genitalia.

112

Ammatho (*Delineatia*)
Ammatho delineata
China, Zhejiang prov., slide AV1892



113

Ammatho (*Delineatia*)
Ammatho sublucana
N Thailand, slide AV2361

114

Ammatho (*Rugosine*)
Ammatho salakia
Indonesia, Bali Isl., slide AV3645

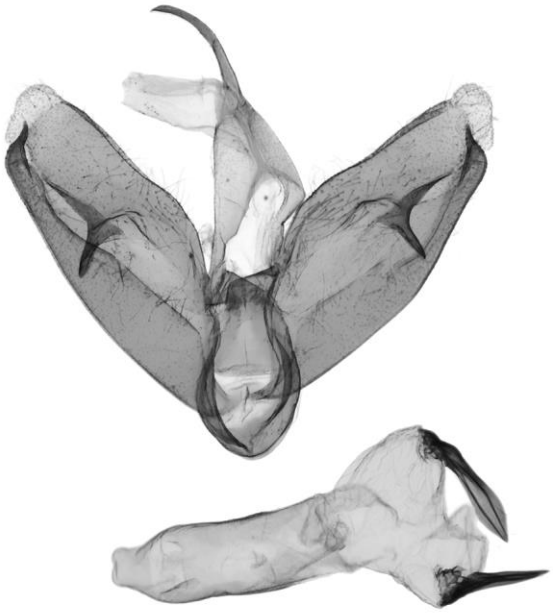
115

Ovipennis (*Ovipennis*)
Ovipennis dudgeoni
Nepal, slide MWM 35754

Figures 112–115. Members of the *Asura* / *Miltochrista* generic complex: male genitalia. 112, 114, 115 – type species of genera and subgenera.

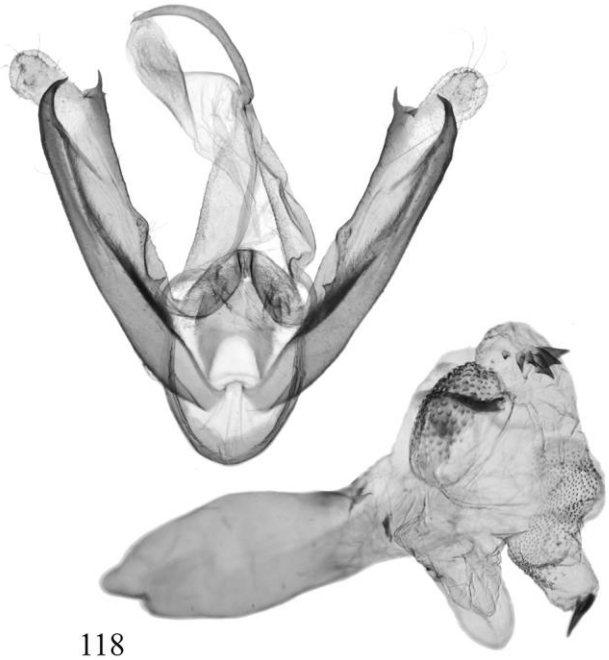
116

Ovipennis (*Barsipennis*)
Ovipennis joshii, HT
NE India, slide MWM 31457



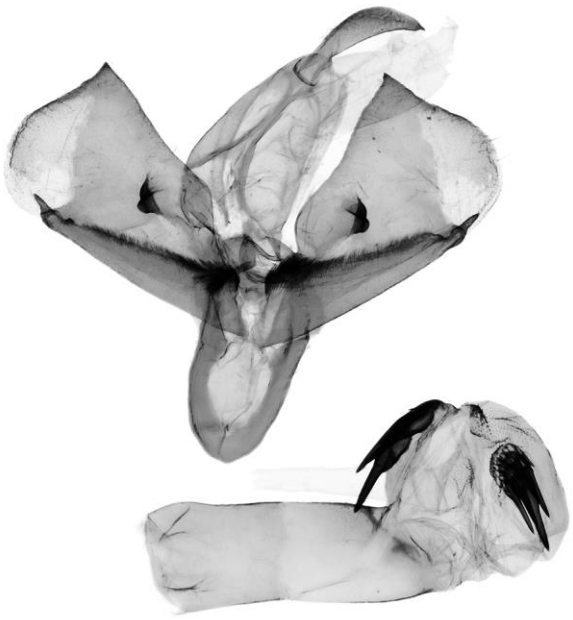
117

Ovipennis (*Nebulene*)
Ovipennis nebulosa
China, Sichuan, slide MWM 31370



118

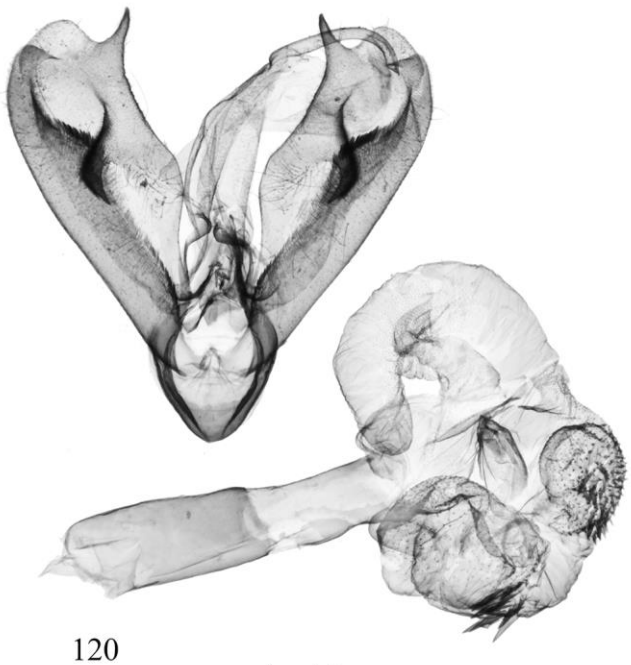
Ovipennis (*Coccinigripennis*)
Ovipennis incompletostriga, HT
NE India, slide MWM 31435



119

Barsochrista
Barsochrista kishidai
S India, slide MWM 31563

Figures 116–119. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: male genitalia.

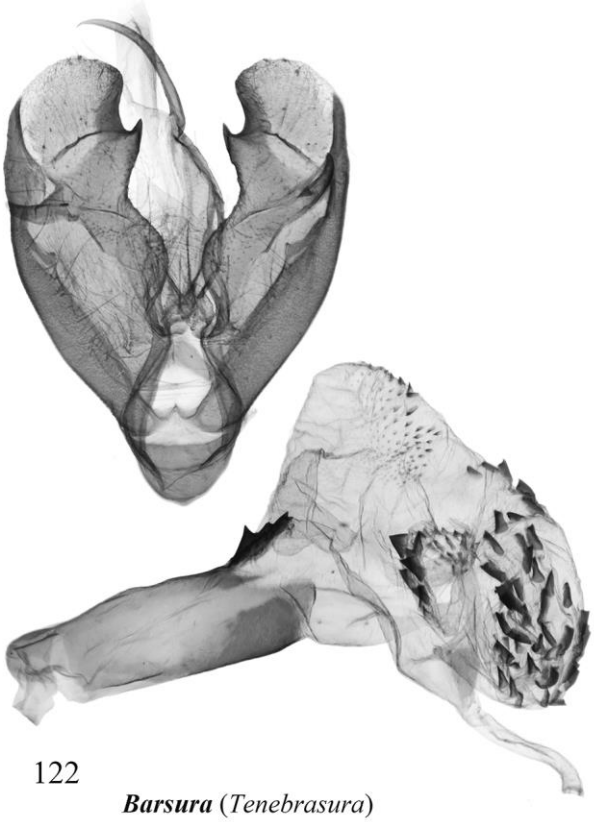


120

Asuridia
Asuridia carnipicta
China, Zhejiang, slide NHMUK010313432

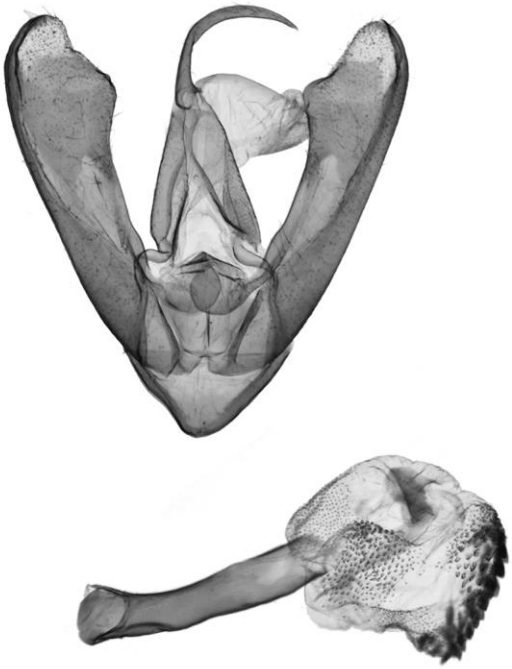
121

Barsura (*Barsura*)
Barsura nubifascia
C Nepal, slide MWM 31358



122

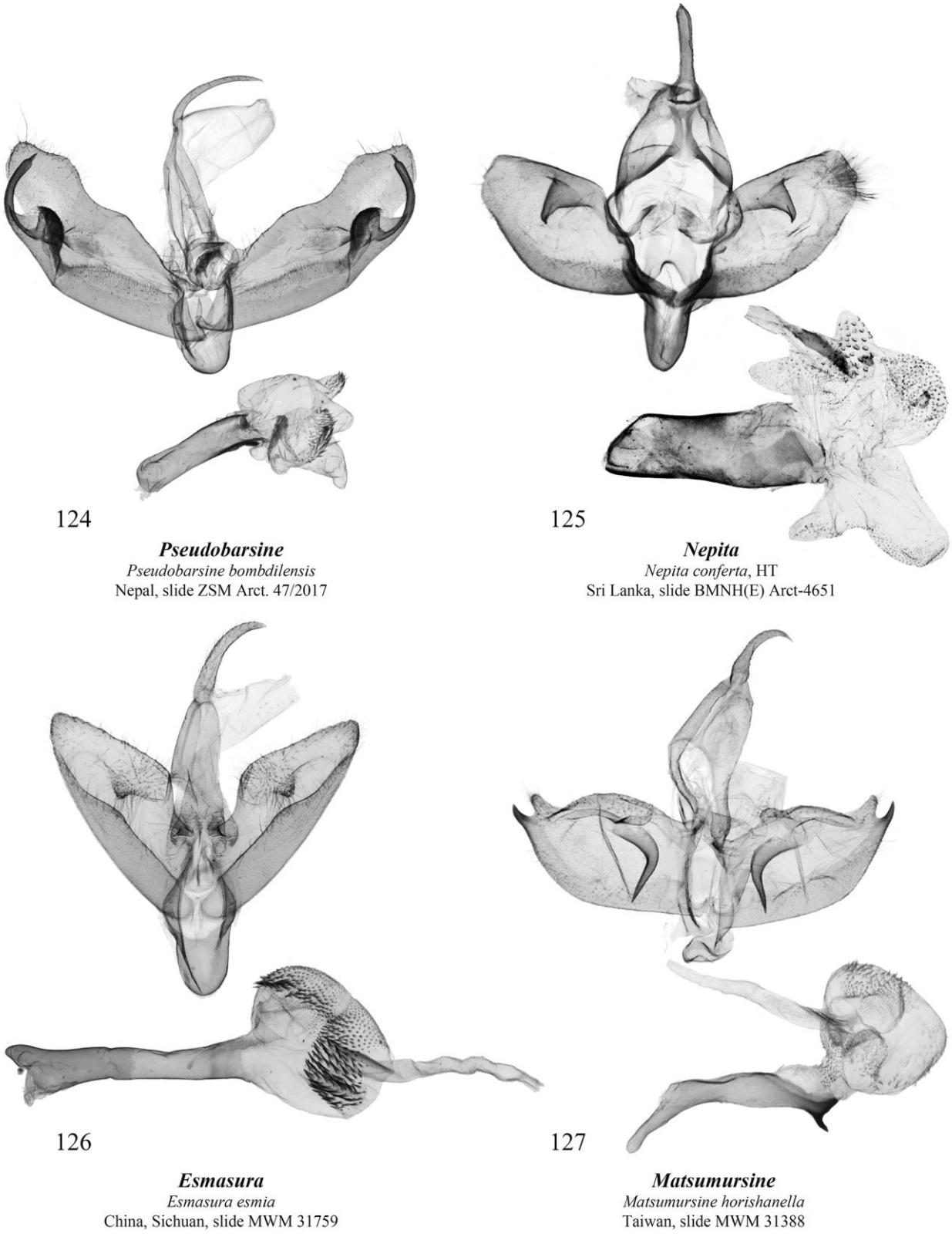
Barsura (*Tenebrasura*)
Barsura albidorsalis
Taiwan, slide AV3653



123

Argentosine
Argentosine proleuca
Nepal, Annapurna Himal, slide MWM 33579

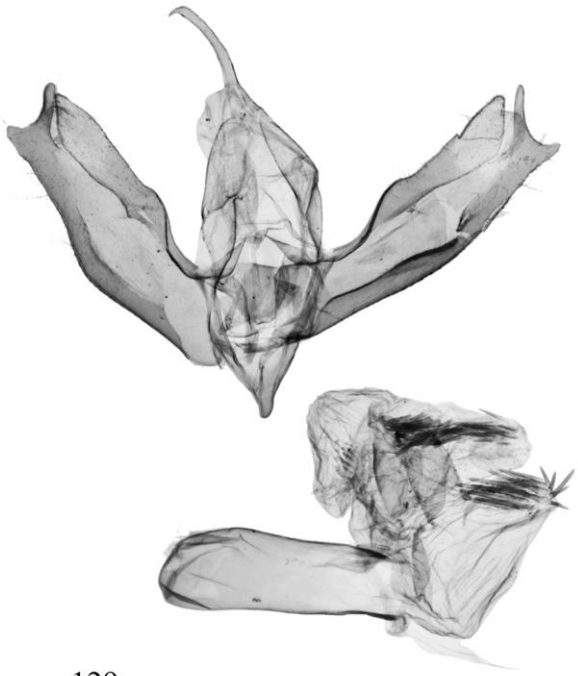
Figures 120–123. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: male genitalia.



Figures 124–127. Type species of genera of the *Asura* / *Miltochrista* generic complex: male genitalia.

128

Asura (*Asura*)
Asura cervicalis
N Australia, slide AV2380

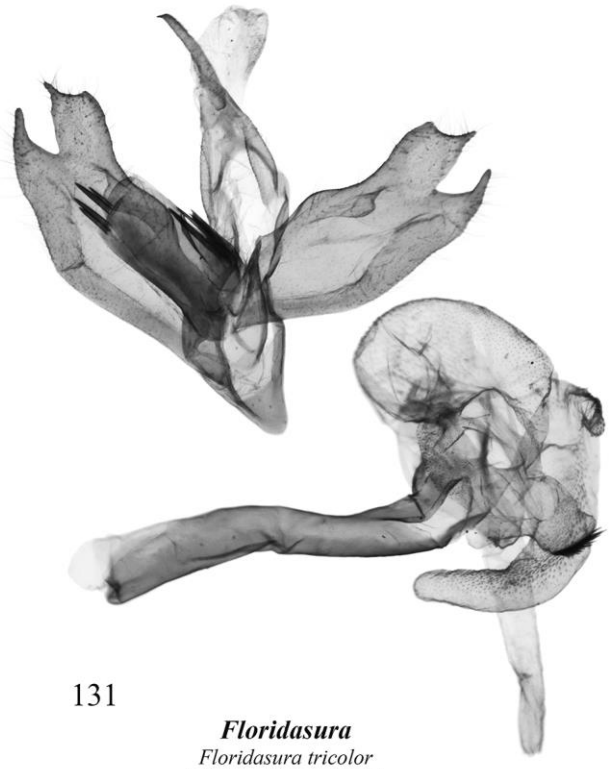


129

Asura (*Asura*)
Asura saginaea, type species of *Gymnasura*
N Australia, slide BMNH(E) Arct.-5265

130

Asura (*Eutane*)
Asura terminalis
Australia, Queensland, slide BMNH(E) Arct.-4573



131

Floridasura
Floridasura tricolor
Taiwan, slide AV2385

Figures 128–131. Type species of genera, subgenera and their synonyms of the *Asura* / *Miltochrista* generic complex: male genitalia.

132

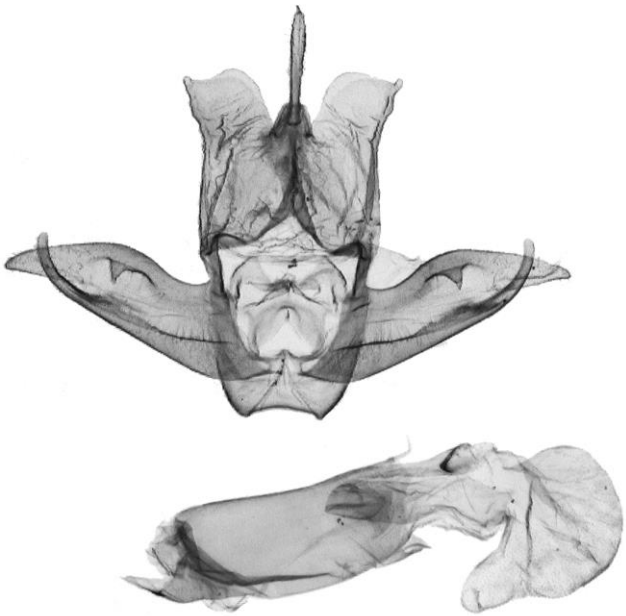
Graptasura
Graptasura polygrapha
Indonesia, Sulawesi, slide BMNH(E) Arct.-5022

133

Xanthetis
Xanthetis luzonica
The Philippines, Luzon Isl., slide NHMUK010313483

134

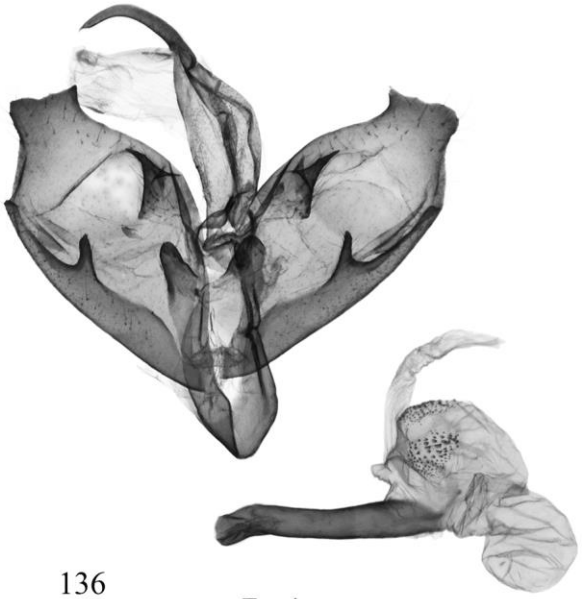
Disasuridia
Disasuridia rubida
N Thailand, Chiang Mai Prov., slide AV2384



135

Quadrasura
Quadrasura ktimuna, ST
W Borneo, slide RMNH.INS.1104137-JDH20

Figures 132–135. Type species of genera of the *Asura* / *Miltochrista* generic complex: male genitalia. Figure 135: photo by R. de Vos.

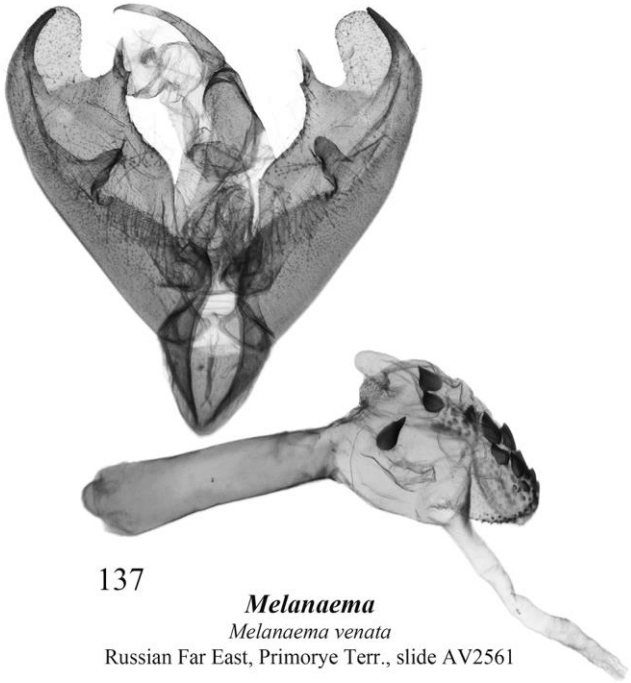


136

Fossia

Fossia melanandra, PT

SW Thailand, Chumphon prov., slide AV2392



137

Melanaema

Melanaema venata

Russian Far East, Primorye Terr., slide AV2561

138

Wittasura

Wittasura lineatus

Borneo, slide BMNH(E) Arct.-4752

139

Disparsine

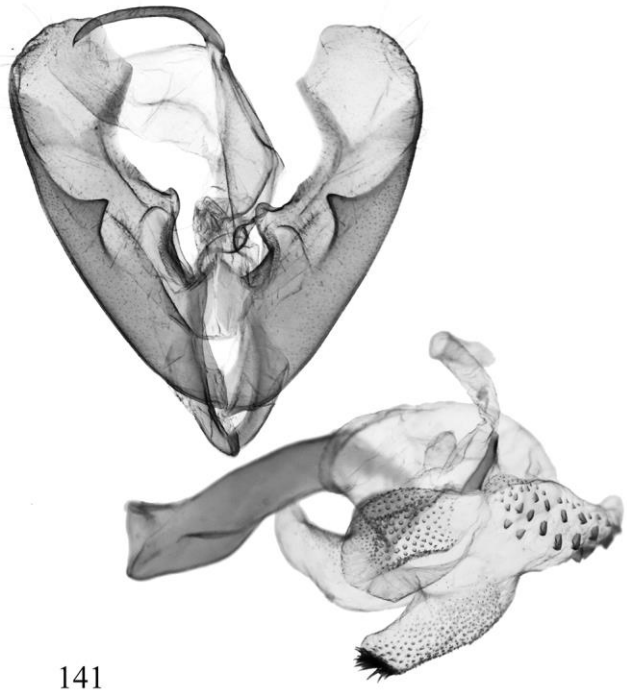
Disparsine crustata

Borneo, slide BMNH(E) Arct.-4741

Figures 136–139. Type species of genera of the *Asura* / *Miltochrista* generic complex: male genitalia.

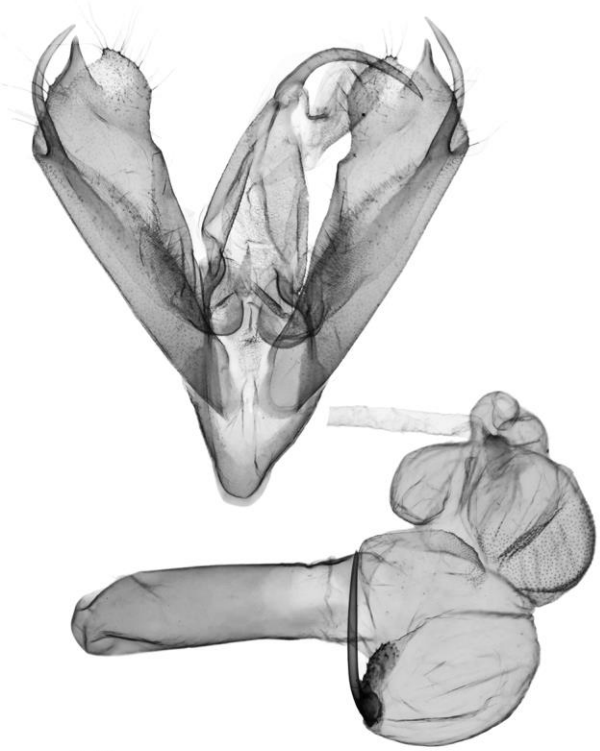
140

Moorasura
Moorasura gloriosa
NE India, slide MWM 33699



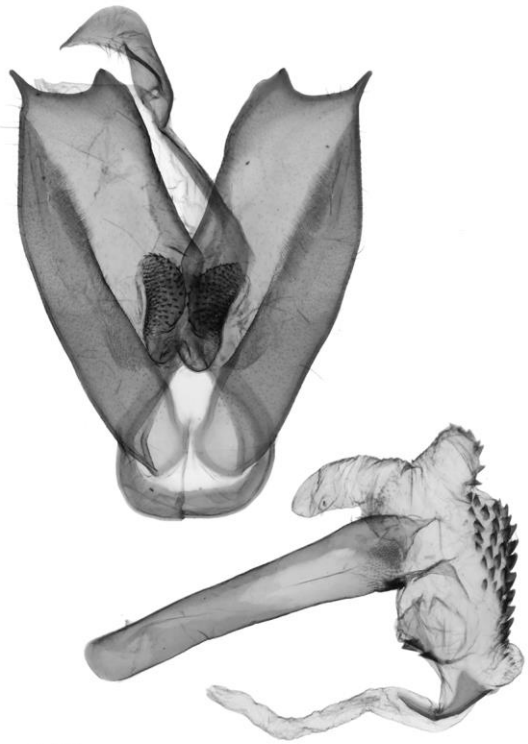
141

Sarbine (*Sarbine*)
Sarbine flavodiscalis
Borneo, slide MWM 33693



142

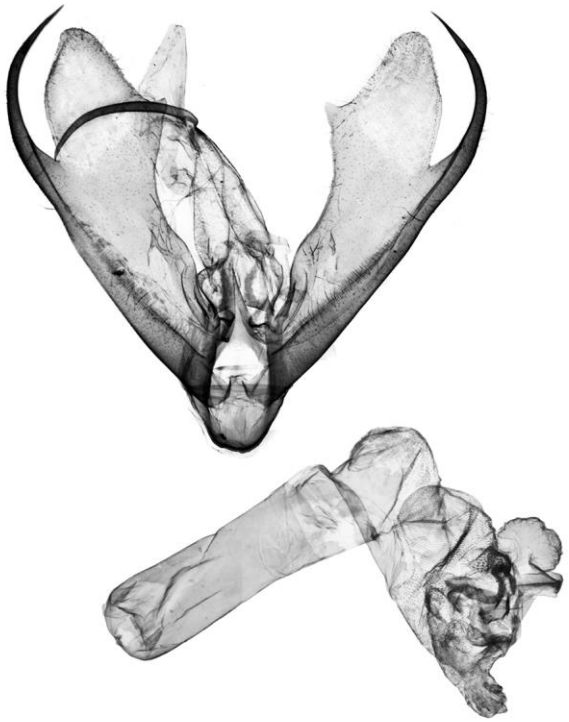
Sarbine (*Processine*)
Processine cruciata
Malay Peninsula, slide MWM 33601



143

Longarista
Longarista longaria
China, Guangdong, slide AV4241

Figures 140–143. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: male genitalia.



144

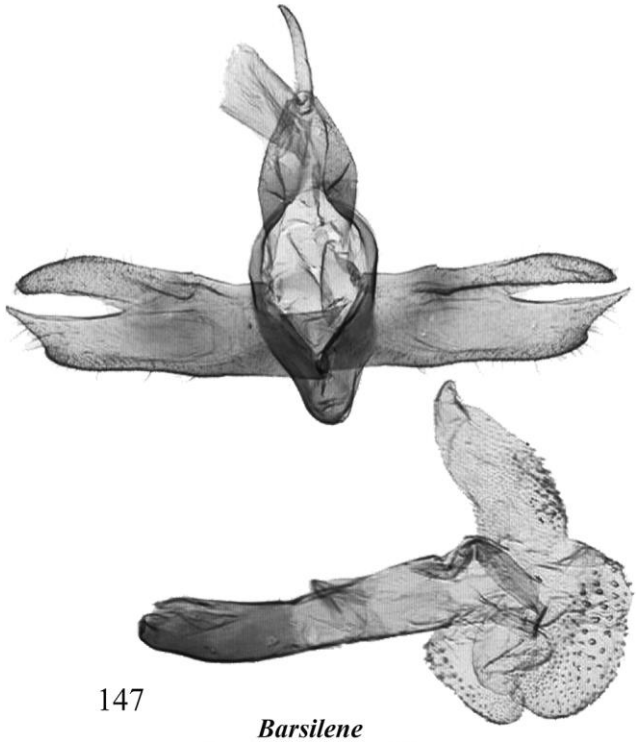
Hampsonascia
Hampsonascia dentifascia
NE India, Khasis, slide NHMUK010313499

145

Cernysura
Cernysura orbis, PT
The Philippines, Luzon Isl., slide AV3739

146

Karolia
Karolia stefaniae
Indonesia, Sumatra Isl., slide MWM 31754



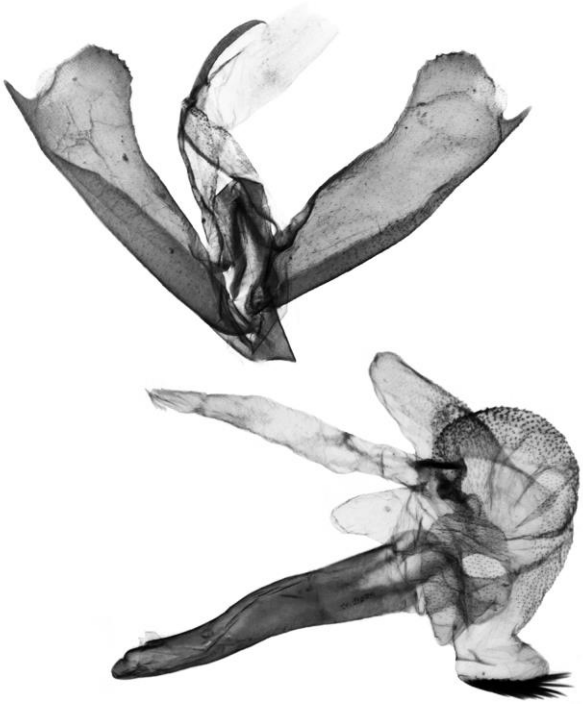
147

Barsilene
Barsilene pallinflexa, HT
Borneo, slide BMNH(E) Arct.-4750

Figures 144–147. Type species of genera of the *Asura* / *Miltochrista* generic complex: male genitalia.

148

Nanarsine
Nanarine porphyrea
Borneo, slide BMNH(E) Arct.-4754

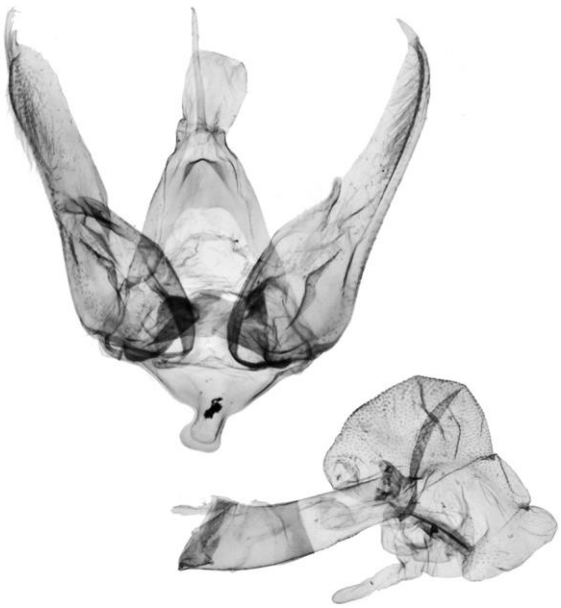


149

Nanarsine
Nanarine senara
Indonesia, Sumatra Isl., slide AV3641

150

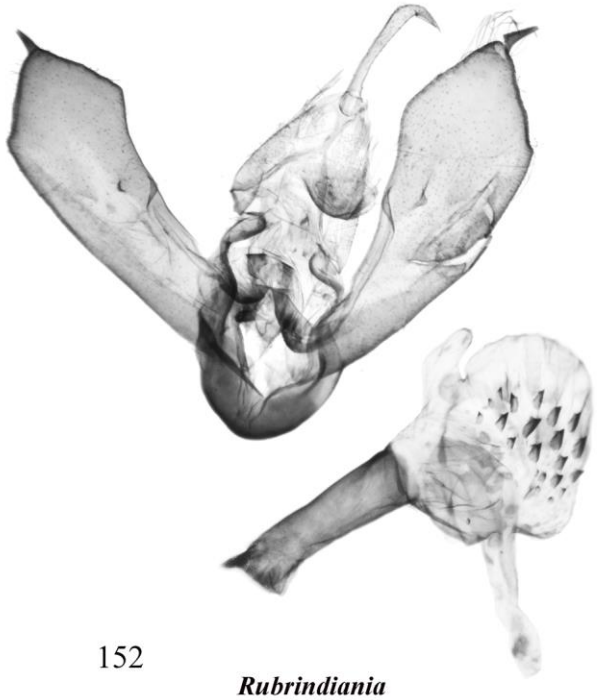
Amphisine
Amphisine perpusilla
Indonesia, Sumatra Isl., slide AV3394



151

Niveutane
Niveutane nivea
Borneo, slide BMNH(E) Arct.-5009

Figures 148–151. Members of the *Asura* / *Miltochrista* generic complex: male genitalia. 148, 150, 151 – type species of genera.

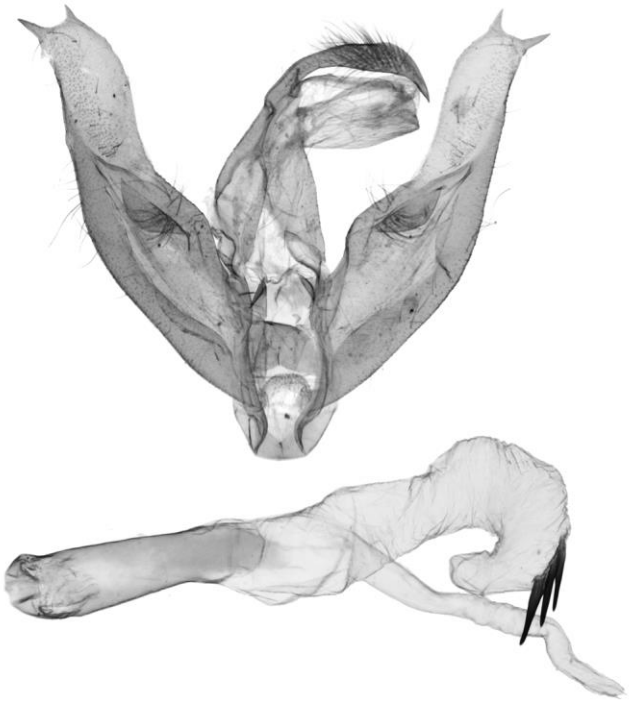


152

Rubrindiania
Rubrindiania cardinalis
NE India, slide BMNH(E) Arct-6487

153

Indiania
Indiania eccentropis
N Thailand, slide AV2736



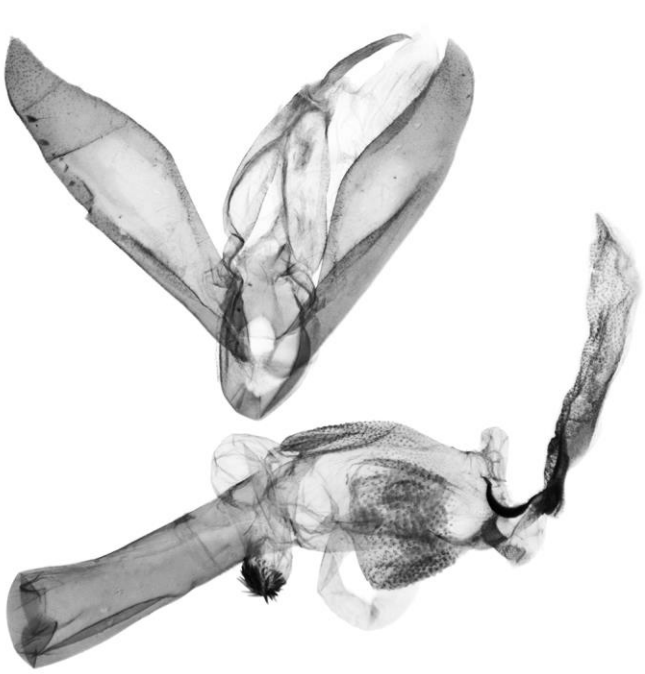
154

Barsaurea
Barsaurea phaeoxanthia
NE India, Khasis, slide AV1898

155

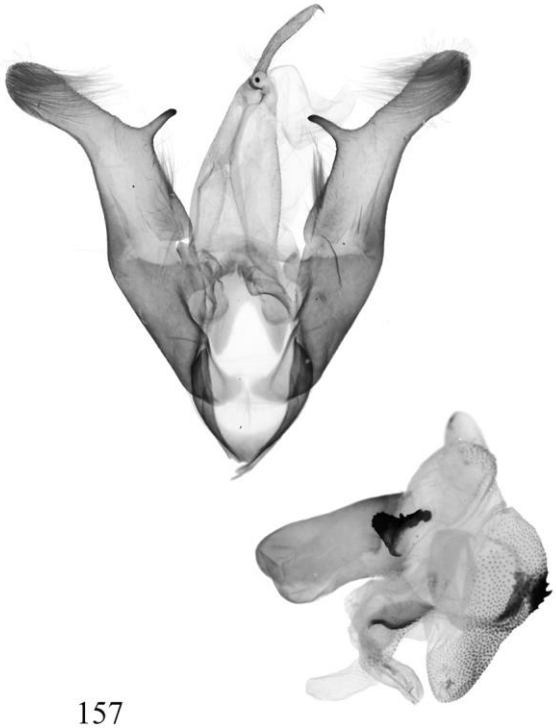
Barsaurea
Barsaurea diehli, HT
Indonesia, Sumatra, slide AV1716

Figures 152–155. Members of the *Asura* / *Miltochrista* generic complex: male genitalia. 152–154 – type species of genera.



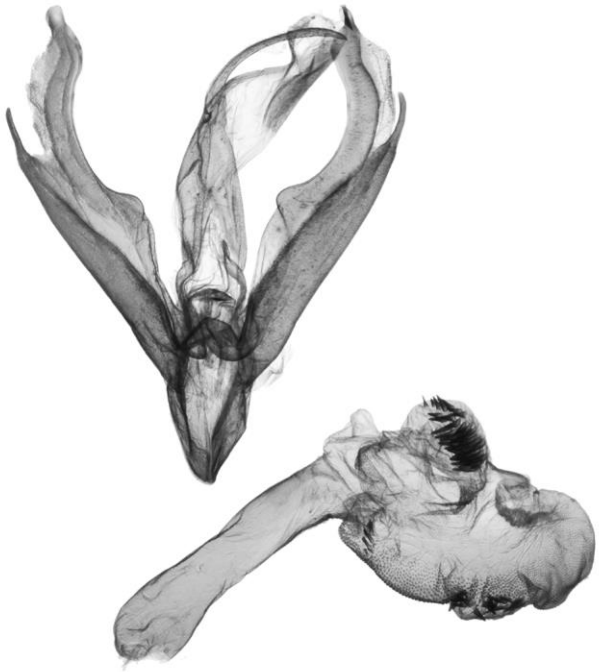
156

Integrivalvia
Integrivalvia exclusa
The Philippines, Palawan Isl., slide AV3621



157

Albarrania
Albarrania podbolotskayae
Indonesia, Flores Isl., slide MWM 34597



158

Aberrasine
Aberrasine aberrans
Japan, Honshu, slide AV2101

159

Aberrasine
Aberrasine collina
S Vietnam, slide AV2327

Figures 156–159. Members of the *Asura* / *Miltochrista* generic complex: male genitalia. 156–158 – type species of genera.

Genus ***Longarista*** Volynkin, 2019
(Figs 76, 77, 143, 198)

Longarista Volynkin, 2019, *Ecologica Montenegrina* **20**: 98 (Type species: *Miltochrista longaria* Daniel, 1951, by original designation).

Diagnosis. Small moths with orange or red colouration and blackish pattern. Species of the genus resemble superficially some species of *Ammatho* and females of *Fossia*. Antennae of both sexes are ciliate. The autapomorphic feature of *Longarista* male genitalia is the unique structure of its juxta, which is swollen, heavily sclerotised and densely covered with small denticles, whereas other members of the generic complex have flattened juxta, only *Ammatho* (*Idopterum*) has lateral swollen dentate lobes, and *Fossia* has the juxta with serrulate apical processes. In addition, the male genitalia of *Longarista* are characterised by the combination of such features as (1) the absence of a medial costal process; (2) the absence of a distal membranous lobe of valva; (3) the prominent and heavily sclerotised distal costal and saccular processes; and (4) the aedeagus vesica having several diverticula armed with clusters of small but stout trigonal cornuti. The female genitalia of *Longarista* are characterized by the presence of a swollen postvaginal plate, which is absent or flattened in other genera of the *Asura / Miltochrista* generic complex.

Distribution. Southern China and northern Indochina.

Number of species. The genus comprises two valid species reviewed by Volynkin (2019a).

Genus ***Hampsonascia*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:2B50E4B5-9600-4DB9-BC47-34AC692C75CA
(Figs 78, 79, 144, 199)

Type species: *Miltochrista dentifascia* Hampson, 1894.

Etymology. The generic name is a combination of the Sir George Hampson's surname, who is an author of its type species, and the type species name *dentifascia*. Gender feminine.

Diagnosis. Small moths with reddish colouration and blackish 'miltochristoid' pattern. Antennae of both sexes are ciliate. The male genital capsule of *Hampsonascia* is similar to that of *Miltochrista*, by the well-developed distal costal and saccular processes and the distal membranous lobe as well, but differ by the presence of a very small, denticle-like medial costal process. The autapomorphic feature of the male genitalia of *Hampsonascia* is the vesica structure, which has several very long and narrow granulated diverticula which are usually almost impossible to evert. The autapomorphic feature of the female genitalia is the extremely long, narrow and hook-like curved anteriorly appendix bursae with spinulose scobination inwardly, which corresponds to the long diverticula of male aedeagus vesica. In addition, the female genitalia of *Hampsonascia* are characterised by (1) the presence of a rugose antevaginal plate having two broadened lateral lobes; (2) the relatively broad ductus bursae being dorso-ventrally flattened and evenly sclerotised; and (3) the corpus bursae being conspicuously shorter than the appendix bursae and having several fields of various sized spinules.

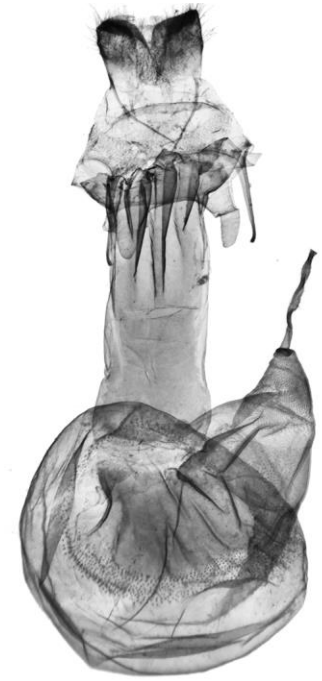
Distribution. Northeastern India, southern China and northern Indochina.

Number of species. The genus is monobasic.

Genus ***Cernysura*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:61D91D84-A088-48C3-A2AC-775A66B7FA19
(Figs 80, 81, 145, 200)

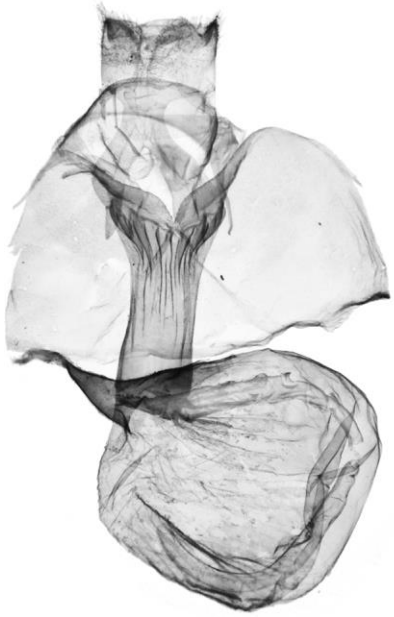
Type species: *Miltochrista orbis* Černý, 1995.

Etymology. The generic name is a combination of the Dr Karel Černý's surname, who is an author of its type species, and the generic name *Asura*. Gender feminine.



160

Barsine
Barsine defecta
NE India, slide NHMUK010313302

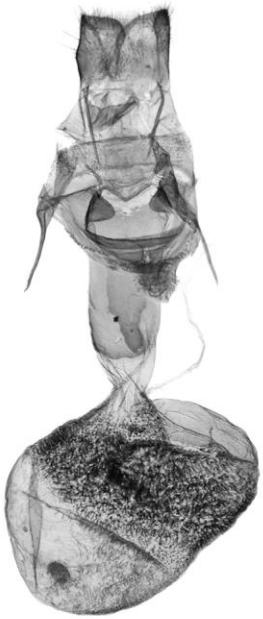


161

Cyme
Cyme reticulata
The Maluku Isl., slide BMNH(E) Arct.-4654

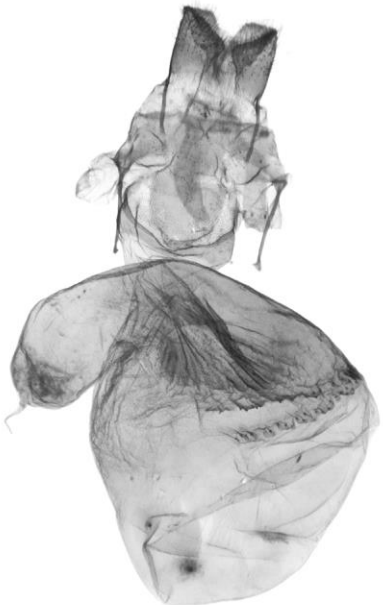
162

Ammatho (*Ammatho*)
Ammatho cuneonotatus
NE Thailand, slide AV2528



163

Ammatho (*Ammathella*)
Ammatho garo, PT
NE India, slide MWM 31770



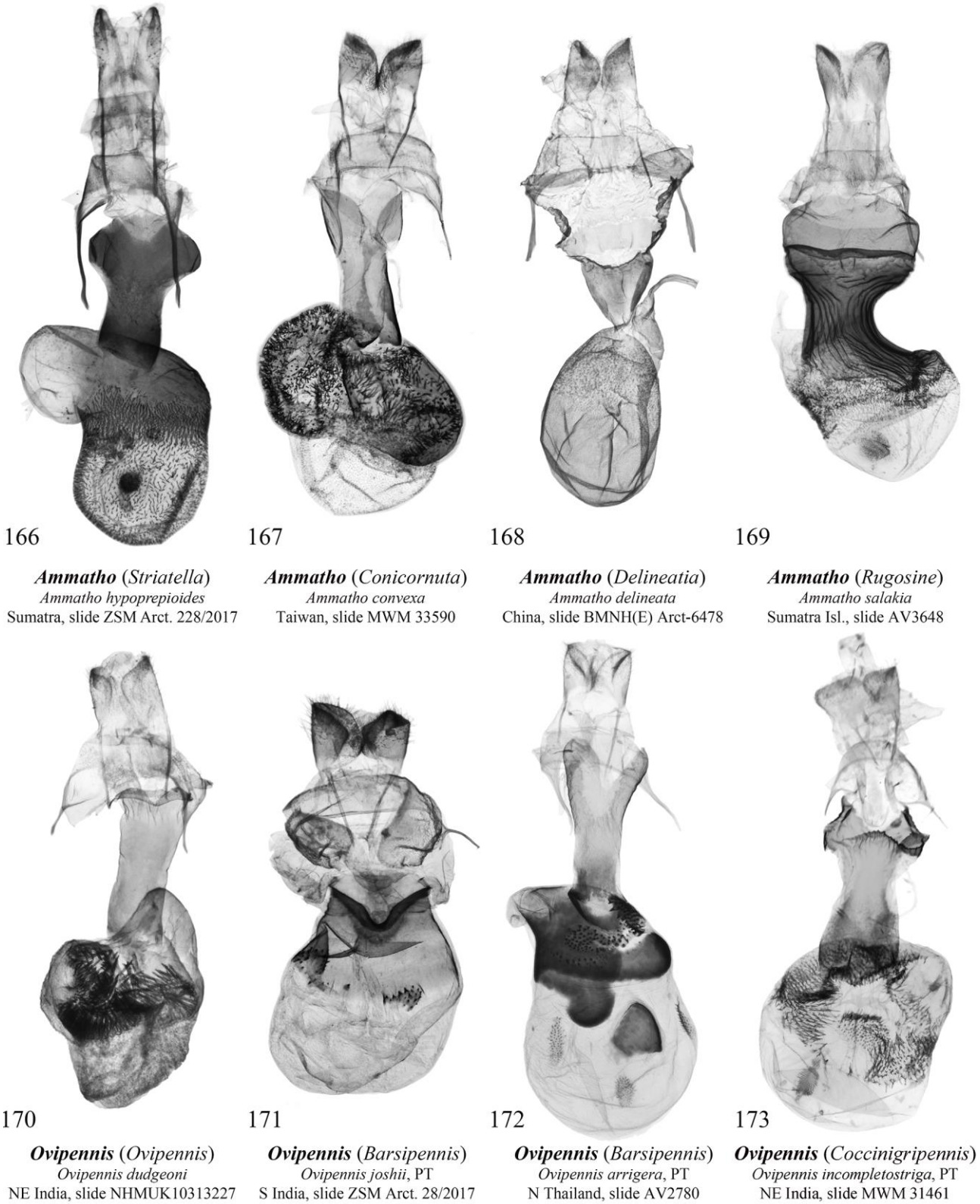
164

Ammatho (*Composine*)
Ammatho complicata
Malay Peninsula, slide MWM 33903

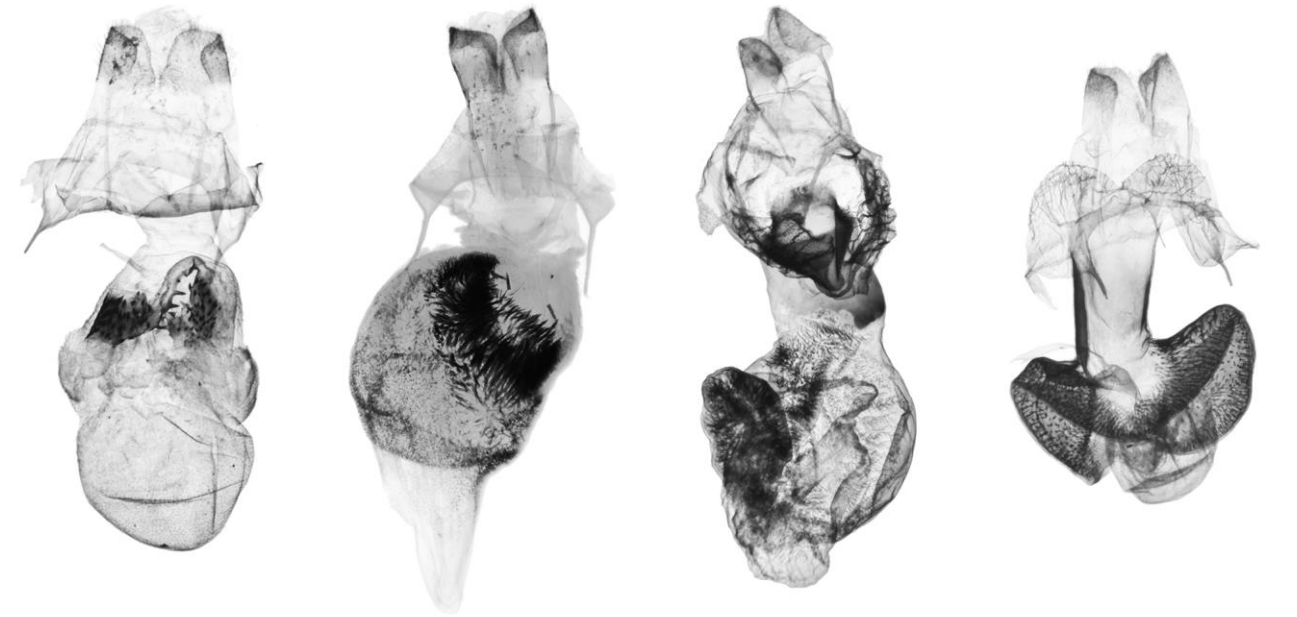
165

Ammatho (*Idopterum*)
Ammatho ovale
N Thailand, slide MWM 31450

Figures 160–165. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: female genitalia.



Figures 166–173. Members of the *Asura* / *Miltochrista* generic complex: female genitalia. 166–171, 173 – type species of genera and subgenera.

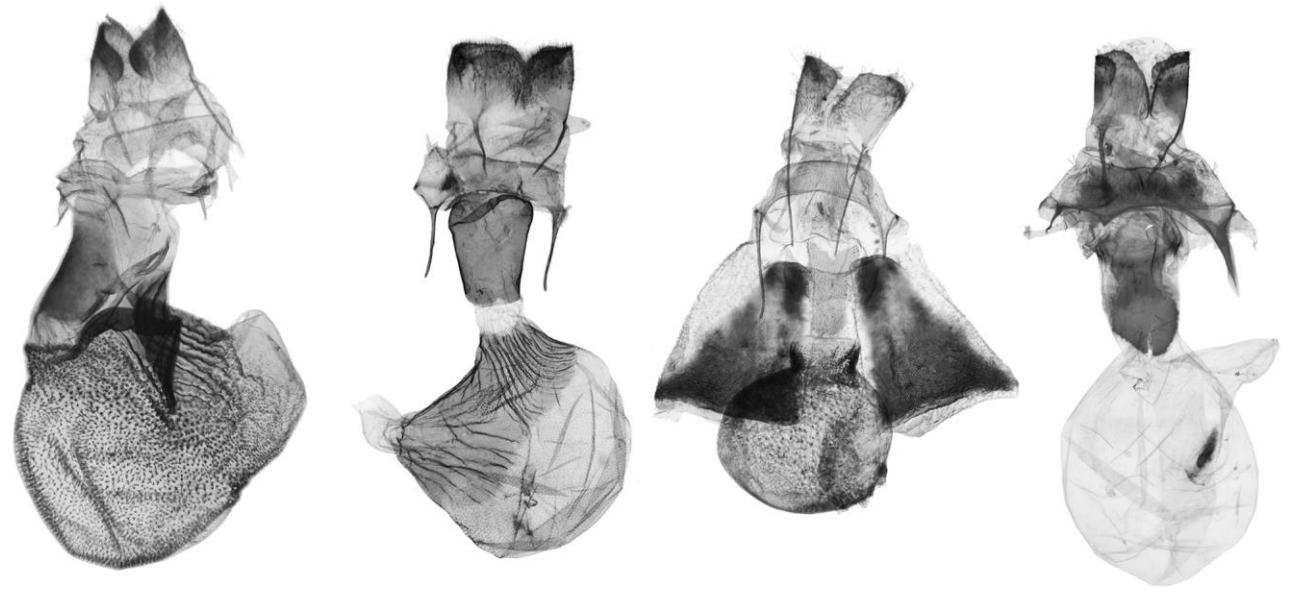


174 **Ovipennis** (*Nebulene*)
Ovipennis nebulosa
NE India, slide BMNH(E) Arct-6519

175 **Barsochrista**
Barsochrista kishidai
S India, slide MWM 31350

176 **Asuridia**
Asuridia carnipicta
China, slide NHMUK010313433

177 **Barsura** (*Barsura*)
Barsura nubifascia
NE India, slide BMNH(E) Arct-6502



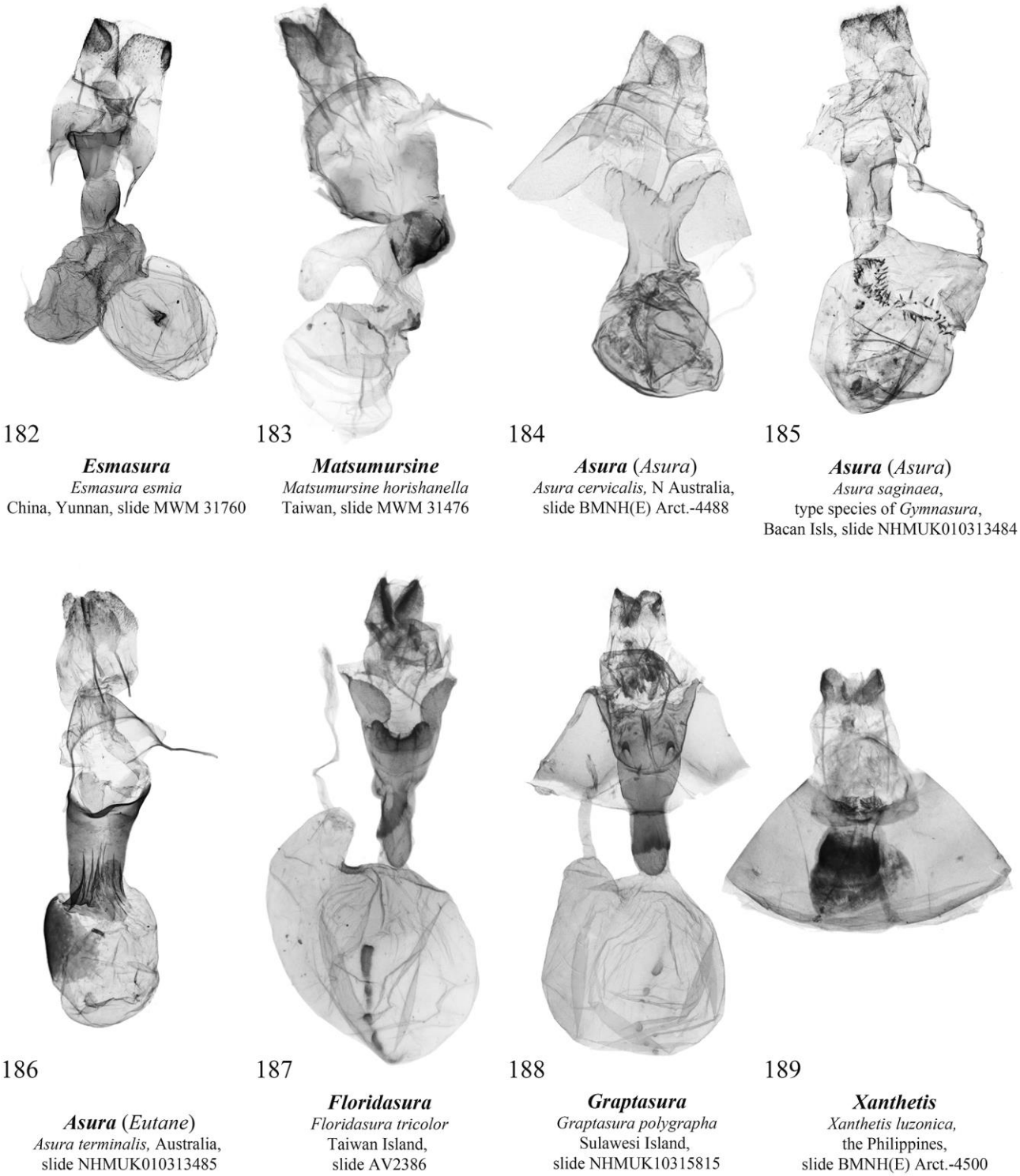
178 **Barsura** (*Tenebrasura*)
Barsura albidorsalis
Taiwan, slide AV3654

179 **Argentosine**
Argentosine proleuca
Nepal, slide MWM 33580

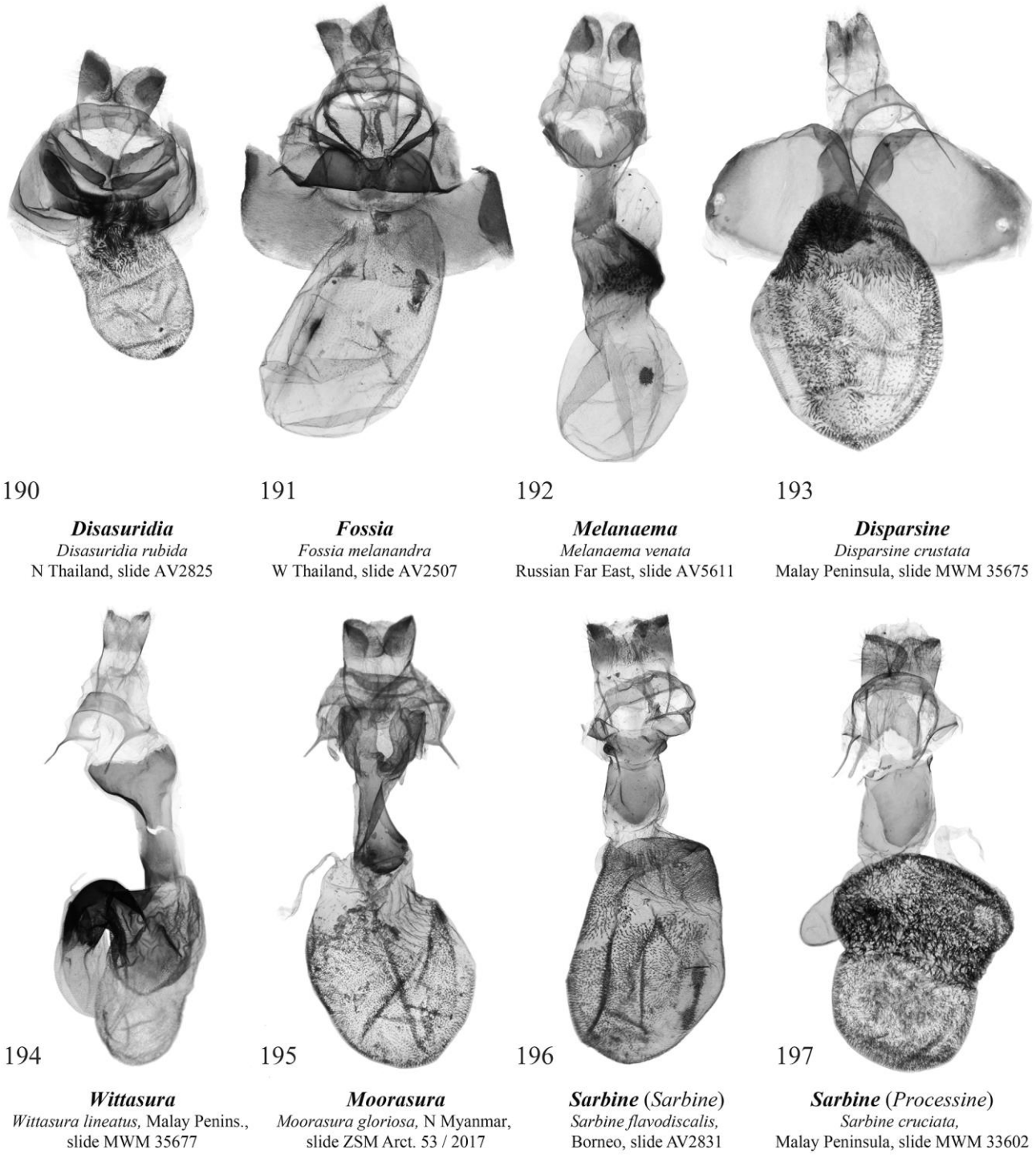
180 **Pseudobarsine**
Pseudobarsine bombdilensis
Nepal, slide ZSM Arct. 2019-643

181 **Nepita**
Nepita conferta
S India, slide MWM 34582

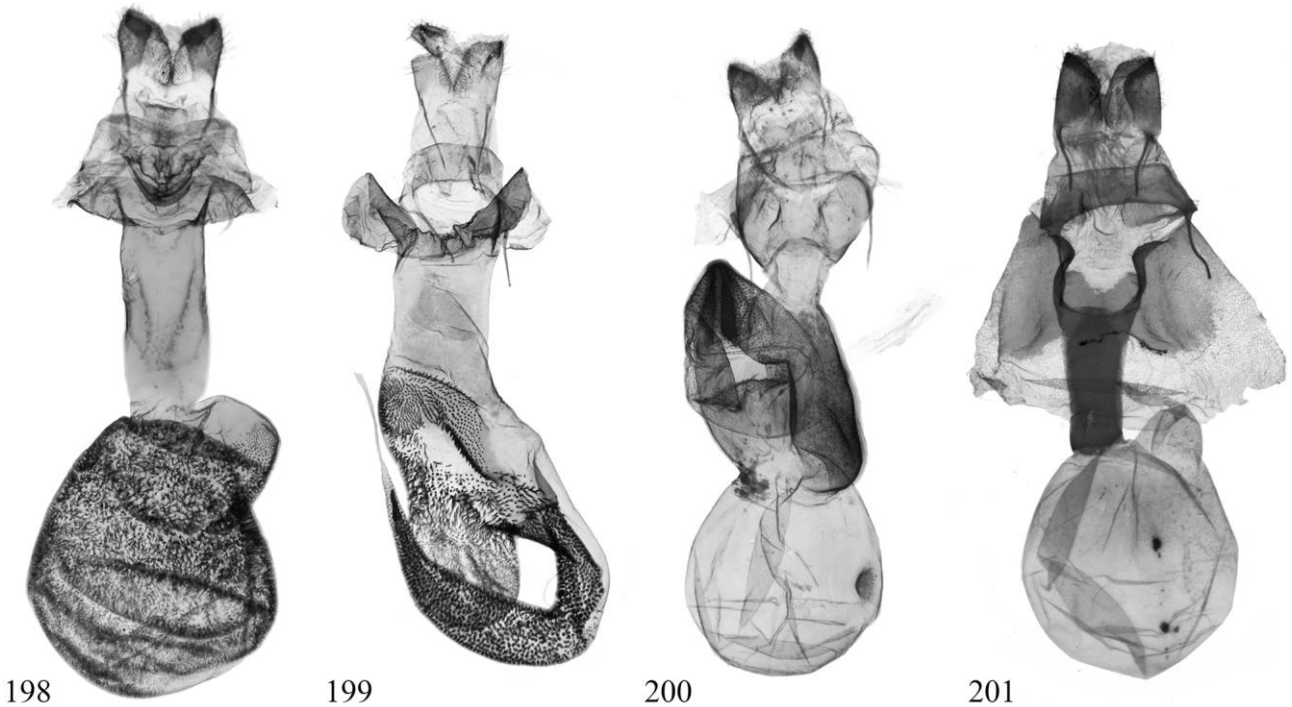
Figures 174–181. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: female genitalia.



Figures 182–189. Type species of genera, subgenera and their synonyms of the *Asura* / *Miltochrista* generic complex: female genitalia.



Figures 190–197. Type species of genera and subgenera of the *Asura* / *Miltochrista* generic complex: female genitalia.

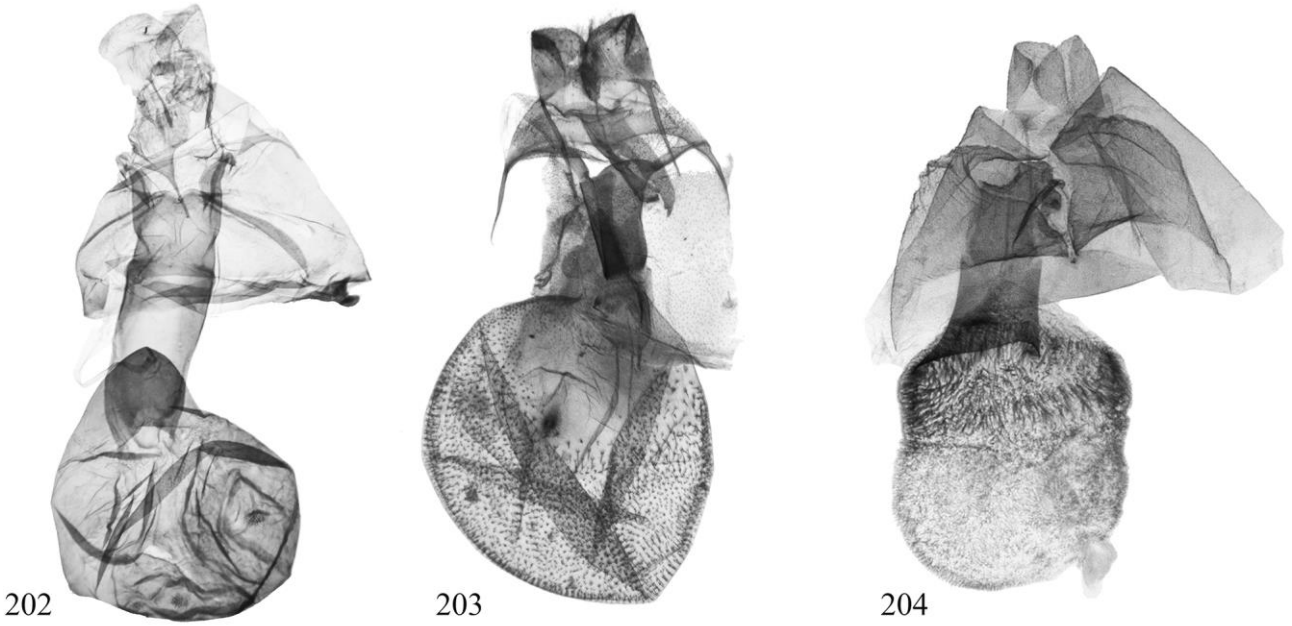


198 **Longarista**
Longarista longaria
SE China, slide AV4242

199 **Hampsonascia**
Hampsonascia dentifascia
NE India, slide NHMUK010313500

200 **Cernysura**
Cernysura orbis, PT
The Philippines, slide AV3739

201 **Barsilene**
Barsilene pallinflexa
Sumatra Isl., slide MWM 34582

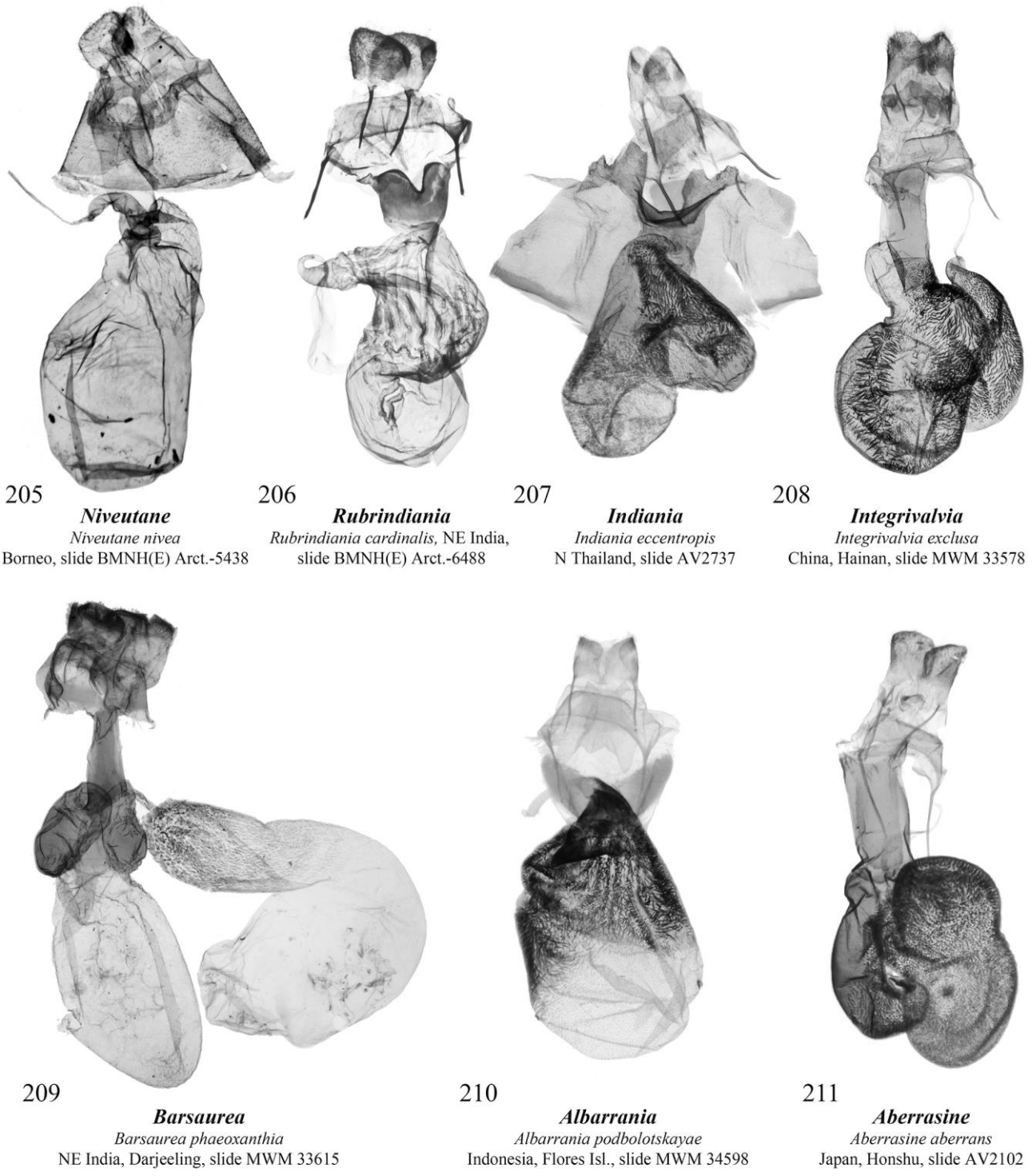


202 **Nanarsine**
Nanarsine porphyrea
Borneo, slide BMNH(E) Arct.-5275

203 **Amphisine**
Amphisine perpusilla
Indonesia, Sumatra Isl., slide AV3395

204 **Karolia**
Karolia stefaniae
Malay Peninsula, slide Mal054

Figures 198–204. Type species of genera of the *Asura* / *Miltochrista* generic complex: female genitalia.



Figures 205–211. Type species of genera of the *Asura* / *Miltochrista* generic complex: female genitalia.

Diagnosis. Small moths with reddish or brown forewing colouration and pattern similar to that of *Ammatho* s. str. In some species (*C. crucipuncta* and *C. orbis*) a slight sexual dimorphism is expressed as the presence of a white spot in male's forewing cell, which is as ground colour in females. Antennae of both sexes are

ciliate. The male genitalia of the genus *Cernysura* are characterised by the unique combination of the following features: (1) the juxta is broad, sclerotised, with a short medial crest; (2) the valva is relatively broad, with well-developed robust distal saccular process and distal lobe, similar to *Ammatho s. str.*, but (3) in *Cernysura* the costa is weakly sclerotised and has no medial and distal processes, like some groups of *Miltochrista*; (4) the aedeagus vesica with short but broad diverticula ornamented with robust granulation and bearing several robust thorn-like cornuti the number of which varies within species. The female genitalia are characterised by (1) the presence of a well-developed antrum; (2) the short and weakly sclerotised ductus bursae; (3) the heavily sclerotised narrow posterior section of corpus bursae; (4) the elliptical membranous anterior section of corpus bursae having a signum; and (5) the large, sack-like, sclerotised and granulated appendix bursae, the base of which occupies the significant part of the corpus bursae posterior section's left side.

Distribution. Endemic of the Philippines (Černý 1995).

Number of species. The genus comprises four valid species.

Genus ***Barsilene*** Volynkin & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:8F521564-F0B9-4F74-9609-7E57F5D152A9
(Figs 82, 83, 147, 201)

Type species: *Barsine pallinflexa* Holloway, 2001.

Etymology. The new genus name is a combination of the generic names *Barsine* and *Lyclene*. Gender feminine.

Diagnosis. Small moths with yellow or orange colouration and blackish pattern resembling that of *Moorasura*, *Sarbine*, *Integrivalvia* and *Albarrania*. Antennae of both sexes are ciliate. The male genital capsule of *Barsilene* is similar to that of *Sesapa* (illustrated by Volynkin 2017b; 2019b) by the short costa, very large elongated distal membranous lobe of valva, and the well-developed, heavily sclerotised distal saccular process, but, however, in *Barsilene* the distal saccular process is plate-like broadened basally and has a small ventral protrusion, the feature considered here to be autapomorphic. In addition, unlike *Sesapa*, the aedeagus vesica of *Barsilene* has elongated diverticula covered with numerous very small stout cornuti and robust granulation, whereas in *Sesapa* vesica has patches of robust spinules. The female genitalia are characterised by the combination of the following features: (1) the presence of an antrum with sclerotised elongated lateral processes and a deep and broad, more or less calyculate medial concavity, with margins being strongly connected to the 7th abdominal sternite; (2) the evenly sclerotised ductus bursae; (3) the membranous corpus bursae with only granulation; (4) the presence of two signa bursae; (5) the small and membranous appendix bursae situated postero-laterally and directed posteriorly.

Distribution. The genus is widespread from northeastern India through Indochina to Sundaland.

Number of species. The genus comprises two valid species.

Genus ***Nanarsine*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:8BE31058-0884-489B-9EDF-D2BD3CF6D894
(Figs 84, 85, 148, 149, 202)

Type species: *Hypocrita porphyrea* Snellen, 1880.

Etymology. The generic name is a combination of Latin 'nanus' meaning 'dwarf' or 'very small' and the generic name *Barsine*. The name refers to the small size of its moths in comparison with species of *Barsine*, which members of *Nanarsine* previously were associated with. Gender feminine.

Diagnosis. Small moths with reddish, yellow of white colouration and various pattern. Antennae of both sexes are ciliate. The male genital capsule of *Nanarsine* is similar to that of some *Miltochrista* species due to its well-developed distal saccular process and the absence of a medial costal process, but aedeagus vesica is armed with patches of numerous small spinules or very small stout cornuti, the feature common for *Barsine*, *Ammatho*, *Asura* and other related genera. In some species anellus bears two patches of spinules. The distal membranous lobe is reduced, very small or absent. The main autapomorphic feature is the female antrum structure: in *Nanarsine* it has elongated sclerotised stick-like lateral plates, but is membranous medially; the

posterior margin of antrum is sclerotised, strongly connected to the 7th abdominal sternite. The ductus bursae is dorso-ventrally flattened, sclerotised. Corpus bursae membranous, with an area of weak sclerotisation around the signum bursae, or (in *N. postalba*, *N. semilutea* and *N. milani*) covered with spinulose scobination and has a sclerotised lateral plate. The appendix bursae is small and membranous, situated postero-laterally and directed posteriorly.

Distribution. The genus is widespread from Mainland China and Taiwan through Indochina and Sundaland to the Philippines.

Number of species. The genus comprises seven valid species. Three of them (*N. postalba*, *N. semilutea* and *N. milani*) form a separated group with female genitalia significantly different from those of *N. porphyrea* and related species. The status of this group is unclear and will be clarified later.

Genus ***Amphisine*** Volynkin, **gen. nov.**

https://zoobank.org/urn:lsid:zoobank.org:act:986E7804-8C6B-46D8-9106-17C1B4E7D523

(Figs 86, 87, 150, 203)

Type species: *Hypoprepia perpusilla* Walker, 1862.

Etymology. The generic name is a combination of the Greek prefix 'αμφί-' meaning 'intermediate' and the generic name *Barsine*. The name refers to the male genitalia features of the new genus being intermediate between *Barsine sensu lato* and *Miltochrista*. Gender feminine.

Diagnosis. Small moths with reddish or yellow colouration and dark grey or blackish pattern. Antennae of both sexes are ciliate. The male genital capsule resembles that of *Miltochrista* due to the absence of a medial costal process, but is characterised by the combination of the following features: (1) the costa is thickened medially; (2) the costa apex is elongated distally and forming a narrow distal process of valva; (3) the sacculus is narrow, fused with costa apically (in the *A. asaphes* group by a very narrow plate along the outer concave valva margin); (4) the distal saccular process is short, present as a small protrusion; (5) the aedeagus vesica with fields of granulation and (in the *A. asaphes* group) with a cluster of small trigonal cornuti. The female genitalia have the dorso-ventrally flattened and sclerotised ductus bursae, the corpus bursae being densely and evenly scobinate, and the appendix bursae being membranous, situated postero-laterally and directed posteriorly. In the *A. perpusilla* group a signum is present and the posterior end of corpus bursae is weakly sclerotised.

Distribution. Indochina, Sundaland and the Philippines.

Number of species. The genus comprises five valid species.

Genus ***Karolia*** Volynkin, **gen. nov.**

https://zoobank.org/urn:lsid:zoobank.org:act:3E9794A0-B3A6-46B2-9B66-8991167FFA13

(Figs 88, 89, 146, 204)

Type species: *Barsine stefaniae* Bucsek, 2012.

Etymology. The genus is dedicated to Mr Karol Bucsek (Bratislava, Slovakia), an author of its type species. Gender feminine.

Diagnosis. Small moths with yellow colouration and blackish pattern. Male abdomen with black hair-like scales distally, similar to many *Miltochrista*. Antennae of both sexes are weakly ciliate. The male genitalia are characterised by the combination of the following features: (1) the juxta is volumetric, as a large elongated cone with a strongly narrowed apex (an autapomorphic feature; only in *Ammatho* (*Ammathella*) there is a similar volumetric conical structure, but it is present as a medial process of juxta and not an entire juxta); (2) the costa is broadened distally and forms a broadly trigonal distal process; (3) the distal membranous lobe is well-developed; (4) the sacculus is weakly setose, its distal process is broadly trigonal with a concave outer margin; (5) the valva has a robust ampulla medio-subdistally, which basally is connected with the distal section of sacculus (an autapomorphic feature); (6) the aedeagus straight and narrow, with a robust but narrow, serrulate carinal process directed distally (robust carinal processes are known in *Matsumursine* and *Barsura* also, but in *Barsura* that is not separated, and in *Matsumursine* that is

directed ventrally, hook-like); (7) the vesica is broadened, has two short and broad diverticula, weakly granulated only (the similar weakly ornamented vesica is known from *Xanthetis* and *Disasuridia*). The female genitalia are characterised by (1) the ostium bursae with an asymmetrical and dentate margin strongly connected with the 7th abdominal sternite; (2) the dorso-ventrally flattened and heavily sclerotised ductus bursae; (3) the corpus bursae being densely scobinated with more robust spinules posteriorly; (4) the presence of a small membranous diverticulum of corpus bursae anteriorly (only in *Sarbine* (*Processine*) there is a diverticulum of corpus bursae, but that is situated postero-laterally and sclerotised); and (5) the short conical appendix bursae directed posteriorly.

Distribution. Malay Peninsula and Sumatra.

Number of species. The genus is monobasic.

Genus ***Niveutane*** Volynkin, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:B968CED4-E66F-4560-B9A1-667AA636E819
(Figs 90, 91, 151, 205)

Type species: *Eutane nivea* Hampson, 1905.

Etymology. The generic name is a combination of its type species' name *nivea* and the generic name *Eutane*, which the species of the new genus were previously associated with. Gender feminine.

Diagnosis. Small moths with short trigonal wings and white colouration; pattern is reduced and present as yellowish terminal line and cilia with black spots at anal forewing margin. In some species a black discal dot is present. The male genitalia are characterised by the combination of the following characters: (1) the vinculum is basally broad and heavily sclerotised, with a narrow rectangular tip (an autapomorphic feature); (2) the valva is strongly broadened basally and strongly narrowed distally, with ventral margin having a small trigonal process subapically; (3) costa is narrow, its distal half is strongly setose (an autapomorphic feature); (4) the sacculus is broad but short and is half the length of the valva; (5) the distal saccular process is present, small, finger-like; (5) aedeagus is narrowed distally and slightly curved medially; (6) the aedeagus vesica has several short granulated diverticula. In the female genitalia, (1) the sclerotised ductus bursae with an arcuate margin of ostium bursae; (2) the ductus bursae is elongated, pear-shaped, weakly sclerotised and scobinated posteriorly; (3) the appendix bursae is small, narrow, elongated, curved, situated ventro-posteriorly.

Distribution. Malay Peninsula and Borneo.

Number of species. The genus comprises four valid species.

Genus ***Rubrindiania*** Volynkin & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:02A9F785-A02B-4C30-97BB-20F96ED0CAC2
(Figs 92, 93, 152, 206)

Type species: *Miltochrista cardinalis* Hampson, 1900.

Etymology. The generic name is a combination of the Latin word 'ruber' meaning 'red' and the generic name of the new genus' closest relative *Indiania*. Gender feminine.

Diagnosis. Small moths with narrow and elongated wings with rounded apex. Forewing crimson, with a longitudinal black stripe broadened distally; hindwing blackish, semi-hyaline at base. Male is significantly smaller than female, with strongly ciliate antenna, whereas female antenna is weakly ciliate. The male genitalia are characterised by the combination of the following features: (1) the valvae are slightly asymmetrical: the right one is somewhat broader than the left one; (2) the valva is elongated, narrow basally and broadened distally, entire, with the costal margin being convex distally and having a small fold medially; (3) the medial costal process is very small, tubercle-like; (4) the distal costal process is thorn-like, situated at valva apex; (5) the sacculus is narrow, its distal process reduced to a small blunted protrusion; (6) a distal membranous lobe of valva is absent; (7) the aedeagus vesica is globular with several small diverticula, membranous, with a field of short stout cornuti (similar to that of *Ammatho*). The male genitalia of *Rubrindiania* are similar to those of *Indiania* due to the slightly asymmetrical valvae with elongated and

robust distal processes and the absence of a distal membranous lobe, but differ by the presence of a tubercle-like medial costal process, smaller distal costal processes, and the presence of small stout cornuti in aedeagus vesica (whereas in *Indiania* vesica bears patches of spinules or spine-like cornuti). The female genitalia of *Rubrindiania* are characterised by the combination of the following features: (1) the heavily sclerotised antrum with a trigonal medial concavity, connected with the 7th abdominal sternite by a weak membrane (whereas in *Indiania* an antrum is absent, but an antevaginal plate is present); (2) the broadened and strongly rugose posterior section of corpus bursae (in *Indiania* that is narrowed and weakly sclerotised on the left and strongly scobinated on the right); (3) the membranous anterior section of corpus bursae bearing a double rugose signum (in *Indiania* the anterior section of corpus bursae is evenly scobinated and has no signum); (4) the appendix bursae is narrow, conical, rugose basally (in *Indiania* that is large, hook-like curved distally, with broad fields of robust scobination).

Distribution. The genus is distributed in northeastern India and northern Indochina.

Number of species. The genus comprises one valid species subdivided into two subspecies.

Genus ***Indiania*** Kirti, Joshi & N. Singh, 2014
(Figs 94, 95, 153, 207)

Indiania Kirti, Joshi & N. Singh, *in* Kirti, N. Singh & Joshi, 2014, *Tinea* **23** (1): 42 (Type species: *Miltochrista eccentropis* Meyrick, 1894, by original designation).

Diagnosis. Small moths with white, yellow or orange coloration and black pattern similar to that of *Ammatho* (*Striatella*). The male genitalia are also characterised by the elongate, entire, asymmetrical valvae with robust thorn-like apical processes being distal costal processes, the absence of a medial costal process, the basally setose sacculus with short trigonal distal process, and the aedeagus vesica bearing fields of granulation and patches of spinules or thorn-like cornuti. The main autapomorphy in the male genitalia is strongly asymmetrical valvae apexes. The female genitalia are characterised by the presence of an antevaginal plate having a narrow and heavily sclerotised medial plate and broad and strongly dentate lateral lobes. In addition, the posterior section of corpus bursae is narrowed, weakly sclerotised on the left and strongly scobinated on the right; the anterior section of corpus bursae is evenly scobinated; the appendix bursae is large, broad, hook-like curved distally and bears fields of robust spinulose scobination.

Distribution. The genus is distributed in Nepal, northeastern India, Myanmar, southern China, and Indochina including Malay Peninsula.

Number of species. The genus comprises four valid species and one subspecies.

Genus ***Barsaurea*** Volynkin & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:5A0B46A9-893D-42BB-BAE0-E595E5EAFC33
(Figs 96, 97, 154, 155, 209)

Type species: *Miltochrista phaeoxanthia* Hampson, 1900.

Etymology. The generic name is a combination of the generic name *Barsine* and the Latin word ‘aureus’ meaning ‘golden’. The name refers to the golden forewing ground colour of species of the genus. Gender feminine.

Diagnosis. Small moths with narrow and elongated forewing, golden colouration and brown diffuse pattern. The male genitalia are characterised by the combination of the following features: (1) the massive, laterally flattened and setose uncus; (2) the presence of an androconial gland in inner membrane of vinculum (an autapomorphic feature); (3) the presence of a round lobe-like medial costal process (an autapomorphic feature); (4) the absence of a distal membranous lobe of valva; (5) the narrowed valva apex with one or two apical processes; (6) the short sacculus without a distal process; and (7) the aedeagus vesica having elongated diverticula bearing patches of robust spines. The female genitalia of *Barsaurea* are characterised by the combination of the following features: (1) the very broad, rectangular papillae anales; (2) the sclerotised, dorso-ventrally flattened and narrowed posteriorly ductus bursae; (3) the narrow posterior section of corpus bursae with a rounded sclerotised lateral protrusion; (4) the large elliptical membranous anterior

section of corpus bursae; (5) the small elliptical and heavily sclerotised appendix bursae having a huge sack-like bulla, which is setose basally and membranous distally, and connected with the appendix bursae by the narrow membranous tube (an autapomorphic feature).

Distribution. The genus is widespread from northeastern India through Indochina to Malay Peninsula and Sumatra Island, but absent from Borneo and other Greater Sunda Islands.

Number of species. The genus comprises two valid species.

Genus ***Integrivalvia*** Volynkin & Huang, **gen. nov.**

https://zoobank.org/urn:lsid:zoobank.org:act:AB544F1E-96AD-42FC-8955-63BCAF1C3DD4

(Figs 98, 99, 156, 208)

Type species: *Barsine exclusa* Butler, 1877.

Etymology. The generic name refers to its entire valvae without any costal and saccular processes. Gender feminine.

Diagnosis. Small moths with reddish or orange coloration and blackish or brown pattern. The male genital capsule is characterised by the entire, narrow, lobe-like valva without any processes (similar to that of the African genus *Tumicla* and the Indian *Parvuspina* only). The autapomorphic feature of the new genus is the presence of a very long and narrow granulated distal diverticulum of aedeagus vesica having a hook-like curved robust plate at base (unlike *Hampsonascia*, which has long and narrow diverticula also, but without plates at base). In addition, the aedeagus vesica has several short diverticula bearing fields of granulation and patches of spinules. The basal plate of vesica ejaculatorius is absent. The autapomorphic feature in the female genitalia is the presence of a membranous lateral semiglobular protrusion on the connection of the ductus bursae with the corpus bursae. In addition, the female genitalia of *Integrivalvia* are characterised by the combination of the following features: (1) the sclerotised, dorso-ventrally flattened ductus bursae without antrum; (2) the pear-shaped corpus bursae densely covered with small spinulose scobination and having a band-like cluster of large spinules medially and a broad field of large spinules laterally and at the base of appendix bursae; (3) the appendix bursae is directed posteriorly, elongated, conical, curved, densely covered with robust spinules.

Distribution. The genus is widespread from southern China (Hainan) through eastern Indochina and Malay Peninsula to Greater and Lesser Sunda Islands including the Philippine Palawan Island.

Number of species. The genus comprises its type species with one subspecies. In addition, herein we provisionally include to the genus seven New Guinean species and one subspecies treated by de Vos (2019) to be members of the genus *Asuridia* and, in addition, one New Guinean species previously included to the genus *Melanaema*. However, all those New Guinean taxa are different in their genitalia structure from the type species (e.g., the principally different vesica structure having no characteristic elongated diverticulum with a sclerotised plate at base) and their generic placement needs further clarification.

Genus ***Albarrania*** Bolotov, Spitsyn & Kondakov, 2019

(Figs 100, 101, 157, 210)

Albarrania Bolotov, Spitsyn & Kondakov, *in* Bolotov, Spitsyn, Kondakov & Tomilova, 2019, *Ecologica Montenegrina* **20**: 213 (Type species: *Barsine podbolotskayae* Spitsyn & Bolotov, 2018, by original designation).

Diagnosis. Small moths externally very similar to members of the genera *Integrivalvia* and *Sarbine*. The male genitalia of the genus are very characteristic and have the following three autapomorphic features: (1) the presence of a spine-like costal process directed dorsally; (2) the presence of a bunch of hair-like setae on costa medially; (3) the club-shaped valva apex densely covered with long hair-like setae. In addition, the male genitalia of *Albarrania* are characterised by the very short sacculus having no processes and the aedeagus vesica bearing granulated diverticula and two robust serrulate plate-like cornuti. The female genitalia are characterised by the combination of the following features: (1) the very broad but weakly sclerotised antrum; (2) the very short and weakly sclerotised ductus bursae; (3) the sack-like corpus bursae with the weakly scobinated anterior section; (4) the posterior section of corpus bursae with robust spinulose

scobination; (5) the presence of a postero-lateral heavily sclerotised trigonal process at base of appendix bursae (an autapomorphic feature); and (6) the short, trigonal, heavily sclerotised appendix bursae situated postero-ventrally (an autapomorphic feature).

Distribution. Endemic of Flores Island (Lesser Sunda Archipelago).

Number of species. The genus is monobasic.

Genus ***Aberrasine*** Volynkin & Huang, **gen. nov.**
https://zoobank.org/urn:lsid:zoobank.org:act:299CCF0E-887A-4D24-8BC2-93B3FF735B70
(Figs 102, 103, 158, 159, 211)

Type species: *Miltochrista aberrans* Butler, 1877.

Etymology. The new genus' name is a combination of its type species' name *aberrans* and the generic name *Barsine*. Gender feminine.

Diagnosis. Small moths with red or orange colouration and thin black 'miltochristoid' pattern. In earlier modern papers, members of the genus were considered to be belonging to the genera *Barsine* (Dubatolov *et al.* 2012; Hsu *et al.* 2018; Volynkin 2018a) and *Miltochrista* (Černý 2016) as it combines some features common for both, *Barsine s. l.* and *Miltochrista*. The male genitalia of the genus are characterised by the combination of the following features: (1) the presence of a strong medial dorsal protrusion of costa, which may be short and round or large and apically pointed as well (Fig. 159) (a similar protrusion is also known in the genus *Sarbine* only); (2) the presence of a robust distal costal process; (3) the absence of a medial costal process; (4) the presence of a distal membranous lobe of valva; (5) the setose dorsal surface of sacculus; (6) the large and heavily sclerotised distal saccular process, which may be unilobate or bilobate; (7) the sack-like aedeagus vesica having several short diverticula, fields of granulations and clusters of short and stout 'barsinoid' cornuti or robust spinules. The female genitalia of *Aberrasine* are most similar to those of *Wittasura* due to the scobinated corpus bursae and the appendix bursae being curved basally, directed anteriorly and having a heavily sclerotised basal section and a scobinated distal section, but differ by the absence of an antrum, which is well-developed in *Wittasura*.

Distribution. The genus is widespread in eastern Palaearctic including Russian Far East and Japan, and in northern and central Indochina.

Number of species. The genus comprises 14 valid species and one subspecies.

Check-list of valid taxa of the *Asura* / *Miltochrista* generic complex

Barsine Walker, 1854
– *andromeda* Volynkin, Černý & Huang, 2019
– *atypicobarsine* N. Singh, Kirti & Joshi, 2016
– *biformis* Volynkin & Černý, 2019
– *cacharensis* N. Singh, Kirti & Kaleka, 2016
– *callorufa* Wu, Fu & Chang, 2013
– *conicornutata* (Holloway, 1982)
– *cornutodefecta* N. Singh, Kirti & Kaleka, 2016
– *curtisi* (Butler, 1881), **comb. nov.**
– *dao* Volynkin, Černý & Huang, 2019
– *defecta defecta* Walker, 1854
= *magna* (Hampson, 1894)
= *indistincta* Moore, 1878
– *defecta rubella* Volynkin & Černý, 2017
– *delicia* Swinhoe, 1891
– *deliciosa* Volynkin & Černý, 2016
– *devikulensis* N. Singh & Kirti, 2016
– *epixantha* (Meyrick, 1894)
– *euprepia* (Hampson, 1900)

– *eurydice* Volynkin & Černý, 2019
– *excelsa* (Daniel, 1952), **comb. nov.**
– *flammealis* Moore, 1878
– *flavicollis* (Moore, 1878)
– *fossi* Volynkin & Černý, 2017
– *fuscozonata* (Inoue, 1980)
– *germana* (Rothschild, 1913)
– *gilveola* (Daniel, 1952)
– *gratiosa* (Guérin-Méneville, 1843)
– *gratissima gratissima* (de Joannis, 1930)
– *gratissima versicolor* Volynkin & Černý, 2017
– *guangxiensis* (Fang, 1991), **comb. nov.**
– *hausmanni* Volynkin & Černý, 2019
– *hemimelaena* (de Joannis, 1928), **comb. nov.**
– *hoenei* (Reich, 1937), **comb. nov.**
– *hololeuca* (Hampson, 1895)
– *kampoli* Černý, 2009
– *laszloi* Volynkin & Černý, 2019
– *linga linga* Moore, 1859
= *tripartita* Walker, 1864
– *linga spilosomoides* (Moore, 1878)
– *lucibilis* Swinhoe, 1892
– *mactans* Butler, 1877
– *marcelae* Černý, 2016
– *nigrovena* (Fang, 2000)
– *obsoleta* (Reich, 1937)
– *orientalis orientalis* (Daniel, 1951)
– *orientalis bigamica* Černý, 2009
– *pardalis* (Mell, 1922), **comb. nov.**
= *miranda* Kishida & Wang, 2017, **syn. nov.**
– *perlucidula* Bucsek, 2012
– *perpallida* (Hampson, 1900)
= *pallida* Moore, 1878, nec. Bremer, 1864
= *porthesioides* Rothschild, 1910
– *ponlai* Wu, Fu & Chang, 2013
– *pretiosa* Moore, 1879
– *pseudomactans* Volynkin & Černý, 2016
– *pseudoorientalis* N. Singh & Kirti, 2016
– *pulchra* (Butler, 1877)
= *pulcherrima* Staudinger, 1887
= *sapporensis* Matsumura, 1930
– *roseata roseata* (Walker, 1864)
– *roseata augasta* Kishida, 2017
– *rosistriata* (Holloway, 1976)
– *rubrata* (Reich, 1937), **comb. nov.**
= *melanovena* Fang, 1991
– *rubricostata* (Herrich-Schäffer, 1855)
= *rubrigutta* Walker, 1862
– *ruficollis* (Fang, 1991)
– *rufumdefecta* N. Singh & Kirti, 2016
– *sauteri* (Strand, 1917)
– *sieglindae sieglindae* Černý, 2016
– *sieglindae coloraria* Černý, 2016
– *kulingensis* (Daniel, 1952)
– *striata striata* (Bremer & Grey, 1852)

= *kurilensis* Bryk, 1942
= *lanceolata* Walker, 1856
= *quelparta* Okamota, 1924, **syn. nov.**
– *striata hachijoensis* (Inoue & Minami, 1963)
– *thomasi* Kaleka, 2003
– *ustrina* Černý, 2009
– *valvalis* Kaleka, 2003
– *vinhphucensis* Spitsyn, Kondakov, Tomilova, Pham & Bolotov, 2018
– *wangi* Volynkin, Huang, Dubatolov & Kishida, 2019
– *yuennanensis* (Daniel, 1952)

Cyme Felder, 1861
= *Pallene* Walker, 1854
= *Prinasura* Hampson, 1903
= *Miltasura* Roepke, 1946, **syn. nov.**
– *anaemica* (Hampson, 1911)
– *analogus* (Rothschild, 1913), **comb. nov.**
– *aroa* (Bethune-Baker, 1904)
= *fasciolata* Rothschild, 1913
– *asuroides* (Rothschild, 1913)
– *avernalis* (Butler, 1887), **comb. nov.**
= *bougainvillei* Rothschild, 1913
= *isabellina* Rothschild, 1913
= *floridensis* Rothschild, 1913
– *basitesselata* (Rothschild, 1913)
– *biagi* (Bethune-Baker, 1908)
=*coccineoflammeus* Rothschild, 1913
– *celebensis* (Roepke, 1946), **comb. nov.**
– *citrinopuncta* (Rothschild, 1913)
– *coccineotermen* (Rothschild, 1913)
– *crocota* (Hampson, 1900), **comb. nov.**
– *effasciata* (Felder, 1861), **comb. nov.**
= *cinnabarina* Pagenstecher, 1884
– *euprepioides euprepioides* (Walker, 1862), **comb. nov.**
= *inclusa* Snellen, 1877
– *euprepioides interserta* (Moore, 1878), **comb. nov.**
– *feminina* (Rothschild, 1913), **comb. nov.**
– *haemachroa* (Hampson, 1905), **comb. nov.**
– *insularis* (Rothschild, 1913), **comb. nov.**
– *metascota* (Hampson, 1905), **comb. nov.**
= *suffusa* Draudt, 1914
– *miltochristaemorpha* (Rothschild, 1913)
=*aureorosea* Rothschild, 1913
– *miltochristina* (Rothschild, 1913), **comb. nov.**
– *multidentata* (Hampson, 1900), **comb. nov.**
– *phryctopa* (Meyrick, 1889)
– *pyraula* (Meyrick, 1886)
– *pyrostrota* (Hampson, 1914), **comb. nov.**
– *pyrrhauloides* (Rothschild, 1913)
– *quadrilineata moluccensis* (van Eecke, 1929)
– *quadrilineata quadrilineata* (Pagenstecher, 1886)
= *melitaula* Meyrick, 1886
= *simulans* Butler, 1886
= *basitessellata* Rothschild, 1913
– *quadrifasciata* (Rothschild, 1913), **comb. nov.**

– *reticulata* (Felder, 1861)
= *placens* Walker, [1865]
= *cyclota* Meyrick, 1886
= *intrita* Swinhoe, 1892
– *septemmaculata* (Heylaerts, 1891), **comb. nov.**
= *agraphia* Hampson, 1900
– *serratilinea* (Turner, 1940)
– *sexualis* (Felder, 1864)
= *cancellata* Pagenstecher, 1900
= *effulgens* Pagenstecher, 1900
= *quadrifasciata* Rothschild, 1913 (p. 214), nec. Rothschild, 1913 (p. 210), **homonym nov.**
= *mylea* Rothschild, 1916
= *terminodenta* Hulstaert, 1924
– *structa* (Walker, 1854)
= *pyrrhopsamma* Hampson, 1903
– *suavis* (Pagenstecher, 1886)
– *triangularis* (Rothschild, 1936)
– *vepallida* (Holland, 1900), **comb. nov.**
– *vivida* (Walker, [1865]), **comb. nov.**
– *wandammenensae* (Joicey & Talbot, 1916)
– *xantherythra* (Hampson, 1900)

Ammatho Walker, 1855, **stat. nov.**
Ammatho Walker, 1855, **stat. nov.**
– *bornescripta* (Holloway, 2001), **comb. nov.**
– *carbonisata* (Černý, 1995), **comb. nov.**
– *celebesa* (Tams, 1935), **comb. nov.**
– *collivolans* (Butler, 1881), **comb. nov.**
– *cruenia* (Hampson, 1918), **comb. nov.**
– *cuneonotatus* Walker, 1855, **comb. rev.**
= *mindorana* (Semper, 1899)
= *conchyliata* (Draudt, 1914)
– *cuneorotatus* (N. Singh & Kirti), **comb. nov.**
– *dohertyi* (Rothschild, 1913), **comb. nov.**
– *duopunctata* (Semper, 1899), **comb. nov.**
– *flavoplagiata* (Rothschild, 1913), **comb. nov.**
– *roseororatus* Butler, 1877, **comb. rev.**
– *sanguitincta* (Hampson, 1900), **comb. nov.**
– *scripta* (Walker, 1864), **comb. nov.**
= *meander* Snellen, 1879
– *erythropoda* (Roepke, 1946), **comb. nov.**

Ammathella Volynkin, **subgen. nov.**
– *midzhan* (Volynkin, 2018), **comb. nov.**
– *garo* (Volynkin, 2018), **comb. nov.**

Composine Volynkin, **subgen. nov.**
– *complicata* (Butler, 1877), **comb. nov.**

Idopterum Hampson, 1894, **stat. nov.**
– *ovale* (Hampson, 1894), **comb. nov.**

Species *incertae sedis*
– *admirabilis* (Schaus, 1922), **comb. nov.**
– *novaepommeraniae* (Strand, 1922), **comb. nov.**

Striatella Volynkin & Huang, **subgen. nov.**
the *hypoprepioides* species-group
– *amaculata* (Volynkin & Černý, 2016), **comb. n.**
– *amoenissima* (Volynkin & Černý, 2019), **comb. n.**
– *asotoida* (Volynkin & Černý, 2018), **comb. n.**
– *cao* (Volynkin & Černý, 2019), **comb. n.**
– *chi* (Roepke, 1946), **comb. nov.**
– *conformis* (Fang, 1991), **comb. nov.**
– *dubatolovi* (Volynkin & Černý, 2019), **comb. n.**
– *eos* (Volynkin & Černý, 2019), **comb. n.**
– *euryphaessa* (Volynkin & Černý, 2019), **comb. n.**
– *fasciata* (Leech, 1899), **comb. nov.**
– *helenae* (Černý, 2016), **comb. n.**
– *hypoprepioides* (Walker, 1862), **comb. nov.**
– *ivanovamariae* (Volynkin, Černý & Huang, 2019), **comb. n.**
– *karenkensis karenkensis* (Matsumura, 1930), **comb. n.**
– *karenkensis wushipheri* (Volynkin & Černý, 2019), **comb. n.**
– *kishidai* (Volynkin & Černý, 2019), **comb. nov.**
– *kuatunensis* (Daniel, 1951), **comb. nov.**
– *maculifasciata* (Hampson, 1894), **comb. nov.**
– *melanovena* (Černý, 2016), **comb. n.**
– *mene* (Volynkin & Černý, 2019), **comb. n.**
– *mesomene* (Volynkin, Černý & Huang, 2019), **comb. n.**
– *multistriata* (Hampson, 1894), **comb. nov.**
– *nemea* (Volynkin & Černý, 2019), **comb. n.**
– *nigralba* (Hampson, 1894), **comb. nov.**
– *pandeia* (Volynkin & Černý, 2019), **comb. n.**
– *parameia* (Rothschild, 1913), **comb. n.**
– *paraprominens* (Huang & Wang, 2018), **comb. n.**
– *persephone* (Volynkin & Černý, 2018), **comb. n.**
– *prominens* (Moore, 1878), **comb. nov.**
= *delicata* Moore, 1878
– *pseudoradians* (Joshi, N. Singh & Volynkin, 2018), **comb. n.**
– *radians* (Moore, 1878), **comb. nov.**
– *rhipiptera* (Wileman & West, 1928), **comb. n.**
– *salamandra* (Černý, 2009), **comb. n.**
– *selene* (Volynkin & Černý, 2019), **comb. n.**
– *specialis* (Fang, 1991), **comb. nov.**
– *speideli* (Volynkin, Černý & Huang, 2019), **comb. n.**
– *syntypica* (Swinhoe, 1906), **comb. n.**
– *syntypicoida* (Volynkin & Černý, 2019), **comb. n.**
– *takamukui* (Matsumura, 1927), **comb. n.**
– *terminifusca* (Daniel, 1955), **comb. nov.**
– *visaya* (Volynkin & Černý, 2019), **comb. n.**
– *wernerthomasi* (Volynkin, 2018), **comb. n.**

the *zebrina* species-group
– *bucseki* (Volynkin & Černý, 2018), **comb. n.**
– *navneetsinghi* (Volynkin & Černý, 2018), **comb. n.**
– *pluma* (Černý, 2009), **comb. nov.**
– *subzebrina* (Volynkin & Černý, 2018), **comb. n.**
– *zebrina* (Moore, 1878), **comb. nov.**

the *marginis* species-group

– *marginis* (Fang, 1991), **comb. nov.**

the *mesortha* species-group
– *callida* (Fang, 1991), **comb. nov.**
– *mesortha* (Hampson, 1897), **comb. nov.**

the *compar* species-group
– *compar* (Fang, 1991), **comb. nov.**

the *umbrosa* species-group
– *umbrosa* (Hampson, 1896), **comb. nov.**

the *pseudocardinalis* species-group
– *pseudocardinalis* (Volynkin & Černý, 2017), **comb. n.**

the *longstriga* species-group
– *longstriga* (Fang, 1991), **comb. nov.**

the *cernyi* species-group
– *cernyi* (Volynkin, 2018), **comb. nov.**

the *honbaensis* species-group
– *honbaensis* (Dubatolov & Bucsek, 2013), **comb. nov.**

the *dentata* species-group
– *dentata* (Wileman, 1910), **comb. nov.**

Conicornuta Volynkin, **subgen. nov.**
– *convexa* (Wileman, 1910), **comb. nov.**
= *cruciata* Matsumura, 1927

Delineatia Volynkin & Huang, **subgen. nov.**
– *delineata* (Walker, 1854), **comb. nov.**
= *figuratus* Walker, 1855
? = *rhodina* Herrich-Schäffer, [1855]
= *chinensis* C. Felder & R. Felder, 1862
= *fuscescens* Butler, 1877
= *coalescens* Draudt, 1914
= *dimidiata* Fang, 1991
– *sublucana* (Volynkin & Černý, 2017), **comb. nov.**

Rugosine Volynkin, **subgen. nov.**
– *salakia* (Schaus, 1922), **comb. n.**

Ovipennis Hampson, 1900
Ovipennis Hampson, 1900
the *dudgeoni* species-group
– *dudgeoni* (Elwes, 1890)

the *bicolora* species-group
– *bicolora* Fang, 1986

the *binghami* species-group
– *binghami* Hampson, 1903
– *thomasi* Černý, 2009

the *connexa* species-group
– *connexa* (Wileman, 1910), **comb. nov.**
– *insolita* (Volynkin, Černý, Bayarsaikhan & Bae, 2019), **comb. nov.**

the *flavivenosa* species-group
– *flavivenosa* (Moore, 1878), **comb. nov.**
– *y-nigrum* (Dubatolov & Bucsek, 2013), **comb. nov.**

Barsipennis Volynkin, **subgen. nov.**
the *phaeodonta* species-group
– *joshii* (Volynkin & Černý, 2019), **comb. nov.**
– *meyi* (Volynkin & Černý, 2019), **comb. nov.**
– *phaeodonta* (Hampson, 1911), **comb. nov.**

the *arrigera* species-group
– *arrigera* (Volynkin & Černý, 2019), **comb. nov.**
– *mophi* (Volynkin & Černý2019), **comb. nov.**

Nebulene Volynkin & Černý, 2018, **stat. nov.**
– *nebulosa* (Moore, 1878), **comb. nov.**
– *monocornuta* (Volynkin & Černý, 2018), **comb. nov.**

Coccinigripennis Volynkin & Huang, **subgen. nov.**
the *anomala* species-group
– *anomala* (Elwes, 1890), **comb. nov.**

the *fansipana* species-group
– *fansipana* (Volynkin & Černý, 2017), **comb. nov.**

the *miloslavae* species-group
– *nangkwak* (Volynkin & Černý, 2017), **comb. nov.**
– *incompletostriga* (Volynkin & Černý, 2017), **comb. nov.**
– *rawanga* (Volynkin & Černý, 2017), **comb. nov.**
– *miloslavae* (Černý, 2016), **comb. nov.**

Barsochrista N. Singh & Kirti, 2016
– *kishidai* (Kirti & Gill, 2009)
– *ocellata* (Hampson, 1907)

Asuridia Hampson, 1900
– *brevistriata* Fang, 1986
– *carnipicta* (Butler, 1877)
– *inouei* Wu & Fu, 2013
– *jinpingica* Fang, 1986
– *kishidai* Wu & Fu, 2013
– *miltochristoides* Rothschild, 1913
– *nigriradiata* (Hampson, 1896)
– *obscura* Fang, 1986
– *rubrimargo* (Hampson, 1894), **comb. nov.**
– *rubripennis* Inoue, 1988
– *yuennanica* Daniel, 1951

Barsura Volynkin, Dubatolov & Kishida, 2017
Barsura Volynkin, Dubatolov & Kishida, 2017

the *nubifascia* species-group
– *autumnalis* Volynkin, 2017
– *clandestina* Volynkin, Dubatolov & Kishida, 2017
– *contrastata* Volynkin, Dubatolov & Kishida, 2017
– *lineata* Fang, 1993
= *disnubifascia* Fang, 2000, an unnecessary replacement name
– *moxiana* Volynkin, 2018
– *nubifascia* (Walker, [1865])
= *punctifascia* Walker, 1869
– *obscura* Volynkin, Dubatolov & Kishida, 2017
– *vernalis* Volynkin, 2017

the *simplicifascia* species-group
– *simplicifascia* (Elwes, 1890)
– *melanoleuca* (Hampson, 1894)
– *umbrifera* (Hampson, 1900)

Tenebrasura Volynkin, **subgen. nov.**
– *albidorsalis* (Wileman, 1914), **comb. nov.**

Argentosine Volynkin, **gen. nov.**
– *proleuca* (Hampson, 1900), **comb. nov.**

Pseudobarsine N. Singh & Kirti, 2016
– *bombdilensis* N. Singh & Kirti, 2016
– *nainitalensis* N. Singh & Kirti, 2016

Nepita Moore, [1860]
– *conferta* (Walker, 1854)
= *anila* Moore, [1860]
= *signata* Walker, 1864
= *aegrota* Butler, 1877
= *ochracea* Butler, 1877
= *limbata* Butler, 1877
= *conferta* var. *fusca* Hampson, 1891

Esmasura Volynkin & Huang, **gen. nov.**
– *esmia* (Swinhoe, 1894), **comb. nov.**
= *callinoma* Meyrick, 1894

Matsumursine Volynkin & Huang, **gen. nov.**
– *horishanella* (Matsumura, 1927), **comb. nov.**
– *speciosa* (Fang, 1993), **comb. nov.**

***Asura* Walker, 1854**
Asura Walker, 1854
= *Stonia* Walker, [1865]
= *Gymnasura* Hampson, 1900, **syn. nov.**
– *albigrisea* (Rothschild, 1913)
– *amabilis* Rothschild & Jordan, 1901
– *arenaria* Rothschild, 1913
– *bipars* Walker, [1865]
= *habrotis* Meyrick, 1886
– *bipartita* Rothschild, 1916
– *brunneofasciata* Bethune-Baker, 1904

– *catameces* Turner, 1940
– *cervicalis* Walker, 1854
= *obliterans* Draudt, 1914
– *chrypsilon* (Semper, 1899), **comb. nov.**
– *chrysomela* Hampson, 1905
= *reducta* Rothschild, 1913
– *coccinocosma* Turner, 1940
– *compsodes* Turner, 1940
– *confina* Hampson, 1900
= *conflua* Hampson, 1914
= *liparidia* Rothschild, 1913
– *costaesignata* (Gaede, 1925), **comb. nov.**
– *crocopepla* Turner, 1940
– *crocoptera* Turner, 1940
– *dentifera* Hampson, 1900
– *dentiferoides* (Rothschild, 1915), **comb. nov.**
– *eichorni* Rothschild, 1936
– *elongata* (Rothschild, 1913)
– *euproctis* Schaus, 1922
– *flavagraphia* van Eecke, 1929
– *flaveola* Bethune-Baker, 1904
=*sagittaria* Bethune-Baker, 1904
=*percurrens* Hampson, 1914
– *flavescens* (Rothschild, 1912)
=*asuroides* Rothschild, 1916
– *flavia* (Hampson, 1900), **comb. nov.**
= *homoea* Turner, 1940
– *flavida* (Butler, 1887)
– *fulguritis* Hampson, 1900
= *birivula* Hampson, 1900
– *geminata* (Pagenstecher, 1900)
– *hemixantha* Hampson, 1900
– *hieroglyphica* Rothschild, 1913
– *lutea* (Bethune-Baker, 1908)
= *mediofascia* (Rothschild, 1913)
– *lydia* (Donovan, 1805)
= *mediastina* Hübner, 1827
= *gaudens* Walker, 1854
= *pectinata* Wallengren, 1860
= *incompleta* Draudt, 1914
= *confluens* Draudt, 1914
– *manusi* Rothschild, 1916
– *marginatana* Strand, 1922
=*marginata* Rothschild, 1913, nec. Walker, [1865]
– *mimetica flagrans* Fletcher, 1957
– *mimetica mimetica* Rothschild, 1913
– *monospila* Turner, 1940
– *nigriciliata* Hampson, 1900
– *obscurodiscalis* Rothschild, 1936
– *ochreomaculata* Bethune-Baker, 1904
=*biplagiata* Rothschild, 1913
– *ocnerioides* Rothschild, 1913
– *octiger* van Eecke, 1929
– *pallida* (Rothschild, 1913), **comb. nov.**
– *pallidana* (Strand, 1922), **comb. nov.**

– *phaeobasis* (Hampson, 1900), **comb. nov.**
– *pilcheri* (Lucas, 1891)
– *polyspila* Turner, 1940
– *pseudojosiodes* Rothschild, 1913
– *pyropa* Tams, 1935
– *reversa dampierensis* Rothschild, 1916
– *reversa reversa* Rothschild, 1916
– *rhodina* (Rothschild & Jordan, 1905), **comb. nov.**
– *ruenca* (Swinhoe, 1892)
– *saginaea* (Turner, 1899), **comb. nov.**
= *limonis* Lucas, 1900
= *eldola* Swinhoe, 1901
– *semiorbis* Schaus, 1922
– *semivitrea* (Rothschild, 1913)
– *simillima* Rothschild, 1936
– *toxodes* Hampson, 1907, **comb. rev.**
– *trizonata* Rothschild, 1913
– *unicolora* Bethune-Baker, 1904
– *zebrina* (Hampson, 1914)

Eutane Walker, 1854, **stat. nov.**
– *aglaea* (Hampson, 1914), **comb. nov.**
– *terminalis* (Walker, 1854), **comb. nov.**
= *maculata* Butler, 1877
= *gradata* Lucas, 1890
– *triplagiata* (Pagenstecher, 1900), **comb. nov.**
– *trimochla* (Turner, 1940), **comb. nov.**

Floridasura Volynkin, **gen. nov.**
– *tricolor* (Wileman, 1910), **comb. nov.**
= *magica* Strand, 1917
= *coccinea* Moore, 1886, **syn. nov.**

Graptasura Hampson, 1900
– *mesilau* Holloway, 2001
– *polygrapha* (Felder, 1874)
– *snelleni snelleni* (Roepke, 1943)
– *snelleni duplicata* (Nieuwenhuis, 1948)
– *trilacunata* Holloway, 2001
– *witti* Bucsek, 2012

Xanthetis Hampson, 1900
– *luzonica* (Felder, 1875)
= *naringa* Swinhoe, 1892
– *obiensis* Rothschild, 1913

Disasuridia Fang, 1991
– *conferta* Fang, 1991
– *confusa* Fang, 1991
– *fangae* Kirti, Joshi & N. Singh, 2013
– *metaphaea* (Hampson, 1900), **comb. nov.**
= *flava* Fang, 1991, **syn. nov.**
– *nakaoi* Černý, 2016
– *rubida* Fang, 1991

– *virgula* Fang, 1991

Quadrasura Holloway, 2001
– *ktimuna* (van Eecke, 1920)

Fossia Volynkin, Ivanova & Huang, **gen. nov.**
– *bachma bachma* (Volynkin & Černý, 2018), **comb. nov.**
– *bachma bolovena* (Volynkin & Černý, 2018), **comb. nov.**
– *elongata* (Černý, 2016), **comb. nov.**
– *melanandra* (Černý, 2009), **comb. nov.**
– *punicea punicea* (Moore, 1878), **comb. nov.**
= *postnigra* Hampson, 1894
– *punicea kachina* (Volynkin & Černý, 2018), **comb. nov.**
– *punicea rothschildi* (Draudt, 1914), **comb. nov.**
= *coccinea* Rothschild, 1913, nec Moore, 1886
= *rothschildi* Hulstaert, 1924
– *sirikitae* (Volynkin & Černý, 2018), **comb. nov.**

Melanaema Butler, 1877
– *venata* Butler, 1877
= *venata kyushuensis* Inoue, 1982
= *venata shikokuensis* Inoue, 1982

Wittasura Volynkin, **gen. nov.**
– *danieli* (Arora, 1983), **comb. nov.**
– *lineatus lineatus* (Walker, 1855), **comb. nov.**
– *lineatus calligenioides* (Snellen, 1879), **comb. nov.**
= *plumbilineata* Hampson, 1900
= *lineata* Walker, 1864, nec. Walker, 1855
– *trifasciata* (Roepke, 1946), **comb. nov.**

Disparsine Volynkin, **gen. nov.**
– *crustata* (Talbot, 1926), **comb. nov.**
– *nigrocincta* (Snellen, 1879), **comb. nov.**

Moorasura Volynkin & Huang, **gen. nov.**
– *inflexa* (Moore, 1878), **comb. nov.**
– *victoria* (Volynkin & Černý, 2019), **comb. nov.**
– *kanchenjunga* (Volynkin & Černý, 2019), **comb. nov.**
– *dejeani* (Volynkin & Černý, 2019), **comb. nov.**
– *gloriosa* (Moore, 1878), **comb. nov.**
– *thagyamin* (Volynkin & Černý, 2019), **comb. nov.**

Sarbine Volynkin, **gen. nov.**
Sarbine Volynkin, **subgen. nov.**
– *hreblayi* (Volynkin & Černý, 2019), **comb. nov.**
– *flavodiscalis* (Talbot, 1926), **comb. nov.**

Processine Volynkin, **subgen. nov.**
– *cruciata* (Walker, 1862), **comb. nov.**
– *siberuta* (Volynkin & Černý, 2019), **comb. nov.**

Longarista Volynkin, 2019
– *longaria* (Daniel, 1951)
– *kareli* Volynkin, 2019

Hampsonascia Volynkin, **gen. nov.**
– *dentifascia* (Hampson, 1894), **comb. nov.**

Cernysura Volynkin, **gen. nov.**
– *crucipuncta* (Černý, 1995), **comb. nov.**
– *lenticulata* (Černý, 1995), **comb. nov.**
– *obscura* (Semper, 1899), **comb. nov.**
– *orbis* (Černý, 1995), **comb. nov.**

Barsilene Volynkin & Huang, **gen. nov.**
– *melaninflexa* (Černý, 2016), **comb. nov.**
– *pallinflexa* (Holloway, 2001), **comb. nov.**

Nanarsine Volynkin, **gen. nov.**
the *porphyrea* species-group
– *porphyrea* (Snellen, 1880), **comb. nov.**
= *subcruciata* Rothschild, 1913
– *sullia* (Swinhoe, 1901), **comb. nov.**

the *senara* species-group
– *emiks* (Černý, 2009), **comb. nov.**
– *senara* (Moore, 1859), **comb. nov.**

the *semilutea* species-group
– *milani* (Černý, 2009), **comb. nov.**
– *postalba* (Fang, 1986), **comb. nov.**
– *semilutea* (Wileman, 1911), **comb. nov.**

Amphisine Volynkin, **gen. nov.**
the *perpusilla* species-group
– *perpusilla* (Walker, 1862), **comb. nov.**
= *clavula* van Eecke, 1920
– *lutivittata* (Wileman & West, 1928), **comb. nov.**

the *asaphes* species-group
– *asaphes* (Hampson, 1900), **comb. nov.**
– *latigrapha* (Černý, 2009), **comb. nov.**
– *cursiva* (Černý, 2009), **comb. nov.**

Karolia Volynkin, **gen. nov.**
– *stefaniae* (Bucsek, 2012), **comb. nov.**

Niveutane Volynkin, **gen. nov.**
– *alba* (Hampson, 1900), **comb. nov.**
– *margarita* (Bucsek, 2012), **comb. nov.**
– *nivea* (Hampson, 1905), **comb. nov.**
– *virginalis* (Bucsek, 2012), **comb. nov.**

Rubrindiania Volynkin & Huang, **gen. nov.**
– *cardinalis cardinalis* (Hampson, 1900), **comb. nov.**
= *erubescens* Rothschild, 1936
– *cardinalis gemina* (Volynkin & Černý, 2017), **comb. nov.**

Indiania Kirti, Joshi & N. Singh, 2014

– *arcana* (Bucsek, 2012)
– *auriflucta auriflucta* (Černý, 2009)
– *auriflucta deaurata* (Černý, 2009)
– *citrona* (Hampson, 1907), **comb. nov.**
– *eccentropis* (Meyrick, 1894)

Barsaurea Volynkin & Huang, **gen. nov.**
– *phaeoxanthia* (Hampson, 1900), **comb. nov.**
– *diehli* (Dubatolov & Bucsek, 2014), **comb. nov.**

Integrivalvia Volynkin & Huang, **gen. nov.**
– *exclusa exclusa* (Butler, 1877), **comb. nov.**
– *exclusa trivittata* (Moore, 1877), **comb. nov.**

Species *incertae sedis*
– *decussa* (Bethune-Baker, 1910), **comb. nov.**
– *dinawa* (Bethune-Baker, 1904), **comb. nov.**
– *intensa* (Rothschild, 1913), **comb. nov.**
– *nigrisparsa* (Hampson, 1914), **comb. nov.**
– *phoenicea novaguinensis* (van Eecke, 1924), **comb. nov.**
– *phoenicea phoenicea* (Hampson, 1914), **comb. nov.**
– *rosacea* (Bethune-Baker, 1904), **comb. nov.**
– *rutila* (Walker, [1865]), **comb. nov.**
– *sanguinea* (Hampson, 1900), **comb. nov.**

Albarrania Bolotov, Spitsyn & Kondakov, 2019
– *podbolotskayae* (Spitsyn & Bolotov, 2018)

Aberrasine Volynkin & Huang, **gen. nov.**
– *aberrans aberrans* (Butler, 1877), **comb. nov.**
= *askoldensis* Oberthür, 1880
= *bivittata* Butler, 1885
= *decussata* (Moore, 1877), **syn. nov.**
– *aberrans okinawana* (Matsumura, 1930), **comb. nov.**
– *atuntseensis* (Daniel, 1951), **comb. nov.**
– *collina* (Černý, 2016), **comb. nov.**
– *dentata* (Wileman, 1910), **comb. nov.**
– *dingjiai* (Hsu, M.-Y. Chen & Buchsbaum, 2018), **comb. nov.**
– *inaequidens* (de Joannis, 1928), **comb. nov.**
– *expressa* (Inoue, 1988), **comb. nov.**
– *marginis* (Fang, 1991), **comb. nov.**
– *nigrociliata* (Fang, 1991), **comb. nov.**
– *peraffinis* (Fang, 1991), **comb. nov.**
– *separans* (de Joannis, 1928), **comb. nov.**
– *sinuata* (Fang, 1991), **comb. nov.**
– *strigivenata* (Hampson, 1894), **comb. nov.**
– *variata* (Daniel, 1951), **comb. nov.**

Arctelene N. Singh, Kirti & Gill, 2008
– *neouncodes* N. Singh & Kirti, 2016
– *patnitopensis* N. Singh & Kirti, 2016
– *rufescens* N. Singh Kirti & Gill, 2008
– *uncodes* N. Singh Kirti & Gill, 2008
– *varians* (Hampson, 1891), **comb. nov.**

Parvuspina N. Singh, Kirti & Datta, 2019
– *tawaghatensis* N. Singh, Kirti & Datta, 2019

Sesapa Walker, 1854
Sesapa Walker, 1854
– *inscripta* Walker, 1854
= *erubescens* Butler, 1877
– *sanguinea* (Moore, 1877)
– *strandiana* Volynkin, 2019
– *taiflamma* Wu, 2019

Nipponasura Inoue, 1965
– *inouei* Volynkin, 2017
= *sanguinea* Inoue, 1965, nec. Moore, 1877
– *kishidai* Wu, 2019

Species *incertae sedis*
– *koshunica* (Strand, 1917)

Chrysasura Hampson, 1914
– *flavopunctata* (Bethune-Baker, 1904)
= *punctatissima* Rothschild, 1913
– *leopardina* (Rothschild, 1913)
– *literata* Wileman & South, 1919
– *meeki meeki* Rothschild, 1915
– *meeki wollastoni* Rothschild, 1915
– *metahyala* (Hampson, 1918), **comb. nov.**
– *pardalina* (Rothschild, 1936), **comb. nov.**
– *postvitreata* (Rothschild, 1913)

Trichocerosia Hampson, 1900
– *atrifulva* Hampson, 1900
= *nigrorufa* Rothschild, 1913
– *truncata* Rothschild, 1913
– *variabilis* Rothschild, 1913
= *antemedialis* Strand, 1922
= *discoidalis* Strand, 1922

Chiretolpis Watson, 1980
= *Tricholepis* Hampson, 1891, nec. Blanchard, [1851]
– *erubescens* (Hampson, 1891)

Species *incertae sedis*
– *bicolorata* (Pagenstecher, 1900)
– *elongata* (Rothschild & Jordan, 1901)
– *melanoxantha* (Hampson, 1911)
– *ochracea* (Rothschild & Jordan, 1901)
= *postdivisa* (Rothschild, 1916)
– *rhodia* (Rothschild & Jordan, 1901)
– *signata* (Rothschild & Jordan, 1901)
– *sinapis* (Rothschild, 1913)
– *unicolor* (Rothschild & Jordan, 1901)
– *woodlarkiana* (Rothschild & Jordan, 1901)
– *xanthomelas* (Hampson, 1900)
= *nigrita* Rothschild & Jordan, 1901

= *keensis* Strand, 1922

Micronyctemera de Vos & van Mastrigt, 2007
– *fojaensis* de Vos & van Mastrigt, 2007

Symmetrodes Meyrick, 1886
= *Habrochroma* Turner, 1940
– *sciocosma* Meyrick, 1888
= *ectophaea* Hampson, 1900
= *atrifusa* Hampson, 1900
= *reducta* Draudt, 1914
= *reductana* (Strand, 1922)
– *platymelas* Turner, 1940

Miltochrista Hübner, [1819]
= *Calligenia* Duponchel, [1845]
= *Cyllene* Walker, 1854
= *Lyclene* Moore, [1860]
= *Setinochroa* Felder, 1874
= *Xanthocraspeda* Hampson, 1894
= *Neasura* Hampson, 1900
= *Asuropsis* Matsumura, 1927, **syn. nov.**
= *Neasuroides* Matsumura, 1927, **syn. nov.**
= *Asuridoides* Daniel, 1951, **syn. nov.**
= *Hypasura* Daniel, 1952
– *acteola* (Swinhoe, 1903), **comb. nov.**
– *acteolina* (Schaus, 1922), **comb. nov.**
– *acutiseriata* (Holloway, 2001), **comb. nov.**
– *alikangiae alikangiae* (Strand, 1917), **comb. nov.**
– *alikangiae intermedia* (Marumo, 1923), **comb. nov.**
– *andamana* (Moore, 1877)
– *angulifera* (Holloway, 2001), **comb. nov.**
– *angulinea* (Holloway, 2001), **comb. nov.**
– *apiseriata* (Holloway, 2001), **comb. nov.**
– *apuncta* Rothschild, 1915
– *aquila* (Černý, 2009)
– *arcuata* (Moore, 1882)
= *rosea* Hampson, 1891
= *aurora* Hampson, 1891
=. *formosicola* Strand, 1917
= *ochrostraminea* Strand, 1917
– *areolifera* (Holloway, 2001)
– *asakurai* (Matsumura, 1927), **comb. nov.**
– *ashleigera* (Holloway, 2001), **comb. nov.**
– *atritermina* (Hampson, 1900), **comb. nov.**
– *atuntseensis* Daniel, 1951
– *atuntseica* (Daniel, 1951), **comb. nov.**
– *aureata* (Rothschild, 1913), **comb. nov.**
– *auritiformis* (Černý, 2009), **comb. nov.**
– *biseriata* (Hampson, 1900), **comb. nov.**
– *brunneata* (Daniel, 1965), **comb. nov.**
– *buruana* (van Eecke, 1929), **comb. nov.**
– *calamaria* (Moore, 1888)
= *punctata* Elwes, 1890
= *celidopa* Meyrick, 1894

= *mediopuncta* Rothschild, 1913
– *calamina calamina* Butler, 1877
– *calamina lutea* Staudinger, 1887
– *calcicola* (Černý, 2016), **comb. nov.**
– *carnea* (Poujade, 1886)
– *cepheus* (Černý, 2009), **comb. nov.**
– *chromatica* (Swinhoe, 1891)
– *cingula* (Černý, 2009), **comb. nov.**
– *circumdata* (Walker, 1864), **comb. nov.**
– *clara* Daniel, 1951, **comb. rev.**
– *classeigera* (Holloway, 2001)
– *congerens* (Felder, 1874)
= *artocarpi* Moore, 1878
= *roseogrisea* Rothschild, 1913
– *conjunctana* (Walker, 1866)
= *tessellata* Butler, 1881
= *eschara* Swinhoe, 1894
– *cornutochrista* N. Singh & Kirti, 2016
– *corrigera* (Volynkin & Bucsek, 2016), **comb. nov.**
= *undulata* Bucsek, 2012, nec. Swinhoe, 1903
– *creatina creatina* (Snellen, 1879), **comb. nov.**
– *creatina javanica* (van Eecke, 1920), **comb. nov.**
– *cruenia* Hampson, 1918
– *cumseriata* (Bucsek, 2012), **comb. nov.**
– *cuneifera* (Walker, 1862)
– *cuneigera* (Walker, 1862), **comb. nov.**
= *lutaroides* van Eecke, 1926
– *cylletona* (Swinhoe, 1893), **comb. nov.**
= *geodetis* Meyrick, 1894
– *dagmarae* (Černý, 2016), **comb. nov.**
– *dasara* (Moore, 1859)
– *delia* (Schaus, 1922), **comb. nov.**
– *dentinebula* (Černý, 2009), **comb. nov.**
– *dharma butleri* (Leech, [1889])
– *dharma dharma* (Moore, 1879)
– *diluta* (Draeseke, 1926), **comb. nov.**
– *discisigna* (Moore, 1878)
= *aurantiaca* Moore, 1878
= *fuscifusa* Hampson, 1894
= *fuscifera* Swinhoe, 1894
= *rufotincta* Rothschild, 1913
= *depuncta* Draudt, 1914
– *discistriga* (Moore, 1878), **comb. nov.**
– *distributa* (Walker, 1862), **comb. nov.**
– *ecmelaena* (Hampson, 1900), **comb. nov.**
– *enormitata* (Bucsek, 2014), **comb. nov.**
– *eos* (Hampson, 1900), **comb. nov.**
– *evae* (Černý, 2016), **comb. nov.**
– *excaviseriata* (Holloway, 2001), **comb. nov.**
– *falcihumilis* N. Singh & Kirti, 2016
– *falciseriata* (Holloway, 2001), **comb. nov.**
– *fascicornuta* Volynkin, 2017
– *flexuosa* Leech, 1899
– *floccosa* (Walker, 1864), **comb. nov.**
= *subcervina* Walker, 1864

– *fruhstorferi* (Aurivillius, 1894), **comb. nov.**
= *scripta* Heylaerts, 1891
– *fulvimarginata* (Hampson, 1909), **comb. nov.**
– *furcata* (Reich, 1936), **comb. nov.**
– *fuscalis* (Hampson, 1891), **comb. nov.**
– *fusciramorum* (Holloway, 2001), **comb. nov.**
– *gilva* Daniel, 1951, **comb. rev.**
– *goaensis* (Kirti & Gill, 2009)
– *grandigilva* Fang, 2000
– *griseata* (Leech, 1899), **comb. nov.**
– *griseirufa* Fang, 1991
– *gyochiana* (Matsumura, 1927), **comb. nov.**
– *hoenei* (Daniel, 1952), **comb. nov.**
– *hollowai* (Kirti & Gill, 2009)
– *humilis* (Walker, 1859)
– *hypophaeola* (Hampson, 1900), **comb. nov.**
– *ila* (Moore, [1860])
= *unifascia* Rothschild, 1913
– *inconspicua* (Moore, 1878), **comb. nov.**
= *obliqua* Hampson, 1891
– *indentata* (Schaus, 1922), **comb. nov.**
– *infumata* (Felder, 1874), **comb. nov.**
= *pallida* Moore, 1878 (*Setinochroa*), nec. Bremer, 1864
= *postica* Moore, 1878
– *inornata* (Wileman & South, 1919), **comb. nov.**
– *irregularis* Rothschild, 1913
– *jaroslavae* (Černý, 2016), **comb. nov.**
– *jeremyhollowayi* (Bucsek, 2014), **comb. nov.**
– *jucunda* Fang, 1991
– *kepica* (Dubatolov & Bucsek, 2013), **comb. nov.**
– *kontumica* (Dubatolov & Bucsek, 2013)
– *kosterini* (Dubatolov & Bucsek, 2013), **comb. nov.**
– *labyrintha* (Bucsek, 2012), **comb. nov.**
– *lateritia* (Černý, 2009), **comb. nov.**
– *latimargo* (Roepke, 1946), **comb. nov.**
– *lignea* (Černý, 1995), **comb. nov.**
– *likiangensis* (Daniel, 1952), **comb. nov.**
– *lineidistincta* (Bucsek, 2012), **comb. nov.**
– *lutara* (Moore, 1859)
= *dividata* Snellen, 1880
– *lutarella* (Kalis, 1934), **comb. nov.**
– *luzonica* (Wileman & South, 1919), **comb. nov.**
– *lyclenoides* Černý, 2016
– *malayproducta* (Bucsek, 2012), **comb. nov.**
– *marginata* (Walker, [1865]), **comb. nov.**
– *margita* (van Eecke, 1926), **comb. nov.**
– *mediobliqua* (Wu, Fu & Chang, 2013), **comb. nov.**
– *megala* (Hampson, 1900), **comb. nov.**
– *melanopyga* (Hampson, 1918), **comb. nov.**
– *mentiens* (Fang, 1993), **comb. nov.**
– *mesilaulinea* (Holloway, 2001), **comb. nov.**
– *miniata miniata* (J.R.Forster, 1771)
= *rubicunda* [Denis & Schiffermüller], 1775
= *rosea* Fabricius, 1775
= *rosea* Donovan, 1793

= *roseana* de Villers, 1789
= *virginea* Delahayne, 1896
= *cinfluens* Lambillion, 1906
= *philippsi* Costantinio, 1922
= *destrigata* Dannehl, 1928
= *nigricirris* Lucas, 1959
= *mosbacheri* Roesler, 1967

– *miniata rosaria* Butler, 1877
– *minibunda* (Černý, 2009), **comb. nov.**
– *modesta* (Leech, 1899)
– *modvena* (Schaus, 1925), **comb. nov.**
– *multiramorum* (Holloway, 2001), **comb. nov.**
– *nataliae* (Černý, 2016), **comb. nov.**
– *nebulosa* (Moore, 1878)
– *neocuneifera* N. Singh & Kirti, 2016
– *neoseriata* N. Singh & Kirti, 2016
– *ni* (Heylaerts, 1891), **comb. nov.**
– *nigrilineata* (Fang, 2000), **comb. nov.**
– *nigrivena* Leech, 1899, **comb. rev.**
– *nigroanalis* (Matsumura, 1927), **comb. nov.**
– *nubilalis* (Hampson, 1894)
– *obliquilinea* (Swinhoe, 1901)
– *obscurilinea* (Holloway, 2001), **comb. nov.**
– *obscuripostica* Dubatolov, Kishida & Wang, 2012
– *obsoleta* (Moore, 1878)
= *assamica* Moore, 1878
– *obsoletaria* (Bucsek, 2014), **comb. nov.**
– *obtusilinea* (Holloway, 2001), **comb. nov.**
– *orsova* (Swinhoe, 1903), **comb. nov.**
– *osthelderi* (Daniel, 1951), **comb. nov.**
– *pallida pallida* (Bremer, 1864)
– *pallida formosana* Daniel, 1951
– *pallida tapaishanica* Daniel, 1951
– *paraarcuata* N. Singh & Kirti, 2016
– *parallelina* Hampson, 1894, **comb. rev.**
– *paraseriata* N. Singh, Kirti & Joshi, 2016
– *pectena* (Bucsek, 2012), **comb. nov.**
– *pellucida* (de Joannis, 1928), **comb. nov.**
– *peloa* (Swinhoe, 1904)
– *perihaemia* (Hampson, 1900), **comb. nov.**
– *phaeoplagia* (Hampson, 1900), **comb. nov.**
– *phantasma* (Hampson, 1907), **comb. nov.**
– *pingera* (Bucsek, 2012), **comb. nov.**
– *platyrhabda* (Tams, 1935), **comb. nov.**
– *poring* (Holloway, 2001), **comb. nov.**
– *postbicolor* (Rothschild, 1913), **comb. nov.**
– *postseriata* (Holloway, 2001), **comb. nov.**
– *poststrigata* Schaus, 1922
– *producta* (Černý, 2009), **comb. nov.**
– *pseudoarcuata* N. Singh & Kirti, 2016
– *pseudobunda* (Holloway, 2001), **comb. nov.**
– *pseudodistributa* (Bucsek, 2014), **comb. nov.**
– *pseudolutara* N. Singh & Kirti, 2016
– *pseudomodesta* Joshi, N. Singh & Volynkin, 2017
– *pseudoseriata* N. Singh & Kirti, 2016

– *pudibunda* (Snellen, 1880)
– *puncakica* (Dubatolov & Bucsek, 2014), **comb. nov.**
– *punctata* (Rothschild, 1913), **comb. nov.**
– *punctilineata* (Wileman & South, 1919), **comb. nov.**
– *quadra* Joshi, N. Singh & Volynkin, 2017
– *quadrata* (Holloway, 2001), **comb. nov.**
– *ranruna* (Matsumura, 1927), **comb. nov.**
– *reflexusa* (Fang, 2000), **comb. nov.**
– *rhabdota* (Rothschild, 1920), **comb. nov.**
– *rigidistria* Candèze, 1927
– *rosacea* (Bremer, 1861)
= *shuotsuensis* Bryk, [1949] 1948
= *undulata* Leech, 1901
– *rosalia* (Hampson, 1914)
= *rosacea* Rothschild, 1913, nec. Bremer, 1861
= *javana* Draudt, 1914
– *rubricans* (Leech, 1890)
– *rubricosa* (Moore, 1878)
= *curvifascia* Hampson, 1891
= *ochracea* Hampson, 1891
– *ruptifascia* (Hampson, 1891), **comb. nov.**
– *semifascia* (Walker, 1854)
= *metamelas* Hampson, 1891
= *postfusca* Draudt, 1914
– *sexpuncta* Hampson, 1894
– *simonka* (Bucsek, 2014), **comb. nov.**
– *simplicior* (Matsumura, 1927), **comb. nov.**
– *sinica* Moore, 1877, **comb. rev.**
– *solita* Walker, 1954
= *punctilinea* Moore, 1878
– *striata* (Wileman, 1910), **comb. nov.**
– *strigatula* (Rothschild, 1913), **comb. nov.**
– *strigibasis* (de Joannis, 1930), **comb. nov.**
– *strigipennis* (Herrich-Schäffer, 1855)
– *synestramena* (Hampson, 1900), **comb. nov.**
– *szetschwanica* (Daniel, 1952)
– *taprobana* (Hampson, 1907), **comb. nov.**
– *terminata* (Moore, 1878), **comb. nov.**
– *terminospota* (N. Singh, Kirti & Joshi, 2015)
– *testata* (Černý, 2016), **comb. nov.**
– *tibeta* Daniel, 1951, **comb. rev.**
– *tronga* (Schaus, 1922), **comb. nov.**
– *tsinglingensis* Daniel, 1951, **comb. rev.**
– *tuta* Fang, 1991
– *uncalis* (Kirti & Gill, 2009)
– *undulata* (Swinhoe, 1903)
– *undulosa* (Walker, 1854)
= *straminea* Walker, 1856
= *excurrens* Walker, 1864
= *decurrens* Walker, 1864
– *undunoides* N. Singh & Kirti, 2016
– *unguifera* (Holloway, 2001), **comb. nov.**
– *uniformeola* (Hampson, 1900), **comb. nov.**
– *unipuncta unipuncta* (Leech, 1890)
– *unipuncta mienshanica* (Daniel, 1952)

– *valdenigra* (Bucsek, 2012), **comb. nov.**
– *venustusa* (Bucsek, 2012), **comb. nov.**
– *vetusta* Snellen, 1904
– *violacea* (Reich, 1936), **comb. nov.**
– *wangmini* Volynkin, Dubatolov & Kishida, 2018
– *weidenhofferi weidenhofferi* (Černý, 2012)
– *weidenhofferi maritima* (Černý, 2016), **comb. nov.**
– *wenchiyehi* (Wu, Fu & Chang, 2013), **comb. nov.**
– *xanthopera xanthopera* (Hampson, 1907), **comb. nov.**
– *xanthopera montana* (Bucsek, 2014), **comb. nov.**
– *x-linea* (Bucsek, 2012), **comb. nov.**
– *yimingcheni* (Wu, Fu & Chang, 2013), **comb. nov.**
– *ziczac* (Walker, 1856)
= *rivalis* Leech, 1890
– *zinchenkoi* (Dubatolov & Bucsek, 2013), **comb. nov.**
– *zorae* (Černý, 2016), **comb. nov.**

Cabarda Walker, 1863
– *nigripuncta* (Wileman & South, 1919)
= *lunilinea* Schaus, 1922, **syn. nov.**
– *sequens* (Walker, 1862);
= *molliculana* Walker, 1863
= *sequens* f. *sumatrana* Draudt, 1914

Gurna Swinhoe, 1892
– *indica* (Moore, 1879)

Adites Moore, [1882]
– *alticurvata* Holloway, 2001
– *bakeri* (Schaus, 1922), **comb. nov.**
– *bifida* Holloway, 2001
– *bizonoides* (Walker, 1862)
= *tabida* Snellen, 1880
– *cornutata* Holloway, 2001
– *curvata* Holloway, 2001
– *impilia* Černý, 2009
– *hilaris* (Walker, 1854)
– *hosei* Holloway, 2001
– *longispina* Holloway, 2001
– *luangensis* Černý, 2009
– *maculata* (Poujade, 1886), **comb. nov.**
= *limbata* (Wileman, 1911)
= *pica* Wileman, 1911
= *maculata* var. *formosibia* Strand, 1917
= *maculata* [f.] *shirakii* Matsumura, 1930
= *karenkonis* Matsumura, 1930
– *mindanensis* (Schaus, 1922), **comb. nov.**
– *ocellata* (Wileman & South, 1919)
– *paraimpilia* N. Singh, Kirti & Kaleka, 2016
– *parafrigida* Černý, 2009
– *parang* Holloway, 2001
– *pisina* Černý, 2009
– *pseudofrigida* Holloway, 2001
– *pseudohilaris* N. Singh & Kirti
– *pseudolyclene* Holloway, 2001

– *retusa* Černý, 2009
– *sandakan* Holloway, 2001
– *temburong* Holloway, 2001
– *thanhi* Dubatolov & Bucsek, 2016
– *trusmadi* Holloway, 2001
– *unilinea* (Wileman & South, 1919)
– *vagilinea* (Walker, 1862)

Pseudoadites N. Singh & Kirti, 2016
– *frigida* (Walker, 1854)
= *diffusa* Walker, 1862
= *griseotincta* Rothschild, 1913

Afrasura Durante, 2009
– *aetheria* Durante, 2012
– *amaniensis* (Cieslak & Häuser, 2006)
– *camilla* Durante, 2012
– *craigii* (Holland, 1893)
– *crenulata* (Bethune-Baker, 1911)
– *discocellularis discocellularis* (Strand, 1912)
– *discocellularis saotomensis* Volynkin & László, 2018
– *discreta* Durante, 2009
– *dubitabilis* Durante, 2009
– *duplex* Durante, 2012
– *emma* Durante, 2009
– *fracta* Durante, 2012
– *friederikeae* (Kühne, 2007)
– *hieroglyphica* (Bethune-Baker, 1911
= *thomensis* (Rothschild, 1913)
– *hyporhoda* (Hampson, 1900)
– *ichorina* (Butler, 1877)
– *indecisa indecisa* (Walker, 1869)
= *rufostria* Plötz, 1880
= *gabunica* Holland, 1893
= *xantha* Bethune-Baker, 1911
= *guentheri* Strand, 1912
– *indecisa orientalis* Durante, 2009
– *neavi* (Hampson, 1914)
– *numida* (Holland, 1893)
– *obliterata* (Walker, 1864)
– *pallescens* Durante, 2009
– *pectinatissima* Volynkin & László, 2018
– *peripherica peripherica* Strand, 1912
– *peripherica hilara* (Kiriakoff, 1958)
– *pinkurata* (Kühne, 2007), **comb. nov.**
– *rivulosa* (Walker, 1854)
= *fulvia* (Hampson, 1900)
= *obsolescens* (Hampson, 1914)
– *rivulosa ethiopica* Durante, 2009
– *subfulvia* (Kiriakoff, 1954), **comb. nov.**
– *submarmorata* (Kiriakoff, 1958)
– *tanzaniae* Volynkin, 2019
– *terlineata* Durante, 2009
– *trunca* Durante, 2012
– *violacea* (Cieslak & Häuser, 2006)

Species *insertae sedis*
– *clara* (Holland, 1893)
– *erythrias* (Holland, 1893)
– *temperata* (Holland, 1893)

Palaeugoa Durante, 2012
– *camerunensis* (Strand, 1912), **comb. nov.**
– *secunda* Volynkin, 2017
– *spurrelli* (Hampson, 1914)

Tumicla Wallengren, 1863
= *Asuroides* Durante, 2008
– *admiranda* Volynkin & László, 2019
– *arktos* Volynkin & László, 2018
– *atricraspeda* (Hampson, 1914)
– *calimerae* (Durante, 2008)
– *congoensis* (Kühne, 2007)
– *dimidiata* (Durante, 2008)
– *distyi* (Kühne, 2007)
– *doa* (Kühne, 2007)
– *eala* (Kühne, 2007)
– *elephantina* Volynkin & László, 2019
– *fasciata* (Durante, 2008)
– *gigantea* (Kühne, 2007)
– *hermanni* (Kühne, 2007)
– *mbeghai* Volynkin & László, 2019
– *mutabilis* (Kühne, 2007)
– *retromaculata* (Durante, 2008)
– *rosea* (Durante, 2008)
– *rubea* (Durante, 2008)
– *russula* (Kiriakoff, 1963)
– *sagenaria* (Wallengren, 1860)
– *similis* (Durante, 2008)
– *smithi* Volynkin & László, 2019
– *spinata* (Kühne, 2007)
– *tsonga* Volynkin & László, 2018
– *versicolor* (Kühne, 2007)

Parafrasura Durante, 2012
– *pectinella* Strand, 1922

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