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Summary

24 • Despite the important role that biocrust communities play in maintaining ecosystem structure and functioning in drylands worldwide, few studies have evaluated how climate change will affect them.

• Using data from an eight-yr old manipulative field experiment located in Central Spain, we evaluated how warming, rainfall exclusion and their combination affected the dynamics of biocrust communities in areas that initially had low (<20%, LIBC plots) and high (>50%, HIBC plots) biocrust cover.

31 • Warming reduced the richness $(35\pm6\%)$, diversity $(25\pm8\%)$ and cover $(82\pm5\%)$ of biocrusts in HIBC plots. The presence and abundance of mosses increased with warming through time in these plots, although their growth rate was much lower than the rate of lichen death, resulting in a net loss of biocrust cover. On average, warming caused a 35 decrease in the abundance $(64\pm7\%)$ and presence $(38\pm24\%)$ of species in the HIBC plots. Over time, lichens and mosses colonized the LIBC plots, but this process was hampered by warming in the case of lichens.

• The observed reductions in the cover and diversity of lichen-dominated biocrusts with warming will lessen the capacity of drylands such as that studied here to sequester 40 atmospheric $CO₂$ and to provide other key ecosystem services associated to these communities.

Key words: biocrust cover, biological soil crust, climate change, drylands, evenness, lichens, mosses, richness

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Introduction

Understanding how drylands (comprised of hyperarid, arid, semiarid and dry subhumid ecosystems) are responding to ongoing climate change is critical for establishing effective sustainability policies. Drylands currently cover ~45% of the Earth's land surface (Prăvălie, 2016), and forecasted increases in aridity will expand them by up to 23% globally by the end of this century (Huang *et al*., 2016). Biocrusts, communities living in the uppermost few millimeters of the soil and composed of autotrophic (e.g., lichens, mosses, liverworts, cyanobacteria and green algae) and heterotrophic (e.g., bacteria, fungi and microfauna) organisms, are prevalent in drylands worldwide (Weber *et al*. 2016a). These communities grow intimately linked to soils from hyperarid to subhumid ecosystems. In the most extreme climate regions, biocrusts are only composed by the microscopic constituents, but as aridity decreases, macroscopic lichens, mosses and liverworths become more prevalent (Bowker *et al*., 2016). They are involved in many 67 important ecosystem processes, including carbon (C) and nitrogen (N) cycling (Evans $\&$ Lange, 2003; Hu *et al*., 2015; Barger *et al*., 2016; Sancho *et al*., 2016), surface energy balance (Rodríguez-Caballero *et al*., 2015; Couradeau *et al*., 2016; Rutherford *et al*., 2017), erosion (Zhao & Xu, 2013; Cantón *et al*., 2014; Chamizo *et al*., 2017) and water redistribution (Bowker *et al*., 2013a; Kidron & Büdel, 2014; Chamizo *et al*., 2016), just to name a few. Biocrusts have also been found to modulate the magnitude of responses of C and N cycling to climate change in experimental studies (Delgado-Baquerizo *et al*., 2014; Maestre *et al*., 2013; Hu *et al*., 2014). Recent surveys have also found that their relative importance as a provider of functions related to nutrient cycling increases along aridity gradients (Delgado-Baquerizo *et al*., 2016). Therefore, biocrusts will be critical for maintaining the multiplicity of functions and services provided by drylands in a warmer and more arid Earth. Nevertheless, it is not clear how the abundance and composition of these communities will respond to climate change. This limits our ability to understand and predict its impacts on ecosystem functioning in ecosystems where biocrusts are a prevalent biotic community.

Expected decreases in the cover of vascular plants under increased aridity scenarios (Delgado-Baquerizo *et al*., 2013) could provide biocrusts new available surface for colonization. However, short- and long-term experiments conducted in cold (e.g., Wertin *et* *al*., 2012; Zelikova *et al*., 2012; Ferrenberg *et al*., 2015) and hot (Escolar *et al*., 2012; Maestre *et al*., 2013, 2015) drylands have shown detrimental effects of climate change treatments on biocrust cover and development, as well as important shifts in their composition. For example, in the Colorado Plateau (USA), a single season of increased summer rainfall frequency caused a drastic mortality of mosses and an increase in the abundance of cyanobacteria (Zelikova *et al*., 2012), whereas the implementation of nearly six years of experimental warming was necessary to record negative effects on mosses and lichens (Ferrenberg *et al*., 2015). In Central and SE Spain, important declines (up to 40%) in the cover of biocrust-forming lichens appear after two/three years of warming (Escolar *et al*. 2012, Maestre *et al*., 2015). These results highlight the importance of evaluating multiple biocrust constituents in long-term experiments conducted under different environmental settings when assessing climate change impacts on biocrust communities.

97 In this study, we describe how simulated climate change $(2-3^{\circ}C \text{ warming}, -35^{\circ})$ rainfall reduction and the combination of both) and the degree of biocrust development affect the cover, richness, diversity, evenness and composition of mosses and lichens in a semiarid ecosystem from Central Spain during the first eight years after the setup of the experiment in 2008. This experiment, still ongoing, is the only of its kind being carried out in an European dryland and, together with that of Ferrenberg *et al*. (2015), is one of the very few experiments that have evaluated climate change impacts on biocrusts for more than six years. Our goal is to expand the previous findings reported by Escolar *et al*. (2012) with new data and statistical approaches aiming to assess how simulated climate change is affecting biocrust community attributes at the short (2008–2011) and medium (2011–2016) term. We also aim to determine the species-specific responses of lichens, the dominant biocrust constituent in our study area, and mosses to simulated climate change.

Materials and Methods

Study area and experimental design

The study was conducted at the Aranjuez Experimental Station, located in central Spain (40°02´N–3°32´W; 590 m a.s.l.). The climate is Mediterranean semi-arid, with mean 114 annual temperature and rainfall values of 15^oC and ~350 mm, respectively. The soil is classified as Gypsiric Leptosol (IUSS Working Group WRB, 2006). Perennial plant

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coverage is lower than 40%, and is dominated by the perennial herbaceous species *Macrochloa tenacissima*. To a lesser extent, there are also isolated individuals of shrubs such as *Helianthemum squamatum, Gypsophila struthium* and *Retama sphaerocarpa*. Open areas between plants contain a well-developed biocrust community dominated by lichens such as *Diploschistes diacapsis*, *Squamarina lentigera*, *Fulgensia subbracteata* and *Buellia zoharyi*. Moss-dominated crust with species such as *Pleurochaete squarrosa* and *Didymodon acutus* are also present (mostly under the canopy of *M. tenacissima*), as well as cyanobacteria of the genera *Microcoleus*, *Schizothrix*, *Tolypothrix*, *Scytonema* and *Nostoc* (Cano-Díaz *et al*., 2018). See Maestre *et al*. (2013) for a complete species checklist of visible biocrust constituents at our study site.

In 2008, we established a full factorial experiment with three factors, each with two levels: initial biocrust cover (IBC; <20% *vs.* >50%), warming (WARM; control *vs.* temperature increase), and rainfall exclusion (RE; control *vs.* rainfall reduction). Ten replicates per combination of treatments were randomly established in the study area, resulting in 80 experimental plots in total. A minimum separation distance of 1 m between plots was kept to minimize the risk of lack of independence between treatments. The applied warming by 2–3ºC is in line with several Atmosphere-Ocean General Circulation Models for the second half of this century in Spain (De Castro *et al*., 2005; Collins *et al*., 2013). Although precipitation predictions are subject to a higher level of uncertainty in the Mediterranean Basin, projected changes point to an intensification of water scarcity in this region (Christensen *et al*., 2007, 2013; Giorgi & Lionello, 2008). To achieve a temperature increase within the forecasted range, we used open top chambers (OTCs) with hexagonal 138 design and sloping sides of $40 \times 50 \times 32$ cm. These chambers were built with methacrylate sheets, a material which ensures high transmittance in the visible spectrum (92%, according to the manufacturer; Decorplax S.L., Humanes, Spain) and a very low emission in the infrared wavelength. They are suspended 3–5 cm over the ground by a metal frame to allow free air circulation at the soil surface level to avoid an excessive overheating and to minimize obstacles for propagule dispersion. Passive rainfall shelters (RS) based on the design of Yahdjian & Sala (2002) were used to achieve a reduction of rainfall amount of \sim 35% without changing the frequency of rainfall events. Each RS has an area of 1.44 m² $(1.2 \times 1.2 \text{ m})$ and a roof composed by three gutters of methacrylate with an inclination of 147 20 \degree and a mean height of 1 m, which cover \sim 37% of this area. For logistic reasons, OTCs

and RS were set up in July and November of 2008, respectively. More details of these

infrastructures and their microclimatic effects are available in Escolar *et al*. (2012) and

Maestre *et al*. (2013).

Quantification of water inputs

The duration, nature and timing of water inputs are key determinants of the physiological status of biocrust constituents such as lichens and mosses (Pintado *et al*., 2010; Green *et al*., 2011; Ladrón de Guevara *et al*., 2014). As such, the interaction of our treatments with features of water inputs could be essential for interpreting their effects on biocrust communities. We recorded rainfall events using an on-site meteorological station (Onset Corp., MA, USA). Periods with data gaps due to technical problems of this station were filled with records from of another station near to this area provided by the Spanish Agency of Meteorology (AEMET; Aranjuez station, located 4 km from our experiment). To consider non-rainfall water inputs (NRWIs), potential fog events —likely the main NRWI in this area— were estimated with relative air humidity sensors (Hobo U23 Pro v.2 Temp/RH, Onset Corp., MA, USA; and DS1923 iButton Temp/RH Logger, Dallas 163 Semiconductor, TX, USA) and consecutive filter criteria programmed in R (version. 3.2.4; R Foundation for Statistical Computing, Vienna, Austria) for their differentiation from rainfall inputs (see more details in Supporting Information Fig. S1).

Monitoring changes in biocrust assemblages through time

During the setup of the experiment, we inserted a PVC collar of 20 cm diameter in each plot to monitor changes in the cover, richness, diversity, evenness and composition of biocrust communities through time. We recorded the presence and frequency of all visible biocrust constituents (mosses and lichens) at the beginning of the experiment (June 2008) and later in May 2011 and 2016 using the point-sampling method. A wire mesh of 12 cm 172 each side with 1×1 cm grid (144 sampling points) was placed over the collars, and the species located under each of the sampling points was recorded. Although four different species of bryophytes have been identified in this site (Maestre *et al*., 2013), they were clustered in a single category (mosses) due to the difficulty of identifying them with confidence in the field. For the same reason, some lichen species of the same genus present in the area were clustered in this last taxonomic level (i.e., *Fulgensia* spp., *Collema* spp.,

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Endocarpon spp.). Using data from these surveys, we calculated species richness, diversity (using the exponential Shannon index with the natural logarithm in the formula; Shannon, 1948) evenness (using Pielou's index; Pielou, 1966) and composition (measured using both the relative abundance and presence/absence of each species). In each collar, annual changes in the total cover of the whole biocrust community were also estimated throughout the experiment using repeated photographs and the software GIMP (http://www.gimp.org), as explained in Maestre *et al*. (2013). Cover estimates obtained by this method correlate well with the values obtained from the point-sampling method (Supporting Information Fig. S2).

Statistical analyses

We used general linear mixed models (LMMs) to analyze the temporal effects of the climate change treatments on the cover, richness, diversity and evenness of the biocrust community. WARM, RE, IBC and year of survey (Year), as well as their interactions, were included in the models as fixed factors. The influence of the variability between replicates was also tested by including them in the model as a random factor and by supervising the effect of this inclusion on the Akaike's Information Criterion (AIC, Akaike, 1973) of the model. Validation of the assumptions of the models were performed by checking the autocorrelation function plot, the Pearson standardized residuals *vs.* fitted values plot and the distribution of these residual by a QQ-plot. After running the best fitted model (considering AIC values) and checking the compliance of the model assumptions, the number of interactions in the fixed part was reduced to gain degrees of freedom, and thus to increase the power of the model. The exclusion started from the interaction with more terms up to the most complex interaction with a significant (< 0.05) *P* value. The final models were run using a restricted maximum likelihood (REML) procedure, and when significant differences were detected in interactions or in the factor Year (the only one with more than two levels), Fisher's least significant difference (LSD) test was applied.

Changes in the relative abundance and presence/absence of species through time caused by the treatments were analyzed using permutational multivariate analysis of variance (PERMANOVA). These analyses were performed on Bray-Curtis similarity matrices of fourth-root transformed species abundances and dichotomous species presence/absence data, respectively. Very rare species not detected in any of the three

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point-sampling surveys were not retained in the analyses. Since our dataset contained samples with very few species records, thus significantly increasing the variability of the 211 data, a dummy variable (weight 1; i.e., a fictitious species with value $= 1$ in all plots) was added to the matrix (Clarke & Gorley, 2006). For balanced designs such as ours, PERMANOVA is a robust method to the heterogeneity of multivariate dispersions (Anderson & Walsh, 2013). As in the LMM analyses, the four factors and all their interactions were introduced in a first step, and a reduction of interactions was applied in a second step following the criterion explained above. *P*-values were calculated using 999 permutations of the residuals under a reduced model, which is the method with the best power and the most accurate Type I error for complex designs (Anderson, 2005). Post-hoc pair-wise comparisons were done with all significant interactions and the factor Year.

The magnitude of the temporal effects of each treatment on the relative abundance (fourth-root transformed) and presence/absence of species were also modeled and visualized using Principal Response Curves (PRC, Van den Brink & Ter Braak, 1998, 1999). This method evaluates the differences between the values of the variables in the treatments and those in the control (standardized to zero-value) in each survey. In addition to scores for each treatment through time, PRC also offers scores for each species, which allow assessing their weight on the overall response (i.e., changes in the relative abundance and presence) of the whole community to the climatic treatments evaluated. In addition, we conducted SIMPER analyses to disentangle the contribution of each species to the overall compositional shifts caused by the factors with significant interactions detected in the PERMANOVA analyses. SIMPER analyses show the percentage of contribution of each species to the average Bray-Curtis dissimilarity or Euclidean squared distance between the levels of the factors evaluated (Clarke & Warwick, 1994).

LMMs, LSD tests and the plots for the validation of the assumptions were performed using the software InfoStat Version 2014 (Di Rienzo *et al*., 2014). PERMANOVA and SIMPER analyses were carried out with the PERMANOVA+ 1.0.1 for the PRIMER 6.1.11 statistical package (PRIMER-E Ltd, Plymouth Marine Laboratory, UK). PRC analyses using the package 'vegan' (Oksanen *et al*., 2016) in the R statistical environment. The data used in our analyses are available from figshare (Ladrón de Guevara *et al*., 2017).

Results

Responses of biocrust cover to climatic treatments

The analysis of cover data revealed two significant interactions among the experimental 243 factors (IBC \times Year and WARM \times Year, Table S1). As only two-term interactions were significant in the saturated models, in a second step of the analyses the more complex interactions were eliminated. This analysis did not detect any statistically significant effect of rainfall exclusion on cover, neither as a main effect nor as a term in the interactions (Table S1). Plots with low IBC (LIBC) and without warming experienced a significant 248 increase of their cover between 2008–2011 (from $5\pm0.9\%$ to $15\pm2.2\%$ on average), had a static period between 2011–2015 and started to increase their cover again in 2016 (Fig. 1a). The cover of these plots also increased by 2011 under warming, but at a significantly lower rate than without this treatment, and after this year, decreased to values similar to those observed in 2008. Changes in cover through time in plots with high IBC (HIBC) followed a distinct pattern (Fig. 1b). From 2008 to 2013, the cover of plots without warming 254 decreased by $41\pm3.9\%$. This tendency was reversed after 2013, but without reaching the initial cover values (~80% on average). The decrease observed in WARM plots was more abrupt and did not stop in 2013. As a result, the loss of cover in these plots was large over the course of the experiment, with an 82±5% of reduction over the course of the experiment and with final cover values lower than those found in the plots with low IBC and without warming. Significant differences in cover with warming were detected three years after the setup of the experiment in both LIBC and HIBC plots (Fig. 1).

Treatment effects on biocrust community attributes through time

As found with cover, only two-term interactions were statistically significant when analyzing the richness, diversity and evenness of biocrust assemblages (Table S2). The 264 IBC \times Year interaction reflected natural trends on their richness and diversity, although they are intermixed with the effects of the climate change treatments. Biocrust richness and diversity decreased during the period 2008–2011, but recovered thereafter in the controls with HIBC (Fig. 2) and in all HIBC plots (Fig. 3). Such a decrease did not occur in the LIBC plots; the significant rise of richness and diversity observed after 2011 was the result 269 of the intensification of the natural colonization of these plots by biocrusts. The WARM \times Year interaction observed in the LMM analyses also indicated that biocrust colonization did not happen in WARM plots. Warming promoted a fast and significant decline in biocrust richness and diversity throughout the duration of the experiment (Fig. 3). Rainfall exclusion negatively affected biocrust diversity only in the HIBC plots, as noted by the 274 statistically significant IBC \times RE interaction (Table S2), but the magnitude of this effect was lower than that promoted by WARM (Fig. 2). Changes in evenness were driven by 276 IBC \times WARM and WARM \times RE interactions. In the first case, species evenness only decreased in the LIBC plots subjected to warming, and in the second such a reduction was 278 only observed in $WARM + RE$ plots (Fig. 3).

Warming was the primary treatment promoting differences on species composition when compared to control plots, an effect that increased through time (Fig. 4). PERMANOVA analyses indicated that the effects of warming became significant during the first experimental period (2008–2011). The magnitude of this effect was similar in LIBC and HIBC plots, but was greater when analyzing presence/absence than relative abundance data (46% and 53% greater in LIBC and HIBC plots, respectively). However, the main driver of community shifts differed between LIBC and HIBC plots. As found when evaluating changes in cover, an increase of the values of the species composition attributes without warming took place in the LIBC plots during the period 2008–2016 (Fig. 4a,c). This process did not occur under warming. This treatment negatively affected the growth of biocrusts, and therefore the values of species composition attributes were similar throughout the course of the experiment in LIBC plots (Fig. 4a,c). The major shifts in species composition took place in plots without warming, as also indicated by the positive values of the parameter taxon weights observed in the PRC analysis. In the LIBC plots, most species increased their presence and relative abundance, but such an increase was particularly noticeable for *Fulgensia* spp., *D. diacapsis*, *S. lentigera* and *Toninia sedifolia*. Only *Squamarina cartilaginea* had a negative score in the PRC analyses (Fig. 4a,c). On the other hand, warming was a main driver of changes in the composition of high HIBC plots, especially when evaluating changes in the relative abundance of species. The abundance of species in plots without warming was stable over the monitoring time, while the number of species present in these plots increased (Fig. 4b,d). Warming decreased the relative abundance and presence of most of the studied species. The opposite trends in the presence of species promoted the greater differences between the control and warming treatments

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detected in the PRC analysis. Across all treatments, we found a clear decline in the relative abundance of *D. diacapsis*, in the presence of *T. sedifolia* and in both the relative abundance and presence of *Fulgensia* spp. and *Psora decipiens* (Fig. 4b,d). However, we found an increase in both the relative abundance and presence of mosses through time.

306 PERMANOVA analyses revealed a significant IBC \times Year interaction when analyzing both relative abundance and species presence data (Table S3). The Bray-Curtis similarity index showed an increase in the homogeneity of the compositional attributes between LIBC and HIBC plots throughout the 2008–2016 period (Table S4). SIMPER analyses showed temporal changes of main species leading the compositional variations at both IBC levels (Table S5). This last analysis highlighted *D. diacapsis* as the most dynamic species throughout the experiment, as it showed the highest colonizing capacity on LIBC plots, and was the third species with the largest losses in both the presence and relative abundance in the HIBC plots (Table S5). As a result, in 2016 the average relative abundance of this species was slightly higher in the LIBC than in the HIBC plots. *Fulgensia* spp. largely contributed to the observed changes in species relative abundance throughout the experiment, whereas *T. sedifolia* had a relevant role in the changes of presence/absence observed during the same period (Table S5). Reductions in the relative abundance of *Fulgensia* spp. in the HIBC plots were much more important than increases on its relative abundance in the LIBC plots. The contribution of this species to presence/absence changes was only relevant in the 2011–2016 period, with important declines in their presence at both LIBC and HIBC plots. The dynamics of the presence of *T. sedifolia* changed during the experiment. It suffered a large decline during the 2008– 2011 period in the HIBC plots, and did not increase in the LIBC plots. However, its presence increased in the 2011–2016 period at both LIBC and HIBC plots, particularly in the former. SIMPER analyses also suggested that the decline in richness and diversity in the 2008–2011 period was mainly driven by reductions in both the relative abundance and presence of *Acarospora nodulosa*, *D. diacapsis*, *Fulgensia* spp. and *T. sedifolia* (Table S5), and their recovery in the 2011–2016 period was led by mosses and *T. sedifolia.* Mosses and lichens such as *B. zoharyi*, *D. diacapsis* and *T. sedifolia* were main actors in the colonization process observed in the LIBC plots through time.

The Bray-Curtis index showed a low degree of similarity in the compositional attributes within WARM plots and between the 2008–2016 period (Table S4). Overall, both the PERMANOVA and SIMPER results revealed that, over the course of the experiment, *D. diacapsis*, *Fulgensia* spp. and *T. sedifolia* were the species most affected by warming (Table S5). *Fulgensia* spp. showed the largest reductions in presence and relative abundance, although this response was particularly evident in the 2011–2016 period. *T. sedifolia* and *D. diacapsis* had a greater contribution to changes in species presences/absences and relative abundance, respectively. Unlike *Fulgensia* spp., these 340 species had a faster response to warming. The WARM \times Year interaction also elucidated in part the changing dynamics explained above for *T. sedifolia*; in the 2011–2016 period, a greater expansion of this species took place in the non-WARM plots along with its neutral response to warming, likely because during 2008-2011 this species disappeared from most 344 WARM plots. The observed WARM \times RE interaction revealed that the negative effect of warming observed in *D. diacapsis* was enhanced under rainfall exclusion. Warming caused greater declines of both the relative abundance and presence of *Fulgensia* spp. in plots without rainfall exclusion, as well as in the presence of *P. decipiens*. PERMANOVA 348 analyses revealed that the significant IBC \times RE interaction detected when analyzing species diversity was largely driven by reductions in both the relative abundance and presence of *P. decipiens* in the high IBC plots subjected to rainfall exclusion.

Discussion

Warming drastically reduced the cover of lichen-dominated biocrusts but promoted moss growth

In contrast with the widespread perception over the past several decades that biocrust constituents, especially crustose lichens, conform a fairly static community due to their slow growth and resistance to extreme environments (Garvie *et al*., 2008; Armstrong & Bradwell, 2010; Meeßen *et al*., 2013), our findings highlight the dynamism of biocrust cover and its fast response to environmental changes. Similar results have been found with biocrust-forming lichens in drylands from other parts of the world (e.g., Belnap *et al*., 2006, 2007; Jimenez Aguilar *et al*., 2009; Read *et al*., 2011; Maestre *et al*., 2015). Cover or biomass of these communities can stay static or decline substantially during drought events

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or other stressful conditions. But terricolous lichens can have pulses of fast growth during wet periods (Belnap *et al*., 2006; Weber *et al*., 2016a). In addition, another feature of some terricolous lichen species is the possession of certain capacity of mobility by dispersion of thallus fragments (Heinken, 1999), such as *D. diacapsis*, the most dynamic species detected in this study. This species can have different degrees of attachment in the soil, sometimes being it able to detach easily (Ballesteros *et al*., 2017). Thus, the fast losses of cover of this species detected in the warming treatment are likely due to a detachment of fragments from the soil and their dispersion through wind to other places. Similarly, we have also detected new, but well established, large thallus of *D. diacapsis* from one year to another in the other treatments, which likely have an external origin (i.e. they come from outside the plots). Therefore, the semi-erratic behaviour of this species, together with its growth capacity when conditions are optimal (Lázaro *et al*., 2008), are presumably the two features that allow the dynamic response to the climate change treatments evaluated. The fast response of these communities to environmental conditions caused a contrasted cover dynamic in the two IBC levels of the control plots. During the experimental period, there were two consecutive wet seasons (2010–2011 and 2011–2012) where soil moisture was 379 below the usual average values $(\sim 15\%)$ for this season in this area (Fig. S1). This decrease in water availability could be not too strong to limit lichen colonization in areas with low competition for this resource (LIBC), but enough to trigger a decline of lichens in HIBC, where competition interactions were very likely stronger. In addition, we estimated a very low number of fog events in the wet season 2012–2013, a period that coincided with the greatest cover decline in HIBC.

The great loss of biocrust cover observed (Fig. 1) was clearly driven by warming, probably due to its indirect negative effect on soil surface moisture. Although we did not directly measure this effect due to the lack of appropriate sensors to accurately measure soil moisture in the first mm of soil depth, which have been developed very recently (Weber *et al*., 2016b), our continuous monitoring of soil subsurface moisture (0–5 cm depth) showed a highly significant negative effect of warming on soil water gains after rainfall events (Lafuente *et al*., 2017). Other studies have also reported negative effects of passive warming by OTCs on soil moisture (e.g., Klein *et al*., 2005; Bokhorst *et al*., 2016). In addition, we have already observed that reductions in soil surface moisture negatively

affect the C fixation rate of biocrust-dominated areas during the first years of our experiment (Maestre *et al*., 2013; Ladrón de Guevara *et al*., 2014). We cannot rule out a possible direct effect of warming on lichen physiology because the increase of dark respiration with temperature is well documented in these organisms (e.g., Lange *et al*., 1998; Lange & Green, 2004; Ouyan & Hu, 2017). Nevertheless, we think that it was not the main physiological impact since mosses had a positive response to warming, and increases in their respiration with warming can be similar or even greater than those observed in lichens (Weber *et al*., 2012; Ouyang & Hu, 2017). During late autumn and winter, the periods when biocrust-forming lichens are most active in Mediterranean drylands (Pintado *et al*., 2010; Raggio *et al*., 2014), net photosynthesis also increases with temperature because the optimal temperature values for the photosynthetic activity of these organisms are not surpassed. However, in an annual basis, the periods of C losses are in general greater than those of C gains (Evans & Lange, 2003), and the combination of both processes (increment of C losses and reductions in C gains directly and indirectly caused by warming, respectively) can produce C starvation in biocrust-forming lichens. Reductions in the production of photoprotective pigments are another documented effect of a lower availability of C for metabolic functions (Belnap *et al*., 2004). Although we did not 411 measure pigment concentrations, fluorescence values (F_v/F_m) recorded in this experiment and in another site in southern Spain indicated that radiation did not damage the photosynthetic system of biocrust constituents (Escolar *et al*., 2012; Ladrón de Guevara *et al*., 2014). Therefore, we discard declines in pigment production as a main cause of the biocrust mortality observed in our experiment.

The observed declines in cover are in line with those recorded after ten years of warming and rainfall manipulations in a cold desert of North America (Ferrenberg *et al*., 2015), although with differences in the main biocrust constituents affected, the drivers of the responses observed and the timing when significant effects were detected. In the study of Ferrenberg *et al*. (2015), an increase of small rainfall events during summer caused a fast response of the biocrust community driven by the high mortality of mosses during the first year of climatic manipulations. Warming and rainfall manipulation separately had variable effects on the cover of lichens throughout the experiment, but their interaction caused a significant decrease in their cover nearly five years after the beginning of the

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experiment, with no signs of recovery after that period. Conversely, warming was the main driver of lichen cover losses in both our study and in another similar experiment conducted in SE Spain (Maestre *et al*., 2015). In both cases, warming promoted a significant decline 428 in the cover of lichens compared to control plots $(\sim]35-45\%)$ just after three years of the treatment implementation, while the cover of mosses was not negatively impacted by warming. Even in our study, a clear increase of moss growth was promoted by this treatment (Fig. 4 and Table S5). Thus, the main drivers of changes in cover differed in these studies —warming in our study and the increment of small rainfall events during summer in Ferrenberg *et al*. (2015)— primarily due to differences in the dominant biocrust constituents and in the climatic treatments applied. In addition, it is possible that the different warming methods used in both studies (OTCs in this study *vs.* infrared lamps in Ferrenberg *et al*. 2015) have significantly influenced in the timing of the lichen cover responses, since OTCs could have a larger impact on air and soil moisture than infrared lamps (Reed *et al*., 2016; Young *et al*., 2017).

Effects of OTCs on other microclimatic variables (e.g., reduction in PAR and greater diurnal variability in air temperature) have also been reported (e.g., Klein *et al*., 2005; Bokhorst *et al*., 2013). However, we believe that such effects are having minor impacts on our results given the transparency of the plastic material used and the lack of significant effects of OTCs on the variability of air temperature registered in our experiment (Maestre *et al.*, 2013). We also consider that other possible disturbances unrelated to the treatments evaluated, such as those created by ourselves during the surveys had a minimal impact in the results due to the precautions adopted (something that was also indicated by the increases in cover observed in the LIBC control plots). However, we must acknowledge that the imposed rate of warming in our experiment is faster than that occurring naturally. The fact that we simulate in years a process that will happen in decades is likely causing a higher loss of maladapted species than that expected under the more gradual warming. Therefore, it is probably that in the abrupt biocrust decline detected in this study will be ameliorated, at least in part, by changes in increases in the cover of mosses, which we have found to respond positively to warming, and cyanobacteria, which have already been found to increase their abundance in response to climate change simulations in Ferrenberg *et al*. experiment.

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Climate change treatments altered biocrust community attributes

Understanding the effects of climate drivers on the assemblage of biotic communities is essential to predict how climate change will impact ecosystem dynamics and functioning (Jing *et al*., 2015; Maestre *et al*., 2016; Pecl *et al*., 2017). In our experiment, warming was involved four times in three different interactions that caused significant responses in the richness, diversity and/or evenness of biocrusts. However, rainfall exclusion had only two significant effects as a term of two different interactions, and did not significantly affect species richness. Warming reduced all community attributes (Figs. 1–4). These warming-induced changes were evident as early as three years since the setup of the experiment, a similar time span as that observed by Maestre *et al*. (2015) but faster than the changes detected by Ferrenberg *et al*. (2015), as noted above. In addition, the divergent results regarding the response of mosses between this last study and ours were likely because our warming treatment decreased the frequency of very short wetting events, which in mosses promote C losses. Observed responses in both experiments are in agreement with the different strategies used by lichens and mosses to cope with desiccation/rehydration cycles (reviewed by Green *et al*., 2011), and therefore, changes in biocrust drying–wetting cycles was the main modulator in the responses of the community assemblage in both studies. Succinctly, lichens are less tolerant to long desiccation periods, but have higher ability to fix C under suboptimal water content conditions. Both characteristics make frequent NRWIs critical for their C economy. On the other hand, mosses have a higher maximal water content, which enables them to continue photosynthesis for longer periods when the soil surface is saturated after rainfall events. However, they also have higher respiration rates than lichens during dehydration processes. Furthermore, mosses have a reduced ability to exploit water resources below their optimum water content, and, therefore, reducing the frequency of very small wetting events decreases the number of metabolic activation processes that could result in a negative C balance.

Rainfall exclusion did not have any negative effect on species richness, but reduced the diversity of the high HIBC plots and the evenness of both LIBC and HIBC plots when interacted with warming (Fig. 3). These effects were not detected during the first years of the experiment (Escolar *et al*., 2012), highlighting the importance of conducting long-term experiments when assessing the responses of biocrusts to climate change. It is plausible to

speculate that rainfall exclusion, which has a much lower negative impact on biocrusts than warming (Fig. 4), will also affect species richness in the future. Our findings show that the frequency of species (incorporated in diversity and evenness attributes) has a faster response to rainfall exclusion than richness because the former varies instantaneously with the dynamics of the most sensitive species, whereas changes in richness require the total mortality of these species. Although warming was the main factor driving changes in surface soil moisture in our experiment, the most critical water source for biocrusts, our rainfall exclusion probably had an effect on the infiltration depth of the large rainfall events. This treatment presumably reduced this infiltration depth, decreasing also the water stored in the soil after these rainfall events. This could have negative effects on the activity periods of biocrust, as this water is available for these organisms when it rises by capillarity to the surface due to evaporative forces (Colesie *et al*., 2016). This effect was observed previously in our experiment (Ladrón de Guevara *et al*., 2014), as rainfall exclusion reduced the duration of photosynthetic events in a day with antecedent rainfalls, whereas this treatment did not affect such duration in days without antecedent water inputs.

Although warming reduced the relative abundance and presence of all lichen species (Fig. 4), this effect was clearly less negative in *Fulgensia* spp, and to a lesser extent in *P. decipiens*, when rainfall exclusion was also present (Table S5). Within the biocrust community studied, the presence of the organic compound anthraquinone is an exclusive chemical trait of these two species (Concostrina-Zubiri *et al*., 2014). Anthraquinones are metabolites that can act as UV-protectors (Nguyen *et al*., 2013) and thus these pigments could have a different photoprotective effectiveness than those presented in the other species under the combination of warming and rainfall exclusion. However, this possible advantage of anthraquinones against other photoprotective pigments has not been studied as far as we know, and therefore we can only speculate that this is the mechanism underlying the responses observed. They could also be related to the depression of net photosynthesis observed at high thallus water contents. Species from the same habitat can show a gradient in the intensity of this depression (Lange *et al*., 1993), ranging for example from 0% in *Cladonia convoluta* to 100% in *Fulgensia fulgens* (Lange *et al*., 1995; Lange & Green, 2003). Intraspecific differences in the magnitude of this depression can also take place between habitats due to the important role that the attachment to substratum plays in the distribution of water within lichen thalli (Souza-Egipsy *et al*., 2002; Lange & Green, 2004). Thus, a reduction of the events that could cause a water saturation of the thallus with our rainfall exclusion treatment might have favored species that, in this study area, suffer a total net photosynthesis depression under water suprasaturation. We also found temporal species-specific responses to warming. The presence and/or relative abundance of *Collema* spp., *D. diacapsis*, *P. decipiens*, *S. lentigera* and *T. sedifolia* showed important reductions during the 2008–2011 period (Table S5), while these responses were especially relevant in the 2011–2016 period for *B. zoharyi* and *Fulgensia* spp. This suggests that these two species have a combination of specific functional traits that provided them short-term resistance to warming, although when the temporal threshold of three years was crossed the negative effects of this treatment on their abundance were dramatic.

Concluding remarks

Our results indicate that climate change, and warming in particular, will trigger a fast loss of late successional biocrust constituents and will reduce the richness, diversity and evenness of lichen-dominated biocrust communities. The impacts of biocrusts on ecosystem functioning, and especially on nutrient cycling, are largely driven by these community attributes (Maestre *et al*., 2012; Bowker *et al*., 2013b). In addition, biocrust-forming species have unique contributions to individual ecosystem functions (Bowker *et al*., 2011; Gotelli *et al*., 2011; Liu *et al*., 2017), suggesting that changes in the composition of biocrusts can have large impacts on ecosystem functioning (Maestre *et al*., 2012). As such, the changes in the community attributes of biocrusts observed in this study will lessen the capacity of drylands such as the studied one here to perform key ecosystem functions associated to these communities in a warmer, and more arid, world.

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Author contributions

F.T.M. planned and designed the research, M.L.G., B.G., J.R. and M.P. conducted

fieldwork, M.L.G., B.G., J.R. and A.L. processed and analysed data, M.L.G. and F.T.M.

wrote the manuscript and all authors contributed in the final review.

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Brief legends of figures and tables available in Supporting Information

- **Fig. S1** Characterization of water input events throughout the experiment and average soil
- subsurface moisture in control plots during lichens growth season.
- **Fig. S2** Relationship between the cover estimates obtained with the two methods used.
- **Table S1** Results of a general linear mixed model showing the effects of the treatments on
- biocrust cover.
- **Table S2** Results of general linear mixed models showing the effects of the treatments on
- biocrust attributes (richness, diversity and evenness).
- **Table S3** Results of PERMANOVA models showing the effects of the treatments on biocrust composition.
- **Table S4** Results of PERMANOVA models showing the average values between/within groups of the Bray-Curtis similarity index.
- **Table S5** Results from SIMPER analysis showing the contribution of lichen species and
- mosses to the Euclidean squared distance in species relative abundance and presence.
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Figure legends in the main text

- **Fig. 1** Temporal changes in total biocrust cover (including mosses and lichens) as response to climate treatments in plots with low (a) and high (b) initial biocrust cover (IBC). Data 864 are means \pm SE ($n = 10$). Asterisks indicate the first year where significant differences in total biocrust cover between the plots with and without warming are detected in each level 866 of IBC according to the post-hoc test Fisher's least significant difference (LSD). WARM = Warming and RE = Rainfall exclusion.
- **Fig. 2** Temporal changes in richness, diversity and evenness in all the treatments. Data are 869 means \pm SE ($n = 10$). WARM = Warming, RE = Rainfall exclusion and IBC = Initial biocrust cover.

871 **Fig. 3** Results of the Fisher's LSD post-hoc test conducted after significant $(P < 0.05)$ two-term interactions observed when analyzing species richness, diversity and evenness. Data 873 are means \pm SE (*n* varies with the interaction of factors). Panels with white background indicate that the significant interactions resulted from the linear mixed model analyses (see Table S2). In each panel, factor levels without letters in common show significant 876 differences among them $(P < 0.05)$. WARM = Warming, RE = Rainfall exclusion and IBC 877 = Initial biocrust cover (LIBC = Low IBC and HIBC = High IBC).

Fig. 4 Principal Response Curves showing the regression coefficients and weights obtained for particular taxa. These coefficients indicate the magnitude of the treatment effects on species composition (i.e., the spatial distance of the treatments regarding the control with 881 constant 0 values in the y-axis). Asterisks indicate significant P values in the Year \times 882 Warming (WARM) interaction, as revealed by PERMANOVA analyses $(* = P < 0.01; ** =$ $P \le 0.001$). Numbers in blue represent the mean of the sum of all relative abundance (ΣRA) 884 and presence (ΣP) values per plot at each time for the control treatment. The same symbols in red are showing these values for the warming treatment. Taxon weights show the relative contribution of each taxon to community changes, and can be interpreted as the weight of each taxon in the response to the treatments. Positive values indicate an increase in their relative abundance (a, b) or presence (c, d). Negative values indicate a reduction in these community attributes. For example, in panel a *Fulgensia* spp. was the species with the highest increment in relative abundance considering the whole study period and all observed responses to the treatments, suggesting that its colonization of control plots was a process with greater effects on its abundance than those promoted by the other treatments. RE = Rainfall exclusion and IBC = Initial biocrust cover.

Fig. 1 Temporal changes in total biocrust cover (including mosses and lichens) as response to climate treatments in plots with low (a) and high (b) initial biocrust cover (IBC). Data are means \pm SE (n = 10). Asterisks indicate the first year where significant differences in total biocrust cover between the plots with and without warming are detected in each level of IBC according to the post-hoc test Fisher's least significant difference (LSD). WARM = Warming and $RE =$ Rainfall exclusion.

171x70mm (300 x 300 DPI)

Fig. 2 Temporal changes in richness, diversity and evenness in all the treatments. Data are means \pm SE (n $= 10$). WARM = Warming, RE = Rainfall exclusion and IBC = Initial biocrust cover.

171x186mm (300 x 300 DPI)

Fig. 3 Results of the Fisher's LSD post-hoc test conducted after significant (P < 0.05) two-term interactions observed when analyzing species richness, diversity and evenness data. Panels with white background indicate that the significant interactions resulted from the linear mixed model analyses (see Table S2). In each panel, factor levels without letters in common show significant differences among them (P < 0.05). IBC $=$ Initial biocrust cover, WARM $=$ Warming and RE $=$ Rainfall exclusion.

342x157mm (300 x 300 DPI)

Fig. 4 Principal Response Curves showing the regression coefficients and weights obtained for particular taxa. These coefficients indicate the magnitude of the treatment effects on species composition (i.e., the spatial distance of the treatments regarding the control with constant 0 values in the y-axis). Asterisks indicate significant P values in the Year x Warming (WARM) interaction, as revealed by PERMANOVA analyses ($* = P < 0.01$; $** = P < 0.001$). Numbers in blue represent the mean of the sum of all relative abundance (Σ RA) and presence (Σ P) values per plot at each time for the control treatment. The same symbols in red are showing these values for the warming treatment. Taxon weights show the relative contribution of each taxon to community changes, and can be interpreted as the weight of each taxon in the response to the treatments. Positive values indicate an increase in their relative abundance (a, b) or presence (c, d). Negative values indicate a reduction in these community attributes. For example, in panel a Fulgensia spp. was the species with the highest increment in relative abundance considering the whole study period and all observed responses to the treatments, suggesting that its colonization of control plots was a process with greater effects on its abundance than those promoted by the other treatments. RE = Rainfall exclusion and IBC = Initial biocrust cover.

342x213mm (300 x 300 DPI)