



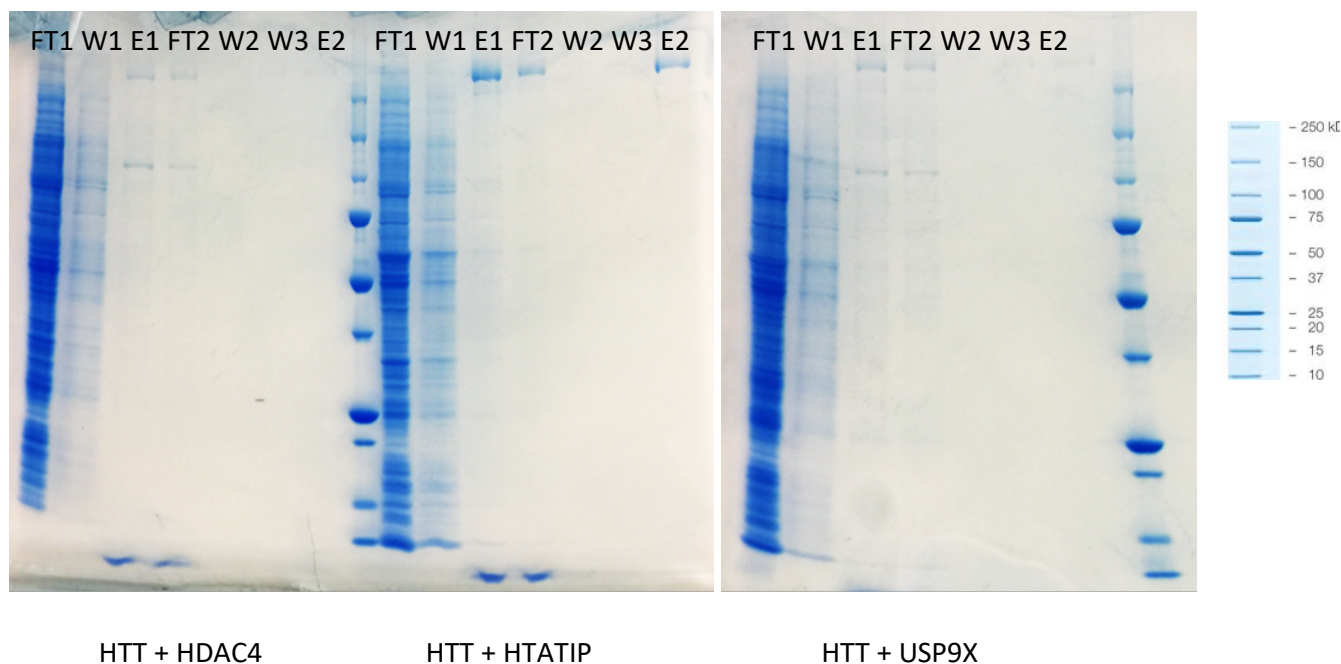
### **Growth:**

3:1 HTT Q23 (FLAG-tagged, TOC009:D01) to binding partner (His-tagged) virus ratios used for 1 L BVES sf9 production. Cells were harvested by centrifugation at 4000 rpm, 10 mins, 4 °C (Beckman JLA 8.1000). HTT cell pellets were resuspended in ~80 mL of 50 mM Tris pH 8, 300 mM NaCl. Cell resuspensions were spiked with protease inhibitor mix and then stored at -80 °C prior to purification in 40 mL aliquots. Full for BVES production methods are here: <https://zenodo.org/record/154611>

### **Purification round 1 – HDAC4/USP9X/HTATIP:**

Cell pastes were thawed and diluted to 250 mL with 50 mM Tris pH 8, 300 mM NaCl and supplemented with benzonase and 1 x protease inhibitors. NB: freeze-thaw cycle for cells is sufficient for lysis. Lysates were clarified by centrifugation at 15,000 rpm, 1 h, 4 °C (Beckman JLA16.2500) and then bound to 2 mL anti-FLAG resin (Sigma M2) at 4 °C with rocking for 2 hours (flow through – FT1). Lysate-resin mix was transferred to an open column (BioRad) and the resins then washed with 250 mL 50 mM Tris pH 8, 300 mM NaCl (wash – W1). HTT protein was eluted with ~5 mL resuspension buffer supplemented with 200 µg/mL 3xFLAG peptide (elution – E1). The sample was then rocked with 1 mL Ni-NTA at 4 °C with rocking for 30 mins (flow through – FT2). Ni-NTA beads were washed with 50 mM Tris pH 8, 300 mM NaCl then 50 mM Tris pH 8, 300 mM NaCl supplemented with 15 mM imidazole (wash – W2 and W3) and then eluted with 50 mM Tris pH 8, 300 mM NaCl, 300 mM imidazole (elution – E2). Samples were analysed by 4-20 % tris-glycine SDS-PAGE.

### **SDS-PAGE:**

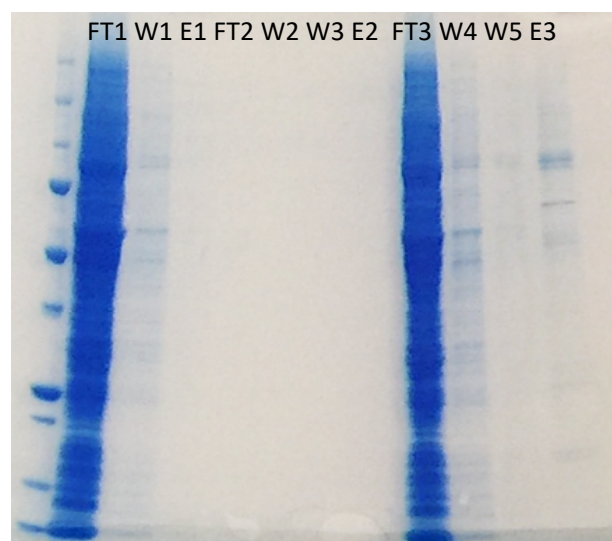


### **Purification round 2 – PRC2 3-mer/PRC2 5-mer/PRMT5-MEP50/USP9X:**

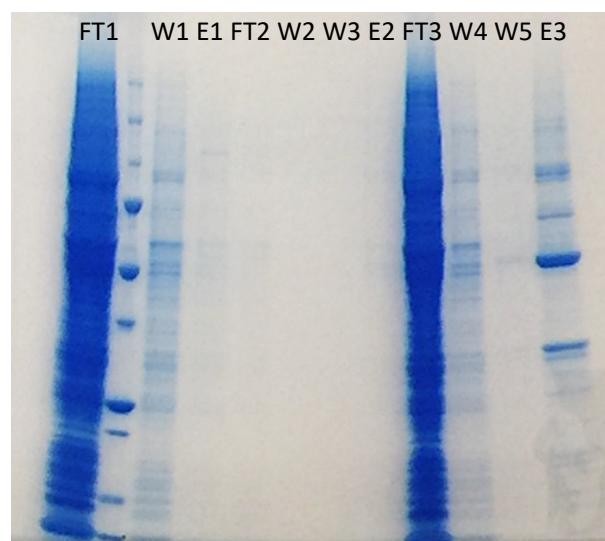
Cell pastes were thawed and diluted to 250 mL with 50 mM Tris pH 8, 300 mM NaCl and supplemented with benzonase and 1 x protease inhibitors. NB: freeze-thaw cycle for cells is sufficient for lysis. Lysates were clarified by centrifugation at 15,000 rpm, 1 h, 4 °C (Beckman JLA16.2500) and then bound to 2 mL anti-FLAG resin (Sigma M2) at 4 °C with rocking for 2 hours (flow through – FT1). Lysate-resin mix was transferred to an open column (BioRad) and the resins then washed with 250 mL

50 mM Tris pH 8, 300 mM NaCl (wash – W1). HTT protein was eluted with ~5 mL resuspension buffer supplemented with 200 µg/mL 3xFLAG peptide (elution – E1). The sample was then rocked with 1 mL Talon at 4 °C with rocking for 30 mins (flow through – FT2). Ni-NTA beads were washed with 50 mM Tris pH 8, 300 mM NaCl then 50 mM Tris pH 8, 300 mM NaCl supplemented with 5 mM imidazole (wash – W2 and W3) and then eluted with 50 mM Tris pH 8, 300 mM NaCl, 300 mM imidazole (elution – E2). Samples were analysed by 4-20 % tris-glycine SDS-PAGE. FT1 rocked with 1 mL Talon at 4 °C with rocking for 30 mins (flow through – FT3). Ni-NTA beads were washed with 50 mM Tris pH 8, 300 mM NaCl then 50 mM Tris pH 8, 300 mM NaCl supplemented with 5 mM imidazole (wash – W4 and W5) and then eluted with 50 mM Tris pH 8, 300 mM NaCl, 300 mM imidazole (elution – E3).

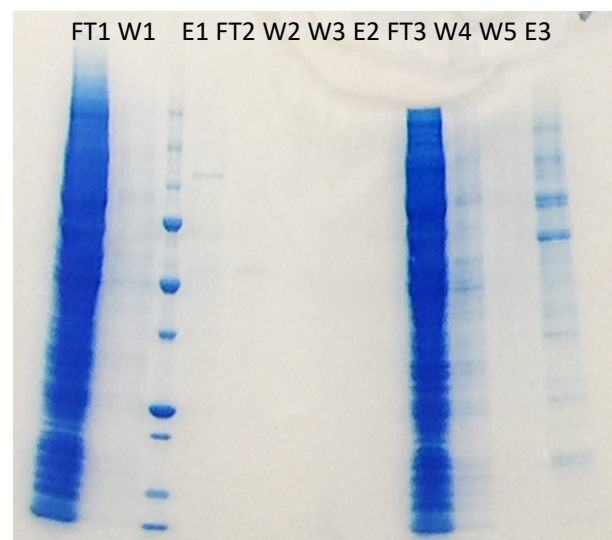
**SDS-PAGE:**



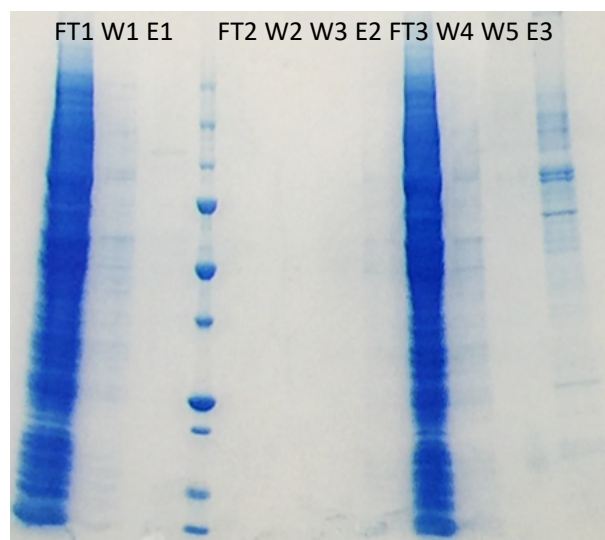
HTT + PRC2 trimer



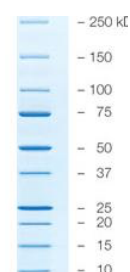
HTT + PRC2 pentamer



HTT + PRMT5



HTT + ELP1



**Comments:**

- HTT-HAP40 complex can be readily purified in FLAG affinity chromatography, Ni affinity chromatography, gel filtration 3-step purification protocol. Therefore same protocol was used to explore other putative HTT interaction partners
- Following run #1 with putative binders – fix by using FT1 to bind to Talon to validate expression of binding partner in subsequent pull-down experiments as not visible in other fractions on SDS-PAGE
- Second run of HTT co-expression purifications – cannot see clear band for HTT (~350 kDa) in FLAG elution so initial pull-down has not worked
- How to improve in future experiments
  - New FLAG beads – current beads have been used to purify more than 200 L of BVES production
  - Run small-scale first using 3 mL TestX protocol as per BVES production methods are here: <https://zenodo.org/record/154611>
  - Plans for 3 mL production of different combos plus controls in 24-well block:

	1	2	3	4	5	6
A	TOC009-D01 + TOC011-C01, 3 to 1 ratio	TOC009-D01 + TOC011-C01, 3 to 1 ratio	TOC009-D01 + (JMC023-T-A04 + JMC023-T-B06), 3 to 1 ratio (HTT:PRC2)	TOC009-D01 + (JMC023-T-A04 + JMC023-T-B06), 3 to 1 ratio (HTT:PRC2)	TOC011-C01	JMC023-T-A04 + JMC023-T-B06
B	TOC009-D01 + JMC110-A07, 3 to 1 ratio	TOC009-D01 + JMC110-A07, 3 to 1 ratio	TOC009-D01 + (JMC023-T-A04 + JMC023-T-B06 + MVC001-B02 + APC062-D03), 3 to 1 ratio (HTT:PRC2)	TOC009-D01 + (JMC023-T-A04 + JMC023-T-B06 + MVC001-B02 + APC062-D03), 3 to 1 ratio (HTT:PRC2)	JMC110-A07	JMC023-T-A04 + JMC023-T-B06 + MVC001-B02 + APC062-D03
C	TOC009-D01 + (JMC046:A09 + JMC015:F03), 3 to 1 ratio (HTT:PRMT5)	TOC009-D01 + (JMC046:A09 + JMC015:F03), 3 to 1 ratio (HTT:PRMT5)	TOC009-D01 + JMC035-C03, 3 to 1 ratio	TOC009-D01 + JMC035-C03, 3 to 1 ratio	JMC046:A09 + JMC015:F03	JMC035-C03
D	TOC009-D01 + YTC039-G10, 3 to 1 ratio	TOC009-D01 + YTC039-G10, 3 to 1 ratio	TOC009-D01 + JMC118-F04, 3 to 1 ratio	TOC009-D01 + JMC118-F04, 3 to 1 ratio	YTC039-G10	JMC118-F04
	HTT + interactor in duplicate		HTT + interactor in duplicate		Interactors without HTT	