# Comparison of Free Radical Scavenging Capacities of Methanol Extract of Two Mint Species by Fixed Point and Kinetic Methods 

Shadia Sirry ${ }^{1, *}$ and Hind Babiker ${ }^{1,2}$<br>${ }^{1}$ Chemistry Department, Faculty of Science, Taibah University, P.O. Box 30002, Saudi Arabia<br>${ }^{2}$ Biochemistry and Molecular Genetic Department, Faculty of Science, Al-Neelain University, Sudan<br>*Author to whom correspondence should be addressed; smrry90@hotmail.com.edu.sa Tel.: +966502008141

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#### Abstract

Mentha Longifolia (ML) and Mentha Puleguim (MP) are usually used as a flavor and medicinal plants in Saudi Arabia. The present study was carried out to compare the potential of methanol extract of fresh leaves of ML and MP as free radical (DPPH) scavenger by fixed point and kinetic methods. In a fixed point method, free radical scavenging capacity was expressed by effective concentration, $\mathrm{EC}_{50}$. However, in kinetic method, capacity expressed by rate constant value. Some other antioxidants parameters like total flavonoids content and total phenolic content were also estimated. Flavonoid content was obtained as catechin equivalent and total phenolic content as gallic acid equivalent. The obtained results indicate that methanol extract of MP possess a high concentration of flavonoids ( $6.55 \pm 0.42 \mathrm{mg} / 100 \mathrm{ml}$ ) and of phenolic ( $3.72 \pm 0.29$ $\mathrm{mg} / 100 \mathrm{ml}$ ) compared to that of ML flavonoids ( $3.22 \pm 0.43 \mathrm{mg} / 100 \mathrm{ml}$ ) and of phenolic ( $1.91 \pm 0.06 \mathrm{mg} / 100 \mathrm{ml}$ ).


Keywords: Free radical scavenging capacity; Antioxidant, DPPH, Kinetic; Total phenolic content; Total flavonoids

## I. Introduction

Free radicals and other reactive species (oxidants) are produced in the body as a by-product of aerobic metabolism or as a result of environmental stress. It is characterized by the presence of unusually high concentrations of reactive species like superoxide, hydroxyl alkoxyl, hydroperoxyl, nitric oxide and nitrogen dioxide radicals [1]. Excessive in vivo production of these radicals induce oxidative damage in protein, lipids, and DNA, which leads to chain reaction and oxidative stress.

Dietary sources of antioxidants are secondary metabolites from plants, which are react with free radicals to terminate the chain reaction and prevent the oxidative stress. However, these secondary metabolites are not essential for basic growth and development of plants, but they are synthesized mainly to face biotic and abiotic stresses. Usually secondary metabolites that have potential effects as antioxidants include phenolic and flavonoid compounds.

The free radical scavenging capacity has been measured by several methods [2] among those DPPH (2,2-Diphenyil-picrylhydrazyl) assay is considered as a simple method to evaluate the antioxidant activity of both pure compound and plant extracts [3]. The free radical DPPH ${ }^{*}$ is a stable free radical with deep violet color and easily measured by UV-Vis spectrometry at 515 nm . When

DPPH ${ }^{\bullet}$ react with antioxidant, the deep violet color was bleached and the percentage of inhibition of color is determined as:

$$
\begin{equation*}
\% I=100 \frac{[D P P H]}{[D P P H]_{0}} \tag{1}
\end{equation*}
$$

$[\mathrm{DPPH}]_{\circ}$ : is the initial concentration, $[\mathrm{DPPH}]$ is the concentration of DPPH after reacting with antioxidant. DPPH assay usually carry out at fixed time and variable concentrations of antioxidant. Antioxidants capacity was measured as $\mathrm{IC}_{50}$, which means concentration of antioxidants necessary to inhibit DPPH concentration by $50 \%$. DPPH assay give an information about antioxidants capacity at definite time. Some researchers reported $\mathrm{IC}_{50}$ after 15 minutes incubation and others after 30 minutes [4]. However, the estimations of $\mathrm{IC}_{50}$ at fixed-point time does not take into consideration the effect of kinetic parameter [5]. Plant extracts may contain more than one type of antioxidants, which react with DPPH radical, some antioxidants are fast and others are slow. In order to overcome this problem some workers suggested that kinetic parameters is an important to determine the actual capacity of antioxidants [6-9]. The kinetic of the reaction between antioxidants and DPPH radical were reported as first order and other as second order reaction [9].

The Genus Mentha, belonging to the family Lamiaceae, contains many species. Mentha species are usually utilized as herbal tea in medicine, taste and aroma, folk remedy, raw material for pharmaceutical, cosmetic, perfume and as food industry [10]. Many studies on antioxidant activity of mint species had been performed and revised that the activity is mainly due to phenolic components, such as flavonoids [11-12], phenolic acids and phenolic terpenes [13]. Mentha Longifolia (ML) and Mentha Pulegum (MP), are usually utilized as flavoring in Saudi Arabia and their local names are AIMadina hasawy mint and Mugrabi mint, respectively [14-15]. The aim of this study is to compare the free radical (DPPH) scavenging capacities of fresh leaves methanol extract of MP and ML by fixed point and kinetic methods. Total phenolic content and total flavonoids were also determined to support the study.

## II. Experimental section <br> II.1. Apparatus:

A grinding machine, a shaker device (GFL), reduced pressure rotatory evaporator, (Buchi R 210), UV/Visible spectrophotometer (CINTRA 6 GBC)

## II.2. Materials

DPPH, gallic acid and absolute methanol were from Sigma Aldrich (USA). Folin-Ciocalteu phenol reagent was obtained from Merck (Darmstadt, Germany). Sodium carbonate, sodium nitrite and aluminum was purchased from chemical pure company.

## II.3. Preparation mint extracts

Fresh leaves of mint species were purchased from local market, washed and grounded. 2 g of the grounded mints were shaken with 100 ml methanol in mechanical shaker at 200 rpm for 2 hours. The extracts were kept at $5^{\circ} \mathrm{C}$ for 24 hours and evaporated under reduced pressure rotatory evaporator, (Buchi R 210)

## II.4. Free radical scavenging capacity

The ability of free radical scavenging of mint species was assayed by 1,1-Diphenyl-2picrylhydrazyl radicals (DPPH) by adding constant concentration of DPPH to different concentrations of mint extracts and measuring the degradation of color intensity of DPPH 3 (Brand-Wiliam et al.,
1995) at wavelength of maximum absorbance, $\lambda_{\text {max }}$.equal 516 nm by UV/Visible spectrophotometer (CINTRA 6 GBC)

## II.5. Kinetic of free radical scavenging

Kinetic of free radical scavenging ability of mint extracts was determined by mixing of DPPH ( 0.18 mmole ) and mint extracts ( $0.0634 \mathrm{mg} / \mathrm{mL}$ ) and measuring the absorbance at wavelength 516 nm at time interval 0-120 minutes. Remaining DPPH concentration was calculated from calibration curve of DPPH. kinetic studies were investigated three times at room temperature.

## II.6. Determination of Total Flavonoids

In 10 mL volumetric flask, 4 mL of water, 2 mL of mint extracts and 0.3 mL of $\mathrm{NaNO}_{2}(5 \%)$ were mixed. After 5 min 3 mL of $\mathrm{AICl}_{3}$ (10\%) was added then after 6 min 1 mL of $\mathrm{NaOH}(4 \%)$ was added. The mixture was completed to 10 mL by water and the absorbance was measured at 510 nm against blank with spectrophotometer. Catechin was used as standard and the calibration curve was obtained by the same manner of extract utilizing $0.2-1 \mathrm{~mL}$ catechin solution $(500 \mu \mathrm{~g} / \mathrm{mL})$ [16-17].

## II.7. Determination of total phenolic compound

Total phenolic contents in mint extracts were quantified by Folin-Ciocalteu reagent [18] as Gallic acid equivalent GAE.

## III. Results and Discussion

Methanol is polar solvent commonly used as extraction of flavonoids and other phenolic compounds. In this study, it was utilized as extractant for fresh ML, MP mint leaves (Fig.1) and the extract yields were calculated and given in table 1. It is clear that the yield of ML extract is slightly higher than MP.


Figure 1: Mentha longifolia (ML) and Mentha pulegium (MP) leaves

Table 1: Percent of yield of ML and MP extract

| type of mint | Weight,g | \%yeild |
| :---: | :--- | :--- |
| ML | 0.0862 | 4.31 |
| MP | 0.0792 | 3.96 |

## III.1. Free Radical Scavenging Capacity (DPPH assay): <br> III.1.1.Fixed point method

Radical scavenging ability is usually evaluated by the ability of antioxidant to scavenge DPPH ${ }^{\bullet}$ radical. The violet color of DPPH ${ }^{\bullet}$ is due to the transferring of free electron around its molecule. After reacting with antioxidant, it is change to yellow due to scavenging of free electron. Free radical scavenging ability are deduced spectrophotometrically by following the color inhiption of DPPH ${ }^{\bullet}$ at mximum absorbance wavelength $(\lambda \max =517 \mathrm{~nm})$. The percent of inhiption of $\mathrm{DPPH}^{\circ}, \%$, is given as:

$$
\begin{equation*}
\% I=\frac{\left(A_{\text {original }}-A_{\text {final }}\right)}{A_{\text {original }}} \tag{2}
\end{equation*}
$$

$\%$ I depend on concentration and time [3,19].The values of $\%$ I of constant concentration of DPPH $(0.08 \mathrm{M})$ as a function of concentration of mint extract at constant time ( 15 minutes) were shown at Fig 2. The amount of free radical inhibnition increased as the concentration of mint extract increased. The concentration of antioxidant that scavenge $50 \%$ of original DPPH radical called effective concentration ( $\mathrm{EC}_{50}$ ) and its characteristic to free radical scavenging ability for antioxidants. The lower value of $\mathrm{EC}_{50}$, the larger of scavenging capacity. $\mathrm{EC}_{50}$ values of ML and MP extracts are given at table 2 and compared with effective concentrations of some standards which were reported previously.


Figure 2: Effect of effective concentration for DPPH radical scavenging

Table 2: Comparison of Efficient concentration of ML, MP extract with previous studies of ML, MP and standards

| Type of mint | $\mathrm{Ec}_{50}, \mathrm{mg} / \mathrm{ml}$ | Reference |
| :--- | :--- | :--- |
| ML | 0.085 | Present study |
| MP | 0.034 | Present study |
| ML | 0.057 | $[20]$ |
| Mentha spicata | 0.021 | $[21]$ |
| ML | 0.086 | $[22]$ |
| Gallic acid | 0.001 | $[23]$ |
| Catechin | 0.007 | $[24]$ |

The lower value of $\mathrm{EC}_{50}$ of MP extract $(0.034 \mathrm{mg} / \mathrm{mL})$ with respect to ML extract $(0.085 \mathrm{mg} / \mathrm{mL})$ prove that MP extract is effective as radical scavenging than ML. The values of $\mathrm{IC}_{50}$ of ML in the present study was cocordant with ML of Özgen et al. [22].

## III.1.2. Kinetic method

The reaction between antioxidants and free radical is time dependent and reached a steady state after a period [9].Thus, reaction kinetics between antioxidant and DPPH radical give more accurate results about the impact of antioxidants than fixed-point antioxidant capacity.

To study rate of free radical scavenging of two mint methanol extracts, a constant concentration of mint extracts $(0.0634 \mathrm{mg} / \mathrm{mL})$ and DPPH $(0.182 \mathrm{mmole} / \mathrm{L})$ were mixed and the reaction was followed spectrophotometrically at $\lambda_{\max } 517 \mathrm{~nm}$ from 0 minute to 150 minutes (Fig.3)


Figure 3: Rate of free radical scavenging of ML and MP mint extracts
From figure, it was obvious that the rate of both extracts as free radical scavenging have two stages initial fast then slow. The reaction of antioxidant in mint and DPPH radical may occur according to two independent parallel mechanism: Hydrogen atom transfer, HAT (fast stage) and sequential proton loss electron transfer, SPLET, (slow stage) [5,7]. However according to the time of reaching to steady state, the kinetic reaction between antioxidants and DPPH has been classified as fast ( $<5 \mathrm{~min}$ ), intermediate ( $5-30 \mathrm{~min}$ ) and slow ( $>30 \mathrm{~min}[3,19]$. The steady state of ML and MP were $>120 \mathrm{~min}$ therefore, the reaction between mint extract antioxidants and DPPH is classified as slow. The rate of MP extract as free radical scavenging more than rate of ML extract. Order of the reaction between DPPH and mint extracts gives an information about the rate constants of the reaction and number of active molecules take part. The scavenging kinetic of DPPH radicals by antioxidants may follow second order or pseudo first order according to the ratio of amount of antioxidant: DPPH [2425].The order of the reaction are obtained by fitting the experimental data to both first and second order kinetic equations.

The reaction between DPPH radical and antioxidants in mint extracts is represented by:

$$
\begin{equation*}
A+D P P H \rightarrow P \tag{3}
\end{equation*}
$$

A: Antioxidant in mint extracts $P$ : product
This reaction may fit either first or/and second order kinetic models:
Frist order kinetic model:

$$
\begin{equation*}
\frac{-\mathrm{d}[\mathrm{DPPH}]}{\mathrm{dt}}=k_{1}[D P P H] \tag{4}
\end{equation*}
$$

Second order kinetic model:

$$
\begin{equation*}
\frac{-\mathrm{d}[\mathrm{DPPH}]}{\mathrm{dt}}=k_{2}[\mathrm{DPPH}][A] \tag{5}
\end{equation*}
$$

Assuming the presence of same amount of A and DPPH

$$
\begin{equation*}
\frac{-\mathrm{d}[\mathrm{DPPH}]}{\mathrm{dt}}=k_{2}[\mathrm{DPPH}]^{2} \tag{6}
\end{equation*}
$$

Integration form of the equations 4 and 6 are

$$
\begin{gather*}
\ln [D P P H]=\ln [D P P H]_{0}-k t  \tag{7}\\
\frac{1}{[D P P H]}-\frac{1}{[D P P H]_{0}}=K_{2} \tag{8}
\end{gather*}
$$

$\mathrm{K}_{1}$ and $\mathrm{K}_{2}$ are the rate constants for first and second order, respectively
[DPPH] ${ }_{0}$ is the original concentration of DPPH and [DPPH] is the concentration at time $t$ and determined from the calibration curve of DPPH, Fig. 4


Figure 4: Calibration Curve of DPPH
Equation 7 and 8 were applied to explore the fitness of $\mathrm{DPPH}^{\bullet}$ scavenging by ML and MP mint extracts to first or second order (Fig. 5 and 6).


Figure 5: First order kinetic equation of free radical scavenging of ML and MP mint extracts


Figure 6: Second order kinetic equation of free radical scavenging of ML and MP mint extracts
From the results it was investigated that both first and second order kinetic equations were good fitted with linear regression constant $R^{2}$ more than $98 \%$. The rate constants for first and second order equations ( $\mathrm{K}_{1}$ and $\mathrm{K}_{2}$ ) are obtained from the slopes data and given at table (6). The applicability of both first and second order reaction may be due to the presence of two groups of antioxidants, fast and slow antioxidants. A previous studies [26-27] reveals that various mentha species contain antioxidants like phenolic (rosmarinic acid, caffeic and luteolin derivatives) as major antioxidants and ascorbic and carotenoids as minor antioxidants. According to Brand William et al., [3, 28] ascorbic acid is a rapid antioxidant, rosmarinic acid is classified as intermediate antioxidant and caffeic acid compounds as slow antioxidants. The overall rate of interaction between the same amounts of plant extracts and DPPH were used as a measure of antioxidants capacity of mint species. The rate constant values, $\mathrm{k}_{1}$ and $\mathrm{k}_{2}$, of MP extract more than ML, thereby, the antioxidants ability of methanol extracts of MP more than ML mint.

Table 3: Kinetic Constants of ML and MP

|  | $M L$ | $M P$ |
| :--- | :--- | :--- |
| $\mathrm{~K}_{1} \mathrm{~min}^{-1}$ | 0.0014 | 0.0095 |
| $\mathrm{R}^{2}$ | 0.986 | 0.984 |
| $\mathrm{~K}_{2}(\mathrm{mg} / \mathrm{ml})^{-1} \mathrm{~min}^{-1}$ | 0.0079 | 0.1035 |
| $\mathrm{R}^{2}$ | 0.985 | 0.993 |

## III.2. Antioxidant characteristics

The antioxidant characteristics of methanolic extraction of fresh MP and ML leaves was achieved by total flavonoids and total phenolic.

## III.2.1. Determination of Total Flavonoid Content

Flavonoids are considered as antioxidants by neutralizing hydroxyl and superoxide radicals and by chelation. Several investigator have established that Mentha species contains wide range of flavonoids [29-31]. Flavonoids in mint extracts was determined according to catechin content The calibration curve of catechin was performed and given in Fig.7. Total flavonoids of methanolic extracts of ML and MP were determined spectrophotometrically as standard catechin content (CE) in $\mathrm{mg} / 100 \mathrm{~mL}$ extract (Table 4)

MP extract have higher flavonoid content than ML extract. The same trend was observed in a previous work [32] and opposite observation for the result was reported by [33].


Figure 7: Standard curve of catechin

Table 4: Flavonoids content of methanolic extract ML and MP as catechin equivalent CE (mg/100mL)

| type of mint | A | CE, <br> mg/100ml <br> extract | Average <br> (standard deviation) |
| :---: | :---: | :---: | :---: |
|  | 0.293 | 3.212 |  |
| ML | 0.270 | 2.799 |  |
|  | 0.318 | 3.662 |  |
|  | 0.454 | 6.107 | $6.557 \pm 0.426$ |
| MP | 0.501 | 6.953 |  |
|  | 0.482 | 6.612 |  |

## III.2.2. Determination of total phenolic content, TPC

Phenolic compounds are the most important compounds in mint species as account of their relation as antioxidants. Total phenolic content was estimated spectrophotometrically as gallic acid equivalent (GAE) by Folin reagent at 760 nm . (Fig.8), and Table 5 .Total phenolic content in MP is more than that of ML. Similar trend was obtained in previous study of same type of Mentha species [34].


Fig. 8 Standard curve of gallic acid

Figure 7: Standard curve of Gallic acid

Table 5: Determination of total phenolic as miligram gallic acid per 100 mL extract ( $\mu \mathrm{g}$ GAE/ml).

| type of <br> mint | A | TPC, <br> $\mathrm{mg} / 100 \mathrm{ml}$ extract | Average <br> (standard deviation) |
| :--- | :--- | :---: | :---: |
|  | 0.194 | 1.986 | $1.914 \pm 0.062$ |
| ML | 0.187 | 1.886 |  |
|  | 0.186 | 1.871 |  |
|  | 0.197 | 4.057 | $3.724 \pm 0.292$ |
| MP | 0.181 | 3.600 |  |
|  | 0.178 | 3.514 |  |

## IV. Conclusion

This study is a comparative study of free radical scavenging capacity of mentha Longifolia, ML and Mentha Puleguim extract by fixed point and kinetic method. In Both methods, the free radical scavenging capacity of methanol extract of MP was found to be higher than that of ML.MP extract also exhibited a higher concentration of total phenol, and total flavonoids than that of ML extract.

## V. References

[1] Fang, Yun-Zhong, Sheng Yang, and Guoyao Wu. "Free radicals, antioxidants, and nutrition." Nutrition 18.10 (2002): 872-879.
[2] Apak, Reşat, et al. "Methods of measurement and evaluation of natural antioxidant capacity/activity (IUPAC Technical Report)." Pure and Applied Chemistry 85.5 (2013): 957-998.
[3] Brand-Williams, Wendy, Marie-Elisabeth Cuvelier, and C. L. W. T. Berset. "Use of a free radical method to evaluate antioxidant activity." LWT-Food science and Technology 28.1 (1995): 25-30.
[4] Fadda, Angela, et al. "Reaction time and DPPH concentration influence antioxidant activity and kinetic parameters of bioactive molecules and plant extracts in the reaction with the DPPH radical." Journal of Food Composition and Analysis 35.2 (2014): 112-119.
[5] Bondet, V., W. Brand-Williams, and C. Berset. "Kinetics and mechanisms of antioxidant activity using the DPPH. free radical method." LWT-Food Science and Technology 30.6 (1997): 609-615.
[6] [6] Li, Yan-feng, Pei-ze Li, and Zai-qun Liu. "Novel kinetic method for expressing the ability of antioxidant to scavenge radicals." Chemical Research in Chinese Universities 29.5 (2013): 947-951.
[7] Volkov, V. A., N. A. Dorofeeva, and P. M. Pakhomov. "Kinetic method for studying the antiradical activity of medicinal plant extracts." Pharmaceutical chemistry journal 43.6 (2009): 333.
[8] Haddadi, H., N. Alizadeh, and M. Shamsipur. "Stoichiometric and free radical-scavenging kinetic studies of extractable polyphenols from pomegranate husk and pistachio hull." Journal of the Iranian Chemical Society 8.3 (2011): 694-707.
[9] Anissi, Jaouad, et al. "A comparative study of the antioxidant scavenging activity of green tea, black tea and coffee extracts: a kinetic approach." Food chemistry 150 (2014): 438-447.
[10] Mimica-Dukic, N., and B. Bozin. "Mentha L. species (Lamiaceae) as promising sources of bioactive secondary metabolites." Current Pharmaceutical Design14.29 (2008): 3141-3150.
[11] Kähkönen, Marja P., et al. "Antioxidant activity of plant extracts containing phenolic compounds." Journal of agricultural and food chemistry 47.10 (1999): 3954-3962.
[12] Riachi, Liza G., and Carlos AB De Maria. "Peppermint antioxidants revisited." Food chemistry 176 (2015): 72-81.
[13] Shahidi, Fereidoon, P. K. Janitha, and P. D. Wanasundara. "Phenolic antioxidants." Critical reviews in food science \& nutrition 32.1 (1992): 67-103.
[14] Osman, I. H. "In vitro antioxidant activity of Mentha pulegium from Saudi Arabia." Bioscience Research 10.1 (2013): 33-37.
[15] Ahmed, Ahmed M., Hani A. Ozbak, and Hassan A. Hemeg. "Effect of essential oil of traditional two Saudi mint types and its possible role in cardiovascular and throat health." International journal of clinical and experimental medicine 8.5 (2015): 8060.
[16] Zhuang, X. P., Y. Y. Lu, and G. S. Yang. "Extraction and determination of flavonoid in ginkgo." Chinese Herbal Medicine 23 (1992): 122-124.
[17] Zhishen, Jia, Tang Mengcheng, and Wu Jianming. "The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals." Food chemistry 64.4 (1999): 555-559.
[18] Singleton, Vernon L., Rudolf Orthofer, and Rosa M. Lamuela-Raventós. "[14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent." Methods in enzymology 299 (1999): 152-178.
[19] Sánchez-Moreno, Concepción, Jose A. Larrauri, and Fulgencio Saura-Calixto. "A procedure to measure the antiradical efficiency of polyphenols." Journal of the Science of Food and Agriculture 76.2 (1998): 270-276.
[20] Gulluce, M., et al. "Antimicrobial and antioxidant properties of the essential oils and methanol extract from Mentha longifolia L. ssp. Iongifolia." Food chemistry103.4 (2007): 1449-1456.
[21] Scherer, Rodrigo, et al. "Antioxidant and antibacterial activities and composition of Brazilian spearmint (Mentha spicata L.)." Industrial crops and products 50 (2013): 408-413.
[22] Özgen, U., et al. "Antioxidant properties of some medicinal Lamiaceae (Labiatae) species." Pharmaceutical biology 44.2 (2006): 107-112.
[23] Scherer, Rodrigo, and Helena Teixeira Godoy. "Antioxidant activity index (AAI) by the 2, 2-diphenyl-1picrylhydrazyl method." Food chemistry 112.3 (2009): 654-658.
[24] Espín, Juan Carlos, et al. "Anthocyanin-based natural colorants: a new source of antiradical activity for foodstuff." Journal of Agricultural and Food Chemistry48.5 (2000): 1588-1592.
[25] Suja, Kizhiyedathu Polachira, Anathasankaran Jayalekshmy, and Chami Arumughan. "Free radical scavenging behavior of antioxidant compounds of sesame (Sesamum indicum L.) in DPPH• system." Journal of Agricultural and Food Chemistry 52.4 (2004): 912-915.
[26] Brahmi, Fatiha, et al. "Chemical Composition and Biological Activities of Mentha Species." Aromatic and Medicinal Plants-Back to Nature. InTech, 2017.
[27] National Toxicology Program. "Toxicology and carcinogenesis studies of pulegone (CAS No. 89-82-7) in F344/N rats and B6C3F1 mice (gavage studies)." National Toxicology Program technical report series 563 (2011): 1.
[28] Savatović, Slađana M., et al. "Kinetic behaviour of DPPH radical scavenging activity of tomato waste extracts." Journal of the Serbian Chemical Society77.10 (2012): 1381-1389.
[29] Al-Jaber, Nabilah A., Amani S. Awaad, and John E. Moses. "Review on some antioxidant plants growing in Arab world." Journal of Saudi Chemical Society15.4 (2011): 293-307.
[30] Justesen, Ulla, and Pia Knuthsen. "Composition of flavonoids in fresh herbs and calculation of flavonoid intake by use of herbs in traditional Danish dishes." Food chemistry 73.2 (2001): 245-250.
[31] Riahi, Leila, et al. "Phytochemistry, antioxidant and antimicrobial activities of the essential oils of Mentha rotundifolia L. in Tunisia." Industrial crops and products49 (2013): 883-889.
[32] Fialová, S., D. Tekelová, and D. Grančai. "The content of phenolic compounds in underground and aerial parts of different Mentha species." Acta Facultatis Pharmaceuticae Universitatis Comenianae 59.1 (2012): 30-38.
[33] Hajlaoui, Hafedh, et al. "Biological activities of the essential oils and methanol extract of tow cultivated mint species (Mentha longifolia and Mentha pulegium) used in the Tunisian folkloric medicine." World Journal of Microbiology and Biotechnology 25.12 (2009): 2227-2238.

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