#### Sugar-sensing swodkoreceptors

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#### Abstract

Understanding how cells sense sugar is a fundamental question in biology and is pivotal for the evolution of life. In numerous organisms, sugar molecules constitute a primary source of energy generation. Consequently, the mechanisms governing sugar sensation in various microorganisms and animals have been experimentally elucidated. However, sugar sensation has primarily been investigated in specialized sensory cells such as taste buds, taste organs, or sensory neurons. These cells detect extracellular sugar through membranebound 'sugar or sweet receptors' or 'gustatory receptors'. However, in addition to these membrane receptors, sugar molecules can also be sensed via other sugar-binding nonmembrane signaling proteins. Utilizing a supermarket employee analogy, I present my rationale for why glucokinase may not be the optimal glucose sensor. Additionally, to encapsulate all sugar-sensing proteins capable of detecting and signaling irrespective of their cellular location, I propose the term 'swodkoreceptor', derived from the Polish word 'Słodkie' meaning 'Sweet'. This proposal aims to facilitate the exploration of the identity and function of all swodkoreceptors for all the sugar molecules, akin to research that identified the bacterial Lac repressor as the allolactose swodkoreceptor and the Liver X receptor as one of the mammalian glucose swodkoreceptor.

2 | Anbalagan, Savani. Sugar-sensing swodkoreceptors. Zenodo. 2024. doi: <u>10.5281/zenodo.10938497</u>; Submission date: 07-04-2024 Due to the priority in human interests, the glycolytic pathway (glucose metabolism) was initially a focus of early biochemical research due to its significance in the production of alcoholic beverages and its commercial value.<sup>1,2</sup> Nevertheless, since then, the mechanisms underlying sugar synthesis, transport, storage, metabolism and homeostasis has been extensively studied due to its role in energy production, diseases such as diabetes, and the development of real-time biosensors for measuring glucose levels in patients.<sup>3-6</sup>

Glycolysis has been proposed to have its origin in thermophilic archaea.<sup>7-9</sup> Both eubacteria and archaea possess the enzymatic ability to synthesize trehalose, with the evolutionary purpose of these molecules largely attributed to stress response mechanisms.<sup>10</sup> Trehalose serves as a virulence factor for bacteria and fungi in colonizing plant and animal cells, with its synthesis triggered by various stressors.<sup>11-14</sup> Nonetheless, owing to the significance of sugar, the identification of gustatory sugar receptors responsible for sugar sensing has been addressed in numerous organisms.<sup>15-17</sup> The trehalose-sensing gustatory receptor in Drosophila was identified as the G protein-coupled receptor Gr5a, whereas Gr5a, Gr64a, and Gr64f are among the gustatory receptors identified for other sugars such as sucrose and maltose.<sup>18-20</sup> In silkworm *Bombyx mori*, D-fructose is sensed by an ionotropic gustatory receptor known as Gr9, while in Drosophila, it is sensed by the ionotropic gustatory receptor Gr43a.<sup>21</sup> Drosophila Gr64a has also been identified as a receptor for D-fructose in the brain, enabling the sensing of D-fructose in the hemolymph.<sup>22</sup> In humans and mice, taste-specific G protein-coupled receptors T1R2/T1R3 have been identified and reported as the "only sweet taste receptor".<sup>23-25</sup> However, in fish, while grass carp T1R2s can sense sugar, zebrafish and medaka rely largely on T1R1/3 and T1R2/3 for amino acid sensing.<sup>26-28</sup> In mice, while glucose sensing was initially attributed to POMC (pro-opiomelanocortin) neurons, it has been reported that adhesion G-protein coupled receptor 1 (ADGRL1) functions as the glucose-sensing receptor in the ventromedial nucleus of the hypothalamus.<sup>29,29-31</sup>

All the aforementioned studies on gustatory sweet taste receptors focus on receptors expressed in sensory cells. However, the sensing of sugar is not confined solely to sensory cells or at the plasma membrane level. This capacity offers a specific advantage to the organism or cell, enabling it to detect sugar in its environment or blood or hemolymph and respond accordingly. The mechanism by which mammalian cells sense sugar molecules within their intracellular environment remains largely unexplored. To illustrate, consider the analogy of the month preceding Christmas Eve. The number of trucks loaded with candies waiting outside a supermarket doesn't determine the availability of candies for sale inside.

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What truly matters is the quantity of candies already inside the supermarket, ready to be sold to customers who are shopping within. Even candies waiting to be unwrapped from their delivery boxes are not sufficient. If customers cannot find readily available candies, they may choose to shop elsewhere or request assistance from salespersons to open the boxes. In more extreme cases, impatient customers, such as parents with highly demanding young children, may even take matters into their own hands and open the wrapped delivery boxes themselves.

Moreover, some animals, such as domestic cats, cannot even sense sweetness due to pseudogenization of one of the gustatory sweet receptors.<sup>32</sup> Does this imply that all sugar sensing in domestic cats occurs solely in the pancreas and the brain? However, even the identity of glucose-sensing receptors in the pancreas is subject to debate, as glucokinase has been suggested as the 'glucose sensor'.<sup>33-36</sup> Returning to our analogy, if one employee in the supermarket must serve as a 'candy sensor' or 'candy receptor', their role would be solely to monitor candy stock levels and take action if stock is low, either by receiving candy from trucks outside, unwrapping delivery crates or signaling to other employees to act. In contrast, the function of glucokinase is to catalyze the first step in glucose metabolism.<sup>2,37</sup> The supermarket employee's responsibility would be limited to checking candy stocks, without involvement in labeling candies for disposal or engaging in unrelated tasks like playing 'candy crush' during peak demand periods such as Christmas.

Since the majority of animal cells utilize glucose for energy production, or other microorganisms may employ sugar molecules like trehalose for stress response, it's logical to assume that cells and microorganisms possess mechanisms to sense sugar within their cytoplasm or organelles where such sugar molecules can be found. This raises the question of whether there are other sugar-sensing receptors and if they have already been identified. A promising candidate for such a sugar-sensing receptor is the nuclear receptor Liver X receptor (LXR), a transcription factor well-known for its role as an oxysterol-sensing.<sup>38,39</sup> In addition to its role in regulating genes related to cholesterol and fatty acid metabolism through oxysterol binding, LXR has also been reported to function as a glucose sensor. The binding of D-glucose and D-glucose-6-phosphate at physiological concentrations can stimulate the transcriptional activity of both LXR-α and LXR-β.<sup>40</sup> In my view, besides the term 'glucose sensor' used by the authors to describe LXR, alternative terms such as 'glucose-sensing receptor,' 'glucoreceptor,' 'sweet-sensing receptor,' or 'sweet receptor' could also be applicable to LXR. However, it may not be suitable to refer to LXR as a 'sweet taste

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receptor' or a 'gustatory receptor,' as these terms typically denote receptors specifically involved in taste perception rather than intracellular sensing of sugars.

It seems there's a lack of unity in the sugar-sensing scientific literature regarding the terms 'sweet receptor', 'sweet taste receptor', and 'sugar receptor', which are often used interchangeably. Additionally, there's a division between the use of the terms 'sensor' and 'receptor.' In my recent proposal of gasoreceptors, I have emphasized the challenges associated with this lack of consistency, drawing parallels to the blind men and the elephant parable.<sup>41-43</sup> It appears that in the context of estrogen- or vitamin D-sensing transcription factors, they are commonly referred to as 'receptors'. However, when it comes to gases like oxygen or sugars such as glucose, the term 'sensor' is often utilized.<sup>38,44</sup> Therefore, to consolidate all sugar-sensing proteins under a unified term, I propose the term "swodkoreceptor" (derived from "Słodkie," meaning "Sweet" in Polish). Embracing the concept of swodkoreceptor will facilitate the identification of receptors akin to LXR in every microorganism and within every cell of organisms.

The term 'swodkoreceptor' will encompass all subclasses of sugar/sweet/gustatory receptors, including trehalose receptors, fructose receptors, mannitol receptors, sucrose receptors, galactose receptors, lactose receptors, maltose receptors, and so forth. Identifying the roles and identities of swodkoreceptors may provide insights into the etiology of diseases related to diabetes and/or immune systems. The bacterial lac repressor (Lacl) is simply a swodkoreceptor for allolactose.<sup>45</sup> Similarly, just as β-galactosidase is necessary for the conversion of lactose to allolactose (via transgalactosylation), which is then sensed by the Lacl swodkoreceptor, the activity of glucokinase in glucose conversion might also fulfill a comparable role.<sup>46</sup> Therefore, the role of glucokinase in glucose sensing may simply be upstream to swodkoreceptors such as LXR, which have the capability to sense glucose-6-phosphate.<sup>40,47</sup>

Finally, based on their capacity for glucose-binding and potential additional roles, putative glucose swodkoreceptors may encompass proteins such as Metalloprotease TRABD2B (TraB Domain Containing 2B/TIKI2) and transcription factors like ZNF737 (Zinc Finger Protein 737).<sup>48</sup>

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## **AUTHOR CONTRIBUTIONS**

Savani Anbalagan: conceptualization, writing of the original draft, and review and editing.

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# **CONFLICT OF INTEREST**

None.

## DISCLOSURE

The author used ChatGPT for correcting the scientific English. The author takes full responsibility for the content of this manuscript.

#### REFERENCES

- 1. Grüning, N.-M., and Ralser, M. (2021). Glycolysis: How a 300yr long research journey that started with the desire to improve alcoholic beverages kept revolutionizing biochemistry. Current Opinion in Systems Biology *28*, 100380. 10.1016/j.coisb.2021.100380.
- 2. Kresge, N., Simoni, R.D., and Hill, R.L. (2005). Otto Fritz Meyerhof and the Elucidation of the Glycolytic Pathway. Journal of Biological Chemistry *280*, e3. 10.1016/S0021-9258(20)76366-0.
- 3. Grmek, M.D. (1968). First Steps in Claude Bernard's Discovery of the Glycogenic Function of the Liver. Journal of the History of Biology *1*, 141–154.
- 4. Olmsted, J.M.D. (1953). Claude Bernard, 1813-1878: A Pioneer in the Study of Carbohydrate Metabolism. Diabetes *2*, 162–164. 10.2337/diab.2.2.162.
- 5. Miyamoto, T., and Amrein, H. (2017). Gluconeogenesis: An ancient biochemical pathway with a new twist. Fly (Austin) *11*, 218–223. 10.1080/19336934.2017.1283081.
- Jia, B., Yuan, D.P., Lan, W.J., Xuan, Y.H., and Jeon, C.O. (2019). New insight into the classification and evolution of glucose transporters in the Metazoa. FASEB J *33*, 7519–7528. 10.1096/fj.201802617R.
- 7. Potter, S., and Fothergill-Gilmore, L.A. (1993). Molecular evolution: The origin of glycolysis. Biochemical Education *21*, 45–48. 10.1016/0307-4412(93)90018-U.
- 8. Bräsen, C., Esser, D., Rauch, B., and Siebers, B. (2014). Carbohydrate Metabolism in Archaea: Current Insights into Unusual Enzymes and Pathways and Their Regulation. Microbiol Mol Biol Rev *78*, 89–175. 10.1128/MMBR.00041-13.
- 9. Verhees, C.H., Kengen, S.W.M., Tuininga, J.E., Schut, G.J., Adams, M.W.W., De Vos, W.M., and Van Der Oost, J. (2003). The unique features of glycolytic pathways in Archaea. Biochem J *375*, 231–246. 10.1042/BJ20021472.
- 10. Avonce, N., Mendoza-Vargas, A., Morett, E., and Iturriaga, G. (2006). Insights on the evolution of trehalose biosynthesis. BMC Evolutionary Biology *6*, 109. 10.1186/1471-2148-6-109.
- 11. Vanaporn, M., and Titball, R.W. Trehalose and bacterial virulence. Virulence *11*, 1192–1202. 10.1080/21505594.2020.1809326.
- 12. McBride, M.J., and Ensign, J.C. (1987). Effects of intracellular trehalose content on Streptomyces griseus spores. J Bacteriol *169*, 4995–5001. 10.1128/jb.169.11.4995-5001.1987.
- 13. Pilonieta, M.C., Nagy, T.A., Jorgensen, D.R., and Detweiler, C.S. (2012). A glycine betaine importer limits Salmonella stress resistance and tissue colonization by reducing trehalose production. Mol Microbiol *84*, 296–309. 10.1111/j.1365-2958.2012.08022.x.
- Foster, A.J., Jenkinson, J.M., and Talbot, N.J. (2003). Trehalose synthesis and metabolism are required at different stages of plant infection by Magnaporthe grisea. EMBO J 22, 225–235. 10.1093/emboj/cdg018.
- 15. Kent, L.B., and Robertson, H.M. (2009). Evolution of the sugar receptors in insects. BMC Evol Biol *9*, 41. 10.1186/1471-2148-9-41.
- 7 | Anbalagan, Savani. Sugar-sensing swodkoreceptors. Zenodo. 2024. doi: <u>10.5281/zenodo.10938497</u>; Submission date: 07-04-2024

- 16. Shi, P., and Zhang, J. (2006). Contrasting modes of evolution between vertebrate sweet/umami receptor genes and bitter receptor genes. Mol Biol Evol *23*, 292–300. 10.1093/molbev/msj028.
- 17. Temussi, P. (2006). The history of sweet taste: not exactly a piece of cake. J Mol Recognit *19*, 188–199. 10.1002/jmr.767.
- Chyb, S., Dahanukar, A., Wickens, A., and Carlson, J.R. (2003). Drosophila Gr5a encodes a taste receptor tuned to trehalose. Proc Natl Acad Sci U S A *100 Suppl 2*, 14526–14530. 10.1073/pnas.2135339100.
- 19. Dahanukar, A., Lei, Y.-T., Kwon, J.Y., and Carlson, J.R. (2007). Two Gr genes underlie sugar reception in Drosophila. Neuron *56*, 503–516. 10.1016/j.neuron.2007.10.024.
- Jiao, Y., Moon, S.J., Wang, X., Ren, Q., and Montell, C. (2008). Gr64f is required in combination with other gustatory receptors for sugar detection in Drosophila. Curr Biol *18*, 1797–1801. 10.1016/j.cub.2008.10.009.
- 21. Sato, K., Tanaka, K., and Touhara, K. (2011). Sugar-regulated cation channel formed by an insect gustatory receptor. Proc Natl Acad Sci U S A *108*, 11680–11685. 10.1073/pnas.1019622108.
- 22. Miyamoto, T., Slone, J., Song, X., and Amrein, H. (2012). A Fructose Receptor Functions as a Nutrient Sensor in the Drosophila Brain. Cell *151*, 1113–1125. 10.1016/j.cell.2012.10.024.
- 23. Hoon, M.A., Adler, E., Lindemeier, J., Battey, J.F., Ryba, N.J., and Zuker, C.S. (1999). Putative mammalian taste receptors: a class of taste-specific GPCRs with distinct topographic selectivity. Cell *96*, 541–551. 10.1016/s0092-8674(00)80658-3.
- 24. Li, X., Staszewski, L., Xu, H., Durick, K., Zoller, M., and Adler, E. (2002). Human receptors for sweet and umami taste. Proc Natl Acad Sci U S A *99*, 4692–4696. 10.1073/pnas.072090199.
- 25. Nelson, G., Hoon, M.A., Chandrashekar, J., Zhang, Y., Ryba, N.J., and Zuker, C.S. (2001). Mammalian sweet taste receptors. Cell *106*, 381–390. 10.1016/s0092-8674(01)00451-2.
- Oike, H., Nagai, T., Furuyama, A., Okada, S., Aihara, Y., Ishimaru, Y., Marui, T., Matsumoto, I., Misaka, T., and Abe, K. (2007). Characterization of ligands for fish taste receptors. J Neurosci 27, 5584–5592. 10.1523/JNEUROSCI.0651-07.2007.
- 27. Yuan, X.-C., Liang, X.-F., Cai, W.-J., He, S., Guo, W.-J., and Mai, K.-S. (2020). Expansion of sweet taste receptor genes in grass carp (Ctenopharyngodon idellus) coincided with vegetarian adaptation. BMC Evol Biol *20*, 25. 10.1186/s12862-020-1590-1.
- 28. Hidaka, I., and Yokota, S. (1967). Taste receptor stimulation by sweet-tasting substances in the carp. Jpn J Physiol *17*, 652–666. 10.2170/jjphysiol.17.652.
- 29. Ritter, R.C., Slusser, P.G., and Stone, S. (1981). Glucoreceptors controlling feeding and blood glucose: location in the hindbrain. Science *213*, 451–452. 10.1126/science.6264602.
- Chhabra, K.H., Bathina, S., Faniyan, T.S., Samuel, D.J., Raza, M.U., de Souza Cordeiro, L.M., Viana Di Prisco, G., Atwood, B.K., Robles, J., Bainbridge, L., et al. (2024). ADGRL1 is a glucose receptor involved in mediating energy and glucose homeostasis. Diabetologia *67*, 170–189. 10.1007/s00125-023-06010-6.

<sup>8 |</sup> Anbalagan, Savani. Sugar-sensing swodkoreceptors. Zenodo. 2024. doi: <u>10.5281/zenodo.10938497</u>; Submission date: 07-04-2024

- 31. Yoon, N.A., and Diano, S. (2021). Hypothalamic glucose-sensing mechanisms. Diabetologia *64*, 985–993. 10.1007/s00125-021-05395-6.
- Li, X., Li, W., Wang, H., Cao, J., Maehashi, K., Huang, L., Bachmanov, A.A., Reed, D.R., Legrand-Defretin, V., Beauchamp, G.K., et al. (2005). Pseudogenization of a sweet-receptor gene accounts for cats' indifference toward sugar. PLoS Genet 1, 27–35. 10.1371/journal.pgen.0010003.
- Moede, T., Leibiger, B., Vaca Sanchez, P., Daré, E., Köhler, M., Muhandiramlage, T.P., Leibiger, I.B., and Berggren, P.-O. (2020). Glucokinase intrinsically regulates glucose sensing and glucagon secretion in pancreatic alpha cells. Sci Rep *10*, 20145. 10.1038/s41598-020-76863-z.
- Basco, D., Zhang, Q., Salehi, A., Tarasov, A., Dolci, W., Herrera, P., Spiliotis, I., Berney, X., Tarussio, D., Rorsman, P., et al. (2018). α-cell glucokinase suppresses glucose-regulated glucagon secretion. Nat Commun *9*, 546. 10.1038/s41467-018-03034-0.
- Heimberg, H., De Vos, A., Moens, K., Quartier, E., Bouwens, L., Pipeleers, D., Van Schaftingen, E., Madsen, O., and Schuit, F. (1996). The glucose sensor protein glucokinase is expressed in glucagon-producing alpha-cells. Proc Natl Acad Sci U S A *93*, 7036–7041. 10.1073/pnas.93.14.7036.
- 36. MacDonald, P.E., Joseph, J.W., and Rorsman, P. (2005). Glucose-sensing mechanisms in pancreatic β-cells. Philos Trans R Soc Lond B Biol Sci *360*, 2211–2225. 10.1098/rstb.2005.1762.
- 37. Walker, D.G., and Rao, S. (1964). The role of glucokinase in the phosphorylation of glucose by rat liver. Biochem J *90*, 360–368. 10.1042/bj0900360.
- Janowski, B.A., Willy, P.J., Devi, T.R., Falck, J.R., and Mangelsdorf, D.J. (1996). An oxysterol signalling pathway mediated by the nuclear receptor LXR alpha. Nature 383, 728–731. 10.1038/383728a0.
- Lehmann, J.M., Kliewer, S.A., Moore, L.B., Smith-Oliver, T.A., Oliver, B.B., Su, J.L., Sundseth, S.S., Winegar, D.A., Blanchard, D.E., Spencer, T.A., et al. (1997). Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. J Biol Chem 272, 3137–3140. 10.1074/jbc.272.6.3137.
- Mitro, N., Mak, P.A., Vargas, L., Godio, C., Hampton, E., Molteni, V., Kreusch, A., and Saez, E. (2007). The nuclear receptor LXR is a glucose sensor. Nature 445, 219–223. 10.1038/nature05449.
- 41. Anbalagan, S. (2023). "Blind men and an elephant", the need for animals in research, drug safety studies, and understanding civilizational diseases. Animal Model Exp Med *6*, 627–633. 10.1002/ame2.12364.
- 42. Anbalagan, S. (2024). Heme-based oxygen gasoreceptors. Am J Physiol Endocrinol Metab *326*, E178–E181. 10.1152/ajpendo.00004.2024.
- 43. Anbalagan, S. (2024). Oxygen is an essential gasotransmitter directly sensed via protein gasoreceptors. Animal Model Exp Med. 10.1002/ame2.12400.

<sup>9 |</sup> Anbalagan, Savani. Sugar-sensing swodkoreceptors. Zenodo. 2024. doi: <u>10.5281/zenodo.10938497</u>; Submission date: 07-04-2024

- 44. Gray, J.M., Karow, D.S., Lu, H., Chang, A.J., Chang, J.S., Ellis, R.E., Marletta, M.A., and Bargmann, C.I. (2004). Oxygen sensation and social feeding mediated by a C. elegans guanylate cyclase homologue. Nature *430*, 317–322. 10.1038/nature02714.
- 45. Lewis, M. (2005). The lac repressor. C R Biol *328*, 521–548. 10.1016/j.crvi.2005.04.004.
- 46. Hall, B.G. (1982). Transgalactosylation activity of ebg beta-galactosidase synthesizes allolactose from lactose. J Bacteriol *150*, 132–140. 10.1128/jb.150.1.132-140.1982.
- Otero-Rodiño, C., Velasco, C., Álvarez-Otero, R., López-Patiño, M.A., Míguez, J.M., and Soengas, J.L. (2016). In vitro evidence in rainbow trout supporting glucosensing mediated by sweet taste receptor, LXR, and mitochondrial activity in Brockmann bodies, and sweet taste receptor in liver. Comp Biochem Physiol B Biochem Mol Biol 200, 6–16. 10.1016/j.cbpb.2016.04.010.
- 48. Chen, W., Zhong, Y., Shu, J., Yu, H., Chen, Z., Ren, X., Hui, Z., and Li, Z. (2021). Characterization of glucose-binding proteins isolated from health volunteers and human type 2 diabetes mellitus patients. Proteins *89*, 1413–1424. 10.1002/prot.26163.