

Drosophila has only been presented in thesis form (Schmidt 1975) though ocellar ultrastructure has been published for the fleshfly (Toh et al. 1971).

We thank Allen Shearn for providing the mutant and the HVEM Laboratory for beam time. This work was supported by a Faculty Development Award to WSS from UMC, a Summer Research Fellowship to WSS from the UMC Graduate School Research Council and Hatch grant 2100 to SDC. We thank H.-M. Chu from the HVEM laboratory for technical assistance.

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Toda, M.J. Hokkaido University, Sapporo, Japan. The northernmost subarctic *Drosophilidae*.

The northernmost areas of the Holarctic region are the most interesting for consideration of biogeographical relationships between the two continents, Eurasia and North America, for it is there that the two continents were sometimes

connected in the past and are the closest even at the present time, through Beringia and several Arctic islands. In a recent monograph on drosophilid biogeography (Ashburner et al. eds. 1981), Wheeler (1981) and Bächli & Rocha Pite (1981) reviewed Nearctic and Palaeartic drosophilids, respectively, but did not specify the northernmost fauna in the two regions.

The strong cohesion of drosophilid distribution to woodland areas has been confirmed not only latitudinally (Basden 1956; Wheeler & Throckmorton 1960) but also altitudinally (Burla 1951; Basden & Harnden 1956; Bächli 1977), except for some specimens sporadically collected far beyond the forest boundary. It can be, therefore, concluded that the northernmost drosophilid fauna as a biogeographical entity is virtually confined to the subarctic forest zone, never deeply entering the real tundra.

Basden (1956) listed a total of 23 arctic species by choosing arbitrarily the Arctic Circle as the southern limit of the area, though this is obviously artificial and biologically meaningless as recognized by himself. Since then, considerable information on northern drosophilid fauna has been brought from several subarctic localities, Alaska (Wheeler & Throckmorton 1960), northern Finland (Lumme et al. 1979), and Mackenzie Delta, N.W.T., Canada (Takada & Toda 1981). By reviewing these reports, the northernmost subarctic drosophilid fauna are listed below. The chorological types are classified into four: Palaeartic (P), Nearctic (N), Holarctic (H) and Cosmopolitan (C); and are given before the specific number.

P 1 <i>Cacoxenus</i> (<i>Paracacoxenus</i>) <i>argyreator</i> Frey	N 22 Sc. (<i>Hemiscaptomyza</i>) <i>terminalis</i> (Loew)
P 2 <i>Stegana</i> (<i>Stegana</i>) <i>furta</i> (Linne)	H 23 Sc. (<i>Hsc.</i>) <i>trochanterata</i> Collin
P 3 St. (<i>Steganina</i>) <i>strobilii</i> Mik	H 24 Sc. (<i>Hsc.</i>) <i>unipunctum</i> (Zetterstedt)
H 4 St. (<i>Stn.</i>) <i>coleoptrata</i> (Scopoli)	C 25 Sc. (<i>Parascaptomyza</i>) <i>pallida</i>
P 5 <i>Amiota</i> (<i>Amiota</i>) <i>alboguttata</i> (Wahlberg)	(Zetterstedt)
N 6 A. (<i>A.</i>) <i>quadrata</i> Takada et Toda	P 26 Sc. sp. Lumme et al. 1979
N 7 A. (<i>A.</i>) sp. Wheeler & Throckmorton 1960	N 27 Sc. sp. Wheeler & Throckmorton 1960
P 8 <i>Chymomyza</i> <i>fuscimana</i> (Zetterstedt)	P 28 <i>Drosophila</i> (<i>Sophophora</i>) <i>alpina</i> Burla
N 9 Ch. <i>aldrichii</i> Sturtevant	P 29 D. (<i>So.</i>) <i>bifasciata</i> Pomini
N 10 Ch. <i>coxata</i> Wheeler	P 30 D. (<i>So.</i>) <i>eskoii</i> Lakovaara et Lankinen
N 11 Ch. <i>tetonensis</i> Wheeler	P 31 D. (<i>So.</i>) <i>obscura</i> Fallen
N 12 Ch. <i>wirthi</i> Wheeler	P 32 D. (<i>So.</i>) <i>subsilvestris</i> Hardy et
H 13 Ch. <i>caudatula</i> Oldenberg	Kaneshiro
H 14 Ch. <i>costata</i> (Zetterstedt)	N 33 D. (<i>So.</i>) <i>athabasca</i> Sturtevant et
P 15 <i>Scaptomyza</i> (<i>Scaptomyza</i>) <i>flava</i> (Fallen)	Dobzhansky
P 16 Sc. (<i>Sc.</i>) <i>griseola</i> (Zetterstedt)	N 34 D. (<i>So.</i>) <i>populi</i> Wheeler et Throckmorton
N 17 Sc. (<i>Sc.</i>) <i>nigrita</i> Wheeler	C 35 D. (<i>So.</i>) <i>melanogaster</i> Meigen
H 18 Sc. (<i>Sc.</i>) <i>graminum</i> (Fallen)	P 36 D. (<i>Lordiphosa</i>) <i>fenestrarum</i> Fallen
H 19 Sc. (<i>Sc.</i>) <i>montana</i> Wheeler	C 37 D. (<i>Dorsilopha</i>) <i>busckii</i> Coquillett
H 20 Sc. (<i>Sc.</i>) <i>teinoptera</i> Hackman	P 38 D. (<i>Hirtodrosophila</i>) <i>lundstroemi</i> Duda
P 21 Sc. (<i>Sc.</i>) sp. (= Finnish Sc. ? <i>montana</i>	P 39 D. (<i>H.</i>) <i>subarctica</i> Hackman
Basden 1956)	C 40 D. (<i>Drosophila</i>) <i>funebri</i> (Fabricius)

P 41 D. (D.) ezoana Takada et Okada	H 47 D. (D.) testacea von Roser*
P 42 D. (D.) littoralis Meigen	P 48 D. (D.) phalerata Meigen
P 43 D. (D.) lummei Hackman	N 49 D. (D.) rellima Wheeler
N 44 D. (D.) borealis Patterson	H 50 D. (D.) transversa Fallen
H 45 D. (D.) montana Patterson et Wheeler	N 51 D. (D.) melanderi Sturtevant
C 46 D. (D.) immigrans Sturtevant	

* Takada & Toda (1981) reported *D. putrida* from MacKenzie Delta, but that was a misidentification of *D. testacea*.

The northernmost subarctic drosophilid fauna is characterized by the relative richness in species number of the following taxa: *Chymomyza*, *Scaptomyza*, the *obscura* group (Nos. 28-33) and the *virilis* group (Nos. 41-45). It is noteworthy that the southernmost antarctic drosophilid fauna is monopolized by *Scaptomyza* (Brncic & Dobzhansky 1957). The relative percentages of the four chorological elements, calculated by excluding unidentified species, are as follows: Palaearctic (19 spp., 40.4%), Nearctic (12 spp., 25.5%), Holarctic (11 spp., 23.4%) and Cosmopolitan (5 spp., 10.6%). The relatively high percentage of Holarctic elements suggests that the intercontinental faunal exchange, possibly across Beringia, repeatedly occurred until relatively recent times in the northernmost subarctic region.

This work is No. 2357 contributed from the Institute of Low Temperature Science, Hokkaido University.

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Tolchkov, E.B. and V.A. Gvozdev. Institute of Molecular Genetics, USSR Academy of Sciences, Moscow USSR. The structure of two rearrangements resulting in the *Pgd* gene position effect in *Drosophila melanogaster*.

The study shows that the previously described rearrangements $T(1;4)pn2$ and $Tp(1)pn3$ (Ilyina et al. 1980) are pericentric inversions, designated $In(1LR)pn2a$ and $In(1LR)pn2b$, respectively, with very similar genetic structures.

Analysis of recombination in $pn2a/y$ *cv v f* car females has shown the genetic map of the rearrangement to differ from that of the normal X chromosome. The $pn2a$ rearrangement is characterized by the following order of the markers: *cv-v-f-car-y* (cf. *y-cv-v-f-car* in the normal chromosome). The distances between the *y* gene and the markers nearest to it, *car* and *f*, in the rearrangements are in good agreement with the reported (Schalet & Lefevre 1976) distances between these markers and the centromere. The easiest way to explain these results is to assume that the distal section of the X chromosome carrying the y^+ gene is transferred to the centromeric region of the X chromosome and not to the 4th chromosome, as formerly believed. The genetic maps of the rearrangements $pn2a$ and $pn2b$ do not differ. Analysis of the polytene chromosome shows the distal end of the rearranged chromosome to break off in the 2DE region. The telomere of the rearranged chromosome consists of heterochromatic material, as attested by its meta-chromasy (bluish staining with azure-eosine, as opposed to the violet staining of the bulk of the chromosomes) and the presence of highly repetitive sequences, probably satellite DNA, revealed by *in situ* hybridization with total labelled DNA in a set-up where the hybridization of highly repetitive DNA is selectively favoured. The 1A-2DE region is associated with the chromocenter through the 2DE segment. The metaphase chromosomes show an enlarged XR the size of the 4th chromosome, which probably corresponds to the 1A-2DE fragment. Comparative analysis of the data on the recombination and structure of the polytene and metaphase chromosomes suggests that the rearrangements are pericentric inversions of the X chromosome (Figure 1).

The euchromatic break point of the inversions lies in the 2D-F region, whose fine genetic structure has been studied earlier (Gvozdev et al. 1973; Gvozdev et al. 1975). Genetic analysis has demonstrated that in both inversions the genes corresponding to