

# Study of Optical Properties of a Glutathione Capped Gold Nanoparticles using Linker (MHDA) by Fourier Transform Infra Red Spectroscopy and Surface Enhanced Raman Scattering

A. Deregowska, J. Depciuch, R. Wojnarowska, J. Polit, D. Broda, H. Nechai, M. Gonchar, and E. Sheregii

**Abstract**—16-Mercaptohexadecanoic acid (MHDA) and tripeptide glutathione conjugated with gold nanoparticles (Au-NPs) are characterized by Fourier Transform Infrared (FTIR) spectroscopy combined with Surface-enhanced Raman scattering (SERS) spectroscopy. Surface Plasmon Resonance (SPR) technique based on FTIR spectroscopy has become an important tool in biophysics, which is perspective for the study of organic compounds. FTIR-spectra of MHDA shows the line at 2500 cm<sup>-1</sup> attributed to thiol group which is modified by presence of Au-NPs, suggesting the formation of bond between thiol group and gold. We also can observe the peaks originate from characteristic chemical group. A Raman spectrum of the same sample is also promising. Our preliminary experiments confirm that SERS-effect takes place for MHDA connected with Au-NPs and enable us to detected small number (less than 106 cm<sup>-2</sup>) of MHDA molecules. Combination of spectroscopy methods: FTIR and SERS – enable to study optical properties of Au-NPs and immobilized bio-molecules in context of a bio-nano-sensors.

**Keywords**—Glutathione; gold nanoparticles, Fourier transform infrared spectroscopy, MHDA, surface-enhanced Raman scattering.

## I. INTRODUCTION

IT is well known fact that metal nanoparticles attract an extensive attention in various field of physics, chemistry, biology, due to their unique properties [1]. Among colloids of metallic, gold nanoparticles, which are one of the most stable colloid, does not form a stable oxide layer under ambient conditions. Further, the properties of gold nano-scale, which are dominated by electromagnetic resonances of plasmons and high surface to volume ratio are important in the context of direct electron transfer between bio-molecules and electrode surface and is particularly advantageous for biosensing technology [2]. Moreover, gold have strong binding affinity toward thiols, amines and disulfides [3].

M. Gonchar, D. Broda, A. Deregowska, J. Depciuch are with Department of Biotechnology, University of Rzeszow, Werynia 502, 36-100, Kolbuszowa, Poland (phone: 48178723261; fax: 48178723261; e-mail: deregowskaanna@tlen.pl).

R. Wojnarowska, J. Polit are with Center of Microelectronics and Nanotechnology, University of Rzeszow, 35-959 Rzeszow, Poland(phone: 48178721154; fax: 48178721283; e-mail: sheregii@univ.rzeszow.pl).

M. Gonchar, E. Sheregii are with Institute of Cell Biology, NAS of Ukraine, Dragomanov St. 14/16, 79005 Lviv, Ukraine (phone: 380322612108, fax: 380322612148, e-mail: myg52@yahoo.com).

The infrared (IR) optical properties of gold nanoparticles are determinate by the plasmons - collective oscillations of electrons within a metal nanoparticles, activated by an incident electromagnetic field. Surface plasmon resonance (SPR) consists on an electromagnetic wave propagating along the surface of a thin metal layer stimulated by incident light [4]. The IR spectra are formed as a consequence of the absorption of electromagnetic radiation at frequencies that correlate to the vibration of specific sets of chemical bonds from within a molecule. It is assumed that the intensity of the band is proportional to the concentration of the functional chemical groups in a molecule.

There are two types of vibrational motions, which are defined by stretching and bending modes. The stretching vibrations are characterized by rhythmic movement along the axis of tying, so the distance between atoms increases and decreases. The second one is bending mode, where an angle between atoms bond is changed [5].

Surface-enhanced Raman scattering (SERS) is a special phenomenon within the field of Raman spectroscopy and involves generation of enhanced Raman signal (by as much as 10<sup>8</sup>) from analyzed molecules adsorbed on roughened metal surfaces (usually nanoparticles of silver and gold) [6]. SERS technique, attributed to large electromagnetic fields near nanostructured metallic surfaces and short range chemical effect, has the capability to detect a single molecule and provide information on molecular structure. Although some enhancement in SERS is due to chemical contribution (electronic interaction between metal and adsorbed molecules), the main reason why SERS produces extraordinary enhancements are surface plasmons - the electromagnetic property of nanostructures.

In a brief preliminary account of the present work, we have studied the immobilization on a surface of gold nanoparticles (Au-NPs) using a surfactant- 16-Mercaptohexadecanoic acid (MHDA), as a space arm. MHDA is a linker for covalent binding of proteins by condensation of carboxylic group and amino group of the protein. The surfactant bind to the gold surface through their thiols. As a protein we used a reduced glutathione, which is a tripeptide (Glu-Cys-Gly), as a model for proteinaceous ligand. Their physical properties were studied using FTIR and SERS.

## II. EXPERIMENTAL METHOD

### A. Samples

Au-NPs were prepared by the Turkevich method [8]. This can be done by using hydrogen tetrachloroaurate ( $\text{HAuCl}_4$ ) as source of gold ions and trisodium citrate ( $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ ) as a reduction and stabilized agent. Tetrachlorauric acid is heated to its boiling on a magnetite stirrer, to which reducing agent sodium citrate is injected. Heating is continued till the color of the solution changed from colorless to pink/red. The colloidal gold is formed because the citrate ions act as both a reducing agent, and a capping agent. As a result of the citrate molecules settle on the particle surface, Au-NPs are stabilized through electrostatic repulsion. The size of the produced particles can be controlled by varying the amount of available gold in the solution.

In order to attach the proteins to the citrate stabilized gold nanoparticles mercaptoalkane linkers are used. 16-mercaptohexadecanoic acid. Au-structures were incubated in MHDA in ethanol for overnight at  $4^\circ\text{C}$ . During incubation MHDA replace the citrate molecules and the thiol group of the mercaptoalkanes interact with the gold surface through an Au-S interaction. After rinsing with dimethylformamide (DMF), the MHDA-covered Au-structures were incubated in DMF solution of pentafluorophenyl (PFP) and *N,N*-diisopropylethylamine (DIPEA) and *N*-cyclohexyl-*N'*-(2-morpholinoethyl) carbodiimide metho-*p*-toluenesulfonate (CMC), during 30 min at  $25^\circ\text{C}$ . After repeated rinsing with DMF, condensation of the activated Au-linked carboxylic groups with first amine group of the reduced glutathione was carried out. On the surfaces of the electrode, solution with enzyme were put and incubated for 1 hour at  $25^\circ\text{C}$ . After rinsing with phosphate buffer, blocking of un-reacted carboxylic groups with 2-(2-aminoethoxy) ethanol (AEE) was performed, using solution of AEE in bicarbonate buffer. The bio-functionalized Au-structure was rinsed with phosphate buffer [7]-[8].

### B. Measurements

The Fourier spectrometer Nicolet 6700 provided measurements of optical reflectivity in the range of far- as well as middle-infrared: from  $500\text{ cm}^{-1}$  to  $4000\text{ cm}^{-1}$  with resolution about  $2\text{ cm}^{-1}$ . Reflectivity was measured using silver mirror as reference. The FTIR spectra of the samples were collected by this way at room temperature. The resolution of  $1\text{--}4\text{ cm}^{-1}$  at 32 scans during one minute characterized these measurements.

The Raman-spectra were examined using Smart Raman DXR, which has a laser  $780\text{ nm}$  and maximal power of this laser is  $14\text{ mW}$ . This spectrometer enables us to measure the Raman shift spectra in the range from  $50$  to  $3300\text{ cm}^{-1}$ .

## III. RESULT AND DISCUSSION

### A. X-ray Diffraction

The synthesized gold nanoparticles by Turkevich method were then checked for the structure and phase purity based on the X-ray diffraction (XRD) analysis. The XRD pattern (Fig.

1) shows five intense peaks in the whole spectrum of  $2\theta$  values ranging from  $20$  to  $100$  degree. The presence of intense peaks of nanoparticles (111), (200), (220), (222) and (311) confirms that the gold nanoparticles are crystalline in nature way and suggest that the Au-NPs are stable and do not aggregate [9]-[10].

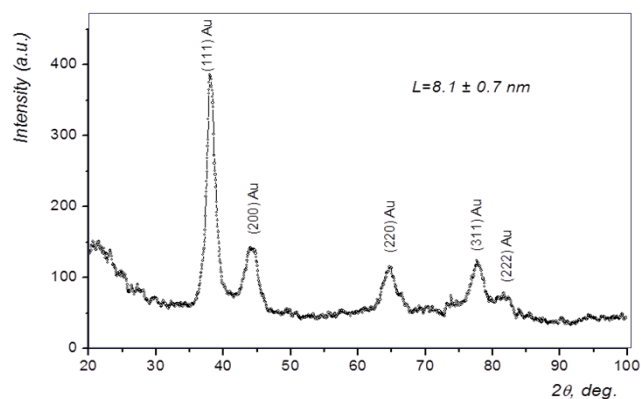


Fig. 1 XRD of Au-NPs

### B. Surface Enhanced Raman Scattering

The effect of size of Au-NPs on signal SERS intensity is demonstrated in Fig. 2. It could be seen that the SERS signal became larger with the size of nanoparticles decreasing, so the size of Au-NPs have significant effect on signal SERS [11].

It can be seen in Fig. 4 that the group of peaks in the range of  $1200\text{--}1600\text{ cm}^{-1}$  is characteristic for glutathione (spectrum a). The band at  $1296\text{ cm}^{-1}$  and a group of bands in the region from  $1400$  to  $1500\text{ cm}^{-1}$  are observed due to (C-C) stretching vibrations. The other spectra represent glutathione capped Au-NPs of different size:  $127\text{ nm}$  (spectrum b) and  $90\text{ nm}$  (spectrum c). So, the 5-times enhancement of Raman-intensity is observed in case of  $90\text{ nm}$  size Au-NPs.

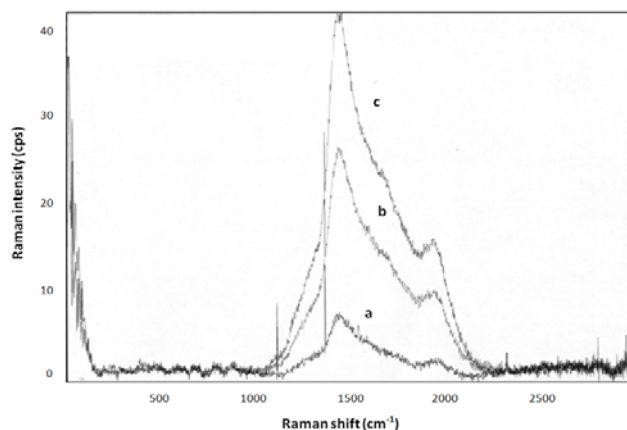


Fig. 2 Raman spectra of glutathione (a), Au-NPs of  $127\text{ nm}$  diameter + glutathione (b) and Au-NPs of  $90\text{ nm}$  diameter + glutathione (c)

### C. FTIR

IR reflectance spectra in Fig. 3 shows several groups of lines associated with the citrate molecules linked with surface of gold nanoparticles. The peak at the wavenumber  $1077\text{ cm}^{-1}$  is assigned to the stretching vibrations of C-O whereas peak

at  $1680\text{ cm}^{-1}$  is attributed to C=O [12]. The peak at the wavenumber at  $1250\text{ cm}^{-1}$  is characteristic for the sample investigated. It can also be found that there are two bands at  $2853$  and  $2914\text{ cm}^{-1}$  which are attributed to stretching vibrations of  $\text{CH}_2$ , and band at  $1412\text{ cm}^{-1}$  which can be

assigned to bending vibrations of CH as well as the line at  $1507\text{ cm}^{-1}$  – to the bending vibrations of OH.

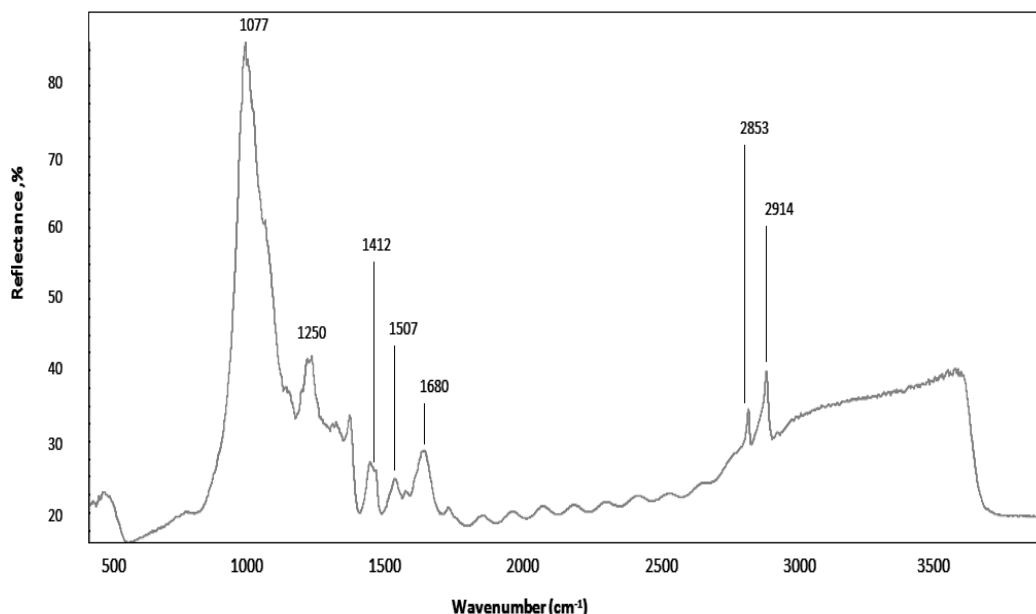


Fig. 3 IR reflectance spectra of citrate-stabilized Au-NPs

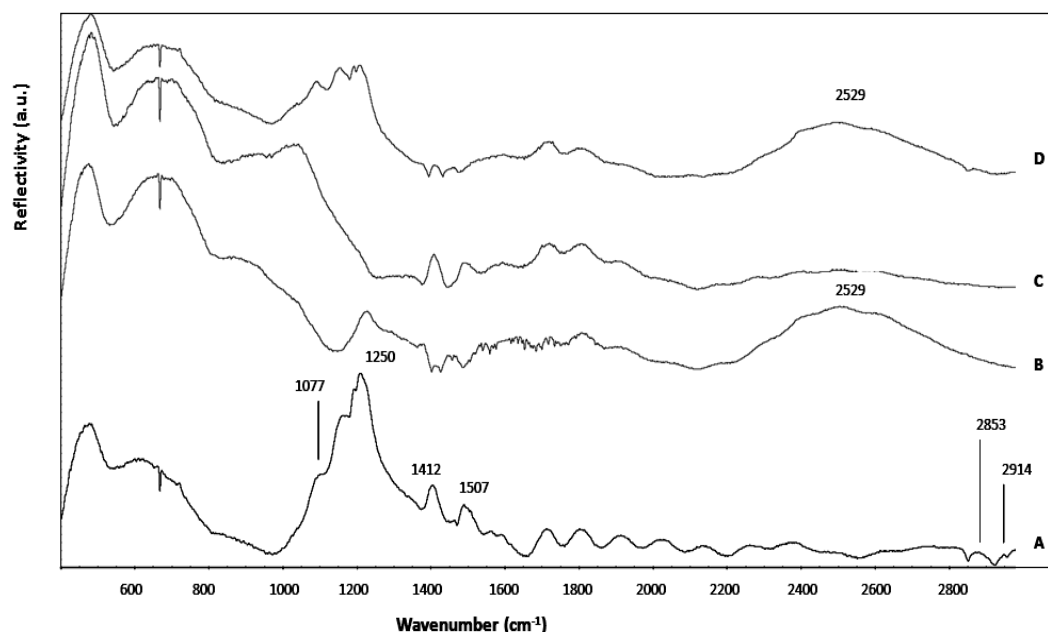


Fig. 4 Infrared spectra of different samples: the citrate stabilized gold nanoparticles (spectrum A), the pure MHDA (spectrum B), the Au-NPs + MHDA (spectrum C), Au-NPs + MHDA + glutathione (spectrum D)

Fig.4 shows the curves of the citrate stabilized Au-NPs (spectrum A), the pure MHDA (spectrum B), the MHDA linked with Au-NPs (spectrum C) and the glutathione capped gold nanoparticles (spectrum D). In the curve B is observed the group of specific broad bands at  $2529\text{ cm}^{-1}$  caused by thiol vibrations which are appeared obviously in MHDA [12]. This group of lines is suppressed in MHDA linked with Au-NPs

giving evidence, that MHDA anchors on the gold surface through the sulfur atom in the thiol group (see the C spectrum) and suggests that the bonding of the biofunctional ligands to the Au surface takes place through the S-H group. The Au-NPs-MHDA researched is a model system for the binding of biofunctional surfactants onto the Au-NPs surface. Thiol bonds are again shown in the AuNPs + MHDA + Glutathione

spectrum (at  $2529\text{cm}^{-1}$ , see curve D) because glutathione is composed of glycine, glutamic acid and cysteine, which also has a mercaptan group [13]-[14].

#### IV. CONCLUSION

FTIR enable us to monitor the interaction of MHDA and glutathione with the gold matrix. The technique in mid-IR can achieve sensitivity that is comparable and even higher than in the visible range and used to detect of biomolecules according to their "fingerprints".

FTIR and Raman spectra can be used to identify unknown molecules, determine their concentrations, and to study bond strengths in molecules, so both of these spectroscopy techniques give a lot of possibilities for study chemical and physics properties of thiol stabilized Au-NPs and proteinaceous ligand.

The colloidal-bound proteins enable to build bio-nano-sensors with great control over functionality and sensitivity.

#### ACKNOWLEDGMENT

This work is supported by the Project No.\*UDA-POKL.04.01.02-00-038/09-00.

#### REFERENCES

- [1] S.J. Guo, E.K. Wang, "Synthesis and electrochemical applications of gold nanoparticles," *Analytica Chimica Acta*, Vol. 598, 2007, pp. 181-192.
- [2] Liu, D.Leech, H. Ju, "Application of Colloidal Gold in Protein Immobilization, Electron Transfer, and Biosensing," *Analytical Letters*, Vol. 36, , 2003, pp. 1-19.
- [3] I. H. El-Sayed, X.Huang, M.A. El-Sayed, "Surface plasmon resonance scattering and absorption of anti-EGFR antibody conjugated gold nanoparticles in cancer diagnostics: applications in oral cancer," *Nano Letters*. Vol. 5, 2005, pp. 829-834.
- [4] M. Hu, J. Chen, ZY. Li, L. Au, G.V. Hartland, X. Li, M. Marquez, Y. Xia, "Gold nanostructures: engineering their plasmonic properties for biomedical applications," *Chemical Society Reviews*, Vol. 35, 2006, pp.1084-1094.
- [5] R.J. Silbey, R.A. Alberty, and M. G. Bawendi, *Physical Chemistry*, John Wiley & Sons Ltd., New York, 2004.
- [6] R. Aroca, *Surface Enhanced Vibrational Spectroscopy*, John Wiley & Sons Ltd., Chichester, 2006.
- [7] A.B. Descalzo, R. Martínez-Mañez, F. Sancenón, K. Hoffmann, K. Rurack, "Hybrid Nanoapatite by Polysaccharide Nanogel-templated Mineralization," *Angewandte Chemie International Edition*, Vol.45, 2006, pp. 5924-5948 2006.
- [8] J. Kimling, M. Maier, B. Okenve, V. Kotaidis, H. Ballot, A. Plech, "Turkevich Method for Gold Nanoparticle Synthesis Revisited," *Journal Physical Chemistry B*, Vol. 110, 2006, pp. 15700-15707.
- [9] N. Stasyuk, R. Serkiz, S. Mudry, G. Gayda, A. Zakalskiy, Y. Koval'chuk, M. Gonchar, M. Nisnevitch, "Recombinant human arginase I immobilized on gold and silver nanoparticles: preparation and properties," *Nanotechnology Development*, Vol.1, 2011, p. 11-15. S.
- [10] V. Leff, L. Brandt, J. R. Heath, "Synthesis and Characterization of Hydrophobic, Organically-Soluble Gold Nanocrystals Functionalized with Primary Amines," *Langmuir*, Vol. 12, 1996, pp. 4723-4730.
- [11] H.Shayani-Jam,D.Nematollahi,"Electrochemical evidences in oxidation of acetaminophen in the presence of glutathione and N-acetylcysteine," *Chemical Communications*, Vol. 46, 2009, pp. 409-411.
- [12] M. Hasan, D. Bethell, M. Brust, "The fate of sulfur-bound hydrogen on formation of self-assembled thiol monolayers on gold: (1)H NMR spectroscopic evidence from solutions of gold clusters," *Journal of the American Chemical Society*, Vol. 124, 2002, pp. 1132-1133.
- [13] M. J. Hostetler, J.J. Stokes, R.W. Murray, "Infrared Spectroscopy of Three-Dimensional Self-Assembled Monolayers: □ N-Alkanethiolate

Monolayers on Gold Cluster Compounds," *Langmuir*, Vol. 12, 1996, pp. 3604-3612.

- [14] Y. Sahoo, H. Pizem, T. Fried, D. Golodnitsky, L. Burstein, C.N. Sukenik, G. Markovich, "Alkyl Phosphonate/Phosphate Coating on Magnetite Nanoparticles: □ A Comparison with Fatty Acids," *Langmuir*, Vol. 17, 2001, pp.7907-7911.