



**anses**

# MEASUREMENT UNCERTAINTY FOR QUANTITATIVE ANALYSES: EN ISO 19036 **TECHNICAL UNCERTAINTY**



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**CONNAÎTRE, ÉVALUER, PROTÉGER**

**EURLs JOINING WEBINAR ON MEASUREMENT UNCERTAINTY**

# EN ISO 19036:2019

## Microbiology of the food chain - Estimation of measurement uncertainty for quantitative determinations

→ 3 components of measurement uncertainty



**Technical  
Uncertainty**

**Matrix  
Uncertainty**

**Distributional  
Uncertainty**

## TECHNICAL UNCERTAINTY; $U_{tech}(1)$

Uncertainty resulting from operational variability associated with the technical steps of the analytical procedures

- associated with the main stages in microbiological method
- considered as a performance characteristic when the method is implemented in a given laboratory
- usually, the largest of the three uncertainty components

## TECHNICAL UNCERTAINTY; $U_{\text{tech}}$ (2)

- Characteristics of the method; technical uncertainty estimated for one method cannot be applied to other methods
- Estimated from the standard deviation of reproducibility on the final result of the measurement process; preferably based on “intralaboratory reproducibility”
- Estimated by performing experiment – Data may be collected in a short period of time as a special exercise

## Measurements conditions

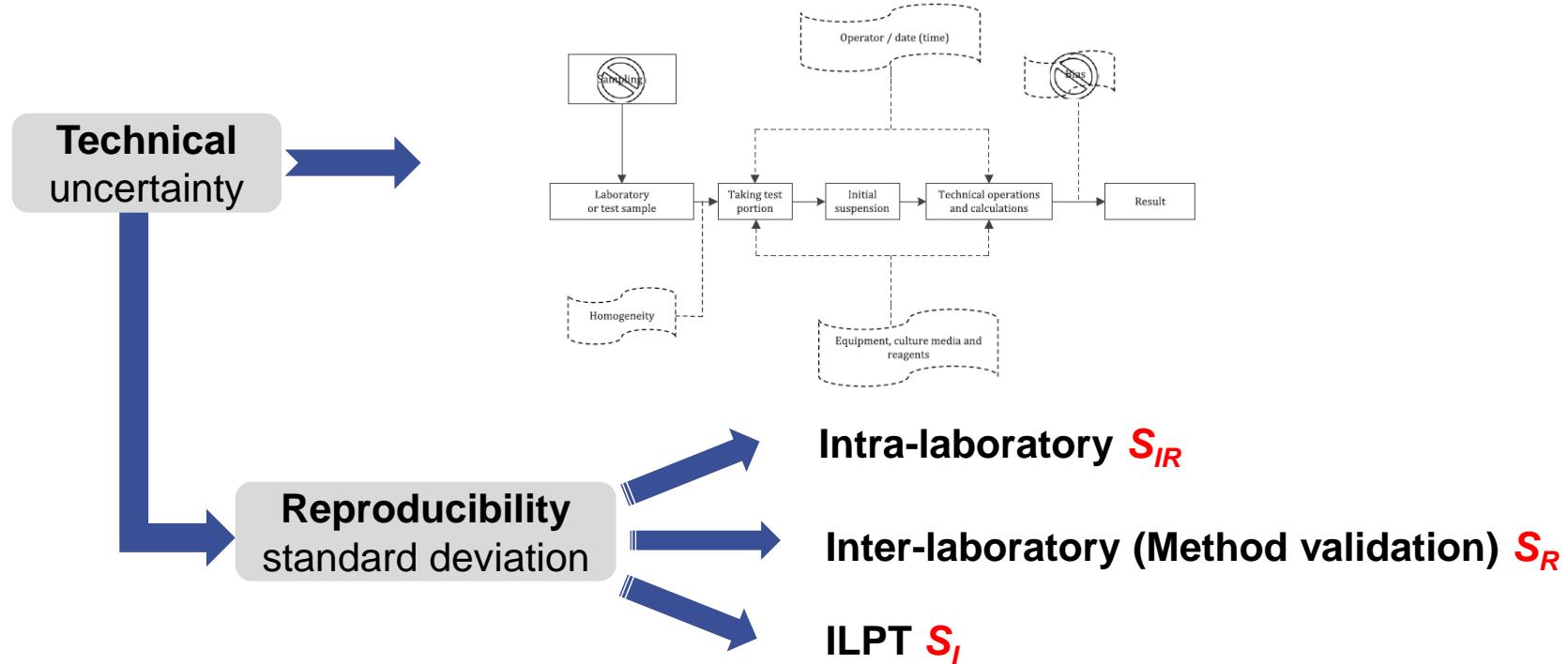
- the measurement conditions (e.g., A and B) for each test portions should differ in as many ways as possible
- the laboratory sample, where possible, should cover the expected natural variation in contamination level
- the pattern of variation should not be the same for all laboratory (test) samples
- If artificial contamination is needed, spike initial suspension

# TECHNICAL UNCERTAINTY; $U_{tech}$ (3)

## Typical sources of uncertainty are:

- stages of the test: e.g., weight of test portion; preparation of initial suspension, serial dilutions, inoculation, incubation, colony counting, confirmation
- batches of culture media, reagents, Equipment: e.g., weighing equipment; vortex/mixers; volumetric measuring/dispensing equipment; pipettes; incubators/baths
- tolerances within method: e.g., temperature range; incubation times
- technicians/operators

# TECHNICAL UNCERTAINTY; $U_{tech}$ (4)



# OPTION 1: INTRA-LABORATORY REPRODUCIBILITY

$S_{IR}(2)$



- Preferred option for deriving technical uncertainty:
- ✓ Reproducibility conditions are varied in the laboratory
- ✓ Provides uncertainty values that is linked to the laboratory
- To be determined for each method for at least 2 test portions
- Samples subjected to artificial contamination (if needed)

# OPTION 1: INTRA-LABORATORY REPRODUCIBILITY

## S<sub>IR</sub>(2)

### 1. Experimental protocol

For each test method, perform the protocol for

- **At least 10 laboratory samples**
- **At least 2 acceptable measurements for each laboratory sample**
- **Repeat design for each laboratory sample**

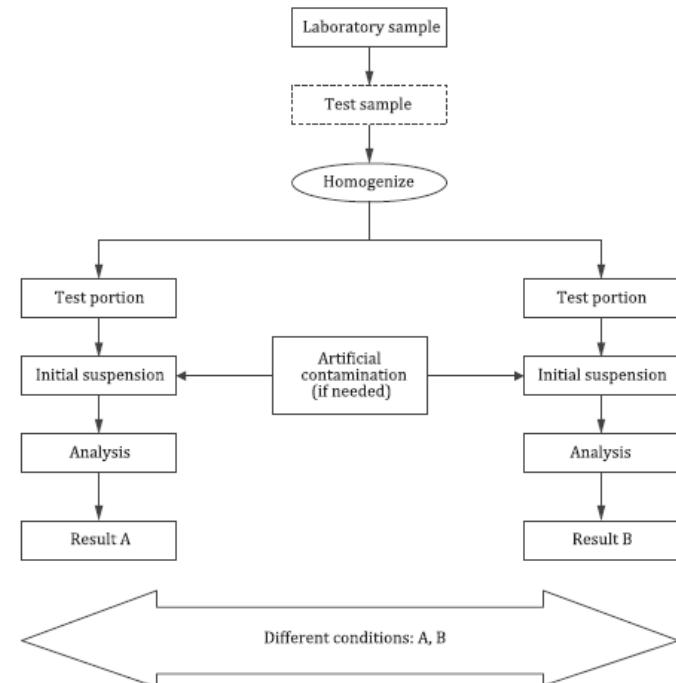


Figure 2 — Experimental protocol for estimation of intralaboratory reproducibility — Two determinations on each laboratory sample

# OPTION 1: INTRA-LABORATORY REPRODUCIBILITY

## $S_{IR}(2)$

## 2. Criteria for excluding and calculation

- The sum of the total colonies on all retained plates

$30 \leq \sum C \leq 300$  cfu/plate or lower as specified in the specific standard

- Transform the result from each test portion into  $\log_{10}$  before calculations
- Eg. for 10 lab samples from operators A and B

$$s_{IR} = \sqrt{\frac{1}{2n} \sum_{i=1}^n (y_{iA} - |y_{iB}|)^2}$$

where

$i$  is the index of the sample,  $i = 1$  to  $n$  ( $n \geq 10$ );

$y_{iA}, y_{iB}$  are the log-transformed data, in  $\log_{10}$  cfu/g or ml, from conditions A and B

# OPTION 1: INTRA-LABORATORY REPRODUCIBILITY

## $S_{IR}(2)$

## 2. Acceptable results and calculation

Description of data:		Utech LCSV pour méthode CFA 2009LR28 (données issues du dossier d'admission)					
Standard uncertainty	$s_{IR}$	Optional; uncertainty components <u>subtracted</u> from $s_{IR}$ to give $s_{IR:corr}$					
	$s_{IR:corr}$	Matrix	Poisson		Confirmation		
Laboratory Sample ID	Result $\log_{10}$	$u_{matrix}$	$\Sigma C$	$u_{Poisson}$	$n_p$	$n_c$	$u_{conf}$
19Q002517	4,832508913						
19Q002517	4,812913357						
19Q002518	4,913813852						
19Q002518	4,799340549						
19Q002326	3,806179974						
19Q002326	3,672097858						
19Q002327	3,770852012						
19Q002327	3,612783857						
19Q002832	3,342422681						
19Q002832	3,230448921						
19Q002979	2,949390007						
19Q002979	3,230448921						
19Q002566	2,380211242						
19Q002566	2,255272505						
19Q002746	2,477121255						

## OPTION 2: INTER-LABORATORY $S_R$ (METHOD VALIDATION)

Method used routinely submitted to inter-laboratory validation study

### Conditions to meet:

**Repeatability and reproducibility estimates of precisions with the lab shall not be larger than those obtained in the inter-lab study**

- Advantages = use precision data already generated
- Drawbacks

Difficulty to generalise values from the inter-lab sutdy to routine samples.

Unavaalaibility of reproducibility parameters from inter-lab method validation study for all methods.

This is why inter-laboratory SR from validation method is only an

**OPTION 2**

## OPTION 3: INTER-LABORATORY $S_R$ (PROFICIENCY TEST)

Laboratory taken part in ILPT can use  $S_R$  to deduce its technical uncertainty

**Conditions to meet:**

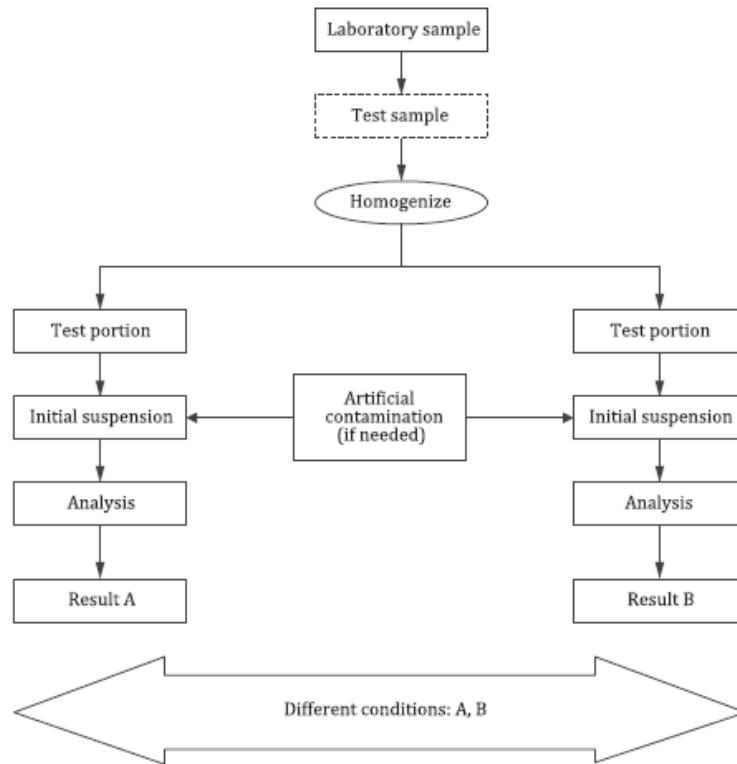
**Method used by the lab in PT = Same method that the laboratory uses in routine analyses**

- Advantages = available values, large number of PT schemes organized in food microbiology
- Drawbacks

Difficulty to generalise from ILPT to routine analyses performed by the laboratory.

# PRACTICAL APPROACH TO ESTIMATE THE STANDARD DEVIATION OF REPRODUCIBILITY ( $S_R$ )

# EXAMPLE OF INTRALABORATORY $S_R$



**Take 10 test portions from the same sample**

**Artificial or naturally contaminated sample**  
**Artificial contamination should be performed in initial suspension and should cover natural variation (**

**Homogenize before and after contamination**

**Implement the method on all the suspensions by inoculating the appropriate dilutions**

Figure 2 — Experimental protocol for estimation of intralaboratory reproducibility — Two determinations on each laboratory sample

# CALCULATION OF INTRALABORATORY $S_R$

Example of CPS using EN ISO 6888-1 in cheese (Mozzarella)

Lab sample	Test portion	Inoculated volume	Dilution factor and copunted colonies					Total colonies counted	Result (EN ISO 7218)	$\log_{10}$ CFU/g	Criteria compliance
			Nombre de	d1	C1	d2	C2				
1	1A	0,1	1	-1	57	-2	4	61	5,5E+03	3,74	conform
2	2A	0,1	1	-2	56	-3	2	58	5,3E+04	4,72	conform
3	3A	0,1	1	-2	54	-3	4	58	5,3E+04	4,72	conform
4	4A	0,1	1	-2	71	-3	5	76	6,9E+04	4,84	conform
5	5A	0,1	1	-3	65	-4	6	71	6,5E+05	5,81	conform
6	6A	0,1	1	-3	35	-4	7	42	3,8E+05	5,58	conform
7	7A	0,1	1	-3	48	-4	8	56	5,1E+05	5,71	conform
8	8A,	0,1	1	-4	63	-5	5	68	6,2E+06	6,79	conform
9	9A	0,1	1	-4	48	-5	4	52	4,7E+06	6,67	conform
10	10A	0,1	1	-4	57	-5	8	65	5,9E+06	6,77	conform
11	11A	0,1	1	-5	64	-6	11	75	6,8E+07	7,83	conform
12	12A	0,1	1	-5	53	-6	4	57	5,2E+07	7,71	conform

\*: Dilution "-1" corresponds to intial solution

## Criteria for the acceptability of the result

- 1- For each test portion, the sum of colonies retained for counting must be  $\geq 30$ .
- 2- Methods including partial confirmation: at least half of the tested colonies confirmed

# CALCULATION OF INTRALABORATORY $S_R$

Example of CPS using EN ISO 6888-1 in cheese (Mozzarella)

Lab sample	Test portion	Inoculated volume	Dilution factor and counted colonies					Total colonies counted	Result (EN ISO 7218)	$\log_{10}$ CFU/g	Criteria compliance
			Nombre de	d1	C1	d2	C2				
1	1B	0,1	1	-1	84	-2	19	103	9,4E+03	3,97	conform
2	2B	0,1	1	-2	84	-3	8	92	8,4E+04	4,92	conform
3	3B	0,1	1	-2	86	-3	9	95	8,6E+04	4,94	conform
4	4B	0,1	1	-2	108	-3	9	117	1,1E+05	5,03	conform
5	5B	0,1	1	-3	104	-4	3	107	9,7E+05	5,99	conform
6	6B	0,1	1	-3	86	-4	6	92	8,4E+05	5,92	conform
7	7B	0,1	1	-3	122	-4	15	137	1,2E+06	6,10	conform
8	8B	0,1	1	-4	135	-5	11	146	1,3E+07	7,12	conform
9	9B	0,1	1	-4	123	-5	11	134	1,2E+07	7,09	conform
10	10B	0,1	1	-4	96	-5	18	114	1,0E+07	7,02	conform
11	11B	0,1	1	-5	110	-6	6	116	1,1E+08	8,02	conform
12	12B	0,1	1	-5	109	-6	20	129	1,2E+08	8,07	conform

\*: Dilution "-1" corresponds to initial solution

## Criteria for the acceptability of the result

1- For each test portion, the sum of colonies retained for counting must be  $\geq 30$ .

2- Methods including partial confirmation: at least half of the tested colonies confirmed

# CALCULATION OF INTRALABORATORY $S_R$

Description of data:		Optional; uncertainty components <u>subtracted</u> from $s_{IR}$ to give $s_{IR:corr}$						MPN		
Too few Results		$s_{IR}$	$n_i$				$m_i$			
Standard uncertainty	$s_{IR:corr}$	Matrix	Poisson	Confirmation		$u_{MPN}$	$x_i$			
Laboratory Sample ID	Result log <sub>10</sub>	$u_{matrix}$	$\Sigma C$	$u_{Poisson}$	$n_p$	$n_c$	$u_{conf}$	$u_{MPN}$	$x_i$	
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# CALCULATION OF INTRALABORATORY $S_R$

Example of total microflora using EN ISO 4833-1 (Amd1 2022) in cheese (Mozzarella)

Lab sample	Test portion	Inoculated volume	Dilution factor and counted colonies					Total colonies counted	Result (EN ISO 7218)	$\log_{10}$ CFU/g	Criteria compliance
			Nombre de	d1	C1	d2	C2				
1	1A	1	1	-5	11	-6	0	11	1,0E+06	6,00	not conform
2	2A	1	1	-5	5	-6	0	5	4,5E+05	5,66	not conform
3	3A	1	1	-5	16	-6	1	17	1,5E+06	6,19	not conform
4	4A	1	1	-5	10	-6	0	10	9,1E+05	5,96	not conform
5	5A	1	1	-3	155	-4	12	167	1,5E+05	5,18	conform
6	6A	1	1	-3	98	-4	7	105	9,5E+04	4,98	conform
7	7A	1	1	-4	58	-5	10	68	6,2E+05	5,79	conform
8	8A	1	1	-3	229	-4	21	250	2,3E+05	5,36	conform
9	9A	1	1	-4	198	-5	28	226	2,1E+05	5,31	conform
10	10A	1	1	-4	69	-5	7	76	6,9E+05	5,84	conform
11	11A	1	1	-3	102	-4	14	116	1,1E+05	5,02	conform
12	12A	1	1	-3	183	-4	17	200	1,8E+05	5,26	conform

\*: Dilution "-1" corresponds to initial solution

### Criteria for the acceptability of the result

- 1- For each test portion, the sum of colonies retained for counting must be  $\geq 30$ .
- 2- Methods including partial confirmation: at least half of the tested colonies confirmed

# CALCULATION OF INTRALABORATORY $S_R$

Example of total microflora using EN ISO 4833-1 (Amd1 2022) in naturally contaminated frozen melon

Lab sample	Test portion	Inoculated volume	Dilution factor and counted colonies					Total colonies counted	Result (EN ISO 7218)	$\log_{10}$ CFU/g	Criteria compliance
			Nombre de	d1	C1	d2	C2				
1	1B	1	1	-5	6	-6	1	7	6,4E+05	5,80	not conform
2	2B	1	1	-5	10	-6	0	10	9,1E+05	5,96	not conform
3	3B	1	1	-5	15	-6	0	15	1,4E+06	6,13	not conform
4	4B	1	1	-5	10	-6	0	10	9,1E+05	5,96	not conform
5	5B	1	1	-3	158	-4	25	183	1,7E+05	5,22	conform
6	6B	1	1	-3	149	-4	14	163	1,5E+05	5,17	conform
7	7B	1	1	-4	35	-5	5	40	3,6E+05	5,56	conform
8	8B	1	1	-3	222	-4	21	243	2,2E+05	5,34	conform
9	9B	1	1	-3	31	-4	2	33	3,0E+04	4,48	conform
10	10B	1	1	-4	43	-5	1	44	4,0E+05	5,60	conform
11	11B	1	1	-3	177	-4	14	191	1,7E+05	5,24	conform
12	12B	1	1	-3	197	-4	25	222	2,0E+05	5,30	conform

\*: Dilution "-1" corresponds to initial solution

## Criteria for the acceptability of the result

- 1- For each test portion, the sum of colonies retained for counting must be  $\geq 30$ .
- 2- Methods including partial confirmation: at least half of the tested colonies confirmed

# CALCULATION OF INTRALABORATORY $S_R$

Description of data:		Optional; uncertainty components <u>subtracted</u> from $s_{IR}$ to give $s_{IR:corr}$						MPN		
Too few Results		$s_{IR}$	<u>uncertainty components subtracted</u> from $s_{IR}$ to give $s_{IR:corr}$				$n_i$	$m_i$		
Standard uncertainty	$s_{IR:corr}$	Matrix	Poisson	Confirmation		$n_p$	$n_c$	$u_{conf}$	$u_{MPN}$	$x_i$
Laboratory Sample ID	Result log <sub>10</sub>	$u_{matrix}$	$\Sigma C$	$u_{Poisson}$						
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