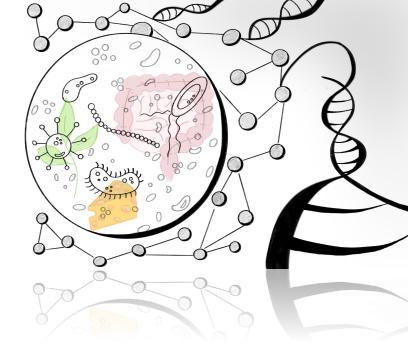
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In silico exploration of metabolism in microbial ecosystems:

from the metabolic network to the model



Clémence Frioux

8th EBAME Workshop

28/10/2023

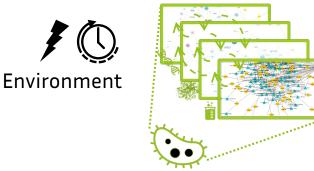
clemence.frioux@inria.f

Inria centre at the University of Bordeaux



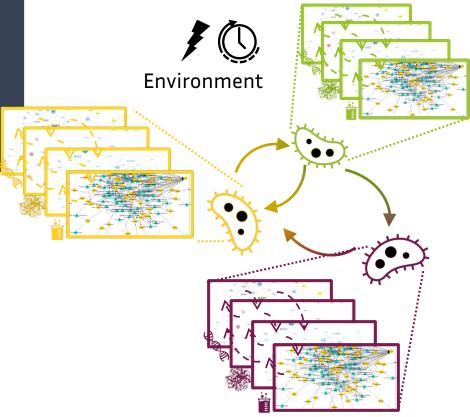
From (microbial) systems biology to systems ecology

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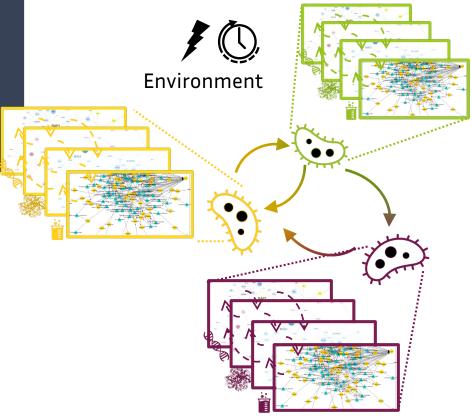


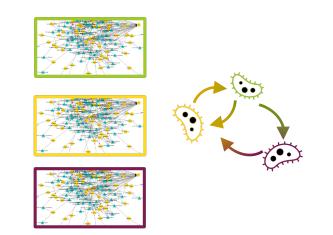
From (microbial) systems biology to systems ecology

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From (microbial) systems biology to systems ecology



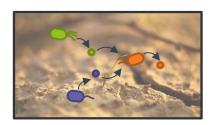


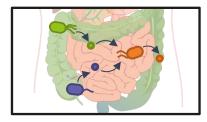
Inría-

Most interactions are mediated by metabolism

Integrating the biological data flood into models

Ecosystems







"Omics" data

"Who" is there? What can they do? What do they actually do? What molecules are present?

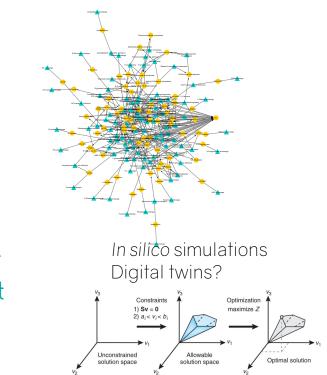
(Meta)genomics Metataxonomics

Model generation

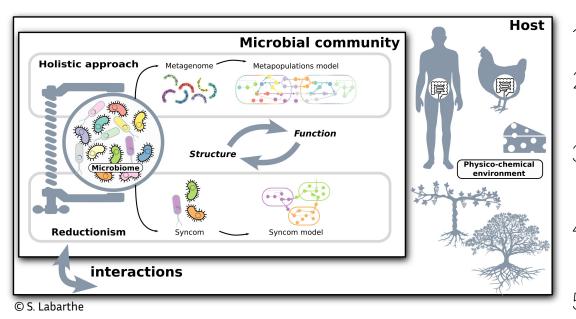
Metatranscriptomics Metabolomics...

Model refinement

Models



Outline



1. Metabolic networks

2. Reconstruction of metabolic networks

- 3. A discrete model of metabolism
- 4. Mining metabolic models of large communities
- 5. Tutorial hands on

Innin -

1. Metabolic networks

Ínría_

Genome-scale metabolic reconstruction of 7,302 human microorganisms for personalized medicine

Received: 9 November 2021 Accepted: 30 November 2022 Published online: 19 January 2023 Almut Heinken ©¹²³, Johannes Hertel¹⁴, Geeta Acharya⁵, Dmitry A. Ravcheev¹², Malgorzata Nyga⁶, Onyedika Emmanuel Okpala ©⁷, Marcus Hogan¹³, Stefania Magnúsdóttir ©⁸, Filippo Martinelli¹³, Bram Nap¹³, German Preciat ©⁹, Janaka N. Edirisinghe⁰³¹, Christopher S. Henry¹⁷, Ronan M. T. Fleming¹⁹ &

Personalized whole-body models integrate metabolism, physiology, and the gut microbiome

Ines Thiele^{1,2,3,4,*}⁽⁰⁾, Swagatika Sahoo^{4,†}, Almut Heinken¹⁽⁰⁾, Johannes Hertel^{1,5}, Laurent Heirendt^{4,0}, Maike K Aurich⁴ & Ronan MT Fleming^{1,6,*}⁽⁰⁾

RESEARCH ARTICLE

Predicting gastrointestinal drug effects using contextualized metabolic models

Marouen Ben Guebila¹, Ines Thiele©^{1,2,3}*

1 Luxembourg Centre for Systems Biomedicine, University of Luxembourg, Esch-sur-Alzette, Luxembourg, 2 School of Medicine, National University of Ireland, Galway, University Road, Galway, Ireland, 3 Discipline of Microbiology, School of Natural Sciences, National University of Ireland, Galway, University Road, Galway, Ireland

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Systematic assessment of secondary bile acid metabolism in gut microbes reveals distinct metabolic capabilities in inflammatory bowel disease Check for updates

Longitudinal flux balance analyses of a patient with episodic colonic inflammation reveals microbiome metabolic dynamics

Arianna Basile^{a,b}, Almut Heinken^{b,c}, Johannes Hertel^d, Larry Smarr^e, Weizhong Li^f, Laura Treu^a, Giorgio Valle^a, Stefano Campanaro^{a§}, and Ines Thiele^{[b,c,g,h§}

Almut Heinken¹, Dmitry A. Ravcheev¹, Federico Baldini², Laurent Heirendt², Ronan M. T. Fleming³ and Ines Thiele^{1,2,4}

Metabolic Network Analysis Reveals Altered Bile Acid Synthesis and Metabolism in Alzheimer's Disease

Bile enide in brain

Graphical Abstract

Altered cholesterol and bile acid metabolism in Alzheimer's disease Metabolic profile of Priyanka Baloni, Cory C. Funk, Jingwen Yan, ..., The Alzheimer's Disease Metabolomics Consortium, Rima Kaddurah-Daouk, Nathan D. Price

Authors

RESEARCH ARTICLE Predicting gastrointestinal drug effects using contextualized metabolic models

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Metabolic dependencies govern microbial syntrophies during methanogenesis in an anaerobic digestion ecosystem

Xinyu Zhu^{1,2}, Stefano Campanaro^{3,4}, Laura Treu^{1,3*}, Rekha Seshadri², Natalia Ivanova², Panagiotis G. Kougias^{1,5*}, Nikos Kyrpides² and Irini Angelidaki¹

Check for

Revealing metabolic mechanisms of interaction in the anaerobic digestion microbiome by flux balance analysis

Arianna Basile^a, Stefano Campanaro^{a,b,*}, Adam Kovalovszki^c, Guido Zampieri^{a,d}, Alessandro Rossi^a, Irini Angelidaki^c, Giorgio Valle^{a,1}, Laura Treu^{a,1}



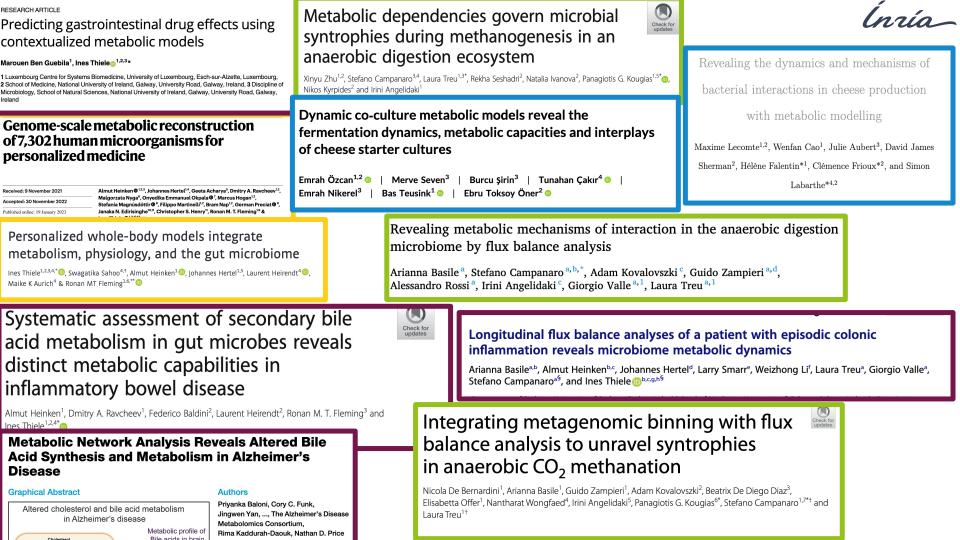
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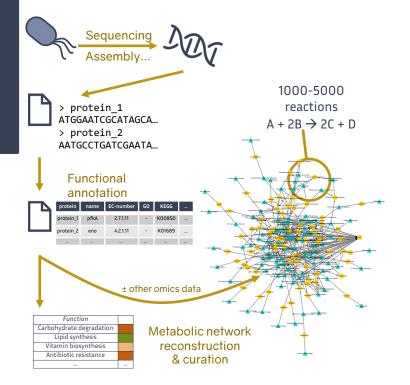
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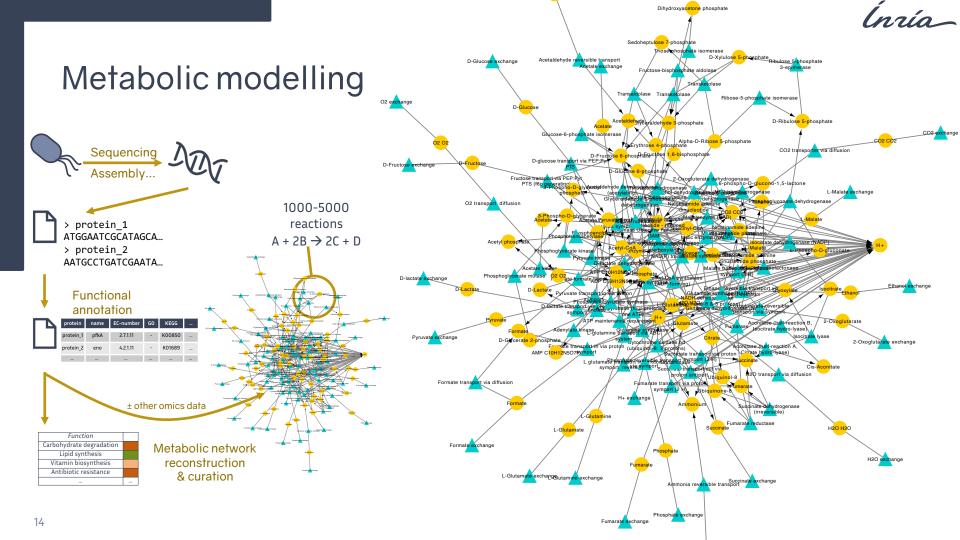
Integrating metagenomic binning with flux balance analysis to unravel syntrophies in anaerobic CO₂ methanation

Nicola De Bernardini¹, Arianna Basile¹, Guido Zampieri¹, Adam Kovalovszki², Beatrix De Diego Diaz³, Elisabetta Offer¹, Nantharat Wongfaed⁴, Irini Angelidaki⁵, Panagiotis G. Kougias^{6*}, Stefano Campanaro^{1,7*†} and Laura Treu^{1†}

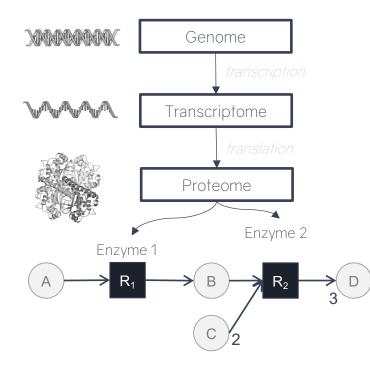


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Connecting genomes to functions

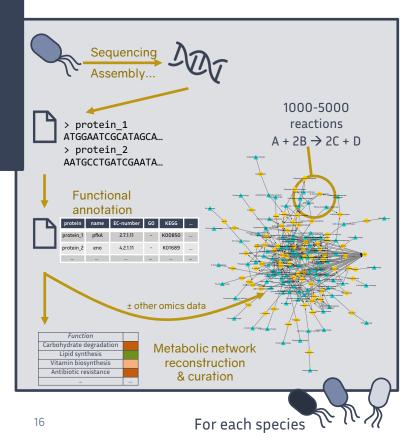


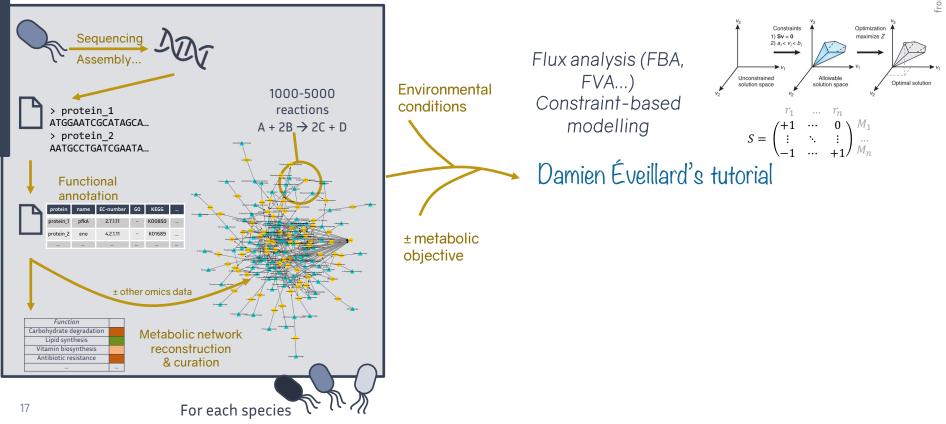
1 genome \rightarrow 1,000 to 5,000 reactions

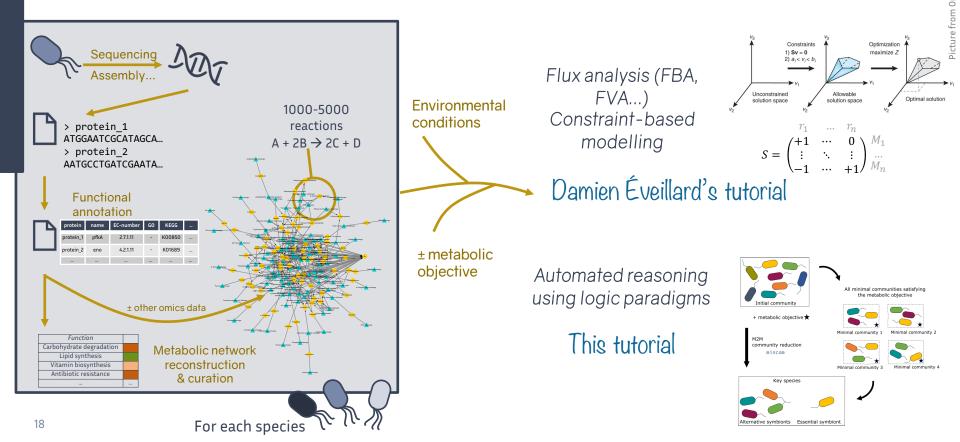
Abstraction of genomic information in genome-scale metabolic networks (GSMNs)

Functional repertoire of the cell

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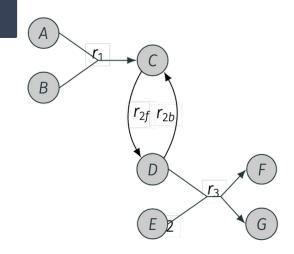




Innín 1

 $\begin{array}{ll} r_1: & A+B \to C \\ r_2: & C \leftrightarrow D \\ r_3: & D+2E \to F+G \end{array}$

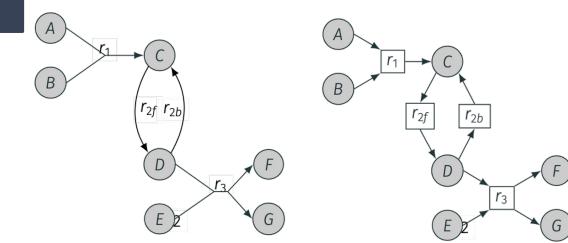
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20 Hypergraph

Innía

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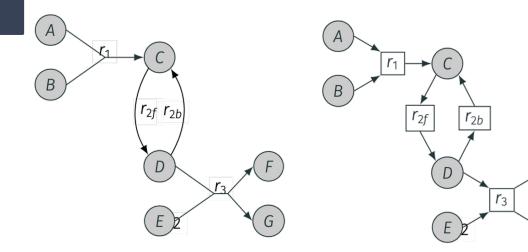


Hypergraph

21

Bipartite graph

 $\begin{array}{ll} r_1: & A+B \to C \\ r_2: & C \leftrightarrow D \\ r_3: & D+2E \to F+G \end{array}$



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Hypergraph

22

Bipartite graph

G

Stoichiometric matrix

Tutorial content

1. Metabolic network exploration:

- a. Visualisation of metabolic networks
- b. Exploration of the exchange format: SBML
- 2. Metabolic network reconstruction
 - a. Structural annotation with Prodigal
 - b. Reconstruction with gapseq
 - c. Reconstruction from Kegg knowledge base
 - d. Exploration of metabolic networks reconstructed with Pathway Tools
- 3. Metabolic network modelling of a unique species
 - a. Using toy data
 - b. Using real data
- 4. Metabolic network modelling of a community
 - a. Using toy data
 - b. Using real data

https://gitlab.inria.fr/cfrioux/ebame

2. Reconstruction of metabolic networks

Input data

- minimal input: a genome or a proteome

- a reference genome
- a Metagenomic-Assembled Genome (MAG)



- Genomes must be structurally annotated

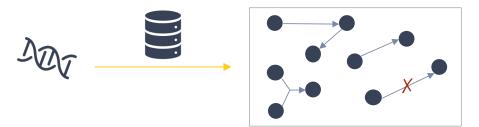
e.g. with Prodigal [Hyatt et al 2010] for prokaryotes

- Some tools require also a functional annotation

e.g.with EggNOG mapper [Cantalapiedra et al 2021] or Prokka [Seeman 2014]

Metabolic network inference: bottom-up procedure

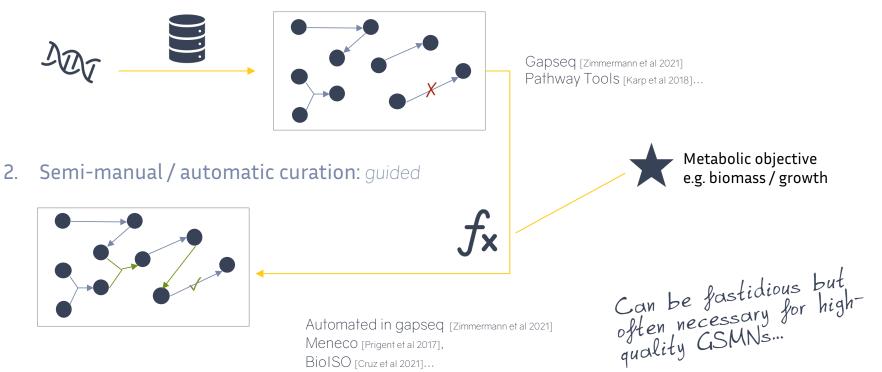
1. Automatic inference from sequences: non-guided



Gapseq [Zimmermann et al 2021] Pathway Tools [Karp et al 2018]...

Metabolic network inference: bottom-up procedure

1. Automatic inference from sequences: non-guided

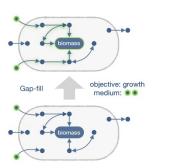


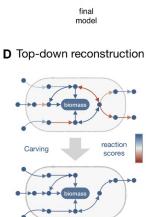
Alternative: top-down reconstruction

A Classic reconstruction workflow

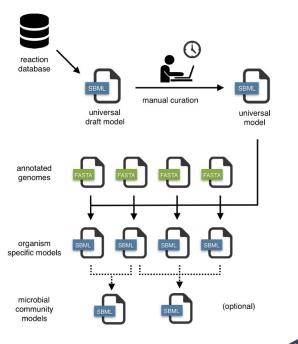
reaction database

C Bottom-up reconstruction





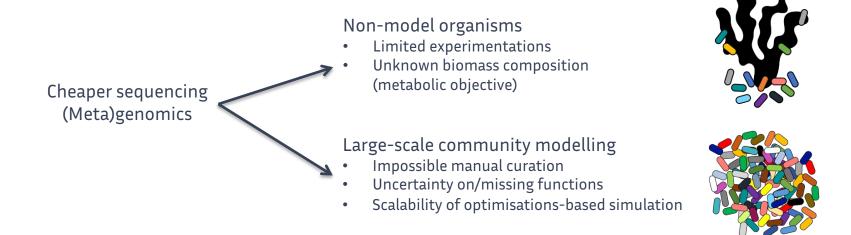
B CarveMe reconstruction workflow



Metabolic models ready for constraint-based simulation \leq

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A rise in non-optimal use-cases



→ The choice of the mathematical/computational model for simulation must account for the data and the question to be answered

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Resources for (draft) reconstruction

Functional annotation

- eggNOG mapper [Cantalapiedra et al 2021]
- Prokka [Seeman 2014]

Tools scalable to microbial communities

- Pathway Tools [Karp et al 2019]
- Carveme [Machado et al 2018]
- gapseq [Zimmermann et al 2021]
- AutoKEGGRec [Karlsen et al 2018]

Collections of reconstructions

- KEGG [Kanehisa et al 2021]
- BioCyc [Karp et al 2019]
- Bigg [King et al 2016]
- Biomodels [Malik-Sheriff et al 2019]
- Virtual metabolic human [Noronha et al 2018]
- AGORA 2 [Heinken et al 2023]
- EMBL GEMs [Machado et al 2018]

A network, then what?

Exploration

- reactions, metabolic pathways, association of genes to reactions
- visualisation (usually complicated)

Modelling

- predicting the behaviour of the microbe/community in given conditions
- finding members of interest within the community
- screen the metabolic potential

Tutorial content

1. Metabolic network exploration:

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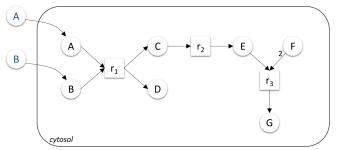
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https://gitlab.inria.fr/cfrioux/ebame

3. A discrete model of metabolism

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Boolean abstraction of metabolic producibility



extracellular

Network expansion [Ebenhöh et al 2004]

- → Scope of seeds
- \rightarrow Discrete modelling

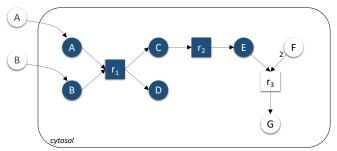
$$scope(G, S) = \bigcup_{i} M_{i},$$

where $M_{0} = S$ and
 $M_{i+1} = M_{i} \cup products(\{r \in R | reactants(r) \subseteq M_{i}\})$

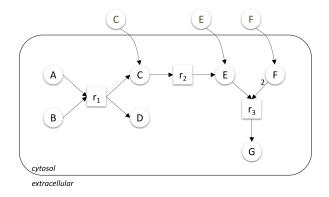
Qualitative simulation of metabolic producibility ignoring stoichiometric coefficients and w/o objective function

Ínría_

Boolean abstraction of metabolic producibility



extracellular



Network expansion [Ebenhöh et al 2004]

- → Scope of seeds
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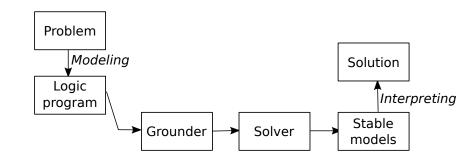
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Qualitative simulation of metabolic producibility ignoring stoichiometric coefficients and w/o objective function

[Gebser et al 2012]

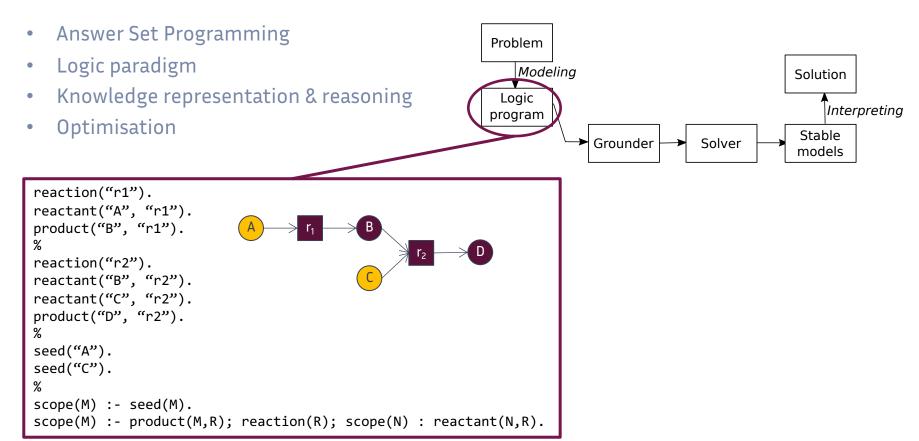
A flexible implementation system

- Answer Set Programming
- Logic paradigm
- Knowledge representation & reasoning
- Optimisation

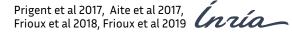


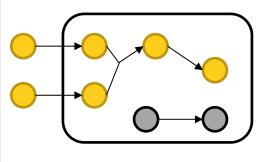
A flexible implementation system

[Gebser et al 2012]



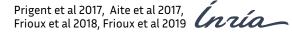
Applications of the Boolean abstraction of producibility

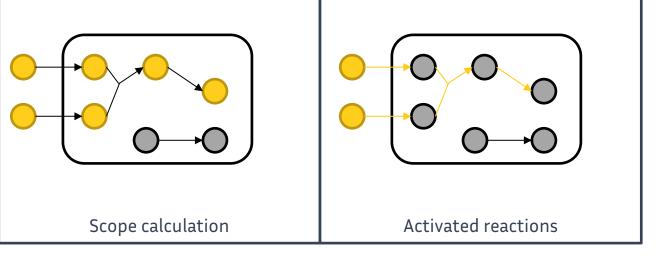




Scope calculation

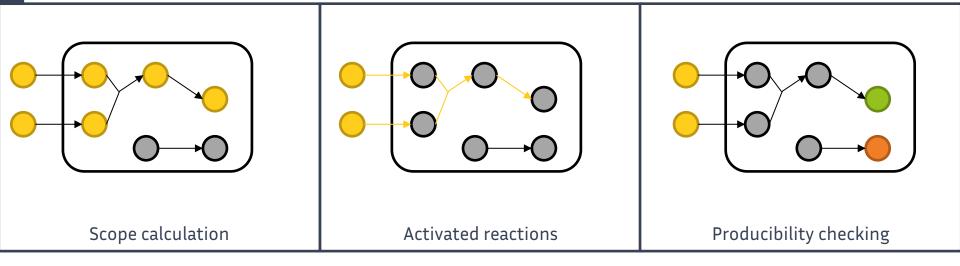
Applications of the Boolean abstraction of producibility





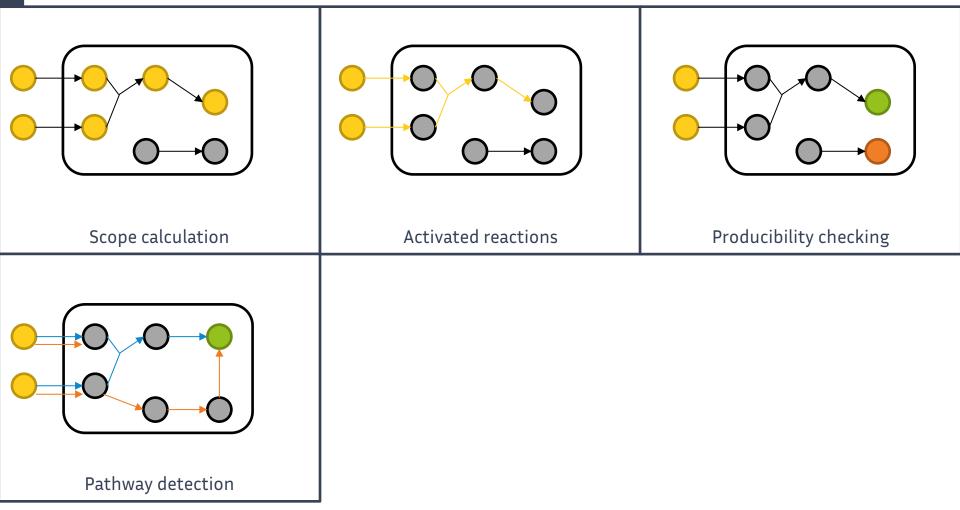
Applications of the Boolean abstraction of producibility

Prigent et al 2017, Aite et al 2017, Frioux et al 2018, Frioux et al 2019 **Única**



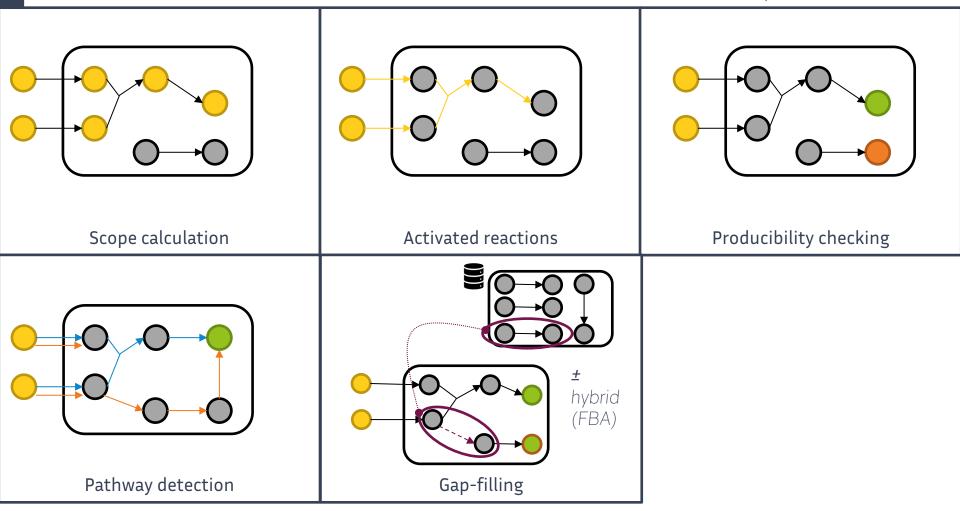
Applications of the Boolean abstraction of producibility

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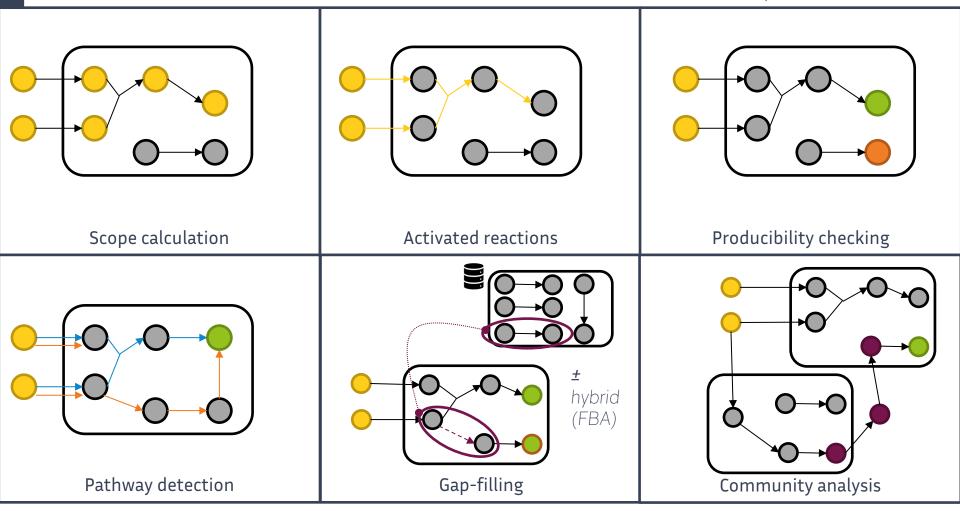
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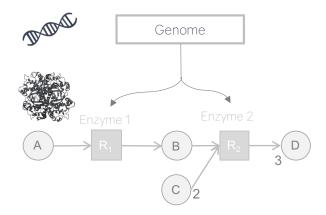
Applications of the Boolean abstraction of producibility

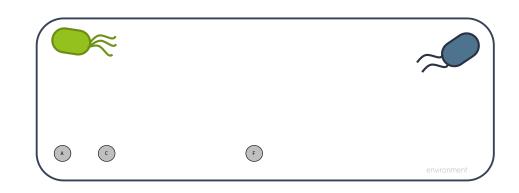
Prigent et al 2017, Aite et al 2017, Frioux et al 2018, Frioux et al 2019



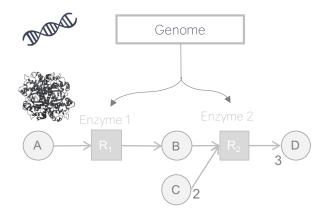
4. Mining metabolic models of large communities

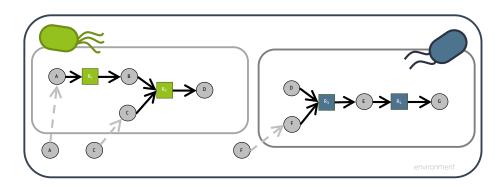
Scale, Screen, Simplify





1 genome → 1,000 to 5,000 reactions Genome-scale metabolic network (GSMN) **Functional** repertoire of the cell

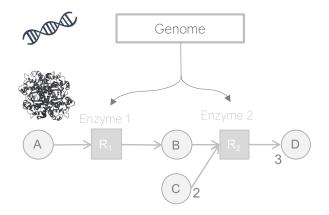




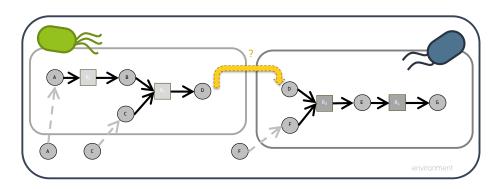
1. Reconstruct GSMNs for all genomes

1 genome \rightarrow 1,000 to 5,000 reactions Genome-scale metabolic network (GSMN)

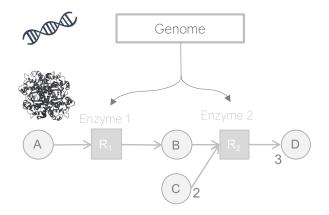
Functional repertoire of the cell



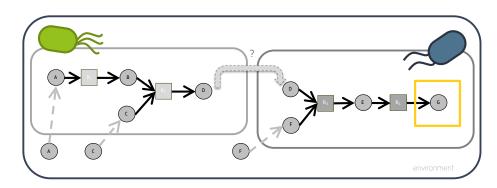
1 genome → 1,000 to 5,000 reactions Genome-scale metabolic network (GSMN) **Functional** repertoire of the cell



- 1. Reconstruct GSMNs for all genomes
- 2. Assess putative complementarity between them



1 genome → 1,000 to 5,000 reactions Genome-scale metabolic network (GSMN) **Functional** repertoire of the cell



- 1. Reconstruct GSMNs for all genomes
- 2. Assess putative complementarity between them
- 3. Identify key species associated to a function

Microbiota

- MAGs

- Reference genomes

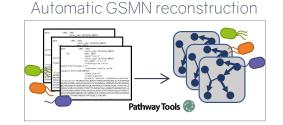
Github Aureme/Metage2Metabo

> [Belcour*, Frioux* et al. eLife 2020] 4[Karp et al. Bioinformatics 2002] [Frioux et al. Bioinformatics 2018]



Microbiota

- MAGs
- Reference genomes



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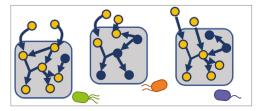


Microbiota

- MAGs
- Reference genomes

Automatic GSMN reconstruction

Individual metabolic capabilities



Github Aureme/Metage2Metabo

[Belcour*, Frioux* et al. eLife 2020] 5[Karp et al. Bioinformatics 2002] [Frioux et al. Bioinformatics 2018]



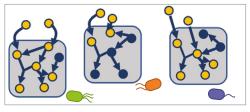
Microbiota

- MAGs
- Reference genomes

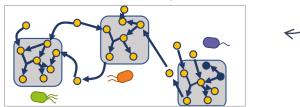
Automatic GSMN reconstruction



Individual metabolic capabilities



Collective metabolic capabilities





Systematic screening of **metabolic potential** and **mutualistic** potential in a microbiota

Github Aureme/Metage2Metabo

[Belcour*, Frioux* et al. eLife 2020] 5[Karp et al. Bioinformatics 2002] [Frioux et al. Bioinformatics 2018]

Microbiota

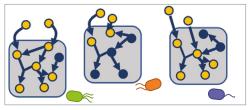
- MAGs
- Reference genomes



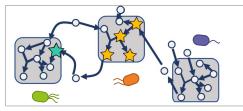
Pathway Tools 🔇



Individual metabolic capabilities



Added value of cross feeding

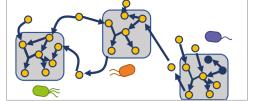


Github Aureme/Metage2Metabo

[Belcour*, Frioux* et al. eLife 2020] 5[Karp et al. Bioinformatics 2002] [Frioux et al. Bioinformatics 2018]



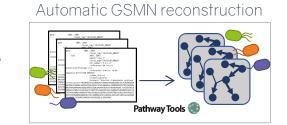
Collective metabolic capabilities



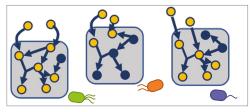


Microbiota

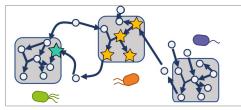
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Individual metabolic capabilities

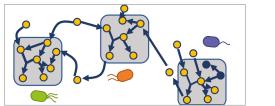


Added value of cross feeding

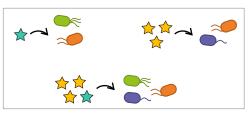




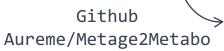
Collective metabolic capabilities



Minimal community selection



Systematic screening of **metabolic potential** and **mutualistic** potential in a microbiota



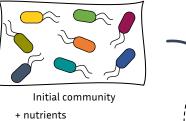
[Belcour*, Frioux* et al. eLife 2020] 5[Karp et al. Bioinformatics 2002] [Frioux et al. Bioinformatics 2018]

[Belcour*, Frioux et al. eLife 2020] [Frioux et al. Bioinformatics 2018]



Key species

- Minimal community selection: combinatorial problem solving
- However... 1 solution (= 1 minimal community) among millions? **Huge combinatorics**
- *Key species* = species found in at least one of the predicted minimal communities (MC)
 - > Essential symbionts : species predicted in every MC
 - > Alternative symbionts : species predicted is some but not all MC



+ metabolic objective ★



Minimal community

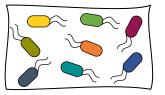
The concept of key species addresses the combinatorial challenge brought by functional redundancy in microbiomes

[Belcour*, Frioux et al. eLife 2020] [Frioux et al. Bioinformatics 2018]



Key species

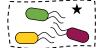
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- Initial community
- + nutrients + metabolic objective ★



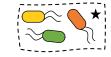
All minimal communities satisfying the metabolic objective



Minimal community 1

Minimal community 2





Minimal community 3

Minimal community 4

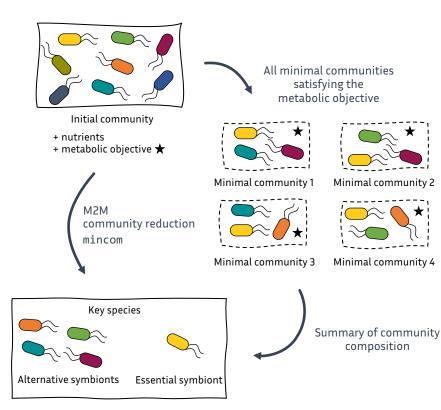
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Key species

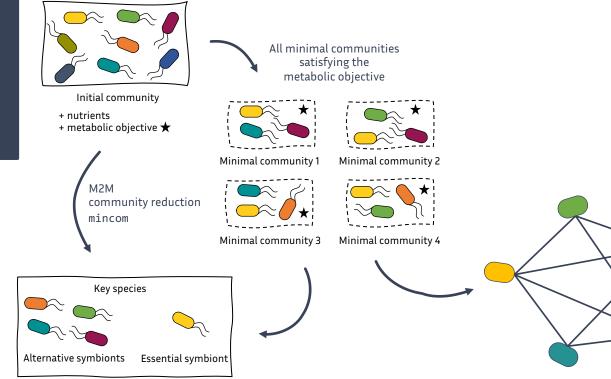
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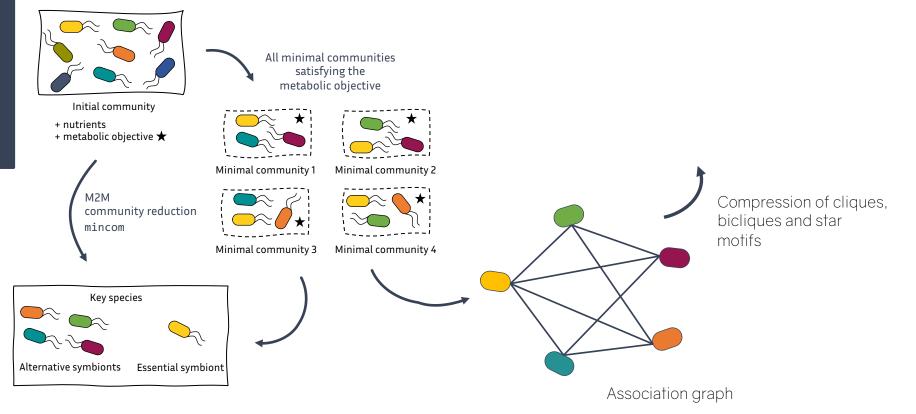
From key species to association analyses



Association graph

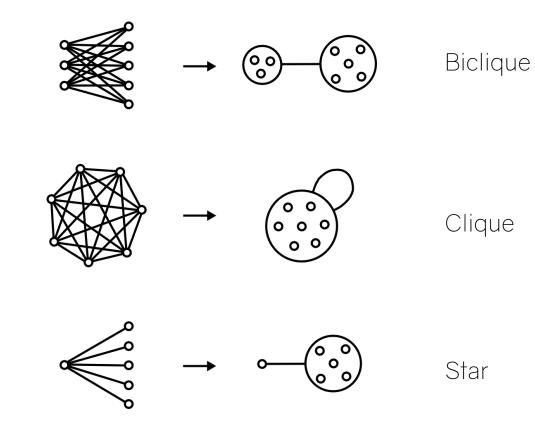
[Belcour*, Frioux et al. eLife 2020] [Frioux et al. Bioinformatics 2018]

From key species to association analyses



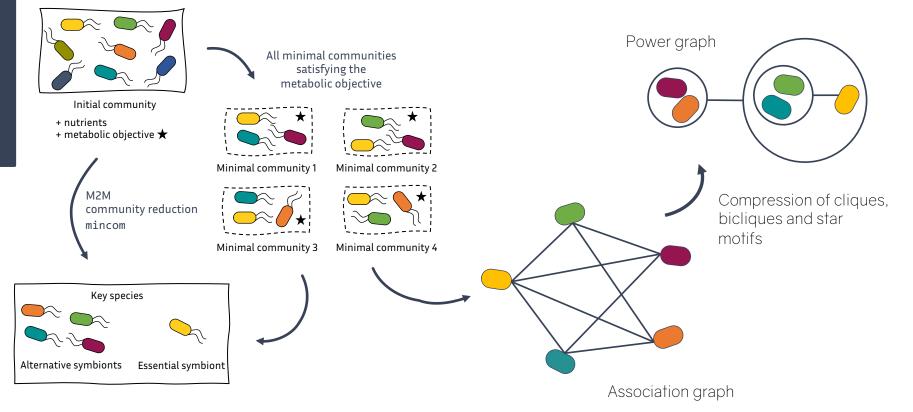
[Royer et al. PLoS CB 2008]

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From key species to association analyses

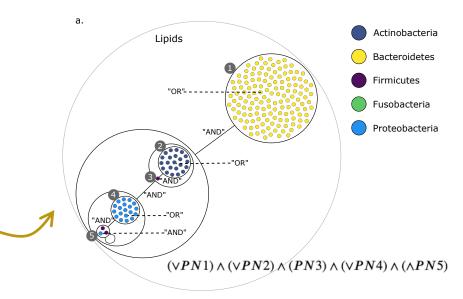


Key species by groups of metabolic end-products

| | | Firm. | Bact. | Acti. | Prot. | Fuso. | total |
|--|----|-------|-------|-------|-------|-------|-------|
| aminoacids and derivatives (5 targets) | KS | 142 | 52 | 0 | 27 | 6 | 227 |
| 4 bact. per community | ES | 0 | 0 | 0 | 0 | 0 | 0 |
| 120,329 communities | AS | 142 | 52 | 0 | 27 | 6 | 227 |
| aromatic compounds (11 targets) | KS | 52 | 0 | 0 | 20 | 0 | 72 |
| 5 bact. per community | ES | 2 | 0 | 0 | 1 | 0 | 3 |
| 950 communities | AS | 50 | 0 | 0 | 19 | 0 | 69 |
| carboxyacids (14 targets) | KS | 16 | 13 | 0 | 28 | 2 | 59 |
| 9 bact. per community | ES | 2 | 0 | 0 | 2 | 0 | 4 |
| 48,412 communities | AS | 14 | 13 | 0 | 26 | 2 | 55 |
| coA derivatives (10 targets) | KS | 106 | 0 | 50 | 17 | 1 | 174 |
| 5 bact. per community | ES | 0 | 0 | 0 | 0 | 1 | 1 |
| 95,256 communities | AS | 106 | 0 | 50 | 17 | 0 | 173 |
| lipids (28 targets) | KS | 3 | 140 | 22 | 20 | 0 | 185 |
| 7 bact. per community | ES | 3 | 0 | 0 | 1 | 0 | 4 |
| 58,520 communities | AS | 0 | 140 | 22 | 19 | 0 | 181 |
| sugar derivatives (58 targets) | KS | 11 | 30 | 78 | 23 | 0 | 142 |
| 11 bact. per community | ES | 5 | 0 | 0 | 0 | 0 | 5 |
| 7,860,528 communities | AS | 6 | 30 | 78 | 23 | 0 | 137 |

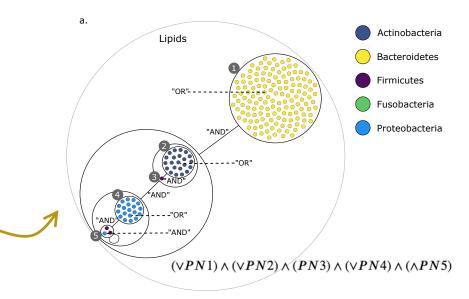
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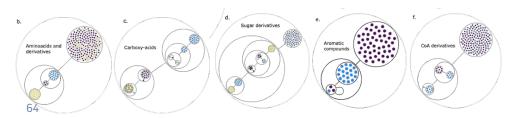
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Key species by groups of metabolic end-products

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Visualisation of key species and their association in minimal communities highlights groups of equivalent roles in the community

Innía -

5. Tutorial

https://gitlab.inria.fr/cfrioux/ebame

Tutorial content

1. Metabolic network exploration:

- a. Visualisation of metabolic networks
- b. Exploration of the exchange format: SBML

2. Metabolic network reconstruction

- a. Structural annotation with Prodigal
- b. Reconstruction with gapseq
- c. Reconstruction from Kegg knowledge base
- d. Exploration of metabolic networks reconstructed with Pathway Tools

3. Metabolic network modelling of a unique species

- a. Using toy data
- b. Using real data

4. Metabolic network modelling of a community

- a. Using toy data
- b. Using real data

https://gitlab.inria.fr/cfrioux/ebame

Acknowledgements



Discrete modelling of metabolism

∮[§]IRISA



INRAO Université BORDEAUX Quadram Distitute Science + Health + Food + Innovation Earlham Institute Decoding Living Systems

- Arnaud Belcour
- Méziane Aite

CNrs

- Anne Siegel
- Chabname Ghassemi Nedjad
- Maxime Lecomte
- Coralie Muller

- Falk Hildebrand

EBAME organisation team

- Lois Maignien
- Stéphanie Renard
- Christophe Blanchet







