

Viromics: what, how, why?

Evelien Adriaenssens
Andrea Telatin

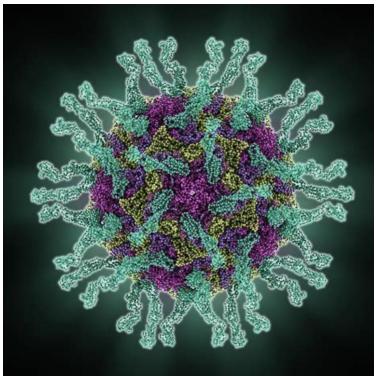


What is a virus?

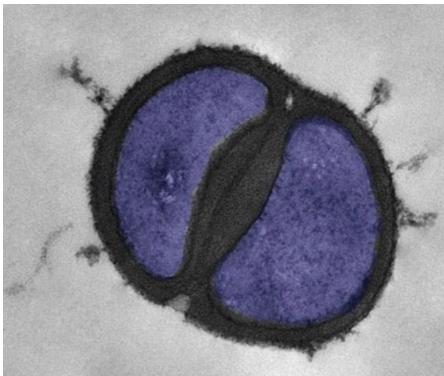


A virus or not a virus?

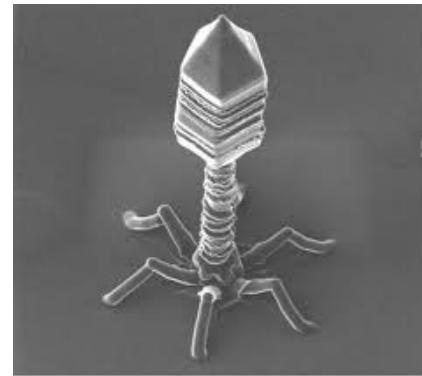
A



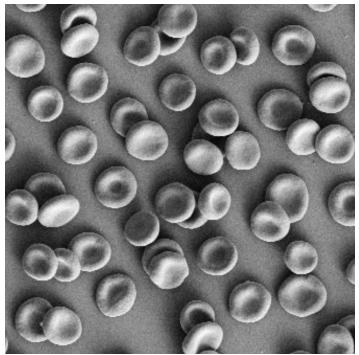
B



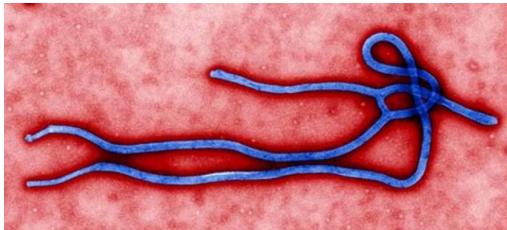
C



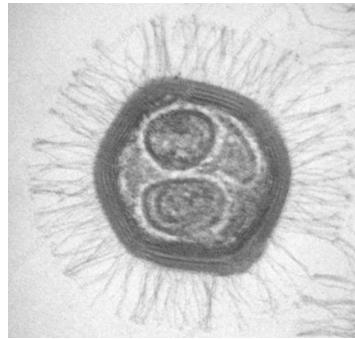
D



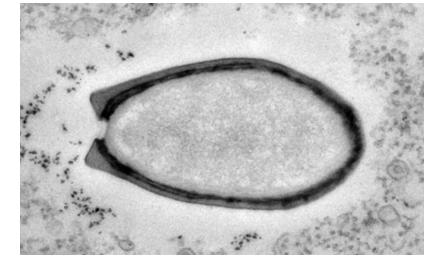
E



F

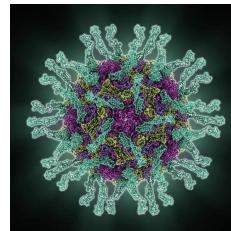


G

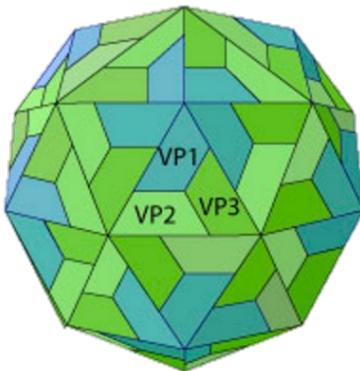
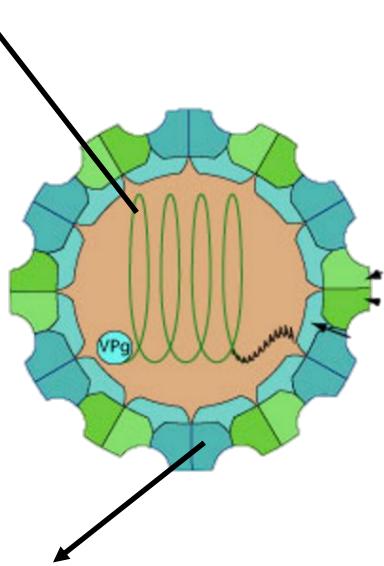


<https://tinyurl.com/ebame-virus>

Example A: poliovirus



genetic information as single segment of RNA

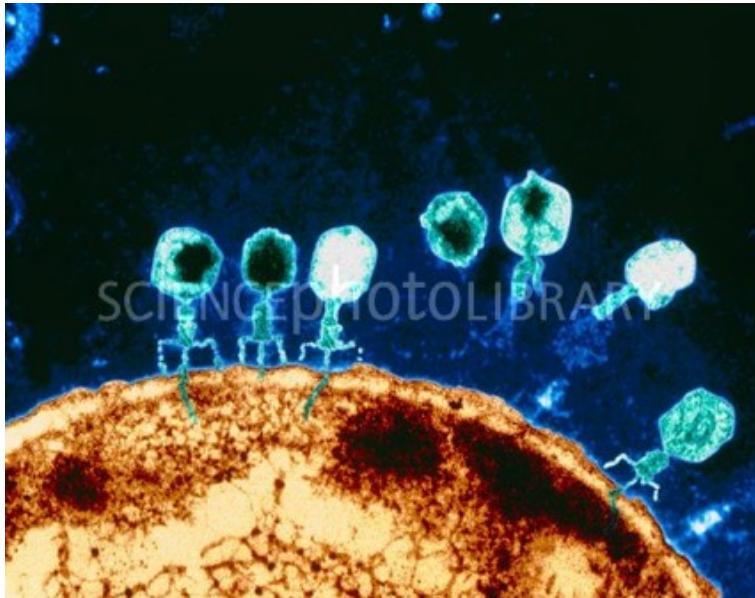
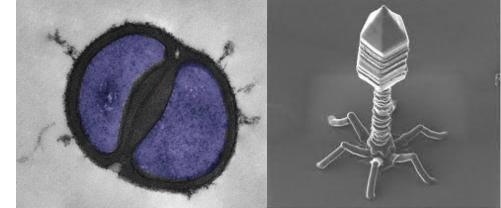


(+)ssRNA molecule
enteroviruses 7-
8.5 kb
polyprotein

capsid or coat made of different proteins

viralzone.expasy.org

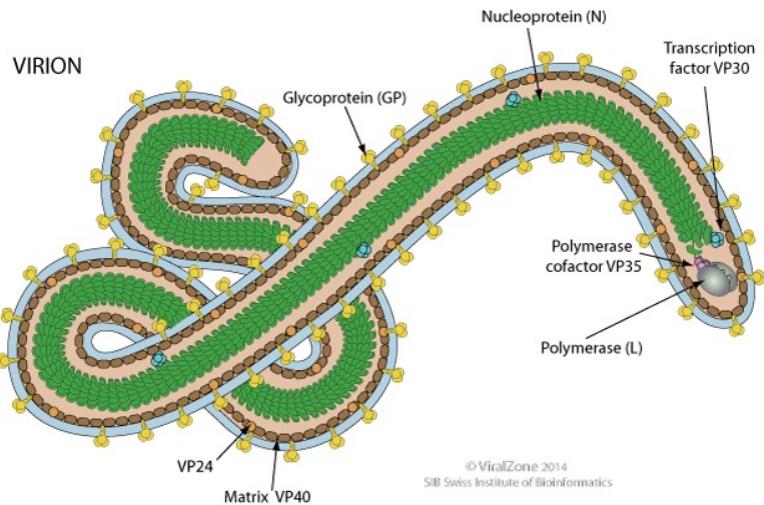
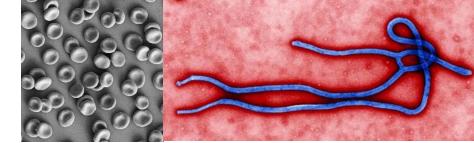
Examples B & C: *Staphylococcus* cell and bacteriophage



virus = bacteriophage
most commonly detected virus in metagenomics

Tailed bacteriophages (*Caudoviricetes*)
protein coat + genetic material (dsDNA)
linear dsDNA molecules
can be circularly permuted, have cohesive overhangs, or terminal repeats
~15 - > 500 kb

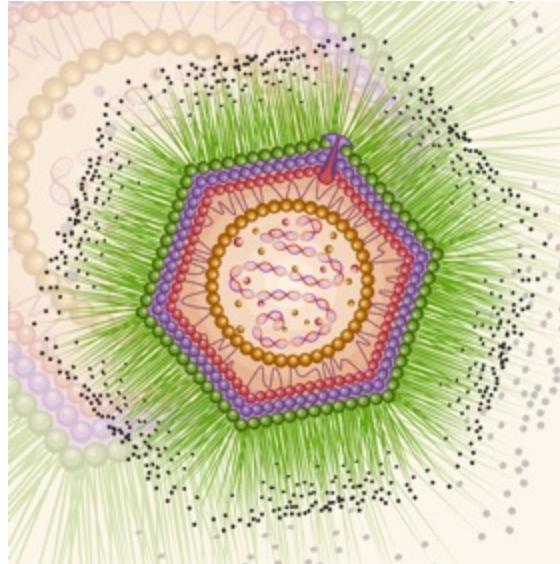
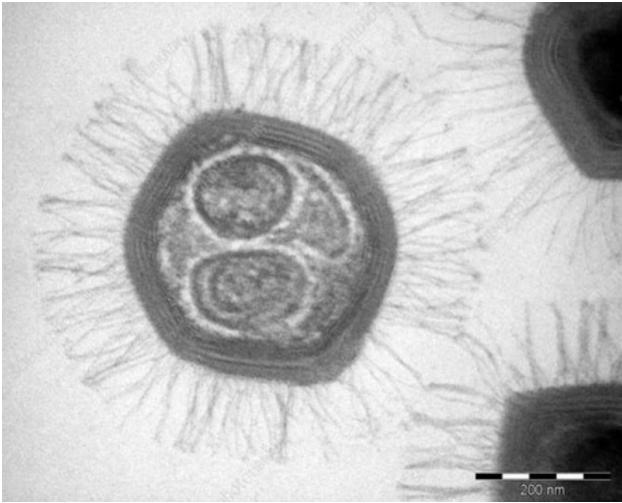
Examples D & E: red blood cells and ebolavirus



very large virion (970 nm long)
(-)ssRNA genome
18-19 kb

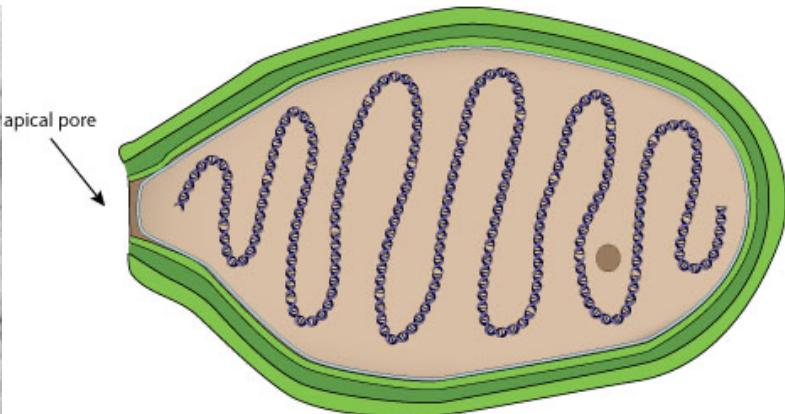
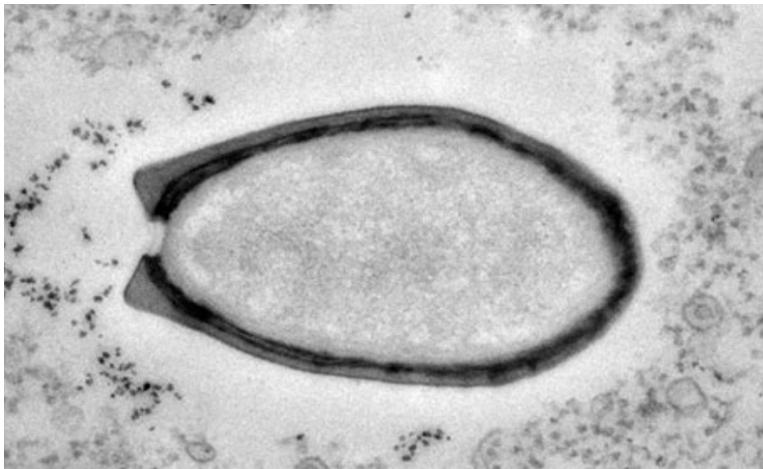


Example F: Mimivirus



Giant virus structure (visible with normal light microscope)
dsDNA genome, linear molecule, *Mimiviridae* genomes 350 kb – 1.2 Mb

Example G: Pandoravirus



particle 1 μm long

Pandoravirus salinus has the largest known virus genome: 2.4 Mb
linear dsDNA

How does this diversity impact
viromics or viral metagenomics?

Problems!

Everything you do creates a bias

Different particle sizes: 20 nm - > 1 μ m

Different capsid structures

Different genome types: ssDNA, dsDNA, (+/-)ssRNA, dsRNA

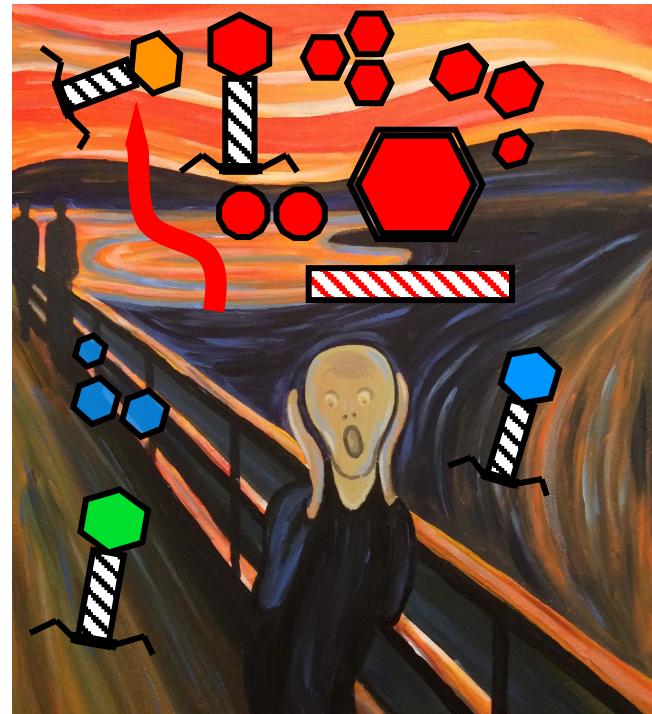
Different genome sizes: ~1,300 - > 2,000,000 nt

Segmented genomes

Linear or circular

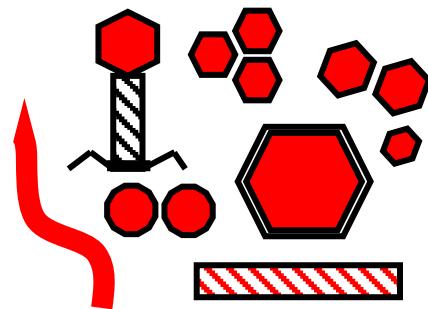
No conserved marker gene across the virosphere

No shared evolutionary origin

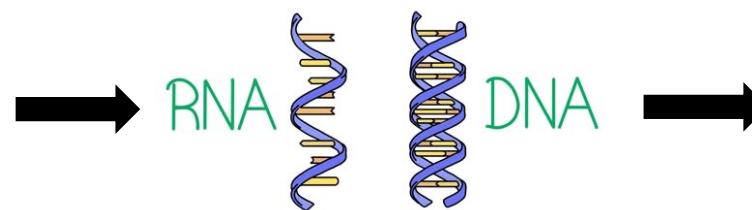


BUT, majority of viruses <200 nm and genomes <200 kb

Viromics



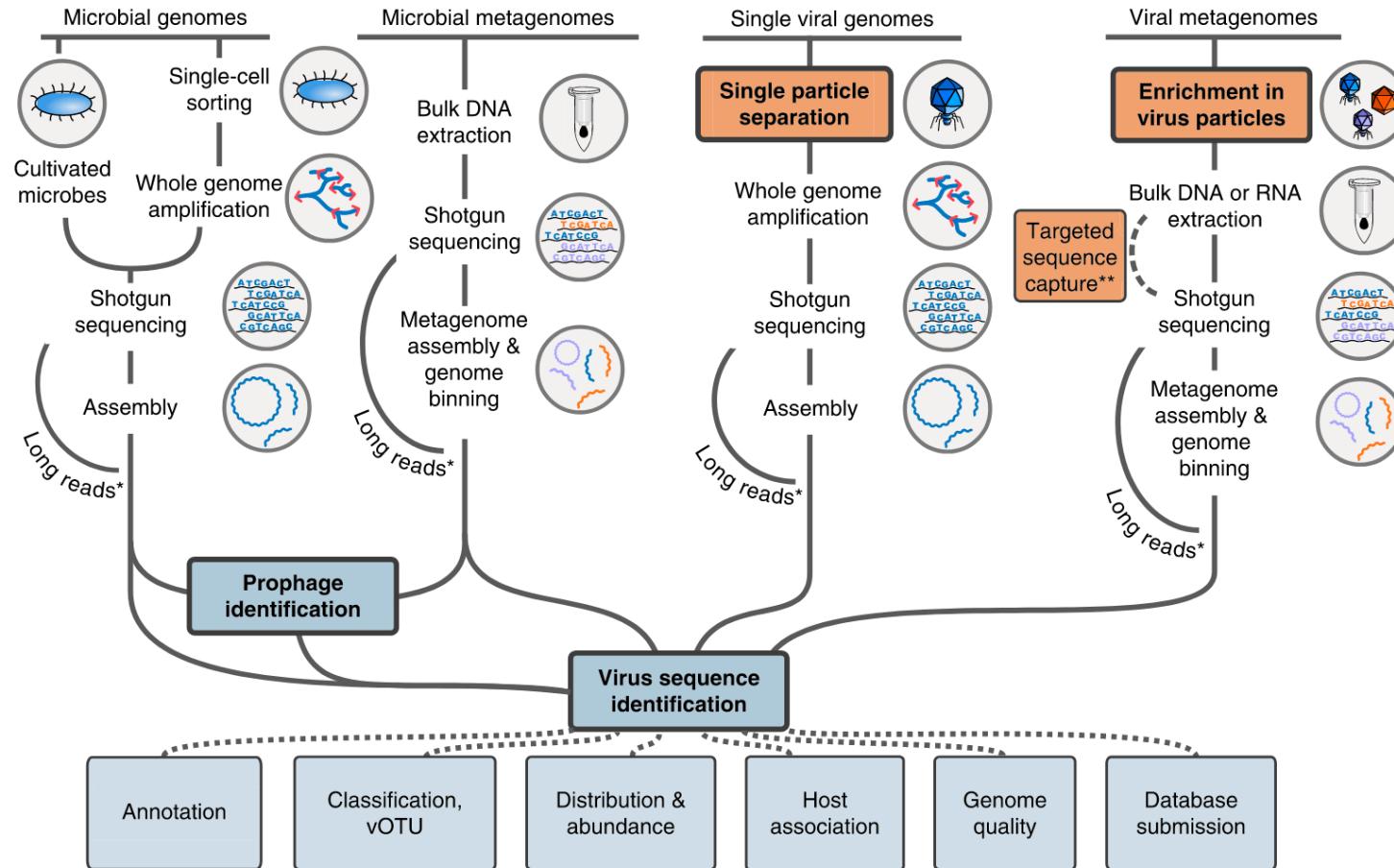
separate virus community
from cellular material



nucleic acid extraction



sequencing



Quick & “dirty” analyses: read profiling

Phage-inclusive profiling of human gut microbiomes with Phanta

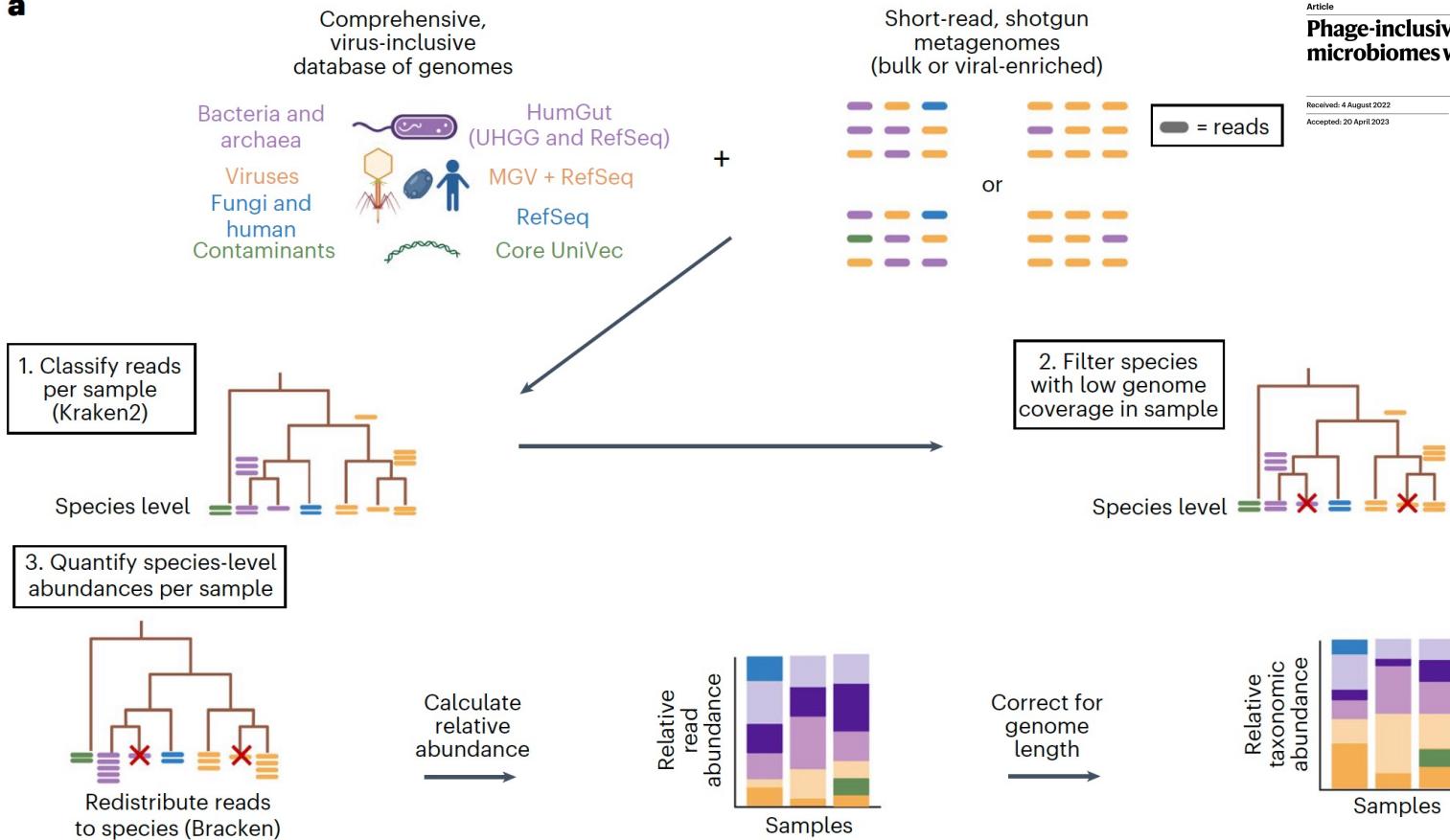
Received: 4 August 2022

Yishay Pinto^{1,2,3}, Meenakshi Chakraborty   ^{1,3}, Navami Jain^{1,2} & Ami S. Bhatt   ^{1,2}

Accepted: 20 April 2023

Published online: 25 May 2023

Due to technical limitations, most gut microbiome studies have focused

a

Article

Phage-inclusive profiling of human gut microbiomes with Phanta

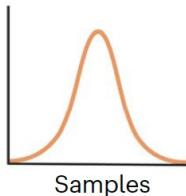
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Yishay Pinto^{1,2}, Meenakshi Chakraborty^{1,2}, Navami Jain^{1,2} & Ami S. Bhatt^{1,2,3}

4. Postprocessing: determine cross-domain relationships

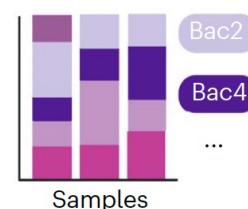
Viral
lifestyle
statistics



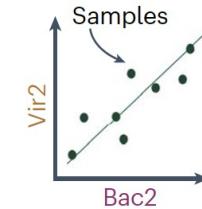
Identification of
likely integrated
prophages



Viral abundances
collapsed by
predicted host



Cross-domain
abundance
correlations



b

1:1
Bacteria:Virus
Species



27.5:1
Bacteria:Virus
Genome length



1:1
Bacteria:Virus
Genome coverage



Correct relative
read abundance for
genome length



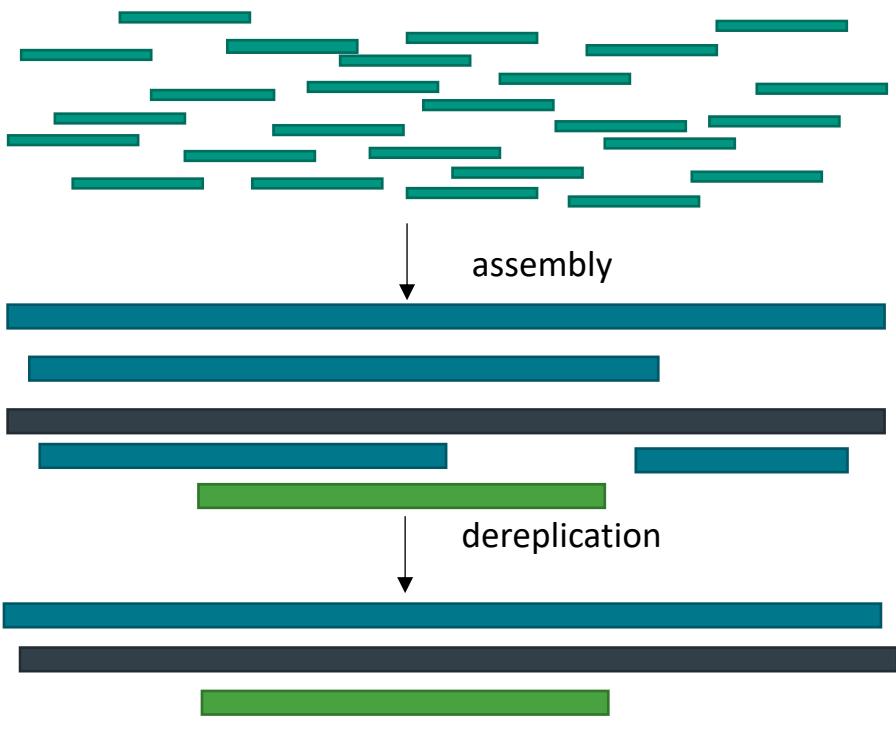
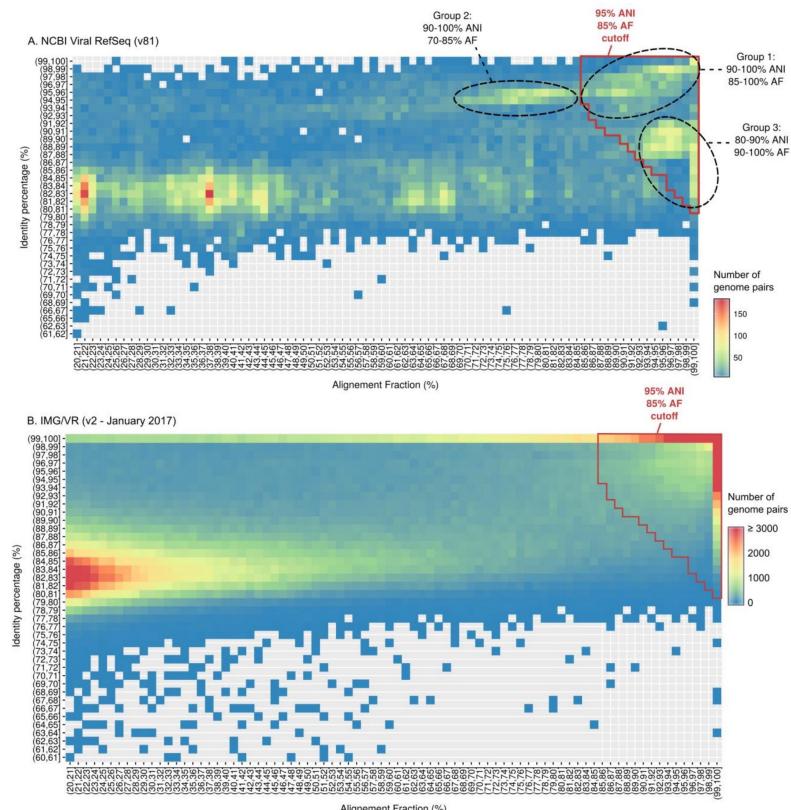
T4 phage



Bacteria Viruses
Taxonomic abundance

Gold standard: vOTU approach

What is a vOTU?



Viromes:
>95% ANI over >85% of alignment: vOTU
~ tailed phage species

What if not all contigs are viral?

→ virus mining

< PeerJ

VirSorter: mining viral signal from microbial genomic data

Research article

Bioinformatics

Genomics

Microbiology

Virology

Simon Roux^{1,*}, Francois Enault^{2,3}, Bonnie L. Hurwitz⁴, Matthew B. Sullivan✉^{1,*}

Published May 28, 2015

METHODOLOGY

Open Access

VirFinder: a novel k -mer based tool for identifying viral sequences from assembled metagenomic data

Jie Ren^{1†}, Nathan A. Ahlgren^{2,4*}, Yang Young Lu¹, Jed A. Fuhrman² and Fengzhu Sun^{1,3*}

Quantitative Biology 2020, 8(1): 64–77
<https://doi.org/10.1007/s40484-019-0187-4>



CrossMark

RESEARCH ARTICLE

Identifying viruses from metagenomic data using deep learning

Jie Ren^{1,1*}, Kai Song^{2,†}, Chao Deng¹, Nathan A. Ahlgren³, Jed A. Fuhrman⁴, Yi Li⁵, Xiaohui Xie⁵, Ryan Poplin⁶, Fengzhu Sun^{1,*}

METHODOLOGY

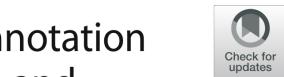
Open Access

VIBRANT: automated recovery, annotation and curation of microbial viruses, and evaluation of viral community function from genomic sequences

Kristopher Kieft, Zhichao Zhou and Karthik Anantharaman*

Guo et al. *Microbiome* (2021) 9:37
<https://doi.org/10.1186/s40168-020-00990-y>

Microbiome



SOFTWARE ARTICLE

Open Access

VirSorter2: a multi-classifier, expert-guided approach to detect diverse DNA and RNA viruses



Jiarong Guo¹, Ben Bolduc¹, Ahmed A. Zayed¹, Arvind Varsani^{2,3}, Guillermo Dominguez-Huerta¹, Tom O. Delmont⁴, Akbar Adjie Pratama¹, M. Consuelo Gazitúa⁵, Dean Vik¹, Matthew B. Sullivan^{1,6,7*} and Simon Roux^{8*}



Article

<https://doi.org/10.1038/s41587-023-01953-y>

Identification of mobile genetic elements with geNomad

Received: 6 March 2023

Accepted: 17 August 2023

Published online: 21 September 2023

Antonio Pedro Camargo , Simon Roux¹, Frederik Schulz ,
Michal Babinski², Yan Xu , Bin Hu², Patrick S. G. Chain , Stephen Nayfach ,
& Nikos C. Kyripides

input data: assemblies

virus mining magic

outputs:
list of contigs
list of probable contigs
fasta files with predictions

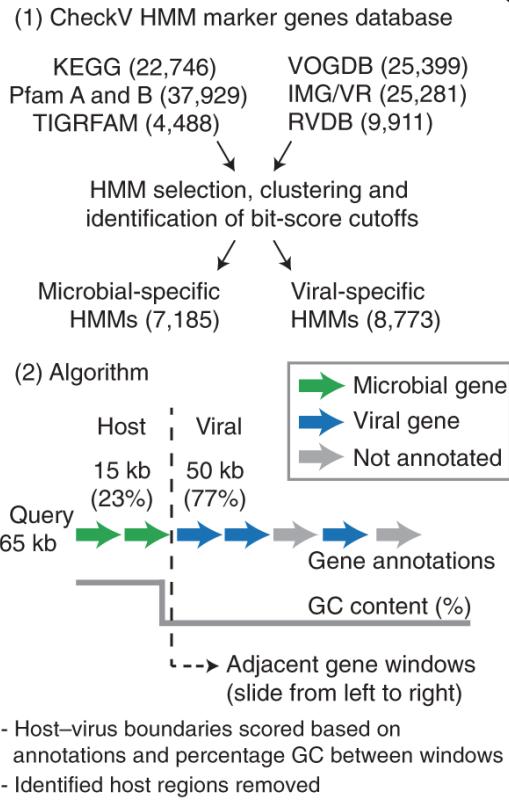
How do we know the virus genomes are complete?

Functional potential, host prediction, taxonomic classification*, diversity & distribution*	New taxonomic groups	New reference species
Finished genome Complete genome with extensive annotation		
High-quality draft genome Predicted ≥90% complete		
Genome fragment(s) Predicted <90% complete or no estimated genome size		

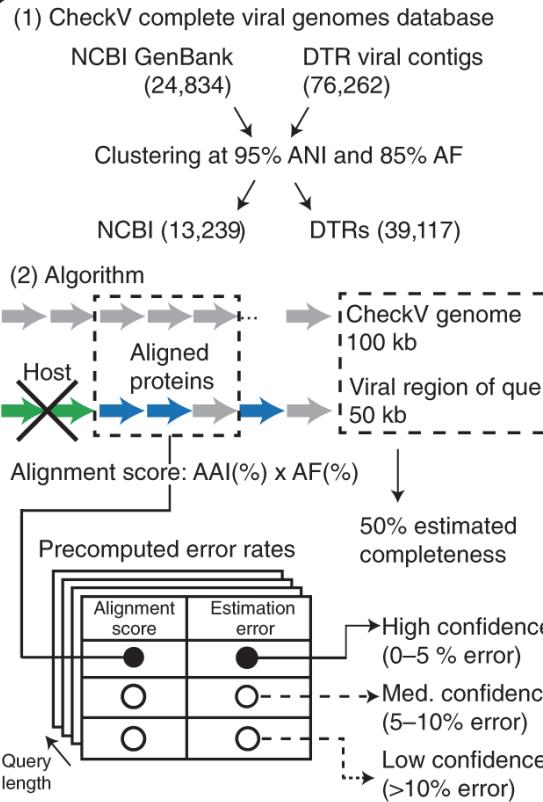


CheckV

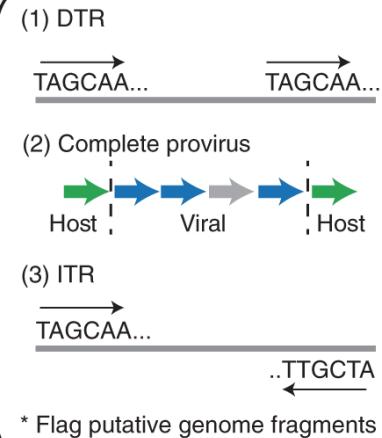
a Remove host contamination



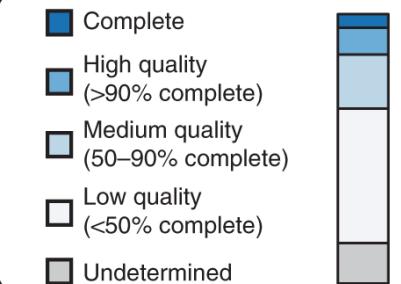
b Estimate genome completeness



c Predict closed genomes



d Summarize quality

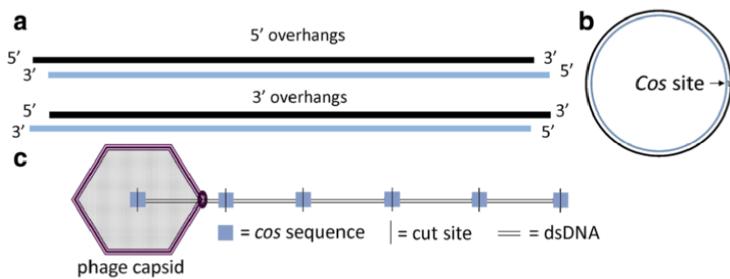


<https://bitbucket.org/berkeleylab/checkv/src/master/>

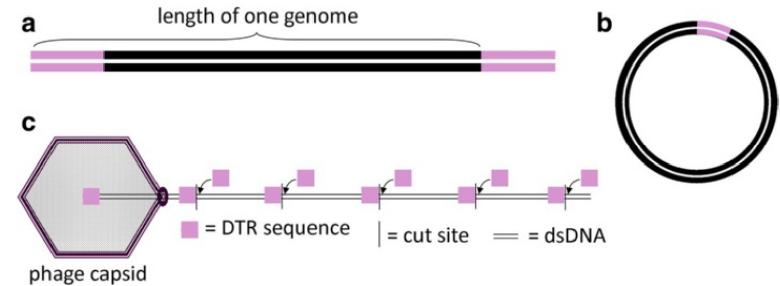
Nayfach et al, 2020, Nature Biotechn

Phage genome examples: defined ends

Cohesive ends

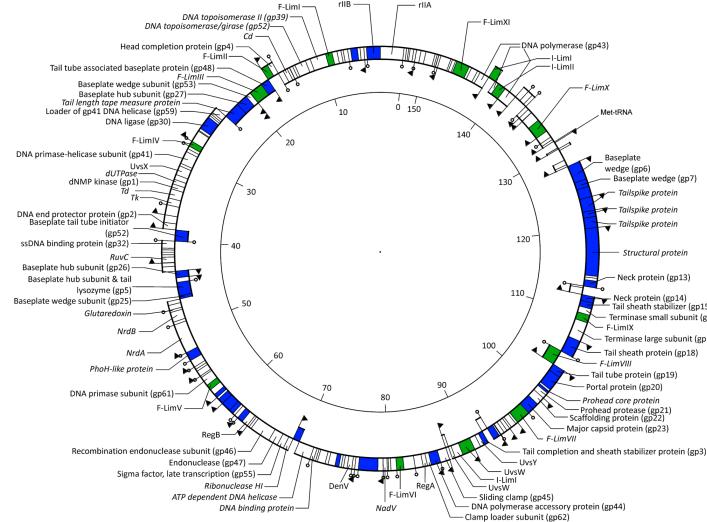


Terminal repeats

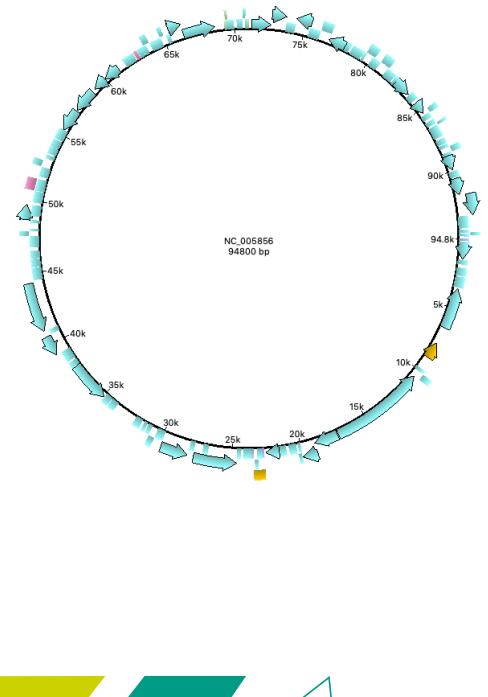
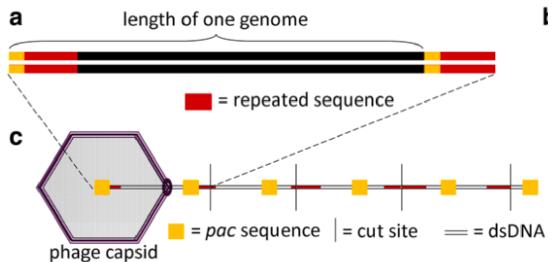


Phage genome examples: circularly permuted genomes

headful packaging: random ends



pac sites: start fixed, end random

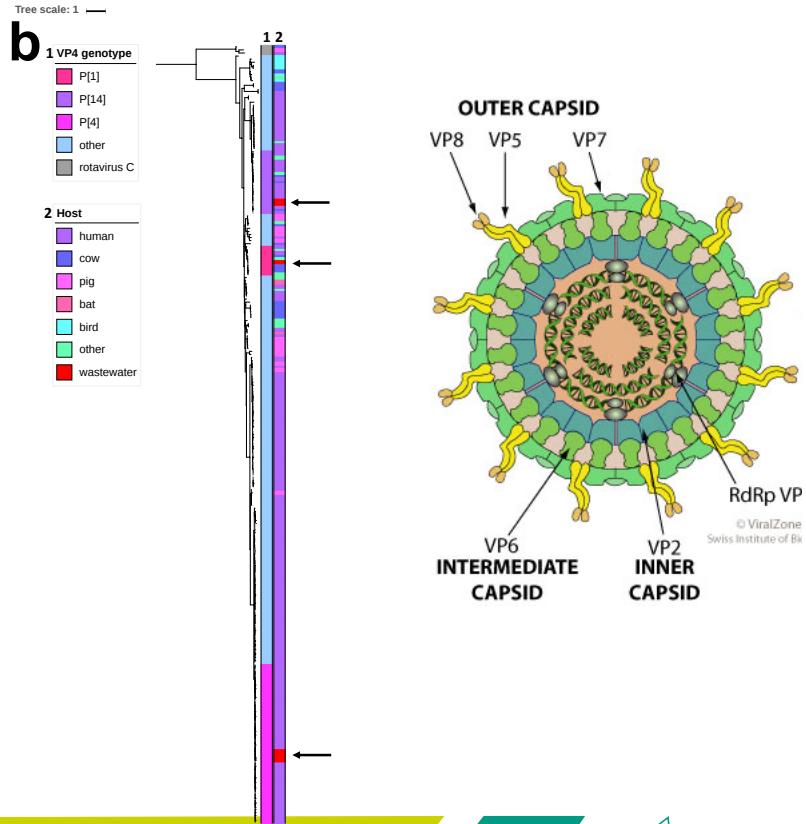
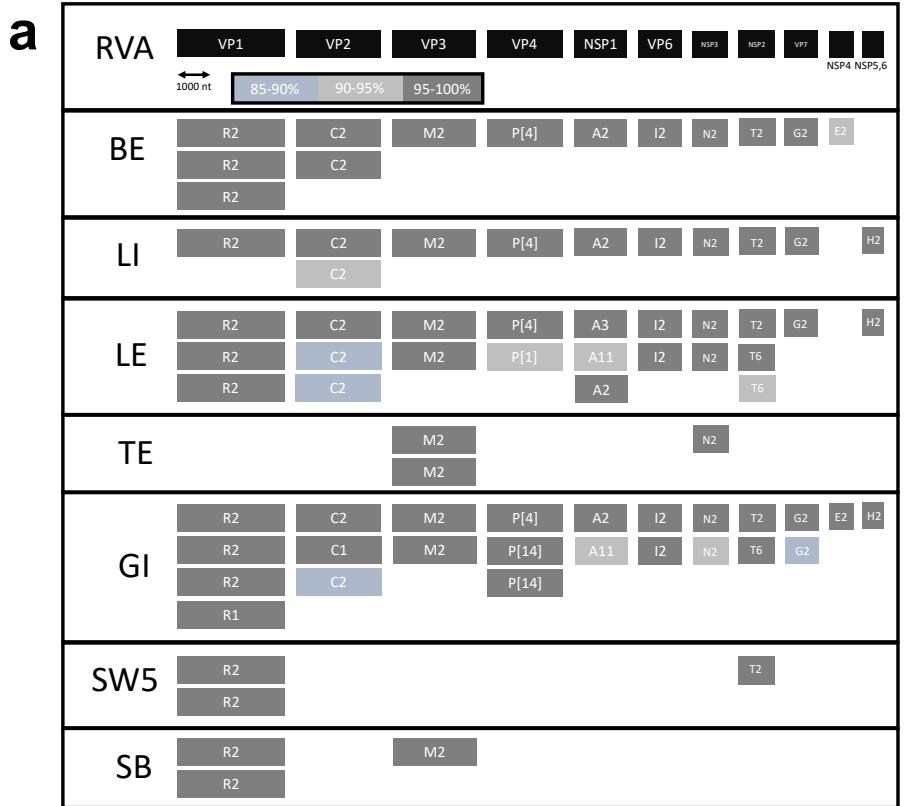


Merrill et al, 2016, BMC Genomics

Dickeya phage LIMEstone1, Adriaenssens et al 2012, PLoS ONE

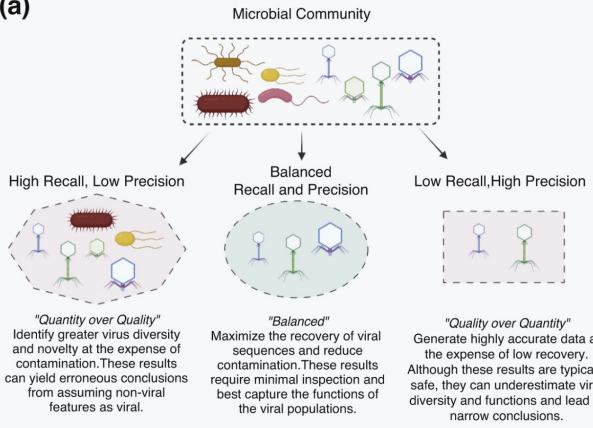
Escherichia phage P1, Lobocka et al 2004, J. Bact.

Segmented genomes: what the...?

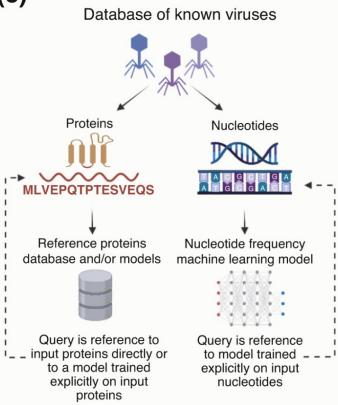


No tool is perfect!
Bias is everywhere

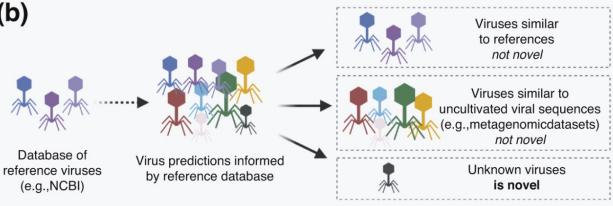
(a)



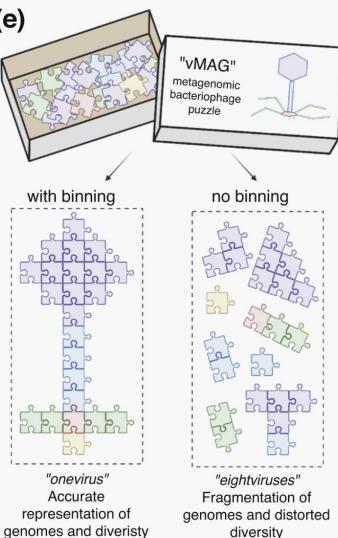
(c)

Available online at www.sciencedirect.com**ScienceDirect****Current Opinion in
Virology****Virus genomics: what is being overlooked?**Kristopher Kieft^{1,2} and Karthik Anantharaman¹

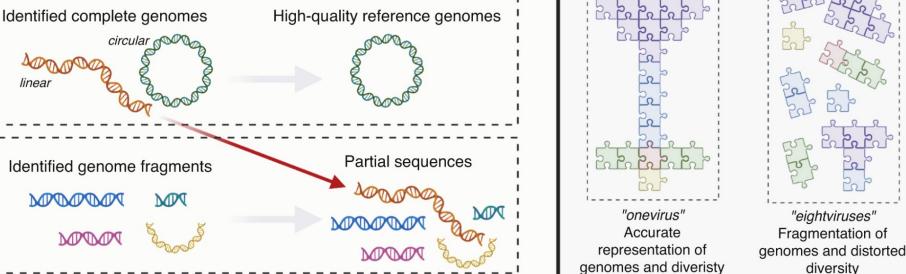
(b)



(e)



(d)



Kieft & Anantharaman, 2022

Open Peer Review | Bacteriophages | Methods and Protocols | 7 September 2022

f t ir

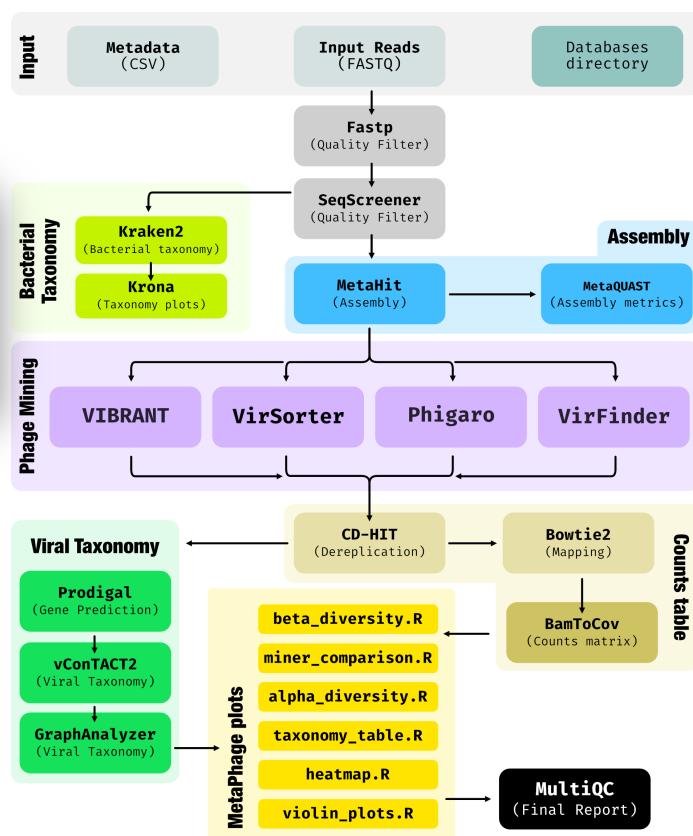
MetaPhage: an Automated Pipeline for Analyzing, Annotating, and Classifying Bacteriophages in Metagenomics Sequencing Data

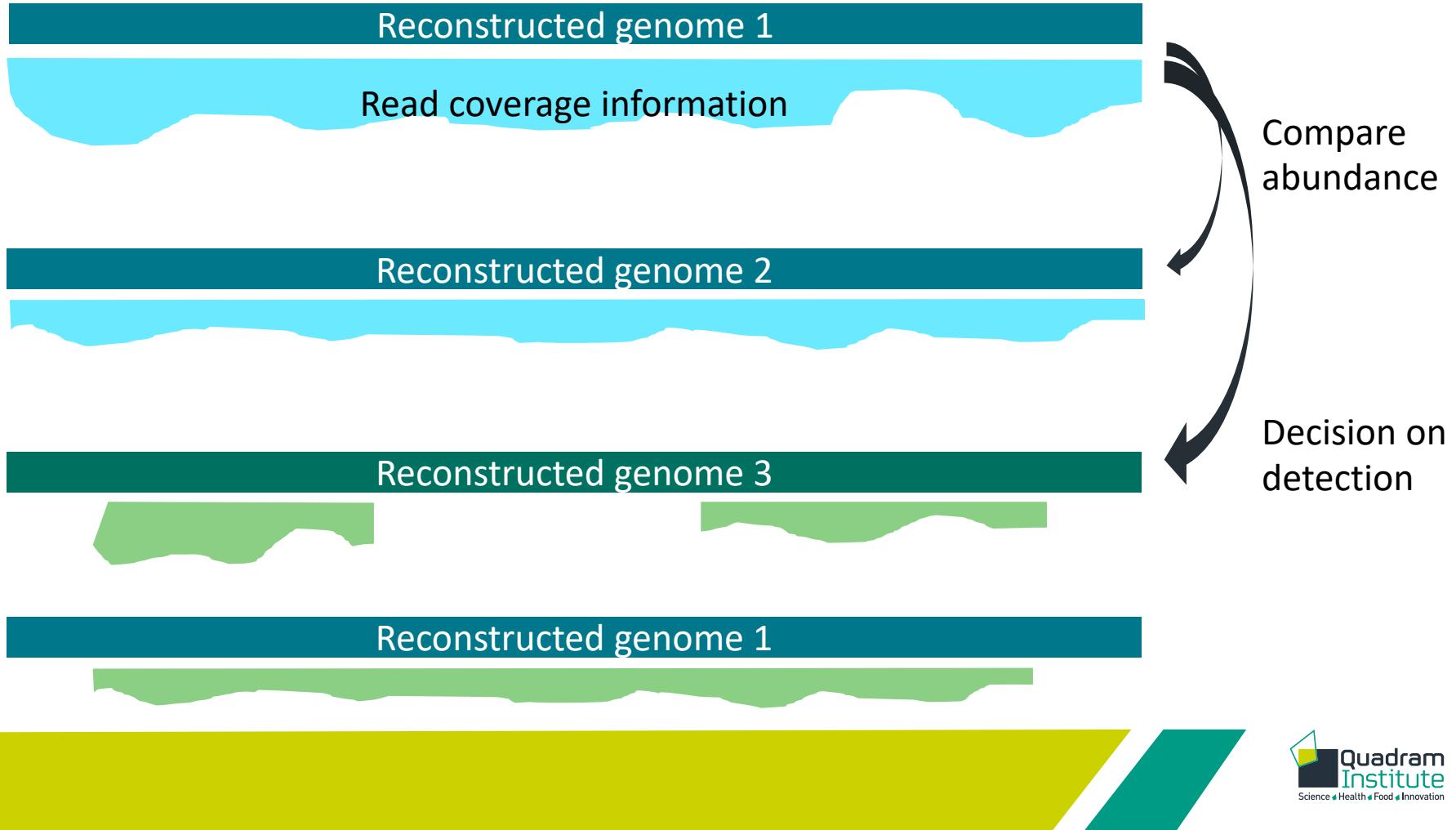
Authors: Mattia Pandolfo , Andrea Telatin , Giuele Lazzari , Evelien M. Adriaenssens , Nicola Vitulo  | AUTHORS INFO &

AFFILIATIONS

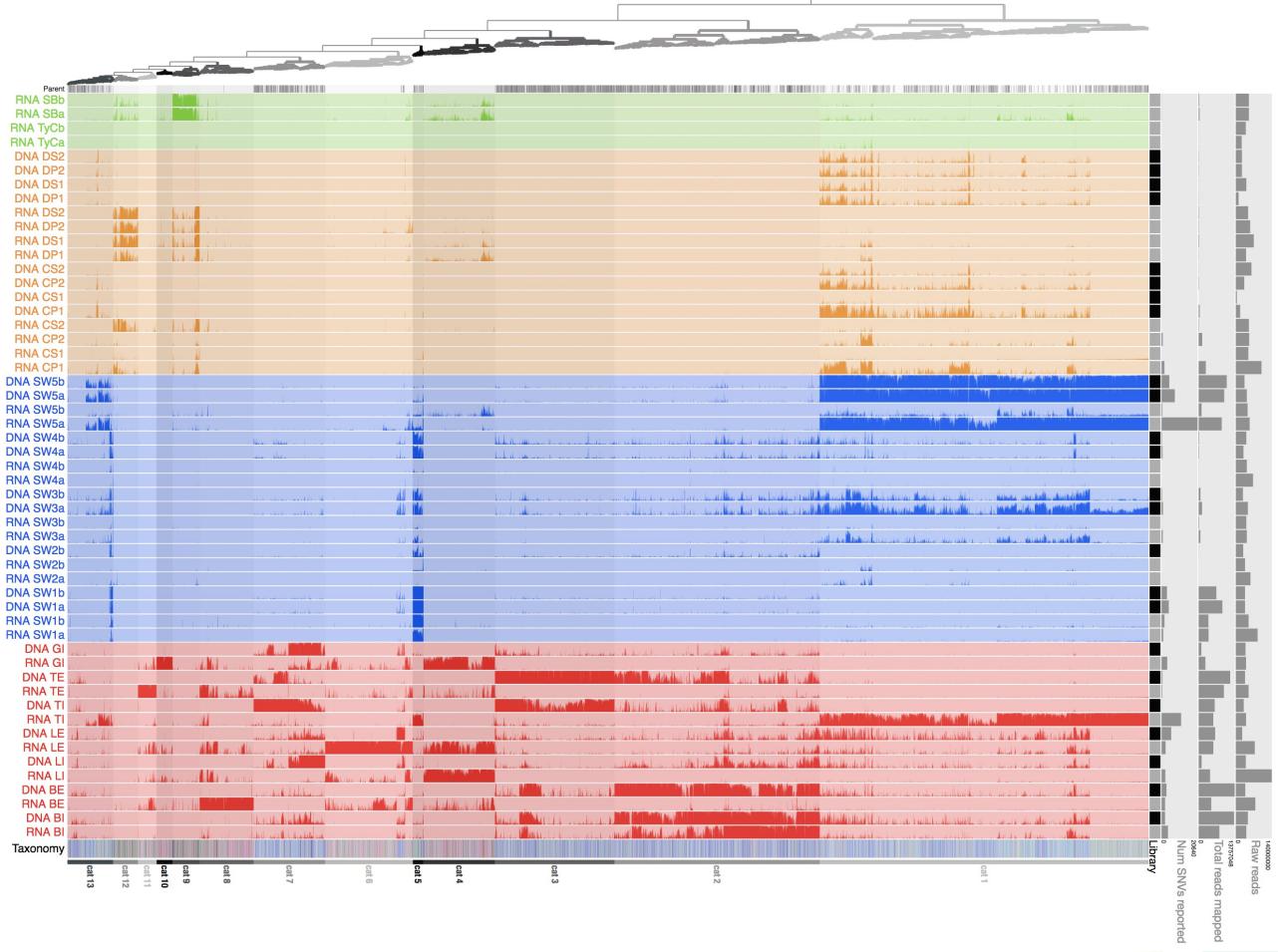
A reads-to-report pipeline for the analysis of the viral component of metagenomes

<https://mattiapandolfov.github.io/MetaPhage>





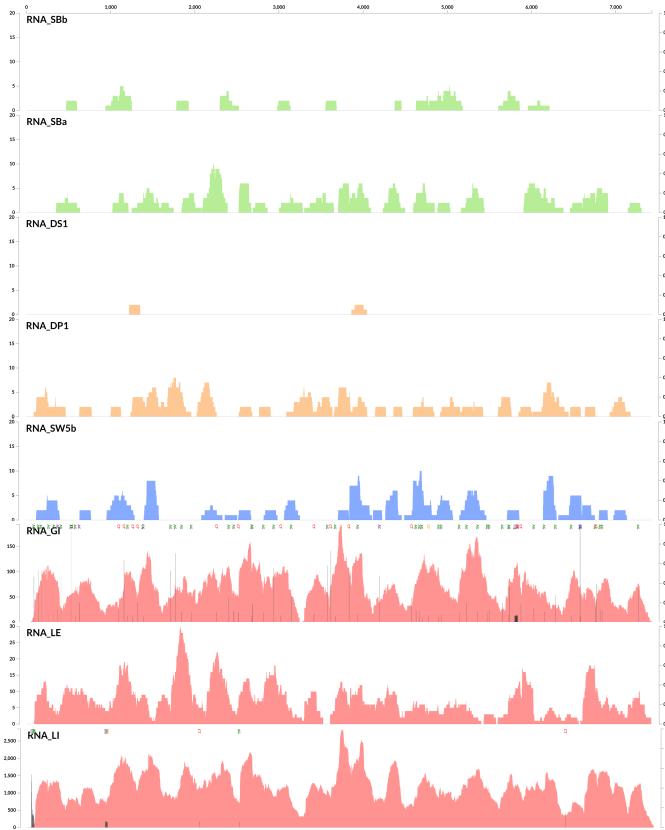
Can we use these outputs in anvi'o?



Adriaenssens et al, 2021, Water Research: analysis with Anvi'o v5 and v6

LI_NODE_9_length_7431_cov_255.051_split_00001 detailed

Sapovirus



Outputs so far:

- viral profile from reads
- set of viral contigs/genomes
- estimates of completeness
- relative abundances