

Alkaloids as Important Scaffolds in Therapeutic Drugs for the Treatments of Cancer, Tuberculosis, and Smoking Cessation

Prasat Kittakoop^{1,2,3,*}, Chulabhorn Mahidol^{1,2} and Somsak Ruchirawat^{1,2,3}

¹Chulabhorn Research Institute, Kamphaeng Phet 6 Road, Laksi, Bangkok 10210, Thailand; ²Chulabhorn Graduate Institute, Chemical Biology Program, Kamphaeng Phet 6 Road, Laksi, Bangkok 10210, Thailand; ³Center of Excellence on Environmental Health and Toxicology (EHT), CHE, Ministry of Education, Thailand

Abstract: Alkaloid molecules can act, depending on a type of amine functionality present in alkaloids, as either hydrogen-acceptor or hydrogen-donor for hydrogen bonding that is critically important for the interaction (binding) between targets (enzymes, proteins and receptors) and drugs (ligands). Because of this unique property, alkaloid scaffolds are therefore present in several drugs and lead compounds. This review highlights alkaloid scaffolds in drugs, particularly those recently approved in 2012; it also covers the scaffolds in leads and drug candidates which are in clinical trials and preclinical pipeline. The review focuses on three therapeutic areas including treatments of cancer, tuberculosis, and tobacco cessation. Alkaloid scaffolds in drugs and leads are inspired by those of naturally occurring alkaloids, and these scaffolds include pyridine, piperidine, quinoline, quinolinone, quinazoline, isoquinoline, indole, indolinone, isoindole, isoxazole, imidazole, indazole, thiazole, pyrazole, oxazolidinone, oxadiazole, and benzazepine. In addition to medicinal chemistry aspects, natural products possessing an individual alkaloid scaffold, as well as the mechanism of action of drugs and leads, are also discussed in this review.

Keywords: Alkaloids, natural products, drugs, lead compounds, cancer, anticancer drugs, cytotoxic activity, antitubercular activity, antimycobacterial activity, tuberculosis, smoking cessation.

INTRODUCTION

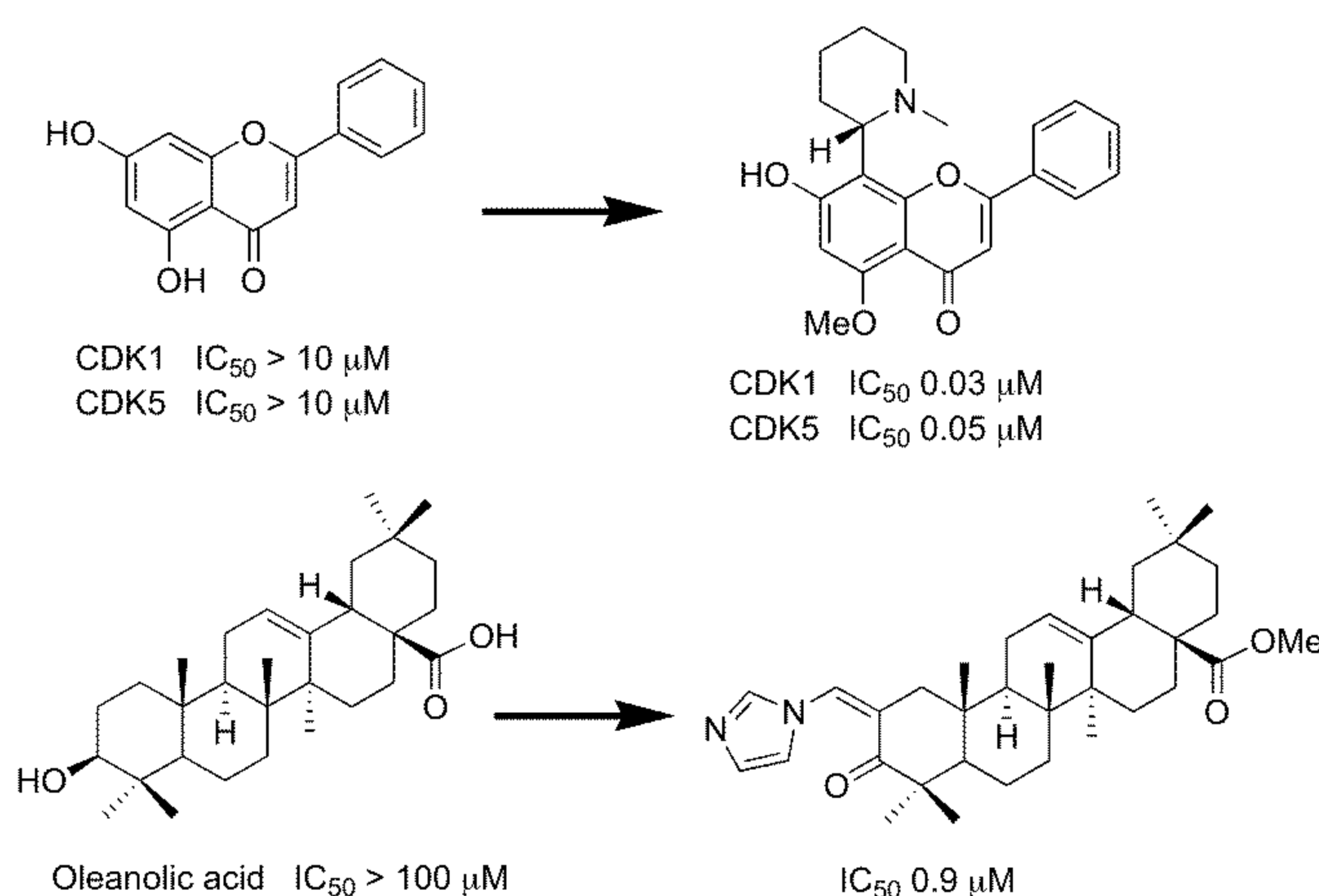
In 2012, the US Food and Drug Administration (FDA) approved 39 new drugs including 33 organic molecules and 6 biological drugs [1]. Among 33 small molecule drugs, there are at least 16 drugs whose structures are alkaloids or related to alkaloid molecules, and this underscores the importance of alkaloid structural features in drug discovery. While natural alkaloids isolated from various living sources provide many new leads with different drug targets, many synthetic alkaloids generated from medicinal chemistry research, through rational drug design (both computer-aided and structure-based drug designs) and lead optimization, prove to be potential drug candidates. The “-omics” technologies have accelerated drug discovery process; genomics and proteomics have revealed new potential drug targets [2-4]. Chemical proteomics enabling the direct analysis of protein activities, or proteins binding the drug, prove to be useful for the elucidation of drug targets as well as lead verification [5,6]. Computer-based docking and chemoinformatic screens are widely used to discover new ligands for targets of known structure, and to predict new substrates for enzymes of unknown function [7]. *In silico* prediction of drug-target interactions from heterogeneous biological data (chemical, genomic, and pharmacological data) [8], translational bioinformatics [9], and network-based relating

pharmacological and genomic spaces for drug target identification [10] have also accelerated the drug discovery processes. Upon the recent technologies mentioned above, it is anticipated that drug leads would be rapidly identified, and thus speeding the drug development processes. Therefore, these advanced technologies would consequently increase number of drugs approved in the future. Among the new small molecule drugs approved in the future, the numbers of molecules decorated with alkaloid scaffolds will also increase.

Alkaloids are generally known as compounds that contain nitrogen and heterocyclic rings, and they are relatively basic. Natural alkaloids are widely found in plants, animals, and microorganisms. Since alkaloids contain nitrogen atom(s) in their molecules, the most common functional groups in alkaloids are primary, secondary, and tertiary amines. The nitrogen atom(s) in alkaloids significantly contributes to the drug properties; a lone pair of electrons on the nitrogen usually accepts the protons (H-acceptor), while hydrogen(s) in primary and secondary amines act as a proton donor (H-donor) necessary for hydrogen bonding. It should be noted that the target-ligand interaction (or protein-ligand binding) heavily employs such hydrogen bonding. These H-acceptor and donor properties of nitrogen and hydrogen mentioned above, together with H-acceptor and donor of other functional groups (i.e. phenolic hydroxyl and polycyclic moieties), enable alkaloids to be ideal bioactive molecules which have perfect ability to bind (or interact) with drug targets (proteins, enzymes, and receptors); accordingly most natural alkaloids usually exhibit biological activities

*Address correspondence to this author at the Chulabhorn Research Institute, Kamphaeng Phet 6 Road, Laksi, Bangkok 10210, Thailand;
Tel: +66 86 9755777; Fax: +66-2-5538545;
E-mail: prasat@cri.or.th

which are of pharmaceutical interest. Therefore, it is not surprised that in some cases, the installation of alkaloid scaffolds into non-alkaloid molecules could lead to the increase of biological activities. Two recent studies have demonstrated that the transformation of non-alkaloids into alkaloidal compounds leads to dramatic increase of biological activities (Scheme 1). The installation of an alkaloid scaffold (a piperidine derivative) into flavonoids to furnish flavonoidal alkaloids led to the markedly increase in cyclin-dependent kinases (CDKs) inhibitory activity (at least 333 folds for CDK1 and 200 folds for CDK5) [11], while the attachment of an imidazole unit to a common plant triterpene, oleanolic acid, significantly improved antiproliferative activity in solid tumour cells up to 111 folds [12] (Scheme 1).



Scheme 1. Transformation of non-alkaloids into alkaloidal compounds leads to significant increase of biological activities.

This review highlights the importance of alkaloid scaffolds in therapeutic drugs, particularly those approved in 2012, as well as the scaffolds in leads and drug candidates that are being evaluated in clinical and preclinical trials. The present review focuses on the drugs and leads for cancer and tuberculosis therapies, and for the treatment of smoking cessation. Moreover, details on the mechanism of action of an individual drug and lead are also provided.

ALKALOID SCAFFOLDS IN ANTICANCER DRUGS

In recent years, there has been increase in the number of new anticancer drugs approved by the US FDA [1], and their numbers are likely to increase in the future. In 2012, the first anticancer drug, GDC-0449 (**1**) or Vismodegib, with a novel mechanism of action of the Hedgehog signaling pathway, is approved for the treatment of metastatic or locally advanced basal cell carcinoma (Fig. (1)). The Hedgehog signaling pathway was first discovered in 1980 [13], however, the landmark discovery in 2002 that the steroidal alkaloid cyclopamine (**2**) (Fig. (1)) from the plant (*Veratrum californicum*) inhibited the Hedgehog pathway has flashed the light on cancer therapeutics [14]. Details for the mechanism of action of Hedgehog signaling pathway have been conclusively established [15-17], and thus enabling medicinal chemistry to generate several potent inhibitors as anticancer drug candidates. Apart from GDC-0449 (**1**), four small molecule inhibitors of Hedgehog signaling pathway including XL-139

(BMS-833923, structure not disclosed), PF0444913 (structure not disclosed), IPI-926 (**3**), and LDE-225 (**4**) are under clinical evaluation (Fig. (1)) [18-21]. Moreover, the search for new synthetic and natural inhibitors of Hedgehog signaling pathway has also been intensively studied [22-25].

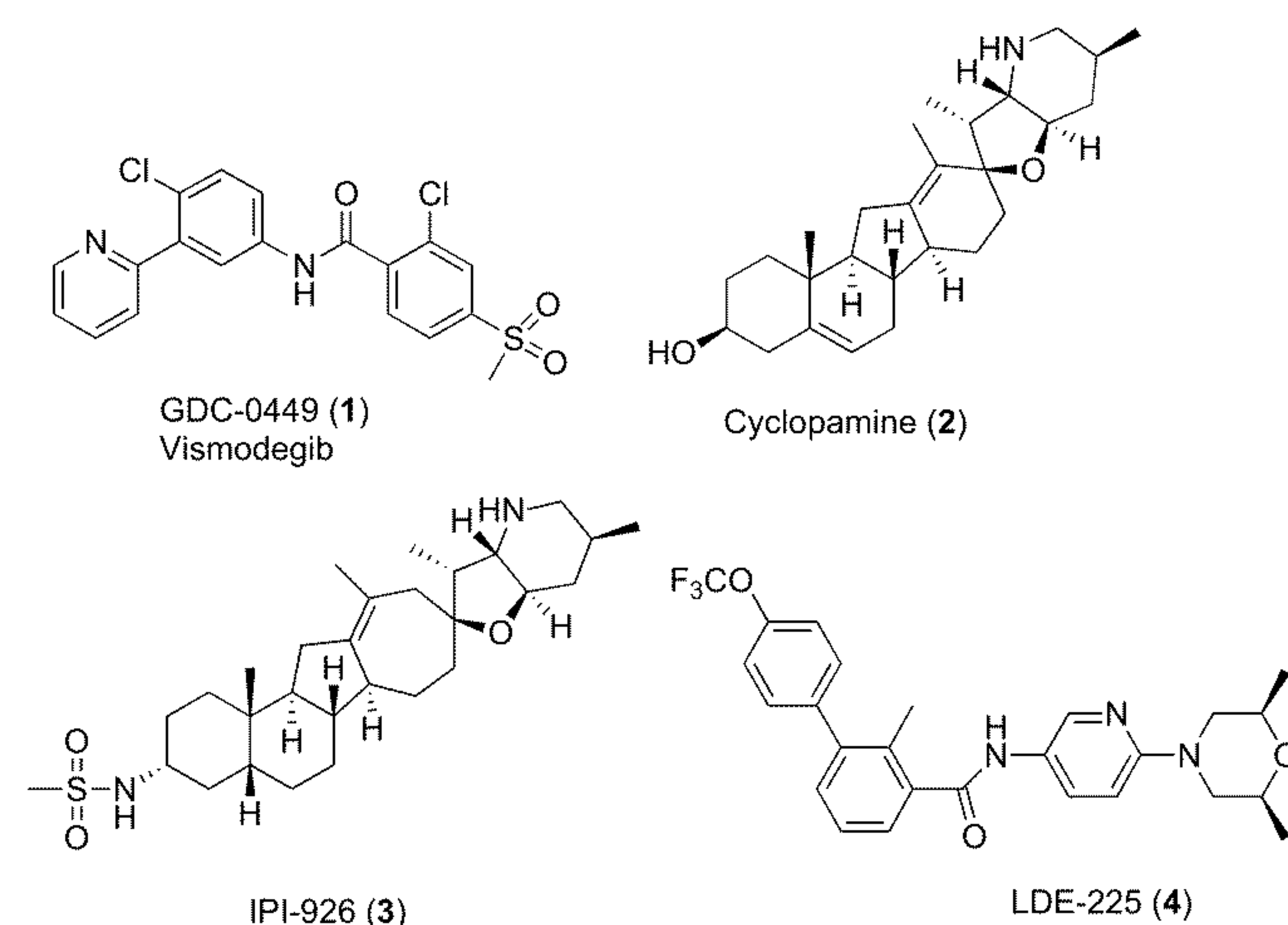


Fig. (1). Inhibitors of Hedgehog signaling pathway as anticancer drugs.

Inhibition of tyrosine kinases, the enzymes that catalyze the transfer of a phosphate group from adenosine triphosphate to target proteins, is one of the effective target therapies for cancer. There are a few anticancer drugs developed from this mechanism of action. Some tyrosine kinase inhibitors (drugs) have the alkaloid scaffolds including quinazoline for Erlotinib (**5**), CI-1033 (**6**) (Canertinib), and ZD1839 (**7**) (Gefitinib); pyridine alkaloids for BAY 43-9006 (**8**) (Sorafenib) and PTK787 (**9**) (Vatalanib); indolinone for SU5416 (**10**) (Semaxinib) and SU11248 (**11**) (Sunitinib or Sutent); and isoxazole for SU101 (**12**) (Leflunomide) (Fig. (2)). Normally, natural alkaloids including quinazoline [26], pyridine (e. g., nicotine derivatives) [27-29], indolinone [30-33], and isoxazole [34,35] are widely found in plant and animal kingdoms, as well as microorganisms.

As mentioned earlier that the inhibition of tyrosine kinases is one of the successful target therapies for cancer, therefore, in 2012, the US FDA approved five tyrosine kinase inhibitors for the treatment of cancers [1]. These drugs include BAY 73-4506 (**13**) (Regorafenib) for the treatment of metastatic colorectal cancer, XL184 (**14**) (Cabozantinib) for the treatment of progressive, metastatic medullary thyroid cancer, AP24534 (**15**) (Ponatinib) for chronic, accelerated or blast-phase chronic myeloid leukaemia, SKI-606 (**16**) (Bosutinib) for the treatment of chronic myelogenous leukemia, and AG013736 (**17**) (Axitinib) for advanced renal cell carcinoma (Fig. (3)) [1]. Again, these new anticancer drugs also possess the alkaloid scaffolds, for example, pyridine in BAY 73-4506 (**13**), quinoline in XL184 (**14**) and SKI-606 (**16**), imidazopyridazine in AP24534 (**15**), and indazole in AG013736 (**17**). While natural alkaloids including pyridine [27-29], quinoline [36-39], and imidazole [40-42] are widely found in Nature, natural indazole alkaloids are rare; so far only a few indazoles have been isolated from a plant source [43-46].

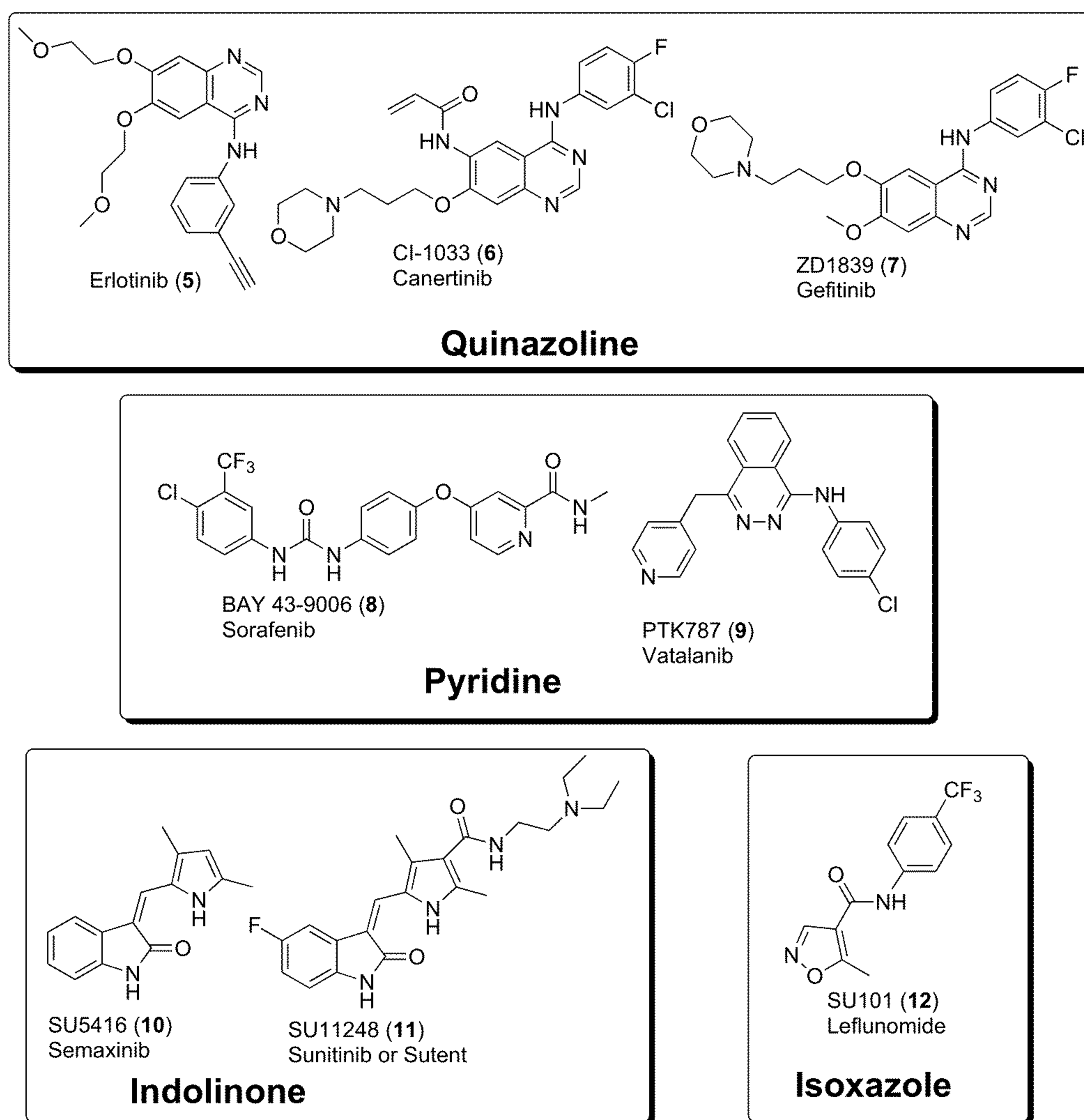


Fig. (2). Alkaloid scaffolds in tyrosine kinase inhibitors (anticancer drugs).

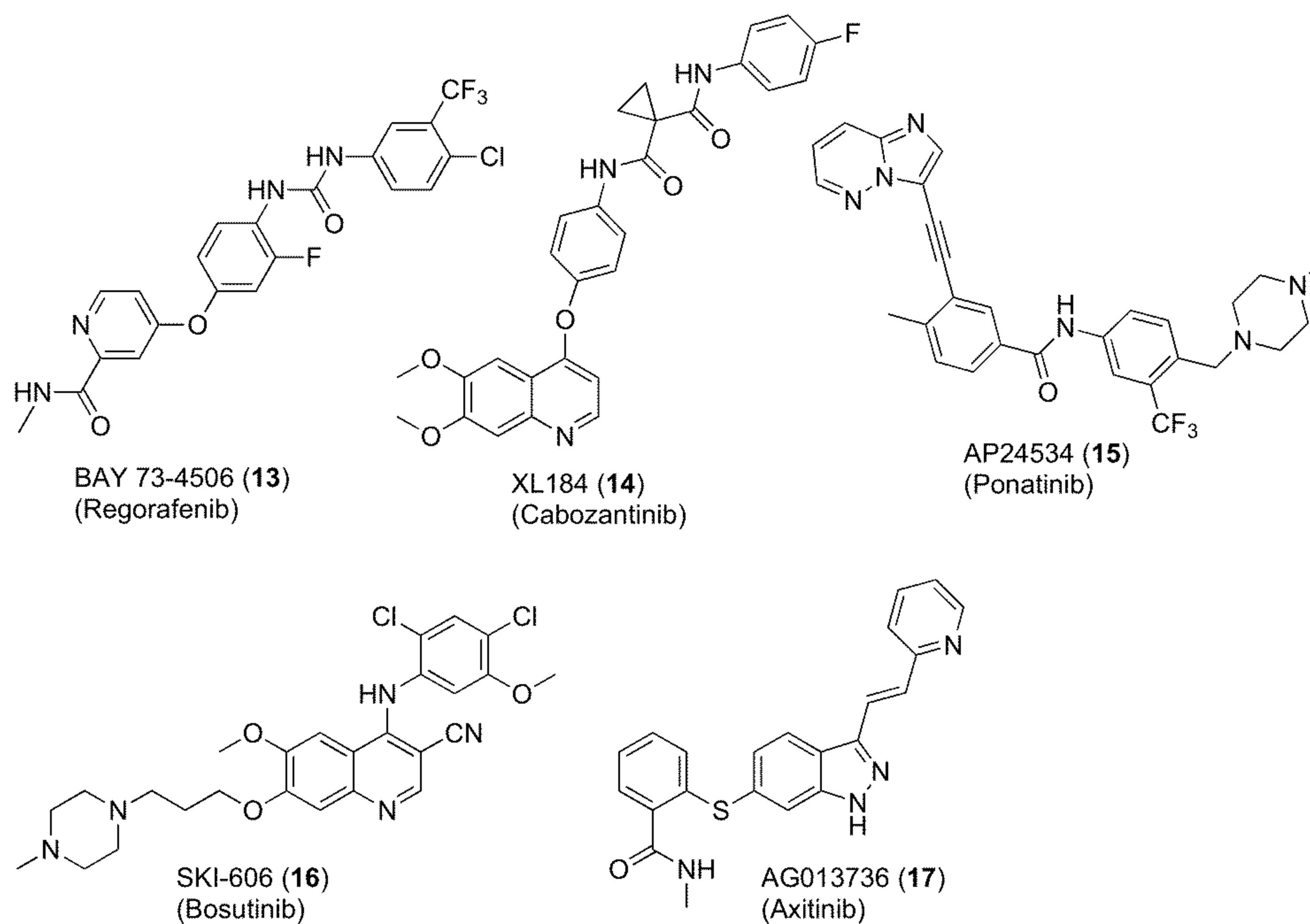
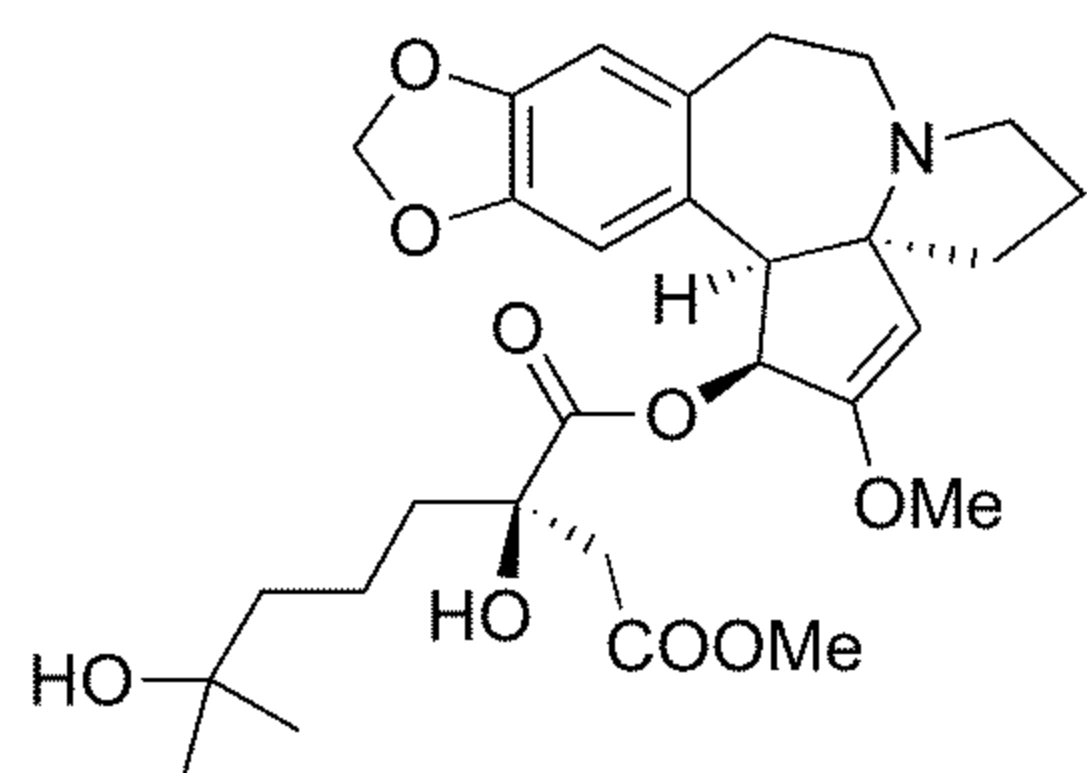


Fig. (3). Tyrosine kinase inhibitors as anticancer drugs approved in 2012.

The plant alkaloid omacetaxine mepesuccinate (**18**), or homoharringtonine, is a natural product approved as an anticancer drug in 2012 (Fig. (4)). Omacetaxine mepesuccinate (**18**) was first isolated in 1970 from *Cephalotaxus harringtonia*, however, at that time this class of alkaloids was collectively called as “harringtonine” [47]. This alkaloid class is also known as “cephalotaxine”. The structure of omacetaxine mepesuccinate (**18**) and related alkaloids was elegantly characterized by Powell and his colleagues [48,49]. Several alkaloids from *C. harringtonia* exhibited anticancer activity against leukemia cell lines [50]. It worth noting that homoharringtonine or omacetaxine mepesuccinate (**18**) takes more than 40 years from the first isolation (and the discovery of anticancer activity) to become a clinically used drug. Omacetaxine mepesuccinate (**18**) has the mechanism of action in preventing the initial elongation step of protein synthesis, and thus inhibiting protein translation [51], and it is used to treat chronic or accelerated phase chronic myeloid leukemia with resistance and/or intolerance to two or more tyrosine kinase inhibitors.



Omacetaxine mepesuccinate (or Homoharringtonine) (**18**)

Fig. (4). The plant alkaloid, omacetaxine mepesuccinate (**18**), approved as an anticancer drug.

Thiazole alkaloids are widely found in Nature [40-42], particularly in marine invertebrates [52,53]. Epothilones are thiazole alkaloids isolated from Myxobacteria [54], and they are leads for anticancer drug. Epothilones A (**19**) and B (**20**) were first isolated in 1993 [55] and described as antifungal and cytotoxic compounds [54,55] (Fig. (5)). In 1995, epothilones A (**19**) and B (**20**) were found to be microtubule-stabilizing agents, both *in vitro* and in cultured cells, sharing the same mechanism of action as that of paclitaxel (taxol) [56]. The pronounced antiproliferative activity of epothilones A (**19**) and B (**20**), together with a potent ability to stabilize microtubules (also active against multiple drug-resistant cancer cells), has led to extensive research on (medicinal) chemistry and chemical biology of these natural thiazole alkaloids. Consequently, several epothilone analogs have been synthesized and tested for anticancer properties [57-59]. Among the new epothilone derivatives, BMS-247550 (**21**) (Ixabepilone), the lactam analog of epothilone B (**20**) known as azaepothilone B, was an anticancer drug approved in 2007 for the treatment of metastatic or locally advanced breast cancer in patients whose tumors are resistant to taxane and anthracycline anticancer drugs.

Another thiazole alkaloid lead is largazole (**22**), a cyclic depsipeptide isolated from a cyanobacterium of the genus *Symploca* (Fig. (6)) [60]. Largazole (**22**) potently inhibits class I histone deacetylases [61], and it also exhibits a broad spectrum of biological activities [62,63]. Several largazole

derivatives have been prepared, and some are promising anticancer drug candidates [62].

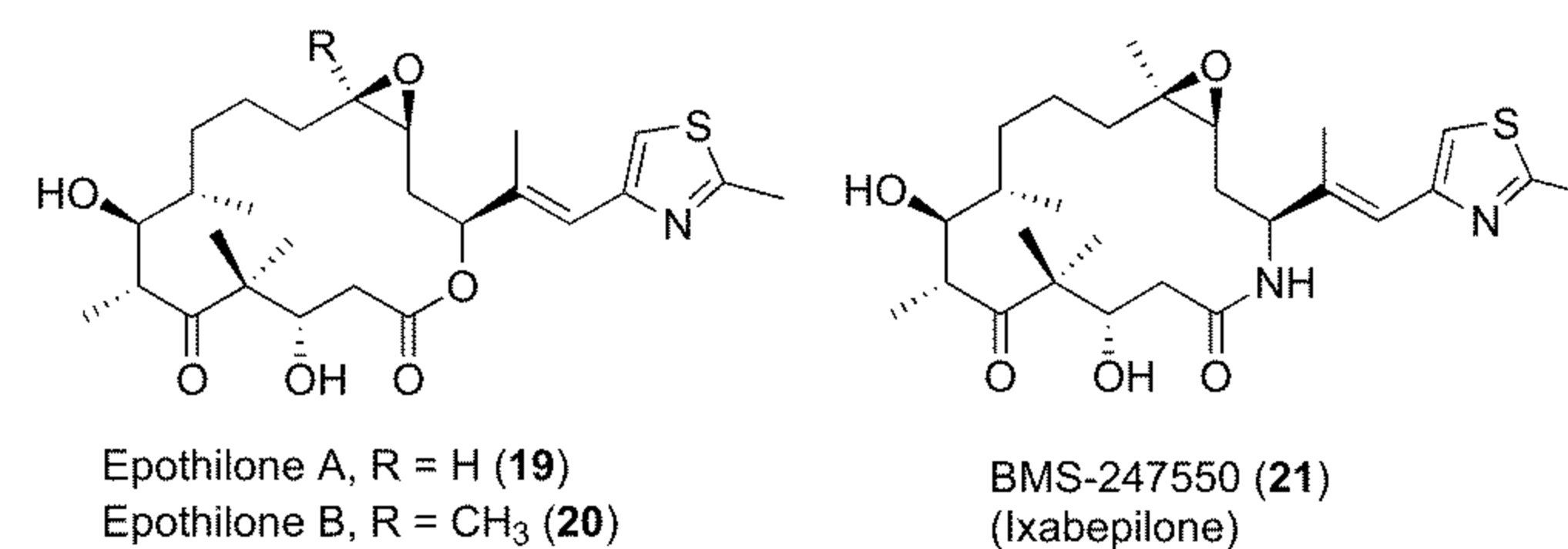


Fig. (5). Epothilones **19** and **20** isolated from Myxobacteria and an anticancer drug **21**.

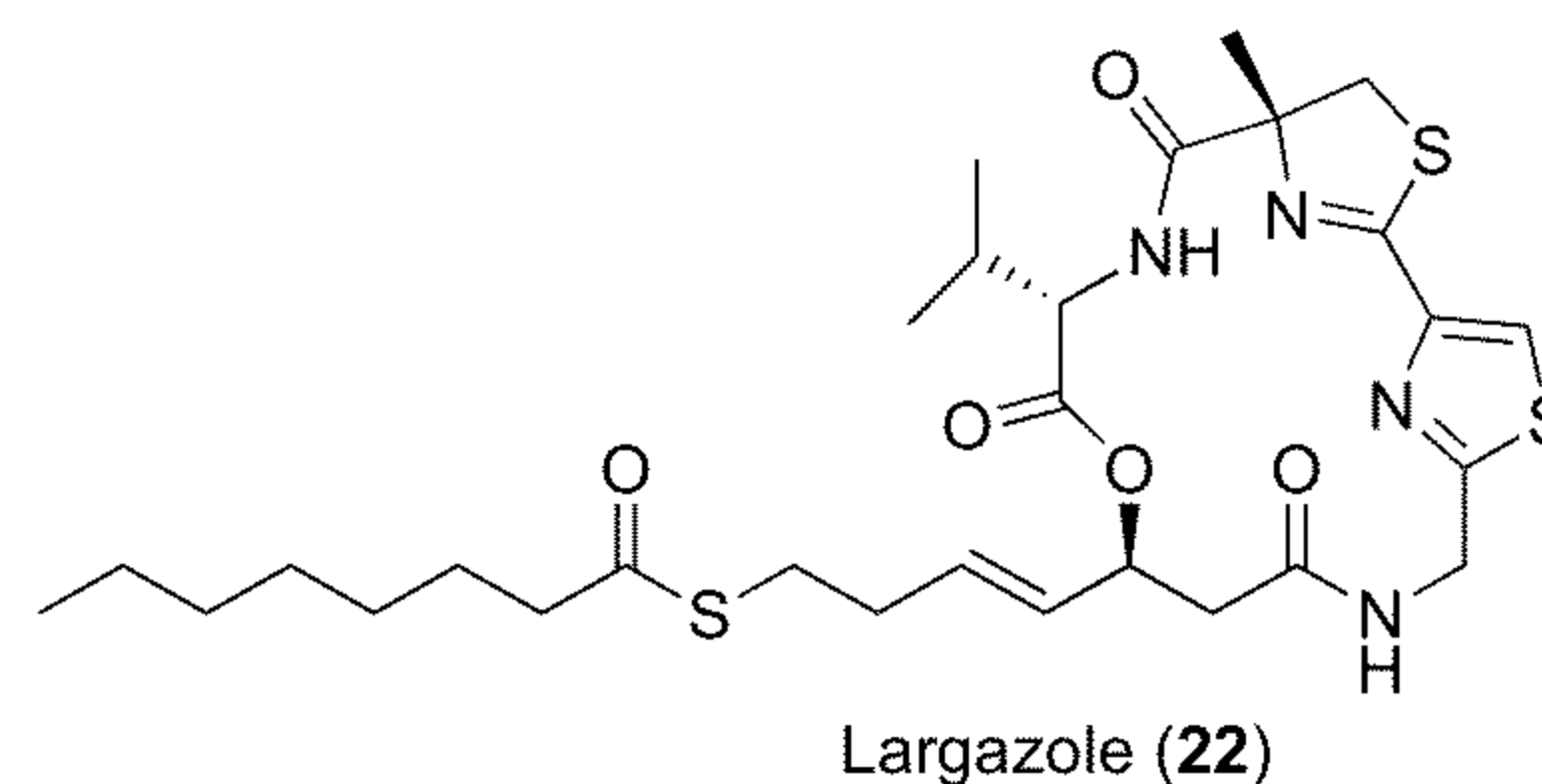


Fig. (6). Largazole (**22**) from a cyanobacterium of the genus *Symploca*

Heat shock protein 90 (Hsp90) is a chaperone protein that cancer cells use to facilitate the function of many proteins required for tumor growth. The fact that Hsp90 is essential for the tumor growth makes the Hsp90 to be one of the hot targets for cancer therapy. Undoubtedly, there are 17 Hsp90 inhibitors recently entering clinical trials [64]. Structures of several Hsp90 inhibitors are mainly inspired by two natural products, geldanamycin (**23**) and radicicol (**26**) (Fig. (7)). It should be noted that both geldanamycin (**23**) and radicicol (**26**) are macrolactam and macrolactone polyketides, respectively, and they do not have alkaloid scaffolds in their molecules. However, a structural transformation of the natural products **23** and **26** into alkaloid-like structures significantly improves anticancer property. As shown in Fig. (7), a chemical modification of the non-alkaloid geldanamycin (**23**) to the alkaloid-like structures, e. g. 17-AAG (**24**) and 17-DMAG (**25**), provided good drug properties, and the derivatives **24** and **25** are anticancer drug candidates being evaluated in clinical trials [65,66]. The resorcinol part of radicicol (**26**) is important for the inhibition of Hsp90, and the attachment of the alkaloid scaffolds to a benzene-1,3-diol derivative (a resorcinol-like unit) provides anticancer drug candidates NVP-AUY922 (**27**), CCT018159 (**28**), and AT13387 (**29**) (Fig. (7)), which are in clinical trials [67-69]. The alkaloids scaffolds, isoxazole, pyrazole, and isoindole are attached to the derivatives **27**, **28**, and **29**, respectively. Isoxazole alkaloids [34,35] are widely present in Nature, while pyrazole [70-74] and isoindole [75-77] alkaloids are rare metabolites.

Vinca alkaloids are probably one of the most-used classes of anticancer drugs; they are dimeric indole alkaloids found in the leaves of the Madagascar periwinkle plant *Catharanthus roseus*. Vincristine (**30**) and vinblastine (**31**) were isolated from *C. roseus* (formerly known as *Vinca rosea*) in the late 1950s, and both are currently used as anticancer

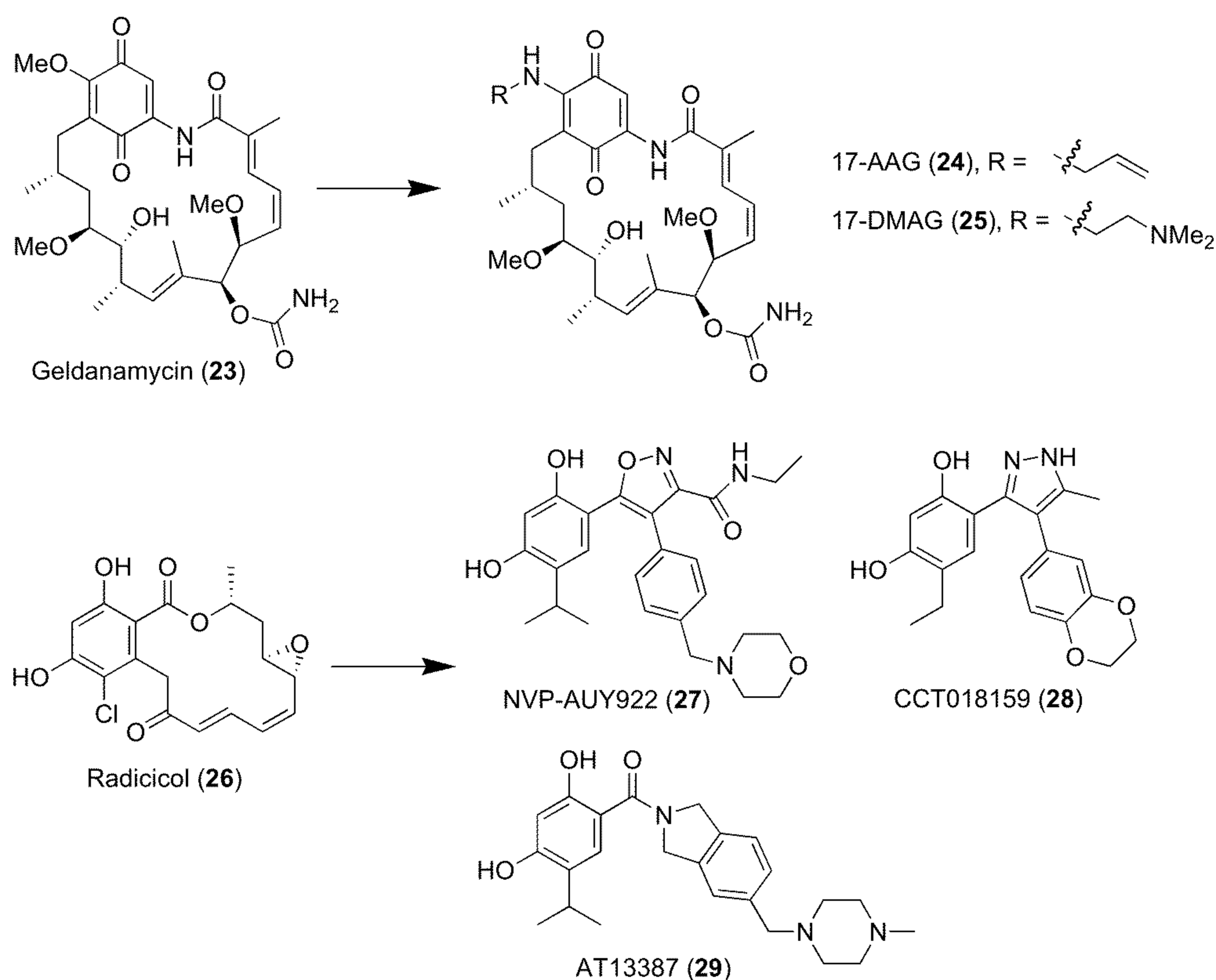


Fig. (7). Geldanamycin (23) and radicicol (26) and their derivatives decorated with alkaloid scaffolds.

drugs (Fig. (8)). Semi-synthetic vinca alkaloid analogs, vinorelbine and vinflunine, also play an important role in cancer chemotherapy. Vinca alkaloids exert their cytotoxicity through the inhibition of microtubule assembly by preventing tubulin polymerization. Recently, Boger and co-workers prepared a series of C20' urea derivatives (e.g. 32) whose potency matched or exceeded that of the parent compound, vinblastine (31). The findings that the hydrogen bond donor on the C20' position is the key role for the potent vinblastine analogues open new prospects for vinblastine analogues (e.g. 32) as anticancer leads [78]. Moreover, several potent vinca alkaloid analogs were recently prepared and extensively studied for anticancer property [79-82].

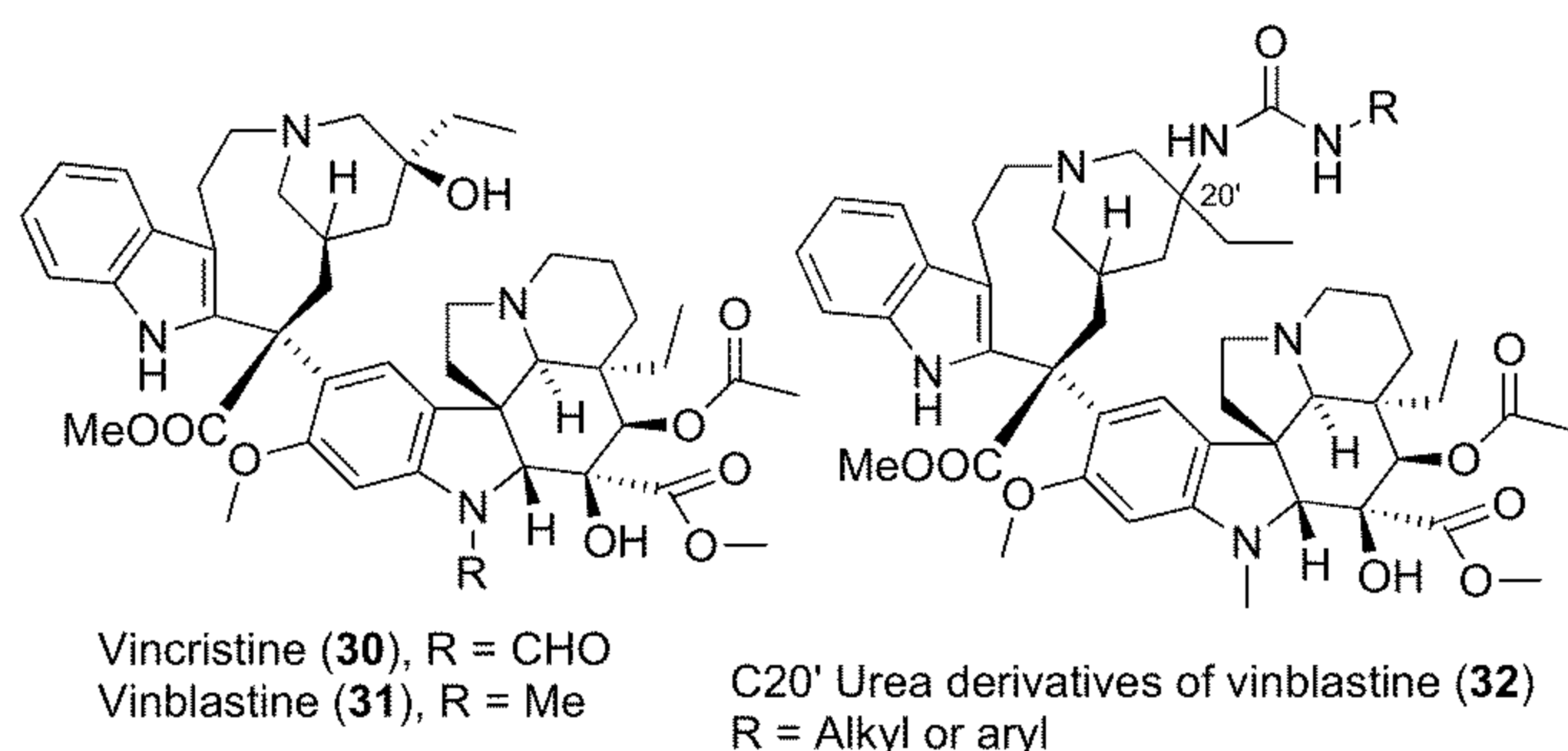


Fig. (8). Structure of vinca alkaloids (30 and 31) and a derivative 32.

Ecteinascidin 743 (33), a tetrahydroisoquinoline alkaloid, isolated from a marine tunicate *Ecteinascidia turbinata*, was approved as an anticancer drug in 2007 (Fig. (9)). Ecteinascidin 743 (33) exhibits anticancer activity by the selective

alkylation of the minor groove of DNA with preference for GC-rich triplets through covalently bonding at the N2-position of guanine [83].

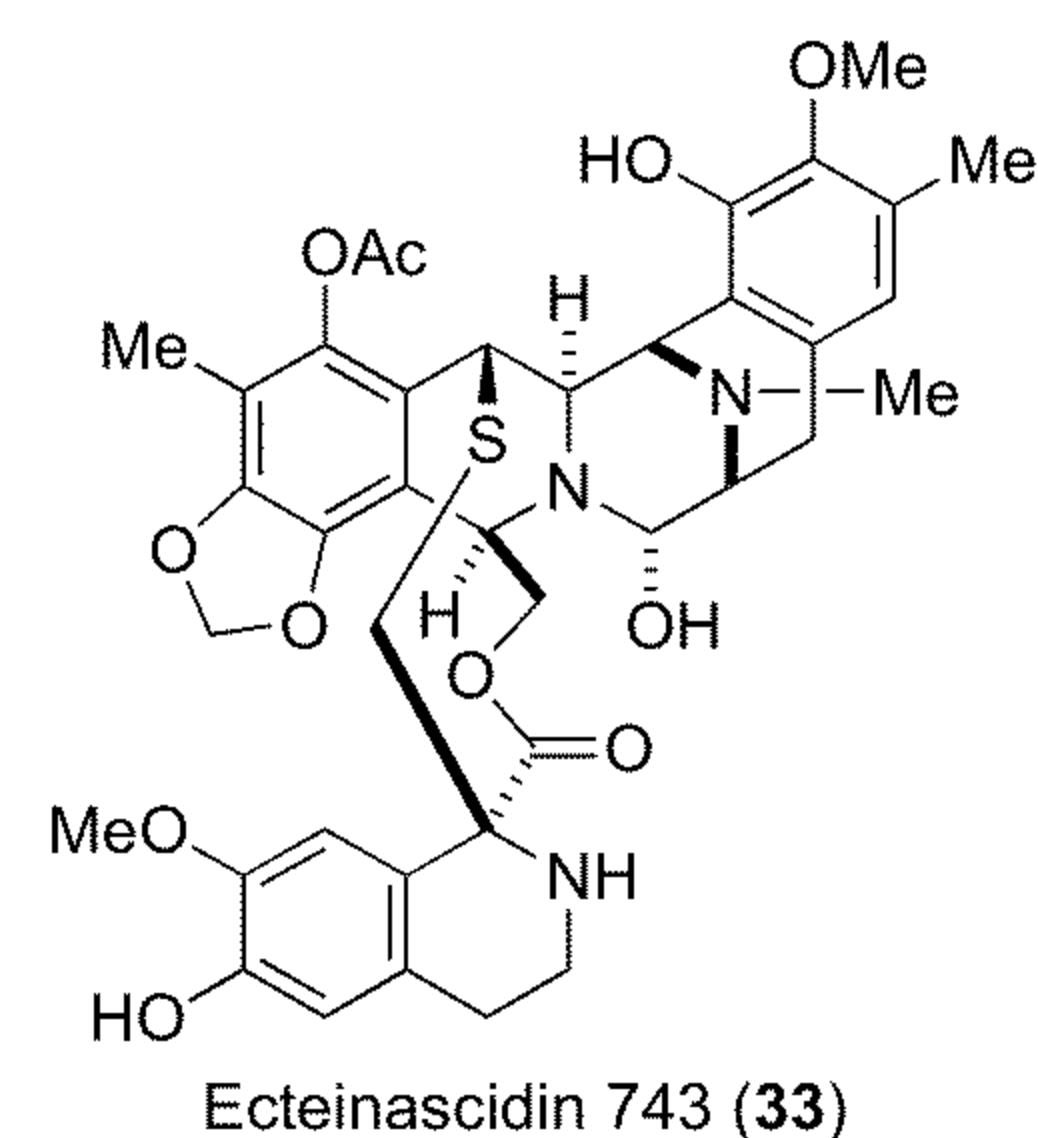


Fig. (9). Structure of ecteinascidin 743 (33).

Duocarmycins, i.e. duocarmycin SA (34), are indole alkaloids produced by the bacterium *Streptomyces* sp., and they exhibit potent cytotoxic activity by binding to the minor groove of DNA and alkylating the AT-rich adenine at the N3 position [84]. CC-1065 (35) and its derivatives, i.e. (+)-CBI-indole₂, are promising anticancer drug candidates (Fig. (10)) [85]. Recently, duocarmycin carbamate prodrug (36) was found to exhibit *in vivo* efficacy exceeded that of (+)-CBI-indole₂, and it also had higher therapeutic window of efficacy versus toxicity than (+)-CBI-indole₂ [86].

Inhibition of topoisomerases, the enzymes that control DNA supercoiling and entanglements, is an effective mechanism of action for anticancer drugs [87]. Camptothecin (37),

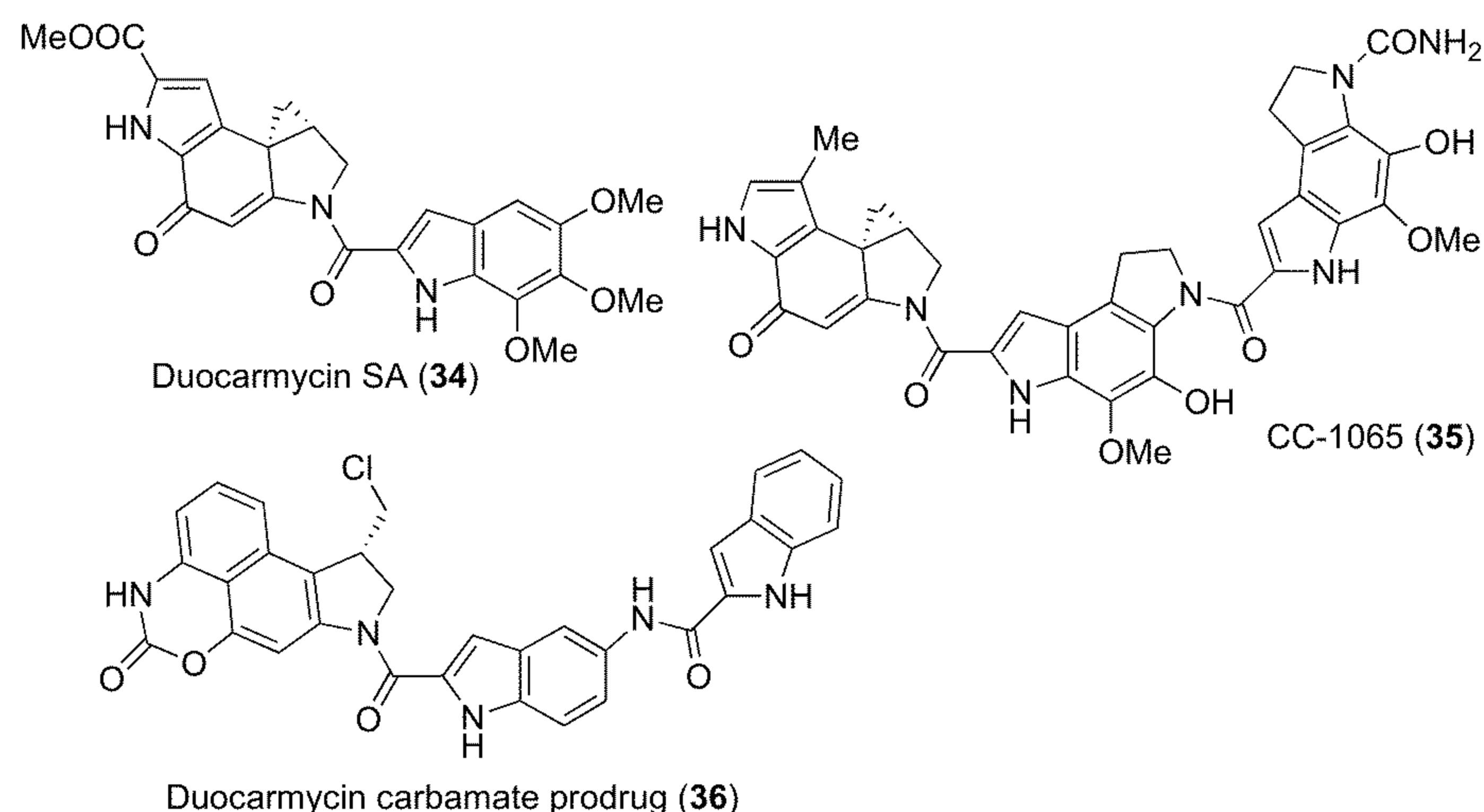


Fig. (10). Structure of duocarmycin SA (34) and derivatives (35 and 36).

a quinoline alkaloid from *Camptotheca acuminata*, displays the inhibition of topoisomerase type I (Fig. (11)). Camptothecin (37) is an anticancer lead, and its derivatives (Topotecan and Irinotecan) are approved as anticancer drugs. Several camptothecin derivatives were found to exhibit pronounced anticancer activity [88,89]. Podophyllotoxin (38), a lignan from many plant species of the Podophyllum family, is an anticancer lead. Two semi-synthetic podophyllotoxin derivatives, etoposide and teniposide, are currently used for the treatment of various types of cancer. It is worth mentioning that the transformation of a non-alkaloid lignan podophyllotoxin (38) to alkaloid-like structures, GL-331 (39) and F14512 (40), led to pronounced increase of cytotoxic activity; compounds 39 and 40 are in clinical trials for cancer treatment (Fig. (11)) [87,90,91]. Moreover, podophyllotoxin derivatives decorated with a pyrazole alkaloid moiety are recently found to exhibit significant antiproliferative activity [92]. Lamellarins, e.g., lamellarin D (41) (Fig. (11)), are polycyclic pyrrole alkaloids isolated from marine tunicates and mollusks, and they have recently gained great attentions from medicinal chemists due to unique structural features and potent cytotoxicity. Total synthesis of lamellarins has been well established [93-96]; several lamellarin derivatives exhibited potent anticancer activity [97-99], and were selective inhibitors of HIV-1 integrase [99,100]. Lamellarin D (41) is particularly of interest, because it is an anticancer lead inhibiting topoisomerase type I [101]. Several natural topoisomerase inhibitors have been isolated from natural sources [102]; some inhibitors show potent activity and become anticancer leads, for example, taspine (42) from *Croton lechleri* and evodiamine (43) from *Evodiae fructus* (Fig. (11)) [103,104]. Taspine (42) (also known as thaspine) and its derivatives showed promising anticancer properties [105-108]. The taspine analog, HMQ1611, inhibited growth of breast cancer and non-small cell lung cancer [109,110]. The derivatives of evodiamine (43) showed good *in vivo* antitumor efficacy with low toxicity, and thus representing a new class of anticancer drug candidate [104].

ALKALOID SCAFFOLDS IN DRUGS FOR THE TREATMENT OF TUBERCULOSIS

While a number of patients infected with tuberculosis and a number of multi- and extensively-drug resistant tuber-

culosis strains have gradually increased each year, only few first-line drugs including isoniazid, rifampicin, ethambutol, pyrazinamide, and streptomycin are available for the treatment of tuberculosis. In 2011, the World Health Organization (WHO) reported that 8.7 million people fell ill with tuberculosis and 1.4 million people died from tuberculosis [111]. The last drug (rifampicin) approved for the treatment of tuberculosis was in 1960s. However, due to the emergence of multi- and extensively-drug resistant tuberculosis strains and the increase number of tuberculosis infection worldwide, there is an urgent need for new anti-tuberculosis (anti-TB) drugs. Recently, there are a few anti-TB drug candidates in clinical trials, as well as in the drug pipeline. Alkaloid scaffolds play an important role in the tuberculosis drug development pipeline, as they always parts of the molecules of anti-TB agents in clinical trials. The alkaloid scaffolds in the anti-TB drug candidates are quinoline, imidazole, quinolinone, and oxazolidinone. The scaffolds of quinoline [36-39], imidazole [34,40-42], and quinolinone [112-116] are widely found in Nature, however, the oxazolidinone scaffold [117,118] is rarely seen in natural products.

TMC207 (44) (also known as R207910) is a new anti-TB drug approved in 2012 by the US FDA (Fig. (12)), and its molecule has a quinoline alkaloid scaffold. TMC207 (44) is not only the first anti-TB drug in four decades, but also a new anti-TB drug with a new mechanism of action. TMC207 (44) targets the proton pump of adenosine triphosphate (ATP) synthase of *Mycobacterium tuberculosis*, and thus specifically inhibiting the mycobacterial ATP synthase [119]. OPC-67683 (45) or Delamanid is a nitroimidazole showing effectiveness in patients with multidrug-resistant pulmonary tuberculosis (Fig. (12)) [120], and it may be approved as an anti-TB drug in 2013 (in Europe). OPC-67683 (45) exerts anti-TB activity with the same mechanistic action as that of an anti-TB isoniazid drug, which inhibits mycolic acid biosynthesis that is essential process for the formation of mycobacterial cell wall [121]. The anti-TB imidazole namely PA-824 (46) (Fig. (12)), exhibiting the same mechanism of action as that of OPC-67683 (45), is now in Phase II clinical trials [122]. Gatifloxacin (47) and moxifloxacin (48), the fourth-generation fluoroquinolone antibiotic drugs (Fig. (12)), are currently in clinical trials for the treatment of

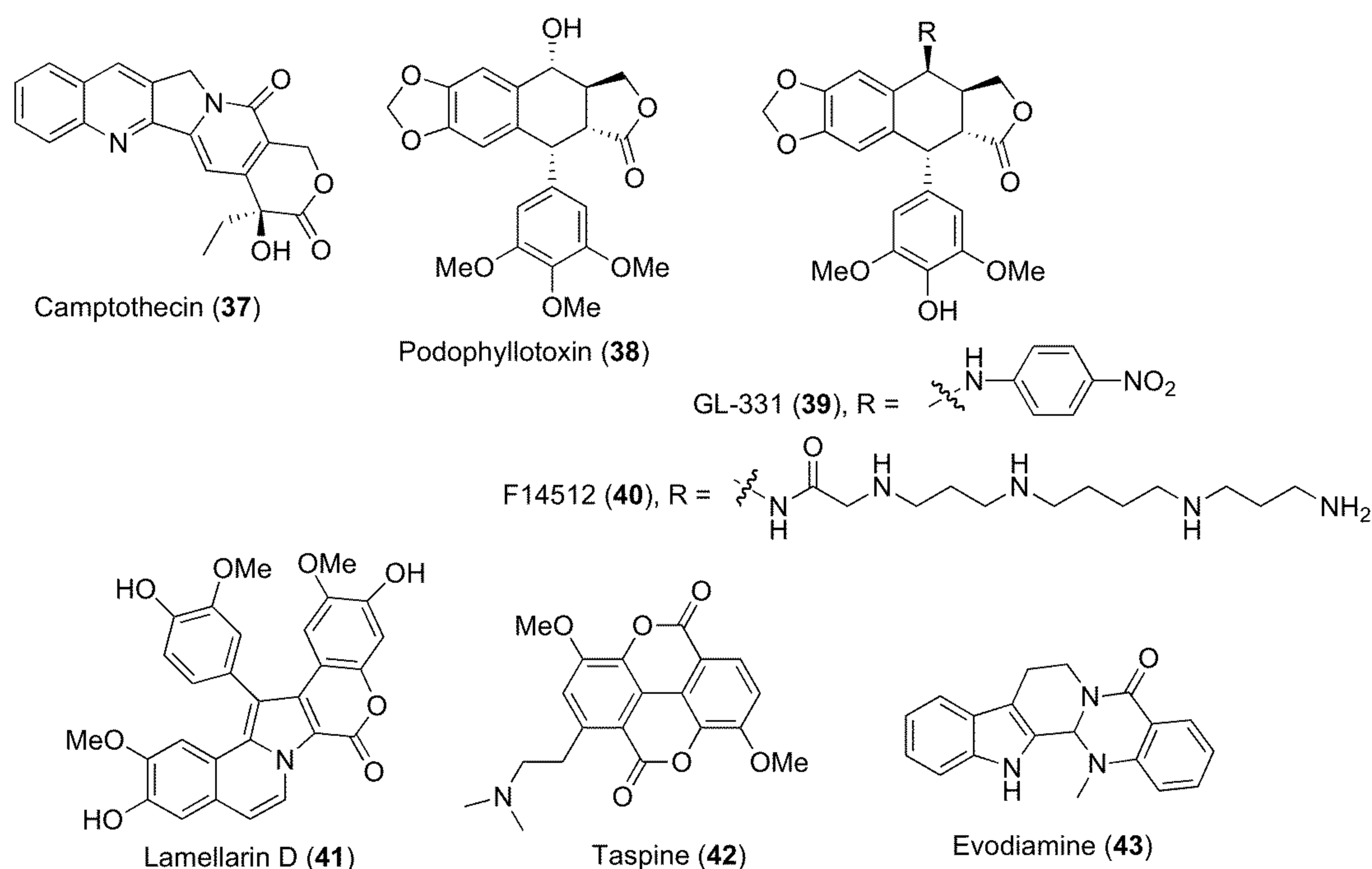


Fig. (11). Inhibitors of topoisomerases as anticancer drugs (or leads).

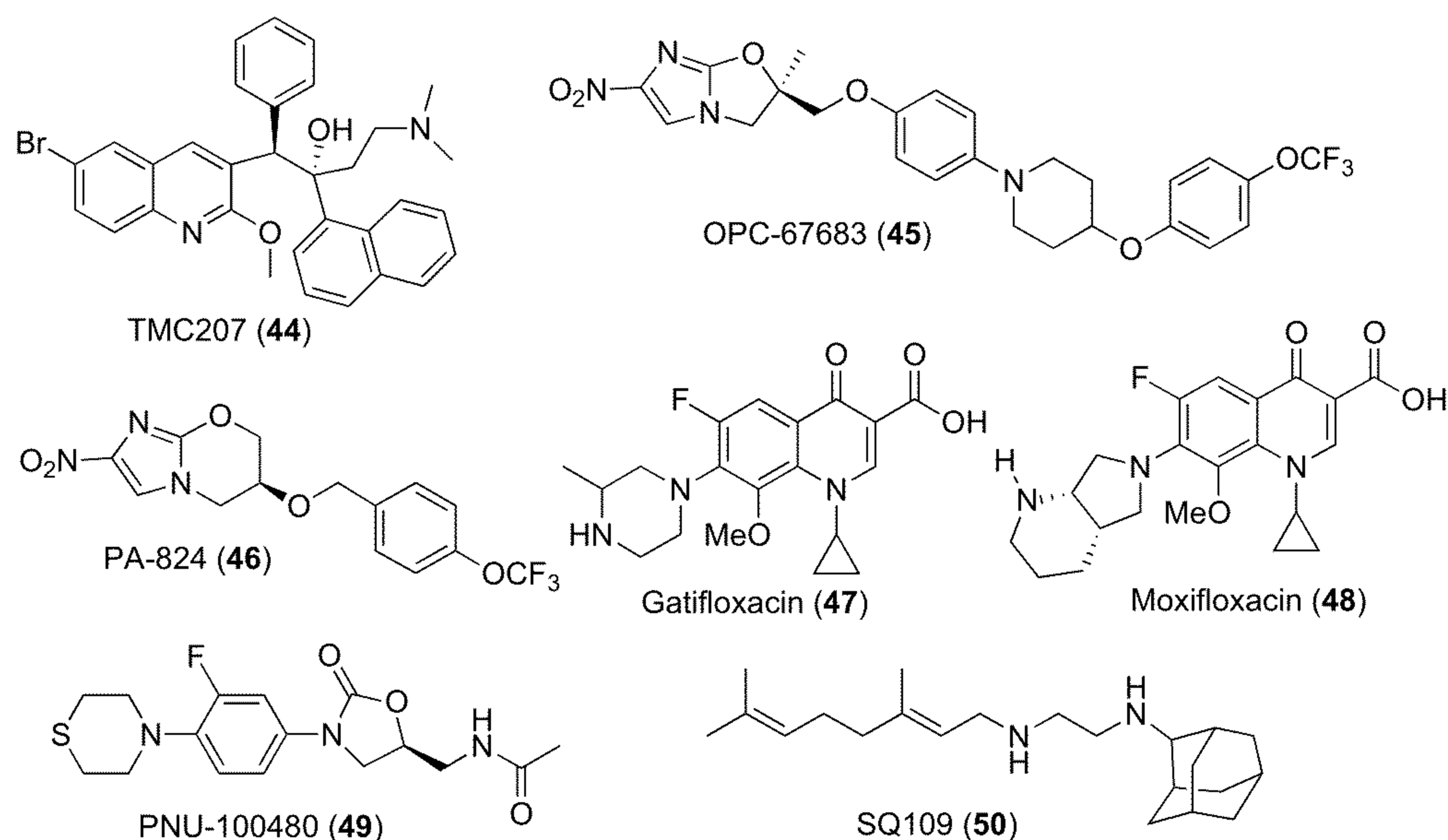


Fig. (12). Structures of a new drug TMC207 (44) and promising anti-TB drug candidates 45-50.

tuberculosis [123,124]. The quinolones **47** and **48** inhibit *M. tuberculosis* DNA gyrase. PNU-100480 (**49**) is an oxazolidinone anti-TB agent whose mechanism of action is through the inhibition of an early step in the initiation phase of protein synthesis [125]. PNU-100480 (**49**) exhibited promising drug properties in Phase II clinical trials [126]. SQ109 (**50**) has an alkaloid-like structure with a diamine-containing geranyl moiety (Fig. (12)). SQ109 (**50**) affects mycobacterial cell wall synthesis, targeting at a membrane transporter of trehalose monomycolate (MmpL3) that involved in mycolic acid donation to the cell wall core of *M. tuberculosis* [127]. SQ109 (**50**) is in Phase II clinical trials [128]. The new anti-TB drug TMC207 (**44**) and the drug candidates **45-50** are likely to play a crucial role in future tuberculosis therapy.

There are also few potent anti-TB agents in preclinical phase. Caprazamycin and capuramycin, pyrimidine-derived antibiotics isolated from actinomycete bacteria of the genus *Streptomyces* [129,130], are anti-TB natural product leads. CPZEN-45 (**51**) is a caprazamycin analog which is in pre-clinical pipeline for anti-TB drug development [131], while SQ641 (**52**), a capuramycin derivative (Fig. (13)), exhibits significant anti-TB activity in a mouse model [132]. BDM31343 (**53**), an oxadiazole compound, is a potent inhibitor of EthR, a repressor protein of the ethionamide (a second-line anti-TB drug) resistance in mycobacteria (Fig. (13)) [133]. BDM31343 (**53**) is in preclinical pipeline. Alkaloids with an oxadiazole moiety are rare in Nature, only few natural oxadiazole alkaloids have been reported so far

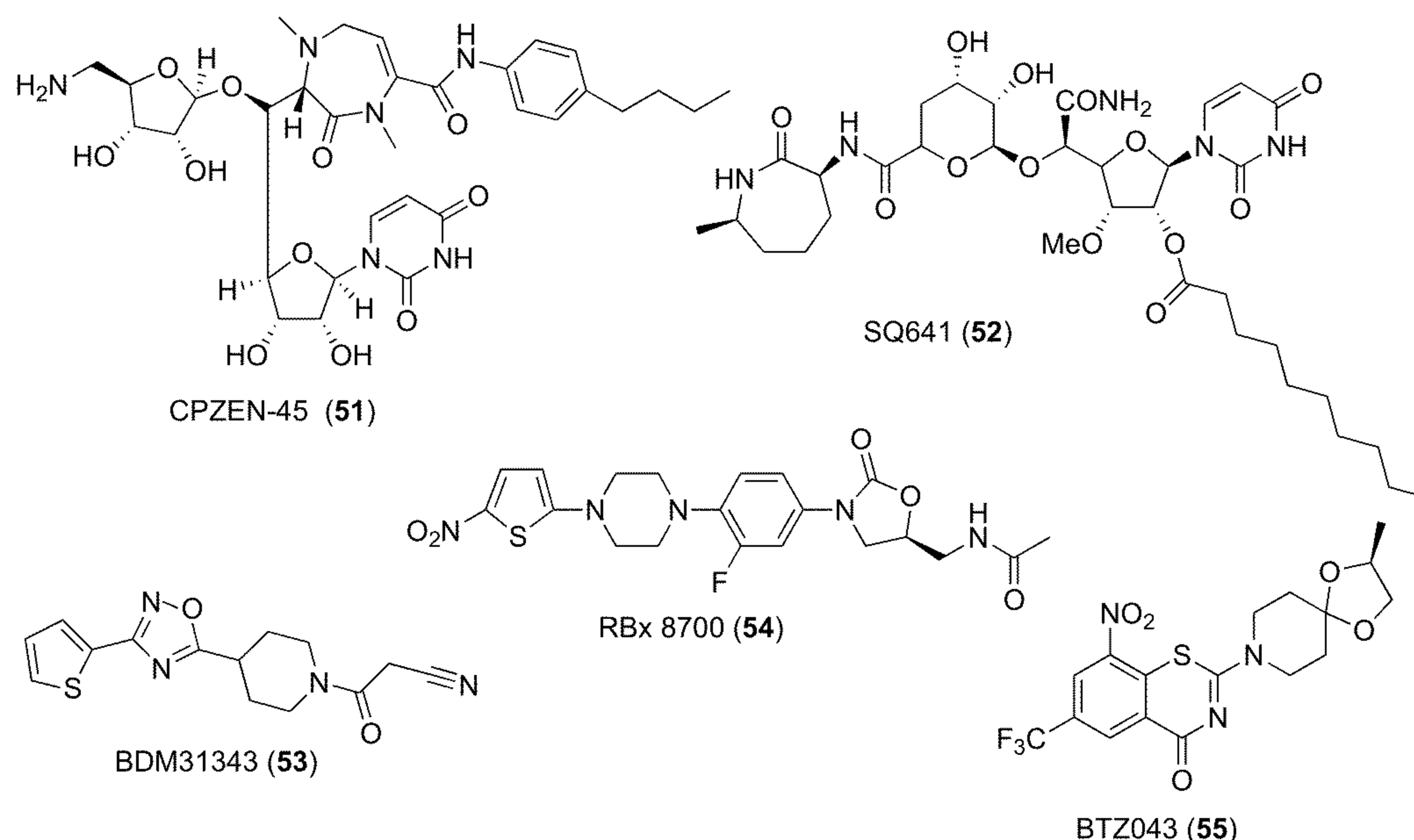


Fig. (13). Anti-TB leads (51-55) in preclinical phase.

[134,135]. An oxazolidinone RBx 8700 (**54**) showed excellent antimycobacterial and antibacterial activities (Fig. (13)) [136,137], and thus entering a preclinical phase. Natural oxazolidinone alkaloids are rarely found in Nature [117,118]. Among anti-TB drug candidates being evaluated in preclinical phase, BTZ043 (**55**), an anti-TB lead decorated with an alkaloid-like scaffold known as benzothiazinone (Fig. (13)), is considered as a new class of antimycobacterial agents, because it is a prodrug that generates a nitroso intermediate *in vivo* [138]. The nitroso intermediate in turn causes a suicide inhibition of decaprenylphosphoryl- β -D-ribose 2' oxidase, which is the enzyme responsible for cell wall arabinogalactan biosynthesis of *M. tuberculosis* [139]. Benzothiazinone is extremely rare in Nature [140].

ALKALOID SCAFFOLDS IN SMOKING CESSATION DRUGS

Tobacco smoking is a serious threat to human health, and causes several health risks and particular diseases, for example, lung cancer, coronary heart disease, and chronic obstructive pulmonary disease. Nicotine (**56**), a psychoactive alkaloid in tobacco, is known to cause smoking addiction (Fig. (14)). Actions of nicotine (**56**) are mediated by nicotinic acetylcholine receptors (nAChRs), which lead to the release of neurotransmitters such as dopamine, glutamate, and GABA (γ -aminobutyric acid) [141], and thus giving behavioral effects. The fact that the nicotine withdrawal syndrome that accompanies smoking cessation would lead to negative effects (such as anxiety, depressed mood, difficulty concentrating, disrupted cognition, and irritability) urges the need for the medication of tobacco cessation [141]. Apart from the nicotine replacement therapy (in the forms of transdermal patches, chewing gums, nasal sprays, and inhalers) that is widely employed as a substitution for tobacco smoking, non-nicotine drugs are also used for smoking cessation. The drugs for smoking cessation commonly target at nAChRs, monoamine oxidase B, dopamine D3 receptor, and cannabinoid receptor type 1.

Alkaloid scaffolds play a major role in drugs for smoking cessation therapy. Bupropion (**57**) and varenicline (**58**) are non-nicotine drugs (Fig. (14)) using as a first-line treatment for smoking cessation. Both bupropion (**57**) and varenicline (**58**) were initially designed as antidepressant drugs, however, they showed efficacy equal to or better than nicotine replacement therapy [142]. The mechanism of action of bupropion (**57**) is through the inhibition of dopamine and norepinephrine reuptake [143,144]. The structure of varenicline (**58**) is inspired by a plant alkaloid, cytisine (**59**) (Fig. (14)). Varenicline (**58**) exhibits its activity as a partial agonist of nAChRs, and thus causing the release of a constant low level of dopamine (a reward neurotransmitter after tobacco smoking) [145]. It is worth mentioning that the discovery of varenicline (**58**) skeleton was inspired by morphine (**60**) and its antinociceptive activity; it was previously found that morphine derivatives with the [3.3.1]-bicyclic skeleton (i.e. **60a**) exhibited morphine-like antinociceptive activity, as did the modified [3.2.1]-bicyclic derivative (**60b**) (Fig. (14)) [146]. Surprisingly, this analogy could also be applied for cytisine (**59**) and its nAChRs binding activity; the [3.2.1]-bicyclic cytisine analog (i.e. **59b**) exhibited the same activity as that of the [3.3.1]-bicyclic skeleton (**59a**) (Fig. (14)) [145]. Varenicline (**58**) is the [3.2.1]-bicyclic version of a natural product cytisine (**59**). A bicyclic alkaloid cytisine (**59**) is normally found in few plants including *Laburnum anagyroides*, *Sophora alopecuroides*, *Thermopsis lanceolata*, and *Caragana sinica* [147]. Although cytisine (**59**) has been widely used for smoking cessation in some countries in Eastern Europe, it has not yet been approved in U.S.A and Western Europe. Cytisine (**59**) is now in Phase III clinical trials for smoking cessation in U.S.A. [148]. Nalmefene (**61**) and naltrexone (**62**), morphine derivatives acting as opioid-system modulators (Fig. (14)), are in phase II clinical trials for smoking cessation [149]. Both nalmefene (**61**) and naltrexone (**62**) are originally designed for the management of alcohol dependence (alcoholism treatment).

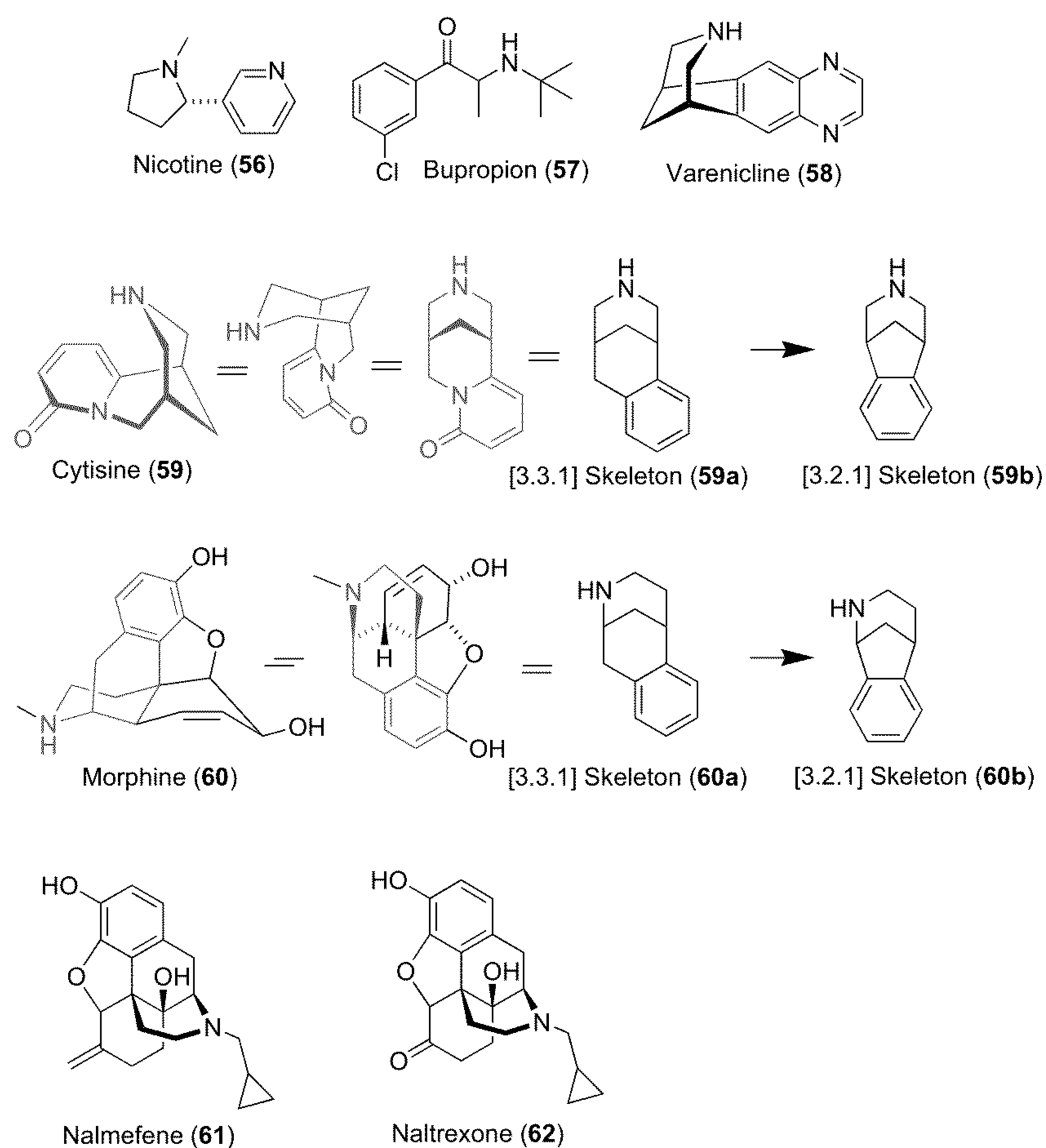


Fig. (14). Structures of nicotine (56), bupropion (57), varenicline (58), cytisine (59), and morphine (60); [3.3.1]- and [3.2.1]-bicyclic skeletons (59a, 59b, 60a, and 60b); and drug candidates 61 and 62.

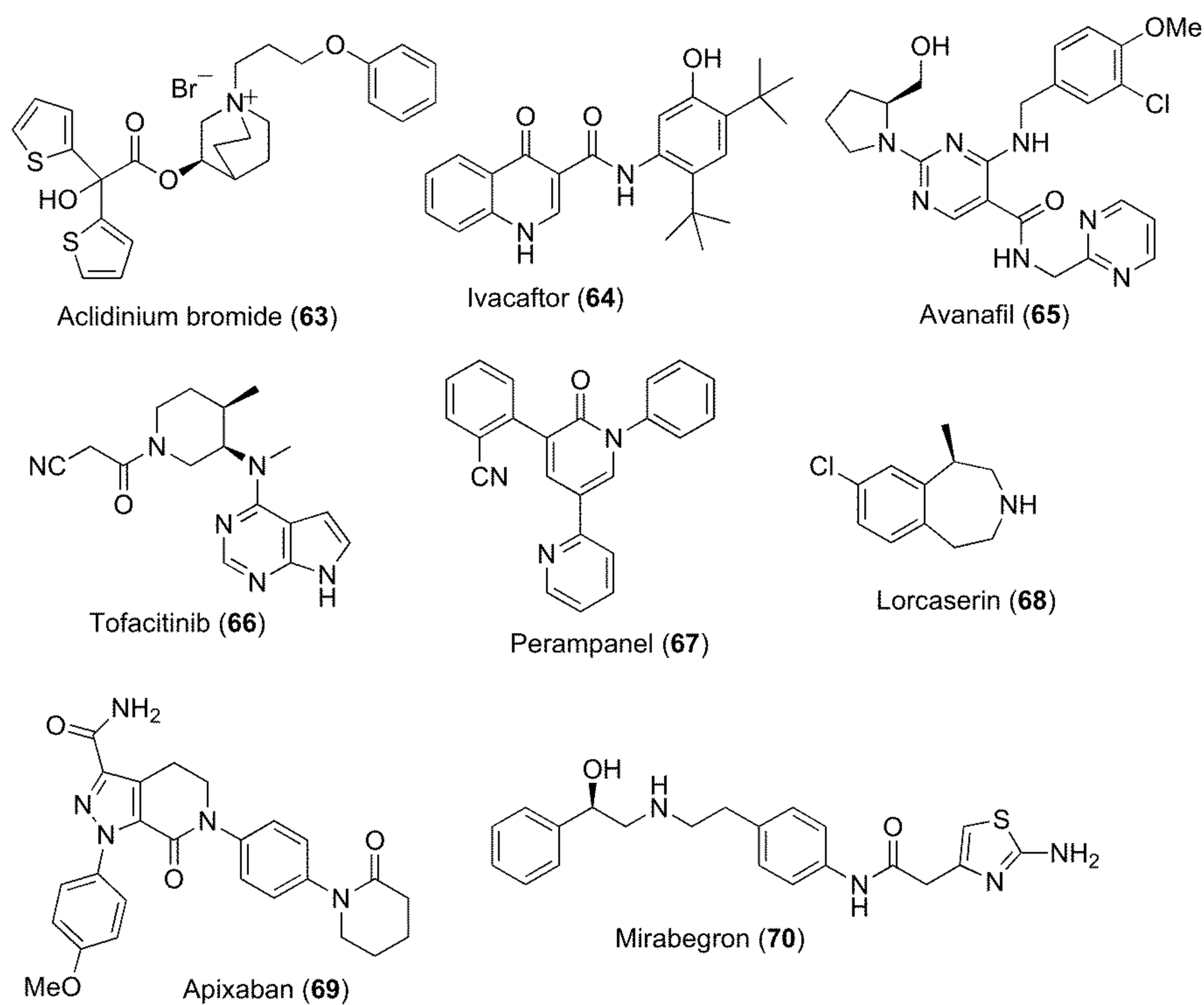


Fig. (15). Structure of aclidinium bromide (63), ivacaftor (64), avanafil (65), tofacitinib (66), perampanel (67), lorcaserin (68), apixaban (69), and mirabegron (70).

MISCELLANEOUS DRUGS (APPROVED IN 2012) WITH ALKALOID SCAFFOLDS

Acidinium bromide (**63**) is a new drug approved in 2012 for the treatment of chronic obstructive pulmonary disease (COPD) (Fig. (15)). Acidinium bromide (**63**) is a muscarinic receptor antagonist which is inspired by natural occurring atropine and scopolamine alkaloids [150]. Traditional medicine used atropine and scopolamine for centuries for the treatment of asthma [150]. Ivacaftor (**64**), a quinolin-4-one derivative, was approved for the treatment of cystic fibrosis (Fig. (15)). Several natural alkaloids with a quinolin-4-one scaffold are found in living organisms [36,37]. Ivacaftor (**64**), also known as Vx-770, enhances ATP-independent activity and increases the open time of wild-type cystic fibrosis transmembrane conductance regulator chloride channel in an ATP-dependent manner [151]. Avanafil (**65**), a pyrimidine derivative (Fig. (15)), was approved in 2012 for the treatment of erectile dysfunction; it is a potent and highly selective phosphodiesterase-5 inhibitor [152].

Tofacitinib (**66**) is an inhibitor of Janus protein tyrosine kinase family (JAK) approved in 2012 for the treatment of rheumatoid arthritis (Fig. (15)) [153]. The drug tofacitinib (**66**), also known as CP-690,550, is also in clinical trials for the treatment of psoriasis [154], chronic inflammatory disease (ulcerative colitis) [155], and organ transplant rejection [156]. Perampanel (**67**) was approved for the treatment of epilepsy; it is a selective α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor antagonist that modulates AMPA-mediated excitatory neurotransmission (Fig. (15)) [157]. Lorcaserin (**68**), a benzazepine alkaloid (Fig. (15)), is a selective serotonin 5-hydroxytryptamine_{2C} (5-HT_{2C}) receptor agonist [158]. The benzazepine scaffold is found in many naturally occurring alkaloids, for example, harringtonine or cephalotaxine [48,49], isopavine [159], and lennoxamine [160]. Lorcaserin (**68**) was approved in 2012 for the treatment of obesity (weight management). Apixaban (**69**) is an anticoagulant drug for the treatment of stroke and systemic embolism in patients with non-valvular atrial fibrillation (Fig. (15)). Apixaban (**69**) is a potent, direct, selective, and orally active inhibitor of coagulation factor Xa [161]. Mirabegron (**70**), a thiazole derivative (Fig. (15)), is β_3 -adrenoceptor agonist [162] approved for the treatment of overactive bladder.

CONFLICT OF INTEREST

The author(s) confirm that this article content has no conflicts of interest.

ACKNOWLEDGEMENTS

P. K. is supported by the Center of Excellence on Environmental Health and Toxicology, Science & Technology Postgraduate Education and Research Development Office (PERDO), Ministry of Education.

REFERENCES

- [1] Mullard, A. 2012 FDA drug approvals. *Nat. Rev. Drug. Discov.*, **2013**, *12* (2), 87-90.
- [2] Kramer, R.; Cohen, D. Functional genomics to new drug targets. *Nat. Rev. Drug. Discov.*, **2004**, *3* (11), 965-72.
- [3] Burbaum, J.; Tobal, G. M. Proteomics in drug discovery. *Curr. Opin. Chem. Biol.*, **2002**, *6* (4), 427-33.
- [4] Wierzba, K.; Muroi, M.; Osada, H. Proteomics accelerating the identification of the target molecule of bioactive small molecules. *Curr. Opin. Chem. Biol.*, **2011**, *15* (1), 57-65.
- [5] Miao, Q.; Zhang, C. C.; Kast, J. Chemical proteomics and its impact on the drug discovery process. *Expert. Rev. Proteomics*, **2012**, *9* (3), 281-91.
- [6] Raida, M. Drug target deconvolution by chemical proteomics. *Curr. Opin. Chem. Biol.*, **2011**, *15* (4), 570-5.
- [7] Kolb, P.; Ferreira, R. S.; Irwin, J. J.; Shoichet, B. K. Docking and chemoinformatic screens for new ligands and targets. *Curr. Opin. Biotechnol.*, **2009**, *20* (4), 429-36.
- [8] Yu, H.; Chen, J.; Xu, X.; Li, Y.; Zhao, H.; Fang, Y.; Li, X.; Zhou, W.; Wang, W.; Wang, Y., A systematic prediction of multiple drug-target interactions from chemical, genomic, and pharmacological data. *PLoS One*, **2012**, *7* (5), e37608.
- [9] Buchan, N. S.; Rajpal, D. K.; Webster, Y.; Alatorre, C.; Gudivada, R. C.; Zheng, C.; Sansseau, P.; Koehler, J. The role of translational bioinformatics in drug discovery. *Drug Discov. Today*, **2011**, *16* (9-10), 426-34.
- [10] Zhao, S.; Li, S. Network-based relating pharmacological and genomic spaces for drug target identification. *PLoS One*, **2010**, *5* (7), e11764.
- [11] Nguyen, T. B.; Lozach, O.; Surpateanu, G.; Wang, Q.; Retailleau, P.; Iorga, B. I.; Meijer, L.; Gueritte, F. Synthesis, biological evaluation, and molecular modeling of natural and unnatural flavonoidal alkaloids, inhibitors of kinases. *J. Med. Chem.*, **2012**, *55* (6), 2811-9.
- [12] Leal, A. S.; Wang, R.; Salvador, J. A.; Jing, Y. Synthesis of novel heterocyclic oleanolic acid derivatives with improved antiproliferative activity in solid tumor cells. *Org. Biomol. Chem.*, **2013**, *11* (10), 1726-38.
- [13] Nusslein-Volhard, C.; Wieschaus, E. Mutations affecting segment number and polarity in *Drosophila*. *Nature*, **1980**, *287* (5785), 795-801.
- [14] Chen, J. K.; Taipale, J.; Cooper, M. K.; Beachy, P. A. Inhibition of Hedgehog signaling by direct binding of cyclopamine to Smoothened. *Genes Dev.*, **2002**, *16* (21), 2743-8.
- [15] Kar, S.; Deb, M.; Sengupta, D.; Shilpi, A.; Bhutia, S. K.; Patra, S. K. Intricacies of hedgehog signaling pathways: a perspective in tumorigenesis. *Exp. Cell Res.*, **2012**, *318* (16), 1959-72.
- [16] Heretsch, P.; Tzagkaroulaki, L.; Giannis, A. Modulators of the hedgehog signaling pathway. *Bioorg. Med. Chem.*, **2010**, *18* (18), 6613-24.
- [17] Gould, A.; Missailidis, S. Targeting the hedgehog pathway: the development of cyclopamine and the development of anti-cancer drugs targeting the hedgehog pathway. *Mini Rev. Med. Chem.*, **2011**, *11* (3), 200-13.
- [18] Lee, M. J.; Hatton, B. A.; Villavicencio, E. H.; Khanna, P. C.; Friedman, S. D.; Ditzler, S.; Pullar, B.; Robison, K.; White, K. F.; Tunkey, C.; LeBlanc, M.; Randolph-Habecker, J.; Knoblaugh, S. E.; Hansen, S.; Richards, A.; Wainwright, B. J.; McGovern, K.; Olson, J. M. Hedgehog pathway inhibitor saridegib (IPI-926) increases lifespan in a mouse medulloblastoma model. *Proc. Natl. Acad. Sci. USA*, **2012**, *109* (20), 7859-64.
- [19] Tremblay, M. R.; McGovern, K.; Read, M. A.; Castro, A. C. New developments in the discovery of small molecule Hedgehog pathway antagonists. *Curr. Opin. Chem. Biol.*, **2010**, *14* (3), 428-35.
- [20] Mahindroo, N.; Punchihewa, C.; Fujii, N. Hedgehog-Gli signaling pathway inhibitors as anticancer agents. *J. Med. Chem.*, **2009**, *52* (13), 3829-45.
- [21] Peukert, S.; Miller-Moslin, K. Small-molecule inhibitors of the hedgehog signaling pathway as cancer therapeutics. *ChemMedChem*, **2010**, *5* (4), 500-12.
- [22] Castanedo, G. M.; Wang, S.; Robarge, K. D.; Blackwood, E.; Burdick, D.; Chang, C.; Dijkgraaf, G. J.; Gould, S.; Gunzner, J.; Guichert, O.; Halladay, J.; Khojasteh, C.; Lee, L.; Marsters, J. C. Jr.; Murray, L.; Peterson, D.; Plise, E.; Salphati, L.; de Sauvage, F. J.; Wong, S.; Sutherlin, D. P. Second generation 2-pyridyl biphenyl amide inhibitors of the hedgehog pathway. *Bioorg. Med. Chem. Lett.*, **2010**, *20* (22), 6748-53.
- [23] Hovhannisyan, A.; Matz, M.; Gebhardt, R. From teratogens to potential therapeutics: natural inhibitors of the Hedgehog signaling network come of age. *Planta Med.*, **2009**, *75* (13), 1371-80.

- [24] Yang, B.; Hird, A. W.; Russell, D. J.; Fauber, B. P.; Dakin, L. A.; Zheng, X.; Su, Q.; Godin, R.; Brassil, P.; Devereaux, E.; Janetka, J. W. Discovery of novel hedgehog antagonists from cell-based screening: Isosteric modification of p38 bisamides as potent inhibitors of SMO. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (14), 4907-11.
- [25] Rifai, Y.; Arai, M. A.; Sadhu, S. K.; Ahmed, F.; Ishibashi, M. New Hedgehog/GLI signaling inhibitors from *Excoecaria agallocha*. *Bioorg. Med. Chem. Lett.*, **2011**, *21* (2), 718-22.
- [26] D'Yakonov, A. L.; Telezhenetskaya, M. V. Quinazoline alkaloids in nature. *Chem. Nat. Compd.*, **1997**, *33* (3), 221-267.
- [27] Santos, V. A.; Regasini, L. O.; Nogueira, C. R.; Passerini, G. D.; Martinez, I.; Bolzani, V. S.; Graminha, M. A.; Cicarelli, R. M.; Furlan, M. Antiprotozoal sesquiterpene pyridine alkaloids from *Maytenus ilicifolia*. *J. Nat. Prod.*, **2012**, *75* (5), 991-5.
- [28] Furukawa, M.; Makino, M.; Uchiyama, T.; Ishimi, K.; Ichinohe, Y.; Fujimoto, Y. Sesquiterpene pyridine alkaloids from *Hippocratea excelsa*. *Phytochemistry*, **2002**, *59* (7), 767-77.
- [29] Jinbo, Z.; Mangan, W.; Wenjun, W.; Zhiqing, J.; Zhaonong, H. Insecticidal sesquiterpene pyridine alkaloids from *Euonymus* species. *Phytochemistry*, **2002**, *61* (6), 699-704.
- [30] Ngadjui, B. T.; Tamboue, H.; Ayafor, J. F.; Connolly, J. D. Thomandersine and isothomandersine, 2-indolinone alkaloids from *Thomandersia laurifolia*. *Phytochemistry*, **1995**, *39* (5), 1249-1251.
- [31] Mohn, T.; Potterat, O.; Hamburger, M. Quantification of active principles and pigments in leaf extracts of *Isatis tinctoria* by HPLC/UV/MS. *Planta Med.*, **2007**, *73* (2), 151-6.
- [32] Rateb, M. E.; Ebel, R. Secondary metabolites of fungi from marine habitats. *Nat. Prod. Rep.*, **2011**, *28* (2), 290-344.
- [33] Wu, X.; Liu, Y.; Sheng, W.; Sun, J.; Qin, G. Chemical constituents of *Isatis indigotica*. *Planta Med.*, **1997**, *63* (1), 55-7.
- [34] Lewis, J. R. Muscarine, oxazole, isoxazole, thiazole, imidazole, and peptide alkaloids and other miscellaneous alkaloids. *Nat. Prod. Rep.*, **1990**, *7* (5), 365-375.
- [35] Arunotayanun, W.; Gibbons, S. Natural product 'legal highs'. *Nat. Prod. Rep.*, **2012**, *29* (11), 1304-16.
- [36] Michael, J. P. Quinoline, quinazoline and acridone alkaloids. *Nat. Prod. Rep.*, **2008**, *25* (1), 166-87.
- [37] Michael, J. P. Quinoline, quinazoline and acridone alkaloids. *Nat. Prod. Rep.*, **2007**, *24* (1), 223-46.
- [38] Yang, J. L.; Liu, L. L.; Shi, Y. P. Limonoids and quinoline alkaloids from *Dictamnus dasycarpus*. *Planta Med.*, **2011**, *77* (3), 271-6.
- [39] Varamini, P.; Javidnia, K.; Soltani, M.; Mehdipour, A. R.; Ghaderi, A. Cytotoxic activity and cell cycle analysis of quinoline alkaloids isolated from *Haplophyllum canaliculatum* Boiss. *Planta Med.*, **2009**, *75* (14), 1509-16.
- [40] Jin, Z. Muscarine, imidazole, oxazole, and thiazole alkaloids. *Nat. Prod. Rep.*, **2011**, *28* (6), 1143-91.
- [41] Jin, Z. Muscarine, imidazole, oxazole and thiazole alkaloids. *Nat. Prod. Rep.*, **2009**, *26* (3), 382-445.
- [42] Jin, Z. Muscarine, imidazole, oxazole and thiazole alkaloids. *Nat. Prod. Rep.*, **2005**, *22* (2), 196-229.
- [43] Ali, Z.; Ferreira, D.; Carvalho, P.; Avery, M. A.; Khan, I. A. Nigellidine-4-O-sulfite, the first sulfated indazole-type alkaloid from the seeds of *Nigella sativa*. *J. Nat. Prod.*, **2008**, *71* (6), 1111-1112.
- [44] Atta ur, R.; Malik, S.; Sadiq Hasan, S.; Iqbal Choudhary, M.; Ni, C. Z.; Clardy, J. Nigellidine - A new indazole alkaloid from the seeds of *Nigella sativa*. *Tetrahedron Lett.*, **1995**, *36* (12), 1993-1996.
- [45] Atta ur, R.; Malik, S.; Cun-heng, H.; Clardy, J. Isolation and structure determination of nigellicine. A novel alkaloid from the seeds of *Nigella sativa*. *Tetrahedron Lett.*, **1985**, *26* (23), 2759-2762.
- [46] Schmidt, A.; Beutler, A.; Snovydyovych, B. Recent advances in the chemistry of indazoles. *Eur. J. Org. Chem.*, **2008**, (24), 4073-4095.
- [47] Perdue, R. E.; Spetzman, L. A.; Powell, R. G. Cephalotaxus-Source of harringtonine, a promising new anti-cancer alkaloid. *American Horticultural Magazine*, **1970**, *49*, 19-22.
- [48] Powell, R. G.; Weisleder, D.; Smith Jr, C. R.; Rohwedder, W. K. Structures of harringtonine, isoharringtonine, and homoharringtonine. *Tetrahedron Lett.*, **1970**, *11* (11), 815-818.
- [49] Powell, R. G.; Weisleder, D.; Smith Jr, C. R.; Wolff, I. A. Structure of cephalotaxine and related alkaloids. *Tetrahedron Lett.*, **1969**, *10* (46), 4081-4084.
- [50] Powell, R. G.; Weisleder, D.; Smith, C. R. Antitumor alkaloids from *Cephalotaxus harringtonia*: Structure and activity. *J. Pharm. Sci.*, **1972**, *61* (8), 1227-1230.
- [51] Cortes, J.; Lipton, J. H.; Rea, D.; Digumarti, R.; Chuah, C.; Nanda, N.; Benichou, A. C.; Craig, A. R.; Michallet, M.; Nicolini, F. E.; Kantarjian, H. Phase 2 study of subcutaneous omacetaxine mepesuccinate after TKI failure in patients with chronic-phase CML with T315I mutation. *Blood*, **2012**, *120* (13), 2573-80.
- [52] Chill, L.; Rudi, A.; Benayahu, Y.; Kashman, Y. Violatinctamine, a new heterocyclic compound from the marine tunicate *Cystodytes cf violatinctus*. *Tetrahedron Lett.*, **2004**, *45* (42), 7925-7928.
- [53] Ojika, M.; Nemoto, T.; Nakamura, M.; Yamada, K. Dolastatin E, a new cyclic hexapeptide isolated from the sea hare *Dolabella auricularia*. *Tetrahedron Lett.*, **1995**, *36* (28), 5057-5058.
- [54] Gerth, K.; Bedorf, N.; Hofle, G.; Irschik, H.; Reichenbach, H. Epothilons A and B: antifungal and cytotoxic compounds from *Sorangium cellulosum* (Myxobacteria). Production, physico-chemical and biological properties. *J. Antibiot. (Tokyo)*, **1996**, *49* (6), 560-3.
- [55] Hoefle, G.; Bedorf, N.; Gerth, K.; Reichenbach, H. German Patent Disclosure, DE 4138042. *Chem. Abstr.*, **1993**, *120*, 52841.
- [56] Bollag, D. M.; McQueney, P. A.; Zhu, J.; Hensens, O.; Koupal, L.; Liesch, J.; Goetz, M.; Lazarides, E.; Woods, C. M. Epothilones, a new class of microtubule-stabilizing agents with a taxol-like mechanism of action. *Cancer Res.*, **1995**, *55* (11), 2325-33.
- [57] Pfeiffer, B.; Hauenstein, K.; Merz, P.; Gertsch, J.; Altmann, K. H. Synthesis and SAR of C12-C13-oxazoline derivatives of epothilone A. *Bioorg. Med. Chem. Lett.*, **2009**, *19* (14), 3760-3.
- [58] Altmann, K. H. The merger of natural product synthesis and medicinal chemistry: on the chemistry and chemical biology of epothilones. *Org. Biomol. Chem.*, **2004**, *2* (15), 2137-52.
- [59] Sefkow, M.; Kiffe, M.; Hofle, G. Derivatization of the C12-C13 functional groups of epothilones A, B and C. *Bioorg. Med. Chem. Lett.*, **1998**, *8* (21), 3031-6.
- [60] Taori, K.; Paul, V. J.; Luesch, H. Structure and activity of largazole, a potent antiproliferative agent from the Floridian marine cyanobacterium *Symploca* sp. *J. Am. Chem. Soc.*, **2008**, *130* (6), 1806-7.
- [61] Ying, Y.; Taori, K.; Kim, H.; Hong, J.; Luesch, H. Total synthesis and molecular target of largazole, a histone deacetylase inhibitor. *J. Am. Chem. Soc.*, **2008**, *130* (26), 8455-9.
- [62] Hong, J.; Luesch, H. Largazole: from discovery to broad-spectrum therapy. *Nat. Prod. Rep.*, **2012**, *29* (4), 449-56.
- [63] Ungermannova, D.; Parker, S. J.; Nasveschuk, C. G.; Wang, W.; Quade, B.; Zhang, G.; Kuchta, R. D.; Phillips, A. J.; Liu, X. Largazole and its derivatives selectively inhibit ubiquitin activating enzyme (E1). *PLoS One*, **2012**, *7* (1), e29208.
- [64] Neckers, L.; Workman, P. Hsp90 molecular chaperone inhibitors: are we there yet? *Clin. Cancer Res.*, **2012**, *18* (1), 64-76.
- [65] Gartner, E. M.; Silverman, P.; Simon, M.; Flaherty, L.; Abrams, J.; Ivy, P.; Lorusso, P. M., A phase II study of 17-allylamino-17-demethoxygeldanamycin in metastatic or locally advanced, unresectable breast cancer. *Breast Cancer Res. Treat.*, **2012**, *131* (3), 933-7.
- [66] Pacey, S.; Wilson, R. H.; Walton, M.; Eatock, M. M.; Hardcastle, A.; Zetterlund, A.; Arkenau, H. T.; Moreno-Farre, J.; Banerji, U.; Roels, B.; Peachey, H.; Aherne, W.; de Bono, J. S.; Raynaud, F.; Workman, P.; Judson, I., A phase I study of the heat shock protein 90 inhibitor alvespimycin (17-DMAG) given intravenously to patients with advanced solid tumors. *Clin. Cancer Res.*, **2011**, *17* (6), 1561-70.
- [67] Okui, T.; Shimo, T.; Hassan, N. M.; Fukazawa, T.; Kurio, N.; Takaoka, M.; Naomoto, Y.; Sasaki, A. Antitumor effect of novel HSP90 inhibitor NVP-AUY922 against oral squamous cell carcinoma. *Anticancer Res.*, **2011**, *31* (4), 1197-204.
- [68] Barril, X.; Beswick, M. C.; Collier, A.; Drysdale, M. J.; Dymock, B. W.; Fink, A.; Grant, K.; Howes, R.; Jordan, A. M.; Massey, A.; Surgenor, A.; Wayne, J.; Workman, P.; Wright, L., 4-Amino derivatives of the Hsp90 inhibitor CCT018159. *Bioorg. Med. Chem. Lett.*, **2006**, *16* (9), 2543-8.
- [69] Graham, B.; Curry, J.; Smyth, T.; Fazal, L.; Feltell, R.; Harada, I.; Coyle, J.; Williams, B.; Reule, M.; Angove, H.; Cross, D. M.; Lyons, J.; Wallis, N. G.; Thompson, N. T. The heat shock protein 90 inhibitor, AT13387, displays a long duration of action *in vitro* and *in vivo* in non-small cell lung cancer. *Cancer Sci.*, **2012**, *103* (3), 522-7.

- [70] Aladesanmi, A. J.; Nia, R.; Nahrstedt, A. New pyrazole alkaloids from the root bark of *Newbouldia laevis*. *Planta Med.*, **1998**, *64* (1), 90-1.
- [71] Adesanya, S. A.; Nia, R.; Fontaine, C.; Pais, M. Pyrazole alkaloids from *Newbouldia laevis*. *Phytochemistry*, **1994**, *35* (4), 1053-1055.
- [72] Mitchell, R. E.; Greenwood, D. R.; Sarojini, V. An antibacterial pyrazole derivative from *Burkholderia glumae*, a bacterial pathogen of rice. *Phytochemistry*, **2008**, *69* (15), 2704-2707.
- [73] Parameswaran, P. S.; Naik, C. G.; Hegde, V. R. Secondary metabolites from the sponge *Tedania anhelans*: Isolation and characterization of two novel pyrazole acids and other metabolites. *J. Nat. Prod.*, **1997**, *60* (8), 802-803.
- [74] Marimoto, A.; Noda, K.; Watanabe, T.; Takasugi, H. The total synthesis of withasomine, a unique pyrazole alkaloid. *Tetrahedron Lett.*, **1968**, (54), 5707-10.
- [75] Scherlach, K.; Schuemann, J.; Dahse, H. M.; Hertweck, C. Aspermidine A and B, prenylated isoindolinone alkaloids from the model fungus *Aspergillus nidulans*. *J. Antibiot.*, **2010**, *63* (7), 375-377.
- [76] Zhang, X.; Ye, W.; Zhao, S.; Che, C. T. Isoquinoline and isoindole alkaloids from *Menispermum dauricum*. *Phytochemistry*, **2004**, *65* (7), 929-932.
- [77] Zhang, Y.; Wang, T.; Pei, Y.; Hua, H.; Feng, B. Aspergillin PZ, a novel isoindole-alkaloid from *Aspergillus awamori*. *J. Antibiot.*, **2002**, *55* (8), 693-695.
- [78] Leggans, E. K.; Duncan, K. K.; Barker, T. J.; Schleicher, K. D.; Boger, D. L., A remarkable series of vinblastine analogues displaying enhanced activity and an unprecedented tubulin binding steric tolerance: C20' urea derivatives. *J. Med. Chem.*, **2013**, *56* (3), 628-39.
- [79] Song, W.; Hu, L.; Meng, Y.; Ma, L.; Guo, D.; Liu, X. The effect of vindoline C-16 substituents on the biomimetic coupling reaction: synthesis and cytotoxicity evaluation of the corresponding vinorelbine analogues. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (10), 3485-7.
- [80] Schleicher, K. D.; Sasaki, Y.; Tam, A.; Kato, D.; Duncan, K. K.; Boger, D. L. Total synthesis and evaluation of vinblastine analogues containing systematic deep-seated modifications in the vindoline subunit ring system: core redesign. *J. Med. Chem.*, **2013**, *56* (2), 483-95.
- [81] Song, W.; Lei, M.; Zhao, K.; Hu, L.; Meng, Y.; Guo, D.; Liu, X. Ceric ammonium nitrate-promoted oxidative coupling reaction for the synthesis and evaluation of a series of anti-tumor amide anhydrovinblastine analogs. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (1), 387-90.
- [82] Hu, L.; Song, W.; Meng, Y.; Guo, D.; Liu, X. Synthesis and structure-activity relationship studies of cytotoxic vinorelbine amide analogues. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (24), 7547-50.
- [83] Pommier, Y.; Kohlhagen, G.; Bailly, C.; Waring, M.; Mazumder, A.; Kohn, K. W. DNA sequence- and structure-selective alkylation of guanine N2 in the DNA minor groove by ecteinascidin 743, a potent antitumor compound from the Caribbean tunicate *Ecteinascidia turbinata*. *Biochemistry*, **1996**, *35* (41), 13303-9.
- [84] Boger, D. L.; Johnson, D. S.; Yun, W., (+)- and ent(-)-Duocarmycin SA and (+)- and ent(-)-N-BOC-DSA DNA alkylation properties. Alkylation site models that accommodate the offset AT-rich adenine N3 alkylation selectivity of the enantiomeric agents. *J. Am. Chem. Soc.*, **1994**, *116* (5), 1635-1656.
- [85] Tietze, L. F.; Krewer, B. Novel analogues of CC-1065 and the duocarmycins for the use in targeted tumour therapies. *Anticancer Agents Med. Chem.*, **2009**, *9* (3), 304-325.
- [86] Wolfe, A. L.; Duncan, K. K.; Parelkar, N. K.; Weir, S. J.; Vielhauer, G. A.; Boger, D. L., A novel, unusually efficacious duocarmycin carbamate prodrug that releases no residual byproduct. *J. Med. Chem.*, **2012**, *55* (12), 5878-86.
- [87] Pommier, Y. Drugging topoisomerases: lessons and challenges. *ACS Chem. Biol.*, **2013**, *8* (1), 82-95.
- [88] Liu, Y. Q.; Dai, W.; Wang, C. Y.; Morris-Natschke, S. L.; Zhou, X. W.; Yang, L.; Yang, X. M.; Li, W. Q.; Lee, K. H. Design and one-pot synthesis of new 7-acyl camptothecin derivatives as potent cytotoxic agents. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (24), 7659-61.
- [89] Alloatti, D.; Giannini, G.; Vesci, L.; Castorina, M.; Pisano, C.; Badaloni, E.; Cabri, W. Camptothecins in tumor homing via an RGD sequence mimetic. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (20), 6509-12.
- [90] Chen, Y.; Su, Y. H.; Wang, C. H.; Wu, J. M.; Chen, J. C.; Tseng, S. H. Induction of apoptosis and cell cycle arrest in glioma cells by GL331 (a topoisomerase II inhibitor). *Anticancer Res.*, **2005**, *25* (6B), 4203-8.
- [91] Barret, J. M.; Kruczynski, A.; Vispe, S.; Annereau, J. P.; Brel, V.; Guminski, Y.; Delcros, J. G.; Lansiaux, A.; Guilbaud, N.; Imbert, T.; Bailly, C., F14512, a potent antitumor agent targeting topoisomerase II vectored into cancer cells via the polyamine transport system. *Cancer Res.*, **2008**, *68* (23), 9845-53.
- [92] Kamal, A.; Tamboli, J. R.; Vishnuvardhan, M. V.; Adil, S. F.; Nayak, V. L.; Ramakrishna, S. Synthesis and anticancer activity of heteroaromatic linked 4 β -amido podophyllotoxins as apoptotic inducing agents. *Bioorg. Med. Chem. Lett.*, **2013**, *23* (1), 273-80.
- [93] Ploypradith, P.; Mahidol, C.; Sahakitpichan, P.; Wongbundit, S.; Ruchirawat, S., A highly efficient synthesis of lamellarins K and L by the Michael addition/ring-closure reaction of benzylidihydroisoquinoline derivatives with ethoxycarbonyl-beta-nitrostyrenes. *Angew. Chem. Int. Ed. Engl.*, **2004**, *43* (7), 866-8.
- [94] Ploypradith, P.; Kagan, R. K.; Ruchirawat, S. Utility of polymer-supported reagents in the total synthesis of lamellarins. *J. Org. Chem.*, **2005**, *70* (13), 5119-25.
- [95] Banwell, M.; Flynn, B.; Hockless, D. Convergent total synthesis of lamellarin K. *Chem. Commun.*, **1997**, (23), 2259-2260.
- [96] Ploypradith, P.; Petchmanee, T.; Sahakitpichan, P.; Litvinas, N. D.; Ruchirawat, S. Total synthesis of natural and unnatural lamellarins with saturated and unsaturated D-rings. *J. Org. Chem.*, **2006**, *71* (25), 9440-8.
- [97] Chittchang, M.; Gleeson, M. P.; Ploypradith, P.; Ruchirawat, S. Assessing the drug-likeness of lamellarins, a marine-derived natural product class with diverse oncological activities. *Eur. J. Med. Chem.*, **2010**, *45* (6), 2165-72.
- [98] Chittchang, M.; Batsomboon, P.; Ruchirawat, S.; Ploypradith, P. Cytotoxicities and structure-activity relationships of natural and unnatural lamellarins toward cancer cell lines. *ChemMedChem*, **2009**, *4* (3), 457-65.
- [99] Pla, D.; Albericio, F.; Alvarez, M. Recent advances in lamellarin alkaloids: isolation, synthesis and activity. *Anticancer Agents Med. Chem.*, **2008**, *8* (7), 746-60.
- [100] Yamaguchi, T.; Fukuda, T.; Ishibashi, F.; Iwao, M. The first total synthesis of lamellarin α 20-sulfate, a selective inhibitor of HIV-1 integrase. *Tetrahedron Lett.*, **2006**, *47* (22), 3755-3757.
- [101] Facompre, M.; Tardy, C.; Bal-Mahieu, C.; Colson, P.; Perez, C.; Manzanares, I.; Cuevas, C.; Bailly, C. Lamellarin D: a novel potent inhibitor of topoisomerase I. *Cancer Res.*, **2003**, *63* (21), 7392-9.
- [102] Han, H. J.; Tan, N. H.; Zeng, G. Z.; Fan, J. T.; Huang, H. Q.; Ji, C. J.; Jia, R. R.; Zhao, Q. S.; Zhang, Y. J.; Hao, X. J.; Wang, L. Q. Natural inhibitors of DNA topoisomerase I with cytotoxicities. *Chem. Biodivers.*, **2008**, *5* (7), 1364-8.
- [103] Fayad, W.; Fryknas, M.; Brnjic, S.; Olofsson, M. H.; Larsson, R.; Linder, S. Identification of a novel topoisomerase inhibitor effective in cells overexpressing drug efflux transporters. *PLoS One*, **2009**, *4* (10), e7238.
- [104] Dong, G.; Wang, S.; Miao, Z.; Yao, J.; Zhang, Y.; Guo, Z.; Zhang, W.; Sheng, C. New tricks for an old natural product: discovery of highly potent evodiamine derivatives as novel antitumor agents by systemic structure-activity relationship analysis and biological evaluations. *J. Med. Chem.*, **2012**, *55* (17), 7593-613.
- [105] Zhang, Y.; Zheng, L.; Zhang, J.; Dai, B.; Wang, N.; Chen, Y.; He, L. Antitumor activity of taspine by modulating the EGFR signaling pathway of Erk1/2 and Akt *in vitro* and *in vivo*. *Planta Med.*, **2011**, *77* (16), 1774-81.
- [106] Castelli, S.; Katkar, P.; Vassallo, O.; Falconi, M.; Linder, S.; Desideri, A., A natural anticancer agent thaspine targets human topoisomerase IB. *Anticancer Agents Med. Chem.*, **2013**, *13* (2), 356-63.
- [107] Zhang, J.; Zhang, Y.; Zhang, S.; Wang, S.; He, L. Discovery of novel taspine derivatives as antiangiogenic agents. *Bioorg. Med. Chem. Lett.*, **2010**, *20* (2), 718-21.
- [108] Altieri, A.; Franceschin, M.; Nocioni, D.; Alvino, A.; Casagrande, V.; Scarpati, M. L.; Bianco, A. Total synthesis of taspine and a symmetrical analogue: Study of binding to G-quadruplex DNA by ESI-MS. *Eur. J. Org. Chem.*, **2013**, (1), 191-196.
- [109] Zhan, Y.; Zhang, Y.; Liu, C.; Zhang, J.; Smith, W. W.; Wang, N.; Chen, Y.; Zheng, L.; He, L., A novel taspine derivative,

- HMQ1611, inhibits breast cancer cell growth via estrogen receptor alpha and EGF receptor signaling pathways. *Cancer Prev. Res. (Phila)*, **2012**, *5* (6), 864-73.
- [110] Lu, W.; Dai, B.; Ma, W.; Zhang, Y., A novel taspine analog, HMQ1611, inhibits growth of non-small cell lung cancer by inhibiting angiogenesis. *Oncol. Lett.*, **2012**, *4* (5), 1109-1113.
- [111] World Health Organization, WHO Global tuberculosis report. World Health Organization, Geneva, Switzerland, **2012**.
- [112] Staerk, D.; Kesting, J. R.; Sairafianpour, M.; Witt, M.; Asili, J.; Emami, S. A.; Jaroszewski, J. W. Accelerated dereplication of crude extracts using HPLC-PDA-MS-SPE-NMR: quinolinone alkaloids of *Haplophyllum acutifolium*. *Phytochemistry*, **2009**, *70* (8), 1055-61.
- [113] Rodriguez-Guzman, R.; Fulks, L. C.; Radwan, M. M.; Burandt, C. L.; Ross, S. A. Chemical constituents, antimicrobial and antimalarial activities of *Zanthoxylum monophyllum*. *Planta Med.*, **2011**, *77* (13), 1542-4.
- [114] Wang, T. Y.; Wu, J. B.; Hwang, T. L.; Kuo, Y. H.; Chen, J. J., A new quinolone and other constituents from the fruits of *Tetradium ruticarpum*: effects on neutrophil pro-inflammatory responses. *Chem. Biodivers.*, **2010**, *7* (7), 1828-34.
- [115] Adams, M.; Mahringer, A.; Kunert, O.; Fricker, G.; Efferth, T.; Bauer, R. Cytotoxicity and p-glycoprotein modulating effects of quinolones and indoloquinazolines from the Chinese herb *Evodia rutaecarpa*. *Planta Med.*, **2007**, *73* (15), 1554-7.
- [116] Gressler, V.; Stuker, C. Z.; Dias Gde, O.; Dalcol, II; Burrow, R. A.; Schmidt, J.; Wessjohann, L.; Morel, A. F. Quinolone alkaloids from *Waltheria douradinha*. *Phytochemistry*, **2008**, *69* (4), 994-9.
- [117] Tadesse, M.; Strom, M. B.; Svenson, J.; Jaspars, M.; Milne, B. F.; Torfoss, V.; Andersen, J. H.; Hansen, E.; Stensvag, K.; Haug, T. Synoxazolidinones A and B: novel bioactive alkaloids from the ascidian *Synoicum pulmonaria*. *Org. Lett.*, **2010**, *12* (21), 4752-5.
- [118] Norte, M.; Rodriguez, M. L.; Fernández, J. J.; Eguren, L.; Estrada, D. M. Aplysinadiene and (*R,R*) 5 [3,5-dibromo-4-[(2-oxo-5-oxazolidinyl)] methoxyphenyl]-2-oxazolidinone, two novel metabolites from *Aplysina aerophoba* syntheses. *Tetrahedron*, **1988**, *44* (15), 4973-4980.
- [119] Andries, K.; Verhasselt, P.; Guillemont, J.; Gohlmann, H. W.; Neefs, J. M.; Winkler, H.; Van Gestel, J.; Timmerman, P.; Zhu, M.; Lee, E.; Williams, P.; de Chaffoy, D.; Huitric, E.; Hoffner, S.; Cambau, E.; Truffot-Pernot, C.; Lounis, N.; Jarlier, V., A diarylquinoline drug active on the ATP synthase of *Mycobacterium tuberculosis*. *Science*, **2005**, *307* (5707), 223-7.
- [120] Gler, M. T.; Skripconoka, V.; Sanchez-Garavito, E.; Xiao, H.; Cabrera-Rivero, J. L.; Vargas-Vasquez, D. E.; Gao, M.; Awad, M.; Park, S. K.; Shim, T. S.; Suh, G. Y.; Danilovits, M.; Ogata, H.; Kurve, A.; Chang, J.; Suzuki, K.; Tupasi, T.; Koh, W. J.; Seaworth, B.; Geiter, L. J.; Wells, C. D. Delamanid for multidrug-resistant pulmonary tuberculosis. *N. Engl. J. Med.*, **2012**, *366* (23), 2151-60.
- [121] Matsumoto, M.; Hashizume, H.; Tomishige, T.; Kawasaki, M.; Tsubouchi, H.; Sasaki, H.; Shimokawa, Y.; Komatsu, M. OPC-67683, a nitro-dihydro-imidazooxazole derivative with promising action against tuberculosis *in vitro* and in mice. *PLoS Med.*, **2006**, *3* (11), e466.
- [122] Diacon, A. H.; Dawson, R.; du Bois, J.; Narunsky, K.; Venter, A.; Donald, P. R.; van Niekerk, C.; Erondou, N.; Ginsberg, A. M.; Becker, P.; Spigelman, M. K. Phase II dose-ranging trial of the early bactericidal activity of PA-824. *Antimicrob. Agents Chemother.*, **2012**, *56* (6), 3027-31.
- [123] Pranger, A. D.; van Altena, R.; Aarnoutse, R. E.; van Soolingen, D.; Uges, D. R.; Kosterink, J. G.; van der Werf, T. S.; Alffenaar, J. W. Evaluation of moxifloxacin for the treatment of tuberculosis: 3 years of experience. *Eur. Respir. J.*, **2011**, *38* (4), 888-94.
- [124] Rustomjee, R.; Lienhardt, C.; Kanyok, T.; Davies, G. R.; Levin, J.; Mthiyane, T.; Reddy, C.; Sturm, A. W.; Sirgel, F. A.; Allen, J.; Coleman, D. J.; Fourie, B.; Mitchison, D. A., A Phase II study of the sterilising activities of ofloxacin, gatifloxacin and moxifloxacin in pulmonary tuberculosis. *Int. J. Tuberc. Lung Dis.*, **2008**, *12* (2), 128-38.
- [125] Williams, K. N.; Stover, C. K.; Zhu, T.; Tasneen, R.; Tyagi, S.; Grosset, J. H.; Nuermberger, E. Promising antituberculosis activity of the oxazolidinone PNU-100480 relative to that of linezolid in a murine model. *Antimicrob. Agents Chemother.*, **2009**, *53* (4), 1314-9.
- [126] Wallis, R. S.; Jakubiec, W. M.; Kumar, V.; Silvia, A. M.; Paige, D.; Dimitrova, D.; Li, X.; Ladutko, L.; Campbell, S.; Friedland, G.; Mitton-Fry, M.; Miller, P. F. Pharmacokinetics and whole-blood bactericidal activity against *Mycobacterium tuberculosis* of single doses of PNU-100480 in healthy volunteers. *J. Infect. Dis.*, **2010**, *202* (5), 745-51.
- [127] Tahlan, K.; Wilson, R.; Kastrinsky, D. B.; Arora, K.; Nair, V.; Fischer, E.; Barnes, S. W.; Walker, J. R.; Alland, D.; Barry, C. E., 3rd; Boshoff, H. I. SQ109 targets MmpL3, a membrane transporter of trehalose monomycolate involved in mycolic acid donation to the cell wall core of *Mycobacterium tuberculosis*. *Antimicrob. Agents Chemother.*, **2012**, *56* (4), 1797-809.
- [128] Sacksteder, K. A.; Protopopova, M.; Barry, C. E., 3rd; Andries, K.; Nacy, C. A. Discovery and development of SQ109: a new antitubercular drug with a novel mechanism of action. *Future Microbiol.*, **2012**, *7* (7), 823-37.
- [129] Igarashi, M.; Nakagawa, N.; Doi, N.; Hattori, S.; Naganawa, H.; Hamada, M. Caprazamycin B, a novel anti-tuberculosis antibiotic, from *Streptomyces* sp. *J. Antibiot. (Tokyo)*, **2003**, *56* (6), 580-3.
- [130] Yamaguchi, H.; Sato, S.; Yoshida, S.; Takada, K.; Itoh, M.; Seto, H.; Otake, N. Capuramycin, a new nucleoside antibiotic. Taxonomy, fermentation, isolation and characterization. *J. Antibiot. (Tokyo)*, **1986**, *39* (8), 1047-53.
- [131] Adhvaryu, M.; Vakharia, B. Drug-resistant tuberculosis: emerging treatment options. *Clin. Pharmacol.*, **2011**, *3*, 51-67.
- [132] Nikonenko, B. V.; Reddy, V. M.; Protopopova, M.; Bogatcheva, E.; Einck, L.; Nacy, C. A. Activity of SQ641, a capuramycin analog, in a murine model of tuberculosis. *Antimicrob. Agents Chemother.*, **2009**, *53* (7), 3138-9.
- [133] Willand, N.; Dirie, B.; Carette, X.; Bifani, P.; Singhal, A.; Desroses, M.; Leroux, F.; Willery, E.; Mathys, V.; Deprez-Poulain, R.; Delcroix, G.; Frenois, F.; Aumercier, M.; Loch, C.; Villeret, V.; Deprez, B.; Baulard, A. R. Synthetic EthR inhibitors boost antituberculous activity of ethionamide. *Nat. Med.*, **2009**, *15* (5), 537-44.
- [134] Carbone, M.; Li, Y.; Irace, C.; Mollo, E.; Castelluccio, F.; Di Pascale, A.; Cimino, G.; Santamaria, R.; Guo, Y. W.; Gavagnin, M. Structure and cytotoxicity of phidianidines A and B: first finding of 1,2,4-oxadiazole system in a marine natural product. *Org. Lett.*, **2011**, *13* (10), 2516-9.
- [135] Rane, R. A.; Gutte, S. D.; Sahu, N. U. Synthesis and evaluation of novel 1,3,4-oxadiazole derivatives of marine bromopyrrole alkaloids as antimicrobial agent. *Bioorg. Med. Chem. Lett.*, **2012**, *22* (20), 6429-32.
- [136] Rao, M.; Sood, R.; Malhotra, S.; Fatma, T.; Upadhyay, D. J.; Rattan, A. *In vitro* bactericidal activity of oxazolidinone, RBx 8700 against *Mycobacterium tuberculosis* and *Mycobacterium avium* complex. *J. Chemother.*, **2006**, *18* (2), 144-50.
- [137] Rudra, S.; Yadav, A.; Raja Rao, A. V.; Srinivas, A. S.; Pandya, M.; Bhateja, P.; Mathur, T.; Malhotra, S.; Rattan, A.; Salman, M.; Mehta, A.; Cliffe, I. A.; Das, B. Synthesis and antibacterial activity of potent heterocyclic oxazolidinones and the identification of RBx 8700. *Bioorg. Med. Chem. Lett.*, **2007**, *17* (24), 6714-9.
- [138] Makarov, V.; Manina, G.; Mikusova, K.; Mollmann, U.; Ryabova, O.; Saint-Joanis, B.; Dhar, N.; Pasca, M. R.; Buroni, S.; Lucarelli, A. P.; Milano, A.; De Rossi, E.; Belanova, M.; Bobovska, A.; Dianiskova, P.; Kordulakova, J.; Sala, C.; Fullam, E.; Schneider, P.; McKinney, J. D.; Brodin, P.; Christophe, T.; Waddell, S.; Butcher, P.; Albrethsen, J.; Rosenkrands, I.; Brosch, R.; Nandi, V.; Bharath, S.; Gaonkar, S.; Shandil, R. K.; Balasubramanian, V.; Balganes, T.; Tyagi, S.; Grosset, J.; Riccardi, G.; Cole, S. T. Benzothiazinones kill *Mycobacterium tuberculosis* by blocking arabinan synthesis. *Science*, **2009**, *324* (5928), 801-4.
- [139] Tiwari, R.; Moraski, G. C.; Krchnak, V.; Miller, P. A.; Colon-Martinez, M.; Herrero, E.; Oliver, A. G.; Miller, M. J. Thiolates chemically induce redox activation of BTZ043 and related potent nitroaromatic anti-tuberculosis agents. *J. Am. Chem. Soc.*, **2013**, *135* (9), 3539-49.
- [140] Ross Kelly, T.; Kim, M. H.; Curtis, A. D. M. Structure correction and synthesis of the naturally occurring benzothiazinone BMY 40662. *J. Org. Chem.*, **1993**, *58* (21), 5855-5857.
- [141] Changeux, J. P. Nicotine addiction and nicotinic receptors: lessons from genetically modified mice. *Nat. Rev. Neurosci.*, **2010**, *11* (6), 389-401.
- [142] Gonzales, D.; Rennard, S. I.; Nides, M.; Oncken, C.; Azoulay, S.; Billing, C. B.; Watsky, E. J.; Gong, J.; Williams, K. E.; Reeves, K. R. Varenicline, an $\alpha 4\beta 2$ nicotinic acetylcholine receptor partial agonist, vs sustained-release bupropion and placebo for smoking

- cessation: a randomized controlled trial. *JAMA. J. Am. Med. Assoc.*, **2006**, *296* (1), 47-55.
- [143] Cooper, B. R.; Hester, T. J.; Maxwell, R. A. Behavioral and biochemical effects of the antidepressant bupropion (Wellbutrin): evidence for selective blockade of dopamine uptake *in vivo*. *J. Pharmacol. Exp. Ther.*, **1980**, *215* (1), 127-34.
- [144] Cooper, B. R.; Wang, C. M.; Cox, R. F.; Norton, R.; Shea, V.; Ferris, R. M. Evidence that the acute behavioral and electrophysiological effects of bupropion (Wellbutrin) are mediated by a noradrenergic mechanism. *Neuropsychopharmacology*, **1994**, *11* (2), 133-41.
- [145] Coe, J. W.; Brooks, P. R.; Vetelino, M. G.; Wirtz, M. C.; Arnold, E. P.; Huang, J.; Sands, S. B.; Davis, T. I.; Lebel, L. A.; Fox, C. B.; Shrikhande, A.; Heym, J. H.; Schaeffer, E.; Rollema, H.; Lu, Y.; Mansbach, R. S.; Chambers, L. K.; Rovetti, C. C.; Schulz, D. W.; Tingley, F. D., 3rd; O'Neill, B. T. Varenicline: an $\alpha 4\beta 2$ nicotinic receptor partial agonist for smoking cessation. *J. Med. Chem.*, **2005**, *48* (10), 3474-7.
- [146] Mazzocchi, P. H.; Stahly, B. C. Synthesis and pharmacological activity of 2,3,4,5,-tetrahydro-1,5-methano-1H-3-benzazepines. *J. Med. Chem.*, **1979**, *22* (4), 455-7.
- [147] Perez, E. G.; Mendez-Galvez, C.; Cassels, B. K. Cytisine: a natural product lead for the development of drugs acting at nicotinic acetylcholine receptors. *Nat. Prod. Rep.*, **2012**, *29* (5), 555-67.
- [148] West, R.; Zatonski, W.; Cedzynska, M.; Lewandowska, D.; Pazik, J.; Aveyard, P.; Stapleton, J. Placebo-controlled trial of cytisine for smoking cessation. *N. Engl. J. Med.*, **2011**, *365* (13), 1193-200.
- [149] King, A. C.; Cao, D.; O'Malley, S. S.; Kranzler, H. R.; Cai, X.; deWit, H.; Matthews, A. K.; Stachowiak, R. J. Effects of naltrexone on smoking cessation outcomes and weight gain in nicotine-dependent men and women. *J. Clin. Psychopharmacol.*, **2012**, *32* (5), 630-6.
- [150] Moulton, B. C.; Fryer, A. D. Muscarinic receptor antagonists, from folklore to pharmacology; finding drugs that actually work in asthma and COPD. *Br. J. Pharmacol.*, **2011**, *163* (1), 44-52.
- [151] Jih, K. Y.; Hwang, T. C. Vx-770 potentiates CFTR function by promoting decoupling between the gating cycle and ATP hydrolysis cycle. *Proc. Natl. Acad. Sci. U S A*, **2013**, *110* (11), 4404-9.
- [152] Kotera, J.; Mochida, H.; Inoue, H.; Noto, T.; Fujishige, K.; Sasaki, T.; Kobayashi, T.; Kojima, K.; Yee, S.; Yamada, Y.; Kikkawa, K.; Omori, K. Avanafil, a potent and highly selective phosphodiesterase-5 inhibitor for erectile dysfunction. *J. Urol.*, **2012**, *188* (2), 668-74.
- [153] Cutolo, M. The kinase inhibitor tofacitinib in patients with rheumatoid arthritis: latest findings and clinical potential. *Ther. Adv. Musculoskelet. Dis.*, **2013**, *5* (1), 3-11.
- [154] Papp, K. A.; Menter, A.; Strober, B.; Langley, R. G.; Buonanno, M.; Wolk, R.; Gupta, P.; Krishnaswami, S.; Tan, H.; Harness, J. A. Efficacy and safety of tofacitinib, an oral Janus kinase inhibitor, in the treatment of psoriasis: a Phase 2b randomized placebo-controlled dose-ranging study. *Br. J. Dermatol.*, **2012**, *167* (3), 668-77.
- [155] Sandborn, W. J.; Ghosh, S.; Panes, J.; Vranic, I.; Su, C.; Rouseff, S.; Niezychowski, W. Tofacitinib, an oral Janus kinase inhibitor, in active ulcerative colitis. *N. Engl. J. Med.*, **2012**, *367* (7), 616-24.
- [156] Vincenti, F.; Tedesco Silva, H.; Busque, S.; O'Connell, P.; Friedewald, J.; Cibrik, D.; Budde, K.; Yoshida, A.; Cohnen, S.; Weimar, W.; Kim, Y. S.; Lawendy, N.; Lan, S. P.; Kudlacz, E.; Krishnaswami, S.; Chan, G. Randomized phase 2b trial of tofacitinib (CP-690,550) in de novo kidney transplant patients: efficacy, renal function and safety at 1 year. *Am. J. Transplant.*, **2012**, *12* (9), 2446-56.
- [157] Bialer, M.; White, H. S. Key factors in the discovery and development of new antiepileptic drugs. *Nat. Rev. Drug Discov.*, **2010**, *9* (1), 68-82.
- [158] Martin, C. K.; Redman, L. M.; Zhang, J.; Sanchez, M.; Anderson, C. M.; Smith, S. R.; Ravussin, E. Lorcaserin, a 5-HT(2C) receptor agonist, reduces body weight by decreasing energy intake without influencing energy expenditure. *J. Clin. Endocrinol. Metab.*, **2011**, *96* (3), 837-45.
- [159] Sidjimov, A. K.; Tawara, J. N.; Stermitz, F. R.; Rithner, C. D. An isopavine alkaloid from *Thalictrum minus*. *Phytochemistry*, **1998**, *48* (2), 403-5.
- [160] Fuwa, H.; Sasaki, M. An efficient method for the synthesis of enol ethers and enecarbamates. Total syntheses of isoindolobenzazepine alkaloids, lennoxamine and chilenine. *Org. Biomol. Chem.*, **2007**, *5* (12), 1849-53.
- [161] Luettgen, J. M.; Knabb, R. M.; He, K.; Pinto, D. J.; Rendina, A. R. Apixaban inhibition of factor Xa: Microscopic rate constants and inhibition mechanism in purified protein systems and in human plasma. *J. Enzyme. Inhib. Med. Chem.*, **2011**, *26* (4), 514-26.
- [162] Svalo, J.; Nordling, J.; Bouchelouche, K.; Andersson, K. E.; Korstanje, C.; Bouchelouche, P. The novel β_3 -adrenoceptor agonist mirabegron reduces carbachol-induced contractile activity in detrusor tissue from patients with bladder outflow obstruction with or without detrusor overactivity. *Eur. J. Pharmacol.*, **2013**, *699* (1-3), 101-5.